

Depth distribution and ecological preferences of periphytic algae in Kenyir Lake, the largest tropical reservoir of Malaysia*

ROUF A. J. M. Abdur^{†,**}, PHANG Siew-Moi^{††}, AMBAK M. Azmi^{†††}

[†] Institute of Oceanography, Universiti Malaysia Terengganu, 21030 Kuala Terengganu, Malaysia

^{††} Institute Ocean and Earth Sciences, University of Malaya, 50603 Kuala Lumpur, Malaysia

^{†††} Department of Fisheries and Aquaculture, Universiti Malaysia Terengganu, 21030 Kuala Terengganu, Malaysia

Received May 13, 2009; revision accepted Dec. 22, 2009

© Chinese Society for Oceanology and Limnology, Science Press and Springer-Verlag Berlin Heidelberg 2010

Abstract We studied the depth distribution of periphyton, growing on inundated dead trees in Kenyir Lake, Malaysia in June 1995. The algal floral composition and structure manifested changes down the depth gradient in terms of species richness, abundance, diversity and cell density. In regression analysis, all these algal attributes were negatively correlated with the depth gradients at $P < 0.05$. In terms of species richness, the bacillariophytes showed dominance over the cyanophytes and chlorophytes; whereas with respect to standing crop, the cyanophytes showed dominance over the bacillariophytes and chlorophytes. The chlorophyll *a* was higher at the mid and bottom-depths than the surface-depth in both the downstream and upstream sites, which showed that vertical productivity or biomass accumulation was greater in low light irradiance. The product-moment correlation analysis showed that conductivity, turbidity, dissolved oxygen, reactive phosphate and ammonium-nitrogen were highly correlated with the algal assemblage data. However, photosynthetic active radiation (PAR) showed poor correlation with the community data. These observations have cast some light on the autoecological characteristics, habitat preferences and environmental responses of tropical periphytic communities.

Keyword: depth; periphytic algae; tropical; large reservoir; Kenyir Lake; community structure; biomass

1 INTRODUCTION

Classically, limnological studies have focused on pelagic habitats and few studies have been made on water column profiling of autotrophs, either phytoplankton or periphyton, with relation to their biodiversity, community structure and primary productivity in freshwater bodies. Vertical distribution of periphytic algae in freshwater bodies from temperate regions have been extensively studied (e.g. Loeb et al., 1981, Wulff et al., 2005, Cantonati et al., 2009) compared to their tropical counterparts.

Kenyir Lake or Tasik Kenyir (Fig. 1) is an artificial lake located in the state of Terengganu in northeast Malaysia created in 1985 by the damming of the Terengganu River to create the Sultan Mahmud Power Station. The depth profile study of Kenyir Lake is important in various ways. This is the largest reservoir not only in Malaysia but also in Southeast

Asia. The mean-depth of Kenyir Lake is of 37 m. Periphyton, being autotrophs, are part of the trophic structure of an aquatic ecosystem. Understanding of the depth distribution of microflora and their productivity is crucial for proper management of reservoirs, but as of yet, they remain unexplored in Southeast Asia, particularly in Malaysia. Depth profile studies of periphyton are also linked with biomonitoring, and biodiversity studies. Nutrient cycling, primary production, lake metabolism are also directly linked with the autotrophic dynamics of waterbodies. Theoretical limnology, focusing on light attenuation and photochemistry study, also requires the study of depth profile (e.g. Nakano et al., 2006). For Kenyir Lake, being a large-deep reservoir, understanding of its vertical profiling with periphyton and their ecological preferences is

* Supported by the Government of Malaysia, Intensified Research in Priority Areas (IRPA Project) (No. 50258-J3)

** Corresponding author: ajmrouf@gmail.com

important for developing sustainable fisheries in the context of overall lake management.

The objectives of the present study are firstly to investigate species composition, structure, diversity, ecological preferences, distribution patterns and biomass variations of periphytic algae down the depth gradient, and secondly to determine environmental descriptors that may have influence or impact on the periphytic assemblages in the water column.

2 MATERIALS AND METHODS

2.1 Description of the lake and sampling sites

Kenyir Lake is located between latitudes 4°40'N to 5°15'N and longitudes 102°32'E to 102°55'E in the state of Terengganu, the east coast of Peninsular Malaysia (Fig.1). This reservoir was created in 1986 to generate hydroelectric power and impounded two main rivers, River Terengganu and River Terengan (Furtado et al., 1977). The lake is very irregularly shaped and dendritic in nature. Its total water surface area is 36.900 h, with a mean-depth of 37 m and Top Water Level (TWL) of 145 m above mean sea level.

2.2 Sampling procedure

Innumerable trees, now dead, were flooded by the

impoundment. Periphyton have grown on their submersed surfaces to a depth of 10 m. Periphyton samples were taken from two sites, Dam side (Site 1) and Petang River (Site 2), representing downstream and upstream habitats of the lake, respectively. The description of lake and sampling sites are detailed in Rouf et al. (2008). Periphytic algae were collected from three depth levels by SCUBA (Self-contained Underwater Breathing Apparatus) diving on a single occasion in June, 1995. These depths were 0.5 m (surface), 5 m (middle) and 10 m (bottom) at the Dam side and 0.5 m (surface), 2.5 m (middle) and 5 m (bottom) at the Petang River. For the purpose of statistical analysis, each standing dead tree was considered as one sampling unit, and from each depth at least three sub-samples were taken to make one composite sample.

All samples were preserved in formalin acetic acid Alcohol (FAA) solution for taxonomical study and in 90% Acetone saturated with Magnesium carbonate for chlorophyll measurement. Chlorophyll *a* was determined using the trichromatic equation method (APHA, 1989) after overnight extraction at 4°C temperature.

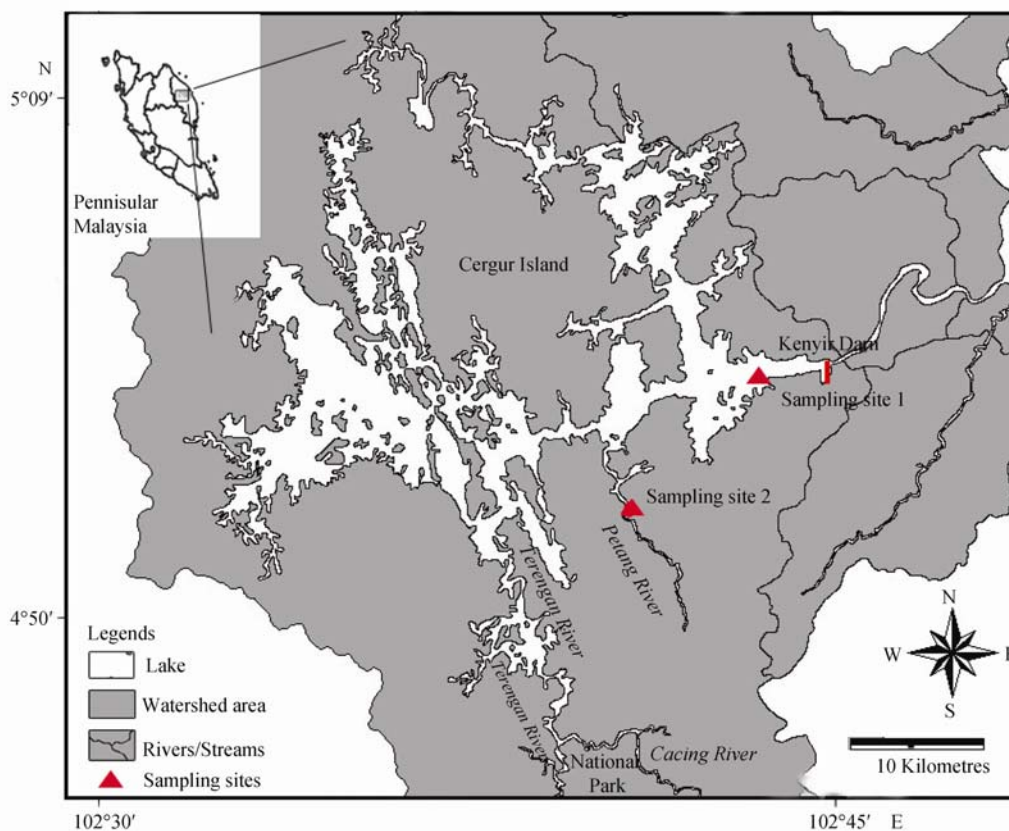


Fig.1 Map of Kenyir Lake showing location of the two sampling sites

Site 1: The dam side; Site 2: Petang River

In the periphytic assemblages, cyanophytes were mostly of filamentous type. The filaments (trichomes) were counted under the microscope; and later converted to cell number by multiplying with the respective *multiplication factor*, which is the average cell number per filament for each individual species or taxon determined by cell counts of at least 7–11 of filaments of the concerned species/taxon. The abundance of diatom taxa was calculated based on the monthly mean value of standing crop (cell density). Calculation of algal cell density per unit area of substrate was done by following APHA (1989).

2.2.1 Water samples

Water samples were collected in acid-washed 1 L polyethylene bottles. In the field, water chemistry data, such as pH, dissolved oxygen and secchi transparency were collected between 12.00 noon and 2.30 pm. Temperature, pH, conductivity and dissolved oxygen levels were measured by HYDROLAB Instrument (Microsonde model). Total dissolved solids (TDS) and turbidity were measured by hand-held TDS and Turbidity metres (DREL/2000) (Hach, 1993). Light was measured by a light meter (LI-COR, model Li-189) with an underwater quantum sensor. All these instruments were calibrated for their corresponding parameters during sampling. For all other chemical analysis, standard methods and procedures outlined in APHA (1989), Golterman et al. (1978) and Mackereth et al. (1978) were followed.

2.2.2 Identification of algae and numerical analysis

Algae were routinely examined and counted with a Nikon research microscope (OPTIPHOT) using $\times 40$ and $\times 100$ magnifications with oil immersion field objective lenses. For identification, reference books

and works of Pascher (1913b), Desikachary (1959), Prowse (1962), Patrick et al. (1966), Whiteford et al. (1973), Mizuno (1978), Ho (1979), Geitler (1985), Bourrelly (1981, 1985, 1990) and Yamagishi et al. (1991) were followed. Diatom nomenclature was updated in accordance with Round et al. (1990). Simple regression, product-moment correlation (Zar, 1984), species richness, abundance, Shannon-Wiener diversity index (H'), Pileuo's evenness and Jaccard similarity index (J') (Ludwig et al., 1988) were deduced for the community and environmental data.

3 RESULTS

The collected algal mats were approximately 1 mm thick with a thin cover of polysaccharide matrix on the top. The mats of the bottom-depth were more fragmented, loose and sparsely distributed on the tree trunks than those of the surface and mid-depth. Over all depths, a total of 177 periphytic algal taxa were found: diatoms (comprising 98 taxa), cyanophytes (61), chlorophytes (13), and dinoflagellates (5) (Appendix 1). In depth-wise distribution at the Dam side, 67 taxa were found at both the 0.5 and 5 m depths and 45 taxa at 10 m. In the Petang River, the total number of taxa found at each depth was 79, 64 and 37 at 0.5 m, 2.5 m and 5 m, respectively (Table 1).

Cyanophytes: The number of cyanophytes did not vary significantly between surface and mid-depths, but declined to one-fourth and one-third at the bottom-depth at the Sites 1 and 2, respectively (Table 1). All the cyanophyte taxa were filamentous except *Chroococcus* sp.; *Microcystis* sp.; *Synechococcus leptosira*; and *Gloeocapsa* sp.. Some taxa also showed depth exclusiveness. For example, *Anabaena torlusa* and *Anabaena variabilis* were found exclusively at 0.5 m depth. *Oscillatoria acuta* and *Oscillatoria amphigranulata*, *Camptylonema indicum*,

Table 1 Changes in species number and cell density of the periphytic algae found on standing dead trees at the three depth levels in June 1995 ($n=7$)

Depth level (m)	Number of taxa				Cell density*			
	Cyanop. (\pm SE)	Bacillario. (\pm SE)	Chloro. (\pm SE)	Whole assemb. (\pm SE)	Cyano. (\pm SE)	Bacillario. (\pm SE)	Chloro. (\pm SE)	Whole assemb. (\pm SE)
Dam side								
0.5	26 (2.10)	36 (1.87)	5 (0.35)	67 (3.7)	5 563 (137)	682 (58)	65 (7)	6 336 (147)
5	26 (2.76)	33 (2.14)	8 (0.75)	67 (3.9)	4 337 (158)	1 046 (95)	45 (7.5)	5 428 (152)
10	5 (1.3)	38 (2.39)	2 (0.12)	45 (2.58)	1 126 (95)	1 230 (115)	24 (5.3)	2 380 (110)
Petang River								
0.5	26 (2.3)	49 (1.74)	4 (0.33)	79 (3.6)	5 020 (174)	669 (70)	188 (24)	5 877 (125)
2.5	23 (2.5)	38 (1.89)	3 (0.35)	64 (3.2)	4 535 (185)	1 553 (115)	121 (12)	6 209 (134)
5	8 (0.86)	26 (1.78)	3 (0.54)	37 (2.8)	1 325 (85)	762 (96)	100 (8)	2 188 (110)

*cell numbers /mm² substrate

Pseudoscytonema malayense, *Scytonema multiramosa* and *Aulosira simplex* were confined to 5 m depth only. At the 10 m depth, only five taxa, viz. *Anabaena cylindrica*, *Haplosiphon* sp., *Lyngbya perelegans*, *Scytonema* sp. and *Tolypothrix* sp., were encountered.

In the Dam side, 14, 18 and 3 cyanophyte genera were encountered at the 0.5, 5 and 10 m depths, respectively. At the 0.5 m depth, *Anabaena* and *Lyngbya* were represented by four taxa each followed by *Phormidium* and *Oscillatoria* (two taxa each); and *Stigonema* and *Scytonema* (two taxa each). At the 5 m depth, *Lyngbya* were comprised of four taxa followed by *Scytonema* (three taxa), *Aulosira*, *Anabaena* and *Oscillatoria* (two taxa each). *Westiella* sp. and *Wollea saceta* were confined exclusively to the 5 m depth.

In the Petang River, a total of 16 cyanophyte genera were encountered each at the surface and middle depths whereas the bottom depth contained only five taxa. Among them, *Lyngbya* was represented by 6 taxa followed by *Phormidium* (four taxa) and *Anabaena* (three taxa). The remaining 13 genera were each comprised of one taxon. The bottom depth taxa were *Lyngbya*, *Phormidium*, *Anabaena* two taxa each; and *Scytonema* and *Schizothrix* one taxon each.

Bacillariophytes: The species richness of the bacillariophytes was almost static along the depths in the Dam side, whereas it gradually declined from upper to lower depths in the Petang River. Here, the number of taxa at the bottom-depth (5 m) was nearly half that of the surface-depth (0.5 m). The cell density varied with depth, gradually increasing from the surface to the bottom depths in the Dam side, while in the Petang River, the mid-depth possessed the highest cell density (Table 1).

In terms of genera, bacillariophytes did not vary widely with depth, however, their species composition varied in the Dam side. Here, 15 genera were found at both the 0.5 m and the 5 m depths and 14 at the bottom (10 m) depth. At the surface-depth (0.5 m), *Navicula* was comprised of seven taxa, followed by *Pinnularia* (four taxa), *Amphora*, *Cymbella*, *Eunotia*, *Frustulia*, *Gomphonema* (three taxa each) and *Achnanthes*, *Neidium* (two taxa each). At the middle-depth (5 m), the genus containing highest number of taxa shifted to *Pinnularia*, with seven taxa, followed by *Gomphonema* and *Eunotia* (four taxa each), *Frustulia*, *Navicula* (three taxa each) and *Cymbella* (two taxa). *Stauroneis pusilla* was exclusively found at the mid-depth in both the sites.

At the bottom depth (10 m), *Eunotia* possessed highest number of taxa (seven) followed by *Navicula* and *Pinnularia* (six taxa each), and *Frustulia* and *Gomphonema* (3 taxa each).

In the Petang River (Site 2), the total numbers of genera were 17, 16 and 11 at the 0.5 m, 2.5 m and 5 m depths, respectively. Like the Dam side, the taxa comprising each genus varied widely. At the surface-depth, *Pinnularia* had seven taxa, followed by *Eunotia* and *Cymbella* (6 each). At the middle-depth, *Navicula* comprised six taxa followed by *Cymbella* and *Achnanthes* (5 taxa each) and *Pinnularia* (three taxa). The taxa composition at the bottom depth was similar to that of the middle-depth. At the bottom depth (5 m), *Cymbella* and *Achnanthes* contained five taxa each followed by *Navicula* and *Gomphonema* (three taxa each) then *Eunotia* and *Pinnularia* (2 taxa each).

Chlorophytes: Chlorophyta was a minor group in terms of both species richness and standing crop (cell density) in this reservoir. Any trends of variation in their vertical distribution were also negligible due to their poor presence. This class constituted only 5%–6% of the total taxa and 1%–3% of the total standing crop of the whole assemblage.

In terms of species richness, diatoms (bacillariophytes) dominated (ranging from 49 to 85% of all species) followed by cyanophytes (ranging from 11% to 39%) and chlorophytes (ranging from 4% to 12%) (Fig.2). The Jaccard similarity indices showed low similarity of floral composition between the depths: the maximum similarity was found to be only 35% (Table 2).

Species abundance

Cyanophyta: The species abundance at different depths varied considerably. In the Dam side, the dominant cyanophyte taxa at the 0.5 m depth, in order of abundance, were: *Schizothrix lamyi*, *Anabaena cylindrica*, *Oscillatoria splendida*, *Microcoleus* sp., *Lyngbya perelegans*, *Scytonema fremyi*, *Lyngbya porphyrosiphonis*, *Stigonema minutum*, and *Fischrella moniliformis* (Fig.3). These nine taxa contributed 80% of total cyanophyte cells. At the mid-depth (5 m), dominance shifted, the dominant taxa being *Scytonema multiramosa*, *Lyngbya perelegans*, *Haplosiphon* sp., *Lyngbya* sp., *Anabaena cylindrica*, *Schizothrix lamyi*, *Microcoleus* sp., *Aulosira bombayensis* and *Lyngbya porphyrosiphonis*. At the 10 m depth, only five taxa were encountered: *Anabaena cylindrica*, *Haplosiphon* sp., *Lyngbya perelegans*, *Scytonema* sp.

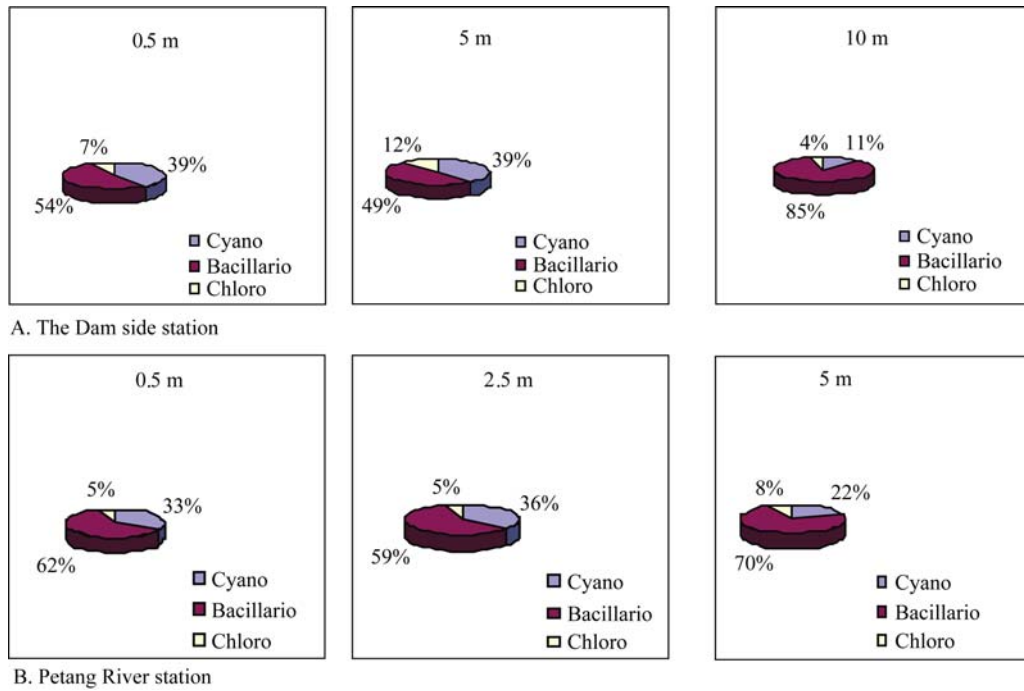


Fig.2 Compositional proportions of algae on natural substrates at different depths in June, 1995

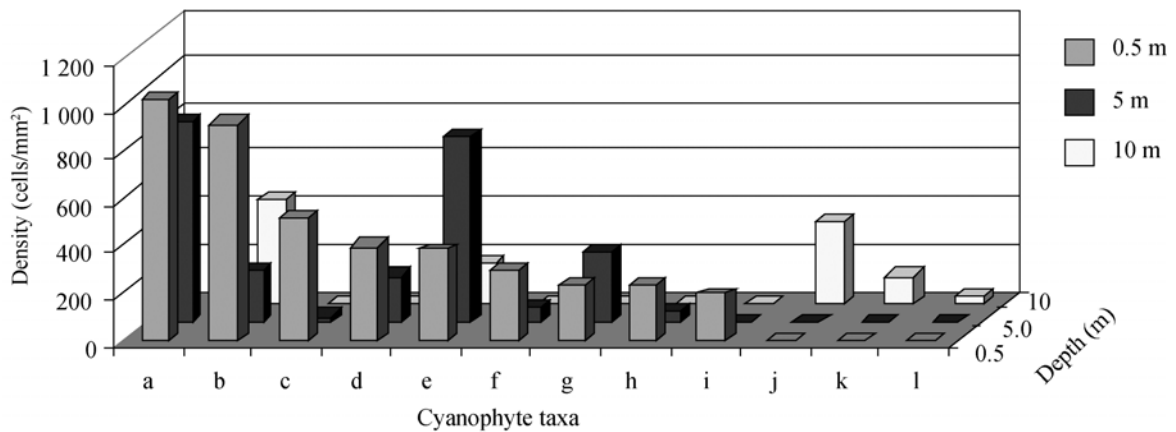


Fig.3 Variation of abundance with depth of the dominant cyanophytes found on natural substrates at the Dam side station

Legend: a. *Schizothrix lamyi*; b. *Anabaena cylindrical*; c. *Oscillatoria splendida*; d. *Microcoleus* sp.; e. *Lyngbya perelegans*; f. *Scytonema fremyi*; g. *Lyngbya porphyrosiphonis*; h. *Stigonema minutum*; i. *Fischerella moniliformis*

Table 2 Jaccard Similarity Indices (C_j) comparing floral assemblages between depths

Station	Floral group	Surface-depth versus mid depth	Mid-depth versus bottom-depth	Bottom-depth versus surface-depth
Dam side station	Cyanophytes	0.35	0.27	0.25
	Bacillariophytes	0.31	0.31	0.16
Petang River station	Cyanophytes	0.32	0.25	0.15
	Bacillariophytes	0.31	0.11	0.05

and *Tolypothrix* sp. In the Petang River, the most abundant cyanophyte taxa were *Oscillatoria splendida*, *Anabaena cylindrical*, *Haplosiphon* sp., *Schizothrix brauni*(?), *Phormidium purpureum*, *Fischerella moniliformis*, *Anabaena circinalis* and

Scytonema sp. at the 0.5 and 2.5 m depths, while *Anabaena cylindrical*, *Phormidium purpureum* and *Phormidium corium* were found at the 5 m depth.

Bacillariophyta: Bacillariophytes also showed variations in abundance with depth. At the Dam side,

at 0.5 m depth, the dominant taxa were *Frustulia saxonica*, *Frustulia rhomboides*, *Encyonema gracilis*, *Fragilaria construens*, *Navicula cryptocephala*, *Frustulia vulgaris*, *Gomphonema longiceps*, *Pinnularia parva*, *Neidium productum*, *Eunotia lunaris* and *Navicula radiosa*. They comprised 70% of the total diatom cell count at this depth. At the 5 m depth, dominance shifted to *Navicula cryptocephala*, *Tabellaria fenestrata*, *Fragilaria construens*, *Frustulia saxonica*, *Encyonema gracile*, *Pleurosigma elongatum*, *Achnanthes linearis*, *Gomphonema gracile*, *Gomphonema parvulum*, and *Navicula radiosa*. The dominance again shifted at the 10 m depth, the representative taxa being *Navicula cryptocephala*, *Navicula radiosa*, *Fragilaria construens*, *Eunotia lunaris*, *Eunotia sudetica*, *Eunotia monodon*, *Eunotia praeurupta*, *Nitzschia rhomboides*, *Tabellaria fenestrata*, *Eunotia major v. indica*, and *Gomphonema parvulum*.

In the Petang River, the dominant taxa at the surface-depth (0.5 m) were *Encyonema gracilis*, *Cymbella minuta*, *Navicula radiosa*, *Cymbella ventricosa*, *Gomphonema longiceps*, *Navicula feuerborni*, *Gyrosigma* sp., *Frustulia rhomboides*, *Pleurosigma elongatum*, *Gomphonema parvulum* and *Navicula gracilis* (Fig.4). At the mid-depth, the dominance shifted to *Navicula radiosa*, *Gyrosigma* sp., *Gomphonema parvulum*, *Navicula gracilis*, *Navicula feuerborni*, *Neidium affinis*, *Achnanthes inflata*, *Achnanthes tropica*, *Fragilaria construens* and *Gomphonema gracile*. At the bottom depth, the dominance again shifted to *Achnanthes crenulata*,

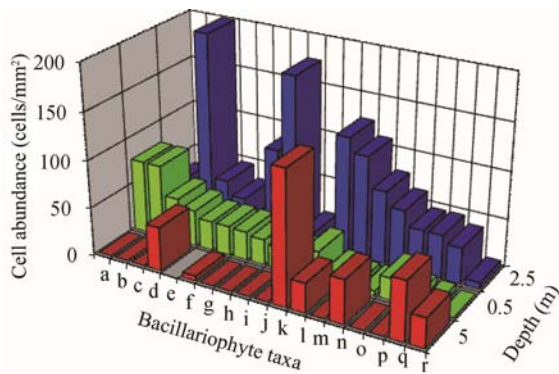


Fig.4 Variation with depth in species abundance of the dominant diatom taxa found on natural substrates at the Petang River station in June, 1995

In the figure the depth order is changed for the sake of better visualization. a. *Encyonema gracilis*; b. *Cymbella minuta*; c. *Navicula radiosa*; d. *Gomphonema gracile*; e. *Cymbella ventricosa*; f. *Gomphonema longiceps*; g. *Navicula feuerborni*; h. *Gyrosigma* sp.; i. *Frustulia rhomboides*; j. *Pleurosigma elongatum*; k. *Gomphonema parvulum*; l. *Navicula gracile*; m. *Neidium affinis*; n. *Achnanthes inflata*; o. *Achnanthes tropica*; p. *Fragilaria construens*; q. *Achnanthes crenulata*; r. *Cocconeis placentula*

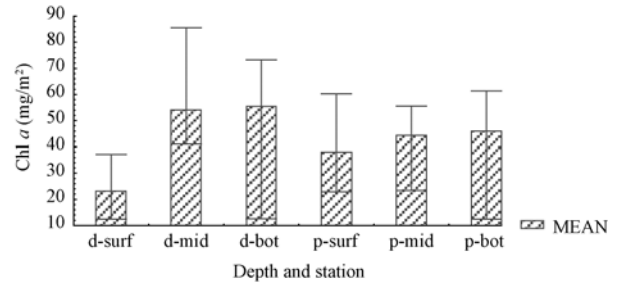


Fig.5 The variation in chlorophyll a concentration of the periphytic algae on standing dead trees at the different depths at the Dam side and Petang River stations, collected in June 1995

Legends: D = Dam side. P = Petang River. Bar = Standard deviation

Achnanthes inflata, *Achnanthes linearis*, *Achnanthes tropica*, *Cocconeis placentula*, *Cyclotella kuetzingiana*, *Cymbella ehrenbergii*, *Cymbella turgida*, *Cymbella javanica*, *Cymbella tumida*, and *Cymbella ventricosa*.

Species diversity: As with taxa richness, the Shannon-Wiener diversity indices (H') of the assemblages decreased from upper to lower depths. The maximum H' value was 3.36. Classwise, diatoms were more diverse than cyanophytes and chlorophytes, the latter being marginalized in the whole assemblages (Table 3).

Chlorophyll a biomass: In both sites, chlorophyll a was found to be much higher at the mid and bottom depths than at the surface depth. Chlorophyll a varied widely at the bottom depth, ranging from 225.7 to 580 mg/m² at the Dam side and from 110 to 610 mg/m² in the Petang River (Fig.5).

Statistical analysis: Different algal attributes, such as species richness, diversity and cell density were regressed against the depth profile. All of the community attributes were found to be highly correlated with the depth gradient at $P < 0.05$ level (Table 4).

Environmental parameters: A total of nine water chemistry and limnological parameters were measured down the depths (Table 5). pH ranged between 6.85 and 7.12, water temperature ranged between 29.05°C and 31.3°C. Conductivity ranged between 25 and 28 μS/cm. PAR ranged between 78.00 and 1781 μmol/m² s in the Dam side, and between 112.78 and 526.3 μmol/m² s in the Petang River. Results of the product-moment correlation analysis between species number and cell density of periphytic algae on standing trees and environmental parameters are cited in Table 6.

Table 3 Number of taxa (*S*), species diversity (*H'*) and evenness (*J'*) of the periphyton assemblages at different depths in June, 1995 (*n*=7)

Depth (m)	Cyanophytes			Bacillariophytes			Chlorophytes			Whole assemblage		
	<i>S</i>	<i>H'</i>	<i>J'</i>	<i>S</i>	<i>H'</i>	<i>J'</i>	<i>S</i>	<i>H'</i>	<i>J'</i>	<i>S</i>	<i>H'</i>	<i>J'</i>
St. 1: Dam side												
0.5	26	2.65	0.81	36	3.21	0.90	5	0.92	0.57	67	3.11	0.74
5	26	2.46	0.76	33	2.68	0.77	8	1.58	0.76	67	3.1	0.74
10	5	1.36	0.98	38	3.23	0.89	2	0.56	0.81	44	3.00	0.79
St. 2: Petang River												
0.5	26	2.6	0.80	49	3.36	0.86	4	0.82	0.59	79	3.13	0.71
2.5	23	2.5	0.82	38	3.11	0.85	5	0.97	0.62	64	2.95	0.74
5	8	1.69	0.87	26	2.99	0.92	3	0.65	0.64	37	2.67	0.75

Table 4 Results of the linear regression between the different algal attributes and the different depth levels (*n*=7; *P*<0.05)

Dependent variables	Independent variables	<i>r</i>	<i>Y</i>
Species Nos.- Dam	Different depth levels	-0.87	$y = 70.95 - 2.32x$
Species Nos.-PR	Different depth levels	-0.99	$y = 81.87 - 8.49x$
Total cell Nos.-Dam	Different depth levels	-0.96	$y = 6711 - 397x$
Total cell Nos.-PR	Different depth levels	-0.83	$y = 6640 - 748x$
Species diversity Dam	Different depth levels	0.91	$y = 3.125 - 0.0111x$
Species diversity-PR	Different depth levels	0.99	$y = 3.15 - 0.093x$

Table 5 Ambient environmental data at the different water depths in June, 1995

Parameters	Depth level (Dam side)			Depth level (Petang River)		
	0.5 m	5 m	10 m	0.5 m	2.5 m	5 m
pH	7.12	7.20	7.10	6.70	7.00	6.85
Temperature (°C)	31.3	30.15	29.98	29.98	29.88	29.5
Conductivity (μS/cm)	26.4	25.00	26.7	26.00	26.00	28.00
DO (mg/L)	7.84	7.64	6.96	7.35	7.24	6.56
Turbidity (NTU)	1.45	1.46	1.51	1.30	1.32	1.34
Reactive phosphate (mg/L)	0.11	0.16	0.18	0.03	0.04	0.04
Ammonia-N (mg/L)	0.04	0.03	0.06	0.07	0.06	0.09
Nitrate-N (mg/L)	0.5	0.4	0.6	0.2	1.00	0.8
Nitrite-N (mg/L)	0.003	0.035	0.003	0.011	0.01	0.01
PAR (μmol/m ² s)	1 781	338	78.20	526.3	242.3	112.78

Table 6 Results of the product-moment correlation analysis between species number and cell density of periphytic algae of standing trees and environmental parameters found at the different depth levels in the Dam side (*P*<0.05 level)

Variables	Cyano. taxa	Bacilla. taxa	Chloro. taxa	Cyano.-cell	Bacilla. cell	Chloro. cell	Whole assem. taxa	Whole assem. cell
pH	0.65	-0.62	0.90	0.43	0.00	0.20	0.50	0.47
Temperature(°C)	0.060	0.90	0.10	0.79	0.98	0.91	0.70	0.76
Conductivity(μS/cm)	-0.94	-0.50	-0.70	1.00	0.93	0.98	1.00	0.99
DO(mg/L)	0.98	0.40	0.70	1.00	0.88	0.96	1.00	1.00
Turbidity (NTU)	-0.99	0.34	0.80	0.99	0.85	0.94	1.00	1.00
Reactive phosphate (mg/L)	-0.72	0.82	0.30	0.88	1.00	0.97	0.80	0.85
Ammonia-N(mg/L)	0.94	0.14	1.00	0.82	0.50	0.67	0.90	0.85
Nitrate-N(mg/L)	-0.87	0.33	1.00	0.70	0.33	0.51	0.80	0.74
Nitrite-N(mg/L)	0.50	0.76	0.90	0.25	0.19	0.01	0.40	0.30
PAR(μmol/m ² s)	0.62	0.89	0.10	0.81	0.98	0.92	0.70	0.77

4 DISCUSSION

This study has revealed some interesting characteristics and attributes of periphytic assemblages down the depth gradient of the Kenyir Reservoir, which has an average depth of 37 m and is a mesotrophic waterbody.

The euphotic zone in Kenyir Lake extends to 15 m depth while the lake littoral depth (Z) was found to be 19 m in June, 1995. The current study showed that periphytic algae did not grow beyond 10 m depth, at which the light irradiance was found to be $78 \mu\text{mol}/\text{m}^2 \text{ s}$. Periphyton assemblages showed gradual reduction with depth in terms of both in species richness and abundance. The species composition also varied considerably, which was depicted by the Jaccard similarity indices (Table 2). The linear regression results (Table 4) showed that species richness, diversity, and standing crop (dependent variables) and the different depth levels (independent variables) were significantly correlated at the $P < 0.05$ level. This signifies that the algal communities have their habitat or ecological preferences along the environmental gradients that are vertically stratified. In other words, they have their own niche occupancies being driven by their inherent autoecological characteristics or behavior. Depth related reduction of the algal assemblages in other large freshwater bodies have also been reported. For example, in Lake Michigan, shallow, mid and deep zones were differentiated on the basis of species composition, abundance and diversity (Stevenson et al., 1981). Another example is Lake Baikal, which showed zonation of benthic algal distribution with depth (Nozaki et al., 2002).

The inter-community composition showed that diatoms thrived better than blue-greens and greens down the depth gradient. In terms of species richness, diatoms dominated at all three depths. In the Dam side, their cell densities were higher at the deep and mid depths (though much lower than that of the cyanophyta at the surface). The situation was similar in the Petang River, with the exception that diatom cell density was highest at the mid-depth. In India, periphyton of submersed plant parts and of glass slides in deep water rice fields showed vertical variations where the bottom and surface levels were dominated by bacillariophytes and chlorophytes, respectively (Das et al., 1994). Poulickova et al. (2006) found that diatom species richness was higher at an oligotrophic site and that of cyanobacteria were often available in the eutrophic fish ponds. The

species richness of diatoms may be linked to the trophic status of Kenyir Lake, it being a mesotrophic water body.

Some interesting information has also been revealed about the habitat or autoecological preferences of cyanophytes. *Schizothrix lamyi* showed a tendency to decline from surface to bottom depths. *Anabaena cylindrica* and *Scytonema fremyii* showed non-rhythmic or heterogeneous abundance in all three depths. *Oscillatoria splendida* was found in negligible amounts at the surface and mid depths. *Fishchrella moniliformis* was only present at the surface-depth. *Lyngbya porphyrosiphonis* was at almost equal abundance at the surface and mid depths, but was absent at the bottom-depth. The trend of *Stigonema minutum* was the same as *Oscillatoria splendida*, whereas *Haplosiphon* sp. and *Lyngbya perelegans* were more abundant at the mid-depth. The oligotrophic lake of Taupo in New Zealand showed three periphytic zones: surface (0–2 m), mid water (2–20 m) and deep (>20 m) where diatoms (*Aulacoseira granulata*, *Rhopalodia novaezealandiae*, *Epithemia sorex*, and *Fragilaria* spp.) dominated the deep assemblage, while cyanophytes, (*Tolypothrix tenuis* and *Mastogloia elliptica*) dominated the mid-depth and *Scytonema*, *Dichothrix*, and filamentous chlorophytes dominated the shallow zone (Hawes et al., 1994). This type of zonation was apparently absent in Kenyir Lake.

In this study, some diatom taxa also showed clear depth preferences. For example, *Gomphonema parvulum* gradually increased from surface to bottom depths, while *G. longiceps* showed the opposite trend to that of *G. parvulum*. It is pertinent to mention that *G. parvulum* was one of the most dominant taxa in the monsoon period (December) when the lake water in Kenyir shifted towards slightly acidic. In the dry period (June), when the lake water became more alkaline, its abundance declined considerably (Rouf, 1997). Another gomphonoid diatom, *Gomphonema gracile* did not show significant change down the depth gradient in the present study. *Cymbella gracilis* gradually decreased from the surface to the mid-depth and was absent at the bottom-depth. *Navicula radiosia* and *Gyrosigma* sp. were dominant in the mid-depth but absent in the bottom-depth, whereas *Navicula gracilis* was present in all the depths and was more dominant in the mid-depth. These phenomena show conspicuous ecological preferences for habitats by individual diatom taxa or species.

Some interesting trends in genera-wise diatom distribution were also revealed in this study. In the

Dam side, at the surface-depth (0.5 m), the dominant genus in terms of species number was *Navicula*, whereas in the mid-depth the dominance shifted to *Pinnularia*, and at the 10 m (bottom depth), to *Eunotia*. *Navicula* is a eurytopic genus; *Pinnularia* and *Eunotia* are mostly acidophilic genera. The trend of decreasing pH with depth coincided with this depth distribution. However, in Petang River, the surface-depth was dominated by *Pinnularia* and *Gomphonema* and the deeper two depths by *Navicula*, *Cymbella* and *Achnanthes*. The light irradiance was less in Petang River, being only one-third to that of the Dam side (Table 5). This would probably have some effect on depth distribution of diatoms. Diatoms have shown conspicuous depth preferences in other studies also. They migrate in response to changes in ambient microhabitat to find their preferred environment. For example, in an experiment in Lake McCaughy, a large reservoir in Nebraska, USA, diatoms responded consistently to switching with manipulation of clay tiles. Out of the 32 most common taxa, 26 showed significant upper (10) or lower (16) depth choices (Hoagland et al., 1990). In another experiment, Mitbavkar et al. (2004) demonstrated that irradiance has a greater role than tide in controlling microscale migration of diatoms. In the temperate environment of Finland, Wulff et al. (2005) found that in the sediment of a microtidal fjord, pelagic forms of diatoms inhabit the deeper locations, while epipelagic and epipsammic forms inhabit the shallower sites.

Many important environmental parameters, such as turbidity, dissolved oxygen, conductivity, reactive phosphate levels and ammonium-nitrogen levels have contributory effects on the depth distribution and growth of periphyton in the Kenyir Lake (Table 6). However PAR showed poor relationship with the whole assemblage data. Wulff et al. (2005) found that light is not the only important environmental factor but rather a combination of temperature, salinity and light explain 57% of the diatom taxa variation in redundancy analysis (RDA). However, Hoagland et al. (1990) found that light regime and wave disturbance were the main controlling factors for vertical zonation of freshwater epilithic algae.

Cells adapted to high light intensity have lower chlorophyll *a* content per cell than those adapted to low light intensities (Wetzel, 1983). Vertical patterns of primary production in Kenyir corroborated this phenomenon. The ecological effects of light and temperature on the photosynthesis and the growth of algae are inseparable because of interrelationships

between metabolism and light saturation. Generally, the intensity of light required to saturate algal photosynthesis increases as water temperature increases. In Kenyir, in the Dam side, the light-irradiance levels were 1 781, 338 and 78 $\mu\text{mol}/\text{m}^2 \text{ s}$ at 0.5 m, 5 m and 10 m depths, respectively (Table 5). Here, the chlorophyll *a* at the mid and bottom depths was more than double that of the surface-depth. In Petang River, light-irradiance values were almost half those of the Dam side being 569.3; 287 and 183 $\mu\text{mol}/\text{m}^2 \text{ s}$ at 0.5 m, 2.5 m and 5 m depths, respectively (Table 9). The chlorophyll *a* values at the mid and bottom depths were slightly higher than that of the surface depth. Nutrients are locked up in the hypolimnion due to thermal stratification in Kenyir Lake (Yusoff et al., 1994). This might also have an impact on primary production in the Dam side.

Horizontal heterogeneity in lakes may also have some effects on vertical distribution or heterogeneity of algal assemblages. Tilzer (1990) emphasized that spatially variable inputs of energy from the overlying atmosphere, in the forms of irradiance and kinetic energy, as well as differences in atmospheric pressure, can cause significant horizontal heterogeneity in large lakes. The Dam side and Petang River sampling sites are, respectively, typical representatives of lentic and lotic habitats in a reservoir ecosystem (Thornton et al., 1990). Petang River (Site 2) is a shallow, narrowed water-channel shaded by the tropical rainforest. The water was more turbid and irradiance was also less than that of the Dam side. The Dam side site is in the main basin of the reservoir with a wide open pelagic zone and with a steep littoral area. These physiogeographic differences may have contributory effects on periphytic algal assemblages.

5 CONCLUSION

In this study, periphytic algae showed significant differences in their depth distribution, which reflects on their niche or habitat preferences down the water column. It was also evident that vertical productivity or biomass accumulation was greater in low light-irradiance. However, depth related trends or distribution patterns of periphytic assemblages warrant further, more extensive (both vertically and horizontally) studies in the reservoir. This information may be immensely helpful in developing sustainable fisheries and for overall lake management particularly in the context of apprehended vulnerability of aquatic ecosystems due to climate changes.

References

- APHA. 1989. Standard Methods for the Examination of Water and Waste Water. 17th Edition. American Public Health Association, Washington D.C. USA.
- Bourrelly P. 1990. Les Algues d'eau Douce, Initiation a la systematique. Tome I. Les algues vertes. Revised ed. Socie'te' Nouvelle des e'ditions Boube'e, Paris.
- Bourrelly P. 1981. Les Algues d'eau Douce, Initiation a la systematique. Tome II. Les algues jaunes et brunes. Revised ed. Socie'te' Nouvelle des e'ditions Boube'e, Paris.
- Bourrelly P. 1985. Les Algues d'eau douce. Initiation a la systematique. Tome III. Les algues bleues et rouges, les Eugle'niens, peridiniens et Cryptomonadines. Ed. N. Socie'te' Nouvelle des e'ditions Boube'e, Paris.
- Cantonati M, Scola S, Angeli N, Guella G, Frassanito R. 2009. Environmental controls of epilithic diatom depth-distribution in an oligotrophic lake characterized by marked water-level fluctuations. *Eur. J. Phycol.*, **44**(1): 15-29.
- Das D N, Mitra K, Mukhopadhyay P K, Chaudhuri D K. 1994. Periphyton of the deepwater rice field at Pearpur Village, Hooghly, west Bengal, India. *Environment and Ecology, Kalyani*, **12**(3): 551-556.
- Desikachary T V. 1959. *Cyanophyta*. Indian Council Agri.Res. New Delhi. 686p.
- Furtado J, Soepadmo I E, Sasekumar A, Lim R P, Ong S, Davidson S L, Liew K S. 1977. Ecological Effects of the Terengganu Hydro-Electric Project (Kenyer Project). Wallaceana, Suppl. 1, p. 51.
- Geitler L. 1985. Dr. Rabenhorst's Kryptogamen-Flora. Reprint 1985. Koeltz Scientific Books. D-6240 Koenigstein/W. Germany. 1 196p.
- Goltermann H L, Clymo R S, Ohnstand M A M. 1978. Method for Physical and Chemical Analysis of Freshwaters. IBP Handbook No. 8. 2nd ed., Blackwell, Oxford.
- Hach Co. 1993. Water Analysis by Spectrophotometer. Hach Co. Loveland, Colorado.
- Hawes I, Smith R. 1994. Seasonal dynamics of epilithic periphyton in oligotrophic Lake Taupo, New Zealand. *New Zealand Journal of Marine and Freshwater Research*, **28**(1): 1-12.
- Ho S C. 1979. Structure, species diversity and primary productivity of epiphytic algal communities in the Schohsee (Holosteen), West Germany. Unpublished Ph.D. Dissertation, Max-Plank Institut fur Limnologie, Universitat Kiel.
- Hoagland K D, Peterson C G. 1990. Effects and wave disturbance on vertical zonation of attached microalgae in a large reservoir. *J. Phycol.*, **26**: 450-457.
- Loeb S L, Reuter J. 1981. The epilithic periphyton community: a few lake comparative study of community productivity, nitrogen metabolism and depth distribution of standing crop. *Verh. Int. Verein Limnol.*, **21**: 346-352.
- Ludwig J A, Reynolds J F. 1988. Statistical Ecology -a Primer on Methods and Computing. John Wiley, NY.
- Mackereth F J H, Heron J, Talling J F. 1978. Water Analysis: Some Revised Methods for Limnologists. FBA scientific Publication No. 36. Freshwater Biological Association, Cumbria, U.K.
- Mitbavkar S, Anil A C. 2004. Vertical migratory rhythms of benthic diatoms in tropical intertidal sand flat: influence of irradiance and tide. *Marine Biology*, **145**: 9-20.
- Mizuno T. 1978. Illustrations of the freshwater plankton of Japan. 2nd ed. Hoikusha Publishing Co. Ltd.
- Nakano Shin-ichi, Takeshita A, Ohtsuka T. 2006. Vertical profiles of current velocity and dissolved oxygen saturation in biofilms on artificial and natural substrates. *Limnology*, **7**: 213-218.
- Nozaki K, Morino H, Munehara H, Sideleva V G, Nakai K, Yamaguchi M, Kozhova O M, Nakanishi M. 2002. Composition, biomass and photosynthetic activity of the benthic algal communities in a littoral zone of lake Baikal in summer. *Limnology*, **3**: 175-180.
- Pascher A. 1913b. Die Susswasser-flora Deutschlands, Osterreichs und der Schweiz. Heft 10. Bacillariales (Diatmeen). Jena Verlag von Gustav- Fisher. 187p.
- Patrick R, Reimer C W. 1966. The diatoms of the United States exclusively of Alaska & Hawaii. Vol. 1 Philadelphia.
- Patrick R, Reimer C W. 1966. The diatoms of the United States exclusively of Alaska and Hawaii. Vol. 1 Monogr. Acad Nat. Sci. Philadelphia. No. 13.
- Pouličková A, Kitner M, Hašler P. 2006. Vertical distribution of attached algae in shallow fishponds of different trophic status. *Versita*, **61**(1), 1-9.
- Rouf A J M Abdur. 1997. Ecology and temporal changes in algal composition and spatial distribution of periphyton community of a drowned tropical forest reservoir in Malaysia. Ph.D dissertation. Universiti Putra Malaysia, Serdang, Malaysia. (unpublished).
- Rouf A J M, Ambak A M A, Lokman Shamsudin, Phang Siew-Moi, Ho S C. 2008. Periphytic algae of Kenyer Lake, Malaysia Temporal changes in the periphytic algal communities in a drowned tropical forest reservoir in Malaysia: Kenyer Lake. *Lakes & Reservoirs: Research and Management*, **13**: 271-287.
- Round F E, Crawford R M, Mann D G. 1990. The Diatoms-Biology and Morphology of the Genera. Cambridge Univ. Press.
- Stevenson R J, Stoermer E F. 1981. Quantitative differences between benthic algal communities along depth gradient in Lake Michigan. *J. Phycol.*, **17**: 29-36.
- Thornton K W, Kimmel B L, Payne F F. 1990. Reservoir Limnology: Ecological Perspectives. John Wiley & Sons Inc.
- Tilzer M M. 1990. Specific Properties of Large Lakes. In: Tilzer M M, Serruya C ed. Large Lakes-Ecological Structure and Function. Springer-Verlag, Berlin.
- Wetzel R G. 1983. Limnology. 2nd edition. Saunders College

- Publishing, Philadelphia.
- Whiteford L A, Schumacher G J. 1973. A manual of Freshwater Algae. Sparks Press.
- Wulff A, Vibaste S, Truu J. 2005. Depth distribution of photosynthetic pigment and diatoms of a microtidal fjord. *Hydrobiologia*, **534**: 117-130.
- Yamagishi T, Kanetsuna Y. 1991. Phytoplankton from Malaysia. *Gen. Edu. Rev., Colle. Agr. & Vet. Med., Nihon Univ.*, **27**: 137-151.
- Yusoff F M, Lock M A. 1994. Thermal stratification and its role in controlling eutrophication in a tropical reservoir, Malaysia. Proceedings of the Internat. Tropical Limnology Conf., Indonesia.
- Zar J H. 1984. Biostatistical Analysis, 2nd edition. Prentice-Hall, Inc., Englewood Cliffs NJ.

APPENDIX-1

List of periphytic algae found at all depths in both the Dam side and the Petang River sites in Kenyir Lake, Malaysia in June 1995

CYANOPHYTES (61 taxa)			
<i>Anabaena circinalis</i>	Rabenh	<i>Oscillatoria splendida</i>	Grev
<i>Anabaena cylindrica</i>	Lemmermann	<i>Phormidium calcicola</i>	Gardner
<i>Anabaena elliptica</i>	Lemmermann	<i>Phormidium corium</i>	(Ag.) Gom.
<i>Anabaena gelatinicola</i>	Ghosh (after Ghosh)	<i>Phormidium mucicola</i>	Hub. - Pest. and
<i>Anabaena torulosa</i>	(Carm) Lagerh. ex. Born et Flah	<i>Phormidium mucosum</i>	Gardner
<i>Anabaena variabilis</i>	Kütz	<i>Phormidium purpurscens</i>	Kütz.
<i>Anabaenopsis elenkinii</i>	Miller	<i>Phormidium valderianum</i> (?)	(Delp.) Gom.
<i>Aphanizomenon flos-aquae</i>	Ralfs	<i>Pseudoanabaena</i> sp.	Koppe
<i>Aulosira bombayensis</i>	Gonzalves	<i>Pseudoscytonema malayense</i>	Biswas
<i>Aulosira simplex</i>	Bharadw	<i>Rivularia</i> sp.	(Roth) Ag.
<i>Calothrix fusca</i>	(Kutx) B and F	<i>Schizothrix braunii</i>	(A. Br.) Gomont
<i>Camptylonema indicum</i>	Schmidle	<i>Schizothrix lamyi</i>	Gomont
<i>Chroococcus</i> sp.	Nag	<i>Scytonema fremii</i>	Nom.nov.
<i>Dasygloea amporpha</i>	Thwaites	<i>Scytonema malayiyensis</i>	Bharadw
<i>Dichothrix</i> sp.	Zanard	<i>Scytonema multiramum</i>	Gardner
<i>Fischerella moniliformis</i>	Fre'my	<i>Scytonema myochrous</i>	(Dillw.) Ag.
<i>Gloeocapsa</i> sp.	Kütz	<i>Scytonema</i> sp.	Ag.
<i>Haplosiphon intricatus</i>	West, W. & G. S.	<i>Stigonema mesentericum</i>	Geitler
<i>Haplosiphon</i> sp.	Nag	<i>Stigonema minutum</i>	Agardh
<i>Lyngbya allorgei</i>	Fre'my	<i>Stigonema panniforme</i>	(Ag) Born. et Flah.
<i>Lyngbya contorta</i>	Lemm.	<i>Synechococcus leptosira</i>	Nageli
<i>Lyngbya lagerheimii</i>	(Möbius) Gom.	<i>Synechococcus</i> sp.	Nag
<i>Lyngbya mesotricha</i>	Skuja	<i>Tolypothrix</i> sp.	Kutz
<i>Lyngbya mucicola</i>	Lemm.	<i>Westiella</i> sp.	Borzi
<i>Lyngbya perelegans</i>	Lemm.	<i>Wollea saceta</i>	Born. et Flah
<i>Lyngbya porphyrosiphonis</i>	Fre'my		
<i>Lyngbya</i> sp.	Ag.	DIATOMS (98 taxa)	
<i>Microchaete elongate</i>	Fre'my	<i>Achnanthes crenulata</i>	Grunow
<i>Microchaete tenera</i>	Thuret	<i>Achnanthes inflata</i>	Kuetzing (Grunow)
<i>Microcoleus lacustris</i>	(Rabenh.) Farlow	<i>Achnanthes linearis</i>	W. Smith
<i>Microcystis</i> sp.	Kütz	<i>Achnanthes microcephala</i> (?)	(Kuetzing) Grunow
<i>Nodularia</i> sp.	Mertens	<i>Achnanthes minutissima</i>	Kützing
<i>Nostoc calcicola</i> (?)	Breb. (after Fre'my)	<i>Achnanthes wolterckii</i>	Hustedt
<i>Oscillatoria acuta</i>	Bruhl and Biswas	<i>Achnanthes tropica</i>	Hustedt
<i>Oscillatoria amphigranulata</i>	Van Goor	<i>Amphipleura</i> sp.	Kuetzing
<i>Oscillatoria</i> sp.	Pollini	<i>Amphora angusta</i>	(Gregory) P.T. Cleve
		<i>Amphora bitumida</i>	Prowse

To be continued

continued

<i>Amphora coffaeiformis</i>	C.A. Agardh	<i>Navicula gracilis</i>	Ehrenberg
<i>Amphora holsatica</i> var. <i>malayana</i>	Prowse	<i>Navicula miniscula</i>	Grunow
<i>Cocconeis placentula</i>	Ehrenberg	<i>Navicula pupula</i>	Kützing
<i>Cyclotella kuetzingiana</i>	Thwaites	<i>Navicula radiosa</i>	Kützing
<i>Cymbella ehrenbergii</i>	Kuetzing	<i>Navicula rhyncephala</i>	Kützing
<i>Cymbella graciles</i>	Rabenhorst (Cleve)	<i>Navicula</i> sp. (new-3 ?)	
<i>Cymbella javanica</i>	Hustedt	<i>Navicula subtilissima</i>	Cleve
<i>Cymbella lanceolata</i>	(Ehrenberg) van Heurck	<i>Navicula tornensis</i>	Cleve
<i>Cymbella microcephala</i>	Grunow	<i>Navicula viridula</i> v. <i>linearis</i>	Hustedt
<i>Cymbella minuta</i>	Hilse. ex. Rabenhorst	<i>Neidium affine amphirhynchus</i> (?)	(Ehrenberg) Cleve
<i>Cymbella parva</i>	(W. Smith) Cleve	<i>Neidium iridis</i>	Ehrenb.
<i>Cymbella prostrate</i>	(Berkeley)	<i>Neidium productum</i>	(W. Smith) Cleve
<i>Cymbella turgida</i>	Gregory	<i>Nitzschia scalaris</i>	
<i>Cymbella ventricosa</i>	Kützing	<i>Nitzschia vermicularis</i>	Grunow
<i>Desmogonium rabenhorstianum</i>	Grunow	<i>Pinnularia microstauron</i>	(Ehrenberg) Cleve
<i>Diatomella</i> sp.	Greville	<i>Pinnularia appendicula</i>	Agardth
<i>Eunotia diodon</i>	Ehrenberg	<i>Pinnularia biceps</i>	Peterson
<i>Eunotia diodon</i> v. <i>minor</i>		<i>Pinnularia bogtensis</i>	Grunow
<i>Eunotia lunaris</i>	(Ehrenberg) Grunow	<i>Pinnularia brauni</i>	Grunow
<i>Eunotia lunaris</i> v. <i>capitata</i>	Grunow	<i>Pinnularia brevicostata</i>	Cleve
<i>Eunotia major</i>	Cleve	<i>Pinnularia microas</i> v. <i>ambigua</i>	Meister
<i>Eunotia major</i> v. <i>indica</i>	Grunow	<i>Pinnularia parva</i>	(Gregory) Cleve
<i>Eunotia monodon</i> v. <i>constricta</i>	Ehrenberg	<i>Pinnularia subsolaris</i>	(Grunow) Cleve
<i>Eunotia monodon</i> v. <i>undulata</i>	Hustedt	<i>Pinnularia tabellaria</i>	Ehrenberg
<i>Eunotia pectinalis</i>	Kützing) Rabenhorst	<i>Pinnularia tabellaria</i>	Ehrenberg
<i>Eunotia pectinalis</i> v. <i>minor</i>	Rabenhorst	<i>Pinnularia trigoncephala</i>	Cleve
<i>Eunotia praemonas</i>	Cleave -Euler	<i>Pinnularia viridis</i>	(Nitzsch) Ehrenberg
<i>Eunotia praemonas</i> v. <i>tibertica</i>	Åke Berg	<i>Pleurosigma elongatum</i>	W. Smith
<i>Eunotia praemonas</i> v. <i>inflata</i>	Cleave	<i>Rhopalodia gibberula</i>	(Ehrenberg) O.F.Muller
<i>Eunotia praerupta</i>	Ehrenberg	<i>Stauroneis pusila</i>	Cleve
<i>Eunotia sudecita</i>	Muller	<i>Stauroneis anceps</i>	Ehrenberg
<i>Fragilaria capucina</i>	Desmazieres	<i>Stauroneis phoenicenteron</i>	(Nitzsch) Ehrenberg
<i>Fragilaria construens</i>	(Ehrenberg) Grunow	<i>Stenopterobia intermedia</i>	(Lewis) Fricke
<i>Fragilaria virescens</i>	Ralfs	<i>Surirella tenera</i>	Gregory
<i>Frustruria vulgaris</i>	(Thwaites) De Toni	<i>Synedra ulna</i>	(Nitz.) Ehrenberg
<i>Frustruria rhomboids</i>	(Ehrenberg) De Toni	<i>Tabellaria fenestra</i>	(Lyng.) Kützing
<i>Frustruria saxonica</i>	(Ehrenberg) De Toni	<i>Tabellaria flocossa</i>	(Roth) Kützing
<i>Gomphonema longiceps</i> v. <i>subclavata</i>	Grunow	CHOLOROPHYTA (13 taxa)	
<i>Gomphonema gracile</i>	Ehrenberg	<i>Crucigenia</i> sp.	Morren
<i>Gomphonema longiceps</i>	Ehrenberg	<i>Euastrum</i> sp.	Ehrenb. ex Ralfs
<i>Gomphonema parvulum</i>	(Kuetzing) van Heurck	<i>Gonatozygon</i> sp.	de Bary
<i>Gomphonema subtile</i>	Ehrenberg	<i>Mougeotia</i> sp.	Agardh
<i>Gyrosigma balticum</i>	(Ehrenberg) Rabenhorst	<i>Oedogonium</i> sp.	Hirn
<i>Gyrosigma distortum</i>	(W. Smith) Cleve	<i>Penium</i> sp.	Brébisson
<i>Gyrosigma elongatum</i>	W. Smith	<i>Pleurotaenium</i> sp.	Nägeli .
<i>Gyrosigma</i> sp.	Hassal	<i>Spirogyra laxa</i>	Kuetzing
<i>Navicula cryptocephala</i>	Kützing	<i>Spondylosium</i> sp.	Kützing
<i>Navicula cuspidate</i>	Kützing	<i>Staurastrum dejectum</i>	Brébisson
<i>Navicula feurborni</i>	Hustedt	<i>Staurastrum glade</i>	(Meyen) Ralfs
<i>Navicula gibbula?</i>	Cleve, P. T	<i>Staurastrum margaritaceum</i>	(Ehrenb.) Ralfs ex Menegh
		<i>Zygonium</i> sp.	Kuetzing