REPORT

Long sediment-laden algal turf likely impairs coral recovery on Florida's coral reefs

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Abstract Coral reefs in the Florida Reef Tract have seen protracted loss of coral over the past several decades due to a variety of disturbances from marine heat waves, cold snaps, and disease events. Corals have not recovered despite abundant herbivorous fishes and relatively low macroalgal cover, two factors thought to facilitate resilience of corals. Thus, factors affecting the replenishment of coral populations may be hindering the recovery of corals. To study the potential factors affecting coral recovery in reefs of the Florida Reef Tract, we assessed benthic abiotic variables (substrate slope, depth, structural complexity, and abundance of sediment), fish assemblages, and benthic composition in three different reef habitats (groove, spur wall, spur top) located on three low-relief reefs and three high-relief spur-and-groove reefs. Herbivorous fish biomass ranged (44.7–107 g m⁻²), which is above average for the Caribbean. Yet there was low coral

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cover (~1%) and low density (~1 coral m²) of small adult corals, which likely reflects the cumulative effects of years of disturbances. The presence and density of juvenile corals were negatively correlated with the depth of the sediment layer trapped within long, sediment-laden algal turfs (LSAT), which are particularly abundant (> 50% cover) in low complexity reef habitats (low-relief groove, low-relief spur top, and high-relief groove). Our results indicate that current unsuitable habitat conditions (high sediment load) for early life stage corals may be an important factor preventing coral recovery. Consequently, the abundance of herbivorous fishes and coral cover trajectories appear decoupled in the region, and additional management initiatives considering LSAT composition are required to aid reef resilience.

Keywords Long sediment-laden algal turf \cdot Coral recruitment \cdot Resilience

Introduction

Both biotic factors (e.g., larval input, primary production, herbivory) and abiotic factors (e.g., light intensity, water temperature, nutrient levels, sedimentation) influence structure, function, productivity, energy cycling and other processes of coral reefs (Done et al. 1996; Harborne et al. 2016) Alterations of one or more factors (e.g., increase of nutrient availability and sediment deposition, reduction of herbivorous fishes), as well as an intensification of natural disturbances, can stress and kill corals, the foundation species of coral reefs, leading to overall ecosystem degradation (Baker et al. 2008; Bozec et al. 2015). Furthermore, anthropogenic stressors such as overfishing herbivores and eutrophication can limit the resilience of western Atlantic reefs after disturbance (Mumby 2006; Hughes et al. 2010).



Therefore, protection of herbivorous fishes that control macroalgae, a major competitor of corals, and reduction of anthropogenic nutrient input, fuel for algal growth, are often suggested as solutions to facilitate coral reef resilience in the region (D'Angelo and Wieddenmann 2014; Adam et al. 2015; Steneck et al. 2019). Nevertheless, some parts of the region, particularly the coral reefs in South Florida, have failed to regain coral cover despite decades of effective protection of herbivorous parrotfishes (Toth et al. 2014). While the decline in corals is likely strongly influenced by repeated disturbances, such as marine heatwaves and disease outbreaks (Toth et al. 2014), resilience still appears lower than expected in such a grazer-rich system, even one without grazing urchins following the disease-induced mortality of *Diadema* urchins (Lessios 2016).

Florida's coral reefs are somewhat atypical because they exist outside the tropics, bounded to the north by a transitional zone from tropical to temperate characteristics that limit their northward expansion (Engle and Summers 1999; Walker 2012; Walker and Gilliam 2013; Toth et al. 2021). Yet, Florida's coral reefs were historically characterized by high, though variable, coral cover and the presence of massive and branching reef-building corals (Dustan 1977; Jaap 1984; Burns 1985; Murdoch and Aroson 1999). Repetitive disease outbreaks in the last 50 years (e.g., black band disease in Orbicella and Dendrogyra, white band disease in Acropora, Porter and Meier 1992; Aronson and Precht 2001; Precht et al. 2016; Lewis et al. 2017), marine heatwaves and cold snaps (Lirman et al. 2011; Manzello 2015) and more recently (after 2014) the spread of stony-coral-tissue-lossdisease (SCTLD) (Precht et al. 2016) have led to low (<5%) coral cover across much of the Florida Keys (Ruzicka et al. 2013; Muller et al. 2020). Coral cover has remained low on these reefs after these mortality events, despite the possible significant if variable input of coral settlers and the implied potential for coral recovery (Toth et al. 2014; van Woesik et al. 2014).

Furthermore, upright macroalgal abundance remains relatively modest (~20% Keys-wide, Ruzicka et al. 2013; Jones et al. 2022), particularly on shallow forereefs, likely because Florida's coral reefs are one of the few places in the Caribbean, where there are still large populations of herbivorous parrotfishes (Shantz et al. 2020). Large scarids such as Scarus guacamaia and S. coelestinus, absent from much of the wider Caribbean as a result of overharvesting, are abundant in the Florida Keys (Jackson et al. 2014; Shantz et al. 2020; Zuercher et al. 2023). Coral-depauperate reefs in the Caribbean have often been associated with overfishing of herbivorous fishes and/ or nutrient pollution, leading to uncontrolled algae growth (e.g., Lobophora variegata and Dictyota spp., Mumby 2009; Lapointe et al. 2011; Jackson et al. 2014). The fact that reefs in the Florida Keys currently have abundant herbivorous fishes, relatively modest algal abundance, and low coral cover despite significant coral larvae input is inconsistent with our general understanding of resilience in the wider Caribbean (Mumby et al. 2007; Steneck et al. 2019). The limited signs of recovery suggest that alternative factors (e.g., natural or human-produced sedimentation) might be compromising processes of coral recovery (Burkepile et al. 2013; Begin et al. 2015; Suchley et al. 2016; Bruno et al. 2019).

The trajectory of recovery for coral reefs following coralkilling disturbances is multifaceted and complex (Doropoulos et al. 2016; Speare et al. 2023). The abundance of algae is often emphasized as a bottleneck to coral recovery as it can limit space for coral larval settlement, and reduce the survival and growth of coral recruits (Kuffner et al. 2006; Box and Mumby 2007; Hoey et al. 2011). Other reef characteristics, such as the presence of facilitating species or the orientation and texture of the benthos, also can strongly influence coral recruitment (Birrell et al. 2005; Davies et al. 2013). For example, the abundance of certain crustose coralline algae that facilitate recruitment (e.g., Titanoderma prototypum) or taxa that inhibit recruitment, such as invertebrates and other species of crustose coralline algae, could significantly influence coral recruitment (Nozawa 2008; Arnold and Steneck 2011; Brandl et al. 2013). Coral larvae also preferentially settle and have higher survival on vertical and rough substrates that often have taxa facilitating recruitment (Arnold and Steneck 2010). In contrast, sediment trapped within algal communities on the benthos appears to be a strong inhibitor of coral recruitment, possibly negatively impacting coral larvae more than abundant macroalgae (Ricardo et al. 2017; Speare et al. 2019). In the Caribbean, however, the impact of sediment on coral settlement, recruitment, and survival is still poorly understood (Rogers and Ramos-Scharron 2022).

Sediment dynamics (e.g., production, suspension, accumulation rate, and spatial distribution) on coral reefs is a complex process driven by several hydrodynamic factors (e.g., current, waves) and habitat characteristics (Schlaefer et al. 2021; 2022; Tebbett et al. 2020a, b; 2023). Of particular importance for coral recovery is the sediment load that settles on the benthos, often trapped within turf algae, creating long sediment-laden algal turf (LSAT hereafter, Goatley et al. 2016) because it could mechanically impede coral settlement (Speare et al. 2019). In Florida, no longterm monitoring programs of coral reefs include LSAT as a category; thus, no information on LSAT abundance or composition is available. Yet the cover by major benthic groups such as stony coral, octocoral, sponge, and macroalgae is usually less than 50% (except for Dry Tortugas National Park; see Toth et al. 2019). Although the composition of the remaining substrate is quite variable (e.g., covered by Peyssonelia spp. and CCA), LSAT is ubiquitous across most habitats, suggesting that sediment accumulation might create unsuitable conditions for the settlement and growth of coral recruits.

Our study investigated the role of abiotic (sediment, substrate slope, and rugosity) and biotic factors (benthic assemblage composition) impacting juvenile corals on reefs in the Upper Florida Keys. We surveyed each reef using a taxonomic resolution suitable for evaluating the relative abundance of potential promoters and detractors of coral recruitment including LSAT, short productive algal turf (hereafter SPAT), juvenile corals, and coral abundance and physical characteristics. Coral juveniles were identified and quantified in situ, across five different reef habitats found within the two most common forereef types in the upper Florida Keys reefs: high-relief and low-relief spur-and-groove reefs. We use these data to 1) evaluate benthic composition among habitats, particularly LSAT and 2) determine potential drivers of juvenile coral densities. We predict that less rugose and flatter habitats on low-relief reefs have deeper sediments than high-relief reefs and are associated with fewer juvenile corals.

Materials and methods

Study sites

Our research was conducted during June-August 2015 on six shallow (5-8 m) forereefs [Conch Reef (24°57.695'W, 80°27.230'N), Davis Reef (80°30.361'W, 24°55.305'N), Pickles Reef (80°24.964'W, 24°59.087'N), Molasses Reef (80°22.374'W, 25°13.632'N), French Reef (80°21.009'W, 25°02.026'N), Elbow Reef (80°26.076'W, 25°00.646'N)] located approximately 10 km offshore from the upper section of the Florida Keys, USA (Fig. 1). Our sites encompassed low-relief and high-relief spur-and-groove reef formations spanning a broad range of benthic assemblages and structural complexity. Considering large-scale complexity (low relief (LR) reef vs. high relief (HR) reef) and within-reef habitats (spur top, spur wall, or groove), we classified the benthos into five habitats: low-relief groove (LR groove), low-relief spur top (LR spur top), high-relief spur groove (HR spur groove), high-relief spur wall (HR spur wall), and high-relief spur top (HR spur top). Spur walls are virtually non-existent in the low-relief reefs and thus were characterized only on high-relief reefs (Fig. 1). Across all sites, sandy substrates (sediment depth over 1 cm equivalent to > 5000 g m⁻²) were excluded from the analysis given that sediment loads of that magnitude are unsuitable for sessile



Fig. 1 Reef types and habitat representations of common Florida Keys forereefs. A site location, B representation of reef habitats within reefs, and C proportion of reef habitats within reefs

organisms like corals (Tebbett et al. 2023). We corroborated the distinction between reef types (low-relief reefs vs. highrelief reefs) using the Rugosity Index (R.I) as a proxy of reef-scale structural complexity (Risk 1972, ESM Fig. 1).

Herbivorous fish biomass and benthic characteristics

We assessed fish community structure at each site via visual surveys along 20 belt transects $(2 \times 30 \text{ m})$ located haphazardly perpendicular to the spur and groove formation. All fishes were identified and their estimated size (total length, to the nearest cm) was recorded. Size estimates were converted to biomass for each fish using published length to weight relationships (Bohnsack and Harper 1988). Biomass of individuals from families Acanthuridae (three surgeonfish species) and scarine labrids were combined (ten parrotfish species) to estimate herbivorous fish biomass at each site. We used one-way ANOVA to compare herbivorous fish biomass across sites.

We surveyed the benthos in 25 (50×50 cm) quadrats placed every two meters along eight (50 m) transects per site (n = 200 plots/site). We used an Olympus camera (Tough TG-5) to photograph each 50×50 cm quadrat and then analyzed the benthic composition in the lab using Coral Point Count with Excel extensions version 4.1 (CPCe, Kohler and Gill 2006) with the following code modifications: long sediment-laden algal turf (LSAT), short productive algal turf algae (see below for description), hereafter "SPAT," were classified as a relatively sediment-free, dense multi-species (e.g., Polysiphonia spp., Ceramium spp. Hypnea spp. Lau*rencia* spp.) algal assemblage forming a layer < 1 cm tall (Connell et al. 2014). We combined all species of crustose coralline algae into a single group (CCA). We recognize that different species of CCA can have positive, neutral, or negative impacts on coral settlement and recruitment (Ritson-Williams et al. 2016), but these taxa are difficult to discriminate in the field making characterization of their distribution at this scale challenging. Species from the genus Peyssonnelia, a non-coralline crustose alga, were classified as a single group (Peyssonnelia). Percent cover from each plot was calculated from 25 points generated following a stratified-random distribution within each photo-quadrat. For each coral colony within a plot, we measured the maximum colony diameter of corals larger than 4 cm.

Since the physical characteristics of the substrate, such as slope, can determine the benthic composition (Fabricius and De'ath 2001), we collected information on rugosity, substrate slope, and depth of the sediment layer within each 0.25 m^2 plot. We estimated R.I. using a 50 cm chain (link ~ 1 cm length) laid within each 50×50 cm quadrat parallel to the transect line and measured the linear length. We calculated habitat-scale R.I by dividing the chain's total length (50 cm) by the linear length covered by the chain

within the plot where R.I = 1 indicates the lowest complexity, and it increases with complexity. The sediment depth (mm) within each plot (n = 200 per site) was measured using a pencil calibrated with 1 mm increments. The pencil was inserted vertically into the sediment layer until it reached the hard substrate to measure the sediment depth. We recorded the slope of the substrate (i.e., the angle from horizontal) using a protractor with a small foam float placed on the 25 cm quadrat side parallel, closest to the transect tape. Small angles correspond to relatively flat (horizontal) substrates, whereas higher degree angles (up to 90°) are associated with more vertical substrates.

Sediment was usually trapped within turf algae, forming a sediment-turf matrix hereafter "long sediment-laden algal turf" (LSAT, Goatley et al. 2016). To quantify the LSAT turf-algae composition, we first measured the sediment layer using the calibrated pencil method from five plots at 1 m, 11 m, 21 m, 31 m, and 41 m along each transect (n = 40/site) of the matrix. Then, we removed sediment by perturbing the water with a manual bilge pump until we fully exposed the turf algae entangled within the sediment matrix. Finally, we used the calibrated pencil to record the length of the exposed turf algae. Here we only report turf length and sediment thickness.

Distribution of residuals and normality was checked using residual Q-Q plots. We used linear mixed models (LMM) with "study site" as a random factor to compare biotic and abiotic variables across reef habitats using R package *lme*4 (Bates et al. 2015). We performed pairwise post hoc analyses tailored for LMM using the R package (multcomp). To contrast benthic assemblage composition by habitat, we used a nonmetric-multidimensional scaling analysis (NMDS) followed by a permutational analysis of variance (PERMANOVA) using the R package *vegan* (Oksanen et al. 2017).

Drivers of juvenile corals

We surveyed coral juveniles (colonies ≤ 4 cm in diameter) in situ within the southwest quarter (25×25 cm) of each plot. The diameter of each juvenile was measured and identified to the genus level. We divided the number of coral juveniles by 0.0625 (area surveyed in m²) to estimate the density of juveniles m⁻². We used GLMM (including study site as a random effect) to compare density of juveniles and adults among reef habitats. Because of the low abundance of corals across all habitats and sites, we tested for the influence of biotic and abiotic variables using simply the presence and absence data of juvenile corals using a multiple logistic regression model (LRM) with a binomial error structure. Collinearity was evaluated using the variation inflation factors (VIF). We included biotic and abiotic variables (e.g., rugosity index, LSAT sediment depth, substrate slope, and abundance of most dominant benthic groups (LSAT, SPAT, *Dictyota* spp. Gorgonian, CCA, Sponge, Coral) in our first logistic regression model. We started from a null model (only the intercept) and built eight models combining biotic and abiotic variables. We selected the model with the lowest AIC. In all cases, we reported the model fit using D², a ratio between the logistic models' residual deviance and null deviance. The estimates reported from logistic regression have not been transformed. We ran all descriptive analyses, graphs, and models with R version 3.2.2 (R Development Core Team 2016).

Results

Herbivorous fish biomass and benthic characteristics

Our study sites displayed a gradient of herbivorous fish biomass and reef habitat characteristics across Florida's offshore coral reefs. Herbivorous fish biomass across all sites ranged from 44.70 (\pm 5.84) g m⁻² at Conch Reef to 107.13 (\pm 13.75) g m⁻² at Molasses Reef (Fig. 2). All three high-relief reefs showed higher herbivorous fish biomass than the low-relief reefs (One-way ANOVA, F_{5,114}=3.965, p=0.003).

Habitat characteristics ranged from almost complete horizontal substrates (average slope ~ 10 degrees) dominated by long sedimented-laden algal turf (LSAT) to more diverse algal communities in habitats with steeper slopes and higher physical complexity (Fig. 3, ESM Table 1). LSAT dominated the LR groove habitat, covering 70% of the benthos. In LR spur and HR groove habitats, LSAT



Fig. 2 Herbivorous fish biomass by study site (mean \pm SE). Dash lines indicate reported parrotfish biomass in some other Caribbean areas (red, Los Roques Venezuela; purple, Saint Vincent; brown, Costa Rica; blue Flower Gardens Bank; green Bonaire). Taken from Jackson et al 2014

covered approximately 50% of the benthos, whereas HR spur wall and HR spur top habitats showed a more heterogenous distribution by taxa and higher diversity compared to LR groove, LR spur, and HR groove habitats (Fig. 3C and D). We observed LSAT abundance decreased with increasing slope (Estimate = -0.26, R² = 0.61, p < 0.001) and physical complexity (Estimate = -32.01, R² = 0.64, p < 0.001) of the reef habitats. The abundance of LSAT increased over three times (from 20 to 60% cover) from high-relief reefs (spur top and wall) to low-relief groove habitats ($F_{4,1030} = 63.85, p < 0.001$). In contrast, *Dictyota* spp., SPAT, and crustose coralline algae became more abundant in high-relief reefs (PERMANOVA, habitat, $R^2 = 0.56$, $F_{4.101} = 30.8$, p = 0.010 (Fig. 3C and D). The NMDS analysis (Fig. 3D) revealed distinct assemblages across habitats where low-relief grooves were dominated by LSAT while high-relief walls had more abundant Dictyota, SPAT, and to a lesser extent CCA. Indeed, differences in assemblage composition among habitats were almost double the variation explained by reef type (PER-MANOVA, habitat, $R^{2=0.32}$, $F_{1,101=}47.7$, p = 0.010).

Coral cover was very low $(1.0 \pm 0.1\%)$ across all reef habitats (LMM, F=1.7, p=0.158). The density of juvenile (≤ 4 cm) corals averaged 13.1 ±0.6 corals m⁻² with assemblages dominated by *Agaricia* (35% of juvenile corals), *Siderastrea* (29% of juvenile corals), and *Porites* (28% of juvenile corals) (Fig. 4A&B). The density of adult corals (>4 cm) was low across all habitats (0.8±0.1 coral m⁻²), which represented approximately 90% fewer adult corals compared to the density of juveniles and was dominated by species of *Siderastrea* (Fig. 4C&D). Noticeably, juvenile and adult colonies of large reef-building corals such as *Orbicella* and *Acropora* represented approximately 3% of both juvenile (n=25) and adult (n=8) coral communities (Fig. 4B&D).

Drivers of juvenile coral abundance

More than 50% of all plots (604 out of 1086 total plots) did not have any juvenile corals. Thus, we analyzed presence/ absence of juvenile corals as a function of benthic abiotic characteristics (rugosity, slope, and LSAT sediment depth) and biotic composition (percent cover of common groups) across all sites and habitats. Sediment depth trapped within LSAT was the best predictor (negative) of juvenile coral presence (ESM Table 2). The probability of finding juvenile coral significantly declined as mean LSAT sediment depth within the plot increased (Fig. 5A, LRM, Estimate = -0.11, p < 0.001). The second significant predictor (positive) of juvenile coral presence was CCA cover (Fig. 5B, LRM, Estimate = 0.03, p < 0.009) with the likelihood of juvenile corals increasing as CCA cover increased.



Fig. 3 Benthic characteristics by reef habitat. A Substrate slope, B Sediment depth of LSAT across habitats, C community composition of major groups by habitat and estimate slope and structural complex-

ity (mean \pm SE). **D** Non-metric multidimensional analysis contrasting reef habitat composition. Notice that colored polygons indicate habitat type



Fig. 4 Coral density and relative species composition. A juvenile coral density, B juvenile coral community composition, C adult coral density, and D adult coral community composition. Points indicate (mean \pm SE) and letters different groups based on Posthoc analysis

Fig. 5 Probability of coral juvenile presence as a function of sediment depth trapped within long sediment-laden algal turf (**A**) and abundance of crustose coralline algae (**B**)



Discussion

Across all reef habitats, we found a very low density of juvenile corals (~13 juvenile m⁻²) with communities dominated by so-called weedy species (from the genera *Porites, Agaricia*, and *Siderastrea;* Knowlton 2001), which might indicate impoverishment of coral settlement substrate. Indeed, reef habitats with lower rugosity and slope, usually dominated by LSAT, showed the lowest juvenile density, likely as a consequence of a deep (> 3.5 mm) LSAT sediment layer. Only the cover of crustose coralline algae, which tended to be more abundant on high-relief reefs, was a significant positive predictor of juvenile coral presence.

Our results show that habitat characteristics, particularly the abundance and depth of sediment, are negatively correlated with juvenile coral densities. There is substantial evidence that sediment has deleterious effects on juvenile and adult corals (Rogers 1990; Jones et al. 2015). Adverse effects of sedimentation include physiological stress and decreased photosynthesis and respiration (Riegl and Branch 1995; Weber et al. 2012; Tuttle and Donahue 2022), reduction of fertilization rates (Jones et al. 2015), recruitment (Birrell et al. 2005), coral growth (Rogers 1990), and total or partial coral colony mortality (Flores et al. 2012). Reef habitats with a high percent cover of LSAT were ubiquitous across our study sites, suggesting that sediment trapped within turf algae might create unsuitable conditions for the settlement and growth of coral recruits (Evans et al. 2020). Previous experiments conducted in the Florida Keys have shown that LSAT, and especially the sediment within LSAT, significantly reduces successful settlement of coral larvae (Speare et al. 2019). Furthermore, given that overall coral cover was very low, it was not surprising that we found few adult corals (approximately 10% of the number of juveniles) on these reefs. These adult coral communities were dominated by the genus Siderastrea. Some species of Siderastrea (e.g.,

Siderastrea radians) are temperature and sediment-tolerant (Lirman and Manzello 2009); thus these species may be the most likely species to thrive under high-sediment conditions.

Common benthic groups (e.g., SPAT, Dictyota spp.) did not significantly predict juvenile coral presence on highrelief reefs. Despite their low abundance, CCA was the only benthic group that showed a positive relationship with the occurrence of juvenile corals. A few CCA species (e.g., *Titanoderma prototypum*) have been proposed to facilitate coral recruitment (Arnold et al. 2010; Ritson-Williams et al. 2010). For the GBR, Abdul Wahab et al. (2023) tested 15 CCA species and reported Titanoderma cf. tessellatum as the most effective inducing coral larval settlement. Other taxa such as Porolithon and Sporolithon have the capacity to promote coral settlement, at least of some coral species (Abdul Wahab et al. 2023). Unfortunately, CCA species from Florida's coral reefs are not well studied, and we are unaware of any work referring to individual species abundances or the composition of CCA assemblage, making it challenging to understand how coral recruitment patterns related to the distribution of CCA taxa. Furthermore, species-level identifications in our quadrats were not possible while underwater or from photographs as they require detailed analyses of the CCA thalli in the lab (Steneck 1986). Such eco-taxonomic analyses are needed in the future, given the importance of this group for coral reef resilience.

On high-relief reefs, *Dictyota* spp. was typically the dominant algal group followed by short productive algal turf (SPAT). Yet, there was no relationship between the abundance of *Dictyota* spp. and patterns of juvenile coral presence. Species within the genus (e.g., *Dictyota pulchella*) have been shown to reduce the growth of newly settled corals, specifically via physical abrasion, shading, or other physical mechanisms (Box and Mumby 2007; Kuffner et al. 2006; Rasher et al. 2011). This result suggests that species of the genus *Dictyota* found in our

surveys might not impact coral juveniles as significantly as elsewhere in the region, perhaps because of the cooler winters or the dominant impact of LSAT. In addition, Dictyota is quite seasonal on Florida's coral reefs (Lirman and Biber 2000; Duran et al. 2016) and maybe less abundant when coral settlement is happening. Other commonly seen algae in Caribbean coral reefs, such as Lobophora variegata, were practically absent. Macroalgae such as Lobophora variegata with prostrate morphology can decrease survival and growth of corals, likely via smothering and creating hypoxic areas at the interface with corals (Box and Mumby 2007; Ferrari et al. 2012). Similar algal species (e.g., Liagora sp) have also been shown to physically inhibit settlement and growth of corals (Box and Mumby 2007; Doropoulos et al. 2014) but were rare on our study reefs.

Considering the high abundance of LSAT and scarcity of CCA, coral larvae are unlikely to find favorable conditions to settle on the studied reefs. Thus, our results support the hypothesis proposed by van Woesik et al. (2014) that larval supply may not be the leading cause of low coral recruitment on Florida's coral reefs, but rather, the lack of substrate with suitable habitat quality for settlement and survival of recruits appears to be a more important bottleneck for recruitment. In addition to our study, results from other work create a concerning picture of coral recruitment on Florida's reefs. Previously, van Woesik et al. (2014) and Tougas and Porter (2002) evaluated coral recruitment across the Florida Keys and found a disproportional amount of hydrocoral recruits compared to scleractinians and the absence of recruits of massive coral species (i.e., Orbicella). Moulding (2005) studied patterns of coral recruitment on patch reefs along the Florida Reef Track and found lower density and diversity in the Upper Keys than Middle Keys and Lower Keys. Studies from Tougas and Porter (2002) and Moulding (2005) show large recruitment variability as a function of depth (deeper sites tend to have more scleractinian recruits), location, and other variables (e.g., tile position and separation from the seafloor). More recently, Harper et al. (2023) reported a dramatic 70-fold difference in coral recruitment between two years (2017–2018), with much of this variability being driven by broadcast spawning Siderastrea spp. These results indicate the urgent need to more fully study coral larval supply and recruitment patterns on these reefs (see King et al. 2023).

Unlike many other Caribbean reefs, the protection of herbivorous fishes in the Florida Keys has maintained a high abundance of large parrotfishes (Shantz et al. 2020) that can substantially impact corals via intentional or incidental predation (Burkepile 2012). For instance, Rotjan et al. (2006) showed that chronic coral predation could delay the process of coral recovery after disturbances. In the Florida Keys, the low coral cover and high abundance of parrotfishes, which often prey on corals (Burkepile et al. 2019), can lead to increases in coral predation with overall detrimental effects on coral communities (Burkepile 2012). Indeed, corallivory by parrotfishes, particularly the stoplight parrotfish (*Sparisoma viride*), is intense enough to hamper current coral restoration efforts in some areas of South Florida (Koval et al. 2020). Although the net effect of parrotfish grazing is generally considered positive for reefs (Mumby 2009), corallivory might lead to low coral recruitment in the Florida Keys. It is worth considering that as abundant LSAT constrains habitat availability to corals for recruitment, and that herbivorous fish tend to feed less (Bonaldo et al. 2012; Goatley and Bellwood 2013), then parrotfishes may be even more likely to negatively impact recruitment patterns.

Florida's coral reefs have experienced severe (>75%)loss of corals in the last 50 years caused by several disturbances, including cold-water stress events (Burns 1985; Lirman et al. 2011), coral diseases (Porter and Meier 1992; Kuta and Richardson 1996), and bleaching events (Manzello 2015), resulting in a dramatic decline in coral cover (Lewis et al. 2017). As a result, the current coral cover is less than 5% across the outer reefs on average across the Keys (Rutten et al. 2008; McClenachan et al. 2017), and is considered in an erosional state with few signs of natural recovery. We proposed that the lack of coral recovery in the last few decades highlights an additional negative feedback loop to incorporate into considerations of reef resilience. This additional feedback loop is that coral cover and complexity were high before the 1980's (Dustan 1977; Jaap 1984; Burns 1985). After the catastrophic impacts of several coral-stress events (e.g., diseases, bleaching, cold snap), the loss of coral cover has reduced physical complexity (Perry and Harborne 2016). Based on our results, reducing physical complexity and flattening reefs facilitate sediment buildup. As sediment accumulates, it protects turf algal communities against herbivory (Gordon et al. 2016; Tebbett et al. 2020a, b, 2021). The sedimented-turf algae grow longer (ESM Fig. 2) and stabilize the turf-sediment matrix (LSAT), creating impoverished habitat conditions for coral settlement and recruitment (Speare et al. 2019, Tebbet et al. 2021).

Complexity has decreased across the region (Alvarez-Filip et al. 2009), but little is known regarding how sedimentation could influence less complex reefs (Tebbett et al. 2023), particularly in Florida (Duran et al. 2018). Further analyses (e.g., seasonal changes of sediment load, sediment sources, sediment composition, and hydrodynamic drivers of reef sediment) are needed to better to understand sediment dynamics on Florida's coral reefs. For instance, sediment composition [grain size, grain type (carbonate vs. silica based), organic composition, and contaminats] might determine the effect of sediment on reef organisms and ultimately on coral reef resilence. Rogers and Ramos-Scharron (2022) argue that sedimentation (sediment delivery) and its impact on Caribbean coral reefs is poorly studied. However, from studies across the globe, we know that sedimentation is a complex process. For instance, Tebbett et al. (2023) showed that sedimentation of coral reefs is related to currents, waves, and parrotfish sediment production (reworked sediment). These investigators also found that the highest sediment accumulation rates in sediment traps and sediment standing stocks in turfs were on the low-complexity reef flat. Such studies are urgently needed to understand sedimentation patterns along Florida's coral reef.

There are still fundamental knowledge gaps regarding the coral larval supply, reef sediment sources, and the overall ecological role of herbivores (corallivory vs. herbivory) on Florida's reefs. It is feasible to think that low coral cover translates into low coral larval production but we still need to understand more about the dynamics of the coral larval pool. The abundance of coral larvae seems to vary significantly from year to year (Harper et al. 2023), some sites in Florida may export (act as sources) and receive (act as sinks) more than others resulting from local connectivity (King et al 2023). However, our study suggests that sediment load within algal turf assemblages could be a key influence on the apparent lack of resilience of corals on these reefs. While such knowledge is critical, it also has important implications for conserving Florida's coral reefs. For example, the role of sediment and its spatial variabiliy suggest that mapping habitat-specific characteristics that make some reefs more resilient than others, improving water quality, and dealing with sediment load as part of restoration activities would benefit management efforts. More generally, our work underscores the need to urgently reduce stressors that drive coral mortality and the loss of structural complexity on reefs.

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Declarations

Conflict of interest On behalf of all authors, the corresponding author states that there is no conflict of interest.

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