REPORT

Clarifying functional roles: algal removal by the surgeonfishes Ctenochaetus striatus and Acanthurus nigrofuscus

Sterling B. Tebbett¹ · Christopher H. R. Goatley^{1,2} · David R. Bellwood¹

Received: 14 July 2016 / Accepted: 14 March 2017 / Published online: 22 March 2017 - Springer-Verlag Berlin Heidelberg 2017

Abstract The lined bristletooth, Ctenochaetus striatus, and the brown surgeonfish, Acanthurus nigrofuscus, are among the most abundant surgeonfishes on Indo-Pacific coral reefs. Yet, the functional role of these species has been the focus of an ongoing debate lasting at least six decades. Specifically, to what extent are C. striatus herbivorous like the visually similar A. nigrofuscus? To address this question, we used natural feeding surfaces, covered with late successional stage reef-grown algal turfs, to examine turf algal removal by the two species. Surfaces exposed to C. striatus in laboratory experiments exhibited no significant reductions in turf length or area covered by turfing algae. In marked contrast, A. nigrofuscus reduced turf length by 51% and area covered by turfing algae by 15% in 1 h. The gut contents of specimens from the reef revealed that A. nigrofuscus predominantly ingests algae (the dominant item in 79.6–94.7% of gut content quadrats), while *C. striatus* ingests detritus and sediments (dominant in 99.6–100% of quadrats). Therefore, C. striatus ingests detritus and sediment, leaving mature algal turfs relatively intact, while A. *nigrofuscus* directly removes and ingests

Communicated by Ecology Editor Dr. Stuart A. Sandin

Electronic supplementary material The online version of this article (doi:[10.1007/s00338-017-1571-z](http://dx.doi.org/10.1007/s00338-017-1571-z)) contains supplementary material, which is available to authorized users.

 \boxtimes Sterling B. Tebbett sterling.tebbett@my.jcu.edu.au

- ¹ ARC Centre of Excellence for Coral Reef Studies and College of Science and Engineering, James Cook University, Townsville, QLD 4811, Australia
- Function, Evolution and Anatomy Research Lab, School of Environmental and Rural Science, University of New England, Armidale, NSW 2351, Australia

turf algae. The function of C. striatus differs from cropping herbivorous surgeonfishes such as A. nigrofuscus. On coral reefs, C. striatus brush detrital aggregates from algal turfs, removing microorganisms, organic detritus and inorganic sediment. Confusion over the functional role of C. striatus may stem from an inability to fit it into a single functional category.

Keywords Coral reef - Herbivory - Detritivory - Epilithic algal matrix - Algal turfs

Introduction

Investigating functional roles on coral reefs and classifying fish species into broad functional groups has received considerable attention in recent years (Graham et al. [2011](#page-9-0); Pratchett et al. [2011;](#page-9-0) Chong-Seng et al. [2012](#page-9-0); Plass-Johnson et al. [2015\)](#page-9-0). The identification of functional components within this complex ecosystem provides a clearer perspective of reef resilience and facilitates ecosystembased management approaches (Bellwood et al. [2004](#page-8-0); Nyström [2006](#page-9-0); Nash et al. [2013\)](#page-9-0). However, broad functional classifications may conceal important interspecific variation that could be vital in understanding the ecology of fishes (Clements et al. [2009;](#page-9-0) Brandl and Bellwood [2014](#page-8-0); Streit et al. [2015](#page-10-0); Kelly et al. [2016](#page-9-0)). This is particularly important when examining common species that play important roles in ecosystems. This issue has come to the fore with the conflicting classifications of two key fishes in Indo-Pacific reef ecosystems, the surgeonfishes Ctenochaetus striatus and Acanthurus nigrofuscus.

The lined bristletooth, C. striatus, and the brown surgeonfish, A. nigrofuscus, are both abundant on Indo-Pacific coral reefs (Randall [2005](#page-10-0); Cheal et al. [2012](#page-8-0)). Together they comprise a core component of the herbivorous/detritivorous fish community that feeds on the epilithic algal matrix (EAM) on these reefs (Russ [1984](#page-10-0); Choat and Bellwood [1985;](#page-9-0) Randall [2005;](#page-10-0) Cheal et al. [2012](#page-8-0)). Their abundance means that the two species are highly influential in quantitative assessments of reef resilience based on functional groups (Cheal et al. [2012](#page-8-0); Johansson et al. [2013](#page-9-0)). Furthermore, with the recent move towards modelling complex coral reef ecosystems, the two species often influence the outputs of models used to assess herbivory on reefs (Brandl and Bellwood [2016;](#page-8-0) Doropoulos et al. [2016](#page-9-0)). However, considerable debate persists over the functional role of these species, especially C. striatus. The key question is: to what extent is C. striatus herbivorous and how does it compare to A. nigrofuscus?

Numerous ecological studies have classified C. striatus as a herbivorous fish along with the other surgeonfishes (e.g. Hiatt and Strasburg [1960](#page-9-0); Bouchon-Navaro and Harmelin-Vivien [1981](#page-8-0); Montgomery et al. [1989](#page-9-0); Polunin and Klumpp [1989](#page-9-0)). Under this classification, the degree to which C. striatus has been considered herbivorous has ranged from being pooled with A. *nigrofuscus* due to their superficial similarity in appearance (Bouchon-Navaro and Harmelin-Vivien [1981\)](#page-8-0), to examples such as Montgomery et al. ([1989\)](#page-9-0) who classified C. striatus as herbivores but noted that they ingested large quantities of sediment and detritus. More recently Marshell and Mumby [\(2012](#page-9-0), [2015\)](#page-9-0) suggested C. striatus could remove more algae than A. nigrofuscus when feeding on early successional algal communities (a maximum of 6 weeks old) on artificial substrata, highlighting its potential role as a functional herbivore on coral reefs. Reports of C. *striatus* bioeroding the reef matrix in the Red Sea (Schuhmacher et al. [2008](#page-10-0); Krone et al. [2011](#page-9-0)) have also been presented as evidence of their ability to remove significant amounts of algae (Marshell and Mumby [2015\)](#page-9-0). These independent lines of evidence all suggest that C. striatus predominantly functions as a herbivore.

There is also evidence suggesting that C. striatus does not remove significant amounts of algae on coral reefs and instead functions primarily as a detritivore (Robertson and Gaines [1986;](#page-10-0) Choat and Clements [1998;](#page-9-0) Choat et al. [2002](#page-9-0); Crossman et al. [2005;](#page-9-0) Clements et al. [2009\)](#page-9-0). Behavioural observations show that C. striatus often feed within the territories of the herbivorous, lined surgeonfish, A. lineatus. This suggests that these fishes do not compete for the same food resource (Choat and Bellwood [1985\)](#page-9-0). Subsequent gut contents and short-chain fatty acid analyses have suggested that A. lineatus and A. nigrofuscus ingest substantial quantities of algal matter, while C. striatus predominantly ingest detritus and sediment (Robertson and Gaines [1986](#page-10-0); Choat [1991](#page-8-0); Choat and Clements [1998;](#page-9-0) Choat et al. [2002](#page-9-0); Crossman et al. [2005\)](#page-9-0). Ctenochaetus striatus was subsequently classified as a detritivore (Robertson and Gaines [1986;](#page-10-0) Choat and Clements [1998;](#page-9-0) Choat et al. [2002](#page-9-0); Crossman et al. [2005](#page-9-0); Clements et al. [2009](#page-9-0)). In addition, comparative examination of the morphology and bite capabilities of C. striatus and A. nigrofuscus have revealed marked differences (Purcell and Bellwood [1993\)](#page-9-0). The key morphological feature of Ctenochaetus species is the possession of highly modified comb-like teeth, which are reportedly used to brush detritus from the EAM (Randall [1955](#page-9-0); Jones [1968;](#page-9-0) Purcell and Bellwood [1993;](#page-9-0) Bellwood et al. [2014](#page-8-0)). Randall [\(1955](#page-9-0)) and Purcell and Bellwood [\(1993](#page-9-0)) suggested that the comb-like teeth of Ctenochaetus species (Fig. 1a) are ineffective at removing algae, particularly when compared to the spatulate teeth of A. nigrofuscus (Fig. 1b). These studies of the behaviour, nutritional ecology and morphology of C. striatus have all suggested that *Ctenochaetus* are unlikely to remove significant amounts of turf algae from coral reefs.

Although research involving Ctenochaetus and Acanthurus has spanned at least six decades, it is evident that considerable disagreement persists over the functional role of C. striatus on coral reefs. As C. striatus is highly abundant and widespread on coral reefs across the Indo-

Fig. 1 Scanning electron micrographs of the teeth of a the lined bristletooth, Ctenochaetus striatus (anterior view of dentary) and b the brown surgeonfish, Acanthurus nigrofuscus (anterior view of dentary)

Pacific, determining its functions is critical to understanding ecological processes such as detritivory and herbivory. Our aim was to examine the functional role of C. striatus and A. nigrofuscus, answering the question: to what extent is C. striatus a herbivore when feeding on mature turf algal communities and how does it compare with the superficially similar species, A. nigrofuscus?

Methods

Algal removal and ingestion by C. striatus and A. nigrofuscus were examined at Lizard Island Research Station (14°40'8.04"S, 145°27'33.84"E), on the mid-shelf of the Great Barrier Reef. To examine algal removal, an aquarium-based before/after control/impact-style experiment was performed using natural feeding surfaces, i.e. late successional stage EAM-covered dead coral rocks. Gut contents analyses were also performed on fishes from the reef to examine ingested material.

Experimental procedures

Fish collection and husbandry

Ten C. striatus and ten A. nigrofuscus were collected using barrier nets. The average total length of the C. striatus and A. *nigrofuscus* specimens was 124.0 ± 4.5 mm (\pm SE) and 127.7 ± 2.3 mm, respectively. The fish were transported to Lizard Island Research Station where they were individually housed in 90-L containers (620 \times 400 \times 380 mm) with flow-through water in an aquarium room. Fish were acclimated to experimental conditions by offering them EAM-covered rocks each day and placing a video camera (GoPro) inside each aquarium to record behaviour. The camera indicator lights and sound were turned off to minimise the effect of the camera on behaviour. This process was repeated for at least 3 d to ensure fish were accustomed to the addition of GoPros to their tanks before treatments were offered. Each afternoon the aquaria were syphoned to remove waste material. The fish readily acclimatised to aquarium conditions (usually within 2 d) and trials were started once they fed repeatedly from EAMcovered rocks.

Feeding surface preparation

To ensure experiments closely replicated field conditions, natural feeding surfaces that supported mature reef-grown algal turfs were used. These feeding surfaces were flat EAM-covered coral rocks measuring approximately 50 cm² (Gordon et al. 2016) collected from a single area of reef. Feeding rocks supported natural algal turfs indistinguishable from adjacent reef EAMs. Rocks were conditioned on the reef for an extra 6 months prior to use to ensure that the algal turfs were similar on all rocks and that rocks supported mature algal turfs (later successional stage turf algal communities, i.e. well-grazed, stable algal turfs $<$ 10 mm high). All rocks were placed in an area measuring approximately 5 m^2 , at approximately chart datum (i.e. submerged all the time apart from the lowest astronomical tides), and away from the territories of damselfish, but within the range of grazing herbivorous/detritivorous fishes. Both C. striatus and A. nigrofuscus were present in this location. Rocks were collected the morning before use in the experiment and held in flow-through aquaria.

Benthic particulate preparation

To standardise the other components of the EAM (sediment and detritus) benthic particulate loads were created. The loads were equivalent to 150 g m^{-2} of sediment with an organic percentage of 14%, to replicate loads found naturally on Lizard Island reef crests (Purcell [2000](#page-9-0); Purcell and Bellwood [2001](#page-9-0)), the preferred reef habitat of the two study species at Lizard Island (Goatley and Bellwood [2010](#page-9-0), [2012](#page-9-0)). A value of 150 g m^{-2} falls mid-way within the range of average sediment loads reported for Lizard Island reef crest EAMs $(75-236 \text{ g m}^{-2})$ (Purcell [2000](#page-9-0); Goatley and Bellwood [2010](#page-9-0), [2012\)](#page-9-0).

To ensure similar properties to sediment found naturally in EAMs, benthic particulates were created from sediment collected from Lizard Island lagoon. Organic material was produced from Hikari Marine A, which approximates the nutritional composition of EAM detrital aggregates (Tenore [1981](#page-10-0); Wilson et al. [2003](#page-10-0)) and acts as a substitute for detritus in reef particulates (Gordon et al. [2016](#page-9-0)). Sediment and Hikari Marine A were prepared following Gordon et al. [\(2016](#page-9-0)). Sediments were bleached using hydrogen peroxide $(H₂O₂)$ to remove residual organic matter. Bleaching continued until no bubbles were released in a 24-h period. Sediment was then dried to a constant weight at 60 \degree C and sieved through a sieve stack $(2000-63 \mu m)$. Hikari Marine A was ground using a pestle and mortar and then passed through a 125-um sieve to ensure similar particle sizes to natural detrital material (Wilson et al. [2003](#page-10-0); Gordon et al. [2016](#page-9-0)). Using this prepared sediment and organic material, benthic particulate loads were created by weighing out individual grain size fractions to simulate grain size distributions in Lizard Island reef crest EAM sediments (Purcell [2000](#page-9-0)). All grain sizes under 2000 μ m were considered sediment (sands-clays; ISO 14688-1:2002). This procedure ensured equivalent sediment and organic loads on all experimental surfaces, as these may influence rates of herbivory/detritivory (Goatley et al. [2016](#page-9-0); Gordon et al. [2016](#page-9-0); Tebbett et al. [2017a](#page-10-0)).

Experimental process

Feeding rocks were rinsed in seawater immediately prior to use to remove existing sediment and detritus, and visually inspected to ensure they were evenly covered with turfing algae and free of macroalgae $(>10 \text{ mm})$ or encrusting organisms. Thirty haphazardly selected algal filaments from the flat upper surface of each rock were measured using the depth probe of vernier calipers (measuring the filament closest to the caliper probe). This distance was immediately recorded by pushing the tips of the calipers into saltwater-resistant pressure-sensitive poster adhesive (Blu-tack). The distance was then measured using digital calipers following Bonaldo and Bellwood ([2009\)](#page-8-0) and Goatley and Bellwood ([2013\)](#page-9-0). Following algal turf measurements, all rocks were photographed to quantify the area covered by turfing algae. A grid of 30 randomly distributed points was overlaid over each photograph, and the presence or absence of turfing algae under each point was recorded.

One rock was placed at the end of each aquarium and concealed within a 500-mm length of 90-mm-diameter polyvinyl chloride (PVC) pipe to prevent feeding by fishes. To ensure sediment and organic loads were consistent, the pre-prepared benthic particulates were wetted and then poured into the PVC pipe and allowed to settle onto the feeding surfaces for at least 12 h (overnight) (following Gordon et al. [2016\)](#page-9-0). At approximately mid-day the following day, a video camera (GoPro, with indicator lights and sound turned off) was placed into each aquarium, to quantify the number of bites taken by each fish. Following the addition of the camera, the PVC pipe was removed and a 10-mm-high 90-mm-diameter PVC ring was placed over the rock to restrict feeding to the upper surface. Following an exposure period of 60 min, the feeding rocks were removed, photographed for quantification of algal coverage and the algal turf lengths remeasured. To control for potential losses of algae due to handling, an additional 20 rocks were exposed to identical experimental procedures in aquaria without fishes. On each rock, the mean length of algae before experimental exposure to C. striatus, A. nigrofuscus or the control was 4.1 ± 0.1 , 4.0 ± 0.2 and 3.8 ± 0.1 mm, respectively (Electronic supplementary material, ESM Table S1). The mean percentage area of algal turfs on each rock before exposure was 50.7 ± 3.4 , 59.7 ± 3.1 and $53.3 \pm 2.7\%$, respectively (ESM Table S1).

Video and statistical analysis

The total number of bites taken by fishes from the surface of the rock over the entire hour was recorded. The difference in average turf length before and after feeding was analysed using paired t tests. Normality was assessed using Shapiro–Wilks tests; all data were normally distributed. The difference in algal turf coverage before and after feeding was analysed using generalised linear mixed-effects models (GLMMs) with a binomial error distribution and logit link. Condition (before vs. after) was treated as a fixed effect, and rock ID was treated as a random effect to account for non-independence arising from measuring from the same rocks. Assumptions of the models were assessed using residual plots. All statistical analyses used the statistical software R (R Core Team 2014) using the *lme4* package (Bates et al. [2015\)](#page-8-0).

Examination of ingested material

Five C. striatus and five A. nigrofuscus were collected using barrier nets and immediately euthanised (clove oil), placed on ice and then frozen for later examination. The average total length of C. striatus and A. nigrofuscus specimens was 188 ± 12.6 and 145.2 ± 8.7 mm, respectively. Ingesta from the anterior portion of the intestines and the stomach were examined separately under a dissecting microscope $(10-40\times)$ for each specimen. The stomach and anterior intestine were examined separately to ensure any differences between species were not a result of variation in the anatomical structure of the stomach (Choat [1991](#page-8-0)). Following Wilson and Bellwood ([1997\)](#page-10-0), a 15×15 square grid with 50 randomly marked quadrats was fixed to the underside of a glass petri dish. Samples were spread evenly over the petri dish, and the dominant item (by area) in each quadrat was recorded as well as any other material present. Material was categorised into detritus, algae or sediment, with the term detritus used in the broad sense to describe amorphous organic material with no visible structure, in all cases consisting of opaque, flocculent material (following Wilson and Bellwood [1997](#page-10-0)). This material is not detritus sensu stricto, as it is likely to have contained some living material such as bacteria, microalgae and fungi (Wilson and Bellwood [1997](#page-10-0)). To analyse the differences in the frequency of occurrence of algal material compared to other matter (sediment or detritus) in the gut contents of C. striatus versus A. nigrofuscus, Pearson's Chi-squared tests with Yates' continuity correction were performed separately on stomach and intestinal data. The tests were performed in the statistical software R (R Core Team [2014](#page-9-0)).

Results

Algal removal experiment

Feeding by C. striatus did not result in a significant decrease in turf length ($t_9 = 1.72$ $t_9 = 1.72$, $p = 0.12$; Fig. 2a) or in

Fig. 2 Mean (percentage \pm SE) reduction of algal turf a length and b area coverage following 1 h of exposure without fish present (control) and after 1 h of feeding by the lined bristletooth, Ctenochaetus striatus, and the brown surgeonfish, Acanthurus nigrofuscus. Asterisk denotes a significant difference before versus after exposure ($\alpha = 0.05$)

the cover of turf algae (GLMM; $z = 0.33$, $p = 0.74$; Fig. 2b; ESM Table S2). On rocks exposed to C. striatus, algal turfs appeared largely unchanged following feeding (Fig. [3](#page-5-0)c, e) with an average decrease in turf length of only 0.2 ± 0.1 mm representing a reduction of $5.2 \pm 2.7\%$. Additionally, the mean proportion of area covered by turfing algae decreased by only $1.3 \pm 3.9\%$. Visually, the only change was a reduction in particulate loads.

By contrast, feeding by A. nigrofuscus significantly decreased turf length ($t_9 = 11.59, p \lt 0.0001$; Figs. 2a, [3d](#page-5-0), f) and the area covered by turf algae (GLMM; $z = 3.59$, $p = \langle 0.001;$ Figs. 2b, [3d](#page-5-0), f; ESM Table S2). On average, A. *nigrofuscus* reduced turf length by 2.1 ± 0.2 mm representing a reduction of $51.2 \pm 2.4\%$ in 1 h of feeding (Fig. 2a). The mean area covered by turf algae was also reduced by $14.7 \pm 4.0\%$. Only short, well-cropped algal turfs remained following exposure to A. nigrofuscus.

There was no significant difference in turf length on control rocks ($t_{19} = 0.62$, $p = 0.55$; Fig. 2a) or proportion of area covered (GLMM; $z = -0.17$, $p = 0.86$; Fig. 2b;

ESM Table S2). During the 1-h feeding trials, C. striatus took an average of 592.5 \pm 108.8 bites on the rocks while A. *nigrofuscus* took an average of 1583.2 ± 159.2 bites.

Ingested material

The material ingested by C. striatus on the reef was dominated by detritus and sediment (Fig. [4](#page-6-0)a, c; ESM Fig. S1). In C. striatus intestinal contents, detritus was the dominant category in $68.0 \pm 8.2\%$ of quadrats. In the stomach of C. striatus, detritus was dominant in 64.4 \pm 2.7% of quadrats. Algae were never the dominant category in intestinal contents quadrats and were only dominant in $0.4 \pm 0.4\%$ of stomach sample quadrats. Although algae were present in 37.6 ± 10.8 and $54 \pm 4.7\%$ of quadrats of C. striatus intestine and stomach samples, respectively, this was generally due to the presence of a single algal filament (ESM Table S3). By contrast, in A. nigrofuscus, algae were the dominant category in 79.6 \pm 3.8 and 94.7 \pm 2.9% of quadrats of intestine and stomach samples, respectively (Fig. [4](#page-6-0)b, d; ESM Table S3). Algal material was significantly more dominant in quadrats in A. nigrofuscus than in C. striatus in both stomach $(\chi^2 = 327.25, df = 1, p = 0.0001)$ and intestinal samples ($\chi^2 = 358.60$, $df = 1$, $p = \langle 0.0001 \rangle$.

Discussion

Ctenochaetus striatus removed little algae from mature algal turfs, even after a period of intensive feeding. In marked contrast, A. nigrofuscus removed substantial quantities of algae, leaving noticeably cropped algal turfs. These differences were mirrored by the material ingested by wild-caught specimens. The gut contents of C. striatus predominantly contained detritus and sediment, while A. nigrofuscus predominantly contained algae. Our findings support conclusions drawn from behavioural (Choat and Bellwood [1985\)](#page-9-0), nutritional (Randall [1955](#page-9-0); Choat et al. [2002](#page-9-0); Crossman et al. [2005](#page-9-0); Clements et al. [2009;](#page-9-0) Brandl et al. [2015\)](#page-8-0) and morphological studies (Randall [1955](#page-9-0); Jones [1968](#page-9-0); Purcell and Bellwood [1993](#page-9-0); Bellwood et al. [2014](#page-8-0)), in that C. striatus did not remove significant amounts of algae from mature algal turfs and therefore are unlikely to be significant herbivores on coral reefs. The apparent inability of Ctenochaetus species to remove mature turfing algae was noted in several early ecological studies of coral reefs (Randall [1955](#page-9-0), [1961](#page-9-0)). Indeed, our findings highlight the fundamental differences in the way two superficially similar surgeonfishes affect algal turfs on coral reefs and the interspecific variability which may occur between fishes that have been classified under the same functional identity, i.e. herbivorous fishes.

Fig. 3 a Lined bristletooth, Ctenochaetus striatus. b The brown surgeonfish, Acanthurus nigrofuscus (photographs by CHRG). c Feeding surface before exposure to C. striatus. d Feedings surface before

Visually, C. striatus and A. nigrofuscus are similar but they interact with algal turfs in distinctly different ways. The spatulate teeth (Fig. [1b](#page-1-0)), small jaw opening (113°) and rapid biting behaviour of A. nigrofuscus are well suited to nipping off algal filaments (Purcell and Bellwood [1993](#page-9-0)). By contrast, C. *striatus* take slower bites, but can open their jaws to nearly 180° (Purcell and Bellwood [1993](#page-9-0)), allowing fish to come in close contact with the substratum and to selectively brush fine particulate material from turfing algae using their comb-like teeth (Fig. [1a](#page-1-0); Purcell and Bellwood [1993](#page-9-0); Tebbett et al. [2017b\)](#page-10-0). This feeding behaviour and morphology does not appear to be consistent with cropping algae.

Ctenochaetus striatus does not crop algal turfs, but it could be argued that their wide gape (178°) , which allows extended contact with the benthos (Purcell and Bellwood [1993\)](#page-9-0), could increase removal of entire algal filaments as in scraping parrotfishes (Bonaldo et al. [2014\)](#page-8-0). However,

the morphology and teeth of C. striatus make this unlikely. e C. striatus and f A. nigrofuscus. Scale bars are approximate

exposure to A. nigrofuscus. Feeding surfaces after 1 h of feeding by

Indeed, no significant reduction in algal turf coverage on rocks exposed to C. striatus was recorded and it appears that mature algal turfs, which are firmly attached to the substratum, are not readily removed by the brushing feeding behaviour. While C. striatus may remove small amounts of loose or long-filamented turfing algae, an expected consequence of feeding on the EAM, such removal is minimal compared to turf-feeding herbivorous fishes like A. nigrofuscus. Observations in both the field and aquaria suggest that when C. striatus dislodge algae it often becomes caught in their teeth, causing visible annoyance to the fish (Randall [1955;](#page-9-0) Purcell and Bellwood [1993](#page-9-0)). This may explain the slight (nonsignificant) reduction in mean algal turf length and proportional area covered on surfaces exposed to C. striatus, as longer algal filaments are more likely to become entangled in their teeth. Such removal appears to be incidental as no significant

Fig. 4 Analysis of ingested material showing the percentage (mean \pm SE) of quadrats in which each dietary category (detritus [brown], sediment [yellow] and algae [green]) was dominant by area in a the gizzard-like stomach of the lined bristletooth, Ctenochaetus striatus ($n = 5$), **b** the stomach of the brown surgeonfish, *Acanthurus* nigrofuscus (n = 3) and in the intestinal tract of **c** C. striatus (n = 5) and **d** A. *nigrofuscus* ($n = 5$)

reductions in algae were recorded even after an average of 592.5 (\pm 108.8) bites in an area of approximately 50 cm² in 1 h.

As commonly reported, the gut contents of C. striatus specimens from the reef did contain some algae (Polunin and Klumpp [1989](#page-9-0); Choat and Clements [1998;](#page-9-0) Choat et al. [2002\)](#page-9-0). This algae may be partly from incidental removal and ingestion, but it may also be the result of secondary ingestion of algal material through coprophagy. Many herbivorous fishes, including A. nigrofuscus, void their faeces over the reef (Fishelson et al. [1985](#page-9-0); Clements [1991](#page-9-0); Bonaldo et al. [2014\)](#page-8-0) and much of the algal material contained within the faeces remains structurally intact and can continue to grow (Vermeij et al. [2013](#page-10-0); Tâmega et al. [2016](#page-10-0)). Ctenochaetus striatus feeds over the same substrata as these fishes, especially herbivorous acanthurids (Choat and Bellwood [1985\)](#page-9-0), and it is likely that C. striatus may ingest a considerable amount of faecal matter (Clements [1991](#page-9-0)). Indeed, of the faeces from the herbivorous surgeonfishes A. nigricans and A. lineatus that was eaten by fishes, 42 and 37%, respectively, was consumed by C. striatus (Robertson [1982\)](#page-10-0). Thus, rather than directly removing mature turf algae from the reef, C. striatus may act as a secondary herbivore ingesting algae and detritus from the faeces of other reef organisms.

In the experiments, A. *nigrofuscus* took substantially more bites during the exposure period than C. striatus; however, this is unlikely to contribute significantly to the differences observed. The bites of each species are

fundamentally different and although a slower feeder, the bite area of C. striatus is much larger than A. nigrofuscus (Purcell and Bellwood [1993](#page-9-0)). A similar disparity in bite rates is seen in the wild, with A. nigrofuscus having a higher bite rate than *C. striatus* (Clements [1991](#page-9-0); Polunin et al. [1995](#page-9-0)). In an experimental study examining algal turf removal, A. nigrofuscus took six times more bites than C. striatus (Marshell and Mumby [2012\)](#page-9-0). Although a disparity in bite rates was reported, similar to the present study, it is interesting to consider the contrasting results in terms of the fishes' effects on algal turfs.

In feeding trials conducted by Marshell and Mumby [\(2012](#page-9-0)), C. striatus removed significantly more algae than A. nigrofuscus. Such contrasting results may be explained by two key differences between our study and the work of Marshell and Mumby. First, the two studies examined different metrics. Marshell and Mumby ([2012\)](#page-9-0) examined algal turf biomass, while we used measurements of algal turf length and area coverage. Although these metrics are not the same, if C. striatus were removing significant amounts of algal biomass it would be expected that at least one of our metrics would show a significant decrease. Clearly, this was not the case.

The second difference lies in the nature of the algae examined. The current study used reef-grown algal turfs that were at least 6 months old growing on planar dead coral rocks. Consequently, the coral rocks supported mature algal turfs. By contrast, the algal turfs used by Marshell and Mumby ([2012\)](#page-9-0) were grown on artificial surfaces (rough "limestone" tiles, orange in colour) which were ''preconditioned on the reef.'' These tiles were, however, scrubbed following conditioning to begin with an algal biomass of zero. They were then placed in flowthrough aquaria to develop sparse algal turfs after 2 weeks and dense algal turfs after 6 weeks (Marshell and Mumby [2012](#page-9-0)). The algal communities in the two studies were likely to differ in two fundamental ways. First, differences in algal communities may arise due to variation in the texture and chemistry between the two settlement substrata (natural coral rocks vs. artificial tiles), which can affect the species of algae that settle and develop (Harlin and Lindbergh [1977](#page-9-0); Borowitzka et al. [1978](#page-8-0); Hixon and Brostoff [1985](#page-9-0); Diaz-Pulido and McCook [2002;](#page-9-0) Smith et al. [2010](#page-10-0)). Indeed, coral rock substrata tend to support more later successional algae compared to artificial settling substrata and, as algal turfs can penetrate coral rock substrata, they may be more firmly attached (Hixon and Brostoff [1996](#page-9-0); Diaz-Pulido and McCook [2002\)](#page-9-0).

The second major difference between these studies lies in the age of the algal turfs used. The algal community used by Marshell and Mumby ([2012\)](#page-9-0) was less than 6 weeks old and consequently was likely to be dominated by early successional algae which are only superseded by more

mature forms after several months (Borowitzka et al. [1978](#page-8-0); Diaz-Pulido and McCook [2002](#page-9-0)). Early successional algal communities are dominated by diatoms, coccoids and bluegreen filamentous algae, while later successional stage turf algal communities are more species rich and include complex algal forms (Borowitzka et al. [1978](#page-8-0); Scott and Russ [1987](#page-10-0); Diaz-Pulido and McCook [2002](#page-9-0)). Early successional algal communities are less firmly attached to substrata (Borowitzka et al. [1978;](#page-8-0) Diaz-Pulido and McCook [2002](#page-9-0)) and consequently may be removed far more easily than later successional algal turfs such as those used in our study. As Ctenochaetus are able to open their jaws to nearly 180° to feed on planar surfaces (Randall 1955 ; Purcell and Bellwood [1993\)](#page-9-0), they are likely to remove loose algae from planar, smooth artificial tiles far more effectively than A. nigrofuscus. Indeed, early successional "algal turfs" (diatoms and cyanobacteria) may form an important component of the nutritional ecology of C. striatus where available (Polunin et al. [1995;](#page-9-0) Choat et al. [2002;](#page-9-0) Wilson et al. [2003](#page-10-0)). This is particularly likely if they feed on smooth surfaces covered with calcareous algae where the removal of microalgal fouling may be important. However, the feeding activity of C. striatus appears to have a minimal effect on later successional stage algal turfs, which are often the dominant benthic covering on coral reefs (Wismer et al. [2009](#page-10-0)). The early successional algae used by Marshell and Mumby ([2012\)](#page-9-0) are also likely to be particularly amenable to the use of biomass as a metric (discussed above), due to the ease with which these algae can be scraped from the substratum for measurement.

In addition to the differences outlined above, other factors may also have contributed to the disparity in the results. In particular, the sediment loads within the algal turfs in the present study were approximately six to thirteen times lower than those in Marshell and Mumby ([2012\)](#page-9-0) and were more similar to loads found in reef crest algal turfs (Goatley and Bellwood [2012](#page-9-0); Tebbett et al. [2017c](#page-10-0)), the predominant feeding habitat of the two surgeonfishes (Russ [1984;](#page-10-0) Goatley and Bellwood [2010\)](#page-9-0). Sediments suppress the feeding rates of herbivorous/detritivorous fishes (Goatley and Bellwood [2012](#page-9-0); Gordon et al. [2016](#page-9-0); Tebbett et al. [2017a,](#page-10-0) [b](#page-10-0)) which may explain why we found higher bite rates in both fish species. Importantly, this means that any effect the fishes may have had on algal turfs in the present study should have been more pronounced.

It should be noted that we did not directly examine whether *C. striatus* was ingesting and assimilating the particulate material used in the experiments. Such an examination may have provided supporting evidence of their ability to brush detritus from algal turfs. However, visual observations and video recordings both strongly suggest that particulates were removed. Nevertheless, the chief aim of this study was to assess the extent of algal removal from mature algal turfs by the two species, which is evidently minimal in the case of C. striatus.

Ctenochaetus striatus may be herbivorous in regards to their ability to remove early successional algal communities from planar surfaces or microalgae from algal turfs, but they do not appear to play a significant role in the removal of algae from mature algal turfs. Inevitably, this raises the question: what are the main functional roles of C. striatus? The most likely answer lies in their contribution to detritivory and EAM sediment dynamics. Detritivory is a central trophic pathway on coral reefs, and given the size, abundance and volume of detritus removed by C. striatus, this species is probably one of the most important detritivorous fish species on Indo-Pacific coral reefs (Wilson et al. [2003](#page-10-0); Crossman et al. [2005](#page-9-0)). However, the term ''detritus'' encompasses items such as diatoms, microalgae, microbes, cyanobacteria and faeces which may also be important nutritional resources (Polunin et al. [1995](#page-9-0); Choat et al. [2002;](#page-9-0) Wilson et al. [2003](#page-10-0); Clements et al. [2016](#page-9-0)). While acknowledging these other resources, given their main feeding mode, we consider C. striatus to be ''detrital aggregate brushers.''

When brushing detrital aggregates from the EAM, C. striatus also removes and ingests inorganic sediments, making it a key player in EAM sediment dynamics (Goatley and Bellwood [2010](#page-9-0); Krone et al. [2011;](#page-9-0) Tebbett et al. [2017b\)](#page-10-0). Unlike many fishes, C. striatus has distinct defecation areas, off the reef and/or in deeper water (Krone et al. [2008;](#page-9-0) Goatley and Bellwood [2010](#page-9-0)). They therefore export ingested sediment away from feeding areas (Krone et al. [2008](#page-9-0), [2011](#page-9-0); Goatley and Bellwood [2010](#page-9-0)). This role may be vital to coral reefs as EAM sediments suppress herbivory (Goatley and Bellwood [2012;](#page-9-0) Gordon et al. [2016](#page-9-0)) and coral recruitment (Birrell et al. [2005;](#page-8-0) Diaz-Pulido et al. [2010;](#page-9-0) Perez et al. [2014](#page-9-0)). By reducing EAM sediment loads, C. striatus may facilitate herbivory in these environments (Choat [1991](#page-8-0); Goatley and Bellwood [2010\)](#page-9-0) underpinning both the preservation of short productive algal turfs (SPATs sensu Goatley et al. [2016](#page-9-0)) and coral replenishment (Brandl and Bellwood [2016\)](#page-8-0). Interestingly, if C. striatus also ingests viable algal material from the faeces of other fishes (as discussed above) their defecation behaviour may also incidentally help limit algal development and expansion. Ctenochaetus striatus may therefore perform a secondary ''herbivory'' function on coral reefs by harvesting loose, but viable, algal material and exporting it off the reef. Exploring the potential for C. striatus to act as a secondary herbivore in this manner may be an important topic for future research.

The importance of the different functional roles that C. striatus fulfil could also change depending on the specific context. Specifically, the ability of C. striatus to remove early successional algae may be important following major disturbance events. By removing early successional algae that colonise dead coral skeletons, C. striatus could slow or prevent the development of mature algal turfs (Hixon and Brostoff [1996;](#page-9-0) Steneck [1997](#page-10-0)). This may assist the recovery of coral reefs, as mature algal turfs impede coral recruitment (Arnold et al. 2010; Diaz-Pulido et al. [2010\)](#page-9-0). However, as C. striatus predominantly feeds on smooth surfaces (Choat and Bellwood [1985](#page-9-0); Brandl et al. 2015), removal of early successional algae from complex or branching dead coral skeletons by C. striatus is unlikely. Once mature algal turfs develop that are not readily removed by C. striatus, the functional roles of C. striatus are predominantly detritivory and sediment transport.

In addition to the functional roles discussed so far, it has also been suggested that C. striatus plays a role in bioerosion in the Red Sea, through the use of a hard palate structure that could be used to rasp the substratum (Schuhmacher et al. [2008;](#page-10-0) Krone et al. [2011\)](#page-9-0), although Krone et al. (2011) (2011) concluded that bioerosion by C. striatus was only a minor role compared to the removal of loose sediments. It is interesting that although C. striatus is abundant, widely distributed and often studied, bioerosion and the presence of a hard palate structure has not been reported outside of the Red Sea. Further investigation of bioerosion and the anatomy of C. striatus in other geographic localities could be worthwhile.

Assigning a single overarching functional role to C. striatus is difficult. Indeed, the debate and confusion surrounding the functional role of this species may stem from a desire to simplify and apply broad categorisations to complex ecosystems. Evidently, C. striatus plays a variable role in many functions on coral reefs; categorising it into a single functional group may overlook the contribution that this species makes to other functions. While functional classifications are useful management tools (Bellwood et al. 2004) care should be taken in their use to assess ecological processes as they may conceal intra-functional group variability (Clements et al. [2009](#page-9-0); Streit et al. [2015](#page-10-0); Kelly et al. [2016](#page-9-0)). Functional classifications fail to consider that the role a fish fulfils is rarely ''black and white'' and in some cases the contribution a fish makes to a particular function is better viewed as a sliding scale, particularly evident in the extent to which C. striatus functions as a herbivore on coral reefs.

The key question in our study was: to what extent is C. striatus herbivorous and how does this compare to A. nigrofuscus? The answer appears to be that compared to A. nigrofuscus, C. striatus has a minimal effect on mature algal turfs with only small amounts of algae ingested under natural settings. The way these two fishes interact with algal turfs and consequently the functional roles they perform on coral reefs are distinctly different. Although C. striatus may remove loosely attached early successional algal communities, predominantly composed of diatoms and cyanobacteria, categorising them as significant herbivores on coral reefs should be done with caution. Ctenochaetus striatus is unlikely to directly remove significant amounts of algae from mature algal turfs. Ctenochaetus striatus appears to be predominantly detritivorous removing particulates from the EAM or reef surface. However, as C. striatus appears to fulfil numerous functional roles on coral reefs, classifying this species into a single functional category may underestimate the extent of its importance in other ecological processes.

Acknowledgements We thank R. Streit, J. Khan, M. McFarland, P. O'Brien and Lizard Island Research Staff for field support; J. Day and K. Miller for assistance with video processing, K. Blake and S. Askew for assistance with scanning electron micrography, two anonymous reviewers for helpful and thought-provoking comments and the Australian Research Council for financial support (DRB).

References

- Arnold SN, Steneck RS, Mumby PJ (2010) Running the gauntlet: inhibitory effects of algal turfs on the processes of coral recruitment. Mar Ecol Prog Ser 414:91–105
- Bates D, Maechler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using lme4. J Stat Softw 67:1–48
- Bellwood DR, Hughes TP, Folke C, Nyström M (2004) Confronting the coral reef crisis. Nature 429:827–833
- Bellwood DR, Hoey AS, Bellwood O, Goatley CHR (2014) Evolution of long-toothed fishes and the changing nature of fish–benthos interactions on coral reefs. Nat Commun 5:3144
- Birrell CL, McCook LJ, Willis BL (2005) Effects of algal turfs and sediment on coral settlement. Mar Pollut Bull 51:408–414
- Bonaldo RM, Bellwood DR (2009) Dynamics of parrotfish grazing scars. Mar Biol 156:771–777
- Bonaldo RM, Hoey AS, Bellwood DR (2014) The ecosystem roles of parrotfishes on tropical reefs. Oceanogr Mar Biol Annu Rev 52:81–132
- Borowitzka MA, Larkum AWD, Borowitzka LJ (1978) A preliminary study of algal turf communities of a shallow coral reef lagoon using an artificial substratum. Aquat Bot 5:365–381
- Bouchon-Navaro Y, Harmelin-Vivien ML (1981) Quantitative distribution of herbivorous reef fishes in the Gulf of Aqaba (Red Sea). Mar Biol 63:79–86
- Brandl SJ, Bellwood DR (2014) Individual-based analyses reveal limited functional overlap in a coral reef fish community. J Anim Ecol 83:661–670
- Brandl SJ, Bellwood DR (2016) Microtopographic refuges shape consumer–producer dynamics by mediating consumer functional diversity. Oecologia 182:203–217
- Brandl SJ, Robbins WD, Bellwood DR (2015) Exploring the nature of ecological specialization in a coral reef fish community: morphology, diet and foraging microhabitat use. Proc R Soc Lond B Biol Sci 282:20151147
- Cheal AJ, Emslie M, Miller I, Sweatman H (2012) The distribution of herbivorous fishes on the Great Barrier Reef. Mar Biol 159:1143–1154
- Choat JH (1991) The biology of herbivorous fishes on coral reefs. In: Sale PF (ed) The ecology of fishes on coral reefs. Academic Press, San Diego, pp 120–155
- Choat JH, Bellwood DR (1985) Interactions amongst herbivorous fishes on a coral reef: influence of spatial variation. Mar Biol 89:221–234
- Choat JH, Clements KD (1998) Vertebrate herbivores in marine and terrestrial environments: a nutritional ecology perspective. Annu Rev Ecol Syst 29:375–403
- Choat JH, Clements KD, Robbins WD (2002) The trophic status of herbivorous fishes on coral reefs 1: dietary analyses. Mar Biol 140:613–623
- Chong-Seng KM, Mannering TD, Pratchett MS, Bellwood DR, Graham NAJ (2012) The influence of coral reef benthic condition on associated fish assemblages. PLoS ONE 7:e42167
- Clements KD (1991) Gut microorganisms of surgeonfishes (family Acanthuridae). Ph.D. thesis, James Cook University, Townsville
- Clements KD, Raubenheimer D, Choat JH (2009) Nutritional ecology of marine herbivorous fishes: ten years on. Funct Ecol 23:79–92
- Clements KD, German DP, Piche´ J, Tribollet AD, Howard Choat J (2016) Integrating ecological roles and trophic resources on coral reefs: multiple lines of evidence identify parrotfishes as microphages. Biol J Linn Soc. doi:[10.1111/bij.12914](http://dx.doi.org/10.1111/bij.12914)
- Crossman DJ, Choat JH, Clements KD (2005) Nutritional ecology of nominally herbivorous fishes on coral reefs. Mar Ecol Prog Ser 296:129–142
- Diaz-Pulido G, McCook LJ (2002) The fate of bleached corals: patterns and dynamics of algal recruitment. Mar Ecol Prog Ser 232:115–128
- Diaz-Pulido G, Harii S, McCook LJ, Hoegh-Guldberg O (2010) The impact of benthic algae on the settlement of a reef-building coral. Coral Reefs 29:203–208
- Doropoulos C, Roff G, Bozec Y-M, Zupan M, Werminghausen J, Mumby PJ (2016) Characterizing the ecological trade-offs throughout the early ontogeny of coral recruitment. Ecol Monogr 86:20–44
- Fishelson L, Montgomery WL, Myrberg AA (1985) A Unique symbiosis in the gut of tropical herbivorous surgeonfish (Acanthuridae: Teleostei) from the Red Sea. Science 229:49–51
- Goatley CHR, Bellwood DR (2010) Biologically mediated sediment fluxes on coral reefs: sediment removal and off-reef transportation by the surgeonfish Ctenochaetus striatus. Mar Ecol Prog Ser 415:237–245
- Goatley CHR, Bellwood DR (2012) Sediment suppresses herbivory across a coral reef depth gradient. Biol Lett 8:1016–1018
- Goatley CHR, Bellwood DR (2013) Ecological consequences of sediment on high-energy coral reefs. PLoS ONE 8:e77737
- Goatley CHR, Bonaldo RM, Fox RJ, Bellwood DR (2016) Sediments and herbivory as sensitive indicators of coral reef degradation. Ecol Soc 21:29
- Gordon SE, Goatley CHR, Bellwood DR (2016) Low-quality sediments deter grazing by the parrotfish Scarus rivulatus on inner-shelf reefs. Coral Reefs 35:285–291
- Graham NAJ, Chabanet P, Evans RD, Jennings S, Letourneur Y, Aaron Macneil M, Mcclanahan TR, Ohman MC, Polunin NVC, Wilson SK (2011) Extinction vulnerability of coral reef fishes. Ecol Lett 14:341–348
- Harlin MM, Lindbergh JM (1977) Selection of substrata by seaweeds: optimal surface relief. Mar Biol 40:33–40
- Hiatt RW, Strasburg DW (1960) Ecological relationships of the fish fauna on coral reefs of the Marshall Islands. Ecol Monogr 30:65–127
- Hixon MA, Brostoff WN (1985) Substrate characteristics, fish grazing, and epibenthic reef assemblages off Hawaii. Bull Mar Sci 37:200–213
- Hixon MA, Brostoff WN (1996) Succession and herbivory: effects of differential fish grazing on Hawaiian coral-reef algae. Ecol Monogr 66:67–90
- Johansson CL, van de Leemput IA, Depczynski M, Hoey AS, Bellwood DR (2013) Key herbivores reveal limited functional redundancy on inshore coral reefs. Coral Reefs 32:963–972
- Jones R (1968) Ecological relationships in Hawaiian and Johnston Island Acanthuridae (surgeonfishes). Micronesica 4:309–361
- Kelly ELA, Eynaud Y, Clements SM, Gleason M, Sparks RT, Williams ID, Smith JE (2016) Investigating functional redundancy versus complementarity in Hawaiian herbivorous coral reef fishes. Oecologia 182:1151–1163
- Krone R, Paster M, Schuhmacher H (2011) Effect of the surgeonfish Ctenochaetus striatus (Acanthuridae) on the processes of sediment transport and deposition on a coral reef in the Red Sea. Facies 57:215–221
- Krone R, Bshary R, Paster M, Eisinger M, van Treeck P, Schuhmacher H (2008) Defecation behaviour of the lined bristletooth surgeonfish Ctenochaetus striatus (Acanthuridae). Coral Reefs 27:619–622
- Marshell A, Mumby PJ (2012) Revisiting the functional roles of the surgeonfish Acanthurus nigrofuscus and Ctenochaetus striatus. Coral Reefs 31:1093–1101
- Marshell A, Mumby PJ (2015) The role of surgeonfish (Acanthuridae) in maintaining algal turf biomass on coral reefs. J Exp Mar Bio Ecol 473:152–160
- Montgomery WL, Myrberg AA, Fishelson L (1989) Feeding ecology of surgeonfishes (Acanthuridae) in the northern Red Sea, with particular reference to Acanthurus nigrofuscus (Forsskål). J Exp Mar Biol Ecol 132:179–207
- Nash KL, Graham NAJ, Bellwood DR (2013) Fish foraging patterns, vulnerability to fishing, and implications for the management of ecosystem function across scales. Ecol Appl 23:1632–1644
- Nyström M (2006) Redundancy and response diversity of functional groups: implications for the resilience of coral reefs. Ambio 35:30–35
- Perez K III, Rodgers KS, Jokiel PL, Lager CV, Lager DJ (2014) Effects of terrigenous sediment on settlement and survival of the reef coral Pocillopora damicornis. PeerJ 2:e387
- Plass-Johnson JG, Ferse SCA, Jompa J, Wild C, Teichberg M (2015) Fish herbivory as key ecological function in a heavily degraded coral reef system. Limnol Oceanogr 60:1382–1391
- Polunin NVC, Klumpp DW (1989) Ecological correlates of foraging periodicity in herbivorous reef fishes of the Coral Sea. J Exp Mar Biol Ecol 126:1–20
- Polunin NVC, Harmelin-Vivien M, Galzin R (1995) Contrasts in algal food processing among five herbivorous coral-reef fishes. J Fish Biol 47:455–465
- Pratchett MS, Hoey AS, Wilson SK, Messmer V, Graham NAJ (2011) Changes in biodiversity and functioning of reef fish assemblages following coral bleaching and coral loss. Diversity 3:424–452
- Purcell SW (2000) Association of epilithic algae with sediment distribution on a windward reef in the northern Great Barrier Reef, Australia. Bull Mar Sci 66:199–214
- Purcell SW, Bellwood DR (1993) A functional analysis of food procurement in two surgeonfish species, Acanthurus nigrofuscus and Ctenochaetus striatus (Acanthuridae). Environ Biol Fishes 37:139–159
- Purcell SW, Bellwood DR (2001) Spatial patterns of epilithic algal and detrital resources on a windward coral reef. Coral Reefs 20:117–125
- R Core Team (2014) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Randall JE (1955) A contribution to the biology of the Acanthuridae (Surgeon Fishes). Ph.D. thesis. University of Hawaii, Hawaii
- Randall JE (1961) Overgrazing of algae by herbivorous marine fishes. Ecology 42:812
- Randall JE (2005) Reef and shore fishes of the South Pacific: New Caledonia to Tahiti and the Pitcairn Islands. University of Hawaii Press, Honolulu
- Robertson D (1982) Fish feces as fish food on a Pacific coral reef. Mar Ecol Prog Ser 7:253–265
- Robertson DR, Gaines SD (1986) Interference competition structures habitat use in a local assemblage of coral reef surgeonfishes. Ecology 67:1372–1383
- Russ GR (1984) Distribution and abundance of herbivorous grazing fishes in the central Great Barrier Reef. II. Patterns of zonation of mid-shelf and outershelf reefs. Mar Ecol Prog Ser 20:35–44
- Schuhmacher H, Krone R, Treeck P Van (2008) Underestimated eroder among reef fishes—experimental comparison between Ctenochaetus striatus and Acanthurus nigrofuscus (Acanthuridae). In: Proceedings of 11th international coral reef symposium, vol 10. pp 331–334
- Scott FJ, Russ GR (1987) Effects of grazing on species composition of the epilithic algal community on coral reefs of the central Great Barrier Reef. Mar Ecol Prog Ser 39:293–304
- Smith JE, Hunter CL, Smith CM (2010) The effects of top-down versus bottom-up control on benthic coral reef community structure. Oecologia 163:497–507
- Steneck RS (1997) Crustose corallines, other algal functional groups, herbivores and sediments: complex interactions along reef productivity gradients. In: Proceedings of the 8th international coral reef symposium, vol 1. pp 695–700
- Streit RP, Hoey AS, Bellwood DR (2015) Feeding characteristics reveal functional distinctions among browsing herbivorous fishes on coral reefs. Coral Reefs 34:1037–1047
- Tâmega FTS, Figueiredo MAO, Ferreira CEL, Bonaldo RM (2016) Seaweed survival after consumption by the greenbeak parrotfish, Scarus trispinosus. Coral Reefs 35:329–334
- Tebbett SB, Goatley CHR, Bellwood DR (2017a) The effects of algal turf sediments and organic loads on feeding by coral reef surgeonfishes. PLoS ONE 12:e0169479
- Tebbett SB, Goatley CHR, Bellwood DR (2017b) Fine sediments suppress detritivory on coral reefs. Mar Pollut Bull 114:934–940
- Tebbett SB, Goatley CHR, Bellwood DR (2017c) Algal turf sediments and sediment production by parrotfishes across the continental shelf of the northern Great Barrier Reef. PLoS ONE 12:e0170854
- Tenore KR (1981) Organic nitrogen and caloric content of detritus: 1. utilization by the deposit-feeding polychaete, Capitella capitata. Estuar Coast Shelf Sci 12:39–47
- Vermeij MJA, van der Heijden RA, Olthuis JG, Marhaver KL, Smith JE, Visser PM (2013) Survival and dispersal of turf algae and macroalgae consumed by herbivorous coral reef fishes. Oecologia 171:417–425
- Wilson SK, Bellwood DR (1997) Cryptic dietary components of territorial damselfishes (Pomacentridae, Labroidei). Mar Ecol Prog Ser 153:299–310
- Wilson SK, Bellwood DR, Choat JH, Furnas MJ (2003) Detritus in the epilithic algal matrix and its use by coral reef fishes. Oceanogr Mar Biol Annu Rev 41:279–309
- Wismer S, Hoey AS, Bellwood DR (2009) Cross-shelf benthic community structure on the Great Barrier Reef: relationships between macroalgal cover and herbivore biomass. Mar Ecol Prog Ser 376:45–54