ONCOLOGY



Distinct patterns of the natural evolution of soft tissue sarcomas on pre-treatment MRIs captured with delta-radiomics correlate with gene expression profiles

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Abstract

Objectives Radiomics of soft tissue sarcomas (STS) is assumed to correlate with histologic and molecular tumor features, but radiogenomics analyses are lacking. Our aim was to identify if distinct patterns of natural evolution of STS obtained from consecutive pre-treatment MRIs are associated with differential gene expression (DGE) profiling in a pathway analysis.

Methods All patients with newly diagnosed STS treated in a curative intent in our sarcoma reference center between 2008 and 2019 and with two available pre-treatment contrast-enhanced MRIs were included in this retrospective study. Radiomics features (RFs) were extracted from fat-sat contrast-enhanced T1-weighted imaging. Log ratio and relative change in RFs were calculated and used to determine grouping of samples based on a consensus hierarchical clustering. DGE and oncogenesis pathway analysis were performed in the delta-radiomics groups identified in order to detect associations between delta-radiomics patterns and transcriptomics features of STS. Secondarily, the prognostic value of the delta-radiomics groups was investigated.

Results Sixty-three patients were included (median age: 63 years, interquartile range: 52.5–70). The consensus clustering identified 3 reliable delta-radiomics patient groups (A, B, and C). On imaging, group B patients were characterized by increase in tumor heterogeneity, necrotic signal, infiltrative margins, peritumoral edema, and peritumoral enhancement before the treatment start (*p* value range: 0.0019–0.0244), and, molecularly, by downregulation of natural killer cell–mediated cytotoxicity genes and upregulation of Hedgehog and Hippo signaling pathways. Group A patients were characterized by morphological stability of pre-treatment MRI traits and no local relapse (log-rank p = 0.0277).

Conclusions This study highlights radiomics and transcriptomics convergence in STS. Proliferation and immune response inhibition were hyper-activated in the STS that were the most evolving on consecutive imaging. **Key Points**

- Three consensual and stable delta-radiomics clusters were identified and captured the natural patterns of morphological evolution of STS on pre-treatment MRIs.
- These 3 patterns were explainable and correlated with different well-known semantic radiological features with an ascending gradient of pejorative characteristics from the A group to C group to B group.
- Gene expression profiling stressed distinct patterns of up/downregulated oncogenetic pathways in STS from B group in keeping with its most aggressive radiological evolution.

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Keywords Magnetic resonance imaging · Sarcoma · Gene expression profiling · Transcript expression analysis · Prognosis

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| AC | Absolute change |
|--------|--|
| CE | Contrast-enhanced |
| CI | Confidence interval |
| DGE | Differential gene expression |
| FNCLCC | French "Fédération Nationale des Centres |
| | de Lutte Contre le Cancer" |
| FS | Fat sat |
| HR | Hazard ratio |
| IQR | Interquartile range |
| IRB | Institutional Review Board |
| LD | Longest diameter |
| LFS | Local relapse-free survival |
| LR | Log ratio |
| MFS | Metastatic relapse-free survival |
| MRI | Magnetic resonance imaging |
| NK | Natural killer |
| OS | Overall survival |
| RF | Radiomics feature |
| RNA | Ribonucleic acids |
| SAM | Significance analysis of microarrays |
| SI | Signal intensity |
| STS | Soft tissue sarcoma |
| TSE | Turbo spin echo |
| TWAC | Time-weighted absolute change |
| TWRC | Time-weighted relative change |
| WI | Weighted imaging |

Introduction

Soft tissue sarcomas (STS) are rare malignant mesenchymal tumors characterized by their clinical, radiological, and molecular heterogeneity, which can lead to diagnostic uncertainties [1]. Consequently, the time elapsed from initial symptoms to final histological diagnosis can reach months during which the tumor can evolve naturally and grow without constraints [2].

MRI is the imaging of reference for the local staging of STS, treatment response assessment, and local relapse detection [3]. MRI enables to evaluate classical prognostic features such as size and depth, which are included in nomograms, as well as the surgical operability that guides the therapeutic strategy [3-5]. Moreover, baseline MRI features (i.e., peritumoral enhancement, necrotic signal, or intra-tumoral heterogeneity on conventional sequences) have been correlated with histologic grade and survivals [6, 7]. However, these "semantic" features are subjective [8] and do not account for the natural changes (i.e., in the absence of any medical

intervention) observed when multiple MRIs are performed during the diagnostic interval.

Yet, significant improvements in the quantification of the tumor radiological phenotype (or "radiophenotype") have been achieved through radiomics [9, 10]. Basically, radiomics correspond to the extensive quantification of the radiophenotype thanks to mathematical operators applied on any imaging modality. A key hypothesis is that the heterogeneity captured by radiomics radiophenotyping reflects the tumor histologic and molecular characteristics. The resulting radiomics features (RFs) are incorporated in supervised machine learning pipelines to predict the patients' outcomes or to identify associations with relevant histologic and molecular status, and unsupervised algorithms (i.e., algorithms that analyze and cluster unlabeled dataset according to the similarity of their RFs) in order to identify hidden patterns and subgroups of similar observations when there is no a priori hypothesis [9].

So far, radiomics approaches have been successfully applied to STS to predict the histologic grade [11–14], metastatic relapse and cancer-related death [12, 15–17], and treatment response [18–20]. Moreover, the interest of evaluating the changes in the STS radiophenotype during treatments through "delta-radiomics" approaches (i.e., quantitative variation in RFs between two evaluations) has been demonstrated during neoadjuvant chemotherapy and radiotherapy [18, 19].

However, as opposed to other neoplasms (such as glioma, urothelial carcinoma, lung adenocarcinoma, clear cell carcinoma, or ovarian cancer), correlations between radiomics, deltaradiomics, and transcriptomics features (i.e., corresponding to the expression levels and profiles of multiple ribonucleic acids [RNA] transcripts) in sarcoma are missing.

Considering the rapid evolution and diagnostic delays seen in STS patients, multidisciplinary boards in sarcoma reference centers sometimes require to update the imaging in order to refine the local staging. Comparing these pretreatment MRIs acquired before any medical intervention highlights distinct patterns of "natural" evolution of STS, from global morphological stability to appearance of strong intra-tumoral heterogeneity and infiltrative margins, which suggests different biological pathways involved in tumorigenesis. Our group has recently investigated the clinical interest of assessing natural radiological and radiomics changes in STS before starting initial treatments [21]. Surprisingly, the natural changes in RFs were not associated with histological type or simple histologic markers of proliferation like Ki-67 or mitotic count, which prompted us to hypothesize that delta-radiomics would rather reflect other oncogenetic pathways and could complement standard radiological and histologic analyses.

Consequently, our aim was to establish a reliable classification of the radiophenotype of STS based on the natural changes in RFs before treatment and using unsupervised algorithms, and to investigate whether those delta-radiomics radiophenotypes could be biologically explained through gene expression profiling.

Materials and methods

Study design

This single-center retrospective study was IRB-approved. Patients were retrieved from our institutional database between 2008 and 2019. The inclusion criteria were age ≥ 16 years, newly diagnosed histopathologically proven STS of extremities, with two available contrast-enhanced (CE) MRIs before the treatment beginning (named MRI₀ and MRI₁), and treated in a curative intent including a surgical resection in our sarcoma reference center. The exclusion criteria were lack of fat-sat (FS) turbo spin echo (TSE) CE-T1-weighted imaging (WI) in at least one of the two pretreatment MRIs, non-diagnostic image quality due to MRI artifacts, delay between pre-treatment MRIs below 3 weeks, and atypical lipomatous tumors. Figure 1 shows the study flowchart. Sixty out of the 63 finally included patients were previously analyzed in another preliminary study [21], which focused on the morphological changes using standard radiological analyses, depiction of delta-radiomics calculation, and associations with basic histologic features.

The data collection consisted of age at diagnosis, sex, World Health Organization Performance Status (WHO-PS), tumor location and depth, pre-treatment histological type, and grade (according to the "*Fédération Nationale des Centres de Lutte Contre le Cancer*" [FNCLCC] grading system) [22]. The initial management was also recorded as follows: surgical margins (R0 versus R1-R2), radiotherapy, and chemotherapy.

726 Adult patients with newly diagnosed STS managed in our sarcoma reference center between May 2008 and December 2018



Fig. 1 Study flowchart. Abbreviation: MRI, magnetic resonance imaging; STS, soft tissue sarcoma

63 Finally included patients

Radiomics pipeline

All MRI examinations were acquired on 1.5-T magnets from 9 radiological centers (with most MRI₁ in our sarcoma reference center) and included T1-WI, T2-WI, and TSE FS-CE-T1-WI. The MR systems were Optima Jem MR450W, Discovery MR450, and SIGNA Explorer (General Electric Healthcare); Achieva and Ingenia (Philips Healthcare); and Magnetom Aera and Amira (Siemens Healthineers).

The radiomics analysis was only achieved on the FS-CE-T1-WI, as this sequence had the most homogeneous acquisition scheme at the two time points over the entire cohort, and was the most informative regarding the tumor radiological architecture. The ranges of acquisition parameters were echo/repetition times: 8-20/350 - 1000 ms for the T1-WI and CE-T1-WI, and 40-120/2000 - 9000 ms for the T2-WI, respectively; in plane resolution: $0.75 \times 0.75 - 1.4 \times 1.4$ mm, and thickness: 1-7 mm. Contrast agents were gadoterate meglumine (Dotarem®, Guerbet), gadobutrol (Gadovist®, Bayer), and gadobenate dimeglunine (Multihance®, Bracco).

Since FS-CE-T1-WI were obtained on different MRI systems and given the lack of standardized units for conventional MRI, a post-processing pipeline was achieved to homogenize the MRI dataset by using the ITK library and, then, to extract 108 3D RFs from the 3D volume of each STS on MRI₀ and MRI₁ by using the Olea Sphere software (v3.0-SP16, Olea Medical) [23–25]. The full radiomics pipeline and quality control, and RF definitions are detailed in Supplementary Data S1 and S2.

Conventional radiological analysis

The morphological changes occurring between MRI₀ and MRI₁ were also evaluated using a standard radiological approach. Three radiologists (2 seniors and 1 fellow) reviewed in consensus the tumor longest diameter (LD) and the 6 following items characterizing the tumor changes in signal, compartments, and surrounding tissues: change in intra-tumoral heterogeneity on FS-CE-T1-WI (categorized as homogeneous or similar heterogeneity versus increase); change in necrotic signal (defined as high, fluid-like signal on T2-WI and no enhancement on FS-CE-T1-WI, and categorized as no necrosis or similar amount versus increase); change in hemorrhagic signal (defined as high signal on T1-WI and no enhancement on FS-CE-T1-WI, and categorized as no hemorrhage or similar amount versus increase); change in MRI growth pattern on FS-CE-T1-WI (categorized as perfectly well-delineated tumor versus < 50% of tumor circumference with ill-defined margins versus $\geq 50\%$ of tumor circumference with ill-defined margins) [26]; change in peritumoral edema on T2-WI (categorized as absent or similar versus increase) [6, 7]; and change in peritumoral enhancement on FS-CE-T1-WI (categorized as absent or similar versus increase) [6, 7]. Visual examples are shown in Supplementary Data S3.

RNA sequencing

All patients with frozen material before treatment and contemporary to MRI_1 were included in this analysis. After whole-RNA sequencing (Integragen), the produced RNA sequences were quality controlled and aligned to the transcriptome. Gene expression was then estimated by the counts of high-quality sequences aligned per gene. Finally gene expression counts were normalized using the Voom method (Supplementary Data S4) [27].

Statistical analysis

Statistical analyses were performed with R (v4.1.0). All tests were two-tailed. A p value < 0.05 was deemed significant.

Methods to calculate delta-radiomics Five mathematical methods have been proposed to calculate the change in RFs during the MRI₀-MRI₁ interval and were applied to the cohort, namely absolute change (AC), time-weighted absolute change (TWAC), relative change (RT), time-weighted relative change (TWRC), and log ratio (LR) (Supplementary Data S5) [21]. We then assessed how the delta-radiomics distributions could be affected by the scale and shape bias potentially associated to each method in order to filter the delta RFs on which the unsupervised algorithms would correctly be applied. Briefly, AC was excluded because it could scale-bias the selection of significant delta RFs by promoting high-scale features and penalizing low-scale ones. Regarding TWAC and TWRC, a bias could result from time delay variability across patients and from how time could potentially non-linearly affect each RF (i.e., plateau effect, cycle effect). RC gives equal odds to both small- and large-scale features and it respects the symmetric and Gaussian nature of the delta-radiomics distribution. Similarly, LR is not affected by scale bias and confers a belllike shape to the distribution. Consequently, RC and LR delta RFs were used in the subsequent analysis while AC, TWAC, and TWRC delta-RFs were removed.

Consensus clustering We performed two consensus clustering independently, one with RF delta-radiomics calculated with LR and one with RC. Each clustering was resampled 10,000 times by leave-one-out of 40% of the samples [28]. Both clustering provided 3 groups. To find a final agreement between the two grouping schemes, we created the confusion matrix between the labeling of each sample generated by the LR and RC clustering analyses. We defined the agreement groups as those that maximized the counts of samples placed in the diagonal of the matrix (Supplementary Data S6).

Explaining the delta-radiomics groups with the radiological features Associations between the delta-radiomics groups and the radiological features, LD on MRI₀, the relative change in LD, and the delays between the two pre-treatment MRIs were assessed using the Kruskal-Wallis test. Associations with the main histological types were assessed with the chi-2 test. Linear trends in the proportion of the radiological features across the delta-radiomics groups were investigated by using the chi-square test for trend in proportion.

Associations with the transcriptomics features Differential gene expression (DGE) and geneset enrichment analyses between the final delta-radiomics groups are detailed in Supplementary Data S7. Exploratory DGE was performed by *t*-test calculation per gene. To discriminate significant up/downregulated genes, the fold change was set to 1.5. The *p* value cut-off of 0.05 was not adjusted as this study was exploratory on a small cohort of heterogeneous histotypes. However, to confirm that the results were anyway statistically significant, we applied an alternative method named significance analysis of microarrays (SAM), which has been designed to investigate significance in the setting of small size cohorts for omics data. The principles and application are detailed in Supplementary Data S8 [29].

We then assessed geneset enrichment in biological pathways based on Broad Institute's Molecular Signature database and the CIBERSORT LM22 immuno-genesets [30, 31].

Survival analysis Metastatic and local relapse-free survivals (MFS and LFS, respectively) were defined as the time elapsed between surgical resection and histologically proven metastatic and local relapses, respectively, or death or last patient contact. Overall survival (OS) was defined as the time elapsed from surgery to death due to disease or last patient contact. The Kaplan-Meier curves for MFS, LFS, and OS depending on the delta-radiomics clusters were drawn and the differences in survivals were tested with the log-rank test. Univariate Cox regressions were performed to estimate the hazard ratio (HR) with 95% confidence interval (CI) of the delta-radiomics groups.

The overall study workflow is presented on Fig. 2.

Results

Study population

The study population finally included 63 patients with complete delta-radiomics data (median age: 63 years, range: 21–



Fig. 2 Study workflow. Abbreviation: MRI, magnetic resonance imaging; RF, radiomics feature. MRI₀ and MRI₁ were performed during the diagnostic interval before the beginning of therapeutic management

95, interquartile range [IQR]: 52.5–70, 24/63 [38.1%] women) (Table 1 and Fig. 1). The most frequent histological type was undifferentiated pleomorphic sarcoma (UPS, 29/63 [46%]).

The median delay between the two pre-treatment MRIs was 74 days (range: 24–939, IQR: 55–105). The median LDs were 98 mm (range: 12–199, IQR: 67–121) on MRI_0 and 120 mm (range: 22–270, IQR: 89–153.5) in MRI_1 , which corresponded to a median increase of 21.8% (range: 0–158.3, IQR: 9.3–44) over the inter-MRIs delay.

Defining and explaining the delta-radiomics groups

We independently performed iterative unsupervised hierarchical clustering on both relative change and on log ratio deltaradiomics as these two methods were theoretically the less prone to distribution and scaling bias (Fig. 3a). To check the consensus given by the two clusterings (Fig. 3b), an agreement table was created, which finally defined 3 consensus groups called A, B, and C (n = 16, 15, and 21, respectively), for a total of 52 patients (Fig. 3c). The 11 patients who did not **Table 1** Characteristics of thestudy population

| Characteristics | Patients | No. of NAs |
|---|-------------------------|------------|
| Age at diagnosis (years) | | |
| Mean (sd) | 60.6 (15.5) | 0 |
| Median (range) [IQR] | 63 (21–95) [52.5–70] | |
| Sex | | |
| Men | 39/63 (61.9%) | 0 |
| Women | 24/63 (38.1%) | |
| Histological type | | |
| Undifferentiated pleomorphic sarcoma | 29/63 (46%) | 0 |
| Myxofibrosarcoma | 6/63 (9.5%) | |
| Dedifferentiated LPS | 3/63 (4.8%) | |
| Myxoid/round cells LPS | 9/63 (14.3%) | |
| Pleomorphic LPS | 2/63 (3.2%) | |
| Synovial sarcoma | 5/63 (7.9%) | |
| Leiomyosarcoma | 4/63 (6.3%) | |
| Rhabdomyosarcoma | 2/63 (3.2%) | |
| Other undifferentiated sarcomas | 2/63 (3.2%) | |
| Malignant peripheral nerve sheath tumor | 1/63 (1.6%) | |
| Tumor depth | | |
| Superficial | 0/63 (0%) | 0 |
| Deep and superficial | 11/63 (17.5%) | |
| Deep | 52/63 (82.5%) | |
| Tumor location | | |
| Shoulder girdle | 4/63 (6.3%) | 0 |
| Upper limb | 9/63 (14.3%) | |
| Trunk | 5/63 (7.9%) | |
| Pelvic girdle | 1/63 (1.6%) | |
| Lower limb | 44/63 (69.8%) | |
| Histological grade [§] | | |
| Surgical grade 1 | 3/58 (5.2%) | 5 |
| Surgical grade 2 | 4/58 (6.9%) | |
| Surgical grade 3 | 5/58 (8.6%) | |
| Biopsy grade 1 | 5/58 (8.6%) | |
| Biopsy grade 2 | 25/58 (43.1%) | |
| Biopsy grade 3 | 16/58 (27.6%) | |
| Delay between MRI0 and MRI1 (days) | | |
| Mean (sd) | 110.8 (136.5) | 0 |
| Median (range) [IQR] | 74 (24–939) [55.5–105] | |
| Longest diameter on MRI0 (mm) | | |
| Mean (sd) | 98.4 (43.2) | 0 |
| Median (range) [IQR] | 98 (12–199) [67–121.5] | |
| Longest diameter on MRI1 (mm) | | |
| Mean (sd) | 123.3 (49.4) | 0 |
| Median (range) [IQR] | 120 (22–270) [89–153.5] | |
| Relative change in LD between MRIs (%) | | |
| Mean (sd) | 32.4 (34.5) | 0 |
| Median (range) [IQR] | 21.8 (0–158.3) [9.4–43] | |
| Chemotherapy | | |
| No | 21/63 (33.3%) | 0 |
| Neoadjuvant | 37/63 (58.7%) | |

Table 1 (continued)

| Characteristics | Patients | No. of NAs |
|--------------------------|---------------|------------|
| Adjuvant | 3/63 (4.8%) | |
| Neoadjuvant and adjuvant | 2/63 (3.2%) | |
| Surgical margins | | |
| R0 | 34/61 (55.7%) | 2 |
| R1-R2 | 27/61 (44.3%) | |
| Radiotherapy | | |
| No | 5/63 (7.9%) | 0 |
| Neoadjuvant | 13/63 (20.6%) | |
| Adjuvant | 45/63 (71.4%) | |

Note — Data are number of patients with percentage in parentheses, except for size, times, and age. Abbreviations: *IQR*, interquartile range; *LD*, longest diameter; *LPS*, liposarcoma; *NA*, non-available; *sd*, standard deviation

[§] Surgical grade was obtained on whole surgical specimen in patients treated with curative surgery first. Biopsy grade was obtained on the initial tumor sample in patients treated with neoadjuvant treatments but are at risk of underestimation due to sampling bias.

achieve consensus were excluded. We did not find associations between the delta-radiomics groups and LD on MRI₀, the relative change in LD, and the MRI₀-MRI₁ delay (p = 0.5912, 0.4503, 0.4256, and 0.4114, respectively), as well as associations with the main histological types (as categorized as UPS, myxoid/round cells liposarcomas, and others, p = 0.3594) and the histologic grade assessed on whole surgical specimen (categorized as grade I-II versus grade III, p = 0.2273).

Confronting the delta-radiomics groups and the conventional radiological analysis stressed the largest proportion of morphological changes in the B group, namely increases in tumor heterogeneity (10/15, 66.7%), necrotic signal (14/15, 93.3%), hemorrhagic signal (10/15, 66.7%), infiltrative margins (9/15, 60%), peritumoral edema (8/15, 53.3%), and peritumoral enhancement (9/15, 60%) (Fig. 4, Supplementary Data S9). Significant ascending linear trends were found from A group, to C group, to B group for the following proportions: increase in tumor heterogeneity (p =0.0199), necrotic signal (p = 0.0244), infiltrative margins (p =0.0032), peritumoral edema (p = 0.0019), and peritumoral enhancement (p = 0.0166).

Correlating delta-radiomics and gene expression

Twenty-one patients had available tumor tissue for RNA sequencing of patients of group A (5/21 [23.8%]), group B (8/21 [38.1%]), and group C (8/21 [38.1%]). To identify the transcriptomics features associated to tumors with aggressive outcome against the others, we performed a supervised analysis by comparing the gene expression patterns of the tumors from group B versus the pool of tumors of group A plus C. This revealed 566 differentially expressed genes and 105 enriched pathways (Fig. 5a, Supplementary Data S10). The alternative *PS*, liposarcoma; *NA*, non-available; *sd*, standard atients treated with curative surgery first. Biopsy ed with neoadjuvant treatments but are at risk of

statistical method, SAM, allowed us to confirm that those genes (except for one, MAGEA3) were statistically significant, with a q value $\leq 2.16\%$ (Supplementary Data S11). Table 2 shows the most significantly enriched genesets. Overall, B group tumors were characterized by a downregulation of natural killer (NK) cell-mediated cytotoxicity genes and an upregulation of genes involved in the glycoproteins, sphingolipids, and peptide hormone biosynthesis, the Hedgehog pathway (cell differentiation during embryogenesis), the Hippo signaling pathway (regulation of cell proliferation and apoptosis), the FAS pathway (regulation of cell death), Ephrin B reverse signaling (cell adhesion and migration) pathway, and the extracellular matrix organization pathway.

To verify whether the trends identified in the overall set of samples were conserved in the most frequent histotype, we reproduced the analysis in the UPS subgroup, which revealed 1466 differentially expressed genes. The enrichment analysis confirmed the trends found in the entire cohort with 162 enriched pathways (Fig. 5c, d, Supplementary Data S12-S13). In summary, typical oncogenesis functions like regulation of cell proliferation and apoptosis, and inhibition of the immune response were upregulated in the patients from the more aggressive B group versus the A+C pool.

Survival analysis

The median follow-up was 82.1 months. There were 24 metastatic relapses, 10 local relapses, and 16 deaths related to disease (Supplementary Data S14). No correlations were found between the delay from MRI₀ and MRI₁ and MFS, LFS, and OS in univariate Cox regression (p = 0.7680, 0.8230, and 0.3680, respectively). No significant differences were found between the MFS, LFS, and OS depending on the



Log ratio change С B' C' A' Total Relative change А 1 2 19 16 В 0 15 5 20 С 2 1 21 24 Total 18 17 28 63

Fig. 3 Definition of the radiomics groups. **a** Assessment of the distribution of the delta-radiomics features (RF) computed according of the five proposed methods. **b** Hierarchical clustering based on relative change and log ratio delta RFs identified 3 groups with good

reproducibility via 10,000 times cross-validation repeated on the patient samples. Relative change groups: A, B, and C. Log ratio groups: A', B', C'. c Agreement table between the relative change and log ratio clustering schemes

three radiomics groups (p = 0.6731, 0.0622, and 0.8970, respectively) (Table 3). However, none of the patients from the A group demonstrated local relapse while the local relapse rates from groups B and C were similar at 2 years (73.3%, 95%CI: 54–99.5, and 78.5%, 95%CI: 61.8–99.7). Thus, significant higher LFS were observed in the group A versus the B-C groups (p = 0.0277) (Fig. 6).

Discussion

Medical imaging enables longitudinal and non-invasive monitoring of the changes occurring in tumor and their surrounding tissues. Since STS are prone to long diagnostic intervals, which require updating the local staging, we proposed an original delta-radiomics-based methodology to quantify the "natural" shape and textural changes arising before any therapeutic intervention. We found three consistent patterns of natural evolution, which were radiologically and biologically explainable, and demonstrated distinct oncogenetic pathway activations.

We transposed bioinformatics tools to the radiomics field to obtain three reliable groups, according to resampling-based consensus hierarchical clustering performed on normally distributed delta RFs from two RF calculation methods. We believe this design counterbalanced the noise due to our small population. These three groups were comparable regarding



Fig. 4 Associations between the radiomics three groups and the changes in the radiological features of interest during the diagnostic interval. (a) Color-coded matrix indicating the proportion of patients in each radiomics groups (column) with the radiological features of interest (row). §: The *p* value indicated in the last column corresponds to the test assessing significant trend from group A to group B to group C. *: p <0.05; **: p < 0.005. Abbreviations: n.s., not significant; CE-T1-WI,

contrast-enhanced T1-weighted imaging. Prototypal changes observed on contrast-enhanced MRI of soft tissue sarcomas from group A (**b**), global stability of the tumor and its surrounding tissues), group C (**c**), increase in necrotic signal and heterogeneity with a global stability of the surrounding tissues), and group B (**d**), increase in necrotic signal, tumor heterogeneity, peritumoral enhancement, and worse definition of the tumor margin, i.e., increase in infiltrative growth pattern)

the main histological types, MRI_0 - MRI_1 delays, initial size, and change in LD from MRI_0 to MRI_1 .

A frequent reproach to radiomics is the lack of explicability with human vocabulary. Thus, to further validate the delta-radiomics groups, we investigated whether they could be explainable by radiologists. Indeed, radiologically, a gradient of morphological evolution was noticed from the radiomics group A to the radiomics group B in terms of changes in the intra-tumoral aspect (increase in visual heterogeneity, complexity, and necrosis) and in the tumor periphery (increase in peritumoral edema, peritumoral enhancement, and more infiltrative margins). At baseline, these radiological features have been linked to higher grade and poorer survivals [6, 7, 25, 32].

Moreover, we observed that the delta-radiomics groups (evaluating the intra-tumoral texture and not the surrounding tissues) were associated with the changes in radiological features evaluating the peritumoral tissues, highlighting the collinearity of intra- and peritumoral evolutions.

Logically, if these features appear or amplify before beginning the treatment, this could have a negative impact on the patients' outcome. Indeed, we found that no local relapse occurred in the A group compared with the two others, whereas the lowest LFS was seen in the B group. The lack of modification of the peritumoral tissue and tumor margins on the MRI of A group patients is in agreement with this better local control. Indeed, abnormal peritumoral MRI findings have been associated with satellite tumor cells spreading [33, 34].





Fig. 5 Transcriptomics analysis. Volcano plots for oncogenetic pathways in the 21 sarcomas and (a) and 8 undifferentiated pleomorphic sarcomas (UPS, b). Main differentially expressed pathways of oncogenesis in UPS from the radiomics group B versus groups A and C (c). Synthesis of the activated oncogenetic pathways from the radiomics groups A and C and from the radiomics group B (d). Abbreviations: CE-T1-WI, contrastenhanced T1-weighted imaging; MRI₀ and MRI₁, magnetic resonance imaging performed during the diagnostic interval and before the beginning of treatment. Dashed gray lines correspond to significance (*p* value < 0.05)

Although a fundamental assumption behind radiomics is the relationship between gene expression and radiophenotype, no studies had directly investigated this link in STS. Since our delta-radiomics groups reflected the natural evolutive patterns before any intervention, we hypothesized that they would also reflect the unconstrained expression of the tumor genotype. Indeed, we identified differentially expressed genes from main oncogenetic pathways associated to the delta-radiomics groups. The B group, with the most aggressive aspect and the worst local outcome, was characterized by an overexpression

 Table 2
 Transcriptomics analysis in the 21 patients with frozen tissue sample contemporary of MRI₁. Only significantly enriched pathways with ratio between the numbers of over-expressed genes in the pathway

of genes involved in the Hedgehog pathway and by a lower expression of genes belonging to the NK cell genesets. Our results are in agreement with previous studies, which demonstrated the important role of developmental pathways including Hedgehog and NK cells in the tumorigenesis and immune surveillance of sarcomas [35, 36].

Our findings could have practical applications. Regarding the poorer LFS in the B group, these patients could benefit from more aggressive locoregional treatments including larger surgical margins, higher dose of peri-operative radiotherapy, and closer local follow-up. Regarding systemic treatments, some of the identified differentially expressed genes and pathways may represent targetable therapeutic vulnerabilities. For instance, anti-angiogenic treatments or Bcl-2 inhibitors may be more effective in the B group. In the future, medical imaging may help to tailor the therapeutic strategy in STS patients, and to perform a "virtual" biopsy inferred from the tumor radiophenotype.

Our study has limitations. First, this was a retrospective single-center study, which implies heterogeneous MRI

and the numbers of DGE genes in the pathway ≤ 0.25 (green) and ≥ 0.75 (red) are shown (§)

| GSEA_go_cc NTERMEDIATE_FLAMENT_CYTOSKELETON 0.001 6 560 18 17817 10.61 (3.4.2.802) 0.53 GSEA_go_cn STRUCTURAL_CONSTITUENT_OF_CYTOSKELETON 0.0016 7 559 49 17786 4.56 (1.7.3 - 10.14) 0.85 GSEA_go_th STRUCTURAL_CONSTITUENT_OF_CYTOSKELETON 0.0016 7 559 49 17786 4.56 (1.7.3 - 10.14) 0.85 GSEA_go_th REACTOME_GLYCOPRENT, HORMONES 0.0052 3 563 9 17262 10.55 (1.83 - 42.46) 1 GSEA_go_th REACTOME_GLYCOPROTEIN_HORMONES 0.0052 3 563 9 17262 10.55 (1.83 - 42.46) 1 GSEA_go_th REACTOME_FEPTIDE_HORMONES 0.0052 3 563 11 17262 10.55 (1.83 - 42.46) 1 GSEA_apo_th REACTOME_FEPTIDE_HORMONES 0.0052 3 563 11 17264 863 (1.54 - 32.76) 1 GSEA_apo_th REACTOME_FEPTIDE_HORMONES 0.0052 3 563 11 17264 863 (1.54 - 32.46) <th>Gene list</th> <th colspan="2">Geneset</th> <th>Significant DGE in geneset</th> <th>Significant DGE not in geneset</th> <th>Not significant DGE in geneset</th> <th>Not significant DGE not in geneset</th> <th>OR (95%Cl - significant DGE vs not significant DGE)</th> <th>Ratio§</th> | Gene list | Geneset | | Significant DGE in geneset | Significant DGE not in geneset | Not significant DGE in geneset | Not significant DGE not in geneset | OR (95%Cl - significant DGE vs not significant DGE) | Ratio§ |
|---|----------------|--|--------|----------------------------------|--------------------------------------|---|---|--|--------|
| CSEA_go_cx GSEA_go_tx S | GSEA go cc | INTERMEDIATE FILAMENT | 0.0001 | 6 | 560 | 18 | 17817 | 10.61 (3.43 - 28.02) | 0.83 |
| CSEE Age_br STRUCTURAL_CONSTITUENT_OF_CYTOSKELETON 0.0016 7 759 49 17766 4.55 (172-10.14) 0.88 GSEA_go_br REACTOME_GLYCOPROTEIN_HORMONES 0.0052 3 563 9 17826 10.55 (18.3-42.46) 1 GSEA_go_br RECATIVE_REGULATION_OF_CYTOKIKE_BIOSYNTHETIC_PRO 0.0052 3 563 9 17826 10.55 (18.3-42.46) 1 GSEA_go_nf RELITAL_AMINO_ACID_TRANSMEMBARAE_TRANSPORTER_ACTIVITY 0.0052 3 563 9 17826 10.55 (18.3-42.46) 1 GSEA_nancina REACTOME_EPTOTE CONTRENT HORMONES 0.0052 3 563 9 17826 10.55 (18.3-42.46) 1 GSEA_noncina REACTOME_EPTOTE CONTRENT HORMONES 0.0082 3 563 11 17824 863 (154 - 32.76) 1 GSEA_noncina REACTOME_EPTOTE CONTRENT HORMONES 0.0082 3 563 11 17824 863 (154 - 32.76) 0.75 GSEA_noncina REACTOME_EPTOTE CONTRENT 0.0127 8 588 93 17742 | GSEA go cc | INTERMEDIATE FILAMENT CYTOSKELETON | 0.0001 | 6 | 560 | 18 | 17817 | 10.61 (3.43 - 28.02) | 0.83 |
| CSEE_Ago.bp SYSTEM_DEVELOPMENT | GSEA go mf | STRUCTURAL CONSTITUENT OF CYTOSKELETON | 0.0016 | 7 | 559 | 49 | 17786 | 4.55 (1.73 - 10.14) | 0.86 |
| CSEA_ganonical REACTOME_GLYCOPROTEIN_HORMONES 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_go_lp MEGATUR_FEDULATION_OF_CYTOKINE_BIOSYNTHETIC_PRO 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_geo_nrf NEUTRAL_AMINO_ACID_TRANSMEMBRANE_TRANSPORTER_ CSEA_canonical 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_anonical REACTOME_GLYCOPROTEIN_HORMORES 0.0052 3 563 11 17824 8.63 (154 - 32.78) 1 GSEA_anonical REACTOME_PEPTIDE_HORMORE BIOSYNTHESIS 0.0082 3 563 11 17824 8.63 (154 - 32.78) 1 GSEA_go_bp GENERATION_OF_A_SIGNAL_INVOLVED_IN_CELL_CELL_SIGN 0.0114 4 562 25 17810 5.07 (128 - 14.75) 0.78 GSEA_go_bp SKEA_tomical PID_EPHRIB_REV_PATHWAY 0.0122 4 563 15 1782 6.33 (117 - 22.46) 1.72 6.73 (117 - 5.3) 0.75 GSEA_go_bp SYMAPTOGENESIS | GSEA go bp | SYSTEM DEVELOPMENT | 0.0052 | 40 | 526 | 791 | 17044 | 1.64 (1.15 - 2.28) | 0.75 |
| GSEA_go_pp NEGATIVE_RESULATION_OF_CYTOKINE_BIOSYNTHETIC_PRO 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_go_ntf NEUTRAL_AMINO_ACID_TRANSMEMBRANE_TRANSPORTER_ ACTIVITY 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_genetic REACTOME_GLYCOPROTEIN_HORMONES 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_reactione REACTOME_FEPTIDE_HORMONE BIOSYNTHESIS 0.0082 3 563 11 17824 8.63 (1.54 - 32.76) 0.75 GSEA_go_bP SECAL_GOLD_OF_ASIGNAL, INVOLVE BIOSYNTHESIS 0.0082 3 563 11 17824 8.63 (1.54 - 32.76) 0.75 GSEA_go_bP SECAL_GOLD_OF_ASIGNAL, INVOLVE DIL_CELL_CELL_SIGN 0.0112 20 563 17 17816 561 (1.2 - 33) 0.75 GSEA_go_bP MANTOMICAL_STRUCTURE_MORPHOGENESIS 0.0113 563 15 17820 6.33 (1.7 - 27.4) 1 GSEA_go_bP SYMAPTOGENESIS 0.0174 5563 17 17818 | GSEA_canonical | REACTOME_GLYCOPROTEIN_HORMONES | 0.0052 | 3 | 563 | 9 | 17826 | 10.55 (1.83 - 42.46) | 1 |
| GSEA_go_mf NEUTRAL_AMINO_ACID_TRANSMEMBRANE_TRANSPORTER_ ACTIVITY 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_reactome GSEA_reactome REACTOME_GLYCOPROTEIN_HORMONE_BIOSVNTHESIS 0.0062 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_reactome REACTOME_PEPTIDE_HORMONE_BIOSVNTHESIS 0.0082 3 563 11 17824 8.83 (1.54 - 32.76) 1 GSEA_reactome GENERATION_OF_A_SIGNAL_INVOLVED_IN_CELL_SIGN ALING 0.0114 4 562 25 17810 5.07 (1.28 - 14.75) 0.75 GSEA_go_bp SKELETA_DEVELOPMENT 0.0127 8 558 93 17742 2.74 (1.14 - 565) 0.75 GSEA_go_bp HORMONE_SIGNALING_EV_PATHWAY 0.0129 4 562 17809 4.88 (1.51 - 32.8) 1 GSEA_go_bp HORMONE_SIGNALING_EV_HIPOGENESIS 0.0143 3 563 14 17821 6.78 (1.12 - 24.9) 1 GSEA_reactome REACTOME_SIGNALING_EV_HIPO 0.0224 3 563 17 17818 5.58 (1.05 - 13.8) 1 | GSEA_go_bp | NEGATIVE_REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PRO CESS | 0.0052 | 3 | 563 | 9 | 17826 | 10.55 (1.83 - 42.46) | 1 |
| CSEA reactome REACTOME_GLYCOPROTEIN_HOMONES 0.0052 3 563 9 17826 10.55 (1.83 - 42.46) 1 GSEA_reactome REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS 0.0082 3 563 11 17824 8.63 (1.54 - 32.78) 1 GSEA_reactome REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS 0.0082 3 563 11 17824 8.63 (1.54 - 32.78) 1 GSEA_reactomic REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS 0.0012 8 558 93 17742 2.74 (1.14 - 5.65) 0.75 GSEA_go_bp SKELETAL_DEVELOPMENT 0.0127 8 558 93 17742 2.74 (1.14 - 5.65) 0.75 GSEA_go_bp MATOMICAL_STRUCTURE_MORPHOGENESIS 0.0123 2.0546 346 17489 1.85 (1.71 - 23.49) 1 GSEA_reactome REACTOME_SIGNALING BY HIPPO 0.0224 3 563 17 17818 558 (1.05 - 19.38) 1 GSEA_reactome REACTOME_SIGNALING BY HIPPO 0.0224 3 563 17 17818 558 (1.05 - 19.38) </td <td>GSEA_go_mf</td> <td>NEUTRAL_AMINO_ACID_TRANSMEMBRANE_TRANSPORTER_ ACTIVITY</td> <td>0.0052</td> <td>3</td> <td>563</td> <td>9</td> <td>17826</td> <td>10.55 (1.83 - 42.46)</td> <td>1</td> | GSEA_go_mf | NEUTRAL_AMINO_ACID_TRANSMEMBRANE_TRANSPORTER_ ACTIVITY | 0.0052 | 3 | 563 | 9 | 17826 | 10.55 (1.83 - 42.46) | 1 |
| CSEA_canonical REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS 0.0082 3 563 11 17824 8.83 (1.54 - 32.78) 1 GSEA_caconical REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS 0.0082 3 563 11 17824 8.83 (1.54 - 32.78) 1 GSEA_go_bp GENERATION_OF_A_SIGNAL_INVOLVED_IN_CELL_CELL_SIGN 0.0114 4 562 25 17810 5.07 (1.28 - 14.75) 0.75 GSEA_go_bp SKELETAL DEVELOPMENT 0.0127 8 568 93 17742 2.74 (1.14 - 5.65) 0.75 GSEA_go_bp ANATOMICAL_STRUCTURE_MORPHOGENESIS 0.0132 20 546 346 17489 1.85 (1.11 - 2.33) 0.75 GSEA_go_bp YNAPTOGENESIS 0.0167 3 563 15 17820 6.33 (1.17 - 22.46) 1 GSEA_acanonical REACTOME_SIGNALING BY, HIPPO 0.0224 3 563 17 17818 5.58 (106 - 19.38) 1 GSEA_canonical REACTOME_SIGNALING BY, HIPPO 0.0224 561 51 17784 3.11 (0.66 - 7.8) | GSEA_reactome | REACTOME_GLYCOPROTEIN_HORMONES | 0.0052 | 3 | 563 | 9 | 17826 | 10.55 (1.83 - 42.46) | 1 |
| CSEA_preactome REACTOME_PEPTIDE_HORMONLE_BIOSYNTHESIS 0.0082 3 553 11 17824 8.63 (154-32.78) 1 GSEA_go_bp ALING GENERATION_OF_A_SIGNAL_INVOLVED_IN_CELL_CELL_SIGN 0.0114 4 562 25 17810 5.07 (1.28-14.75) 0.757 GSEA_go_bp SKELETAL_DEVELOPMENT 0.0127 8 558 93 17742 2.74 (1.14-5.65) 0.757 GSEA_go_bp ANATOMICAL_STRUCTURE_MORPHOGENESIS 0.0129 4 552 26 17809 4.88 (1.23-14.11) 0.750 GSEA_go_bp MANTOMICAL_STRUCTURE_MORPHOGENESIS 0.0143 553 15 17820 6.33 (1.17-22.46) 1 GSEA_go_bp SYNAPTOGENESIS 0.0174 8 558 99 17736 2.57 (1.07-5.3) 0.75 GSEA_reactome REACTOME_SIGNALING_P_HIPPO 0.0224 3 563 17 17818 556 (1.05-19.38) 1 GSEA_canonical PIG_FAS_ALING_BY_HIPPO 0.0224 3 563 17 17818 556 (1.05-19.38) | GSEA_canonical | REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS | 0.0082 | 3 | 563 | 11 | 17824 | 8.63 (1.54 - 32.78) | 1 |
| GSEA_go_bp GENERATION_OF_A_SIGNAL_INVOLVED_IN_CELL_CELL_SIGN ALING 0.0114 4 562 25 17810 5.07 (1.28 - 14.75) 0.75 GSEA_go_bp SKELETAL_DEVELOPMENT 0.0127 8 558 93 17742 2.74 (1.14 - 565) 0.75 GSEA_go_bp ANATOMICAL_STRUCTURE_MORPHOGENESIS 0.0132 20 546 346 17489 1.85 (1.17 - 22.46) 1 GSEA_go_bp HORMONE_SECRETION 0.0167 3 563 17 17818 5.68 (1.05 - 19.39) 1 GSEA_go_cc CELL_PROJECTION 0.0167 3 563 17 17818 5.56 (1.05 - 19.39) 1 GSEA_reactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.56 (1.05 - 19.39) 1 GSEA_reactome REACTOME_SIGNALING BY_HIPPO 0.0224 3 563 17 17818 5.56 (1.05 - 19.39) 1 GSEA_canonical KEGG_SPHINOCULPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) <td< td=""><td>GSEA_reactome</td><td>REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS</td><td>0.0082</td><td>3</td><td>563</td><td>11</td><td>17824</td><td>8.63 (1.54 - 32.78)</td><td>1</td></td<> | GSEA_reactome | REACTOME_PEPTIDE_HORMONE_BIOSYNTHESIS | 0.0082 | 3 | 563 | 11 | 17824 | 8.63 (1.54 - 32.78) | 1 |
| GSEA go.pb SKELETAL DEVELOPMENT 0.0127 8 558 93 17742 2.74 (114 - 565) 0.75 GSEA, go.pb MATOMICAL STRUCTURE_MORPHOGENESIS 0.0122 2 546 346 17489 1.88 (12.3 - 14.1) 0.75 GSEA, go.pb HORMONE SECRETION 0.0132 20 546 346 17489 1.85 (1.1 - 2.2.3) 0.75 GSEA, go.pb SYNAPTOGENESIS 0.0167 3 563 14 17821 6.78 (1.25 - 4.3.9) 1 GSEA, go.pc CGL_PROLECTION 0.0167 3 563 17 17818 558 (1.05 - 19.38) 1 GSEA, reactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 558 (1.05 - 19.38) 1 GSEA, anancial HALLMARK, HEDGEHOG_SIGNALING 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA, canonical KEGG_SPHINSOLIPIO_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 | GSEA_go_bp | GENERATION_OF_A_SIGNAL_INVOLVED_IN_CELL_CELL_SIGN ALING | 0.0114 | 4 | 562 | 25 | 17810 | 5.07 (1.28 - 14.75) | 0.75 |
| GSEA genonical PID_EPHRINE_REV_PATHWAY 0.0129 4 562 26 17809 4.88 (1.23 · 14.11) 0.75 GSEA go bp HORMORE_SECRETION 0.0143 3 563 14 17821 6.78 (1.25 - 24.39) 1 GSEA go bp SYNAPTOGENESIS 0.0167 3 563 15 17820 6.33 (1.17 - 22.46) 1 GSEA go cc CELL_PROJECTION 0.0174 8 563 17 17818 558 (1.05 - 19.38) 1 GSEA canonical REACTOME_SIGNALING BY_HIPPO 0.0224 3 563 17 17818 558 (1.05 - 19.38) 1 GSEA canonical REACTOME_SIGNALING BY_HIPPO 0.0224 3 563 17 17818 558 (1.05 - 19.38) 1 GSEA canonical KEGG SPHINGOLIPID_METABOLISM 0.0285 5 561 51 17781 373 (0.96 - 10.5) 0.75 GSEA canonical NBA CORE_NATRINOME 0.0226 3 632 17801 3.73 (0.96 - 10.5) 0.75 GSEA canonical | GSEA_go_bp | SKELETAL_DEVELOPMENT | 0.0127 | 8 | 558 | 93 | 17742 | 2.74 (1.14 - 5.65) | 0.75 |
| GSEA_go_bp ANATOMICAL_STRUCTURE_MORPHOGENESIS 0.0132 20 546 346 17499 1.85 (1,11 - 2.93) 0.75 GSEA_go_bp FVNAPTOGENESIS 0.0143 3 563 14 17821 6.78 (1.02 - 2.4.39) 1 GSEA_go_bp SYNAPTOGENESIS 0.0167 3 563 15 17820 6.33 (1.17 - 22.46) 1 GSEA_go_cc CELL_PROJECTION 0.0174 8 563 17 17818 558 (1.05 - 19.38) 1 GSEA_genomical REACTOME_SIGNALING BY_HIPPO 0.0224 3 563 17 17818 558 (1.05 - 19.38) 1 GSEA_genomica NEACTOME_SIGNALING 0.0244 4 562 32 17801 3.73 (0.96 - 10.5) 0.75 GSEA_genomica HOL, FAS PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_bp REGULATION_OP_CYTOKINE BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 <td< td=""><td>GSEA_canonical</td><td>PID_EPHRINB_REV_PATHWAY</td><td>0.0129</td><td>4</td><td>562</td><td>26</td><td>17809</td><td>4.88 (1.23 - 14.11)</td><td>0.75</td></td<> | GSEA_canonical | PID_EPHRINB_REV_PATHWAY | 0.0129 | 4 | 562 | 26 | 17809 | 4.88 (1.23 - 14.11) | 0.75 |
| GSEA_op.bp HORMONE_SECRETION 0.0143 3 563 14 17821 6.78 (125-24.39) 1 GSEA_op.cc CELL_PROJECTION 0.0167 3 563 15 17820 6.33 (1.17-22.64) 1 GSEA_gencc CELL_PROJECTION 0.0174 8 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_reactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_canonical KEGG_SPHINGOLIPID_METABOLISM 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_canonical KEGG_SPHINGOLIPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NBA_CORE_NATIRBOME 0.0325 3 563 20 17818 356 (1.01 - 11.22) 0.75 GSEA_gen_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0325 3 563 20 17810 3.73 (0.96 - 10.5) 0.75 | GSEA_go_bp | ANATOMICAL_STRUCTURE_MORPHOGENESIS | 0.0132 | 20 | 546 | 346 | 17489 | 1.85 (1.11 - 2.93) | 0.75 |
| CSEA_go_bp SYNAPTOGENESIS 0.0174 3 563 15 17820 6.33 (1.7 - 22.46) 1 GSEA_go_cc CELL_PROJECTION 0.0174 8 558 99 177.36 2.57 (1.07 - 5.3) 0.75 GSEA_areactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_neactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_anonical MARK_HEDGENGO_SIGNALING 0.0244 4 562 34 17801 3.73 (0.96 - 1.05) 0.75 GSEA_anonical KCells activated 0.0286 4 562 34 17801 3.73 (0.96 - 1.05) 0.75 GSEA_anonical NABA_CORE_MATRISOME 0.0286 4 562 34 17801 3.73 (0.96 - 1.05) 0.75 GSEA_anonical NABA_CORE_MARKATISOME 0.0312 15 515 258 17577 1.85 (1.02 - 3.14) 0.80 GSEA_ag | GSEA go bp | HORMONE SECRETION | 0.0143 | 3 | 563 | 14 | 17821 | 6.78 (1.25 - 24.39) | 1 |
| GSEA_go_cc CELL_PROJECTION 0.017 8 558 99 17736 2.57 (1.07 - 5.3) 0.75 GSEA_canonical REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_reactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_reactome REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_reanonical KEGG_SPHINGOLIPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical PID_FAS_PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NABA_CORE_MATRISOME 0.0312 15 551 258 17577 1.85 (102 - 1.80) 0.75 GSEA_go_bp NGGAN_ZERGANIZATION_AND_BIOGENESIS 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 | GSEA go bp | SYNAPTOGENESIS | 0.0167 | 3 | 563 | 15 | 17820 | 6.33 (1.17 - 22.46) | 1 |
| GSEA_reaction REACTOME_SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_nellmark HALLMARK_HEDGEHOG_SIGNALING 0.024 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_nellmark HALLMARK_HEDGEHOG_SIGNALING 0.024 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_nellmark HALLMARK_HEDGEHOG_SIGNALING 0.024 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_anonical KEGUS SPHINGOLIPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical PID_FAS_PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NRGOLATINO,OF_CYTOKINE_BIOSYNTHETIC_PROCESS 0.0312 15 551 258 17577 185 (102 - 10.07) 1 GSEA_go_bp SYNAPSE_ORGANZATION_AND BIOGENESIS 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) <td>GSEA go cc</td> <td>CELL PROJECTION</td> <td>0.0174</td> <td>8</td> <td>558</td> <td>99</td> <td>17736</td> <td>2.57 (1.07 - 5.3)</td> <td>0.75</td> | GSEA go cc | CELL PROJECTION | 0.0174 | 8 | 558 | 99 | 17736 | 2.57 (1.07 - 5.3) | 0.75 |
| GSEA_freactome REACTOME SIGNALING_BY_HIPPO 0.0224 3 563 17 17818 5.58 (1.05 - 19.38) 1 GSEA_hallmak HALLMARK_HEDGEHOG_SIGNALING 0.0224 4 562 32 17803 3.96 (1.01 - 11.22) 0.75 GSEA_canonical KEGG_SPHINGOLIPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical PID_FAS_PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_bp REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_bp REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_bp NEGATWE_REGULATION_O_TRANSPORT_AND_A 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGANIZATION_AND_BIOCENESIS 0.0325 3 563 20 17815 | GSEA canonical | REACTOME SIGNALING BY HIPPO | 0.0224 | 3 | 563 | 17 | 17818 | 5.58 (1.05 - 19.38) | 1 |
| GSEA_inimumark HALLMARK_HEDGEHOG_SIGNALING 0.024 4 562 32 17803 3.96 (1.01 - 11.22) 0.75 LM22_immuno NK_oells_activated 0.0285 5 561 51 17784 3.11 (0.96 - 7.78) 0 GSEA_canonical KEGG_SPHINGOLIPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NAB_CORE_MATRISOME 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NABA_CORE_MATRISOME 0.0312 15 551 258 17577 1.85 (1.02 - 3.14) 0.8 GSEA_go_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0346 26 540 535 17300 1.56 (1 - 2.33) 0.77 GSEA_go_bp ORGAN_DEVELOPMENT 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 <t< td=""><td>GSEA reactome</td><td>REACTOME SIGNALING BY HIPPO</td><td>0.0224</td><td>3</td><td>563</td><td>17</td><td>17818</td><td>5.58 (1.05 - 19.38)</td><td>1</td></t<> | GSEA reactome | REACTOME SIGNALING BY HIPPO | 0.0224 | 3 | 563 | 17 | 17818 | 5.58 (1.05 - 19.38) | 1 |
| LM22_immuno NK_cells_activated 0.0285 5 561 51 17784 3.11 (0.96 - 7.78) 0 GSEA_canonical PID_FAS_PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical PID_FAS_PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical PID_FAS_PATHWAY 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NABA_CORE_MATRISOME 0.0312 15 551 258 17577 1.85 (1.02 - 3.14) 0.8 GSEA_go_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_reactome MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_b | GSEA hallmark | HALLMARK HEDGEHOG SIGNALING | 0.024 | 4 | 562 | 32 | 17803 | 3.96 (1.01 - 11.22) | 0.75 |
| GSEA_canonical GSEA_canonical SEA_canonical PID_FAS_PATHWAY KEG_G_SPHINGOLIPID_METABOLISM 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_gan_pb GSEA_ga_bp REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_ga_bp REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_ga_bp NABA_CORE_MATRISOME 0.0312 15 551 258 17577 1.85 (1.02 - 3.14) 0.8 GSEA_go_bp SYNAPSE_ORGANIZATION_AND_BIOGENESIS 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0326 2 564 8 17827 7.9 (0.82 - 39.7) 0 GSEA_gao_mf ANTIPORTER_ACTIVITY REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.7) 0 GSEA_go_ph CYTOKINE_BIOSYNTHETIC_PROCESS <td>LM22 immuno</td> <td>NK cells activated</td> <td>0.0285</td> <td>5</td> <td>561</td> <td>51</td> <td>17784</td> <td>3.11 (0.96 - 7.78)</td> <td>0</td> | LM22 immuno | NK cells activated | 0.0285 | 5 | 561 | 51 | 17784 | 3.11 (0.96 - 7.78) | 0 |
| GSEA_canonical PID_FAS_PATHWAY O 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_pb REGULATION_OF_CTYCKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_pb REGULATION_OF_TCYCKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_go_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0312 15 551 258 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORAN_DEVELOPMENT 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORAN_DEVELOPMENT 0.0346 26 540 535 17300 1.56 (1 - 2.3) 0.77 GSEA_reactome REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0361 2 564 8 | GSEA canonical | KEGG SPHINGOLIPID METABOLISM | 0.0286 | 4 | 562 | 34 | 17801 | 3.73 (0.96 - 10.5) | 0.75 |
| GSEA_go_bp REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PROCESS 0.0286 4 562 34 17801 3.73 (0.96 - 10.5) 0.75 GSEA_canonical NABA_CORE_MATRISOME 0.0312 15 551 258 17777 1.85 (1.02 - 3.14) 0.8 GSEA_go_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0346 26 540 555 17300 1.56 (1 - 2.33) 0.77 GSEA_canonical REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A 0.0346 26 540 555 17300 1.56 (1 - 2.33) 0.77 GSEA_canonical REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 2 564 8 17827 7. | GSEA canonical | PID FAS PATHWAY | 0.0286 | 4 | 562 | 34 | 17801 | 3.73 (0.96 - 10.5) | 0.75 |
| GSEA_canonical NABA_CORE_MATRISOME 0.0312 15 551 258 17577 1.85 (1.02 - 3.14) 0.8 GSEA_go_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_canonical REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A 0.0325 3 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 | GSEA ao bp | REGULATION OF CYTOKINE BIOSYNTHETIC PROCESS | 0.0286 | 4 | 562 | 34 | 17801 | 3.73 (0.96 - 10.5) | 0.75 |
| GSEA_go_bp NEGATIVE_REGULATION_OF_TRANSLATION 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp SYNAPSE_ORGANIZATION_AND_BIOGENESIS 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0346 26 540 535 17300 1.56 (1 - 2.33) 0.77 GSEA_canonical REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_reactome REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 37 17798 3.42 (0.88 - 6.58) 0.76 GSEA_go_bp CYTOKINE_METABOLIC_PROCESS 0.0395 4 562 < | GSEA canonical | NABA CORE MATRISOME | 0.0312 | 15 | 551 | 258 | 17577 | 1.85 (1.02 - 3.14) | 0.8 |
| GSEA_go_bp SYNAPSE_ORGANIZATION_AND_BIOGENESIS 0.0325 3 563 20 17815 4.75 (0.9 - 16.07) 1 GSEA_go_bp ORGAN_DEVELOPMENT 0.0346 26 540 535 17300 1.56 (1 - 2.33) 0.77 GSEA_canonical REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_reactome MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_reactome MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 37 17798 3.42 (0.88 - 9.58) 0.75 GSEA_go_bp CYTOKINE_METAGOLIC_PROCESS 0.0395 4 562 38 17977 < | GSEA go bp | NEGATIVE REGULATION OF TRANSLATION | 0.0325 | 3 | 563 | 20 | 17815 | 4.75 (0.9 - 16.07) | 1 |
| GSEA_go_bp ORGAN_DEVELOPMENT 0.0346 26 540 535 17300 1.56 (1 - 2.33) 0.77 GSEA_go_bp REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROACTIVITY 0.0432 2 564 9 17826 7.02 | GSEA go bp | SYNAPSE ORGANIZATION AND BIOGENESIS | 0.0325 | 3 | 563 | 20 | 17815 | 4.75 (0.9 - 16.07) | 1 |
| GSEA_canonical REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_reactome REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mb CYTOKINE_BIOSVNTHETIC_PROCESS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSVNTHETIC_PROCESS 0.03661 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSVNTHETIC_PROCESS 0.03661 2 564 8 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp CYTOKINE_BIOSONTHETIC_PROCESS 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_th SECETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 | GSEA_go_bp | ORGAN_DEVELOPMENT | 0.0346 | 26 | 540 | 535 | 17300 | 1.56 (1 - 2.33) | 0.77 |
| GSEA_reactome REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 0 GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 37 17798 3.42 (0.88 - 9.58) 0.75 GSEA_go_bp CYTOKINE_METABOLIC_PROCESS 0.0366 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp CYTOKINE_ENTABOLIC_PROCESS 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 | GSEA_canonical | REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS | 0.0361 | 2 | 564 | 8 | 17827 | 7.9 (0.82 - 39.71) | 0 |
| GSEA_go_mf ANTIPORTER_ACTIVITY 0.0361 2 564 8 17827 7.9 (0.82 - 39.71) 1 GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 37 17798 3.42 (0.82 - 9.58) 0.75 GSEA_go_bp CYTOKINE_METABOLIC_PROCESS 0.0395 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp CYTOKINE_METABOLIC_PROCESS 0.0395 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp ACUTE_INFLAMMATORY_RESPONSE 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17756 2.41 (0.85 - 5.51) 1 GSEA_go_mf REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp C_PROCESS 0.0489 4 562 41 1779 3.09 (0.8 - 8.57) 1 | GSEA_reactome | REACTOME_GAMMA_CARBOXYLATION_TRANSPORT_AND_A MINO_TERMINAL_CLEAVAGE_OF_PROTEINS | 0.0361 | 2 | 564 | 8 | 17827 | 7.9 (0.82 - 39.71) | 0 |
| GSEA_go_bp CYTOKINE_BIOSYNTHETIC_PROCESS 0.0366 4 562 37 17798 3.42 (0.88 - 9.58) 0.75 GSEA_go_bp CYTOKINE_METABOLIC_PROCESS 0.0395 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp ACUTE_INFLAMMATORY_RESPONSE 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_bm REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0449 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI 0.0489 4 562 41 17794 | GSEA_go_mf | ANTIPORTER_ACTIVITY | 0.0361 | 2 | 564 | 8 | 17827 | 7.9 (0.82 - 39.71) | 1 |
| GSEA_go_bp CYTOKINE_METABOLIC_PROCESS 0.0395 4 562 38 17797 3.33 (0.86 - 9.3) 0.75 GSEA_go_bp ACUTE_INFLAMMATORY_RESPONSE 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_hf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_canonical REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp REACTOME_CATRACELLULAR_PROTEIN_METABOLI 0.0489 4 562 41 17794 | GSEA_go_bp | CYTOKINE_BIOSYNTHETIC_PROCESS | 0.0366 | 4 | 562 | 37 | 17798 | 3.42 (0.88 - 9.58) | 0.75 |
| GSEA_go_bp ACUTE_INFLAMMATORY_RESPONSE 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_canonical GSEA_reactome REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI C_PROCESS 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SULFOTRANSFERASE_ACTIVITY 0.049 3 563 24 17811 3.95 (0.76 - 13.08) 1 | GSEA go bp | CYTOKINE METABOLIC PROCESS | 0.0395 | 4 | 562 | 38 | 17797 | 3.33 (0.86 - 9.3) | 0.75 |
| GSEA_go_mf SECRETIN_LIKE_RECEPTOR_ACTIVITY 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_gonical GSEA_canonical GSEA_reactione REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0432 2 564 9 17826 7.02 (0.74 - 34.06) 1 GSEA_reactione REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI C_PROCESS 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SECONDARY_ACTIVE_TRANSMEMBRANE_TRANSPORTER_AC TVITY 0.049 3 563 24 17811 3.96 (0.76 - 13.08) 1 | GSEA go bp | ACUTE INFLAMMATORY RESPONSE | 0.0432 | 2 | 564 | 9 | 17826 | 7.02 (0.74 - 34.06) | 1 |
| GSEA_canonical GSEA_reactome REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_reactome REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_reactome NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI C_PROCESS 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SECONDARY_ACTIVE_TRANSMEMBRANE_TRANSPORTER_AC TIVITY 0.049 3 563 24 17811 3.99 (0.8 - 8.57) 1 | GSEA go mf | SECRETIN LIKE RECEPTOR ACTIVITY | 0.0432 | 2 | 564 | 9 | 17826 | 7.02 (0.74 - 34.06) | 1 |
| GSEA_reactome REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION 0.0469 6 560 79 17756 2.41 (0.85 - 5.51) 1 GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI C_PROCESS 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SECONDARY_ACTIVE_TRANSMEMBRANE_TRANSPORTER_AC TIVITY 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SULFOTRANSFERASE_ACTIVITY 0.049 3 563 24 17811 3.95 (0.76 - 13.08) 1 | GSEA canonical | REACTOME EXTRACELLULAR MATRIX ORGANIZATION | 0.0469 | 6 | 560 | 79 | 17756 | 2.41 (0.85 - 5.51) | 1 |
| GSEA_go_bp NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI C_PROCESS 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SECONDARY_ACTIVE_TRANSMEMBRANE_TRANSPORTER_AC TIVITY 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SULFOTRANSFERASE_ACTIVITY 0.049 3 563 24 17811 3.95 (0.76 - 13.08) 1 | GSEA_reactome | REACTOME_EXTRACELLULAR_MATRIX_ORGANIZATION | 0.0469 | 6 | 560 | 79 | 17756 | 2.41 (0.85 - 5.51) | 1 |
| GSEA_go_mf SECONDARY_ACTIVE_TRANSMEMBRANE_TRANSPORTER_AC TIVITY 0.0489 4 562 41 17794 3.09 (0.8 - 8.57) 1 GSEA_go_mf SULFOTRANSFERASE_ACTIVITY 0.049 3 563 24 17811 3.95 (0.76 - 13.08) 1 | GSEA_go_bp | NEGATIVE_REGULATION_OF_CELLULAR_PROTEIN_METABOLI C_PROCESS | 0.0489 | 4 | 562 | 41 | 17794 | 3.09 (0.8 - 8.57) | 1 |
| GSEA_go_mf SULFOTRANSFERASE_ACTIVITY 0.049 3 563 24 17811 3.95 (0.76 - 13.08) | GSEA_go_mf | SECONDARY_ACTIVE_TRANSMEMBRANE_TRANSPORTER_ACTIVITY | 0.0489 | 4 | 562 | 41 | 17794 | 3.09 (0.8 - 8.57) | 1 |
| | GSEA_go_mf | SULFOTRANSFERASE_ACTIVITY | 0.049 | 3 | 563 | 24 | 17811 | 3.95 (0.76 - 13.08) | 1 |

| Outcome | Radiomics groups | No. at risk | No. of events | 2-year survival probability (95%CI) | log-rank p value | HR (95%CI) | p value |
|---------|---------------------|-------------|---------------|--|------------------|------------------|-----------|
| MFS | A | 16 | 6 | 75 (56.5–99.5) | 0.6731 | Reference | Reference |
| | В | 15 | 7 | 73.3 (54–99.5) | | 1.29 (0.43–3.85) | 0.650 |
| | С | 20 | 11 | 68.1 (49.8–93) | | 1.56 (0.58-4.23) | 0.379 |
| LFS | А | 16 | 0 | 1 (1–1) | 0.0622 | Reference | Reference |
| | В | 15 | 5 | 73.3 (54–99.5) | | +∞ () | |
| | С | 20 | 5 | 78.5 (61.8–99.7) | | +∞ () | |
| | Α | 16 | 0 | 1 (1–1) | 0.0277* | Reference | Reference |
| | B + C | 35 | 10 | 76.5 (63.5–92.2) | | +∞ () | |
| | A + C | 36 | 5 | 87.5 (76.8–99.8) | 0.0923 | Reference | Reference |
| | В | 15 | 5 | 73.3 (54–99.5) | | 2.79 (0.80-9.68) | 0.107 |
| | A + B | 31 | 5 | 86.9 (75.6–99.8) | 0.5543 | Reference | Reference |
| | С | 20 | 5 | 78.5 (61.8–99.7) | | 1.45 (0.42–5.02) | 0.557 |
| OS | А | 16 | 5 | 80.2 (62.4–1) | 0.897 | Reference | Reference |
| | В | 15 | 5 | 93.3 (81.5–1) | | 1.24 (0.35–4.33) | 0.739 |
| | С | 20 | 6 | 94.7 (85.2–1) | | 0.94 (0.29–3.10) | 0.922 |

 Table 3
 Correlations between patients' outcomes and radiomics groups

Note — Abbreviations: *CI*, confidence interval; *HR*, hazard ratio; *LFS*, local relapse-free survival; *MFS*, metastatic relapse-free survival; *No.*, number; *OS*, overall survival

One patient out of the 52 patients was removed from the survival analysis because of insufficient follow-up

*p < 0.05. Significant results are highlighted in bold

protocols and lack of frozen specimens for RNA-sequencing in a third of patients. Hence, our delta-radiomics analysis was achieved on FS-CE-T1-WI only. Second, dynamic contrast enhancement, spectroscopy and diffusion imaging, and positron emission tomography could have improved the assessment of tumor angiogenesis, cellularity, and metabolism [35, 37] but they are never performed outside reference centers. Third, the small population led to a lack of statistical power for transcriptomics analyses, which prompted us not to adjust the p values with standard method in order to detect biologically meaningful trends; however, our results were confirmed using an alternative method named SAM which is dedicated



Fig. 6 Kaplan-Meier curves for (a) metastatic relapse-free survival (MFS), (b) local relapse-free survival (LFS), and (c) overall survival (OS) depending on the three radiomics groups, A, B, and C

to very imbalanced datasets in terms of number of variables and number of samples [29]. Fourth, there were important variations in the delay between the two pre-treatment MRIs that could have influenced our findings though no association was found between this delay and patients' outcome. Fifth, in order to limit the exclusion of several RFs before the calculation and selection step of delta RFs, we choose to apply a long post-processing pipeline in order to obtain comparable images between patients and between two evaluations per patient and, then, to make the VOIs adjusted and validated by the most experienced radiologists from our sarcoma reference center, this instead of filtering RFs at a single time point depending on their reproducibility over various voxel sizes, numbers of gray levels, or experience of the segmenting radiologist. Sixth, although the final agreement table between the consensus clusterings on LR and RC features was an additional security to avoid tumor misclassification, it led to the additional exclusion of 11 patients (17%) without agreement.

In conclusion, this exploratory study provides new research leads regarding the radiogenomics of STS. We have highlighted original correlations between meaningful delta-radiomicsbased clusters quantifying the natural tumor changes before any treatment, and important oncogenetic pathways and local control of the disease after curative treatment, bridging the gap between quantitative imaging and cancer biology.

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Declarations

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Conflict of interest The authors of this manuscript declare no relationships with any companies whose products or services may be related to the subject matter of the article.

Statistics and biometry Three of the authors have significant statistical expertise (C.L. and F.B. are bioinformaticians; A.C. has a PhD in applied mathematics).

Informed consent Written informed consent was waived by the Institutional Review Board.

Ethical approval Institutional Review Board approval was obtained.

Study subjects or cohorts overlap Some study subjects or cohorts have been previously reported in the study by Fadli et al [20].

Methodology

- retrospective
- · diagnostic or prognostic study/observational/experimental
- · performed at one institution

References

- Fletcher CDMF, Bridge JA, Hogendoorn PCW, Martens F (2020) WHO classification of soft tissue and bone tumours, 5th edn. IARC Publications, Lyon, France
- Soomers V, Husson O, Young R et al (2020) The sarcoma diagnostic interval: a systematic review on length, contributing factors and patient outcomes. ESMO Open 5:e000592
- Gronchi A, Miah AB, Dei Tos AP et al (2021) Soft tissue and visceral sarcomas: ESMO-EURACAN-GENTURIS Clinical Practice Guidelines for diagnosis, treatment and follow-up¹/₂. Ann Oncol S0923-7534(21):02184–0
- 4. Callegaro D, Miceli R, Bonvalot S et al (2016) Development and external validation of two nomograms to predict overall survival and occurrence of distant metastases in adults after surgical resection of localised soft-tissue sarcomas of the extremities: a retrospective analysis. Lancet Oncol 17:671–680
- Coindre JM, Terrier P, Bui NB et al (1996) Prognostic factors in adult patients with locally controlled soft tissue sarcoma. A study of 546 patients from the French Federation of Cancer Centers Sarcoma Group. J Clin Oncol 14:869–877
- Zhao F, Ahlawat S, Farahani SJ et al (2014) Can MR imaging be used to predict tumor grade in soft-tissue sarcoma? Radiology 272: 192–201
- Crombé A, Marcellin P-J, Buy X et al (2019) Soft-tissue sarcomas: assessment of MRI features correlating with histologic grade and patient outcome. Radiology 291:710–721
- Peeken JC, Neumann J, Asadpour R et al (2021) Prognostic assessment in high-grade soft-tissue sarcoma patients: a comparison of semantic image analysis and radiomics. Cancers (Basel) 13:1929
- 9. Limkin EJ, Sun R, Dercle L et al (2017) Promises and challenges for the implementation of computational medical imaging (radiomics) in oncology. Ann Oncol 28:1191–1206
- Lambin P, Leijenaar RTH, Deist TM, et al (2017) Radiomics: the bridge between medical imaging and personalized medicine. Nat Rev Clin Oncol. 2017;14:749–762
- Corino VDA, Montin E, Messina A et al (2018) Radiomic analysis of soft tissues sarcomas can distinguish intermediate from highgrade lesions. J Magn Reson Imaging 47:829–840
- Peeken JC, Spraker MB, Knebel C et al (2019) Tumor grading of soft tissue sarcomas using MRI-based radiomics. EBioMedicine 48:332–340
- Yan R, Hao D, Li J et al (2021) Magnetic resonance imaging-based radiomics nomogram for prediction of the histopathological grade of soft tissue sarcomas: a two-center study. J Magn Reson Imaging 53:1683–1696
- Wang H, Chen H, Duan S et al (2020) Radiomics and machine learning with multiparametric preoperative MRI may accurately predict the histopathological grades of soft tissue sarcomas. J Magn Reson Imaging 51:791–797
- Crombé A, Fadli D, Buy X et al (2020) High-grade soft-tissue sarcomas: can optimizing dynamic contrast-enhanced MRI postprocessing improve prognostic radiomics models? J Magn Reson Imaging 52:282–297
- Crombé A, Le Loarer F, Sitbon M et al (2020) Can radiomics improve the prediction of metastatic relapse of myxoid/round cell liposarcomas? Eur Radiol 30:2413–2424

- 17. Spraker MB, Wootton LS, Hippe DS et al (2019) MRI radiomic features are independently associated with overall survival in soft tissue sarcoma. Adv Radiat Oncol 4:413–421
- Crombé A, Périer C, Kind M et al (2019) T2 -based MRI deltaradiomics improve response prediction in soft-tissue sarcomas treated by neoadjuvant chemotherapy. J Magn Reson Imaging 50:497– 510
- Peeken JC, Asadpour R, Specht K et al (2021) MRI-based deltaradiomics predicts pathologic complete response in high-grade softtissue sarcoma patients treated with neoadjuvant therapy. Radiother Oncol 164:73–82
- Chen S, Li N, Tang Y et al (2021) Radiomics analysis of fatsaturated T2-weighted MRI sequences for the prediction of prognosis in soft tissue sarcoma of the extremities and trunk treated with neoadjuvant radiotherapy. Front Oncol 11:710649
- Fadli D, Kind M, Michot A et al (2021) Natural changes in radiological and radiomics features on MRIs of soft-tissue sarcomas naïve of treatment: correlations with histology and patients' outcomes. J Magn Reson Imaging. https://doi.org/10.1002/jmri.28021
- 22. Trojani M, Contesso G, Coindre JM et al (1984) Soft-tissue sarcomas of adults; study of pathological prognostic variables and definition of a histopathological grading system. Int J Cancer 33:37–42
- McCormick M, Liu X, Jomier J et al (2014) ITK: enabling reproducible research and open science. Front Neuroinform 8:13
- 24. Tustison NJ, Avants BB, Cook PA et al (2010) N4ITK: improved N3 bias correction. IEEE Trans Med Imaging 29:1310–1320
- Nyúl LG, Udupa JK (1999) On standardizing the MR image intensity scale. Magn Reson Med 42:1072–1081
- 26. Nakamura T, Matsumine A, Matsubara T et al (2017) Infiltrative tumor growth patterns on magnetic resonance imaging associated with systemic inflammation and oncological outcome in patients with high-grade soft-tissue sarcoma. PLoS One 12:e0181787
- 27. Law CW, Chen Y, Shi W, Smyth GK (2014) voom: precision weights unlock linear model analysis tools for RNA-seq read counts. Genome Biol 15:R29.
- Wilkerson MD, Hayes DN (2010) ConsensusClusterPlus: a class discovery tool with confidence assessments and item tracking. Bioinformatics 26:1572–1573

- Tusher VG, Tibshirani R, Chu G (2001) Significance analysis of microarrays applied to the ionizing radiation response. Proc Natl Acad Sci U S A 98:5116–5121
- Subramanian A, Tamayo P, Mootha VK et al (2005) Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. Proc Natl Acad Sci U S A 102: 15545–15550
- Newman AM, Liu CL, Green MR et al (2015) Robust enumeration of cell subsets from tissue expression profiles. Nat Methods 12: 453–457
- 32. Hanna SL, Fletcher BD, Parham DM, Bugg MF (1991) Muscle edema in musculoskeletal tumors: MR imaging characteristics and clinical significance. J Magn Reson Imaging 1:441–449
- Crombé A, Le Loarer F, Stoeckle E et al (2018) MRI assessment of surrounding tissues in soft-tissue sarcoma during neoadjuvant chemotherapy can help predicting response and prognosis. Eur J Radiol 109:178–187
- White LM, Wunder JS, Bell RS et al (2005) Histologic assessment of peritumoral edema in soft tissue sarcoma. Int J Radiat Oncol Biol Phys 61:1439–1445
- Fortes-Andrade T, Almeida JS, Sousa LM et al (2021) The role of natural killer cells in soft tissue sarcoma: prospects for immunotherapy. Cancers (Basel) 13:3865
- 36. Kelleher FC, Cain JE, Healy JM et al (2012) Prevailing importance of the hedgehog signaling pathway and the potential for treatment advancement in sarcoma. Pharmacol Ther 136:153–168
- Ahlawat S, Fritz J, Morris CD, Fayad LM (2019) Magnetic resonance imaging biomarkers in musculoskeletal soft tissue tumors: review of conventional features and focus on nonmorphologic imaging. J Magn Reson Imaging 50:11–27

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