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Genetic variations in the *CLNK* gene and *ZNF518B* gene are associated with gout in case–control sample sets

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Abstract A genome-wide association study of gout in European populations identified 12 genetic variants strongly associated with risk of gout, but it is unknown whether these variants are also associated with gout risk in Chinese populations. A total of 145 patients with gout and 310 healthy control patients were recruited for a case– control association study. Twelve SNPs of *CLNK* and *ZNF518B* gene were genotyped, and association analysis was performed. Odds ratios (ORs) with 95 % confidence intervals (CIs) were used to assess the association. Overall, we found four risk alleles for gout in patients: the allele "G" of rs2041215 and rs1686947 in the *CLNK* gene by

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National Engineering Research Center for Miniaturized Detection Systems, Xi'an 710069, Shaanxi, China dominant model (OR 1.66; 95 % CI 1.04–2.63; p = 0.031) (OR 2.19; 95 % CI 1.38–3.46; p = 0.001) and additive model (OR 1.39; 95 % CI 1.00–1.93; p = 0.049) (OR 1.67; 95 % CI 1.19–2.32; p = 0.003), respectively, and the allele "A" of rs10938799 and rs10016022 in ZNF518B gene by recessive model (OR 4.66; 95 % CI 1.44–15.09; *p* = 0.008) (OR 4.54; 95 % CI 1.23–16.76; p = 0.020). Further haplotype analysis showed that the TCATTCTGA haplotype of CLNK was more frequent among patients with gout (adjusted OR 0.48; 95 % CI 0.24–0.95; p = 0.036). Additionally, polymorphisms of rs2041215, rs10938799, and rs17467273 were also correlated with clinical pathological parameters. This study provides evidence for gout susceptibility genes, CLNK and ZNF518B, in a Chinese population, which may have potential as diagnostic and prognostic marker for gout patients.

Keywords Single nucleotide polymorphism (SNP) \cdot Gout \cdot *CLNK* \cdot *ZNF518B*

Introduction

Gout, a common form of inflammatory arthritis, is caused by urate crystal precipitates in the joint via an inflammatory reaction. It is increasingly assumed that gout occurs when serum uric acid (SUA) levels exceed the physiological saturation threshold for uric acid [1, 2]. Gout is characterized by persistent pain, nerve compression, and joint destruction and deformities, if left untreated [3]. With general changes in lifestyle and the rise of obesity, epidemiological data clearly demonstrate that the incidence and prevalence of gout are on the rise [4]. One to two percent of adults were affected by gout in developed countries in 2011, with the disease being more prevalent in men [5]. The prevalence rate of gout is also on the rise in China. In the Shandong coastal cities of Eastern China, reports showed that the overall gout prevalence increased to 1.14 % (1.94 % in men and 0.42 % in women) in 2008, from 0.1 % 10 years earlier [6].

An elevated concentration of SUA, or hyperuricemia (HUA), is a pivotal "danger signal" for gout and may also be a risk factor for a number of comorbidities including cardiovascular disease and mortality, hypertension, diabetes, obesity, hyperlipidemia, and metabolic syndrome [7–9]. In addition, elevated SUA levels can result in chronic kidney disease, by triggering chronic interstitial nephritis and the formation of urinary tract stones composed of uric acid [10]. Understanding the control of uric acid homeostasis is, thus, critical to improving the management and the treatment of patients with not only HUA and gout, but also a number of other diseases, which are linked to elevated SUA levels.

Although the specific pathogenesis of gout is still unclear, there is mounting evidence that both environmental and genetic factors play crucial roles in the etiology of HUA and gout. Observational studies have shown that dietary factors (animal purines, alcohol, and fructose), obesity, metabolic syndrome, hypertension, diuretic use, and chronic kidney disease may be clinically relevant risk factors for HUA and gout [11, 12]. However, epidemiological studies of genes affecting SUA levels have been performed and manifested that SUA levels are highly impacted by hereditary factors. Previous genome-wide association studies (GWAS) identified susceptibility genes, that affected UA levels, located in or near 12 genes and six regions, including hURAT1, SLC2A9, ABCG2, SLC17A1, SLC17A3, and GCKR, especially in populations of European descent [13-15].

In an attempt to discriminate other genes affecting gout, we performed a comprehensive association analysis between gout and 12 susceptible SNPs in the *CLNK* gene and *ZNF518B* gene that were previously reported to involve in the risk of gout [16]. The study sheds light on the association between common SNPs and gout risk in the Chinese population.

Materials and methods

Study participants

We recruited a total of 145 patients diagnosed with gout from 2011 to 2013 among Han Chinese. All the subjects were treated by the Affiliated Hospital of Tibet University for Nationalities and Xianyang Central Hospital. All the patients were recently diagnosed and histologically confirmed to suffer from gout according to the American College of Rheumatology classification criteria (1977) and had no history of cancer, infection, nephropathy, or other autoimmune diseases. All cases were verified, and patients were recruited without age, sex, or disease stage restriction. Moreover, patients did not receive systemic inflammatory treatment including drug control treatment before the blood samples used in this study were obtained.

A number of 310 healthy unrelated individuals were recruited randomly as sample, and the participants were Han Chinese living in Xi'an city and nearby. All of the chosen subjects were from the Medical Center in the Affiliated Hospital of Tibet University for Nationalities. To reduce the potential environmental and therapeutic factors impacting the variation of complex human diseases, we performed detailed recruitment and set exclusion criteria to exclude subjects with chronic disease and conditions involving vital organs (brain, liver, heart, and lung) and more advanced cardiovascular, metabolic, or endocrine diseases.

Clinical data and demographic

At recruitment, each subject gave written informed consent, and was interviewed by a nurse to collect detailed information including region, ethnicity, gender, age, education status, smoking status, alcohol use, occupational radiation exposure, family history of cancer, and other lifestyle factors. The use of samples was approved by the Human Research Committee of the Affiliated Hospital of Tibet University for Nationalities for Approval of Research Involving Human Subjects.

SNP selection and genotyping

Using HapMap database, candidate SNPs in the *CLNK* and *ZNF518B* gene with minor allele frequencies (MAFs) >5 % in Asian were identified in previously published polymorphisms associated with gout, resulting in 12 genotyped SNPs. The phenol–chloroform extraction method was performed to extract genomic DNA from whole blood [17]. DNA concentration was measured by spectrometry (DU530 UV/VIS spectrophotometer, Beckman Instruments, Fullerton, CA, USA). Sequenom MassARRAY Assay Design 3.0 software was used to design multiplexed SNP MassEX-TEND assay, and SNP genotyping was performed utilizing the Sequenom MassARRAY RS1000 recommended by the manufacturer [18]. Sequenom Typer 4.0 software was used to perform data management and analyses [18, 19].

Statistical analysis

We used Microsoft Excel and SPSS 16.0 (SPSS, Chicago, IL, USA) to perform statistical analyses. In this study, all p values were two-sided, and $p \le 0.05$ was considered as

Table 1 Demographic and clinical variables in gout cases		Cases	%	Control	%	p^{a}
and control patients	Total	145		310		
	Sex					0.000*
	Female	18	12.4	113	36.5	
	Male	125	86.2	197	63.5	
	Data unavailable	2	1.4			
TC total abalastaral TC	Mean \pm SD					
triglyceride. HDL-C high-	Age	43.17 ± 14.061		49.09 ± 7.877		0.000^{b}
density lipoprotein cholesterol,	TC (mmol/L)	3.95 ± 1.421				
<i>LDL-C</i> low-density lipoprotein	TG (mmol/L)	1.65 ± 1.492				
cholesterol, <i>UA</i> uric acid	HDL-C (mmol/L)	0.95 ± 0.392				
* $p \le 0.05$ indicates statistical significance	LDL-C (mmol/L)	2.08 ± 0.837				
^a Two sided Chi squared test	Urea (mmol/L)	6.83 ± 4.319				
^b Independent samples <i>t</i> test	UA (µmol/L)	507.15 ± 100.343				

achieving the threshold of statistical significance. Observed genotype frequencies were compared with expected frequencies to test for deviations from Hardy–Weinberg equilibrium (HWE). Chi-squared test/Fisher's exact test was used to calculate the allele and genotype frequencies of cases and controls [20]. Odds ratios (ORs) and 95 % confidence intervals (CIs) were used for unconditional logistic regression analysis with adjustment for age and gender [21]. The possibility of sex differences as a source of population substructure was evaluated by a genotype test for each SNP in male and female, and the number of significant results at the 5 % level was compared with the number expected by the Chi-squared test [20].

Three genetic models (dominant, recessive, and additive) were performed using PLINK software (http://pngu. mgh.harvard.edu/purcell/plink/) to estimate ORs for SNP main effects. We determined *p* values for trend by entering the variable as a single term in the model (i.e., one degreeof-freedom) and testing using the Wald's test. For SNP main effects analysis, we used ordinal variables coded as the number of variant alleles, zero, one, or two, assuming a log-additive genetic model. ORs and 95 % CIs were calculated by unconditional logistic regression analyses adjusted for age and sex [21, 22].

Finally, the Haploview software package (version 4.2) and SHEsis software platform (http://www.nhgg.org/analysis/) were used for estimate the pairwise linkage disequilibrium (LD), haplotype construction, and genetic association at polymorphism loci [23, 24].

Results

A number of 145 cases and 310 controls were enrolled in our study. The demographic and clinical variables in gout cases and control are shown in Table 1. The genotyping rate of 12 SNPs was 98.5 %, and all 12 SNPs were in Hardy–Weinberg equilibrium in control subjects (p > 0.01) (Table 2). We compared the differences in frequency distributions of alleles between cases and controls by Chi-squared test and found that two significant SNPs were associated with gout risk in the *CLNK* gene at a 5 % level (rs2041215 p = 0.033, OR 1.36; 95 % CI 1.02–1.82 and rs16869474 p = 0.002, OR 1.57; 95 % CI 1.17–2.10). To reduce the potential of spurious findings due to multiple testing, a strict Bonferroni correction analysis was applied; we found that rs16869474 (p < 0.05) satisfied the threshold between *CLNK* SNPs and risk of gout (Table 2).

Comparisons of the SNP genotypes and the risk of gout are listed in Table 3. We identified four significant SNP genotypes associated with the risk of gout. They were genotype "AA" of rs10938799 (OR 4.63; 95 % CI 1.41–15.20; p = 0.011), genotype "AA" of rs10016022 (OR 4.51; 95 % CI 1.21–16.82; p = 0.025), genotype "GT" of rs2041215 (OR 1.64; 95 % CI 1.01–2.66; p = 0.049), and genotype "GC" and "CC" of rs16869474 (OR 2.18; 95 % CI 1.34– 3.53; p = 0.002) (OR 2.22; 95 % CI 1.06–4.64; p = 0.035).

The minor allele of each SNP was assumed a risk allele compared to the wild-type allele. Minor allele frequency (MAF) in cases and controls are listed in Table 4. We performed logistic tests to analyze further model association. rs10938799 and rs10016022 were observed to be associated with gout risk by recessive model analyses (OR 4.66; 95 % CI 1.44–15.09; p = 0.008 and OR 4.54; 95 % CI 1.23–16.76; p = 0.020, respectively). We also observed two susceptibility SNPs additionally: rs2041215, dominant model analyses (OR 1.66; 95 % CI 1.04–2.63; p = 0.031) and additive model analyses (OR 1.39; 95 % CI 1.00–1.93; p = 0.049); and rs16869474 dominant model analyses (OR 2.19; 95 % CI 1.38–3.46; p = 0.001) and additive model analyses (OR 1.67; 95 % CI 1.19–2.32; p = 0.003).

SNP ID	Gene(s)	Position	Alleles A/B	HWE p value	p^{a}	padj. ^b	OR (95 % CI)
rs10938799	ZNF518B	3' UTR(10443425)	A/G	0.142	0.374	1	1.17 (0.83–1.65)
rs3217	ZNF518B	3' UTR(10444650)	T/C	0.454	0.501	1	0.89 (0.64–1.24)
rs10016022	ZNF518B	Coding exon(10446906)	A/G	0.092	0.603	1	1.10 (0.77-1.56)
rs7667644	CLNK	Intron(10495072)	C/T	0.465	0.887	1	1.03 (0.72–1.46)
rs10033825	CLNK	Intron(10496792)	T/C	1.000	0.863	1	0.97 (0.69–1.36)
rs6819820	CLNK	Intron(10498830)	A/G	0.646	0.661	1	1.06 (0.80–1.41)
rs17467273	CLNK	Intron(10500431)	C/T	0.875	0.677	1	1.07 (0.77-1.48)
rs13109939	CLNK	Intron(10501625)	C/T	0.369	0.413	1	0.86 (0.60-1.24)
rs13125670	CLNK	Intron(10513072)	T/C	0.341	0.738	1	0.95 (0.70-1.29)
rs2041215	CLNK	Intron(10519982)	G/T	0.900	0.033*	0.369	1.36 (1.02–1.82)
rs16869474	CLNK	Intron(10520972)	C/G	0.225	0.002*	0.024*	1.57 (1.17-2.10)
rs12641877	CLNK	Intron(10528226)	G/A	0.611	0.949	1	0.99 (0.74–1.33)

Table 2 Candidate SNPs examined in the ZNF518B and CLNK gene

SNP single nucleotide polymorphism, OR odds ratio, 95 % CI 95 % confidence interval, HWE Hardy-Weinberg equilibrium

* $p \le 0.05$ indicates statistical significance

^a p value were calculated using two-sided Chi-squared test

^b *p* value was adjusted by Bonferroni correction

Table 3 Distributions of genotypes of prominent SNPs	SNP_ID	Gene type	No. (frequency)		P^{a}	Logistic regression	
and their associations with risk			Case	Control		OR (95 % CI)	p^{b}
orgout	rs10938799	GG	85 (63 %)	198 (63.9 %)	0.044*	1.00	
		GA	40 (29.6 %)	105 (33.9 %)		0.99 (0.61-1.59)	0.957
		AA	10 (7.4 %)	7 (2.3 %)		4.63 (1.41–15.20)	0.011*
	rs10016022	GG	88 (65.2 %)	201 (64.8 %)	0.096	1.00	
		GA	39 (28.9 %)	103 (33.2 %)		0.98 (0.61-1.59)	0.939
* .0.05 11		AA	8 (5.9 %)	6 (1.9 %)		4.51 (1.21–16.82)	0.025*
* $p \le 0.05$ indicates statistical significance	rs2041215	TT	44 (32.6 %)	132 (42.9 %)	0.055	1.00	
^a <i>p</i> value from were calculated from two-sided Chi-squared test/Fisher's exact test		GT	70 (51.9 %)	138 (44.8 %)		1.64 (1.01–2.66)	0.049*
		GG	21 (15.6 %)	38 (12.3 %)		1.73 (0.86–3.52)	0.127
	rs16869474	GG	48 (35.6 %)	157 (50.6 %)	0.003*	1.00	
^b <i>p</i> values were calculated from		GC	68 (50.4 %)	121 (39 %)		2.18 (1.34-3.53)	0.002*
Wald's test adjusted for age and sex		CC	19 (14.1 %)	32 (10.3 %)		2.22 (1.06-4.64)	0.035*

Table 4 F	Frequency	distributions of	f prominent	SNPs and	their a	ssociations	with	the risk o	f devel	oping	gout
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SNP no.	Minor allele	MAF		Dominant model		Recessive model		Additive model	
		Case	Control	OR (95 % CI)	p^{a}	OR (95 % CI)	p^{a}	OR (95 % CI)	p^{a}
rs10938799	A	0.217	0.192	1.16 (0.73–1.83)	0.520	4.66 (1.44–15.09)	0.008*	1.34 (0.90–1.97)	0.150
rs10016022	А	0.200	0.185	1.12 (0.71–1.78)	0.630	4.54 (1.23–16.76)	0.020*	1.27 (0.85-1.90)	0.250
rs2041215	G	0.421	0.347	1.66 (1.04-2.63)	0.031*	1.32 (0.69–2.52)	0.410	1.39 (1.00–1.93)	0.049*
rs16869474	С	0.400	0.298	2.19 (1.38-3.46)	0.001*	1.49 (0.75–2.96)	0.260	1.67 (1.19–2.32)	0.003*

* $p \le 0.05$ indicates statistical significance

^a p values were calculated by unconditional logistic regression adjusted for age and sex

Furthermore, the candidate SNPs in the *CLNK* and *ZNF518B* genes showed strong linkage (Figs. 1, 2). The results for the association between the *CLNK*

haplotype and the risk of gout are listed in Table 5. Haplotype "TCATTCTGA" was found to be associated with a decreased risk of gout (OR 0.48; 95 % CI 0.24–0.95;



Fig. 1 Haplotype block map for all the SNPs of the CLNK gene



Fig. 2 Haplotype block map for all the SNPs of the ZNF518B gene

p = 0.036). We have not found any association between ZNF518B haplotype and the risk of gout.

Finally, correlations of clinicopathological parameters and prominent SNPs in patients with gout are shown in Supplementary Table. For rs2041215, a significant difference exists in total cholesterol (TC) as the genotype changes; for rs10938799 and rs10016022, a difference is seen in urea concentrations. We did not find any statistically significant associations between the clinicopathological parameters examined and rs16869474 genotypes in gout patients and control subjects.

Discussion

In this case-control study, we identified 12 previously reported gout risk loci identified through GWAS in European populations. Four susceptibility locis from chromosomes 4(rs10938799, rs10016022, rs2041215, and rs1686947) were statistically significantly associated with an increased risk of gout. We also observed that a haplotype "TCATTCTGA" of CLNK was associated with a 52 % reduction in the risk of gout. These results suggest that some gout risk variants identified in European population are also associated with risk in Han Chinese populations.

CLNK, a gene mapped in chromosome 4p16.1, is also termed MIST, which is expressed in several cell types, including T cells, natural killer cells, and mast cells, and its expression seems to be strictly dependent on sustained exposure to cytokines such as interleukin (IL)-2 and IL-3. Thus, CLNK may be involved in a cross-talk mechanism between cytokine receptor and immunoreceptor signaling [25, 26]. We have not found any evidence for the role of heredity between CLNK and gout susceptibility in previous studies. In the present study, nine CLNK SNPs (rs7667644, rs10033825,

Table 5 <i>CLNK</i> haplotypefrequencies and the associationwith gout risk in case and	Haplotype	Freq (case)	Freq (control)	χ^2	p^{a}	OR	95 % CI	p ^b
	TCGTTCTGA	0.248	0.243	0.033	0.855	1.00		_
control subjects	TTACTTGCA	0.174	0.175	0.002	0.966	0.89	0.53-1.50	0.67
	CCGTCCTGG	0.108	0.134	1.138	0.286	0.66	0.37-1.20	0.17
	TCATTCTGA	0.092	0.116	1.241	0.265	0.48	0.24-0.95	0.036*
	TCATTCGCG	0.102	0.080	1.204	0.273	1.26	0.66-2.38	0.48
	CCGTCCTGA	0.043	0.054	0.519	0.471	1.02	0.44-2.36	0.96
* $p < 0.05$ indicates statistical	TCGTTTTGA	0.023	0.041	2.029	0.154	0.61	0.22-1.75	0.36
significance	TCGTTTTGG	0.023	0.033	0.56	0.454	0.44	0.16-1.24	0.12
^a <i>p</i> value from were calculated	TTACTCGGG	0.021	0.034	1.16	0.282	0.53	0.19-1.49	0.23
from two-sided Chi-squared test	TCGTTTGCA	0.028	0.014	2.021	0.155	2.61	0.68 - 10.07	0.16
^b p values were calculated by	TCATTCTGG	0.021	0.012	1.103	0.294	1.38	0.34-5.58	0.65
adjusted for age and sex	TCGTTCGCG	0.017	0.008	1.259	0.262	2.24	0.50-10.01	0.29

rs6819820, rs17467273, rs13109939, rs13125670, rs2041215, rs16869474, and rs12641877) were genotyped, and rs2041215 and rs1686947 were identified to be associated with gout risk. Carriers of the rs2041215 G allele exhibited a statistically significant increased 1.36-, 1.66-, and 1.39-fold gout susceptibility by the allele model, dominant model, and additive model, respectively. As for rs1686947, the risk was 1.57-, 2.19-, and 1.67-fold, respectively. Simultaneously, genotype "GT" of rs2041215 predicted an increased 1.64-fold gout risk, and genotype "GC" and "CC" of rs10016022 increased gout risk 2.18and 2.22-fold. One of the most critical characteristics of gout and SUA is to stimulate the inflammatory response. Therefore, these findings indicate that CLNK may play a critical role in arthritis caused by gout. More samples and functional test are required to confirm our result. In subsequent genotype-phenotype analysis (Supplementary Table), TC presented prominent difference as the genotype changed, which may be the reason for the relationship between HUA and gout and levels of plasma lipids (cholesterol) that has been observed in a number of investigations [27].

ZNF518B, which is located in 4p16.1, has been reported in several researches. Observational studies have shown that the WDR1-ZNF518B intergenic region is implicated in a complex mechanism that regulates SCL2A9 function, which may potentially contribute to the SLC2A9-mediated effect on gender differences in human UA concentration levels [28]. However, study based on this ZNF518B gene is rare. As part of this study, the allele "A" of rs10938799 and "A" rs10016022 exhibited increased gout risk. Hence, ZNF518B gene may play an essential function in UA transport metabolism and further affect gout. Subsequently, we performed a detailed genotype-phenotype analysis among gout patient and controls (Supplementary Table). A significant difference exists in the concentration of urea as the genotype changes, which could be explained by the renal dysfunction caused by HUA and gout.

Our study is the first report on association between the SNPs *CLNK* and *ZNF518B* and gout risk. Meanwhile, GWAS is a powerful research strategy that uses SNPs as markers to identify susceptibility genes of many complex diseases. Furthermore, MassARRAY SNP genotyping method provided the reliability of these data as a broad method in the post-GWAS area. Finally, a haplotype-based association approach is an increasingly accepted approach for genetic association studies.

This study has some potential limitations. Firstly, the subgroup analyses testing for age or gender, and genderspecific significant variants were not performed because of the limited sample size. Secondly, the ethnicity of study participants was limited to the Han Chinese population. Hence, whether current research is applicable to other ethnicities still needs to be validated and therefore further meta-analysis is required to confirm our findings. Thirdly, when selecting SNPs, we achieved SNPs with MAF higher than 5 % in HapMap CHB (Chinese Han Bejing) population to ensure that the statistical power was large enough for analyzing data. This approach will leave out some significant SNPs that have been reported in other studies, and therefore, large sample size will be convincing. Finally, the function genetic variants and mechanisms underpinning this association will require additional studies including fine mapping and laboratory studies.

To sum up, we have confirmed that two genes previously reported in Europeans are associated with risk of gout in Han Chinese population for the first time, which may provide new data for screening of gout in Han population and shed light on the new candidate genes and new ideas for the study of subsequent occurrence mechanism of gout.

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Conflict of interest The authors have no conflicts of interest to report.

Ethical standard The use of samples was approved by the Human Research Committee of the Affiliated Hospital of Tibet University for Nationalities for Approval of Research Involving Human Subjects.

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