



Role of the chemical modification of titanium dioxide surface on the interaction with silver nanoparticles and the capability to enhance antimicrobial properties of poly(lactic acid) composites

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Abstract

In this work, the antimicrobial activity of neat and silanized titanium dioxide deposited with silver nanoparticles was evaluated when it was used as filler in a poly(lactic acid) matrix. The silanization and deposition processes were evaluated by scanning transmission electron microscopy and X-ray photoelectron spectroscopy confirming the chemical modification on the titanium dioxide surface by 3-aminopropyltriethoxy silane and the formation of silver nanoparticles. According to the elemental analysis conducted by energy-dispersive X-ray spectroscopy, more silver, 7.4% higher, was deposited on the oxide when this was previously silanized and when 30% w/w of silver nitrate was used as a precursor. The antimicrobial effect was confirmed for the nanoparticles through the disk diffusion method and for the composites by drop test, against *Staphylococcus aureus* and *Escherichia coli* bacteria; the results showed that the inhibition rate increased by 14.2% and 39.1% for nanoparticles and by 57.6% and 38.8% for composites against each bacteria, respectively, when deposition was performed on silanized titanium dioxide. Also, better mechanical properties were obtained in the composites filled with silanized oxide; the best results were obtained in the PLA/sTiO₂-Ag 20% system with an improvement of 45.7% in tensile stress and of 38.73% for Young's modulus. Finally, the toxicity of the composites was evaluated by seeding peripheral blood mononuclear cells; results show evidence that composites filled with these nanoparticles are non-toxic since these do not migrate from the polymeric matrix, which helps to enhance the prolonged surface antibacterial effect and to open a broad perspective of the commercial use of these composites.

Keywords Antimicrobial activity · Silver nanoparticles · Titanium dioxide · Poly(lactic acid) · Polymer composites

Introduction

Poly(lactic acid) (PLA) has increased interest in recent years due to its wide range of applications in the medical, food, and packaging field [1]. PLA is a well-known biodegradable and biocompatible polymer [2], widely considered as a suitable candidate to replace synthetic polymers due to its, thermal stability, high transparency, biocompatibility, ease of processability, lower environmental impact and reasonable price [3, 4]. In recent years, the necessity for both degradable and antimicrobial materials has been increased due to the priority to reduce residues and environmental pollution. Although PLA exhibits essential characteristics, it presents significant disadvantages as its low performance and functionality due to bacterial attack, which limits its use in major industries such as food packaging, preservation, and agriculture.

To open a broad perspective of PLA to more industrial applications, researches have worked to incorporate fillers, which allow improving the antimicrobial and mechanical properties of the film. The fillers that have been shown better microbicidal properties are silver, copper, zinc, titanium, and iron oxides [5, 6]. Silver has been recognized as an effective antimicrobial agent that exhibits low toxicity in humans [7]. Some industrial, biotechnological, and medical applications of silver, against *Escherichia coli* and *Staphylococcus aureus*, are the use of nanoparticles embedded in copolymers [8], in structural control of materials interfaces, and in health care and food packaging fields [9]. It is well known that the antibacterial activity of silver is due to Ag^+ ions, which interact with the three main components of the bacterial cell: the peptidoglycan cell wall and plasma membrane, bacterial DNA, and bacterial protein, particularly enzymes.

Recently, it is known that silver nanoparticles (AgNPs) also possess considerable antimicrobial activity [10, 11], and it is widely believed that AgNPs bind strongly to thiol groups present in the bacterial cell membrane, thereby breaking down the cell wall and, therefore, destroying the cell; these are also incorporated into the cell membrane, which causes the leakage of intracellular substances and then cell death; and finally, AgNPs also manage to penetrate the cells [10–12]. Nevertheless, AgNPs with diameters below 200 nm tend to aggregate spontaneously, and their stability in the air, water, or sunlight is not good enough for long-term applications, which will decrease their antibacterial performance [13, 14]. To solve this problem, a wide range of metal oxides such as TiO_2 , SiO_2 , and Al_2O_3 have been used to support AgNPs; therefore, Ag-deposited materials can be homogeneously formed without aggregation [10–17].

TiO_2 has been proven to be the most suitable for widespread environmental applications thanks to non-toxicity, low cost, and excellent degradation of organic pollutants [12, 13, 18]. TiO_2 has called interest in many industrial fields since it is a light-sensitive type n semiconductor that absorbs electromagnetic radiation; it is also a very chemically stable amphoteric oxide, which makes it the most used photocatalyst to degrade organic molecules, as well as to fill polymeric matrices [12, 19, 20]. Furthermore, it is well known that TiO_2 can provide a new tool for the prevention of bacterial contamination and disinfection [21, 22].

Nevertheless, TiO_2 particles tend to agglomerate and precipitate, causing a high density that limits its stability in suspension. In order to avoid this problem, these particles are modified on their surface with the purpose of making them suitable for applications; these modifications improve interactions among materials [20, 23, 24]. TiO_2 particles coated with silane, named silanization process, presents a great option because it allows the surface of metal oxides to be modified, changing their attractive interaction and helping to reduce the problem of agglomeration, reducing its interaction and dispersing very well [19, 20, 25, 26]. Organosilanes have a functional group based on hydrolytically active silicon; their carbon–silicon bond is very stable and nonpolar, and in the presence of an alkyl group, results in low surface energy and hydrophobic effects [19, 20, 25]. These coupling agents are widely used in industry to improve the performance of polymers, the resistance of the compounds to water attack significantly, and have other applications such as treatment of fillers to increase reinforcement and adhesion [19].

In order to improve mechanical properties, PLA could be filled with metal oxides, as titanium oxide, and with nanoparticles (AgNPs) [10, 13, 14, 27]. Studies of silver/poly lactide nanocomposites confirm a uniform distribution due to different chemical structures and silver–polymer interactions. This is attributed to a strong interaction between AgNPs and PLA chains, which promoted phase separation of polymer that prevents AgNPs agglomeration in the reduction process [28]. According to the literature [29], it is widely demonstrated that interactions between PLA chain molecules and AgNPs are due to the presence of van der Waals interactions between the hydroxyl groups of PLA and the partial positive charge on the surface of AgNPs.

Various works have been performed to improve the antimicrobial activities of PLA. De Silva et al. [30] prepared films with nanoparticles of ZnO deposited on the surface of halloysite nanotubes (Hal) using a novel solvothermal method; nevertheless, although this filler exhibited promising antimicrobial properties, large amounts of filler (5% and 10%) were required to obtain good results, while several regulations allow only 1% as an additive for the food industry, plus the high costs due to expensive NPs and process prices. On the other hand, Li et al. [9] used a blend of nano- TiO_2 and a silver solution; despite having good inhibition results, the method presented two main problems, the first is that these types of fillers had problems of miscibility and interfacial interaction in TiO_2 and PLA; therefore, a coupling agent is necessary to solve this; and the second is the low applicability of this precursor due to its high price and to the significant amount of silver present in the solution, joined to the reduced certainty of silver stability because no new chemical bonds were observed and there was no evidence on the formation of the NPs. Finally, Vallejo-Montesinos et al. [31] proposed a filler made with TiO_2 and AgNPs, finding that the particles modified by silanization support the integration within the matrix. Nevertheless, a considerable amount of AgNO_3 was required as the precursor of AgNPs which could cause saturation of the support, whereas the antimicrobial activity was tested on starch–chitosan films, presenting disadvantages and limitation because this material exhibits high affinity for moisture and low water stability attributed to its highly hydrophilic character, which makes its application not very feasible.

For all this, the aim of this work was to discuss the mechanism of Ag deposition on TiO_2 , evaluating if the use of a coupling agent, organosilane, would

obtain more AgNPs, and indeed, it would enhance the antimicrobial activity of the functionalized metal oxide; for this purpose, Gram-positive and Gram-negative bacteria were tested by the disk diffusion method. In addition, it was assessed the proposal to confer this property to PLA by filling it with these NPs, which was evaluated by the drop test method; and finally how the mechanical properties are influenced with these fillers.

Materials and methods

Materials

Titanium dioxide (TiO_2) particles with an average diameter of 350 nm and a crystalline structure of rutile were obtained from DuPont (R-104 Dupont, Mexico). The coupling agent 3-aminopropyltriethoxysilane (APTES, 97% purity) was supplied by Sigma-Aldrich (Mexico). Calcium hydroxide ($\text{Ca}(\text{OH})_2$, 98%) and poly(lactic acid) (3.3 relative viscosity, 15 Melt flow index, and 1.24 specific gravity) were supplied by 3M Company Mexico; and silver nitrate (AgNO_3 , 99.3%) was supplied by Fermont.

Functionalization of TiO_2 nanoparticles with APTES

TiO_2 was superficially modified with APTES in a 5:1 proportion. TiO_2 was mixed with ethanol and stirred for 30 min and then sonicated for 30 min at 60 Hz in an ultrasonic bath; this procedure was repeated until completing 2.5 h. Subsequently, APTES was added and stirred overnight. The solution was washed by centrifugation with water twice and with methanol five times, with the aim of removing the unreacted agent. Drying was carried out in an oven at 80 °C for 3 h. The dried powder was stored in vials for further testing and labeled as sTiO_2 .

Preparation of Ag-deposited TiO_2 nanoparticles

TiO_2 was mixed with water and sonicated for 5 min at 60 Hz in an ultrasonic bath; then, heated to 80 °C; and while stirring, AgNO_3 was added (at different percentages 10%, 20%, and 30% w/w). The pH was adjusted to 12 with NaOH (0.5 M), and each mixture was stirred for 2 h. Each solution was washed with water by centrifugation five times, in order to remove the unreacted agent. Drying was carried out in an oven at 80 °C until the water evaporated. The dried powder was stored in vials for further testing, and the vials were labeled as $\text{TiO}_2\text{-Ag}$ 10%, $\text{TiO}_2\text{-Ag}$ 20%, and $\text{TiO}_2\text{-Ag}$ 30%, respectively. The above procedure was repeated for the TiO_2 functionalized with APTES (sTiO_2), and the obtained products were labeled as $\text{sTiO}_2\text{-Ag}$ 10%, $\text{sTiO}_2\text{-Ag}$ 20%, and $\text{sTiO}_2\text{-Ag}$ 30%, respectively.

Characterization of Ag-deposited TiO₂ nanoparticles

Scanning transmission electron microscopy (sTEM) analysis was performed to confirm the superficial modification and the Ag deposition. The eight samples TiO₂, sTiO₂, TiO₂-Ag (at 10%, 20%, and 30% w/w), and sTiO₂-Ag (at 10%, 20%, and 30%) were dispersed in an aqueous medium, making serial dilutions until achieving the optimum particle concentration. A drop was placed on a copper grid and observed in a JEOL equipment (model JEM 1230 microscope) with a resolution of 0.4 nm and an acceleration voltage of 100 kV.

X-ray photoelectron spectroscopy (XPS) was used to measure the chemical surface composition of the material and the chemical and electronic state of the elements that compose the surface. For the XPS analysis, an ultra-high vacuum (UHV) system from PerkinElmer PHI5100 was used with an SCA 10-360 analyzer detector. A dual X-ray source of Mg K α ($h\nu = 1256$ eV) at 300 W and 20 mA beam intensity was used, with a polarized anode at 15.0 kV. The XPS spectrum was obtained at 54° from normal to the surface with a constant energy step (CAE) $E_0 = 71.55$ eV for the full scan spectrum and $E_0 = 22.36$ eV for high-resolution spectra. The pressure was maintained, during measurement, at 1×10^{-8} Torr. The energy position was calibrated with the Ag 3d_{5/2} orbital at the position of 368.20 eV with a resolution (FWHM) of 1.10 eV, Au 4f_{7/2} at 84.00 eV, and C 1s at 284.75 eV. Elemental composition analysis was performed using as a basis for the relative sensitivity factor reported by Scofield [30].

Antimicrobial activity Ag-deposited TiO₂ nanoparticles by disk diffusion method

The antimicrobial activity of the eight systems TiO₂, sTiO₂, TiO₂-Ag (at 10%, 20%, and 30%), and sTiO₂-Ag (at 10%, 20%, and 30%) was evaluated by disk diffusion method [32] against bacteria Gram-positive (*Staphylococcus aureus*) and Gram-negative (*Escherichia coli*). The eight samples were sterilized in a 1 ppm solution with ethanol at 70% v/v. The disks were placed in vials and soaked with each freshly prepared system. Then, the disks were left to impregnate. The microbial strains of bacteria, *E. coli*, and *S. aureus*, were inoculated into nutrient broth and kept overnight for incubation in a furnace. The turbidity of broth cultures was compared with 0.5 McFarland solutions, corresponding to $1-2 \times 10^8$ colony-forming units (CFU mL⁻¹).

The solid growth medium used for *E. coli* and *S. aureus* was eosin methylene blue (EMB) and mannitol agar, respectively. Petri dishes were inoculated using a sterile swab with the microorganisms, rotating the box 60° for each striatum application. The inoculated dishes were let settle and dried for 15 min with the lid closed. Finally, the disks previously impregnated in each of the TiO₂ systems were placed on each dish. Petri dishes were incubated in a furnace at 37 °C for 24 h. Finally, the zones of inhibition (halo) of each system were measured under an extraction hood. The assays were performed in triplicate, and mean values of zone diameter were taken.

Preparation of PLA films with Ag-deposited TiO₂ nanoparticles

PLA/TiO₂, PLA/sTiO₂, PLA/TiO₂-Ag (at 10%, 20%, and 30%), and PLA/sTiO₂-Ag (at 10%, 20%, and 30%) nanocomposites at 1% w/w were prepared. 500 mg of PLA was dissolved in 20 mL of chloroform; once a homogeneous solution was obtained, 5 mg of NPs was added, and then stirred and sonicated for 30 min at 60 Hz in an ultrasonic bath. The suspension obtained was placed on a glass container (silica), and the solvent was allowed to evaporate at room temperature overnight with the purpose of obtaining the film. This procedure was made for each system of nanoparticles.

Antibacterial analysis of PLA films by drop test method

PLA and its composites, PLA/TiO₂, PLA/TiO₂-Ag 10%, PLA/TiO₂-Ag 20%, PLA/TiO₂-Ag 30%, PLA/sTiO₂, PLA/sTiO₂-Ag 10%, PLA/sTiO₂-Ag 20%, and PLA/sTiO₂-Ag 30%, were tested against *E. coli* or *S. aureus* using the antibacterial drop test method with some modifications [22]. The same inoculated microbial strains of bacteria were used as for the disk method. A film of each respective composite was introduced in a different test tube, in triplicate, each one containing nutrient broth inoculated with the respective bacteria, Gram-positive and Gram-negative. Moreover, a film of pure PLA was used as a positive control, and a culture of each microorganism was grown without any treatment as a negative control. Then, the tubes were incubated for 24 h, and the solutions were read with a spectrophotometer. The optical absorptions $[A]$ were obtained, and the inhibition rate (%) was calculated as follows:

$$\text{Inhibition rate (\%)} = \frac{[A]_i - [A]_c}{[A]_i} \times 100 \quad (1)$$

where $[A]_i$ is the optical absorption of untreated bacteria and $[A]_c$ is the absorption of each composite.

Effect of Ag-deposited TiO₂ nanoparticles in PLA films under mechanical properties

The mechanical tests of PLA and composites PLA/TiO₂, PLA/TiO₂-Ag 10%, PLA/TiO₂-Ag 20%, PLA/TiO₂-Ag 30%, PLA/sTiO₂, PLA/sTiO₂-Ag 10%, PLA/sTiO₂-Ag 20%, and PLA/sTiO₂-Ag 30% were determined using an INSTRON universal machine model 3369 with a charge of 50 kN and a jaw separation speed of 25 m min⁻¹. Tensile stress and Young's modulus values were recorded with the aim of knowing the capacity at which materials will maintain the integrity and resistance during handling.

Evaluation of ion release by ICP-MS

One sample of each composite (PLA and PLA/TiO₂ and PLA/sTiO₂ at each concentration) was immersed in deionized water and sonicated for 30 min at 60 Hz in an ultrasonic bath, and then 2% of HNO₃ was added. The amounts of released titanium and silver ions were determined by analyzing the resulting solutions using an inductively coupled plasma mass spectroscopy (Model X, Series 2 ICP-MS by Thermo Electron Corporation); this spectrophotometer detects ions in each mass in a rapid sequence, allowing the individual isotopes of an element to be determined. The electrical signals resulting from the detection of the ions are processed within the digital information that is used to indicate the intensity of the ion and, subsequently, the concentration of the element.

Evaluation of cell viability

Cell viability was investigated by seeding peripheral blood mononuclear cells (PBMC) in 96-well plates (Costar, Corning, NY, USA) at a cell density of 3×10^4 cells per well in RPMI-1640 medium (supplemented with 10% fetal bovine serum, 2 mM l⁻¹ glutamine, 50 U ml⁻¹ penicillin, and 50 mg ml⁻¹ streptomycin; Gibco-Invitrogen Corp., Carlsbad, CA, USA). The mononuclear cells were exposed to different concentrations (5% and 11% v/v) of the solutions resulting from each composite; these solutions were obtained the same as in the ICP-MS analysis and were cultured for 24 h at 37 °C in an atmosphere of 5% CO₂ and 95% relative humidity. After this time, the cells were treated with resazurin (Sigma-Aldrich, St. Louis, MO, USA) at 30 µg ml⁻¹ diluted in RPMI medium in a final volume of 100 µl per well. Plates were incubated immediately for 2 h at 37 °C and 5% CO₂ atmosphere and protected from light. Fluorescent measurements with an excitation wavelength $\lambda = 560$ nm and an emission wavelength $\lambda = 590$ nm were performed with the objective of determining the degree of metabolism of resazurin in a Spectrofluorometer Synergy H1 (BioTek Instruments Inc. Winooski, VT, USA). Untreated PMBC was used as a negative control, and cells treated with acetone (10%) were set as a positive control (Sigma-Aldrich, St. Louis, MO, USA).

Cell viability of all dosing treatments was determined in triplicate using the following equation:

$$\text{Cell viability (\%)} = \frac{F_S}{F_C} \times 100\% \quad (2)$$

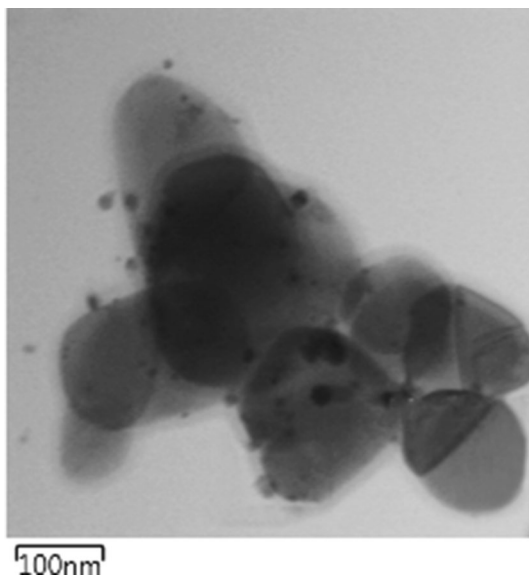
where F_S is the fluorescence value of each sample and F_C is the fluorescence of the control.

Results and discussion

sTEM analysis of Ag-deposited TiO₂ nanoparticles

sTEM was used to confirm both the Ag deposition and the chemical modification of TiO₂ with APTES. The image corresponding to the system TiO₂-Ag 30% is shown in Fig. 1; as can be seen, there are TiO₂ particles with a size of 100–180 nm, forming agglomerates due to their high density; also, it is possible to observe small particles of Si, incrustated on the TiO₂ particles of size between 15 and 35 nm, and finally small dots of Ag of 5–11 nm are observed in all the agglomerates. With the goal of quantifying the amount of Ag deposited, an elemental analysis with the help of energy-dispersive X-ray spectroscopy (EDS) was performed; the results are observed in Fig. 2, and the amount of each element is listed in Table 1, in which 1.2% of Ag was measured in the whole system of TiO₂-Ag 30%. On the other hand, Fig. 3 shows the image for sTiO₂-Ag 30%, which reveals an APTES organic coating on the TiO₂ surface of around 25 and 30 nm; actually, these particles present a better dispersion since no large agglomerates are observed, agreeing with previous investigations [25]; finally, more dots of Ag are observed than in the previous system but of equal size. The results of EDS analysis (Fig. 4) showed that more Ag was deposited in the sTiO₂-Ag 30% system, measuring up to 7.4%. According to both tests, sTEM and EDS, Ag was observed attached to the Si atoms; therefore, the silanized system, sTiO₂-Ag 30%, improved the deposition of Ag since more Si atoms (22.6% according to EDS) are presented in the surface of the oxide. It is important to measure that sTEM at high magnification is a more or less surface-related analytical tool. As a result, only a limited number of particles can be detected or visualized in such a small volume, and the limit of detection is therefore high. Energy-dispersive X-ray

Fig. 1 sTEM images for TiO₂-Ag 30%



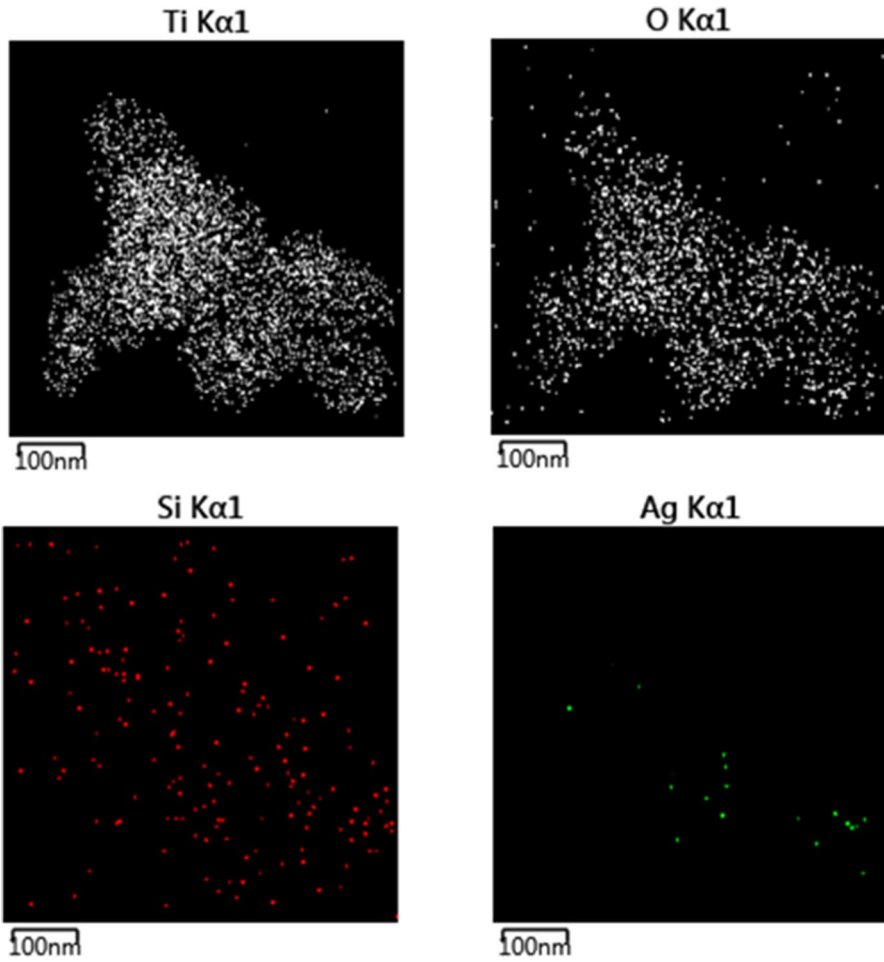
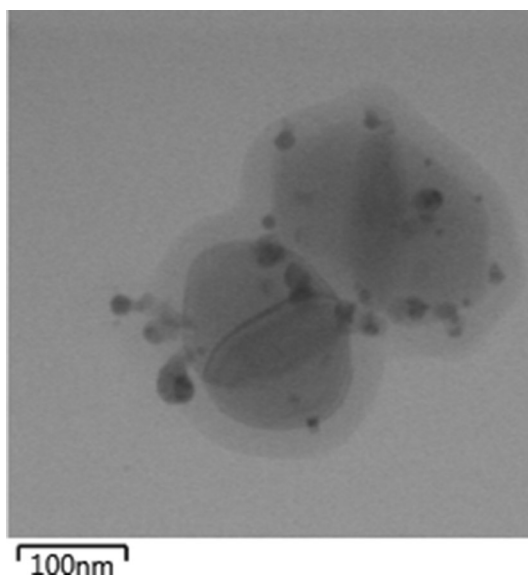


Fig. 2 EDS analysis for TiO₂-Ag 30%

Table 1 Weight % of elements in TiO₂-Ag 30% and sTiO₂-Ag 30% systems obtained from EDS

Element	System	
	TiO ₂ -Ag 30% (%)	sTiO ₂ -Ag 30% (%)
Ti	62.2	48.8
O	34.4	21.3
Si	2.2	22.6
Ag	1.2	7.4

Fig. 3 sTEM images for sTiO₂-Ag 30%



spectroscopy (EDS or EDX) become even more important tool for the determination of elemental composition of the observed samples [33]. This means that through this technique (sTEM), it is possible to obtain an approximate concentration of the analyzed surface. Nevertheless, the main concern for this investigation is to fully know how the AgNPs were deposited and how TiO₂ was superficially modified with the coupling agent APTES and with the variation of the AgNO₃ amount. This was possible to observe by means of X-ray photoelectron spectroscopy (XPS), an analysis that allows to measure the chemical surface composition, the binding, and the chemical and electronic state of the elements that compose the surface.

Characterization of Ag-deposited TiO₂ nanoparticles by XPS

X-ray photoelectron spectrometry was carried out in order to further investigate the binding between Ti and Ag atoms on the surfaces of the TiO₂ and sTiO₂ systems: TiO₂-Ag 20%, TiO₂-Ag 30%, sTiO₂-Ag 20%, and sTiO₂-Ag 30%. Deconvolution analysis of XPS raw spectra was performed to estimate the presence of all species of each element. First, the XPS results for TiO₂ (Fig. 5a) and sTiO₂ (Fig. 5d) are compared, which show the Ti 2*p* core-level spectrum as two peaks at 464.5 and 458.8 e.V. for neat TiO₂ and at 464.3 and 458.7 e.V. for silanized TiO₂. These intense peaks can be assigned to the spin-orbital splitting of the Ti 2*p*_{1/2} and Ti 2*p*_{3/2}, respectively; and it is important to point out that the distance between them is ~5.7 e.V.; all this proves clearly that this element consists of Ti⁴⁺ species on the sample surface [34–37]. This oxidation state for titanium is the most common in TiO₂; and according to the literature [38, 39], this is due to its route of synthesis (chlorination). In Fig. 5d, it is possible to observe that after the functionalization with APTES the titanium signals shift to a slightly lower B.E. (~0.2 eV), which is indicative of

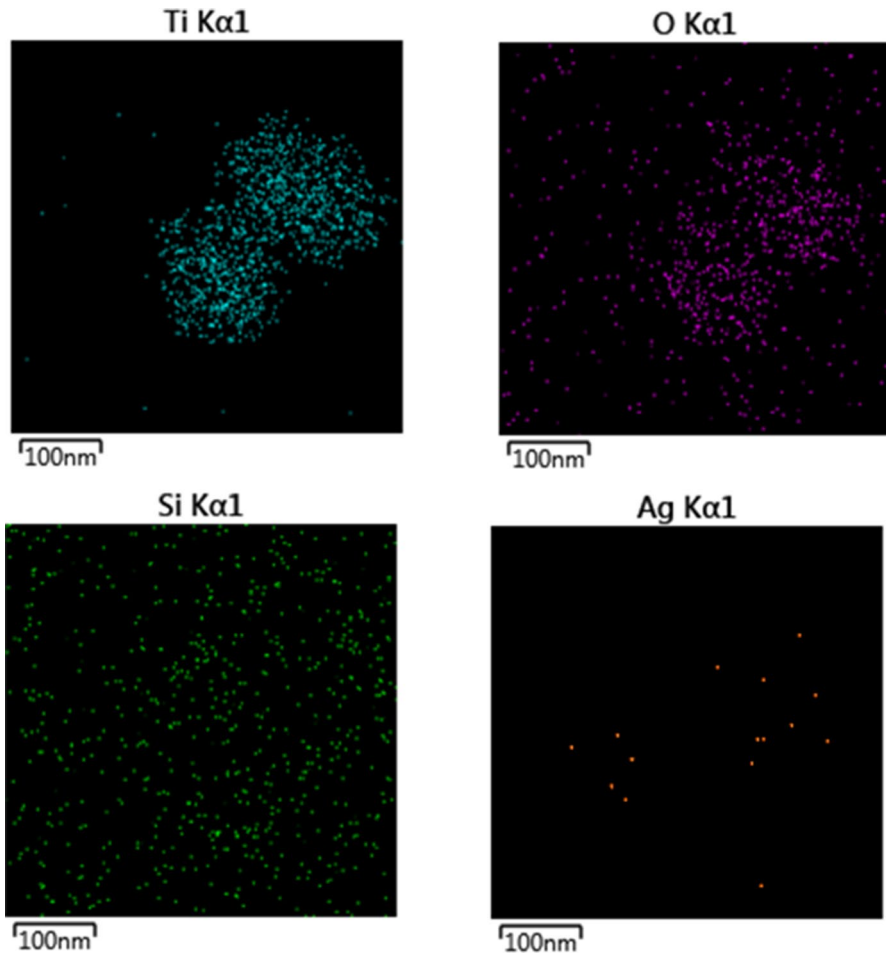


Fig. 4 EDS analysis for sTiO₂-Ag 30%

more electron-rich Ti atoms [40], and therefore, this surface improves its capacity to donate electrons allowing more deposition of AgNPs, which is shown in Fig. 8.

When Fig. 5a is compared to the results obtained for the Ag-deposited systems, the binding energy at Ti 2*p* increases as the amount of AgNO₃ (precursor) is added in the system, shifting in 0.3 e.V. for TiO₂-Ag 20% (Fig. 5b) and in 1.2 e.V. for TiO₂-Ag 30% (Fig. 5c); and for the silanized system (Fig. 5d) in 0.9 e.V. for sTiO₂-Ag 20% (Fig. 5e) and in 1.2 e.V. for sTiO₂-Ag 30% (Fig. 5f). According to the literature [41, 42], this phenomenon is because the Fermi level of Ag is lower than that of TiO₂, which results in a decrease in the outer electron cloud density of Ti ions and therefore in an increase in binding energy. On the other hand, it is possible to see that the Ti spectra for the Ag-deposited systems (Fig. 5b, c, e, f) present a new peak at ~461 e.V. and a flattening in the signals at ~464 and ~468 e.V.; this

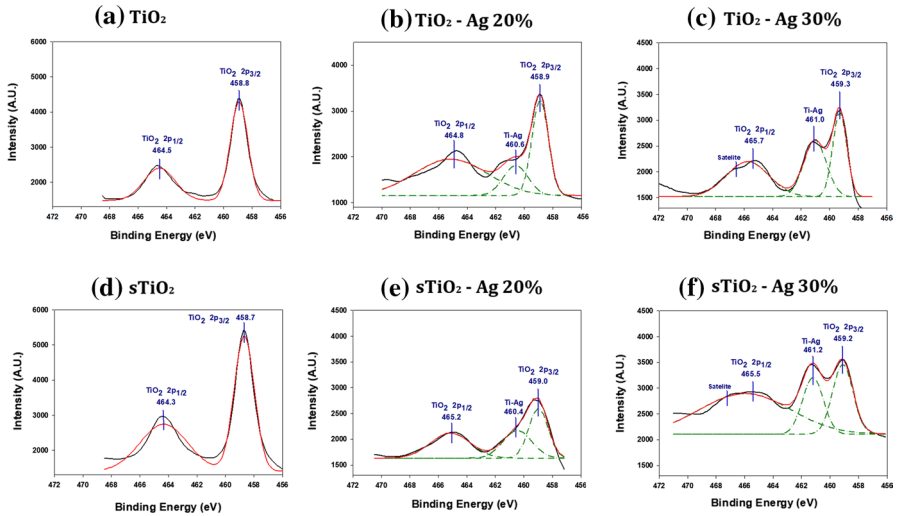


Fig. 5 XPS spectra for Ti element

could be interpreted as the partial reduction of the Ti on the surface after metal position, which is the production of a metal–metal bonding between Ag and Ti [43, 44].

Meanwhile in Fig. 6, the XPS spectrum of O1s for pure TiO₂ shows an asymmetric peak related to different chemical states. The characteristic peaks indicate that the value at ~530 e.V. is attributed to metal oxides (Ti–O–Ti) [42], and the peak at ~532 e.V. corresponds to the hydroxyl group (–OH) [45]. After silanization, it is observed a decrease in the B.E. (~0.2 eV) and in the intensity of the Ti–O–Ti

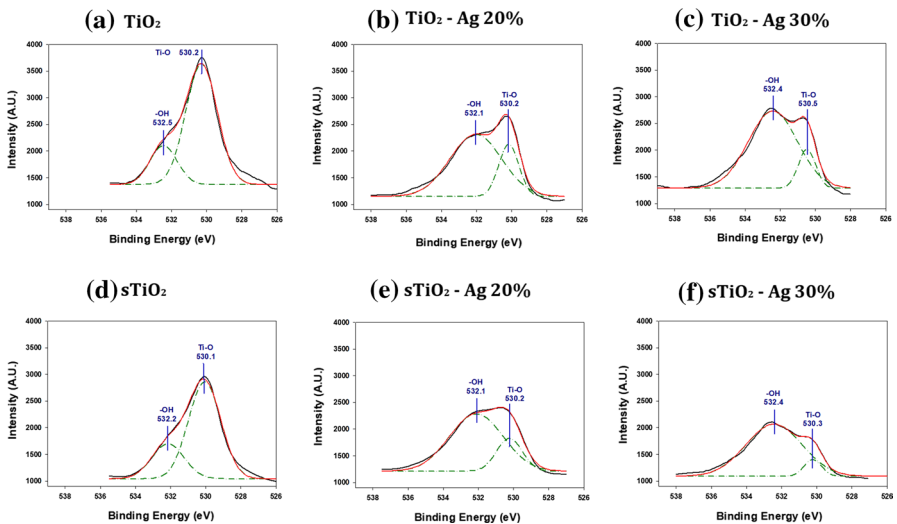


Fig. 6 XPS spectra for O element

Table 2 XPS results of TiO₂, sTiO₂ and Ag-deposited TiO₂ systems

Element	Specie	System					
		TiO ₂ (%)	sTiO ₂ (%)	TiO ₂ -Ag 20% (%)	TiO ₂ -Ag 30% (%)	sTiO ₂ -Ag 20% (%)	sTiO ₂ - Ag 30% (%)
C	Adventitious carbon	36.3	20.5	–	–	–	–
	C–O–C	39.7	43.7	71.2	46.1	39.0	50.0
	Oxidant carbon	24.0	35.7	17.4	23.7	36.9	19.0
	Carboxyl carbon	–	–	11.4	30.3	24.1	31.0
O	–OH	21.9	26.7	75.4	84.9	78.3	90.8
	O–Ti	78.1	73.3	24.6	15.1	21.7	9.2
Ti	TiO ₂ 2p _{1/2}	36.9	45.2	56.6	37.9	36.2	58.5
	TiO ₂ 2p _{3/2}	63.1	54.8	31.4	30.0	34.9	22.7
	Ti–Ag	–	–	15.0	32.2	28.9	18.8
Ag	Ag 3d _{3/2}	–	–	18.9	15.3	15.5	17.3
	Ag 3d _{5/2}	–	–	30.0	21.5	31.8	22.5
	Ag–Ti 3d _{5/2}	–	–	26.1	35.4	21.8	33.2
	Ag–Ti 3d _{3/2}	–	–	25.0	27.8	30.9	27.0

signal for the oxide functionalized with APTES, while according to the results in Table 2, the relative intensity of the hydroxyl group (–OH) increased by 4.8%; this indicates that the oxygen was changed from Ti–O–Ti into the chemical state of the hydroxyl state (–OH) [45]. This is likely due to surface band bending, which determines the formation of a negative space charge layer and implies that Ti⁴⁺ ions achieve a more reduced state [40]. Regarding the Ag-deposited systems (20% and 30%), the intensity of –OH peaks enlarged as the w/w concentration of AgNO₃ (precursor) increased, in 53.5% for TiO₂–Ag 20%, 63.0% for TiO₂–Ag 30%, and for the silanized systems in 56.5% for sTiO₂–Ag 20% and 68.9% for sTiO₂–Ag 30%, which possibly favors the oxidation processes on the TiO₂ surface with the production of reactive oxygen species (ROS) [46].

Regarding the C1s core-level spectra in Fig. 7, three peaks were shown at ~283, ~285, and ~286 e.V. in the TiO₂ and sTiO₂ systems, which are attributed to an adventitious carbon, C–O–C bond, and oxidant carbon such as alcohols, respectively, which are particularly frequent on reactive titanium surfaces [47]. The relative intensity of the C–O–C increased by four units after the silanization process; according to this, it is inferred that the carbon that exists near the oxygen in the (Ti–O) on the TiO₂ surface changed into the carbon that exists near the hydrogen in the Ti–OH bound [45].

As can be seen in Fig. 8, Ag signals were found only in the deposited systems exhibiting two characteristic peaks for the Ag 3d spectrum. The binding energy at ~374 e.V. corresponds to Ag 3d_{3/2}, while the second peak at ~368 e.V. is for Ag 3d_{5/2}, suggesting the presence of Ag⁰ (368.2 eV) [42, 48]. The results of this study, together with sTEM and EDS analysis, indicate the presence of metal Ag-deposited systems. It is important to mention that there are no remains of precursor AgNO₃

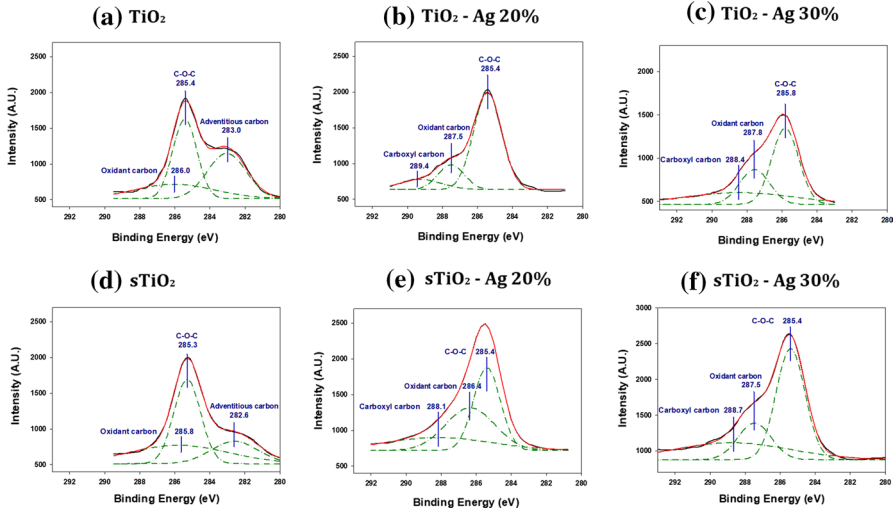


Fig. 7 XPS spectra for C element

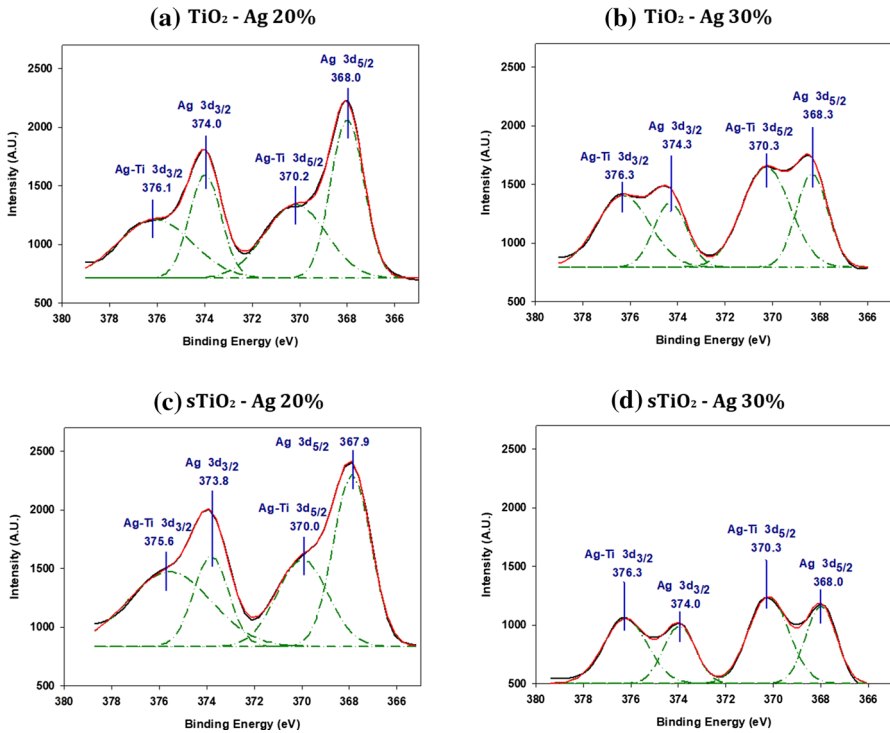


Fig. 8 XPS spectra for Ag element

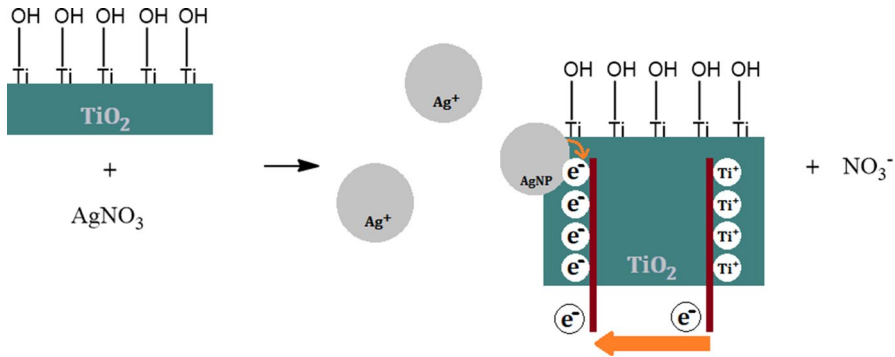


Fig. 9 Proposed reaction mechanism of Ag deposition on neat titanium dioxide (TiO₂)

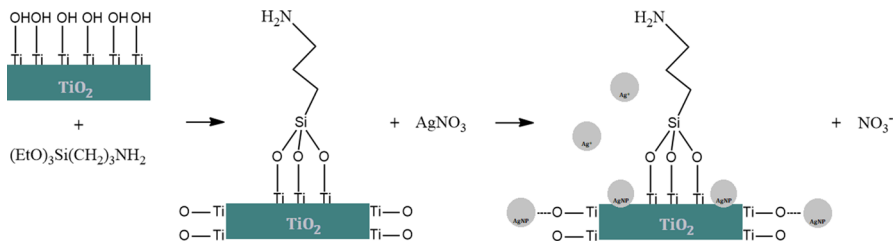
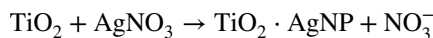


Fig. 10 Proposed reaction mechanism of Ag deposition on silanized titanium dioxide (sTiO₂)

since nitrogen was not detected at ~400 e.V. [49]. On the other hand, for the 20% and 30% systems, it was possible to observe two forming peaks at ~376 and ~370 e.V., respectively; these peaks are not an indicator of silver oxidation, but the Ti partial reduction due to the bond of Ag–Ti [44, 50]. This can be explained as the breaking of the original bonds with metal oxides, and the subsequent formation of new ones with silver (AgO), as confirmed by the appearance of peaks at ~376 and ~370 e.V., which is confirmed in Fig. 8b and d with the systems exhibiting an increase in these Ag–Ti peaks and higher ratios regarding Ag species (Ag–Ti/Ag): 51.1% for TiO₂–Ag 20%, 63.2% for TiO₂–Ag 30%, 52.7% for sTiO₂–Ag 20%, and 60.2% for TiO₂–Ag 30%).

Proposed reaction mechanism of Ag-deposited TiO₂

According to the results of the sTEM and XPS tests, it is possible to propose two reaction mechanisms for Ag deposition. The first one is for non-functionalized titanium dioxide, illustrated in Fig. 9, where the Ag nanoparticles have been deposited on the TiO₂ surface through a reduction in an AgNO₃ solution:



Silver ions in the vicinity of the TiO_2 surface capture electrons from titanium and deposit on the surface in the form of silver nanoparticles, creating Ti–Ag bonds.

When TiO_2 was silanized with APTES (Fig. 10), hydroxyl groups ($-\text{OH}$) were formed originating more active sites; therefore, the original bond Ti–O–Ti was broken, and silver oxide was formed (Ag–O). With this mechanism, more Ag could be deposited on the surface of TiO_2 .

Thus, the presence of AgNPs on the TiO_2 surface favors oxidation processes on the TiO_2 surface with the production of reactive oxygen species (ROS), including the superoxide ion (O_2^-) and hydroxyl radical (OH) [46]. It is well known that the cell membrane of the bacteria has a negative charge due to the presence of lipids, carboxyl, phosphate, and amino groups; therefore, it attracts Ag^+ ions released from AgNPs due to electrostatic forces. This interaction of positive ions with phospholipid bilayer results in altering cell membrane permeability and reduction in the elastic modulus, extracellular products leak in, and intracellular contents leak out, ROS are generated, and finally, the microorganism dies (Fig. 11).

Antimicrobial activity of Ag-deposited TiO_2 nanoparticles by disk diffusion method

The antimicrobial activity of the disks impregnated with the systems was assessed on two bacteria, *E. coli* and *S. aureus*, under the disk diffusion method. The paper disk diffusion method is excellent for determining the susceptibility of bacteria to antibiotics and bactericidal agents [51], this is based on the presence or the absence of a zone of inhibition. For determining antimicrobial susceptibility, an acceptable disk diffusion method should be based not only on the presence or absence of a zone of inhibition but also on the size of the inhibition zone. Reliable results can only be obtained with disk diffusion tests that use the principle of standardized methodology

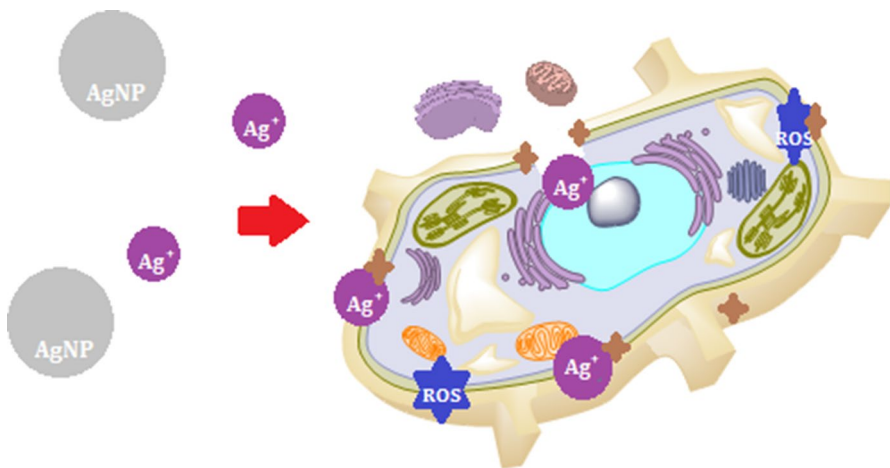


Fig. 11 Schematic mechanisms for antimicrobial effect of the AgNPs

and zone diameter measurements with strains known to be susceptible or resistant to various antimicrobial agents [52].

The measurements of inhibition zone in diameter (mm) around the impregnated disks are shown in Table 3. The results revealed that the Ag-deposited systems had an antimicrobial effect when compared to titanium dioxide, TiO₂, sTiO₂. It is clearly observed an increment in the inhibition zone for the silanized systems, and the best results were obtained for the sTiO₂-Ag 30%, system with inhibitions diameter of 10.6 mm for *S. aureus* and 18.4 mm for *E. coli*. This is attributable to this system that had more Ag content, according to the sTEM and EDS analysis, in comparison with the other systems. Yallappa et al. [53] performed a disk diffusion method to measure the antibacterial activity of Au, Ag, and Au–Ag alloy nanoparticles, using erythromycin (10 µg/disk) as organic antimicrobial standard. The antimicrobial zone of inhibition of the standard was 8 and 13 mm to *S. aureus* and *E. coli*, respectively. This means that the results of sTiO₂-Ag 30% (silanized system) on *S. aureus* (10.6 mm) and *E. coli* (18.4 mm) are above the inhibition range.

This bactericidal effect was caused by Ag⁺ ions, which interact with bacterial cells and adhere to the bacterial walls [17] affecting the peptidoglycan cell wall and plasma membrane, bacterial DNA, and bacterial protein, particularly enzymes [5, 13]; on the other side, the effect of concentration in the system is clearly noted; a higher percentage (20–30%) of Ag increased the inhibition zone for both bacteria. It is noteworthy that silanized systems enhanced antimicrobial effect for Ag-deposited TiO₂; this can be attributed to silane agent that provides a better dispersion of the Ag and TiO₂, not only generating a high surface area, which caused a high antimicrobial efficiency of these nanoparticles [53] but also electrostatic forces between bacteria and Ag⁺ ions improving their adhesion [17].

Analysis of films by antibacterial drop test method

The films obtained following “Preparation of PLA films with Ag-deposited TiO₂ nanoparticles” section for the different systems are shown in Fig. 12. According to the drop test method described in “Antibacterial analysis of PLA films by drop test method” section, it was possible to observe clearly the antibacterial activity for the PLA composites filled with Ag-deposited TiO₂. As observed in Fig. 13, the results indicated that the antimicrobial activity of PLA/TiO₂-Ag is attributed to the AgNPs deposited on TiO₂ surface, since a better inhibition rate is observed

Table 3 Antibacterial activity of sTiO₂, TiO₂ and Ag-deposited TiO₂ by Disk diffusion method

Microorganisms	Mean zone of inhibition (mm)							
	TiO ₂	sTiO ₂	10% w/w		20% w/w		30% w/w	
			TiO ₂ -Ag	sTiO ₂ -Ag	TiO ₂ -Ag	sTiO ₂ -Ag	TiO ₂ -Ag	sTiO ₂ -Ag
<i>S. aureus</i>	ND	ND	ND	ND	ND	ND	9.1	10.6
<i>E. coli</i>	ND	ND	ND	ND	ND	12.4	11.2	18.4

ND not detected

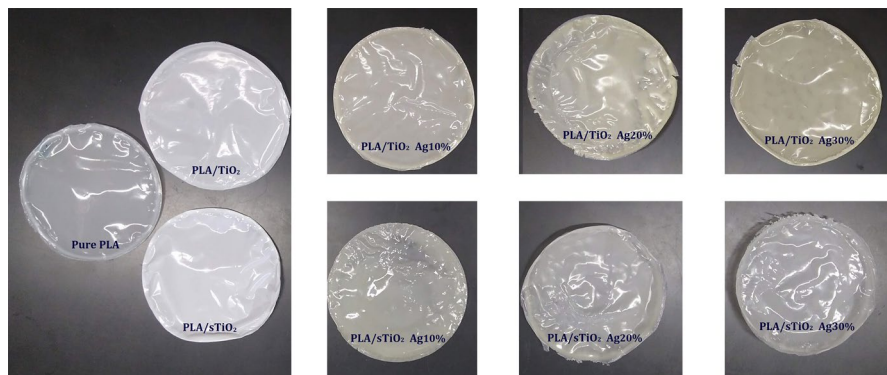


Fig. 12 Films of PLA and its composites

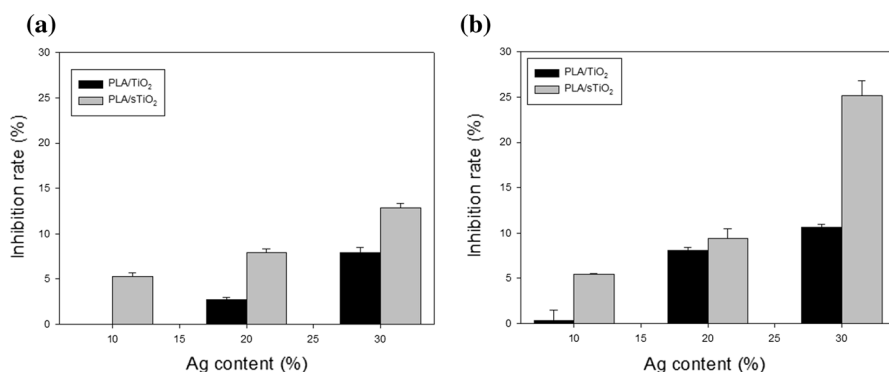


Fig. 13 Inhibition rates for **a** *E. coli* and **b** *S. aureus* as a function of Ag content for PLA composites

when the Ag content increased. The results of the inhibition rate are shown in Fig. 13a for *E. coli* and in Fig. 13b for *S. aureus*.

As observed, the inhibitory effect of PLA/TiO₂-Ag and PLA/sTiO₂-Ag films for *S. aureus* was stronger than that for *E. coli*. This can be attributed to the fact that the wall of Gram-positive cells contains a thick layer and, conversely, Gram-negative cell walls are more complex since this cell contains a thin membrane and an outer membrane, which cover the wall of the surface and confer resistance to hydrophobic compounds. Therefore, the structure of the cell wall plays an important role in the susceptibility of bacteria in the presence of AgNPs [22, 54]. Finally, it is possible to conclude that the best inhibitory rate was obtained again for the systems with silanized TiO₂, especially for the PLA/sTiO₂-Ag at 30%, thanks to the greater presence of AgNPs in the filler. In this case, the antibacterial effect was caused by contact with silver, since previous trials of disk diffusion method using the polymer composites as disks on agar did not show inhibition zone, while in the drop test method, a clear inhibition was found.

The antimicrobial effect of AgNPs is known to be greatly influenced by the type and size of the AgNPs and the uncontrolled release of AgNPs or silver ions from the carrier matrices. Therefore, in the PLA/TiO₂-Ag composites, non-functionalized and silanized, the polymer matrices can help to control the release of silver ions by altering the interactions between AgNPs and polymeric materials [55]. These findings are consistent with other studies. In LDPE-silver nanocomposites, clear inhibition zone of 10.71 and 1.52 mm was observed by agar diffusion test to *S. aureus* and *E. coli*, respectively [56]. In cellulose-AgNPs nanocomposites, the inhibition zone of nanocomposites active films was 3.11 and 2.75 mm to *S. aureus* and *E. coli*, respectively; they indicated that Gram-positive strains are more sensitive to silver compounds than Gram-negative bacteria, which attributed to binding of silver ions to peptidoglycan layer [57]. More oxygen vacancies on the TiO₂ surface increase the nucleation of metallic AgNPs, besides TiO₂ contributing to the improvement in physical and mechanical properties on fillers. Previous studies reported the enhancement of antibacterial activity by synergistic effect between Ag and metal oxides, which could be mainly due to the increase in the generation of the reactive oxidative species (especially superoxide) and the increased damage to plasmid DNA induced by AgNPs [58].

Effect of Ag-deposited TiO₂ nanoparticles under mechanical properties

In order to investigate the effect of nanoparticles on the mechanical properties of PLA film, especially the effect of silanization on the TiO₂ particles in the tensile stress and Young’s modulus tests, all composites were evaluated and the results are represented in Fig. 14. In this, it is possible to observe that the presence of NPs within the polymeric matrix increased the rigidity of the resulting composites, even just with the inclusion of TiO₂ without AgNPs; this improvement is due to the strong interfacial interaction between TiO₂ and polymeric matrix [22, 30].

In Fig. 14a, the values indicate that the presence of Ag has influence in the tensile stress test (Fig. 14a) since higher values were obtained when the concentration increased. However, for the highest percent of Ag (30%), there was a decrease in the value, probably to the excess of AgNPs that tend of agglomerate in the filler; also,

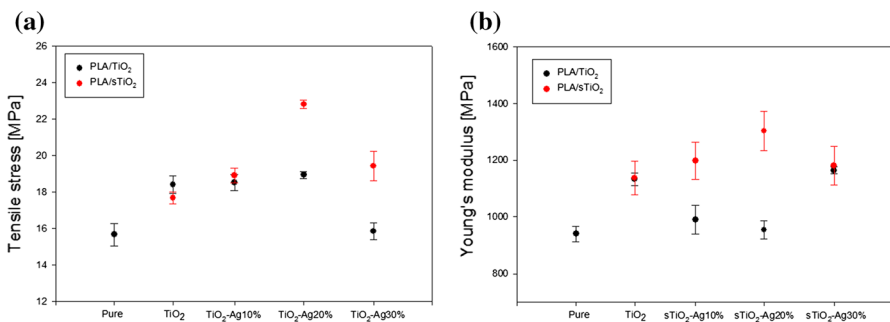


Fig. 14 Variation of **a** tensile stress and **b** Young’s modulus as function of Ag content for PLA composites

this can be explained by the aggregation of TiO_2 in the matrix due to the interaction between the Ti-OH and Ti-Ag groups on the particle surfaces [9], especially for the non-silanzed particles. It is essential to point out the higher values found for the systems silanzed with APTES because this coupling agent improves the integration of the particles within the matrix thanks to the strong interfacial adhesion and better dispersion between the sTiO_2 nanoparticles [19, 30]. The best result was found for the composite $\text{PLA/sTiO}_2\text{-Ag}$ 20% with an improvement of 45.7% in relation to pure PLA.

Similar behavior was observed for Young's modulus (Fig. 14b), in which the elasticity decreased as Ag content increased, exclusive for the composites with silanzed particles, with the exception of $\text{PLA/sTiO}_2\text{-Ag}$ 30%, while the non-silanzed systems had less stiffness [59, 60] and practically exhibit the same values than pure PLA, except for the $\text{PLA/TiO}_2\text{-Ag}$ 30%. The high values found in composites with silanzed particles are an indicator that these particles make films harder [61], which makes the composite ideal for applications where a resistant material is required. The best improvement was in 38.73% for $\text{PLA/sTiO}_2\text{-Ag}$ 20% composite. For all this, it is possible to state that it is widely demonstrated that the proper interactions between PLA chain molecules and AgNPs are due to the presence of van der Waals interactions between the hydroxyl groups of PLA and the partial positive charge on the surface of AgNPs [29] and that the silanzed systems had better mechanical properties thanks to their terminal NH_2 groups, which form intermolecular hydrogen bonds with the OH- groups of the PLA [31].

Evaluation of ion release by ICP-MS

Table 4 shows the concentration of Ti and Ag ions released from each composite sample measured by ICP-MS. The presence of these ions is observed only in four composites and in very low concentrations. In $\text{PLA/TiO}_2\text{-Ag}$ at 10 and 20%, the Ag release maybe is due to the incomplete efficiency reaction on metal nanostructures, such as long reaction time, for example, in previous investigation; there was observed oxidations of p-amino thiophenol (PATP) to p,p'dimercaptoazobenzene

Table 4 Concentration of ion release for composites by ICP-MS analysis

Composite	Ion concentration (ppb)	
	Ti	Ag
PLA	0	0
PLA/TiO_2	0	0
$\text{PLA/TiO}_2\text{-Ag}$ 10%	0	3.4
$\text{PLA/TiO}_2\text{-Ag}$ 20%	0	0.5
$\text{PLA/TiO}_2\text{-Ag}$ 30%	0	0
PLA/sTiO_2	0	0
$\text{PLA/sTiO}_2\text{-Ag}$ 10%	1.2	0
$\text{PLA/sTiO}_2\text{-Ag}$ 20%	0	0
$\text{PLA/sTiO}_2\text{-Ag}$ 30%	5.2	11.3

(DMAB) in Ag nanoparticles (AgNPs) and an incomplete reduction of 4-nitrobenzenethiol (4NBT) to DMAB on Ag nanoparticle film hybrid system or Ag nanowire-Au nanoparticle nanostructures [62]. While only Ti was slightly detected in the composites with sTiO₂-Ag but in scarce concentrations, the highest value of Ag ion was detected in the PLA/sTiO₂-Ag 30%; the composite with the highest amount of silver, but in a scarce value of ~11 ppb, may be due to excess silver not properly anchored to the TiO₂ surface. Although the silanized particle improves the AgNPs deposition, this reaction is limited by a certain number of sites in which the nanoparticle can interact with the OH residues of APTES, so the rest of the silver is distributed in the polymer matrix and migrate because it is not anchored to the support material. These results suggest that an intermediate concentration between 20 and 30% of AgNPs could give a better deposited-composite relation. All this achieves not only the non-toxicity of the composites but also a prolonged surface antibacterial effect [63].

Evaluation of cell viability

The cytotoxic effect of each resulting solution was evaluated on PBMC at two different concentrations (5% and 11%) through Resazurin assay. Figure 15 shows the results for each composite, and according to this, it is possible to state that the resulting solutions do not have a negative effect on the PBMC viability for any treatment during 24 h of exposure, compared to the control sample. It is well known that the intrinsic properties of Ag-deposited TiO₂ nanoparticles, such as size, shape, and chemical composition, could correlate with the development of cytotoxic effects [64]. In this context, it has been observed that the nanoparticles that may enter the bloodstream and have direct contact with proteins and cells generate a reaction that decreases cell viability through oxidative stress and damages DNA, apoptosis, and others [65]; nevertheless, these resulting solutions did not exert a toxic effect since cell viability was above 80%, representing non-toxic conditions for cells, in accordance with ISO 10993-5:2009.

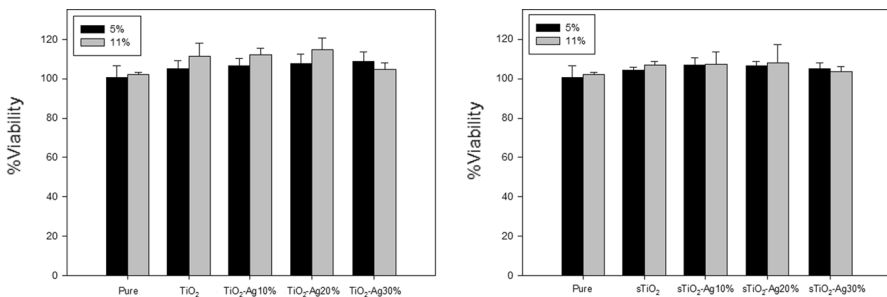


Fig. 15 Cell viability of PLA and its composites with a neat TiO₂ and b silanized TiO₂

Conclusions

In this work, pure and silanized titanium dioxide was deposited with AgNPs by reduction of AgNO_3 , testing three different concentrations (10%, 20%, and 30% w/w) of this precursor. The results of the sTEM analysis confirm the chemical modification of TiO_2 with APTES, since an organic coating on the TiO_2 surface of around 25 and 30 nm is observed in the s TiO_2 systems, which was not previously observed in pure TiO_2 . The Ag deposition on the oxide particles was confirmed by the sTEM, XPS, and EDS analysis. With these results, it was possible to conclude that a higher amount of silver (7.4%) was present when the amount of AgNO_3 precursor was incremented and when the oxide was previously silanized, thanks to the silanization process that created more active sites on the TiO_2 surface, favoring the bound of silver.

Respect to the antimicrobial effect, favorable results were found in Ag-deposited TiO_2 nanoparticles since they showed antimicrobial activity against microorganisms such as *Staphylococcus aureus* and *Escherichia coli* at 20% and 30% w/w of AgNO_3 (precursor). It is clearly shown that there were better inhibition results when deposition was performed on titanium dioxide previously silanized, 14.2% for *S. aureus*, and 39.1% for *E. coli* in % inhibition rate. According to the results, it was possible to confer the antimicrobial property to PLA composites by incorporating Ag-deposited TiO_2 as filler, and the inhibition behavior was replicated according to what was observed for the silanized particles, showing enhancement in 57.6% for *S. aureus* and 38.8% for *E. coli*. Regarding mechanical properties, better properties were obtained when silanized particles were used as fillers, since higher results were obtained in the functionalized composites, in comparison with pure PLA. The best result was found for the PLA/s TiO_2 -Ag 20% composite with an improvement of 45.7% for tensile stress and of 38.73% for Young's modulus. Therefore, these films are harder, making them ideal for applications where a tough material is required.

Finally, it was found that PLA composites with these nanoparticles are non-toxic because there is practically no migration of ions from the polymeric matrix, which helps to enhance the prolonged surface antibacterial effect. Therefore, in accordance with the favorable results of the research, a broad perspective is opened on the use of Ag-deposited TiO_2 , previously silanized, to create antimicrobial and non-toxic PLA composites that can be used at the industrial and commercial levels in the medical, food, and agricultural field, in areas as water purification, the stability of liposomes, systems with photoactivation, biomolecules, drug encapsulation, environment, among others.

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Compliance with ethical standards

Conflict of interest The authors declare no conflicts of interest.

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