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Kinetic study of chitosan degradation by an electrochemical process

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Abstract The kinetics of chitosan degradation by an electrochemical process was studied in this work. The order of degradation reaction was determined according to the dependence of degradation rate constant on initial chitosan concentration. For electrochemical degradation of chitosan, the apparent rate constant varied inversely with initial chitosan concentration, suggesting that the degradation reaction was zeroth-order in chitosan concentration. The influence of experimental conditions on the degradation rate constant was also investigated in detail. The degradation rate constant increased with current density, acetic acid concentration, and temperature. The influence of temperature on the degradation rate was modeled using the Arrhenius equation and the activation energy was determined to be 14.16 kJ/mol under the experimental conditions examined. The variation of sodium acetate concentrations had a negligible influence on degradation rate of chitosan.

Keywords Chitosan · Electrochemical · Degradation · Kinetics

Introduction

Chitosan is a copolymer of 1-4 linked β -D-glucosamine and β -D-N-acetylglucosamine. Chitosan and its derivatives have a special set of interesting properties: nontoxicity, biocompatibility, controllable biodegradability, and nonantigenicity. These properties make chitosan an attractive biopolymer for applications in wide

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areas such as biotechnology, pharmaceuticals, wastewater treatment, cosmetics, agriculture, food science, and textiles [1-3]. However, the applications of chitosan are limited by its high molecular weight and poor solubility. When chitosan is degraded, its solubility and some special biological, chemical and physical functions can be improved [4, 5].

Many methods have been applied to degrade chitosan such as acid hydrolysis [6], oxidative degradation [7, 8], enzymatic method [9], ultrasonic degradation [10], gamma radiation [11], ultraviolet degradation [12, 13], microfluidization [14], and microwave irradiation [15]. We recently communicated a novel, electrochemically based approach in which chitosan was treated by electrolysis using Ti/TiO₂–RuO₂ electrode without additions of chemical and biochemical reagents [16]. The results showed that the molar mass of chitosan dramatically decreased with the increase of electrolysis time and current density, and the chemical structure of degraded chitosan was not obviously modified. The main part of this previous short communication [16] focused on demonstration of the feasibility of degradation of chitosan by an electrochemical process, while this work mainly aims to investigate two fundamental aspects which have not yet been investigated in detail: (1) the kinetic behavior of electrochemical degradation of chitosan; (2) the influence of experimental conditions including chitosan concentration, current density, reaction temperature, acetic acid concentration and sodium acetate concentration on the degradation rate constant.

Experimental

Materials

Chitosan, with degree of deacetylation (DD) of 89.8% and viscosity average molecular weight (M_v) of 327 kDa, was purchased from Jinke Biochemical Co. (Zhejiang, China). All other chemicals were of analytical grade. Distilled water was used. Ti/TiO₂-RuO₂ electrode (5 cm × 7 cm) was prepared by thermal deposition of TiO₂ and RuO₂ (Ti/Ru molar ratio = 7:3) on Ti plate [17].

Analytical methods

Viscosity of dilute chitosan solutions in 0.1 mol/L sodium acetate–0.2 mol/L acetic acid was measured at 30.0 \pm 0.1 °C using a glass capillary Ubbelohde viscometer with a capillary diameter of 0.58 mm. The dilute solutions had relative viscosities, $\eta_{\rm rel}$, from about 1.2 to 2.0 to assure good accuracy and linearity of extrapolation to zero concentration. Flow times were measured in triplicate, for each sample, and their average values were used for the calculations. The limiting viscosity number ("intrinsic viscosity"), [η], was obtained by double extrapolation to zero concentration of Huggins' and Kraemer equations, respectively

$$\frac{\eta_{\rm sp}}{C} = [\eta] + k' \cdot [\eta]^2 \cdot C \tag{1}$$

$$\frac{\ln \eta_{\rm rel}}{C} = [\eta] + k'' \cdot [\eta]^2 \cdot C \tag{2}$$

where η_{rel} and η_{sp} are the (dimensionless) relative and specific viscosities, k' and k'' are the Huggins' and Kraemer's coefficients, respectively, and *C* is the solution concentration.

The M_v of chitosan was calculated using the Marke–Houwink–Sakurada relationship (3):

$$[\eta] = K \cdot M_{\rm v}^a \tag{3}$$

where a and K are constants for the buffer solution and chitosan system.

These constants were calculated using the two following equations, proposed by Kasaai [18] as a model to calculate a and K for chitosan in any solvent–temperature system using viscometric constant data previously reported by several research groups:

$$a = 0.6202 + \left(\frac{0.699x}{0.4806 + x}\right) \tag{4}$$

$$\log K \cdot 10^{-5} = -5.7676a + 5.9232 + 0.003(T - 20) \tag{5}$$

where $x = [DA/(pH \cdot \mu)]$, with DA, degree of acetylation of chitosan (DA = 1 - DD), pH of chitosan solution in a solvent with ionic strength of μ , *T*, temperature of chitosan solution [18].

The potentiometry method was used to determine the DD of original chitosan and its degraded products [19]. Dried chitosan of 0.2 g was accurately weighed and dissolved in 20 mL of 0.1 mol/L hydrochloric acid standard solution, and stirred for 2 h at 25 ± 0.1 °C. The mixture was diluted with 20 mL double distilled water, and then was titrated with 0.1 mol/L caustic soda standard solution. The degree of deacetylation (DD) was calculated as follows:

$$w = \frac{0.016(c_1V_1 - c_2V_2)}{G(1 - W)} \times 100\%$$
(6)

where c_1 is the concentration of HCl (mol/L); c_2 , the concentration of NaOH (mol/L); V_1 , the volume of HCl (mL); V_2 , the volume of NaOH (mL); G, the sample weight (g); W, the water percentage of sample (%) and 0.016 is the weight of NH₂ equal to 1 mL of 1 mol/L HCl (g). Then:

$$DD = \frac{w}{9.94\%} \times 100\%$$
 (7)

where 9.94% is the theoretical NH₂ percentage of chitosan.

Electrolysis of chitosan

Chitosan and sodium acetate were completely dissolved in acetic acid solution firstly. The electrolysis of 250 mL chitosan solution was performed in a reactor (diameter 7 cm × height 12 cm) with Ti/TiO₂–RuO₂ as anode, stainless steel as cathode (5 cm × 7 cm) and the spacing of 20 mm between two electrodes. A DC potentiostat was used as the power supply for the electrolysis. During the electrochemical treatment the solution was stirred constantly and the temperature was controlled continuously with a thermostat. After degradation, the resulting solution was

neutralized with 2 mol/L NaOH solution to pH 8–9, added ethanol (twice volume of the solution), filtered off, washed thoroughly with ethanol, and the products were collected after drying overnight in a vacuum at 60 °C.

Theoretical model

In this study, the electrochemical degradation of chitosan might be zero-, first-, or second-order degradation reaction. In order to investigate the kinetic behavior, *n*th-order kinetic equation was used to describe the electrochemical degradation of chitosan.

The *n*th-order kinetic is given by [20]:

$$\frac{\mathrm{d}L}{\mathrm{d}t} = -kL^n \tag{8}$$

where L is the total number of degradable linkages, t is the reaction time, k is the rate constant, and n is the order of reaction.

When n = 1, the degradation reaction is first-order process and its kinetic is given by

$$\frac{\mathrm{d}L}{\mathrm{d}t} = -kL\tag{9}$$

L can be substituted to N(M/m - 1) where *N* is the total number of polymer chains, *M* is the average molecular weight, and *m* is the molecular weight of the repeating unit. Assuming that $M/m \gg 1$, Eq. 9 can be obtained as follows:

$$\frac{1}{M_t} = \frac{1}{M_0} + \frac{kt}{m} \tag{10}$$

where M_0 and M_t are average molecular weight of polymer at time zero and time t, respectively.

When $n \neq 1$, the following result is obtained by integrating Eq. 8 and making appropriate substitutions:

$$\left[\left(1 - \frac{m}{M} \right)^{1-n} - \left(1 - \frac{m}{M_0} \right)^{1-n} \right] = \frac{k(n-1)}{N_0^{1-n}} t \tag{11}$$

Thus, for zeroth order, n = 0

$$\frac{1}{M_t} = \frac{1}{M_0} + \left(\frac{k}{mN_0}\right)t \tag{12}$$

And, for second order, n = 2

$$\frac{1}{M_t} \cong \frac{1}{M_0} + \left(\frac{kN_0}{m}\right)t \tag{13}$$

Here, the approximation made for the second-order case is similar to that made for the first-order analysis and is valid as long as the degradation is not carried out to the extent that only very small polymer molecules remain. In previous studies, the inverse relation between molecular weight and reaction time was traditionally considered as evidence for first-order kinetics [20–22]. However, Eqs. 10, 12, and 13 reveal that, regardless of the order of the reaction, the same functional dependence of molecular weight on reaction time is predicted for zero-, first-, and second-order degradation reaction. Therefore, this linear relationship between $1/M_v$ and *t* is not enough evidence to determine the true reaction order [20].

Instead, the true reaction order can be determined by examining the effect of initial polymer concentration (c_0) on the apparent rate constant, *S* (slope of 1/*M* vs. reaction time plot). According to Eqs. 10, 12, and 13, the equation of *S* can be derived as follows [20]:

$$S = \frac{kN_0^{n-1}}{m} \tag{14}$$

Substituting

$$c_0 = \frac{N_0 V}{N_{\rm av} m} \tag{15}$$

where $N_{\rm av}$ is the Avogadro number and V is the sample volume, then

$$S \propto c_0^{n-1} \tag{16}$$

Thus, for a zeroth-order reaction (n = 0), S is proportional to $(1/c_0)$; for a first-order reaction (n = 1), S is independent of c_0 ; for a second-order reaction (n = 2), S is proportional to c_0 . It is, therefore, critical that the relationship between S and c_0 is checked before any conclusions on reaction kinetics are made.

Results and discussions

Determination of reaction order

In order to determine the reaction order of electrochemical degradation of chitosan, we first plotted $1/M_v$ as a function of reaction time at different chitosan concentrations under conditions of 0.2 mol/L acetic acid, 0.3 mol/L sodium acetate, 120 mA/cm², and 60 °C (Fig. 1). As shown in Fig. 1, the inverse M_v changed linearly with reaction time. In previous studies, this linear relationship between $1/M_v$ and t was traditionally considered as evidence for first-order kinetics [20–22]. Our above theoretical analysis reveals that this linear relationship is expected regardless of the reaction order and the true order can be determined according to the dependence of degradation rate constant on initial polymer concentration.

For determining the relationship of *S* and c_0 , the *S* value was obtained from the slope of the straight line in Fig. 1 and then the plot of *S* as a function of c_0 was shown in Fig. 2. As can be seen from Fig. 2, *S* is proportional to $(1/c_0)$ in the range of 0.25–0.45% (w/v) chitosan concentrations. According to the above theoretical analysis, this relationship of *S* and c_0 confirms that the electrochemical degradation is zeroth-order in chitosan concentration. A possible explanation of the observed



Fig. 1 $1/M_v$ of chitosan versus treatment time at different initial chitosan concentrations of 0.25% (w/v) (*a*), 0.3% (w/v) (*b*), 0.35% (w/v) (*c*), 0.4% (w/v) (*d*), and 0.45% (w/v) (*e*)



Fig. 2 Apparent rate constant as a function of initial chitosan concentration

zeroth-order degradation reaction presents as follows. In this study, there are sufficient chitosan molecules for the electrochemical degradation at 0.25-0.45% (w/v) chitosan concentrations. Thus, the increase of chitosan concentration did not increase the number of degraded bonds of chitosan molecules, implying that the electrochemical degradation is zeroth-order in chitosan concentration under the experimental conditions examined. Similar kinetic behavior was observed for enzymatic catalysis of simple substrates at high substrate/enzyme ratios and was termed "substrate saturation" [20, 23].

Effect of current density on degradation rate constant

Current density is an important factor affecting the electrolysis kinetics. Thus, the electrolysis experiments at different current densities of 40, 80, 120 and 160 mA/ cm² were conducted under conditions of 0.3% (w/v) chitosan, 0.2 mol/L acetic acid, 0.3 mol/L sodium acetate and 60 °C. The variation of $1/M_v$ of chitosan with reaction time at different current densities is shown in Fig. 3. The *S* for each experiment was calculated from the slopes of best fit lines in Fig. 3 and then summarized in Table 1. As shown in Table 1, significant differences between the *S* values obtained at different current densities. When the current densities varied from 40 to 160 mA/cm², the *S* increased from 1.13×10^{-7} to 7.47×10^{-7} mol/ (g min), demonstrating that the increase in current density would result in an increase in *S* values. A similar effect of current density on an electrochemical degradation was also observed in other studies [24–26].

Effect of temperature on degradation rate constant

Like other electrochemical reactions, temperature could have a significant effect on the rate of degradation. Therefore, the electrolysis experiments at different reaction temperatures of 30, 40, 50 and 60 °C were conducted under conditions of 0.3% (w/v) chitosan, 0.2 mol/L acetic acid, 0.3 mol/L sodium acetate and 120 mA/cm². The variation of $1/M_v$ of chitosan with treatment time at different reaction temperatures is shown in Fig. 4. The *S* for each experiment was calculated from the slopes of best fit lines in Fig. 4 and then summarized in Table 2. As shown in Table 2, when the reaction temperatures were varied from 30 to 60 °C, the *S* increased from 2.39×10^{-7} to 3.95×10^{-7} mol/(g min). It indicated that the increase of reaction temperature had a positive effect on degradation rate of



Fig. 3 $1/M_v$ of chitosan versus treatment time at different current densities of 160 mA/cm² (*a*), 120 mA/cm² (*b*), 80 mA/cm² (*c*), and 40 mA/cm² (*d*)

Table 1Apparent rateconstants (S) for electrochemicaldegradation of chitosan at	Current density (mA/cm ²)	S (mol/(g.min))
different current densities	40	1.13×10^{-7}
Experimental conditions: 0.3% (w/v) chitosan, 0.2 mol/L acetic acid, 0.3 mol/L sodium acetate, and 60 °C	80	2.34×10^{-7}
	120	3.95×10^{-7}
	160	7.47×10^{-7}



Fig. 4 $1/M_v$ of chitosan versus treatment time at different reaction temperatures of 60 °C (*a*), 50 °C (*b*), 40 °C (*c*), and 30 °C (*d*)

Table 2 Apparent rateconstants (S) for electrochemicaldegradation of chitosan at	Reaction temperature (°C)	S (mol/(g min))
different reaction temperatures ^a	30	2.39×10^{-7}
^a Experimental conditions:	40	2.89×10^{-7}
0.3% (w/v) chitosan, 0.2 mol/L	50	3.43×10^{-7}
acetic acid, 0.3 mol/L sodium acetate, and 120 mA/cm ²	60	3.95×10^{-7}

chitosan. Hence, the electrochemical degradation of chitosan was accelerated synchronously with temperature increase.

The influence of temperature on the rate of degradation was modeled using the Arrhenius equation [21, 26]:

$$\ln S = \ln A - \frac{E_a}{RT} \tag{17}$$

where A is the frequency factor, R is the universal gas constant, 8.314 J/(K mol), and T is the temperature in Kelvin. The apparent rate constant for the electrolysis of 0.3% (w/v) chitosan in 0.2 mol/L acetic acid–0.3 mol/L sodium acetate solution at

120 mA/cm² was plotted in the Arrhenius plot (Fig. 5). From the data shown in Fig. 5, the activation energy (E_a) for the degradation reaction was determined to be 14.16 kJ/mol which is significantly lower than that for H₂O₂ degradation ($E_a = 88.5$ kJ/mol) or nitrous acid hydrolysis of chitosan ($E_a = 87.1$ kJ/mol) [21, 27].

Effect of sodium acetate concentration on degradation rate constant

The addition of an electrolyte in solution during electrolysis can influence the treatment, since it modifies the conductivity of the solution and facilitates the passage of the electrical current. Thus, the investigation of influence of sodium acetate (used as supporting electrolyte) concentration on the degradation rate constant of chitosan was carried out by electrolysis of 0.3% (w/v) chitosan under conditions of 0.2 mol/L acetic acid and 60 °C. The sodium acetate concentrations varied from 0.1 to 0.6 mol/L and the current density of 120 mA/cm² was held constant during the electrochemical treatment. As shown in Fig. 6, when the sodium acetate concentrations varied from 0.1 to 0.2 mol/L, the S increased from 3.56×10^{-7} to 3.89×10^{-7} mol/(g min). Whereas, when the sodium acetate concentrations varied from 0.2 to 0.6 mol/L, the S was statistically the same. Therefore, the variation of sodium acetate (supporting electrolyte) concentrations had a negligible influence on degradation rate of chitosan under constant current density. A similar effect of supporting electrolyte concentration on an electrochemical degradation was also observed in other studies [26, 28, 29]. Moreover, it can be seen Fig. 6 that the voltage decreased from 17.8 to 7.2 V as the sodium acetate concentrations varied from 0.05 to 0.6 mol/L, indicating that increasing sodium acetate concentration decreased the voltage during the electrochemical treatment.



Fig. 5 Arrhenius plot of the degradation of 0.3% (w/v) chitosan in 0.2 mol/L acetic acid-0.3 mol/L sodium acetate solution at 120 mA/cm²



Fig. 6 Effect of sodium acetate concentration on apparent rate constant and voltage under conditions of 0.3% (w/v) chitosan, 0.2 mol/L acetic acid, 60 °C, 120 mA/cm²

Effect of acetic acid concentration on degradation rate constant

Investigation of the effect of acetic acid concentration on the degradation rate constant of chitosan was carried out by electrolysis of 0.3% (w/v) chitosan under conditions of 0.3 mol/L sodium acetate, 120 mA/cm² and 60 °C. The acetic acid concentrations varied from 0.05 to 0.6 mol/L during the electrochemical treatment. As shown in Fig. 7, when the acetic acid concentrations varied from 0.05 to 0.6 mol/L, the *S* increased from 2.54 × 10⁻⁷ to 5.84×10^{-7} mol/(g min). It indicated that higher acetic acid concentration resulted in a better degradation of



Fig. 7 Effect of acetic acid concentration on apparent rate constant under conditions of 0.3% (w/v) chitosan, 0.3 mol/L sodium acetate, 60 °C, 120 mA/cm²

chitosan. A similar effect of acid concentration on an electrochemical degradation was also observed in other studies [25, 30, 31]. Moreover, these *S* values for the electrochemical degradation of chitosan varied from 1.13×10^{-7} to 7.47×10^{-7} mol/(g min) under the experimental conditions examined. They are less than those for H₂O₂ degradation of chitosan (2.60×10^{-6} – 4.40×10^{-6} mol/(g min)) calculated from the data of Chang et al. [21] and those for nitrous acid hydrolysis of chitosan (4.18×10^{-4} – 4.04×10^{-5} mol/(g min)) calculated from the data of Allan et al. [27], but higher than those for ultrasonic degradation of chitosan ($\sim 0.58 \times 10^{-7}$ mol/(g min)) calculated from the data of Chen et al. [32].

Conclusions

In this work, the degradation rate constant was inverse dependent of initial chitosan concentrations in the range of 0.25-0.45% (w/v) which confirmed that the electrochemical degradation is zeroth-order in chitosan concentration. During the electrochemical treatment, the experimental conditions (current density, reaction temperature and acetic acid concentration) affected the degradation rate of chitosan. The degradation rate constant increased with current density, acetic acid concentration and temperature. The influence of temperature on the rate of degradation was modeled using the Arrhenius equation and the activation energy was determined to be 14.16 kJ/mol under the experimental conditions examined. The variation of sodium acetate concentrations had a negligible influence on degradation rate of chitosan.

References

- Carmen RL, Roland B (1997) Mechanical, water uptake and permeability properties of crosslinked chitosan glutamate and alginate films. J Control Release 44:215–225
- Majeti NV, Kumar R (2000) A review of chitin and chitosan applications. React Funct Polym 46: 1–27
- Zheng LY, Zhu JF (2003) Study on antimicrobial activity of chitosan with different molecular weights. Carbohydr Polym 54:527–530
- Kittur FS, Vishu-Kumar AB, Tharanathan RN (2003) Low molecular weight chitosans preparation by depolymerization with *Aspergillus niger pectinase*, and characterization. Carbohydr Res 338: 1283–1290
- Qin CQ, Li HR, Xiao Q, Liu Y, Zhu JC, Du YM (2006) Water-solubility of chitosan and its antimicrobial activity. Carbohydr Polym 63:367–374
- Rege PR, Block LH (1999) Chitosan processing: influence of process parameters during acidic and alkaline hydrolysis and effect of the processing sequence on the resultant chitosans properties. Carbohydr Res 321:235–245
- Qin CQ, Du YM, Xiao L (2002) Effect of hydrogen peroxide treatment on the molecular weight and structure of chitosan. Polym Degrad Stab 76:211–218
- Huang QZ, Zhuo LH, Guo YC (2008) Heterogeneous degradation of chitosan with H₂O₂ catalysed by phosphotungstate. Carbohydr Polym 72:500–505
- Li J, Du YM, Liang HB (2007) Influence of molecular parameters on the degradation of chitosan by a commercial enzyme. Polym Degrad Stab 92:515–524

- Czechowska-Biskup R, Rokita B, Lotfy S, Ulanski P, Rosiak JM (2005) Degradation of chitosan and starch by 360-kHz ultrasound. Carbohydr Polym 60:175–184
- 11. Choi WS, Ahn KJ, Lee DW, Byun MW, Park HJ (2002) Preparation of chitosan oligomers by irradiation. Polym Degrad Stab 78:533–538
- Andrady AL, Torikai A, Kobatake T (1996) Spectral sensitivity of chitosan photodegradation. J Appl Polym Sci 62:1465–1471
- Yue W, He R, Yao P, Wei Y (2009) Ultraviolet radiation-induced accelerated degradation of chitosan by ozone treatment. Carbohydr Polym 77:639–642
- Kasaai MR, Charlet G, Paquin P, Arul J (2003) Fragmentation of chitosan by microfluidization process. Innov Food Sci Emerg Tech 4:403–413
- Shao J, Yang YM, Zhong QQ (2003) Studies on preparation of oligoglucosamine by oxidative degradation under microwave irradiation. Polym Degrad Stab 82:395–398
- Cai Q, Gu Z, Chen Y, Han W, Fu T, Song H, Li F (2010) Degradation of chitosan by an electrochemical process. Carbohydr Polym 79:783–785
- 17. Beer HB (1972) Electrodes and coating therefore. US Patent 3,632,498
- Kasaai MR (2007) Calculation of Mark–Houwink–Sakurada (MHS) equation viscometric constants for chitosan in any solvent–temperature system using experimental reported viscometric constants data. Carbohydr Polym 68:477–488
- Lin R, Jiang S, Zhang M (1992) The determination of degree of deacetylation of chitosan. Chem Bull 3:39–42
- Tayal A, Kelly RM, Khan SA (1999) Rheology and molecular weight changes during enzymatic degradation of a water-soluble polymer. Macromolecules 32:294–300
- Chang KLB, Tai MC, Cheng FH (2001) Kinetics and products of the degradation of chitosan by hydrogen peroxide. J Agri Food Chem 49:4845–4851
- Lii CY, Chen CH, Yeh AI, Lai VMF (1999) Preliminary study on the degradation kinetics of agarose and carrageenans by ultrasound. Food Hydrocoll 13:477–481
- 23. Cheng Y, Prud'homme RK (2000) Enzymatic degradation of guar and substituted guar galactomannans. Biomacromolecule 1:782–788
- Kim S, Kim TH, Park C, Shin EB (2003) Electrochemical oxidation of polyvinyl alcohol using a RuO₂/Ti anode. Desalination 155:49–57
- Mahmoud SS, Ahmed MM (2009) Electrocatalytic oxidation of phenol using Ni–Al₂O₃ compositecoating electrodes. J Alloys Compd 477:570–575
- Bejankiwar R, Lalman JA, Seth R, Biswas N (2005) Electrochemical degradation of 1,2-dichloroethane (DCA) in a synthetic groundwater medium using stainless-steel electrodes. Water Res 39:4715–4724
- Allan GG, Peyron M (1995) Molecular weight manipulation of chitosan I: kinetics of depolymerization by nitrous acid. Carbohydr Res 277:257–272
- Tran LH, Drogui P, Mercier G, Blais JF (2009) Electrochemical degradation of polycyclic aromatic hydrocarbon in creosote solution using ruthenium oxide on titanium expanded mesh anode. J Hazard Mater 164:1118–1129
- 29. Chen X, Chen G (2006) Anodic oxidation of orange II on Ti/BDD electrode: variable effects. Sep Purif Technol 48:45–49
- Samet Y, Elaoud SC, Ammar S, Abdelhedi R (2006) Electrochemical degradation of 4-chloroguaiacol for wastewater treatment using PbO₂ anodes. J Hazard Mater B 138:614–619
- Aquino Neto S, de Andrade AR (2009) Electrooxidation of glyphosate herbicide at different DSA compositions: pH, concentration and supporting electrolyte effect. Electrochim Acta 54:2039–2045
- Chen RH, Chang JR, Shyur JS (1997) Effects of ultrasonic conditions and storage in acidic solutions on changes in molecular weight and polydispersity of treated chitosan. Carbohydr Res 299:287–294