REVIEW

The pathogenesis and diagnosis of systemic lupus erythematosus: still not resolved

Ole Petter Rekvig · Johan Van der Vlag

Received: 23 October 2013 /Accepted: 1 April 2014 /Published online: 25 April 2014 \oslash Springer-Verlag Berlin Heidelberg 2014

Abstract Systemic lupus erythematosus (SLE) is a systemic autoimmune disease with various clinical manifestations affecting different tissues. A characteristic feature of SLE is the presence of autoantibodies against double-stranded (ds)DNA, histones and nucleosomes, and other chromatin components. SLE is a prototype type III hypersensitivity reaction. Local deposition of anti-nuclear antibodies in complex with released chromatin induces serious inflammatory conditions by activation of the complement system. The severe renal manifestation, lupus nephritis, is classified based on histological findings in renal biopsies. Apoptotic debris, including chromatin, is present in the extracellular matrix and circulation of patients with SLE. This may be due to an aberrant process of apoptosis and/or insufficient clearance of apoptotic cells/chromatin. The non-cleared apoptotic debris may lead to activation of both the innate and adaptive immune systems. In addition, an aberrant presentation of peptides by antigen-presenting cells, disturbed selection processes for lymphocytes, and deregulated lymphocyte responses may be involved in the development of autoimmunity. In the present review, we briefly will summarize current knowledge on the pathogenesis of SLE. We will also critically discuss and challenge central issues that need to be addressed in order to fully understand the pathogenic mechanisms involved in the development of SLE and in order to

This article is a contribution to the special issue on B cell-mediated autoimmune diseases - Guest Editors: Thomas Winkler and Reinhard Voll

O. P. Rekvig (\boxtimes)

Molecular Pathology Research Group, Department of Medical Biology, Faculty of Health Sciences, University of Tromsø, 9037 Tromsø, Norway e-mail: olepr@fagmed.uit.no

J. Van der Vlag (\boxtimes)

have an improved diagnosis for SLE. Disappointingly, in our opinion, there are still more questions than answers for the pathogenesis, diagnosis, and treatment of SLE.

Keywords Systemic lupus erythematosus \cdot Lupus nephritis \cdot Chromatin . Apoptosis . NETs . DNaseI . Renal

Background

Systemic lupus erythematosus (SLE) causes a heavy burden for affected patients and their families, for the society, and for the economy [\[79](#page-8-0)]. Like other chronic inflammatory diseases, SLE is characterized by protracted morbidity and significant mortality. Substantial efforts are needed in order to develop a better insight into the complex molecular disease mechanisms, diagnostic procedures, and in particular development of biomarkers that indicate progression of the disease and also may reflect response to therapy. The research on SLE is intense, and since 1946, ~56,000 papers have been published (search term in Pubmed: Systemic lupus erythematosus), and \sim 140 genes have been associated with SLE (search term in OMIM: Systemic lupus erythematosus). Since 1959, ~34,000 papers have been published on the origin and role of anti-DNA antibodies, which are central in the pathogenesis and diagnosis of SLE (search term in Pubmed: Anti-DNA antibodies). This enormous number of scientific papers, which is still growing, reflects a strong international effort to understand the complex pathogenesis of SLE but indicates also that we still do not fully understand the pathogenesis of the disease. In other words, we do not understand the pathogenesis of autoimmunity in SLE and of its clinical manifestations in various affected tissues. This lack of knowledge and some controversies preclude the development of precise diagnostic tools and in addition precise and causal therapeutic interventions.

Department of Nephrology (480), Radboud University Medical Centre, Geert Grooteplein 10, 6525 GA Nijmegen, The Netherlands e-mail: Johan.vanderVlag@radboudumc.nl

Introducing remaining questions on the role of anti-dsDNA antibodies in SLE

Anti-double-stranded (ds)DNA autoantibodies are the most studied anti-chromatin autoantibodies in SLE. There is still no international consensus that precisely explains why and how the assumed pathogenic anti-dsDNA antibodies are generated in vivo (origin). In particular, what is the source of DNA/ chromatin and how is autoimmunity against DNA/chromatin initiated and developing? There is still debate on the disease mechanisms that causally involve anti-dsDNA antibodies (impact) in the various tissues that can be affected in SLE. Thus, despite tremendous international efforts to understand the nature of SLE and its pathogenic mechanisms, still, our insight is fragmental and elusive [[4,](#page-7-0) [18](#page-7-0), [47,](#page-8-0) [73,](#page-8-0) [74](#page-8-0), [78,](#page-8-0) [84](#page-9-0), [92,](#page-9-0) [96,](#page-9-0) [98](#page-9-0), [101](#page-9-0)]. Additional unanswered questions include the following: what are the characteristics of a pathogenic antidsDNA antibody, are these characteristics related to the mechanisms responsible for their production, and what is the origin and characteristics of the chromatin fragments retained and targeted by anti-dsDNA antibodies in, e.g., the glomeruli in the context of lupus nephritis? In our opinion, answers to these questions will lead to a better insight into the origin and impact of anti-dsDNA antibodies on disease manifestations in SLE and lupus nephritis, which may in a broader sense be translated into fundamental aspects of causal treatment strategies.

In summary, the pathogenic and diagnostic value of antidsDNA antibodies may be dictated by the mechanism from which they originate (transient or sustained) and, secondly, by the mechanism from which the target antigen originates (transient or sustained). In subsequent paragraphs, we will discuss some of the biochemical processes that may induce autoimmunity and ultimately transform a well functional kidney into a destroyed organ.

Breaking of tolerance: apoptotic blebs and apoptotic chromatin in SLE

The source of chromatin, the main autoantigen, in SLE is most likely apoptotic and/or necrotic cells, including the neutrophilic extracellular traps (NETs). Apoptosis is involved in normal tissue homeostasis and is essential in the regulation of the immune response by central deletion of autoreactive B cells and T cells. Apoptosis can be induced by intrinsic and extrinsic factors, for example by DNA damage and by Fas ligand binding to the Fas receptor. Characteristic for apoptosis is the fragmentation of chromatin and the segregation of apoptotic blebs [\[37](#page-8-0)]. Notably, except for the NETs, we do not know the exact origin of apoptotic blebs and chromatin. Importantly, apoptotic blebs contain autoantigens targeted in SLE [[17](#page-7-0)]. Many factors involved in apoptosis have been

associated to SLE. Autoantigens can be modified during apoptosis, whereby these autoantigen modifications may facilitate the initial breaching of tolerance [[26,](#page-7-0) [37](#page-8-0), [88](#page-9-0)]. Autoantigens in SLE can be cleaved by caspases and endonucleases, like DNaseI as will be detailed below [\[16,](#page-7-0) [122](#page-9-0)]. In addition, autoantigens, including chromatin, may be modified through the addition of acetyl, phosphoryl, methyl, ubiquitin, citruline, ADP, or glutamine moieties. Autoantibodies against aforementioned modifications are present in patients with SLE [\[27](#page-7-0), [88,](#page-9-0) [121](#page-9-0)]. Notably, we have identified specific hyperacetylation patterns on histone H2A, H2B, and H4, as well as a specific methylation pattern on H3 to be associated with apoptosis and SLE [\[24,](#page-7-0) [95](#page-9-0), [123](#page-9-0)–[125\]](#page-10-0). In summary, several factors related to apoptosis, including apoptosisinduced chromatin modification, have been associated with SLE.

Breaking of tolerance: clearance defects in SLE

Apoptosis and necrosis explain how normally inaccessible autoantigens can be released and subsequently become exposed to the immune system. In addition, an impaired removal may lead to further accumulation of apoptotic cells and debris. Normally, apoptotic cells are swiftly removed by professional phagocytes, including macrophages, B cells, and dendritic cells. Removal of apoptotic cells can occur in a non-inflammatory and antiinflammatory manner. However, exposure of apoptotic debris, including chromatin, to dendritic cells may also result in a pro-inflammatory response. As mentioned, apoptotic blebs contain clustered SLE autoantigens, like (modified) chromatin [\[17](#page-7-0)]. In addition to apoptotic blebs, NETs can be considered as apoptotic chromatin. Released apoptotic chromatin autoantigens have a dual function: they may lead not only to the induction of autoimmunity but also to the formation of immune complexes in tissues affected by SLE. In the kidney for example, these immune complexes can deposit in the glomerular capillary filter, thereby inciting a severe glomerulonephritis by activation of the complement system. There is convincing evidence for clearance defects of apoptotic cells and debris in SLE, and actually, defects in many factors required for a proper clearance have been described [[25](#page-7-0), [37,](#page-8-0) [45](#page-8-0), [87,](#page-9-0) [88](#page-9-0), [103\]](#page-9-0). Actually, the clearance of apoptotic material by phagocytes is impaired in both lupus mice and patients [[51](#page-8-0), [72](#page-8-0)]. Downregulation of the expression/activity of the endonuclease DNaseI in the kidney further contributes to the persistence of extracellular apoptotic chromatin and to the development of lupus nephritis as will be further detailed below [[105\]](#page-9-0).

Chromatin fragments play a central role in the pathogenesis of lupus nephritis and persist due to silencing of the renal DNaseI gene

We described that early phases of lupus nephritis in lupusprone (NZBxNZW)F1 mice were characterized by chromatin-IgG complex deposition in the mesangial matrix. A striking observation was that this phenomenon correlated with the early detection of serum antibodies to dsDNA and with mild or clinically silent nephritis [\[30](#page-7-0)]. In analogous experiments, injection of monoclonal anti-dsDNA antibodies into BALB/c mice imposed mesangial nephritis [[31](#page-7-0)]. These spontaneous and experimental events were restricted to mesangial nephritis, and for the spontaneously occurring process, this was always preceded by progression of lupus nephritis into endstage organ disease [[30,](#page-7-0) [105](#page-9-0), [107](#page-9-0)]. Apparently, as a consequence of this early lupus nephritis, renal DNaseI mRNA expression levels and enzyme activity were severely (>80 %) reduced. Reduced levels of renal DNaseI were associated in time with deficient fragmentation of chromatin from dead cells. Large chromatin fragments were retained and accumulated in the glomerular basement membrane (GBM). These observations may in fact explain the basis for deposition of chromatin-IgG complexes in glomeruli in early and late stages of nephritis, leading to complement activation and ultimately loss of glomerular integrity and renal failure.

Acquired error of renal chromatin metabolism—a conditio sine qua non for progressive lupus nephritis

As mentioned, lupus nephritis is a prototype immune complex disease where antibodies to dsDNA play a central role [[47,](#page-8-0) [60](#page-8-0)]. Deposition of chromatin-anti-dsDNA antibody complexes is the core factor that imposes renal inflammation in both murine [\[9](#page-7-0), [30](#page-7-0), [62,](#page-8-0) [63](#page-8-0), [127](#page-10-0), [128\]](#page-10-0) and human SLE [\[50](#page-8-0), [61,](#page-8-0) [117](#page-9-0), [126\]](#page-10-0). This is in harmony with observations that most antibodies eluted from nephritic kidneys contain IgG antibodies reactive with components of chromatin, like nucleosomes, dsDNA, and histones [\[19,](#page-7-0) [63,](#page-8-0) [130](#page-10-0), [138,](#page-10-0) [139,](#page-10-0) [141](#page-10-0)]. However, the picture is not quite clear, as eluted antibodies aside from targeting components of chromatin also have the potential to recognize non-chromatin antigens [[1,](#page-7-0) [21,](#page-7-0) [66](#page-8-0), [130](#page-10-0), [141\]](#page-10-0).

Role of chromatin fragments in lupus nephritis

The origin of chromatin bound to glomerular matrices and membranes has for a long time been unknown, as has been the factors that account for progression of lupus nephritis. Recent data have contributed significantly to new insight into these problems. Current results from our studies on the etiology of

murine and human lupus nephritis demonstrate that renal DNaseI, representing >80 % of total endonuclease activity in the kidneys [\[7](#page-7-0)], is profoundly downregulated when mild or clinically silent mesangial nephritis is transformed into endstage organ disease in human SLE [[105](#page-9-0), [107](#page-9-0), [142\]](#page-10-0). With low DNaseI enzyme activity, apoptotic chromatin is not appropriately fragmented and is instead transformed into secondary necrotic chromatin unmasked from apoptotic blebs (reviewed in [\[83,](#page-9-0) [84](#page-9-0)], see also [[8,](#page-7-0) [126,](#page-10-0) [127](#page-10-0)]). Secondary to this, chromatin is exposed to the environment where it binds the GBM and the mesangial matrix at high affinity as is demonstrated in vitro by plasmon surface resonance analyses [\[81\]](#page-9-0). In those experiments, we demonstrated that chromatin fragments bound collagen IV and laminins at robust affinities, while the proteoglycan perlecan did not [\[49](#page-8-0), [81](#page-9-0)]. This has been observed in both murine and human lupus nephritis [[61](#page-8-0), [62,](#page-8-0) [80\]](#page-8-0).

Transcriptional interference—a possible explanation for the silencing of renal DNaseI gene expression

The term "transcriptional interference" is widely used but poorly defined in the literature [[108](#page-9-0)]. Transcriptional interference usually refers to the direct negative impact of transcription of one gene on transcription of another gene provided the genes are transcribed in opposite directions and that the two genes overlap with each other. Transcriptional interference is potentially widespread throughout biology; therefore, it is timely to assess exactly its nature, significance, and operative mechanisms especially in clinical medicine. A bioinformatics analysis using the UCSC browser has led to a working hypothesis based on transcriptional interference between the DNaseI gene and a convergently transcribed gene—Trap 1. We hypothesize that this mechanism affects inversely DNaseI and Trap 1 gene expression in vivo.

This model also explains what happens if transcription of one of the gene pairs is initiated. In that situation, transcription of the convergent gene is blocked by the transcriptional activity of the first gene [\[52](#page-8-0)]. These results provide insight into fundamental mechanisms of gene expression control and point to an unexplored effect of antisense transcription on gene regulation via polymerase collision. This model is also valid if the genes overlap in the untranslated 3′ regions (UTR) since the primary transcript is elongated far beyond UTR [\[109,](#page-9-0) [115](#page-9-0)].

In summary, both experimental and descriptive data in clinical diseases are consistent and demonstrate that transcriptional interference between convergent and overlapping pair of genes may be a new principle for gene regulation. To establish that the Trap 1 and DNaseI gene are mutually regulated by this mechanism may provide important information on impact of the two genes on prognosis and therapy response and how to control their expression. Details on studies on transcriptional interference between DNaseI and Trap 1 have recently been reviewed [[33,](#page-7-0) [106](#page-9-0)].

The role of dendritic cells in SLE

In SLE, two main subsets of dendritic cells have been implicated to contribute to autoimmunity, i.e., myeloid dendritic cells (mDC) and plasmacytoid dendritic cells (pDC). pDC and mDC differ in their lineage and expression of receptors, including the Toll-like receptors. Macrophages can ingest apoptotic cells, blebs, and debris in an anti-inflammatory manner, which is characterized by the production of TGF-β and interleukin (IL)-10. In addition, dendritic cells encountering autoantigens without being activated will induce immunological tolerance. However, the immunological balance is skewed towards autoimmunity in SLE. We and other researchers demonstrated that mDC can be activated by apoptotic blebs and modified chromatin, by showing an increased expression of co-stimulatory molecules (CD86 and CD40) and increased secretion of pro-inflammatory cytokines (IL-1β, IL-6, and tumor necrosis factor (TNF)-α [[12,](#page-7-0) [13](#page-7-0), [35,](#page-8-0) [36\]](#page-8-0). Other studies showed that high-mobility group protein B1 (HMGB1) which is attached to apoptotic chromatin leads to the activation of mDC via TLR2 [[120](#page-9-0)]. Presentation by activated mDC of the ingested modified chromatin to autoreactive T cells may be an early step in breaking the immunological tolerance that may occur in patients with SLE. Co-cultures of mDC matured with apoptotic blebs and T cells produce IL-2, interferon (IFN)-γ, and IL-17, which suggests a mixed Th1/ Th17 response, as has been shown for patients with SLE. Importantly, IL-6 that is produced by activated mDC is increased in patients with SLE, which inhibits the development of regulatory T cells (T_{REG}) and stimulates the development of Th17 cells. Th17 cells activate autoreactive B cells and recruit inflammatory cells to specific organs [[40\]](#page-8-0). Activated autoreactive T cells, specific for apoptosis-modified histone peptides, can also activate B cells which recognize either modified or unmodified parts of chromatin with their receptor, which results in the production of autoantibodies directed to modified and unmodified chromatin (DNA, histones, nucleosomes) via epitope spreading. After the formation of autoantibodies, immune complexes with circulating chromatin are formed that can activate mDC, thereby creating an amplification loop in the immune response against apoptotic chromatin. Immune complexes also can activate pDC via ligation of TLR7 and TLR9, thereby initiating the production of type I IFN, with IFN- α as the central cytokine. In addition to nucleic acid containing immune complexes, NETs also specifically trigger pDC to produce IFN- α [[13](#page-7-0), [39](#page-8-0), [65](#page-8-0), [69](#page-8-0)]. In patients with SLE, a type I IFN response is frequently observed, suggesting that pDC activation is a central event in the

pathogenesis of SLE. IFN- α has several effector functions, such as facilitation of mDC maturation, B cell activation, T cell activation, and stimulation of NETosis, thereby amplifying the autoimmune response against chromatin [\[102](#page-9-0)]. In summary, both mDC and pDC play central roles in early events and amplifying events leading to autoimmunity (see Fig. [1](#page-4-0) for an integrated hypothesis). The two pathways of immune activation via mDC and pDC are central events; however, additional tolerance-breaching mechanisms in SLE include (i) direct activation of autoreactive B cells by (apoptotic) chromatin; (ii) aberrant presentation of selfpeptides by antigen-presenting cells; (iii) defects in the central selection processes for B and T cells; and (iv) defects in the regulatory processes of B and T cell responses, including cytokine regulation [\[46,](#page-8-0) [73,](#page-8-0) [119](#page-9-0)].

Anti-dsDNA antibodies as diagnostic criterion in SLE: critical remarks

As outlined above, SLE is regarded as a systemic autoimmune syndrome [[78,](#page-8-0) [91](#page-9-0), [98](#page-9-0), [135\]](#page-10-0). B cell and T cell autoimmunity to chromatin and particularly to the dominant individual components of nucleosomes that is native dsDNA and histones are important in establishing a diagnosis [\[34,](#page-8-0) [47](#page-8-0), [116](#page-9-0)]. The importance of anti-chromatin antibodies is further underscored by the fact that anti-chromatin, including anti-dsDNA, antibodies have the potential to induce lupus nephritis in SLE [\[28,](#page-7-0) [54,](#page-8-0) [55](#page-8-0), [126,](#page-10-0) [131](#page-10-0)], while the etiology of other clinical manifestations in SLE is largely unknown [\[56](#page-8-0)]. Related to the latter remark, it can be theoretically questioned whether SLE represents one disease entity or is represented by a continuous overlap of etiologically unrelated organ manifestations. The American College of Rheumatology (ACR) classification criteria for SLE [[116](#page-9-0)] do not answer this question, neither do the newly defined Systemic Lupus International Collaborating Clinics Classification Criteria for Systemic Lupus Erythematosus (SLICC) criteria [[91,](#page-9-0) [135\]](#page-10-0). In both the ACR and SLICC classification criteria, the acceptance and inclusion of each criterion are set partly by consensus among experienced clinicians and scientists, partly by statistics, based on statistical co-appearance of organ and laboratory manifestations but also by insight into disease mechanisms for individual organ manifestations. The term "SLE" may, therefore, still theoretically represent an artificial common denominator for a wide variety of intrinsically unrelated disease manifestations. Are we not in fact directing treatments towards major manifestations of the disease? In a provocative way, one can state that there are no established diagnostic criteria for SLE, although the SLICC criteria may be used also to settle the diagnosis SLE. Consequently, two problems concerning the validity of the diagnosis and treatment of SLE can be defined. First, considering the different and complex faces of SLE, it seems not logic to

Fig. 1 Integrated hypothesis for the pathogenesis of SLE and lupus nephritis. (1) Apoptotic blebs and chromatin are ingested by immature myeloid dendritic cells (mDC), which, thereby, (2) are matured and present (apoptosis-modified) chromatin in their MHC to autoreactive T cells. (3) Activated autoreactive T cells assist autoreactive B cells to produce autoantibodies directed against chromatin. (4) Immune complexes between autoantibodies and chromatin are formed. (5A) Immune complexes are ingested by plasmacytoid dendritic cells (pDC) , which thereby are activated and (5B) start to produce type I interferons, including IFN- α . (5C) IFN- α

primes neutrophils, (6A, 6B) autoantibodies against chromatin, and NETassociated proteins LL37 and HNP induce NETosis, which is normally triggered by microbes (7). (8) Chromatin and NET-associated proteins (LL37, HNP) spewed into the extracellular space function as autoantigens for the B cell, which leads to anti-LL37, anti-HNP, and anti-chromatin autoantibodies that may form immune complexes with NET (9), thereby facilitating their uptake by $pDC(10)$. This establishes a loop between pDC and neutrophils that chronify and/or exacerbate the autoimmune response and the inflammatory condition in SLE. Adapted from [[106\]](#page-9-0)

correlate single distinct biomarkers, like anti-dsDNA, with the whole syndrome instead of correlating them with individual clinical manifestations within the syndrome. Secondly, for treatment, the opposite holds true, namely treating defined manifestations and not the syndrome as that seems not logic. In the context of the previous reasoning, it is particularly a problem when "biomarkers" in SLE are analyzed and tried to be associated with SLE per se. Principally, it is not possible to link a given "biomarker" or single genes or several sets of genes to a syndrome that is regarded a heterogeneous collection of organ manifestations or to panels of laboratorydetermined biological parameters. In our opinion, this should only apply to single manifestations.

To illustrate our reasoning, we will now further focus on antibodies to dsDNA. According to current knowledge, antibodies to dsDNA are directly involved in lupus nephritis [[5,](#page-7-0) [30,](#page-7-0) [47](#page-8-0), [106](#page-9-0), [132](#page-10-0)] and lupus dermatitis [\[32](#page-7-0), [45](#page-8-0)] and possibly also involved in certain aspects of cerebral lupus ([[53](#page-8-0)], discussed in [\[4,](#page-7-0) [23\]](#page-7-0)). This makes up 3 criteria of a total of 11 that classify SLE (ACR criteria). In the remaining eight

criteria, the (pathogenic) role of anti-dsDNA antibodies is not demonstrated. This simple reasoning may suggest that SLE classified, and diagnosed, by combinations of these remaining eight criteria may in fact reduce the impact of anti-dsDNA antibodies as marker antibodies for SLE as a syndrome. Thus, to correlate the anti-dsDNA antibody population with SLE per se may at the best be a biased approach since the impact of the antibody in most of these eight criteria is elusive. How then can this antibody remain a criterion for SLE with no further distinction?

There is no unifying definition of the term anti-dsDNA antibodies in SLE

In the context of the discussion in the previous paragraph, it is important to note that anti-dsDNA antibodies are not representing a homogenous antibody population [[48](#page-8-0), [59,](#page-8-0) [64,](#page-8-0) [113](#page-9-0)], both with respect to their molecular and structural specificities. Since nucleosomal DNA is tightly wrapped around

the histone core, DNA in nucleosomes possesses a highly bent structure, in addition to a more extended (linear-shaped) structure in the linker region. The overall twist of nucleosomal DNA is only 10.2 bp/turn, varying from a value of 9.4 to 10.9 bp/turn [[137](#page-10-0)]. This implies that DNA may be targeted by B cells and consequently by antibodies in two ways, either by binding an elongated DNA structure or by binding the (conformational) bent DNA structure. This may explain the fact that many antibodies bind DNA in ELISA, while fewer bind in the Crithidia luciliae assay (see, e.g., [\[48](#page-8-0)]). This is in harmony with the fact that the *Crithidia* kinetoplast DNA has one of the greatest known degrees of stable curvature [\[44\]](#page-8-0). Other specificities of anti-DNA antibodies include nucleotide sequences, synthetic polynucleotides, and Z DNA [[14,](#page-7-0) [112,](#page-9-0) [113\]](#page-9-0).

Of particular interest in this context is the fact that the mechanisms accounting for production of anti-dsDNA antibodies is highly diverse; some are sustained, and some are transient (see below). Even though, the anti-dsDNA antibodies are regarded specific (as a biomarker) for SLE [[47,](#page-8-0) [59](#page-8-0), [75,](#page-8-0) [112\]](#page-9-0), and the ACR [[116](#page-9-0)] and SLICC [\[91\]](#page-9-0) criteria for the disease include this antibody population as a diagnostic criterion by stating that this criterion is validated by detection of the antibodies by any assay at abnormal titers. Important variables are not included in diagnostic testing for antidsDNA, including antibody characteristics like avidity, whether produced transiently or sustained, whether crossreactive or specific for defined chromatin structures. The SLICC criteria ask for a stringent cutoff value in ELISA to link anti-dsDNA antibodies to SLE by stating: "The new antidsDNA antibody criterion, however, requires a stricter cut-off for ELISA assays." Both the ACR and the SLICC recommendations for testing for anti-dsDNA antibodies may be poor approaches, as they do not consider basic knowledge on the highly diverse origins of anti-dsDNA antibodies. Thus, the validation of anti-dsDNA antibodies in both classification systems may represent an oversimplification and an overestimation of the classification, and pathogenic and diagnostic impact of these antibodies.

With the development of new insight into their factual and plural genesis, it is even tentatively hard to accept a strong relationship between the anti-dsDNA antibody and SLE as a syndrome. Antibodies that phenomenologically bind dsDNA as demonstrated by any assay (ACR criteria) or at a certain magnitude (SLICC criteria) may be produced in the context of several quite different mechanisms, some driven by non-self DNA antigens like bacterial DNA, some by cross-reactive antigens, and some by pure autologous autoantigens. Published mechanisms involve processes linked to (i) acquired, infection-related, or true autoimmune mechanisms similar to hapten-carrier systems, where, e.g., dsDNA or pure nucleosomes represent the principally non-immunogenic hapten-like ligand and the DNA-binding protein (like a DNA-binding

viral protein) represents the carrier protein presented to nontolerant T cells [\[6](#page-7-0), [15,](#page-7-0) [22,](#page-7-0) [41,](#page-8-0) [42](#page-8-0), [71](#page-8-0), [76,](#page-8-0) [82,](#page-9-0) [100,](#page-9-0) [114,](#page-9-0) [133,](#page-10-0) [134\]](#page-10-0) (Fig. [2a\)](#page-6-0); (ii) molecular mimicry [\[2,](#page-7-0) [20](#page-7-0), [68](#page-8-0), [85](#page-9-0), [89,](#page-9-0) [90,](#page-9-0) [97,](#page-9-0) [99\]](#page-9-0); (iii) single gene defects or mutations [[10](#page-7-0), [11,](#page-7-0) [29,](#page-7-0) [43,](#page-8-0) [57,](#page-8-0) [58,](#page-8-0) [136\]](#page-10-0); (iv) translocation of a cluster of X-linked genes onto the Y chromosome creating the genetic lesion underlying Yaa [[29](#page-7-0), [94](#page-9-0)]; (v) the stimulation by non-self DNA like bacterial DNA [\[42](#page-8-0), [93,](#page-9-0) [140\]](#page-10-0); and finally (vi) the stimulatory effect of apoptotic and secondary necrotic cell debris like nucleo-somes on the immune system in vivo [\[8,](#page-7-0) [25,](#page-7-0) [26,](#page-7-0) [38](#page-8-0), [67](#page-8-0), [120](#page-9-0)] (Fig. [2b](#page-6-0)). In this latter situation, impaired removal of apoptotic cells may account for exposure of secondary necrotic material as outlined in previous sections [\[8](#page-7-0), [67,](#page-8-0) [86,](#page-9-0) [120\]](#page-9-0).

Whether the insight given above on the multiple and diverse mechanisms accounting for production of anti-dsDNA antibodies fits with the idea that anti-dsDNA antibodies per se are associated with SLE is far from logic. One major argument against the general impact of anti-dsDNA antibodies in SLE and lupus nephritis is the simple perception that in some cases, the stimulus is transient and results in poor antibody affinity maturation, like for example for infectious-related haptencarrier-like complexes that can be terminated by the normal functioning immune system. In other cases, the stimulus is sustained allowing maturation into high affinity antibodies [\[110,](#page-9-0) [111,](#page-9-0) [118](#page-9-0)], like for example when there is a sustained reduced clearance of apoptotic cells or in context of gene defects/mutations as outlined above. Thus, the link between anti-dsDNA antibodies as that, without any distinction, with, in our opinion, insufficiently defined syndrome SLE must from theoretical considerations be seriously questioned. Indeed, the assay principle may be decisive for the detection of clinically significant anti-DNA antibodies, as has been indicated by Haugbro et al. [\[48](#page-8-0)]. By means of different assays to analyze the presence of various anti-dsDNA antibodies and using an unbiased clinical approach, it may be possible to identify anti-dsDNA antibody subpopulations that may show a stronger association with certain organ manifestations that really are imposed by organ-specific pathogenic anti-dsDNA antibodies. In order to obtain that insight, a wide variety of assays must be performed in a blinded, prospective way, to identify antibody subsets that may be directly involved in organ manifestations but not linked to, e.g., the syndrome SLE. One study [[3\]](#page-7-0) showed, in a cohort of soldiers, the presence of anti-chromatin antibodies several years before the onset of clinical manifestations. However, these authors in a retrograde approach first identified contemporary SLE patients and then analyzed backwards in time detection of relevant antibodies. In our opinion, it would have been preferred that they screened for early detection of relevant antibodies in unselected serum samples from the whole military biobank. This approach would include serum samples of subjects that may not develop SLE. This would give a more true picture of the predictive role of autoantibodies in clinical

Fig. 2 Cognate interaction of nucleosome-specific B cells and peptidespecific T cells. The figure presents a classical hapten-carrier model to explain sustained production of arrays of anti-nucleosome antibodies. In this model, chromatin constituents play the role as a hapten, while either heterologous (infectious-derived DNA-binding proteins like polyomavirus large T antigen) or homologous (e.g., histone-derived) peptides play the role as carrier proteins. a In the left panel, T cells are primed by polyomavirus T antigen peptides presented by an antigen-presenting cell (APC). Then, these T cells recirculate, and eventually, they bind the same peptides presented by B cells specific for different nucleosome structures. Here, T cell tolerance is intact as T antigen is a non-self viral protein. The immune responses are sustained as long as T antigen is expressed. b In the right panel, the T cells are primed by histone-derived peptides presented

medicine. Till now, this approach has not been performed but must be done in our opinion in order to try to select valid diagnostic assays that leave behind antibodies that are purely stochastic epiphenomena, i.e., those that evidently are not involved in disease pathogenesis, nor has any diagnostic impact.

How do the anti-dsDNA antibodies exert their assumed pathogenic impact?

Aside from the problems linked to pure existence of processes that impose anti-dsDNA antibody production, there is also a yet controversial problem whether, and how, these antibodies exert their pathogenic potential. From the simple statement that individuals de facto produce anti-dsDNA antibodies without having organ manifestations like, e.g., nephritis, this may mean that there exists a selection principal that determines their pathogenicity. One possibility is that only those antibodies that bind inherently expressed glomerular antigens are

by APC. Subsequently, these T cells encounter the same peptides on nucleosome-specific B cells and provide the required help to transform them into an array of nucleosome-reactive antibody secreting plasma cells. In this situation, T cell tolerance to nucleosomal proteins is terminated, and the immune response is truly autoimmune. The immune responses are sustained as long as histone-specific T cell tolerance is not controlled. Both mechanisms may in fact be operational in vivo and account for a wide variety of nucleosome-reactive antibodies. The principal paradigm for the hapten-carrier models presented in this figure is based on strong experimental evidences (see text for details). Thus, the cognate interaction of chromatin-specific B cells and immune or autoimmune peptide-specific T cells may explain the origin of the comprehensive repertoire of chromatin-reactive IgG antibodies in human patients

pathogenic [[1,](#page-7-0) [21,](#page-7-0) [77](#page-8-0), [85](#page-9-0), [104\]](#page-9-0). Alternatively, anti-dsDNA antibodies are pathogenic only when chromatin fragments are exposed in glomeruli [\[62](#page-8-0), [70,](#page-8-0) [83](#page-9-0), [84](#page-9-0), [128,](#page-10-0) [129](#page-10-0)]. This obviously requires that chromatin structures must be retained and exposed in the kidney (discussed in [[84\]](#page-9-0)). Therefore, not even how anti-dsDNA antibodies exert their pathogenic potential is clearly defined. Recently, it has been indicated that antidsDNA antibodies are a nonpathogenic factor in the absence of exposed chromatin, whereas exposed chromatin represents a structural epiphenomenon in the absence of antibodies to dsDNA [\[30,](#page-7-0) [81](#page-9-0)]. In this context, the observation that exposure of chromatin in glomerulus membranes and matrices correlates in lupus nephritis is closely linked to loss of renal DNaseI [\[106\]](#page-9-0).

Concluding remarks

The pathogenesis of SLE involves apoptotic chromatin, clearance defects including downregulation of renal DNaseI, mDC, pDC, and B and T lymphocytes. The role of anti-DNA antibodies as a criterion for the diagnosis of SLE as a syndrome is questioned in this review. Two important questions need to be resolved in order to understand the role of antidsDNA antibodies as a diagnostic tool and as a pathogenic antibody: Are all mechanisms for production of anti-dsDNA antibodies linked to SLE, and what is the mechanism that accounts for, e.g., glomerular exposure of chromatin that can be targeted by anti-dsDNA antibodies? The latter problem is partly solved for the kidney, as it has been demonstrated that an *acquired* silencing of the renal DNaseI enzyme result in impaired chromatin degradation and a consequent retention in the glomerular tissue [30, [105,](#page-9-0) [107,](#page-9-0) [142](#page-10-0), [143\]](#page-10-0). However, whether similar mechanisms are operative in other tissues affected with SLE remains to be established. In conclusion, with respect to the pathogenesis, diagnosis, and treatment of SLE, we still have much more questions than answers.

Acknowledgments Elmar Pieterse is acknowledged for the help in preparing Fig. [1](#page-4-0). We thank Rod Wolstenholme (Faculty of Health Sciences, Uit) for expert help in preparing Fig. [2.](#page-6-0) This study was supported by Northern Norway Regional Health Authority Medical Research Program (Grant nos. SFP-100-04 and SFP-101-04), the University of Tromsø as Milieu (OPR), and the Dutch Arthritis Association (Grant 09-1-308; JvdV).

Conflict of interest The authors declare no conflict of interest.

References

- 1. Amital H, Heilweil M, Ulmansky R et al (2005) Treatment with a laminin-derived peptide suppresses lupus nephritis. J Immunol 175: 5516–5523
- 2. Andrzejewski C Jr, Rauch J, Lafer E et al (1981) Antigen-binding diversity and idiotypic cross-reactions among hybridoma autoantibodies to DNA. J Immunol 126:226–231
- 3. Arbuckle MR, Mcclain MT, Rubertone MV et al (2003) Development of autoantibodies before the clinical onset of systemic lupus erythematosus. N Engl J Med 349:1526–1533
- 4. Ardoin SP, Pisetsky DS (2008) Developments in the scientific understanding of lupus. Arthritis Res Ther 10:218
- 5. Ardoin SP, Pisetsky DS (2008) The role of cell death in the pathogenesis of autoimmune disease: HMGB1 and microparticles as intercellular mediators of inflammation. Mod Rheumatol 18:319–326
- 6. Ash Lerner A, Ginsberg Strauss M, Pewzner Jung Y et al (1997) Expression of an anti-DNA-associated VH gene in immunized and autoimmune mice. J Immunol 159:1508–1519
- 7. Basnakian AG, Apostolov EO, Yin X et al (2005) Cisplatin nephrotoxicity is mediated by deoxyribonuclease I. J Am Soc Nephrol 16:697–702
- 8. Berden JH (2003) Lupus nephritis: consequence of disturbed removal of apoptotic cells? Neth J Med 61:233–238
- 9. Berden JH, Licht R, Van Bruggen MC et al (1999) Role of nucleosomes for induction and glomerular binding of autoantibodies in lupus nephritis. Curr Opin Nephrol Hypertens 8:299–306
- 10. Boackle SA, Holers VM, Chen X et al (2001) Cr2, a candidate gene in the murine Sle1c lupus susceptibility locus, encodes a dysfunctional protein. Immunity 15:775–785
- 11. Bolland S, Yim YS, Tus K et al (2002) Genetic modifiers of systemic lupus erythematosus in FcgammaRIIB(-/-) mice. J Exp Med 195:1167–1174
- 12. Boule MW, Broughton C, Mackay F et al (2004) Toll-like receptor 9 dependent and -independent dendritic cell activation by chromatinimmunoglobulin G complexes. J Exp Med 199:1631–1640
- 13. Bouts YM, Wolthuis DFGJ, Dirkx MFM et al (2012) Apoptosis and NET formation in the pathogenesis of SLE. Autoimmunity 45:597–601
- 14. Brigido MM, Stollar BD (1991) Two induced anti-Z-DNA monoclonal antibodies use VH gene segments related to those of anti-DNA autoantibodies. J Immunol 146:2005–2009
- 15. Carroll P, Stafford D, Schwartz RS et al (1985) Murine monoclonal anti-DNA autoantibodies bind to endogenous bacteria. J Immunol 135:1086–1090
- 16. Casciola-Rosen L, Andrade F, Ulanet D et al (1999) Cleavage by granzyme B is strongly predictive of autoantigen status: Implications for initiation of autoimmunity. J Exp Med 190:815–825
- 17. Casciolarosen LA, Anhalt G, Rosen A (1994) Autoantigens targeted in systemic lupus-erythematosus are clustered in 2 populations of surfacestructures on apoptotic keratinocytes. J Exp Med 179:1317–1330
- 18. Choi J, Kim ST, Craft J (2012) The pathogenesis of systemic lupus erythematosus—an update. Curr Opin Immunol 24:651–657
- 19. Dang H, Harbeck RJ (1982) A comparison of anti-DNA antibodies from serum and kidney eluates of NZB x NZW F1 mice. J Clin Lab Immunol 9:139–145
- 20. Davies JM (1997) Molecular mimicry: can epitope mimicry induce autoimmune disease? Immunol Cell Biol 75:113–126
- 21. Deocharan B, Qing X, Lichauco J et al (2002) Alpha-actinin is a cross-reactive renal target for pathogenic anti-DNA antibodies. J Immunol 168:3072–3078
- 22. Desai DD, Krishnan MR, Swindle JT et al (1993) Antigen-specific induction of antibodies against native mammalian DNA in nonautoimmune mice. J Immunol 151:1614–1626
- 23. Diamond B, Volpe BT (2012) A model for lupus brain disease. Immunol Rev 248:56–67
- 24. Dieker JW, Fransen JH, Van Bavel CC et al (2007) Apoptosisinduced acetylation of histones is pathogenic in systemic lupus erythematosus. Arthritis Rheum 56:1921–1933
- 25. Dieker JW, Van der Vlag J, Berden JH (2004) Deranged removal of apoptotic cells: its role in the genesis of lupus. Nephrol Dial Transplant 19:282–285
- 26. Dieker JW, Van der Vlag J, Berden JH (2002) Triggers for antichromatin autoantibody production in SLE. Lupus 11:856–864
- 27. Doyle HA, Mamula MJ (2005) Posttranslational modifications of self-antigens. Ann N Y Acad Sci 1050:1–9
- 28. Ehrenstein MR, Katz DR, Griffiths MH et al (1995) Human IgG anti-DNA antibodies deposit in kidneys and induce proteinuria in SCID mice. Kidney Int 48:705–711
- 29. Fairhurst AM, Hwang SH, Wang A et al (2008) Yaa autoimmune phenotypes are conferred by overexpression of TLR7. Eur J Immunol 38:1971–1978
- 30. Fenton K, Fismen S, Hedberg A et al (2009) Anti-dsDNA antibodies promote initiation, and acquired loss of renal Dnase1 promotes progression of lupus nephritis in autoimmune (NZBxNZW)F1 mice. PLoS One 4:e8474
- 31. Fenton KA, Tommeras B, Marion TN et al (2010) Pure anti-dsDNA mAbs need chromatin structures to promote glomerular mesangial deposits in BALB/c mice. Autoimmunity 43:179–188
- 32. Fismen S, Hedberg A, Fenton K et al (2009) Circulating chromatinanti-chromatin antibody complexes bind with high affinity to dermo-epidermal structures in murine and human lupus nephritis. Lupus 18:597–607
- 33. Fismen S, Thiyagarajan D, Seredkina N et al. (2013) Impact of the tumor necrosis factor receptor-associated protein 1 (Trap1) on renal DNaseI shutdown and on progression of murine and human lupus nephritis. Am J Pathol 182:688–700
- 34. Foster MH (2007) T cells and B cells in lupus nephritis. Semin Nephrol 27:47–58
- 35. Fransen JH, Hilbrands LB, Jacobs CW et al (2009) Both early and late apoptotic blebs are taken up by DC and induce IL-6 production. Autoimmunity 42:325–327
- 36. Fransen JH, Hilbrands LB, Ruben J et al (2009) Mouse dendritic cells matured by ingestion of apoptotic blebs induce T cells to produce interleukin-17. Arthritis Rheum 60:2304–2313
- 37. Fransen JH, Hilbrands LB, Koeter CM, Berden JH, Van der Vlag J. (2009) The role of apoptosis and removal of apoptotic cells in the genesis of systemic lupus erythematosus. Arch Med Sci 5:S466-S477
- 38. Gaipl US, Sheriff A, Franz S et al (2006) Inefficient clearance of dying cells and autoreactivity. Curr Top Microbiol Immunol 305:161–176
- 39. Garcia-Romo GS, Caielli S, Vega B et al (2011) Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. Sci Transl Med 3:73ra20
- 40. Garrett-Sinha LA, John S, Gaffen SL (2008) IL-17 and the Th17 lineage in systemic lupus erythematosus. Curr Opin Rheumatol 20: 519–525
- 41. Gilkeson GS, Grudier JP, Karounos DG et al (1989) Induction of anti-double stranded DNA antibodies in normal mice by immunization with bacterial DNA. J Immunol 142:1482–1486
- 42. Gilkeson GS, Ruiz P, Howell D et al (1993) Induction of immunemediated glomerulonephritis in normal mice immunized with bacterial DNA. Clin Immunol Immunopathol 68:283–292
- 43. Green MC, Shultz LD (1975) Motheaten, an immunodeficient mutant of the mouse. I. Genetics and pathology. J Hered 66:250–258
- 44. Griffith J, Bleyman M, Rauch CA et al (1986) Visualization of the bent helix in kinetoplast DNA by electron microscopy. Cell 46:717–724
- 45. Grootscholten C, Van Bruggen MC, Van Der Pijl JW et al (2003) Deposition of nucleosomal antigens (histones and DNA) in the epidermal basement membrane in human lupus nephritis. Arthritis Rheum 48:1355–1362
- 46. Guerra SG, Vyse TJ, Cunninghame Graham DS (2012) The genetics of lupus: a functional perspective. Arthritis Res Ther 14:211
- 47. Hahn BH (1998) Antibodies to DNA. N Engl J Med 338:1359–1368
- 48. Haugbro K, Nossent JC, Winkler T et al (2004) Anti-dsDNA antibodies and disease classification in antinuclear antibody positive patients: the role of analytical diversity. Ann Rheum Dis 63:386–394
- 49. Hedberg A, Fismen S, Fenton KA et al (2011) Heparin exerts a dual effect on murine lupus nephritis by enhancing enzymatic chromatin degradation and preventing chromatin binding in glomerular membranes. Arthritis Rheum 63:1065–1075
- 50. Hedberg A, Mortensen ES, Rekvig OP (2011) Chromatin as a target antigen in human and murine lupus nephritis. Arthritis Res Ther 13:214
- 51. Herrmann M, Voll RE, Zoller OM et al (1998) Impaired phagocytosis of apoptotic cell material by monocyte-derived macrophages from patients with systemic lupus erythematosus. Arthritis Rheum 41:1241–1250
- 52. Hobson DJ, Wei W, Steinmetz LM et al (2012) RNA polymerase II collision interrupts convergent transcription. Mol Cell 48:365–374
- 53. Huerta PT, Kowal C, Degiorgio LA et al (2006) Immunity and behavior: antibodies alter emotion. Proc Natl Acad Sci U S A 103:678–683
- 54. Isenberg D, Lesavre P (2007) Lupus nephritis: assessing the evidence, considering the future. Lupus 16:210–211
- 55. Isenberg DA, Ehrenstein MR, Longhurst C et al (1994) The origin, sequence, structure, and consequences of developing anti-DNA antibodies. A human perspective. Arthritis Rheum 37:169–180
- 56. Isenberg DA, Manson JJ, Ehrenstein MR et al (2007) Fifty years of anti-ds DNA antibodies: are we approaching journey's end? Rheumatology (Oxford) 46:1052–1056
- 57. Izui S (1990) Autoimmune accelerating genes, lpr and Yaa, in murine systemic lupus erythematosus. Autoimmunity 6:113–129
- 58. Izui S, Kelley VE, Masuda K et al (1984) Induction of various autoantibodies by mutant gene lpr in several strains of mice. J Immunol 133:227–233
- 59. Jang YJ, Stollar BD (2003) Anti-DNA antibodies: aspects of structure and pathogenicity. Cell Mol Life Sci 60:309–320
- 60. Balow JE, Boumpas DT, Ausin HA (1999) Systemic lupus erythematosus and the kidney. In: Lahiota RG (ed) Systemic lupus erythematosus, 3rd edn. Academic, San Diego, pp 657–685
- 61. Kalaaji M, Fenton KA, Mortensen ES et al (2007) Glomerular apoptotic nucleosomes are central target structures for nephritogenic antibodies in human SLE nephritis. Kidney Int 71:664–672
- 62. Kalaaji M, Mortensen E, Jorgensen L et al (2006) Nephritogenic lupus antibodies recognize glomerular basement membraneassociated chromatin fragments released from apoptotic intraglomerular cells. Am J Pathol 168:1779–1792
- 63. Kalaaji M, Sturfelt G, Mjelle JE et al (2006) Critical comparative analyses of anti-alpha-actinin and glomerulus-bound antibodies in human and murine lupus nephritis. Arthritis Rheum 54:914–926
- 64. Kalsi JK, Martin AC, Hirabayashi Y et al (1996) Functional and modelling studies of the binding of human monoclonal anti-DNA antibodies to DNA. Mol Immunol 33:471–483
- 65. Kaplan MJ (2011) Neutrophils in the pathogenesis and manifestations of SLE. Nat Rev Rheumatol 7:691–699
- 66. Krishnan MR, Wang C, Marion TN (2012) Anti-DNA autoantibodies initiate experimental lupus nephritis by binding directly to the glomerular basement membrane in mice. Kidney Int 82:184–192
- 67. Kruse K, Janko C, Urbonaviciute Vet al (2010) Inefficient clearance of dying cells in patients with SLE: anti-dsDNA autoantibodies, MFG-E8, HMGB-1 and other players. Apoptosis Int J Program Cell Death 15:1098–1113
- 68. Lafer EM, Rauch J, Andrzejewski C Jr et al (1981) Polyspecific monoclonal lupus autoantibodies reactive with both polynucleotides and phospholipids. J Exp Med 153:897–909
- 69. Lande R, Ganguly D, Facchinetti Vet al (2011) Neutrophils activate plasmacytoid dendritic cells by releasing self-DNA-peptide complexes in systemic lupus erythematosus. Sci Transl Med 3:73ra19
- 70. Lefkowith JB, Gilkeson GS (1996) Nephritogenic autoantibodies in lupus: current concepts and continuing controversies. Arthritis Rheum 39:894–903
- 71. Lerner J, Ginsberg M, Marion TN et al (1997) Analysis of B/W-DNA 16 V(H) gene expression following DNA-peptide immunization. Lupus 6:328–329
- 72. Licht R, Dieker JW, Jacobs CW et al (2004) Decreased phagocytosis of apoptotic cells in diseased SLE mice. J Autoimmun 22:139–145
- 73. Liu Z, Davidson A (2012) Taming lupus-a new understanding of pathogenesis is leading to clinical advances. Nat Med 18:871–882
- 74. Luijten RKMC, Fritsch-Stork RD, Bijlsma JWJ et al (2013) The use of glucocorticoids in systemic lupus erythematosus. After 60 years still more an art than science. Autoimmun Rev 12:617–628
- 75. Khalil M, Spatz L, Diamond B (1999) Anti-DNA antibodies. In: Lahita RG (ed) Systemic lupus erythematosus, 3rd edn. Academic, San Diego, pp 197–217
- 76. Madaio MP, Hodder S, Schwartz RS et al (1984) Responsiveness of autoimmune and normal mice to nucleic acid antigens. J Immunol 132:872–876
- 77. Mageed RA, Zack DJ (2002) Cross-reactivity and pathogenicity of anti-DNA autoantibodies in systemic lupus erythematosus. Lupus 11:783–786
- 78. Mak A, Isenberg DA, Lau CS (2013) Global trends, potential mechanisms and early detection of organ damage in SLE. Nat Rev Rheumatol 9:301–310
- 79. Meacock R, Dale N, Harrison MJ (2013) The humanistic and economic burden of systemic lupus erythematosus: a systematic review. Pharmacoeconomics 31:49–61
- 80. Mjelle JE, Kalaaji M, Rekvig OP (2009) Exposure of chromatin and not high affinity for dsDNA determines the nephritogenic impact of anti-dsDNA antibodies in (NZBxNZW)F1 mice. Autoimmunity 42: 104–111
- 81. Mjelle JE, Rekvig OP, Fenton KA (2007) Nucleosomes possess a high affinity for glomerular laminin and collagen IV and bind nephritogenic antibodies in murine lupus-like nephritis. Ann Rheum Dis 66:1661–1668
- 82. Moens U, Seternes OM, Hey AW et al (1995) In vivo expression of a single viral DNA-binding protein generates systemic lupus erythematosus-related autoimmunity to double-stranded DNA and histones. Proc Natl Acad Sci U S A 92:12393–12397
- 83. Mortensen ES, Fenton KA, Rekvig OP (2008) Lupus nephritis: the central role of nucleosomes revealed. Am J Pathol 172:275–283
- 84. Mortensen ES, Rekvig OP (2009) Nephritogenic potential of anti-DNA antibodies against necrotic nucleosomes. J Am Soc Nephrol 20:696–704
- 85. Mostoslavsky G, Fischel R, Yachimovich N et al (2001) Lupus anti-DNA autoantibodies cross-react with a glomerular structural protein: a case for tissue injury by molecular mimicry. Eur J Immunol 31:1221–1227
- 86. Munoz LE, Janko C, Schulze C et al (2010) Autoimmunity and chronic inflammation—two clearance-related steps in the etiopathogenesis of SLE. Autoimmun Rev 10:38–42
- 87. Munoz LE, Lauber K, Schiller M et al (2010) The role of defective clearance of apoptotic cells in systemic autoimmunity. Nat Rev Rheumatol 6:280–289
- 88. Munoz LE, Van Bavel C, Franz S et al (2008) Apoptosis in the pathogenesis of systemic lupus erythematosus. Lupus 17:371–375
- 89. Ohteki T, Hessel A, Bachmann MF et al (1999) Identification of a cross-reactive self ligand in virus-mediated autoimmunity. Eur J Immunol 29:2886–2896
- 90. Oldstone MB (1987) Molecular mimicry and autoimmune disease [published erratum appears in Cell 1987 Dec 4;51(5):878]. Cell 50: 819–820
- 91. Petri M, Orbai AM, Alarcon GS et al (2012) Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. Arthritis Rheum 64:2677–2686
- 92. Pisetsky DS (2012) Antinuclear antibodies in rheumatic disease: a proposal for a function-based classification. Scand J Immunol 76: 223–228
- 93. Pisetsky DS (1997) Specificity and immunochemical properties of antibodies to bacterial DNA. Methods 11:55–61
- 94. Pisitkun P, Deane JA, Difilippantonio MJ et al (2006) Autoreactive B cell responses to RNA-related antigens due to TLR7 gene duplication. Science 312:1669–1672
- 95. Price JV, Tangsombatvisit S, Xu GY et al (2012) On silico peptide microarrays for high-resolution mapping of antibody epitopes and diverse protein-protein interactions. Nat Med 18:1434−1440
- 96. Punaro MG (2013) The treatment of systemic lupus proliferative nephritis. Pediatr Nephrol 28:2069–2078
- 97. Putterman C, Diamond B (1998) Immunization with a peptide surrogate for double-stranded DNA (dsDNA) induces autoantibody production and renal immunoglobulin deposition. J Exp Med 188: 29–38
- 98. Rahman A, Isenberg DA (2008) Systemic lupus erythematosus. N Engl J Med 358:929–939
- 99. Ray SK, Putterman C, Diamond B (1996) Pathogenic autoantibodies are routinely generated during the response to foreign antigen: a paradigm for autoimmune disease. Proc Natl Acad Sci U S A 93:2019–2024
- 100. Rekvig OP, Moens U, Sundsfjord A et al (1997) Experimental expression in mice and spontaneous expression in human SLE of polyomavirus T-antigen. A molecular basis for induction of antibodies to DNA and eukaryotic transcription factors. J Clin Invest 99:2045–2054
- 101. Rekvig OP, Nossent JC (2003) Anti-double-stranded DNA antibodies, nucleosomes, and systemic lupus erythematosus: a time for new paradigms? Arthritis Rheum 48:300–312
- 102. Ronnblom L, Alm GV, Eloranta ML (2009) Type I interferon and lupus. Curr Opin Rheumatol 21:471–477
- 103. Rumore PM, Steinman CR (1990) Endogenous circulating DNA in systemic lupus erythematosus. Occurrence as multimeric complexes bound to histone. J Clin Invest 86:69–74
- 104. Sabbaga J, Line SR, Potocnjak P et al (1989) A murine nephritogenic monoclonal anti-DNA autoantibody binds directly to mouse laminin, the major non-collagenous protein component of the glomerular basement membrane. Eur J Immunol 19:137–143
- 105. Seredkina N, Rekvig OP (2011) Acquired loss of renal nuclease activity is restricted to DNasel and is an organ-selective feature in murine lupus nephritis. Am J Pathol 179:1120–1128
- 106. Seredkina N, Van der Vlag J, Berden J et al (2013) Lupus nephritis: enigmas, conflicting models and an emerging concept. Mol Med 19: 161–169
- 107. Seredkina N, Zykova SN, Rekvig OP (2009) Progression of murine lupus nephritis is linked to acquired renal Dnase1 deficiency and not to up-regulated apoptosis. Am J Pathol 175:97–106
- 108. Shearwin KE, Callen BP, Egan JB (2005) Transcriptional interference—a crash course. Trends Genet 21:339–345
- 109. Shilatifard A (2004) Transcriptional elongation control by RNA polymerase II: a new frontier. Biochim Biophys Acta 1677:79–86
- 110. Shlomchik M, Mascelli M, Shan H et al (1990) Anti-DNA antibodies from autoimmune mice arise by clonal expansion and somatic mutation. J Exp Med 171:265–292
- 111. Shlomchik MJ, Marshak-Rothstein A, Wolfowicz CB et al (1987) The role of clonal selection and somatic mutation in autoimmunity. Nature 328:805–811
- 112. Stollar BD (1986) Antibodies to DNA. CRC Crit Rev Biochem 20: 1–36
- 113. Stollar BD (1989) Immunochemistry of DNA. Int Rev Immunol 5: $1 - 22$
- 114. Sundar K, Jacques S, Gottlieb P et al (2004) Expression of the Epstein-Barr virus nuclear antigen-1 (EBNA-1) in the mouse can elicit the production of anti-dsDNA and anti-Sm antibodies. J Autoimmun 23:127–140
- 115. Svejstrup JQ (2013) RNA polymerase II transcript elongation. Biochim Biophys Acta 1829:1
- 116. Tan EM, Cohen AS, Fries JF et al (1982) The 1982 revised criteria for the classification of systemic lupus erythematosus. Arthritis Rheum 25:1271–1277
- 117. Thiyagarajan D, Fismen S, Seredkina N et al (2012) Silencing of renal DNaseI in murine lupus nephritis imposes exposure of large chromatin fragments and activation of toll like receptors and the Clec4e. PLoS One 7:e34080
- 118. Tillman DM, Jou NT, Hill RJ et al (1992) Both IgM and IgG anti-DNA antibodies are the products of clonally selective B cell stimulation in (NZB x NZW)F1 mice. J Exp Med 176:761–779
- 119. Tsokos GC (2011) Mechanisms of disease systemic lupus erythematosus. N Engl J Med 365:2110–2121
- 120. Urbonaviciute V, Furnrohr BG, Meister S et al (2008) Induction of inflammatory and immune responses by HMGB1-nucleosome complexes: implications for the pathogenesis of SLE. J Exp Med 205:3007–3018
- 121. Utz PJ, Anderson P (1998) Posttranslational protein modifications, apoptosis, and the bypass of tolerance to autoantigens. Arthritis Rheum Us 41:1152–1160
- 122. Utz PJ, Gensler TJ, Anderson P (2000) Death, autoantigen modifications, and tolerance. Arthritis Res 2:101–114
- 123. Van Bavel CC, Dieker J, Muller S et al (2009) Apoptosis-associated acetylation on histone H2B is an epitope for lupus autoantibodies. Mol Immunol 47:511–516
- 124. Van Bavel CC, Dieker JW, Kroeze Yet al (2011) Apoptosis-induced histone H3 methylation is targeted by autoantibodies in systemic lupus erythematosus. Ann Rheum Dis 70:201–207
- 125. Van Bavel CC, Dieker JW, Tamboer WP et al (2010) Lupus-derived monoclonal autoantibodies against apoptotic chromatin recognize acetylated conformational epitopes. Mol Immunol 48:248–256
- 126. Van Bavel CC, Fenton KA, Rekvig OP et al (2008) Glomerular targets of nephritogenic autoantibodies in systemic lupus erythematosus. Arthritis Rheum 58:1892–1899
- 127. Van Bavel CC, Van Der Vlag J, Berden JH (2007) Glomerular binding of anti-dsDNA autoantibodies: the dispute resolved? Kidney Int 71:600–601
- 128. Van Bruggen MC, Kramers C, Berden JH (1996) Autoimmunity against nucleosomes and lupus nephritis. Ann Med Interne Paris 147:485–489
- 129. Van Bruggen MC, Kramers C, Hylkema MN et al (1994) Pathophysiology of lupus nephritis: the role of nucleosomes. Neth J Med 45:273–279
- 130. Van Bruggen MC, Kramers C, Hylkema MN et al (1996) Significance of anti-nuclear and anti-extracellular matrix autoantibodies for albuminuria in murine lupus nephritis; a longitudinal study on plasma and glomerular eluates in MRL/l mice. Clin Exp Immunol 105:132–139
- 131. Van Bruggen MC, Walgreen B, Rijke TP et al (1997) Antigen specificity of anti-nuclear antibodies complexed to nucleosomes determines glomerular basement membrane binding in vivo. Eur J Immunol 27:1564–1569
- 132. Van der Vlag J, Berden JH (2011) Lupus nephritis: role of antinucleosome autoantibodies. Semin Nephrol 31:376–389
- 133. Van Ghelue M, Moens U, Bendiksen S et al (2003) Autoimmunity to nucleosomes related to viral infection: a focus on hapten-carrier complex formation. J Autoimmun 20:171–182
- 134. Voll RE, Roth EA, Girkontaite I et al (1997) Histone-specific Th0 and Th1 clones derived from systemic lupus erythematosus patients induce double-stranded DNA antibody production. Arthritis Rheum 40:2162–2171
- 135. Weening JJ, D'agati VD, Schwartz MM et al (2004) The classification of glomerulonephritis in systemic lupus erythematosus revisited. Kidney Int 65:521–530
- 136. Westhoff CM, Whittier A, Kathol S et al (1997) DNA-binding antibodies from viable motheaten mutant mice: implications for B cell tolerance. J Immunol 159:3024–3033
- 137. Widom J (1992) A relationship between the helical twist of DNA and the ordered positioning of nucleosomes in all eukaryotic cells. Proc Natl Acad Sci U S A 89:1095–1099
- 138. Williams RC Jr, Malone C, Blood B et al (1999) Anti-DNA and anti-nucleosome antibody affinity—a mirror image of lupus nephritis? J Rheumatol 26:331–346
- 139. Winfield JB, Faiferman I, Koffler D (1977) Avidity of anti-DNA antibodies in serum and IgG glomerular eluates from patients with systemic lupus erythematosus. Association of high avidity antinative DNA antibody with glomerulonephritis. J Clin Invest 59:90–96
- 140. Wu ZQ, Drayton D, Pisetsky DS (1997) Specificity and immunochemical properties of antibodies to bacterial DNA in sera of normal human subjects and patients with systemic lupus erythematosus (SLE). Clin Exp Immunol 109:27–31
- 141. Xie C, Liang Z, Chang S et al (2003) Use of a novel elution regimen reveals the dominance of polyreactive antinuclear autoantibodies in lupus kidneys. Arthritis Rheum 48:2343–2352
- 142. Zykova SN, Seredkina N, Benjaminsen J et al (2008) Reduced fragmentation of apoptotic chromatin is associated with nephritis in lupus-prone (NZB x NZW)F(1) mice. Arthritis Rheum 58:813– 825
- 143. Zykova SN, Tveita AA, Rekvig OP (2010) Renal Dnase1 enzyme activity and protein expression is selectively shut down in murine and human membranoproliferative lupus nephritis. PLoS One 5(8):e12096