REVIEW

The immunopathology of human biliary cell epithelium

Ya-Hui Chuang · Ruth Y. Lan · M. Eric Gershwin

Received: 30 April 2009 /Accepted: 2 June 2009 / Published online: 17 June 2009 \circ Springer-Verlag 2009

Abstract Bile ducts lined with biliary epithelial cells, or cholangiocytes, are the main components of the biliary system in liver. Cholangiocytes participate in the production and transport of bile substances, as well as participate in immune responses. Cholangiocytes protect against pathogens by expressing toll-like receptors and antimicrobial peptides; act as antigen-presenting cells by expressing human leukocyte antigen molecules and costimulatory molecules; recruit leukocytes to the target site by expressing adhesion molecules, cytokines, and chemokines; and induce apoptosis of leukocytes to limit the immune responses. Several cholangiopathies result from dysfunctions of the biliary system. They can broadly be divided into autoimmune, genetic, infectious, drug, and ischemicinjury-induced categories. The pathogenesis of many of these cholangiopathies is unclear and treatment is limited. Further understanding of the complexity of the biliary system is critical for medical advancements in this field.

Keywords Biliary epithelium · Cholangiocytes · Cholangiopathy . Immune response

YH Chuang and RY Lan contributed equally to this work.

Y.-H. Chuang

Department of Clinical Laboratory Sciences and Medical Biotechnology, College of Medicine, National Taiwan University, Taipei, Taiwan e-mail: yahuichuang@ntu.edu.tw

R. Y. Lan *:* M. Eric Gershwin (***) Division of Rheumatology, Allergy and Clinical Immunology, University of California at Davis School of Medicine, 451 E. Health Sciences Drive, Suite 6510, Davis 95616 CA, USA e-mail: megershwin@ucdavis.edu

Abbreviations

Introduction

The biliary system can be divided into extrahepatic and intrahepatic components [[1](#page-6-0)]. The extrahepatic biliary tract is composed of the gallbladder, common hepatic duct, common bile duct, and cystic duct [\[2](#page-6-0)] (Fig. [1\)](#page-1-0). Bile canaliculi, the canals of Hering (intrahepatic bile ductules), interlobular bile ducts, intrahepatic bile ducts, and the left and right hepatic bile ducts comprise the intrahepatic biliary tract [\[1](#page-6-0)]. Bile ducts lined with biliary epithelial cells, or cholangiocytes, are the main components of the biliary system. Cholangiocytes play a number of roles in the biliary system, such as contributing from 30% to 40% of total bile secretion, participating in bile acid reabsorption and drug metabolism, and mediating immune responses [\[3](#page-6-0)–[5](#page-6-0)]. They compose

Fig. 1 The biliary system. The biliary tract is a system of ducts which carry bile from the liver to the gallbladder and the intestines. Bile is secreted by the hepatocytes and drains into bile canaliculi, which is connected to interlobular bile ducts via the canals of Hering. From there, bile flows from the intrahepatic bile ducts, which join the left and right hepatic ducts that merge to form the common hepatic duct. Bile in the common hepatic duct leaves the liver and joins the cystic duct that exits from the gallbladder, forming the common bile duct. Eventually, bile from the common bile duct merges with the pancreatic duct, which joins to form the ampulla of Vater that enters the duodenal lumen and aids in digestion. The sphincter of Oddi is located at the end of the ampulla of Vater and controls secretions into the duodenum

approximately 4–5% of liver mass and exhibit a special capacity to proliferate under disease conditions [\[6](#page-6-0), [7](#page-6-0)]. Due to the exposure of the biliary tract to foreign antigens, cholangiocytes are equipped to respond through various immunological pathways. They also interact with a number of leukocytes in various fashions. However, a balance between inflammatory responses and tolerance is a key in mucosal environments. This review will cover the immunology of the cholangiocytes, as well as associated diseases.

Cholangiocytes protect against pathogens

Toll-like receptors

Toll-like receptors (TLRs) are key responders to pathogenassociated molecular patterns. Cholangiocytes from intrahepatic large bile ducts, septal bile ducts, interlobular bile ducts, and bile ductules express TLRs 2, 3, 4, and 5, which bind to ligands, such as bacterial molecules, double-stranded RNA, gram-negative lipopolysaccharide (LPS), and flagellin, respectively [\[8](#page-6-0), [9\]](#page-6-0). Evidence for the maintenance of tolerance by cholangiocytes was demonstrated by the up-regulation of interleukin-1 receptor-associated kinase M (IRAK-M), a negative regulator of TLR signaling, in freshly isolated human intrahepatic biliary epithelial cells upon stimulation with TLR-2 and TLR-4 [[10](#page-6-0)]. In fact, IRAK-M is expressed in cholangiocytes in the absence of TLR stimulation. However, in the presence of inflammatory cytokines, such as interferon γ (IFN- γ), cholangiocytes also activate the proinflammatory nuclear factor κB (NF-κB) pathway [\[8](#page-6-0)]. This points to the requirement of a coordinated inflammatory effort by multiple inflammatory mediators for the elicitation of immune responses in the biliary tract. In contrast, IRAK-M may be up-regulated as a result of natural negative feedback mechanisms upon TLR stimulation, as demonstrated by the increase in IRAK-M expression in TLR-stimulated macrophages [[11](#page-6-0)]. In addition to TLRs, anti-microbial peptides are also crucial in mucosal defense.

Anti-microbial peptides

Defensins and cathelicidin are anti-microbial peptides belonging to the innate immune system [\[12](#page-6-0)]. They protect the mucosal barrier against gram-positive and -negative bacteria, mycobacteria, fungi, and viruses via mechanisms such as the disruption of microbial membranes. However, they also participate in adaptive immunity through the recruitment of CD4⁺ T cells and immature dendritic cells [\[13](#page-6-0)]. Granulocytes express the highest density of defensins; however, cholangiocytes also produce these peptides in basal and diseased states [\[12,](#page-6-0) [14\]](#page-6-0). Specifically, human βdefensin 1 (hBD-1) is diffusely expressed in the cytoplasm of normal intrahepatic bile duct epithelium. However, hBD-2, although undetected in normal bile ducts, is expressed almost exclusively by diseased large bile duct epithelium in the liver. Cultured human cholangiocytes constitutively express hBD-1, while hBD-2 is up-regulated only upon IL-1β or tumor necrosis factor- α (TNF- α) treatment [\[14](#page-6-0)]. Cathelicidin is expressed by normal biliary epithelial cells, as well as hepatocytes. Moreover, bile salts and therapeutic bile salts, including chenodeoxycholic acid and ursodeoxycholic acid (UCDA), enhance cathelicidin expression through the farnesoid X receptor and the vitamin D receptor [\[15\]](#page-6-0). Again, the response of cholangiocytes to inflammatory cytokines reflects the significance of a concerted immune response. The ability of cholangiocytes to interact with the cellular immune arm is evidence of this concerted effort and important for maintaining mucosal integrity.

Cholangiocyte*–*leukocyte interactions

Adhesion molecules

Cholangiocytes interact with members of the immune system in a number of ways, such as the presentation of foreign antigen, as well as the expression of leukocyte adhesion molecules. In basal conditions, normal cholangiocytes express low levels of lymphocyte adhesion molecules such as intercellular adhesion molecule 1 (ICAM-1) and lymphocyte-associated antigen 3 (LFA-3). However, some studies report a lack of specific ICAM-1 immunohistochemical staining in normal livers [\[16](#page-6-0)]. ICAM-1 is critical for leukocyte migration to inflammatory sites. It binds to lymphocyte function-associated antigen-1 (LFA-1) and macrophage antigen-1 expressed on leukocytes such as neutrophils, macrophages, and lymphocytes [[17\]](#page-6-0). Importantly, during an inflammatory response, cholangiocytes upregulate their expression of these molecules. This is demonstrated via the addition of inflammatory cytokines such as TNF- α , IL-1, or IFN- γ to in vitro cultures [\[18](#page-6-0)]. Interlobular cholangiocytes from patients with primary biliary cirrhosis (PBC) exhibit increased expressions of ICAM-1, with a corresponding expression of LFA-1 on infiltrating lymphocytes [\[16](#page-6-0)]. LFA-3 on cholangiocytes can interact with CD2 molecules on cytotoxic and natural killer T cells [[3](#page-6-0)]. Vascular cell adhesion molecule-1, which binds to very late antigen-4 on leukocytes, can also be upregulated in cholangiocytes during inflammation [[18\]](#page-6-0). Overall, the increase in leukocyte adhesion molecules facilitates tissue-specific migration by the slowing down of leukocyte circulation near the damaged epithelium, encouraging trafficking to the target site. The expression of antigen-presenting molecules on cholangiocytes also contributes to the local immune response.

Cytokines and chemokines

Human cholangiocytes constitutively express IL-6, IL-8, and monocyte chemotactic protein-1 (MCP-1) [[19,](#page-6-0) [20](#page-7-0)], which are important chemotactic agents for neutrophils, monocytes, and T cells. IL-6 and MCP-1 are increased upon TLR-4 stimulation with LPS in the absence of inflammatory cytokines. However, IL-8 expression is unaltered by TLR ligation [[19\]](#page-6-0). Cytokines and chemokines secreted by cholangiocytes could recruit and activate immune cells such as T cells, macrophages, and natural killer cells to protect against biliary infection [\[21](#page-7-0)].

Antigen presentation

Human leukocyte antigen (HLA) classes I and II, human MHC molecules, are essential for antigen presentation to $CD8⁺$ and $CD4⁺$ T cells, respectively. In normal livers, HLA class I is expressed at a low frequency on cholangiocytes, while HLA class II molecules are not detected [\[22](#page-7-0)]. However, upon cytomegalovirus infection, HLA class I expression is significantly augmented, reflecting the role of $CD8⁺$ T cells in viral responses [[23\]](#page-7-0). PBC patients exhibit

increased HLA class II expressions on injured cholangiocytes [[24\]](#page-7-0). Despite the expression of HLA molecules on cholangiocytes in an inflammatory environment, costimulation is required for T cell activation. In vitro cultures of cholangiocytes in the presence of proinflammatory cytokines, such as IFN- γ and TNF- α increased the expression of HLA class II. However, the lack of up-regulation of CD80 or CD86 molecules (B7-1 and B7-2, respectively) resulted in the inability of these cholangiocytes to induce T cell responses in culture [\[25](#page-7-0)]. In contrast, B7-2 expression was found in damaged cholangiocytes of PBC and primary sclerosing cholangitis (PSC) patients [[24,](#page-7-0) [26\]](#page-7-0). The expression of antigen-presenting-cell-related molecules on cholangiocytes suggests a capacity for antigen presentation, although limited by the requirement for the expression of costimulatory molecules. Similarly, the induction of apoptosis in infiltrating leukocytes is another method of controlling the inflammatory response in the biliary tract.

Apoptosis

There is controversial evidence concerning the expression of programmed-death (PD) ligands, other members of the B7 family, in cholangiocytes of diseased livers. PD molecules are expressed on leukocytes, which, when ligated, induce apoptosis in the leukocyte and may be another method of limiting the immune response [\[27](#page-7-0), [28](#page-7-0)]. Ligation of TNF-related apoptosis-inducing ligand (TRAIL) receptors such as death receptors 4 and 5 (DR4, DR5) on target cells such as cancer and immune cells has also resulted in apoptosis [\[29](#page-7-0), [30](#page-7-0)]. Although absent in normal conditions, the expression of TRAIL by diseased cholangiocytes in PBC and PSC may be an attempt by cholangiocytes to control the inflammatory responses in these diseases by targeting DR-expressing leukocytes [\[30](#page-7-0)]. Therefore, cholangiocytes are capable of antigen presentation in the appropriate environment, with the requirement of costimulatory molecules, as well as the expression of apoptosis-inducing molecules preventing unwarranted inflammatory responses.

The induction of apoptosis in cholangiocytes by leukocytes is also important for the clearance of pathogens and may play a role in the induction of cholangiocyte damage in biliary diseases. CD40, a member of the TNF receptor superfamily, is virtually undetected in cholangiocytes from normal livers but is up-regulated in inflammatory settings. It binds to the CD40L found on leukocytes such as T cells, B cells, and macrophages. In the case of PBC, infiltrating macrophages and T cells express the CD40L, with a more pronounced expression on macrophages [[31\]](#page-7-0). In the same study, cultured cholangiocytes constitutively express CD40, which, upon ligation, increased expression of FasL and the transcription of NF-κB and activator protein 1. The ligation of CD40 also led to a three- to fourfold increase in the apoptosis of cultured cholangiocytes, which is Fasdependent. Thus, the binding of CD40 on cholangiocytes by inflammatory leukocytes may cause apoptosis, leading perhaps to the clearance of infectious agents. DR5 is constitutively expressed by cholangiocytes in normal conditions. It is significantly up-regulated in cholangiocytes of some biliary diseases, which may result in their apoptosis via ligation by TRAIL expressed by activated infiltrating leukocytes [\[30](#page-7-0)]. Thus, apoptosis is an important mechanism in the control of immune responses and may contribute to cholangiocyte-related diseases. In addition to the biliary ducts, periductal connective tissue that loosely surround bile duct walls containing nerve bundles, lymphatics, and blood vessels also play an important role in the biliary system [[2\]](#page-6-0).

Transport of secretory IgA

The transport of IgA to the bile duct lumen is a critical component of humoral immunity in the biliary tract and other mucosal surfaces. Bile contains approximately twice the concentration of secretory IgA (SIgA) compared to that found in upper intestinal fluid [\[32](#page-7-0)]. IgA found in bile is mainly in the form of SIgA, which is composed of two IgA molecules, a peptide J chain, and a secretory component [\[33\]](#page-7-0). The secretory component (SC), or polymeric immunoglobulin receptor, is mainly expressed on cholangiocytes composing the bile duct walls of auxiliary ducts off the main duct. These molecules are present on the luminal surface, perinuclear spaces, endoplasmic reticulum, and endocytic vesicles of chonlangiocytes, although faint expression can be found on the basolateral membrane [[32](#page-7-0)]. Polymeric IgA produced by plasma cells binds to the SC on the basolateral side of cholangiocytes and is transported to the luminal surface,

Table 1 The immunological roles of cholangiocytes

where the SC is cleaved and secreted along with the polymeric IgA [\[34](#page-7-0)]. SCs act as a protective component against proteolytic digestion of the IgA in the gastrointestinal tract, and its carbohydrate residues anchor the SIgA to mucus [[33\]](#page-7-0). SIgA functions in a number of ways to protect the biliary tract. For example, it can directly bind and neutralize bacterial toxins. Either in a specific or a nonspecific manner, SIgA can bind to bacteria and prevent their adhesion to the mucosal membrane [\[33\]](#page-7-0). Additionally, IgA has been demonstrated to neutralize intracellular microbes and their products during its transit through mucosal epithelium. Immune complexes of IgA and foreign antigen in the lamina propria may also be transported to the lumen via SCs, excreting pathogens to a proteolytic mucosal environment [[35](#page-7-0)]. The immunological roles of cholangiocytes are summarized in Table 1.

Cholangiopathies

There are a number of cholangiopathies involving autoimmune, genetic, infectious, drug-induced and ischemic injury, malignant, and idiopathic etiologies [[36](#page-7-0), [37\]](#page-7-0). Autoimmune cholangiopathies include diseases such as PBC, PSC, autoimmune cholangitis, allograft rejection, and graftversus-host disease (GVHD). Alagille syndrome, cystic fibrosis, and fibropolycystic diseases are primarily considered genetic cholangiopathies. Infectious cholangiopathies include bacterial, fungal, parasitic, and viral cholangitis. Floxuridine-induced cholangiopathy is an example of druginduced cholangiopathy, while post-liver transplantation hepatic artery stenosis represents an ischemic-injury induced cholangiopathy. Other cholangiopathies include cholangiocarcinomas and those with unknown etiologies, such as biliary atresia, sarcoidosis, and idiopathic adult/childhood ductopenia.

TLRs Toll-like receptors, IRAK-M interleukin-1 receptor-associated kinase M, PD programmed death, TRAIL TNF-related apoptosis-inducing ligand, MCP-1 monocyte chemotactic protein-1, ICAM-1 intercellular adhesion molecule 1, LFA-3 lymphocyte-associated antigen 3, VCAM-1 vascular cell adhesion molecule-1, HLA human leukocyte antigen

Autoimmune cholangiopathies

Autoimmune cholangiopathies are characterized by the involvement of autoreactive elements of the immune system in the destruction of the biliary tract. PBC is histologically identified by portal inflammation and leukocyte infiltration to intrahepatic bile ducts, eventually leading to their destruction. The destruction of intrahepatic bile ducts eventually leads to the accumulation of toxins, resulting in liver cirrhosis and failure [\[38](#page-7-0)–[40](#page-7-0)]. A key diagnostic element of PBC is the presence of antimitochondrial antibodies against members of the 2-oxo acid dehydrogenase family, particularly the E2 subunit of the pyruvate dehydrogenase complex [[41\]](#page-7-0). Other unique characteristics of PBC are the high levels of serum IgM and the high prevalence among women in their fifth decade of life [[38\]](#page-7-0). The pathogenesis of PBC is currently unknown; however, the involvement of genetic factors, xenobiotics, and microbes has been suggested [\[41](#page-7-0)–[45](#page-7-0)]. Recently, murine models of PBC have been identified, including those exhibiting regulatory T cell defects [\[46](#page-7-0), [47](#page-7-0)]. The main treatment for PBC is the administration of UCDA, which is efficacious up to 10 years [[38\]](#page-7-0).

PSC is characterized by the chronic destruction of medium to large intrahepatic and extrahepatic bile ducts leading to eventual hepatic cirrhosis or failure [[37\]](#page-7-0). Cholangiocarcinoma is developed in 10–30% of PSC patients [[48\]](#page-7-0). The presence of lymphocytic infiltrations, the association with other autoimmune diseases such as inflammatory bowel disease, and the presence of autoantibodies suggest an autoimmune etiology. Unlike PBC, PSC is a male-dominant disease. Although serum anti-nuclear antibodies, anti-cardiolipin antibodies, anti-smooth-muscle antibodies, anti-thyroid peroxidase antibodies, and rheumatoid factor can be found in PSC patients, atypical perinuclear-staining, anti-neutrophil cytoplasmic antibodies are the most prevalent despite their presence in other autoimmune diseases [\[49](#page-7-0)]. However, there is disagreement as to the autoimmune nature of the disease due to the presence of bacterial and viral antigens and the induction of TLRs by anti-cholangiocyte antibodies [[49\]](#page-7-0).

Transplant complications such as liver allograft rejection and GVHD often result in cholangitis. Acute allograft rejection occurs between 5 and 30 days after liver transplantation and involves inflammation of portal or terminal hepatic veins and the destruction of bile ducts. Inflammatory infiltrates include mainly lymphocytes, but also neutrophils and eosinophils [[50\]](#page-7-0). Chronic allograft rejection (CR) can occur as early as 2 weeks to 2 months after liver transplantation [\[51](#page-7-0)]. In CR, the loss of small bile ducts less than 60 μm in diameter results primarily from the senescence of these cholangiocytes. Interestingly, a compensatory proliferation of cholangiocytes resulting from

injury is not observed [[52\]](#page-7-0). CR also involves damage to the terminal hepatic venules, zone-three hepatocytes, portal tract hepatic arterioles, large perihilar hepatic artery branches, and large perihilar bile ducts [\[51](#page-7-0)].

Hematopoietic stem cell transplantation (HSCT) can often result in GVHD with the targeted destruction of small bile duct epithelium [\[53](#page-8-0)]. Although most cases of HSCT-induced GVHD-cholangitis (L-GVHD) result from allogeneic donors, it has been observed in patients receiving autologous and syngeneic HSCT [\[53](#page-8-0), [54](#page-8-0)]. Acute GVHD usually occurs within 1 month of transplantation and involves the skin, gastrointestinal tract, and liver [\[50](#page-7-0)]. In the liver, destruction of interlobular bile ducts is the primary observation. On the other hand, chronic GVHD that occurs 80–100 days after transplantation is characterized by symptoms similar to those found in autoimmune connective tissue disorders, including Sjögren's syndrome, scleroderma, and cholestatic liver disease. Liver pathology resembles that of acute L-GVHD. T cells are the predominant effector cells inducing apoptosis via perforin and granzyme. Similar to CR, cholangiocyte regeneration is not as frequently observed as in PBC and PSC. L-GVHD eventually leads to bile duct loss and rarely to fibrosis. One major difference between CR and L-GVHD is the lack of damage to the arterial system in the liver [\[50](#page-7-0)]. Although genetic factors may play a role in many of these cholangiopathies, direct genetic involvement has been identified in a number of diseases involving the biliary tract.

Genetic cholangiopathies

Alagille syndrome (AGS) involves mutations in genes of the Notch pathway, in particular, Jagged1 (JAG1) or NOTCH2. Clinical features of AGS include cholestasis, cardiac disease, ocular abnormality, skeletal abnormality, and characteristic facial features [[55\]](#page-8-0). The paucity of intrahepatic bile ducts is the most prominent feature of AGS and is due to defects in the development of the biliary tree, not bile duct destruction [\[56](#page-8-0)]. Similar to HSCT GVHD and liver allograft rejection, cholangiocyte reactivity or proliferation in response to damage is lacking in AGS. This may be due to a defect in hepatobiliary cells in their capacity to differentiate into biliary progenitors [\[56](#page-8-0)].

In cystic fibrosis (CF), mutations in the CFTR gene result in multi-organ diseases. Although lung disease is primarily responsible for morbidity and mortality in CF, liver disease is diagnosed in approximately one third of all CF patients, with a potential for underreporting due to diagnostic difficulty [\[57](#page-8-0)]. The CFTR molecule is critical for the secretion of bicarbonate by cholangiocytes being present on the apical surfaces of biliary epithelium and gallbladder epithelium, while it is absent on hepatocytes.

The dysregulation of biliary secretions results in the production of bile with increased viscosity, which slowly progresses to plugged bile ducts, periportal fibrosis, and multilobular cirrhosis [[37\]](#page-7-0). UCDA treatment has been demonstrated to improve liver biochemistry, liver histology, hepatic excretory function, biliary drainage, and essential fatty acid status [[57\]](#page-8-0). Another group of genetically induced cholangitic diseases is the fibropolycystic disease family. The disease family includes Caroli's disease, congenital hepatic fibrosis, Von Meyenburg complex, autosomal dominant polycystic kidney disease, autosomal recessive polycystic kidney disease, and mesenchymal hamartoma [[58\]](#page-8-0). The common characteristics of these diseases are the dilation of intrahepatic bile ducts and fibrosis. Fibropolycystic diseases have also been described as examples of ductal plate malformation, or the lack of remodeling by ductal plates into tubular bile ducts. For example, Caroli's disease affects the formation of larger bile ducts while the Von Meyenburg complex involves interlobular bile ducts [[59\]](#page-8-0). Another category of biliary diseases is infectious cholangiopathies.

Infectious cholangiopathies

Fungal infections have been associated with sclerosing cholangitis-like symptoms. For example, infection by Cryptococcus neoformans, which can be found in pigeon excrement and soil, resulted in dilation of bilateral intrahepatic bile ducts, mass-like lesions in portal tracts resembling cholangiocellular carcinoma, and biliary ob-struction [\[60](#page-8-0)]. The presence of *C. neoformans* was found in bile cultures and demonstrated in biopsies in areas of granulomatous inflammation and multinucleated giants cells. Candida infections can lead to bead-like deformation of the intra- and extrahepatic bile duct system, resembling secondary sclerosing cholangitis [[61,](#page-8-0) [62](#page-8-0)]. Bacterial infections of the biliary tract can result in biliary obstruction, which may increase ductular pressures, causing the release biliary bacterial contents systemically [[63\]](#page-8-0). Bacteria in the biliary tract can deconjugate bile acids and bilirubin, as well as hydrolyze phospholipids, resulting in epithelial damage and stone/sludge formation.

The biliary tract is also subject to parasitic infections by organisms such as liver flukes and roundworms. Examples of liver flukes include Clonorchis sinensis and Opisthorchis viverrini, while roundworms such as Ascaris lumbricoides can enter the biliary system via the ampulla of Vater [[64](#page-8-0)]. Fluke infections begin with the ingestion of encysted larva, which are released in the stomach, penetrate to the peritoneal cavity, and enter the liver. Once in the liver, these flukes tunnel through the liver parenchyma until they reach the lumen of bile ducts, where they reside for years, feeding on bile. As they mature, liver flukes move from small bile ducts to larger ducts and the gallbladder, causing chronic inflammation resulting in thickened bile duct and gallbladder walls. Ascaris lumbricoides infections result in intrahepatic stones, recurrent pyogenic cholangitis, cirrhosis, cholelithiasis, pancreatitis, and cholangiocarcinoma [\[64\]](#page-8-0).

Drug and ischemic-injury induced cholangiopathy

Drug-induced cholangiopathies often result in cholestasis. These cholestatic diseases may be divided into hepatocellular cholestasis and ductular/ductal cholestasis, as reviewed previously [\[65](#page-8-0)]. Hepatocellular cholestasis refers to dysfunctions in hepatocyte canalicular bile secretion, while ductular/ductal cholestasis results from the obstruction of bile ducts. In hepatocellular cholestasis, canaliculi dilation and the appearance of cytoplasmic granules in hepatocytes may be observed. Additionally, cholestatic hepatitis, a more severe manifestation of hapatocellular cholestatis, hepatocyte necrosis, inflammatory infiltration by mononuclear, polymorphonuclear, and eosinophils can occur. Ductular/ductal cholestasis usually results in inflammatory infiltration of ducts, edema, and cholangiocyte abnormalities. Treatments involving sex hormones, anabolic steroids, and sex hormone antagonists may result in hepatocellular cholestasis without liver necrosis. Drugs such as phenothiazines, anticonvulsants, anti-microbial agents, antirheumatic drugs, anti-thyroid drugs, cardiovascular drugs, anti-cancer drugs, immunosuppressants, and anti-depressants may cause cholestatic hepatitis. Examples of drugs causing ductular/ductal cholestasis include azathioprine (immunosuppressant), barbiturates, clindamycin (antibiotic), phenytoin (antiepileptic), and sulpiride (antipsychotic). Patients undergoing metastatic liver disease that have been treated with hepatic arterial infusion chemotherapy using floxuridine (FUDR) may develop sclerosing cholangitis and intrahepatic or extrahepatic strictures, potentially leading to biliary cirrhosis [\[66\]](#page-8-0). The effects of FUDR may be directly due to drug toxicity or via ischemic mechanisms caused by damage to common bile duct arterioles [\[65](#page-8-0)]. Post-liver transplantation hepatic artery stenosis can also cause bile duct strictures due to bile duct infection, viral infection, ischemic injury, persistent rejection, and blood group incompatibility [[67\]](#page-8-0).

Cholangiopathies with unknown etiologies

Several cholangiopathies present enigmatic etiologies, such as biliary atresia, sarcoidosis, and idiopathic adult ductopenia. Biliary atresia is a neotatal disease characterized by the inflammation and obstruction of the extrahepatic and intrahepatic bile ducts. Chronic biliary inflammation results in bile duct damage and loss and liver fibrosis [[37](#page-7-0)]. Eighty percent of biliary atresia cases in the Western countries are "acquired" after birth, while the remaining cases exhibit the "embryonic" form, which usually is presented with other congenital anomalies [[68](#page-8-0)]. Intrahepatic bile ducts are infiltrated by lymphocytes, macrophages, and eosinophils. Macrophages compose a major fraction of leukocytic infiltrates, and their frequency correlates with poorer prognosis. Several hypotheses have been proposed as to the etiology of the disease, including viral infection, autoimmunity, and genetic mutation (particularly with the embryonic form). Further interdisciplinary efforts would be required to reveal the etiology of this disease in order to provide effective treatments. Similarly, the pathogenesis of sarcoidosis is unknown despite its recognition over a century ago [[69\]](#page-8-0). Sarcoidosis is a systemic granulomatous disease, which affects all organs, with the lungs being the most frequently afflicted. A recent study demonstrated over 20% of 1,436 patients with sarcoidosis exhibited abnormal liver tests [\[70](#page-8-0)]. Of the available biopsy samples, 85% exhibited portal inflammation, 50% showed bile duct depletion, and 26% had signs of cirrhosis. Idiopathic adult ductopenia is a rare disease characterized by the absence of interlobular bile ducts in over 50% of portal tracts, adult onset, normal cholangiograms, a lack of anti-mitochondrial antibodies, and the absence of inflammatory bowel disease [[71](#page-8-0)]. There is a 2:1 male-to-female ratio and the prognosis is variable. Genetic factors have been investigated due to evidence of familial clustering. Liver transplantation, immunosuppression, and UCDA have been successful in various patients; however, significant investigation would be required to identify actual mechanisms of this enigmatic disease [\[70\]](#page-8-0).

Dysfunctions of the biliary system result in a number of devastating diseases. Unfortunately, a large number of etiologies in these diseases remain unclear. Further study of the functions and mechanisms of the biliary system under both normal and disease states will be required for the elucidation of the pathogenesis of these diseases, resulting in improved treatment development strategies.

Conclusion

The biliary system is a complex network of tissues/organs, as well as intestinal flora, coordinating functions such as digestion, detoxification, immunological defense, and elimination of waste [\[72](#page-8-0)]. Disruptions in this delicately balanced system may result in a range of diseases. Therefore, careful and thorough investigations considering the entire biliary system may eventually shed light on many enigmatic cholangiopathies. Although useful, animal models should be carefully considered due to differences in the composition of the biliary tract. Further study of the

heterogeneity of the biliary epithelium is also important due to the specific targeting by various diseases.

Acknowledgments This work was partly supported by a grant from the National Science Council, Taiwan (NSC97-2320-B-002-001-).

References

- 1. Sherlock S, Dooley J (2002) Diseases of the liver and biliary system, 11th edn. Blackwell Science, Malden
- 2. Nakanuma Y, Hoso M, Sanzen T, Sasaki M (1997) Microstructure and development of the normal and pathologic biliary tract in humans, including blood supply. Microsc Res Tech 38:552–570
- 3. Chen XM, O'Hara SP, Larusso NF (2008) The immunobiology of cholangiocytes. Immunol Cell Biol 86:497–505
- 4. Johnson L, Byrne J (2003) Essential medical physiology, 3rd edn. Academic, London
- 5. Joplin R, Kachilele S (2009) Human intrahepatic biliary epithelial cell lineages: studies in vitro. Methods Mol Biol 481:193–206
- 6. Beuers U (2009) Crosstalk of liver, bile ducts and the gut. Clin Rev Allergy Immunol 36:1–3
- 7. Glaser SS, Gaudio E, Miller T, Alvaro D, Alpini G (2009) Cholangiocyte proliferation and liver fibrosis. Expert Rev Mol Med 11:e7
- 8. Harada K, Isse K, Nakanuma Y (2006) Interferon gamma accelerates NF-kappaB activation of biliary epithelial cells induced by Toll-like receptor and ligand interaction. J Clin Pathol 59:184–190
- 9. Takeda K, Kaisho T, Akira S (2003) Toll-like receptors. Annu Rev Immunol 21:335–376
- 10. Harada K, Isse K, Sato Y, Ozaki S, Nakanuma Y (2006) Endotoxin tolerance in human intrahepatic biliary epithelial cells is induced by upregulation of IRAK-M. Liver Int 26:935–942
- 11. Kobayashi K, Hernandez LD, Galan JE, Janeway CA Jr, Medzhitov R, Flavell RA (2002) IRAK-M is a negative regulator of Toll-like receptor signaling. Cell 110:191–202
- 12. Fellermann K, Stange EF (2001) Defensins—innate immunity at the epithelial frontier. Eur J Gastroenterol Hepatol 13:771–776
- 13. Taylor K, Barran PE, Dorin JR (2008) Structure-activity relationships in beta-defensin peptides. Biopolymers 90:1–7
- 14. Harada K, Ohba K, Ozaki S, Isse K, Hirayama T, Wada A, Nakanuma Y (2004) Peptide antibiotic human beta-defensin-1 and -2 contribute to antimicrobial defense of the intrahepatic biliary tree. Hepatology 40:925–932
- 15. D'Aldebert E, Biyeyeme Bi Mve MJ, Mergey M, Wendum D, Firrincieli D, Coilly A, Fouassier L, Corpechot C, Poupon R, Housset C, Chignard N (2009) Bile salts control the antimicrobial peptide cathelicidin through nuclear receptors in the human biliary epithelium. Gastroenterology 136:1435–1443
- 16. Yokomori H, Oda M, Ogi M, Wakabayashi G, Kawachi S, Yoshimura K, Nagai T, Kitajima M, Nomura M, Hibi T (2005) Expression of adhesion molecules on mature cholangiocytes in canal of Hering and bile ductules in wedge biopsy samples of primary biliary cirrhosis. World J Gastroenterol 11:4382–4389
- 17. Yang L, Froio RM, Sciuto TE, Dvorak AM, Alon R, Luscinskas FW (2005) ICAM-1 regulates neutrophil adhesion and transcellular migration of TNF-alpha-activated vascular endothelium under flow. Blood 106:584–592
- 18. Reynoso-Paz S, Coppel RL, Mackay IR, Bass NM, Ansari AA, Gershwin ME (1999) The immunobiology of bile and biliary epithelium. Hepatology 30:351–357
- 19. Yokoyama T, Komori A, Nakamura M, Takii Y, Kamihira T, Shimoda S, Mori T, Fujiwara S, Koyabu M, Taniguchi K, Fujioka

H, Migita K, Yatsuhashi H, Ishibashi H (2006) Human intrahepatic biliary epithelial cells function in innate immunity by producing IL-6 and IL-8 via the TLR4-NF-kappaB and -MAPK signaling pathways. Liver Int 26:467–476

- 20. Morland CM, Fear J, McNab G, Joplin R, Adams DH (1997) Promotion of leukocyte transendothelial cell migration by chemokines derived from human biliary epithelial cells in vitro. Proc Assoc Am Physicians 109:372–382
- 21. Selmi C, Mackay IR, Gershwin ME (2007) The immunological milieu of the liver. Semin Liver Dis 27:129–139
- 22. Dillon PW, Belchis D, Minnick K, Tracy T (1997) Differential expression of the major histocompatibility antigens and ICAM-1 on bile duct epithelial cells in biliary atresia. Tohoku J Exp Med 181:33–40
- 23. Scholz M, Cinatl J, Blaheta RA, Kornhuber B, Markus BH, Doerr HW (1997) Expression of human leukocyte antigens class I and class II on cultured biliary epithelial cells after cytomegalovirus infection. Tissue Antigens 49:640–643
- 24. Tsuneyama K, Harada K, Yasoshima M, Kaji K, Gershwin ME, Nakanuma Y (1998) Expression of co-stimulatory factor B7-2 on the intrahepatic bile ducts in primary biliary cirrhosis and primary sclerosing cholangitis: an immunohistochemical study. J Pathol 186:126–130
- 25. Leon MP, Bassendine MF, Wilson JL, Ali S, Thick M, Kirby JA (1996) Immunogenicity of biliary epithelium: investigation of antigen presentation to CD4+ T cells. Hepatology 24:561–567
- 26. Selmi C, Lleo A, Pasini S, Zuin M, Gershwin ME (2009) Innate immunity and primary biliary cirrhosis. Curr Mol Med 9:45–51
- 27. Oikawa T, Takahashi H, Ishikawa T, Hokari A, Otsuki N, Azuma M, Zeniya M, Tajiri H (2007) Intrahepatic expression of the co-stimulatory molecules programmed death-1, and its ligands in autoimmune liver disease. Pathol Int 57:485–492
- 28. Mataki N, Kikuchi K, Kawai T, Higashiyama M, Okada Y, Kurihara C, Hokari R, Kawaguchi A, Nagao S, Kondo T, Itoh K, Miyakawa H, Miura S (2007) Expression of PD-1, PD-L1, and PD-L2 in the liver in autoimmune liver diseases. Am J Gastroenterol 102:302–312
- 29. Takeda K, Stagg J, Yagita H, Okumura K, Smyth MJ (2007) Targeting death-inducing receptors in cancer therapy. Oncogene 26:3745–3757
- 30. Takeda K, Kojima Y, Ikejima K, Harada K, Yamashina S, Okumura K, Aoyama T, Frese S, Ikeda H, Haynes NM, Cretney E, Yagita H, Sueyoshi N, Sato N, Nakanuma Y, Smyth MJ (2008) Death receptor 5 mediated-apoptosis contributes to cholestatic liver disease. Proc Natl Acad Sci U S A 105: 10895–10900
- 31. Afford SC, Ahmed-Choudhury J, Randhawa S, Russell C, Youster J, Crosby HA, Eliopoulos A, Hubscher SG, Young LS, Adams DH (2001) CD40 activation-induced, Fas-dependent apoptosis and NF-kappaB/AP-1 signaling in human intrahepatic biliary epithelial cells. FASEB J 15:2345–2354
- 32. Nagura H, Smith PD, Nakane PK, Brown WR (1981) IGA in human bile and liver. J Immunol 126:587–595
- 33. Phalipon A, Cardona A, Kraehenbuhl JP, Edelman L, Sansonetti PJ, Corthesy B (2002) Secretory component: a new role in secretory IgA-mediated immune exclusion in vivo. Immunity 17:107–115
- 34. Kaetzel CS, Robinson JK, Chintalacharuvu KR, Vaerman JP, Lamm ME (1991) The polymeric immunoglobulin receptor (secretory component) mediates transport of immune complexes across epithelial cells: a local defense function for IgA. Proc Natl Acad Sci U S A 88:8796–8800
- 35 Lamm M (2008) Protection of mucosal epithelia by IgA: Intracellular neutralization and excretion of antigens. Mucosal immune defense: Immunoglobulin A. Springer, Berlin Heidelberg New York, pp 173–182
- 36. Lazaridis KN, Strazzabosco M, Larusso NF (2004) The cholangiopathies: disorders of biliary epithelia. Gastroenterology 127:1565–1577
- 37. Xia X, Demorrow S, Francis H, Glaser S, Alpini G, Marzioni M, Fava G, Lesage G (2007) Cholangiocyte injury and ductopenic syndromes. Semin Liver Dis 27:401–412
- 38. Kaplan MM, Gershwin ME (2005) Primary biliary cirrhosis. N Engl J Med 353:1261–1273
- 39. Shimoda S, Miyakawa H, Nakamura M, Ishibashi H, Kikuchi K, Kita H, Niiro H, Arinobu Y, Ono N, Mackay IR, Gershwin ME, Akashi K (2008) CD4 T-cell autoreactivity to the mitochondrial autoantigen PDC-E2 in AMA-negative primary biliary cirrhosis. J Autoimmun 31:110–115
- 40. Sasaki M, Ikeda H, Nakanuma Y (2008) Activation of ATM signaling pathway is involved in oxidative stress-induced expression of mito-inhibitory p21WAF1/Cip1 in chronic nonsuppurative destructive cholangitis in primary biliary cirrhosis: an immunohistochemical study. J Autoimmun 31:73–78
- 41. Lan RY, Leung P, Ansari AA, Coppel RL, Gershwin ME (2005) Solving the primary biliary cirrhosis puzzle: The emerging image of immunopathology in primary biliary cirrhosis. Clin Appl Immunol Rev 5:271–284
- 42. Shimoda S, Harada K, Niiro H, Yoshizumi T, Soejima Y, Taketomi A, Maehara Y, Tsuneyama K, Nakamura M, Komori A, Migita K, Nakanuma Y, Ishibashi H, Selmi C, Gershwin ME (2008) Biliary epithelial cells and primary biliary cirrhosis: the role of liver-infiltrating mononuclear cells. Hepatology 47:958– 965
- 43. Allina J, Stanca CM, Garber J, Hu B, Sautes-Fridman C, Bach N, Odin JA (2008) Anti-CD16 autoantibodies and delayed phagocytosis of apoptotic cells in primary biliary cirrhosis. J Autoimmun 30:238–245
- 44. Salunga TL, Cui ZG, Shimoda S, Zheng HC, Nomoto K, Kondo T, Takano Y, Selmi C, Alpini G, Gershwin ME, Tsuneyama K (2007) Oxidative stress-induced apoptosis of bile duct cells in primary biliary cirrhosis. J Autoimmun 29:78–86
- 45. Rieger R, Gershwin ME (2007) The X and why of xenobiotics in primary biliary cirrhosis. J Autoimmun 28:76–84
- 46. Wakabayashi K, Lian ZX, Moritoki Y, Lan RY, Tsuneyama K, Chuang YH, Yang GX, Ridgway W, Ueno Y, Ansari AA, Coppel RL, Mackay IR, Gershwin ME (2006) IL-2 receptor alpha(−/−) mice and the development of primary biliary cirrhosis. Hepatology 44:1240–1249
- 47. Yang GX, Lian ZX, Chuang YH, Moritoki Y, Lan RY, Wakabayashi K, Ansari AA, Flavell RA, Ridgway WM, Coppel RL, Tsuneyama K, Mackay IR, Gershwin ME (2008) Adoptive transfer of CD8(+) T cells from transforming growth factor beta receptor type II (dominant negative form) induces autoimmune cholangitis in mice. Hepatology 47:1974–1982
- 48. Maggs JR, Chapman RW (2008) An update on primary sclerosing cholangitis. Curr Opin Gastroenterol 24:377–383
- 49. Weismuller TJ, Wedemeyer J, Kubicka S, Strassburg CP, Manns MP (2008) The challenges in primary sclerosing cholangitis aetiopathogenesis, autoimmunity, management and malignancy. J Hepatol 48(Suppl 1):S38–S57
- 50. Demetris AJ (1998) Immune cholangitis: liver allograft rejection and graft-versus-host disease. Mayo Clin Proc 73:367–379
- 51. Inomata Y, Tanaka K (2001) Pathogenesis and treatment of bile duct loss after liver transplantation. J Hepatobiliary Pancreat Surg 8:316–322
- 52. Lunz JG 3rd, Contrucci S, Ruppert K, Murase N, Fung JJ, Starzl TE, Demetris AJ (2001) Replicative senescence of biliary epithelial cells precedes bile duct loss in chronic liver allograft rejection: increased expression of p21(WAF1/Cip1) as a disease marker and the influence of immunosuppressive drugs. Am J Pathol 158:1379–1390
- 53. Quaglia A, Duarte R, Patch D, Ngianga-Bakwin K, Dhillon AP (2007) Histopathology of graft versus host disease of the liver. Histopathology 50:727–738
- 54. Saunders MD, Shulman HM, Murakami CS, Chauncey TR, Bensinger WI, McDonald GB (2000) Bile duct apoptosis and cholestasis resembling acute graft-versus-host disease after autologous hematopoietic cell transplantation. Am J Surg Pathol 24:1004–1008
- 55. McDaniell R, Warthen DM, Sanchez-Lara PA, Pai A, Krantz ID, Piccoli DA, Spinner NB (2006) NOTCH2 mutations cause Alagille syndrome, a heterogeneous disorder of the notch signaling pathway. Am J Hum Genet 79:169–173
- 56. Fabris L, Cadamuro M, Guido M, Spirli C, Fiorotto R, Colledan M, Torre G, Alberti D, Sonzogni A, Okolicsanyi L, Strazzabosco M (2007) Analysis of liver repair mechanisms in Alagille syndrome and biliary atresia reveals a role for notch signaling. Am J Pathol 171:641–653
- 57. Colombo C (2007) Liver disease in cystic fibrosis. Curr Opin Pulm Med 13:529–536
- 58. Yonem O, Bayraktar Y (2007) Is portal vein cavernous transformation a component of congenital hepatic fibrosis? World J Gastroenterol 13:1928–1929
- 59. Desmet VJ (1998) Ludwig symposium on biliary disorders—part I. Pathogenesis of ductal plate abnormalities. Mayo Clin Proc 73:80–89
- 60. Nara S, Sano T, Ojima H, Onaya H, Ikeda M, Morizane C, Esaki M, Sakamoto Y, Shimada K, Kosuge T (2008) Liver cryptococcosis manifesting as obstructive jaundice in a young immunocompetent man: report of a case. Surg Today 38:271–274
- 61. Domagk D, Bisping G, Poremba C, Fegeler W, Domschke W, Menzel J (2001) Common bile duct obstruction due to candidiasis. Scand J Gastroenterol 36:444–446
- 62. Domagk D, Fegeler W, Conrad B, Menzel J, Domschke W, Kucharzik T (2006) Biliary tract candidiasis: diagnostic and therapeutic approaches in a case series. Am J Gastroenterol 101:2530–2536
- 63. Hanau LH, Steigbigel NH (2000) Acute (ascending) cholangitis. Infect Dis Clin North Am 14:521–546
- 64. Rana SS, Bhasin DK, Nanda M, Singh K (2007) Parasitic infestations of the biliary tract. Curr Gastroenterol Rep 9:156– 164
- 65. Erlinger S (1997) Drug-induced cholestasis. J Hepatol 26(Suppl 1): $1 - 4$
- 66. Phongkitkarun S, Kobayashi S, Varavithya V, Huang X, Curley SA, Charnsangavej C (2005) Bile duct complications of hepatic arterial infusion chemotherapy evaluated by helical CT. Clin Radiol 60:700–709
- 67. Sidhu PS, Ellis SM, Karani JB, Ryan SM (2002) Hepatic artery stenosis following liver transplantation: significance of the tardus parvus waveform and the role of microbubble contrast media in the detection of a focal stenosis. Clin Radiol 57:789–799
- 68. Mack CL, Sokol RJ (2005) Unraveling the pathogenesis and etiology of biliary atresia. Pediatr Res 57:87R–94R
- 69. Judson MA (2008) Sarcoidosis: clinical presentation, diagnosis, and approach to treatment. Am J Med Sci 335:26–33
- 70. Kahi CJ, Saxena R, Temkit M, Canlas K, Roberts S, Knox K, Wilkes D, Kwo PY (2006) Hepatobiliary disease in sarcoidosis. Sarcoidosis Vasc Diffuse Lung Dis 23:117–123
- 71. Burak KW, Pearson DC, Swain MG, Kelly J, Urbanski SJ, Bridges RJ (2000) Familial idiopathic adulthood ductopenia: a report of five cases in three generations. J Hepatol 32:159– 163
- 72. Fitz JG (2002) Regulation of cholangiocyte secretion. Semin Liver Dis 22:241–249