

ORIGINAL ARTICLE

Barbara A. Conley · Thomas S. Ramsland
Dorothy L. Sentz · Suhlan Wu · D. Marc Rosen
Megan Wollman · Julie L. Eiseman

Antitumor activity, distribution, and metabolism of 13-*cis*-retinoic acid as a single agent or in combination with tamoxifen in established human MCF-7 xenografts in mice

Received: 4 November 1996 / Accepted: 27 July 1998

Abstract Purpose: The efficacy of 13-*cis*-retinoic acid (13-CRA) given as a single agent or in combination with tamoxifen (TAM) was determined in athymic nude mice bearing advanced s.c. MCF-7 human breast cancers. **Methods:** 13-CRA alone was given by gavage at doses ranging from 26.4 to 200 mg/kg. TAM alone was given by gavage at doses of 7.5, 15, 30, or 60 mg/kg. For combination studies, each dose of TAM was followed 4 h later by 13-CRA at doses of 25, 50, 100, or 200 mg/kg. All treatments began on day 12 and were continued for 3 weeks. **Results:** The median time to two doublings recorded for the control and for 13-CRA and TAM given as single agents at the highest dose were 22.2, 29.2, and 54.7 days, respectively. In combination, 100 and 200 mg/kg 13-CRA with 7.5 mg/kg TAM resulted in a delay in tumor growth at least as high as that achieved

with highest-dose TAM alone, but the effect was not synergistic. Pharmacokinetic analysis of 13-CRA was performed in plasma, liver, and tumor from mice bearing 0.5- to 2.0 g carcinomas following a single dose of 100 mg/kg 13-CRA. Results showed that 13-CRA was metabolized differently in various tissues, but concentrations of 13-CRA detected in tumor were in the range reported to be active *in vitro*. *all-trans*-Retinoic acid (ATRA) concentrations were about 5% of the 13-CRA concentrations detected in plasma, 68% of those found in liver, and 20% of those found in tumor. 4-oxo-CRA represented between 2% and 10% of 13-CRA concentrations detected in plasma and liver but was not detected in tumor. Furthermore there was no difference in peak plasma 13-CRA concentrations found in the same tissues at 30 min after a single dose or after the eighth dose of 100 mg/kg 13-CRA or 13-CRA and TAM. Mean 13-CRA concentrations detected in liver and tumor were 50–90% and 16–30% of plasma peak concentrations, respectively. No difference in 4-oxo-CRA concentration was observed between the treatment groups. **Conclusions:** These data suggest that 13-CRA is not effective against established human breast tumor xenografts despite the stability of the pharmacokinetics of 13-CRA and the generation of ATRA as a metabolite. The addition of 13-CRA to TAM did not improve the efficacy of TAM against these estrogen-receptor-positive xenografts.

This work was supported by contract NO1-CM27752, awarded by the National Cancer Institute

J.L. Eiseman (✉)
Division of Developmental Therapeutics,
University of Maryland Cancer Center,
9th Floor, Bressler Research Laboratory Building,
655 W. Baltimore Street,
Baltimore, MD 21201, USA
Tel.: +1-410-328-3685, Fax: +1-410-328-6559

B.A. Conley · T.S. Ramsland · D.L. Sentz · S. Wu ·
D.M. Rosen · M. Wollman · J.L. Eiseman
Division of Developmental Therapeutics,
University of Maryland Cancer Center,
Program of Oncology, Baltimore,
MD 21201, USA

B.A. Conley
Division of Hematology-Oncology,
Department of Medicine,
University of Maryland School of Medicine,
Baltimore, MD 21201, USA

J.L. Eiseman
Department of Pathology,
University of Maryland School of Medicine,
Baltimore, MD 21201, USA

Key words 13-*cis*-Retinoic acid · Tamoxifen ·
MCF-7 xenografts · Breast cancer

Abbreviations 13-CRA 13-*cis*-retinoic acid ·
TAM Tamoxifen · 4-oxo-CRA 4-oxo-13-*cis*-retinoic
acid · ATRA *all-trans*-retinoic acid · ER estrogen
receptor · NCI National Cancer Institute (USA) ·
DTP Developmental Therapeutics Program · RAR
retinoic acid receptor · HPLC high-performance
liquid chromatography · AUC area under the plasma
concentration × time curve

Introduction

Retinoic acid compounds have recently been investigated as anticancer agents for a variety of cancers. Compounds such as *all-trans*-retinoic acid (ATRA) and 13-*cis*-retinoic acid (isotretinoin, 13-CRA) have shown activity against a wide variety of human malignancies [2, 6, 9, 14, 26–29], including nonmelanoma skin cancer, bladder carcinoma, and acute promyelocytic leukemia. In addition, chemopreventive activity of retinoids has been demonstrated for premalignant skin lesions and oral leukoplakia and in the prevention of second primary carcinomas of the upper aerodigestive tract [13, 23, 26].

Numerous studies have suggested that the retinoic acid compounds may be of use in human breast cancer [10, 11, 17–19, 22, 24–26, 30]. *In vitro*, Fontana [10] demonstrated that ATRA at concentrations of 1 μ M effectively inhibited the growth of estrogen-receptor (ER)-positive MCF-7 cells in culture and that the effect of combined treatment with ATRA and TAM was additive. Wetherell and Taylor [30] also showed that ATRA and 13-CRA were effective against the ER-positive human mammary cell line T47D and that cotreatment of the cells with either 13-CRA or ATRA plus TAM produced additive effects. Toma et al. [28] showed inhibitory effects of 13-CRA in MCF-7 cells *in vitro*, but no study has either documented *in vivo* activity of 13-CRA against ER-positive human breast xenografts or measured the concentrations of 13-CRA achieved in animals. However, one study showed that, although 13-CRA inhibited the growth of the ER-negative human breast carcinoma cell line MDA-MB-231 *in vitro*, it was ineffective when given in feed to mice bearing xenografts of the same tumor [11]. 13-CRA was also inactive as a single agent in a clinical trial in breast cancer [5]. For clinical applications, 13-CRA may represent a better candidate than ATRA because the pharmacokinetics of 13-CRA are stable over time, whereas those of ATRA are variable [1, 7, 12, 20, 26]. In the present study we examined the efficacy of 13-CRA both as a single agent and in combination with TAM in athymic nude mice bearing established MCF-7 human breast carcinoma xenografts. We also determined the pharmacokinetics of 13-CRA in plasma, liver, and tumor after the administration of 13-CRA to both fasted and nonfasted tumor-bearing athymic nude mice. Furthermore we examined peak concentrations of 13-CRA in mice after one or eight doses of the agent given alone or in combination with TAM so as to determine whether its metabolism or absorption would be altered by continued and/or combination treatment.

Materials and methods

Mice

Female athymic nude mice (NCr nu nu, 5–6 weeks of age specific pathogen-free) were obtained from the NCI Animal Production

Branch (Frederick, Md.) and were allowed to acclimate to the University of Maryland animal facility for 1 week prior to initiation of the study. Mice were housed in microisolator caging and allowed autoclaved food and water *ad libitum*. Animal rooms were maintained at 25 ± 2 °C on a 12-h light/dark cycle and at least 12 air changes/h. All animals were handled in accordance with the Guide to the Care and Use of Laboratory Animals (National Institutes of Health, 1985) except that the s.c. tumors were allowed to grow to as large as 5 g in the control groups before the study was ended. Monthly analysis of sentinel mice housed in 1/5 dirty bedding from study mice confirmed that the study mice remained MAP (murine antibody profile)-negative.

Tumor

MCF-7 human breast-carcinoma tumor fragments (G00135) were obtained from the Division of Cancer Treatment Tumor Repository, National Cancer Institute (NCI) Frederick Cancer Research Facility (Frederick, Md.) and were implanted s.c. on the right flank. Using metaflane anesthesia, slow-release 1.7-mg 17- β -estradiol pellets (Innovative Research of America, Sarasota, Fla.) were implanted s.c. above the left flank. Tumors used in these studies were harvested from passage mice using aseptic techniques. Fragments of tumor (approximately 25 mg) and estradiol pellets were implanted in study mice on day 0. Tumors were measured twice weekly, and tumor volumes were calculated from the formula $Length \times (width)^2 / 2$, where *length* is the largest dimension and *width*, the smallest dimension perpendicular to the length. Mice were observed until the tumors measured between 22 and 394 mm³ as determined by digital caliper measurement. At that time, 12 to 18 days postimplantation, the mice were stratified into treatment groups of 8 or 10 mice/treatment group and 20 mice/control group such that the mean and median values recorded for the groups were not different from those noted for the other groups in terms of either body weight or tumor volume. For the pharmacokinetics studies, additional mice were implanted with MCF-7 fragments and estradiol pellets. These mice were dosed when the tumor volumes had reached between 300 and 500 mm³ as determined by caliper measurement.

Drugs

13-CRA (Isotretinoin, NSC 329481) was obtained from the Developmental Therapeutics Program (DTP) of the NCI or was purchased from Toronto Research Chemical Inc. (Downsview, Ontario, Canada). ATRA (NSC 122758) was obtained from the DTP, NCI. TAM was purchased from Sigma Chemical Co. (St. Louis, Mo.) as the acetate salt. Mice received 13-CRA and/or TAM by gavage 5 days a week for 3 weeks beginning on day 12 or 18 post tumor implantation. An initial dose-ranging study was performed at doses of 26.4, 39.6, 59.4, 89, 133.2, and 200 mg/kg per dose of 13-CRA, prepared as fine suspensions in cremophor:ethanol:water (1:1:6, by vol.) such that the animals received 0.01 ml/g body weight. For TAM the initial dose-ranging study was performed at doses of 7.5, 15, 30, or 60 mg/kg in sterile water. For the combination studies, doses of 13-CRA were 25, 50, 100, and 200 mg/kg. TAM doses were 7.5, 15, 30, or 60 mg/kg. Doses were given as oral boluses using a 22-gauge 1.5-inch gavage needle and were based on each day's exact body weights. The positive control groups received the vehicle(s) only at 0.01 ml/g body weight. Single agents and combinations were given to cohorts of mice for 3 weeks, 5 days/week. Mice were weighed and observed daily during the dosing period, and tumor measurements were recorded twice weekly. After treatment had been completed, tumor measurements and body weights were recorded twice weekly until the end of the study. Any moribund animals were euthanized by inhalation of CO₂. Gross necropsies were conducted on all mice.

Pharmacokinetics studies

Mice bearing s.c. MCF-7 tumors ranging in volume between 500 and 2000 mm³ were fasted overnight prior to dosing. 13-CRA at

100 mg/kg in cremophor:ethanol:water (1:1:6, by vol.) was given by oral gavage. Two mice from each group were euthanized with CO₂, and blood was collected by cardiac puncture using heparinized syringes at the following times after dosing: 5, 15, 30, 45, 60, 90, 120, 180, 240, 360, 480, and 960 min. For comparison, mice bearing s.c. MCF-7 xenografts were given 13-CRA without prior fasting to determine if there were differences in 13-CRA absorption between fasting and fed mice. These animals were killed at 5, 10, 15, 30, 45, 60, 90, 120, 180, 240, 360, 480, 960, and 1440 min after dosing, and blood was collected by cardiac puncture. Animals receiving the vehicle only were killed at 5 min postdosing to determine if the vehicle altered the chromatography of the retinoic acid compounds. All procedures were performed in dim light because of the light sensitivity of the retinoic acid compounds. Livers and tumors from all the animals were quickly removed and weighed. The gall-bladder was removed from the liver samples. Both liver and tumor were snap-frozen in liquid nitrogen and stored in aluminum-foil-wrapped cryotubes at -70 °C until analysis. Blood was centrifuged at 1500 g for 10 min, and plasma was stored in foil-wrapped tubes at -70 °C until analysis. Additional mice, dosed with either 13-CRA at 100 mg/kg and/or TAM at 30 mg/kg for either one dose or eight doses, were killed at 0.5 h after the single dose or the eighth dose to determine if there were any changes in peak plasma concentration with repetitive dosing and if administration of TAM interfered with the metabolism of 13-CRA.

Sample preparation

Samples of plasma, liver, and tumor were thawed in a dark environment. Liver and tumor samples were weighed, homogenized in 2 vols of filtered, distilled water; and stored at -70 °C until analysis.

HPLC assay

A 35- μ l aliquot of the internal standard (9.45 μ M Ro 11-5036, kindly provided by Hoffmann LaRoche, Inc., Nutley, N.J.) was added to 100- μ l aliquots of plasma or tissue homogenate. Retinoids and internal standard were extracted from plasma or tissue homogenates with 0.7 vol. acetonitrile: 1-butanol (50:50, v:v) in the presence of 0.3 vol. K₂HPO₄ (pH 11.1). The organic phase was analyzed by reversed-phase gradient HPLC using a modification of the method of Bugge et al. [4]. The HPLC system employed Beckman solvent-delivery model 110B pumps (Beckman Instruments, Fullerton, Calif.) programmed with a 10-min linear gradient from 30% to 100% B at a flow rate of 1.5 ml/min. Mobile phase A was acetonitrile: 0.02 M ammonium acetate:acetic acid (pH 4.6; 50:50:0.5, by vol.), and mobile phase B was acetonitrile:0.2 M ammonium acetate:acetic acid (pH 8.4; 95:5:0.04, by vol.). The column employed was a Zorbax ODS special analytical column (MacMOD Analytical, Chadds Ford, Pa.) of 25 cm \times 4.6 mm inside diameter with 5- μ m spherical particles. The sample injection volume was 100 μ l. Detection of retinoids was accomplished with a UV detector set at 360 nm. The detector output was processed with a SP4290 integrator (Spectra-Physics Inc., San Jose, Calif.). Concentrations of retinoids were determined by the ratio of the peak height of the compound to the peak height of the internal standard, with reference to a concomitantly performed standard curve. Under these conditions, good baseline separation of 4-oxo-13CRA, internal standard, 13CRA, and ATRA was achieved, with retention times being approximately 6, 11, 13.3, and 14.5 min, respectively. There was no endogenous interfering peak, and the inter- and intraday assay CV% were <10% for 13-CRA and ATRA. The lower limit of quantitation was 30 ng/ml (0.10 μ M) for both 13-CRA and ATRA. The assay was linear at concentrations between 0.10 and 10 μ M. Samples expected to contain concentrations higher than 10 μ M were diluted prior to analysis.

Pharmacokinetic analysis

Noncompartmental analysis was performed on the plasma concentrations of 13-CRA and metabolites using the program LaGran

[21], which uses the LaGrange function [31]. Compartmental analysis was performed with ADAPT II [8] using a one-compartment open linear model with first-order absorption.

Optimal T/C percent

The percentage of treated/control value (%T/C) was calculated by division of median treated tumor weight by the median control tumor weight on each observation day and multiplication by 100. For display the optimal value or minimal value obtained after the first course of treatment is shown and the day on which the optimal %T/C occurred is shown in parentheses.

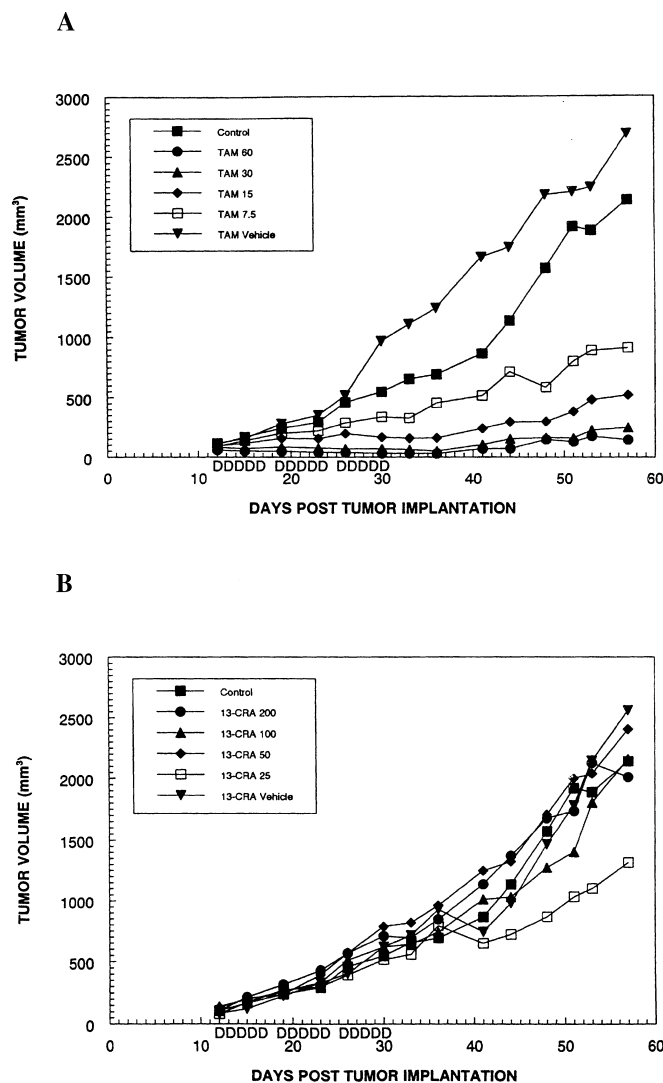


Fig. 1 **A** Effect of TAM as a single agent on MCF-7 human xenografts in athymic nude mice. Mice were implanted with tumor and treated with doses of TAM ranging from 7.5 to 60 mg/kg daily for 5 of 7 days/week for a period of 3 weeks, after tumors had reached a volume of 22–394 mm³. The vehicle control was water. **B** Effect of 13-CRA as a single agent on MCF-7 human xenografts in athymic nude mice. 13-CRA was given at doses ranging from 25 to 200 mg/kg daily for 5 days/week for 3 weeks, after tumors had reached a volume of 150–300 mm³. The vehicle control was cremophor:ethanol:water (1:1:6, by vol.). Results are expressed as median tumor volumes

Growth delay

Growth delay was expressed as the percentage by which the treated-group median tumor volume was delayed in achieving a specified tumor size as compared with the controls. We used the formula $(T - C)/C \times 100$, where T and C are the median times (in days) to a predetermined target tumor volume (in cubic millimeters) for the treated and control groups, respectively.

Net log cell kill

This is an estimate of the number of \log_{10} units of cells killed by the test agent and is calculated using the following formula:

$$\frac{[(T - C) - \text{Duration treatment}] \times 0.301}{\text{Doubling time}}$$

where the doubling time is calculated as the time required for the tumor volume to increase from 500 to 1000 mm³ and T and C are the median numbers of days needed to reach the specified tumor size defined above.

Statistical evaluation

Median tumor volumes and tumor growth delays were calculated. The data were analyzed by one-way analysis of variance with pairwise comparisons using Dunnett's test and by the nonparametric method, Kruskal-Wallis test. Paired comparisons were done using the Mann-Whitney test. Significance was set at $P \leq 0.05$. Calculations were done using the statistical software package MINITAB (Minitab, Inc., State College, Pa.).

Results

Single-agent efficacy

Treatment with TAM for 15 days was very effective in inhibiting the growth of MCF-7 xenografts (Fig. 1A, and Table 1, Addendum 1). The tumor growth was suppressed within 3 days of treatment at all dose levels

tested and regrowth was very slow after treatment had been stopped. The growth delay was greater than 91% for the three highest doses of TAM and 64% for the lowest dose of 7.5 mg/kg per day. The time to two doublings was greater than 49 days for the highest doses of TAM versus 25.6 days for the controls. The vehicle (water) did not result in any growth delay.

When 13-CRA was given as a single agent the effects were much less dramatic (Fig. 1B, Table 2, Addendum 2). The median time to two doublings was increased to 37 days as compared with 23.7 days for the controls. The vehicle controls also exhibited a growth delay, presumably due to the mild anticancer activity of cremophor.

Combination-agent efficacy

When TAM and 13-CRA were given concurrently for 15 days the tumor growth delays were of a magnitude similar to those observed with TAM alone, except when 13-CRA was combined with the lowest dose of TAM. At a daily dose of 7.5 mg/kg TAM the addition of 13-CRA resulted in an increased time to three doublings and in increased growth delays. When 7.5 mg/kg TAM was combined with 200 and 100 mg/kg 13-CRA the median time to three doublings increased from 23.9 days to 56 and 50 days, respectively. The growth delays increased from 8% to 152% and 125%, respectively. At the higher doses of TAM the addition of 13-CRA did not result in efficacy greater than that of TAM alone (Fig. 2, Table 3, Addendum 3–6).

Pharmacokinetic results

The 13-CRA plasma concentration-time profiles obtained from both fasted and nonfasted animals are

Table 1 Response of advanced-stage s.c. MCF-7 breast tumor xenografts to tamoxifen^a

TAM dose ^b	Optimal %T/C (day) ^c	Median days to 2 doublings ^d	Growth delay %T-C/C	NLCK ^e
Control	–	25.6 (4.5–>49)	–	–
60	7(41)	> 49 (9.8–>49)	> 91%	0.3
30	–3(41)	> 49 (10.9–>49)	> 91%	0.3
15	3(41)	> 49 (5.3–>49)	> 91%	0.3
7.5	18(41)	41.9 (16.1–>49)	64%	–0.1
Vehicle	–	19.2 (9.4–>49)	–	–

^aMCF-7 human breast tumor xenografts and 1.7-mg 17- β -estradiol pellets were implanted into athymic nude mice as described in Materials and methods. Mice were randomized for study at 10 mice per treatment group and 20 mice in the control group. Treatment with tamoxifen p.o. as based on exact body weight was begun on day 18, qdX5, and repeated for 3 weeks. Vehicle-treated mice received 0.01 ml/g body weight water p.o. on the same treatment schedule. Tumor length and width were measured twice weekly using digital calipers and tumor volumes were calculated from the formula $\text{length} \times \text{width}^2/2$, where length is the largest dimension and width is the smallest dimension perpendicular to the length

^bDose of tamoxifen is given in mg/kg per day of treatment

^cOptimal %T/C is the minimal %T/C obtained for that treatment group, and the day on which it occurred is indicated in parentheses

^dThe median number of days required for 2 doublings is given, and the range of days needed for 2 doublings is indicated in parentheses

^eNLCK is the net log cell kill as based on the values recorded for the doubling times calculated from tumor-doubling times

Addendum 1 Data for Fig. 1A: median values and their 95% confidence intervals. The number of animals surviving at each time point is given in parentheses under the 95% confidence interval

Treatment	Days post tumor implantation													
	12	15	19	23	26	30	33	36	41	44	48	51	53	57
Control	107.5	168	242	294	460	548	655	693	867	1138	1570	1918	1885	2559
	85.5-175 (20)	117-232 (20)	185-357 (20)	240-413 (20)	347-637 (19)	421-766 (19)	452-1014 (15)	413-1105 (15)	433-1525 (15)	560-1779 (11)	687-2330 (11)	645-2980 (11)	882-3014 (11)	889-3171 (11)
TAM 60	58.5	49.5	49	40.5	38.5	32	32	30	69	71	146	128	174.5	144
	38.5-120.5 (8)	35-140 (8)	35-180 (8)	28-148 (8)	17-130 (8)	13-100 (8)	14-116 (8)	13-92 (8)	50-129 (8)	50-143 (8)	10-202 (8)	91-217 (8)	138.5-237.5 (8)	105-284 (8)
TAM 30	81.5	71	85	76	69	69	63	50	103	153	161	160	226	249
	40-101 (8)	38-179 (8)	54-258 (8)	46-298 (8)	45-372 (8)	39-374 (8)	26-361 (8)	22-444 (8)	34-199 (8)	65-600 (8)	48-851 (8)	52-1014 (8)	97-1285 (8)	106-1636 (8)
TAM 15	91.5	116	156.5	155	197	166.5	158	159.5	240	294	297	383	484	521.5
	33-200 (8)	39-220 (8)	36-303 (8)	50-242 (8)	54-304 (8)	96-250 (8)	83-393 (8)	93-295 (8)	94-359 (8)	144-378 (8)	189-452 (7)	155-558 (7)	264-613 (7)	270-886 (7)
TAM 7.5	91	137.5	200.5	220.5	289	337.5	339	455	516	714	585	804	895	917
	56.5-123 (8)	61-238 (8)	92-321 (8)	101-416 (8)	126-431 (8)	103-667 (8)	83-809 (7)	86-787 (7)	90-1095 (7)	121-1238 (7)	132-1657 (7)	136-1845 (7)	139-2419 (7)	197-2699 (7)
TAM vehicle	116	155	280.5	353.5	518	970	1109	1240	1660	1742	2184	2209	2247	2691
	62-190 (8)	73-266 (8)	72-587 (8)	50-666 (7)	58-867 (6)	230-1070 (6)	560-1150 (6)	640-1891 (5)	690-1960 (5)	750-2363 (5)	920-2541 (5)	921-2521 (5)	1116-2975 (5)	1277-3416 (5)

Table 2 Response of advanced-stage s.c. MCF-7 breast tumor xenografts to 13-CRA^a

13-CRA dose ^b	Optimal %T/C(day) ^c	Median days to 2 doublings ^d	Growth delay %T-C/C	NLCK ^e
Control	–	23.7 (12.2–>38.0)	–	–
200	–9(21)	37.0 (21.8–>38.0)	56%	–0.2
133.2	59(21)	31.0 (24.6–>38.0)	31%	–0.5
89	27(21)	36.9 (13.0–>38.0)	55%	–0.2
59.4	27(21)	22.6 (17.2–37.0)	–5%	–0.9
39.6	80(21)	27.4 (18.5–35.0)	16%	–0.7
26.4	31(21)	36.2 (21.8–>38.0)	53%	–0.3
Vehicle	–	31.8 (19.6–>38.0)	34%	–

^aMCF-7 human breast tumor xenografts and 1.7-mg 17- β -estradiol pellets were implanted into athymic nude mice as described in Materials and methods. Mice were randomized for study at 10 mice per treatment group and 20 mice in the control group. Treatment with 13-CRA p.o. as based on exact body weight was begun on day 12, qdX5, and repeated for 3 weeks. Vehicle-treated mice received 0.01 ml/g body weight cremophor:ethanol:water (1:1:6, by vol.) p.o. on the same treatment schedule. Tumor length and width were measured twice weekly, and tumor volumes were calculated as described in Table 1

^bDose of 13-CRA is given in mg/kg per day of treatment

^cOptimal %T/C is the minimal %T/C obtained for that treatment group, and the day on which it occurred is indicated in parentheses. %T/C is the change in treated tumor over the change in control tumor using the median tumor volumes

^dThe median number of days required for 2 doublings is given, and the range of days needed for 2 doublings is indicated in parentheses

^eNLCK is the net log cell kill as based on the values recorded for the doubling times calculated from tumor-doubling times

shown in Fig. 3. It is apparent that the absorption of 13-CRA is much more variable in fed mice (Fig. 3B) than in fasted animals (Fig. 3A). Furthermore, 13-CRA peak concentrations are lower in the fed animals than in the fasted mice, suggesting that food may interfere with the absorption of 13-CRA.

The decline of 13-CRA concentrations in plasma over time fit a one-compartment open linear model with a half-life of 73 min and first-order absorption. Peak concentrations were obtained at 30–45 min in plasma when mice had been fasted prior to dosing. The area under the plasma concentration-time curve (AUC) averaged 975 $\mu\text{g ml}^{-1} \text{ min}$, with the apparent clearance being 103 $\text{ml min}^{-1} \text{ kg}^{-1}$. A major metabolite, likely the glucuronide of 4-oxo-CRA, was identified at a retention time of approximately 3.3 min (METAB X in Table 4). AUCs were also calculated for 4-oxo-CRA, ATRA, and METAB X and were, respectively, 117, 87, and 3262 $\mu\text{g ml}^{-1} \text{ min}$. The peak 13-CRA concentrations detected in tumors were 20% of the peak plasma concentrations, whereas those found in liver were equivalent to the plasma values (Table 4). Peak ATRA concentrations represented 5% of 13-CRA concentrations detected in plasma, 68% of those found in liver, and 20% of those found in tumor (Table 4). 4-oxo-CRA represented 2–10% of 13-CRA concentrations detected in plasma and liver but was not observed in tumor.

Mean peak plasma 13-CRA concentrations in the nonfasted mice occurred at 30 min and were $4.19 \pm 1.66 \mu\text{g/ml}$. Mean peak plasma 13-CRA concentrations were not different on day 1 or day 8 and were not affected by coadministration of TAM. The peak concentrations obtained on days 1 and 8 of treatment in mice treated with 13-CRA only were, re-

spectively, 4.73 ± 1.92 and 3.10 ± 3.09 , whereas those obtained in mice treated with both 13-CRA and TAM were 3.99 ± 1.17 and $4.53 \pm 0.02 \mu\text{g/ml}$, respectively. Mean concentrations of 13-CRA detected in tumor and liver were equivalent to those found in the initial pharmacokinetics study. No difference in 4-oxo-CRA was observed between mice that had been given 13-CRA alone and those that had been treated with 13-CRA and TAM.

Discussion

Several studies have suggested that retinoids may enhance the anticarcinogenic effects of antiestrogen treatment. In 1983, McCormick et al. [19] noted that oral retinyl acetate combined with ovariectomy significantly reduced the number of second primary breast tumors in rats. Fenretinide and TAM have been shown to be more effective than either agent alone against methyl nitrosourea-induced mammary tumors [18]. Various retinoids have been shown to be growth-inhibitory to mammary tumors in vitro [10, 17–19, 22, 24–26, 29]. Generally, retinoids are inhibitory only to ER-positive tumors [10, 26], although some agents classed with retinoids are effective against either ER-positive or ER-negative cells lines [11, 25]. Sheikh et al. [24] demonstrated that transfection of ER into ER-negative breast-cancer cell lines resulted in increased expression of retinoic acid receptor (RAR) alpha and in sensitivity to growth inhibition by ATRA. Rubin et al. [22] showed that both ATRA, a ligand for RARs, and 9-*cis*-retinoic acid, a ligand for both RARs and RXRs, down-regulate ER mRNA and protein in MCF-7 cells. These agents also down-regulate estrogen

Addendum 2 Data for Fig. 1B: median values and their 95% confidence intervals. The number of animals surviving at each time point is given in parentheses under the 95% confidence interval

Treatment	Days post tumor implantation													
	12	15	19	23	26	30	33	36	41	44	48	51	53	57
Control	107.5	168	242	294	460	548	655	693	867	1138	1570	1918	1885	2559
	85.5-175 (20)	117-232 (20)	185-357 (20)	240-413 (20)	347-637 (19)	421-766 (19)	452-1014 (15)	413-1105 (15)	433-1525 (15)	560-1779 (11)	687-2330 (11)	645-2980 (11)	882-3014 (11)	889-3171 (11)
13-CRA 200	105.5	216	317.5	430	568	710	695	849	1138	1372	1675	1733	2125	2009
	57-223 (8)	118-321 (8)	226-468 (8)	330-608 (8)	388-675 (8)	586-898 (8)	543-945 (7)	725-1175 (7)	795-1586 (7)	824-1753 (7)	1201-2083 (7)	984-2316 (6)	1224-2730 (5)	1203-3479 (5)
13-CRA 100	141.5	197	261	333.5	513	619.5	643	741	1014.5	1035.5	1276	1404.5	1801	2009
	92-244 (8)	111-451 (8)	149-603 (8)	206-682 (8)	256-788 (8)	293-934 (8)	400-1011 (8)	388-1101 (8)	517-1446 (8)	563-1265 (8)	703-1820 (8)	708-2707 (8)	896-2994 (8)	1054-3436 (8)
13-CRA 50	130.5	206	240.5	395.5	568	788	821	965	1249	1325	1703	1998	2037	2405
	58-282 (8)	47-489 (8)	57-488 (8)	90-660 (8)	131-900 (8)	256-967 (8)	372-1229 (7)	395-1341 (7)	322-2179 (6)	347-2237 (6)	544-2678 (6)	564-3014 (6)	789-3485 (6)	831-4043 (6)
13-CRA 25	81.5	173	275.5	298	394	517.5	561	794	651	724	870.5	1038	1107	1319
	68.5-141 (8)	94-275 (8)	120-464 (8)	169-596 (8)	165-965 (8)	236-1091 (8)	254-1114 (8)	342-1399 (6)	341-1516 (4)	391-1865 (4)	518-2386 (4)	602-2410 (4)	700-2516 (4)	715-2976 (4)
13-CRA vehicle	85.5	123	228	330	405	621	715.5	931	744	984	1467	1778	2145	2559
	62.5-104.5 (8)	66-178 (8)	117-310 (8)	145-512 (8)	193-520 (8)	234-667 (7)	83-809 (6)	373-1223 (6)	111-1572 (5)	194-2074 (5)	440-2175 (5)	542-2189 (5)	672-2270 (5)	889-3171 (5)

Table 3 Response of advanced-stage s.c. MCF-7 breast tumor xenografts to combination treatment with tamoxifen and 13-CRA^a

TAM ^b	13-CRA ^b	Optimal %T/C (day) ^c	Median days to 3 doublings ^d	Growth delay %T-C/C	NLCK ^e
Control	Control	–	22.2 (8.1–>56.0)	–	–
–	200	105(33)	29.2 (9.3–>56.0)	32%	–0.6
–	100	76(19)	30.9 (27.9–38.8)	39%	–0.5
–	50	103(19)	30.0 (17.1–41.1)	35%	–0.5
–	25	85(33)	> 22.0 (10.9–>38.2)	–1%	–0.9
–	Vehicle	–	19.1 (15.0–>33.6)	–14%	–
60	–	–49(36)	54.7 (47.0–>56.0)	146%	0.8
30	–	–36(36)	50.6 (21.7–>56.0)	128%	0.5
15	–	10(36)	53.5 (27.9–>56.0)	141%	0.7
7.5	–	60(23)	23.9 (13.2–>56.0)	8%	–0.8
Vehicle	–	–	29.0 (15.0–>33.6)	31%	–
60	200	–17(33)	> 55.9 (46.8–>56.0)	152%	0.8
60	100	–22(30)	> 56.0 (42.0–>56.0)	152%	0.8
60	50	–13(30)	> 56.0 (> 56.0)	152%	0.8
60	25	0(36)	54.1 (45.8–>56.0)	144%	0.7
30	200	–16(19)	48.9 (35.9–>56.0)	120%	0.5
30	100	1(30)	> 56.0 (38.0–>56.0)	152%	0.8
30	50	–43(23)	> 56.0 (54.9–>56.0)	152%	0.8
30	25	–35(33)	> 55.4 (35.5–>56.0)	150%	0.8
15	200	7(33)	53.5 (42.4–>56.0)	141%	0.7
15	100	–28(33)	> 56.0 (46.8–>56.0)	152%	0.8
15	50	12(33)	52.3 (33.6–>56.0)	136%	0.6
15	25	–4(30)	48.5 (34.1–>56.0)	118%	0.4
7.5	200	24(19)	> 56.0 (31.9–>56.0)	152%	0.8
7.5	100	22(30)	50.0 (21.7–>56.0)	125%	0.5
7.5	50	43(36)	40.9 (28.0–>56.0)	84%	0.0
7.5	25	48(33)	39.5 (19.3–53.9)	78%	0.0

^aMCF-7 human breast tumor xenografts and 1.7-mg 17- β -estradiol pellets were implanted into athymic nude mice as described in Materials and methods. Mice were randomized for study at 8 mice in each treatment group and 20 mice in the control group. Treatment with 13-CRA and TAM p.o. was begun on day 12, qdX5, and repeated for 3 weeks. Vehicle-treated mice for TAM received 0.01 ml/g body weight water, whereas vehicle-treated mice for 13-CRA received 0.01 ml/g cremophor:ethanol:water (1:1:6, by vol.) p.o. on the same treatment schedule. Tumor length and width were measured twice weekly, and tumor volumes were calculated as described in Table 1

^bDoses of TAM and 13-CRA are given in mg/kg/day of treatment

^cOptimal %T/C is the minimal %T/C obtained for that treatment group, and the day on which it occurred is indicated in parentheses

^dThe median number of days required for 3 doublings is given, and the range of days needed for 3 doublings is indicated in parentheses

^eNLCK is the net log cell kill as based on the values recorded for the doubling times calculated from tumor-doubling times

response genes. Down-regulation of the ER and/or its response genes may result in an increased TAM effect. If the ER were down-regulated, there would be fewer ER receptors; therefore, at the same concentration, TAM could more effectively compete with endogenous estrogens for binding. Likewise, if response genes were down-regulated, the signal from the ER would be attenuated, possibly resulting in down-regulated tumor growth. Therefore, it is plausible that retinoids, which are thought to act through specific receptors, may well enhance the antiestrogen effect of TAM against breast cancer.

ATRA and fenretinide have been the major retinoids investigated in combination with TAM. In other tissues, most notably in oral premalignancy, 13-CRA has shown cancer-inhibitory effects [13]. 13-CRA may represent a more attractive clinical compound as compared with ATRA because its pharmacokinetics are stable over time

[12], whereas those of ATRA are variable [1, 7, 20]. However, 13-CRA does not bind to the RARs with as much affinity as does ATRA [3]. Fenretinide, a semi-synthetic retinoid analogue, appears to exert its effects by mechanisms other than those involving RARs [25]. Although each of these agents is considered to be a retinoid, it is necessary that we discover which of these agents, if any, may be best suited for combination with TAM for possible therapeutic use in breast cancer.

Our data described experiments in xenografts of human ER-positive breast carcinomas in immune-suppressed mice. Peak tumor concentrations of 13-CRA and ATRA were in the range of effective concentrations reported for both agents in *in vitro* studies [10, 11, 30]. 13-CRA alone was ineffective against established MCF-7 human breast cancer xenografts in these studies. The data demonstrated a benefit in the combination of 13-CRA with TAM only for the lowest dose of TAM

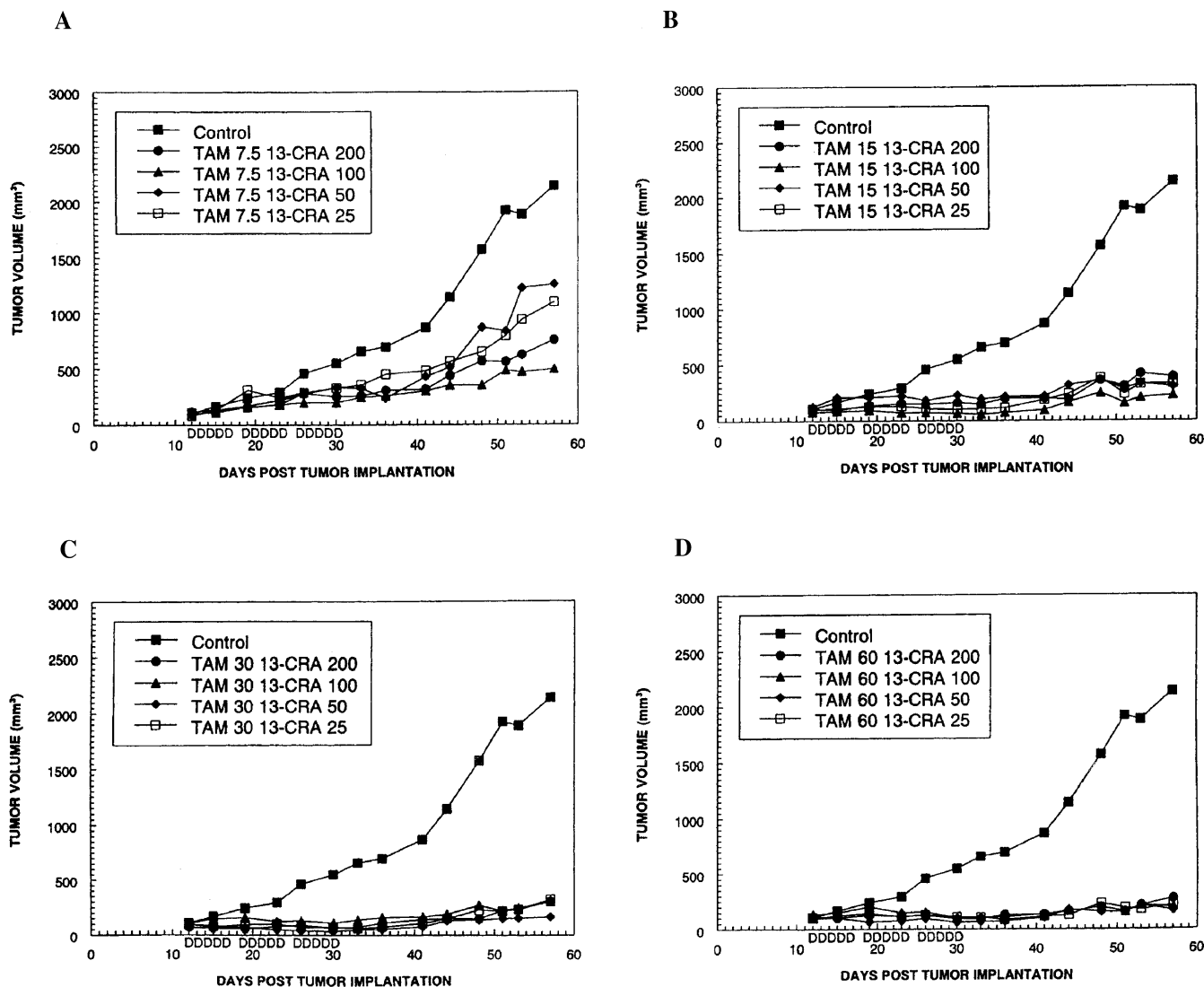


Fig. 2A–D Effect of the combination of daily doses of 25–200 mg/kg 13-CRA and A 7.5, B 15, C 30, or D 60 mg/kg TAM on established MCF-7 human breast cancer xenografts in athymic nude mice. Results are expressed as median tumor volumes

combined with the two highest doses of 13-CRA. Because TAM alone is well tolerated and is effective in ER-positive breast cancer, it is not likely that 13-CRA and TAM would be useful against established breast cancers clinically.

Although a reasonable delivery of 13-CRA to the tumor was demonstrated in these studies, there remain many unanswered questions. Protein binding may play a role in the observation of little activity in vivo as compared with in vitro studies. 13-CRA probably circulates with 90% of the drug bound to albumin in vivo versus about 40% protein binding in culture. Concentrations of TAM were not measured. It is possible that these were altered by the addition of 13-CRA. However, the similarity of the median time to three doublings observed in tumors treated either with TAM alone or with TAM and 13-CRA would argue against this pos-

sibility. At low concentrations, TAM is an estrogen agonist [15]. The increased activity of the 13-CRA and TAM combination as compared with TAM alone at a low dose may be related to estrogen agonist effects rather than to the antagonistic effects of TAM at this dose. Two other possibilities may influence drug interactions in clinical trials but would not be addressed by either in vitro or these in vivo experiments. These possibilities (a) relate to the necessity for estrogen supplementation to the mouse for the xenografts to grow and (b) to the immunosuppressed state of the animal. The relationship of ambient estrogen concentrations to retinoid or TAM activity has not been defined. Likewise, retinoids do have effects on immune function ([26] and references therein) that may or may not influence tumor response. Finally, the active drug may be not 13-CRA but rather ATRA or 9-*cis*-retinoic acid or a combination of these. ATRA concentrations detected in tumor were only about 20% of 13-CRA concentrations found in plasma, and 9-*cis*-retinoic acid was not detectable. Thus, ATRA concentrations would be at or near an effective “in vitro” concentration (300 ng/g, or 1 μ M) for only

Addendum 3 Data for Fig 2A: median values and their 95% confidence intervals. The number of animals surviving at each time point is given in parentheses under the 95% confidence interval

Treatment	Days post tumor implantation														
	12	15	19	23	26	30	33	36	41	44	48	51	53	57	
Control	107.5	168	242	294	460	548	655	693	867	1138	1570	1918	1885	2140	
	85.5-175 (20)	117-232 (20)	185-357 (20)	240-413 (20)	347-637 (19)	421-766 (19)	452-1014 (15)	413-1105 (15)	433-1525 (15)	560-1779 (11)	687-2330 (11)	645-2980 (11)	882-3014 (11)	1028-3935 (11)	
TAM 60 13-CRA 200	114	104.5	126	117	124.5	91	91.5	130	130	162.5	247	173	245.5	275	
	54-190 (8)	49-182 (8)	40-208 (8)	36-193 (8)	41-204 (8)	60-169 (8)	31-186 (8)	46-173 (8)	57-195 (8)	79-246 (8)	119-316 (8)	77-302 (8)	103-405 (8)	95-466 (8)	
TAM 60 13-CRA 100	132.5	150.5	202	148.5	154.5	103.5	107.5	107.5	133.5	162.5	198.5	157	213	199	
	72-161.5 (8)	62-187 (8)	80-265 (8)	83-221 (8)	83-226 (8)	21-154 (8)	52-215 (8)	70-228 (8)	67-222 (8)	93-236 (8)	97-348 (8)	83-304 (8)	100-410 (8)	83-533 (8)	
TAM 60 13-CRA 50	97	102.5	68	75	94.5	61.5	67.5	70.5	102	176	154.5	152	213	169	
	65-152.5 (8)	42-175 (8)	33-179 (8)	32-193 (8)	31-219 (8)	42-341 (8)	30-168 (8)	20-164 (8)	38-198 (8)	46-257 (8)	64-326 (8)	38-327 (8)	60-350 (8)	0-408 (7)	
TAM 60 13-CRA 25	99.5	124	145	113	130.5	109.5	106	84.5	114.5	127	231.5	190.5	177	213	
	72-143.5 (8)	79-216 (8)	83-252 (8)	62-307 (8)	62-310 (8)	19-158 (8)	45-400 (8)	52-343 (8)	67-407 (8)	83-330 (8)	121-434 (8)	100-572 (8)	111-630 (8)	114-767 (8)	
TAM 60 13-CRA vehicle	58.5	49.5	49	40.5	38.5	32	32	30	69	71	146	128	174.5	144	
	38.5-120.5 (8)	35-140 (8)	35-180 (8)	28-148 (8)	17-130 (8)	13-100 (8)	14-116 (8)	13-92 (8)	50-129 (8)	50-143 (8)	10-202 (8)	91-217 (8)	138.5-237.5 (8)	105-284 (8)	

Addendum 4 Data for Fig 2B: median values and their 95% confidence intervals. The number of animals surviving at each time point is given in parentheses under the 95% confidence interval

Treatment	Days post tumor implantation													
	12	15	19	23	26	30	33	36	41	44	48	51	53	57
Control	107.5	168	242	294	460	548	655	693	867	1138	1570	1918	1885	2559
	85.5-175 (20)	117-232 (20)	185-357 (20)	240-413 (20)	347-637 (19)	421-766 (19)	452-1014 (15)	413-1105 (15)	433-1525 (15)	560-1779 (11)	687-2330 (11)	645-2980 (11)	882-3014 (11)	889-3171 (11)
TAM 30	70.5	64	59	77	92	67	74	110	139	150	146	217.5	244	303
13-CRA 200	52.5-123 (8)	43.5-107 (8)	25.5-123.5 (8)	32-154 (8)	21-166 (7)	19-158 (7)	23-178 (7)	34-187 (7)	46-235 (7)	61-353 (7)	93-474 (6)	72-594 (6)	115-525 (6)	115-464 (6)
TAM 30	106.5	146.5	156	122	129	110	137.5	159	164	186	267	215.5	244	301
13-CRA 100	67-230 (8)	57-289 (8)	52-256 (8)	70-208 (8)	52-233 (8)	62-222 (8)	67-279 (8)	78-319 (8)	113-320 (8)	136-319 (8)	193-433 (8)	152-564 (6)	125-862 (5)	97-714 (5)
TAM 30	96.5	87	68.5	48	40	36.5	46	53	74.5	129	137	147	151	163
13-CRA 50	60-131 (8)	40-148 (8)	25-153 (8)	20-122 (8)	15-104 (8)	10.5-102.5 (8)	14.5-92.5 (8)	21-105.5 (8)	29-147 (8)	60-208 (7)	48-194 (7)	43-231 (7)	62-262 (7)	55-282 (7)
TAM 30	94.5	76.5	94	99.5	73	68.5	61	74.5	104.5	149	224	213	229	332
13-CRA 25	49-126 (8)	31-122 (8)	43-166 (8)	52-208 (8)	54-172 (8)	40-213 (8)	37-198 (8)	39-208 (8)	50-282 (8)	90-331 (8)	192-531 (8)	149-486 (8)	170-552 (8)	239-703 (7)
TAM 30	81.5	71	85	76	69	69	63	50	103	153	161	160	226	249
13-CRA vehicle	40-101 (8)	38-179 (8)	54-258 (8)	46-298 (8)	45-372 (8)	39-374 (8)	26-361 (8)	22-444 (8)	34-199 (8)	65-600 (8)	48-851 (8)	52-1014 (8)	97-1285 (8)	106-1636 (8)

Addendum 5 Data for Fig. 2C: Median values and their 95% confidence intervals. The number of animals surviving at each time point is given in parentheses under the 95% confidence interval

Treatment	Days post tumor implantation													
	12	15	19	23	26	30	33	36	41	44	48	51	53	57
Control	107.5	168	242	294	460	548	655	693	867	1138	1570	1918	1885	2559
	85.5-175 (20)	117-232 (20)	185-357 (20)	240-413 (20)	347-637 (19)	421-766 (19)	452-1014 (15)	413-1105 (15)	433-1525 (15)	560-1779 (11)	687-2330 (11)	645-2980 (11)	882-3014 (11)	889-3171 (11)
TAM 15 13-CRA200	108.5	97.5	136.5	153.5	147.5	162	148	198	201	188	364	309	423	393
	67.5-162 (8)	57-212 (8)	70-256 (8)	81-214 (8)	83-241 (8)	68-237 (8)	74-204 (8)	87-245 (7)	102-387 (7)	137-477 (7)	199-629 (7)	118-721 (7)	162-649 (7)	121-891 (7)
TAM 15 13-CRA 100	78	84.5	93.5	71.5	72	65	56	71.5	93.5	164	249	157	207	227
	57-145 (8)	41-186 (8)	42-177 (8)	40-163 (8)	46-127.5 (8)	39-103.5 (8)	34-116 (8)	35.5-119 (8)	60-165 (8)	118-227 (8)	165-298 (8)	95-258 (8)	160-286 (8)	120-955 (7)
TAM 15 13-CRA 50	125.5	210.5	211	222	182	228	193	215	218	318	361	286	333	306
	85-201 (8)	111-347 (8)	125-425 (8)	90-381 (7)	76-418 (7)	93-411 (7)	100-416 (7)	113-425 (7)	148-371 (7)	225-543 (7)	298-987 (7)	224-856 (7)	247-973 (7)	249-1166 (7)
TAM 15 13-CRA 25	104.5	106	125.5	131	108	100	107	114	187	239	385	238	328	332
	66-219 (8)	73-249 (8)	71-258 (8)	82-175 (8)	70-175 (8)	69-215 (8)	62-233 (7)	79-223 (7)	115-330 (7)	175-308 (7)	213-590 (7)	171-473 (7)	222-581 (6)	152-650 (6)
TAM 15 13-CRA vehicle	91.5	116	156.5	155	197	166.5	158	159.5	240	294	297	383	484	521.5
	33-200 (8)	39-220 (8)	38-303 (8)	50-242 (8)	54-304 (8)	98-250 (8)	83-393 (8)	93-295 (8)	94-359 (8)	144-378 (8)	189-452 (7)	155-558 (7)	264-613 (7)	270-886 (6)

Addendum 6 Date for Fig. 2D: Median values and their 95% confidence intervals. The number of animals surviving at each time point is given in parentheses under the 95% confidence interval

Treatment	Days post tumor implantation														
	12	15	19	23	26	30	33	36	41	44	48	51	53	57	
Control	107.5	168	242	294	460	548	655	693	867	1138	1570	1918	1885	2559	
	85.5-175 (20)	117-232 (20)	185-357 (20)	240-413 (20)	347-637 (19)	421-766 (19)	452-1014 (15)	413-1105 (15)	433-1525 (15)	560-1779 (11)	687-2330 (11)	645-2980 (11)	882-3014 (11)	889-3171 (11)	
TAM 7.5 13-CRA 200	118	139.5	156	187	284	254	261	309	319	440	565	560	622	755	
	48-158 (8)	30-235 (8)	37-256 (8)	43-296 (8)	43-345 (8)	67-368 (7)	49-333 (7)	61-385 (7)	75-606 (7)	91-661 (7)	94-844 (7)	182-871 (7)	177-1087 (7)	195-1232 (7)	
TAM 7.5 13-CRA 100	103.5	113.5	163	184	202	202	248.5	260	304	354	355	485	472	495	
	67.5-158.5 (8)	90-194 (8)	131-278 (8)	142-270 (8)	145-282 (7)	133-321 (8)	147-296 (8)	170-388 (8)	204-466 (8)	258-600 (8)	245-657 (8)	279-821 (7)	272-849 (7)	201-955 (6)	
TAM 7.5 13-CRA 50	94	132	174.5	222	280	335	329	240	429	513	868	836	1221	1253	
	68-148.5 (8)	67-202 (8)	109-340 (8)	85-386 (8)	80-517 (7)	81-681 (7)	71-598 (7)	96-777 (7)	137-836 (6)	194-941 (6)	128-1323 (6)	159-1329 (6)	169-1509 (6)	166-1837 (6)	
TAM 7.5 13-CRA 25	86	133	313.5	254.5	290	334	358	451.5	480.5	562	648	793	938	1094	
	78.5-172 (8)	90-245 (8)	149-434 (8)	142-393 (8)	154-413 (8)	177-511 (8)	217-720 (8)	250-645 (8)	330-952 (8)	394-1147 (8)	438-1272 (8)	502-1639 (8)	603-2147 (8)	755-2511 (8)	
TAM 7.5 13-CRA vehicle	91	137.5	200.5	220.5	289	337.5	339	455	516	714	585	804	895	917	
	56.5-123 (8)	61-238 (8)	92-321 (8)	101-416 (8)	126-431 (8)	103-667 (8)	83-809 (7)	86-787 (7)	90-1095 (7)	121-1238 (7)	132-1657 (7)	136-1845 (7)	139-2419 (7)	197-2699 (7)	

Table 4 Mean peak concentrations of 13-CRA and metabolites detected in the plasma, liver, and tumor of fasted athymic nude mice treated with CRA p.o.

Compound	Plasma ($\mu\text{g/ml}$)	Liver ($\mu\text{g/g}$)	Tumor ($\mu\text{g/g}$)	AUC($\mu\text{g min ml}^{-1}$)
13-CRA	9.84 (9.42–10.23) ^b	10.44 (5.76–15.15) ^b	1.2 (0.98–1.36) ^b	975
4-oxo-CRA	0.47 (0.34–0.57)	0.4 (0.38–0.43)	0 (0)	117
ATRA	0.44 (0.39–0.48)	3.3 (2.54–4.08)	0.27 (0.16–0.38)	87
METAB X ^a	9.48 (8.84–10.13)	40.96 (40.66–41.27)	0.75 (0.64–0.86)	3262

^a Concentrations of METAB X are expressed in 13-CRA equivalents

^b Ranges for the values are presented in parentheses

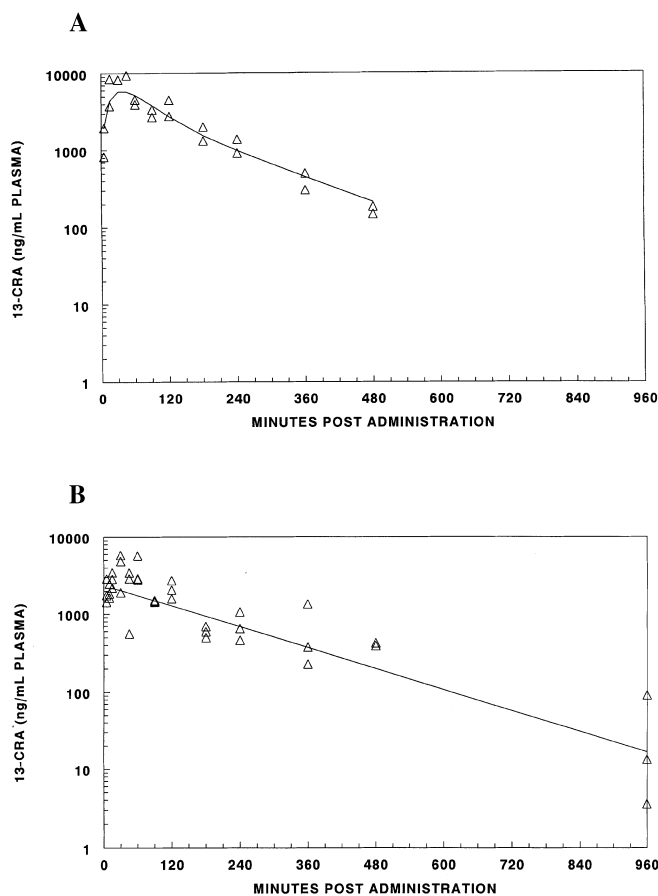


Fig. 3A,B Pharmacokinetics of 13-CRA given as a 100-mg/kg bolus to **A** fasted and **B** fed mice. Measurements of 13-CRA in plasma were obtained in 2 fasted mice at each time point and in 3 fed mice at each time point. Note the wider variability and lower peak in plasma concentration in fed animals

short periods. In general, though, delivery of drug to the tumor or its increased metabolism to inactive agents would not explain the lack of activity of this combination, given the activity of ATRA in acute promyelocytic leukemia [6, 14]. It is also possible that retinoids may be effective only in premalignant cells, not when cells have become fully malignant.

Because TAM and retinoic acid compounds are extensively metabolized in animals by the cytochrome P-450 mixed-function oxidase system, the activities of these agents might be altered by concurrent administration [12, 16]. Furthermore, the absorption of the agents might be altered by the presence of the other

agent or of food in the stomach. Our data demonstrate that 13-CRA was absorbed more variably and to a lesser extent in fed mice than in fasted animals. Coadministration of TAM and continued administration of 13-CRA had no effect on 13-CRA pharmacokinetics or metabolism.

In summary, preclinical studies suggest that the combination of retinoids and antiestrogen therapy are promising as a possible therapy for ER-positive breast cancer. However, further studies are necessary to define better regimens that are likely to be successful in the clinic.

References

1. Adamson PC, Pitot HC, Balis FM, Rubin J, Murphy RF, Poplack DG (1993) Variability in the oral bioavailability of all trans retinoic acid. *J Natl Cancer Inst* 85: 993
2. Alfthan O, Tarkkanen J, Grohn P, Heinonen E, Pyronen S, Salla K (1983) Tigason (etretinate) in the prevention of recurrence of superficial bladder tumors. *Eur Urol* 9: 6
3. Astrom A, Pettersson U, Krust A, Chambon P, Voorhees JJ (1990) Retinoic acid and synthetic analogs differentially activate retinoic acid receptor dependent transcription. *Biochem Biophys Res Comm* 173: 339
4. Bugge CJL, Rodriguez LC, Vane FM (1985) Determination of isotretinoin or etretinate and their major metabolites in human blood by reversed-phase high performance liquid chromatography. *J Pharm Biomed Anal* 3: 269
5. Cassidy J, Lippman M, Lacroix A, Peck G (1982) Phase II trial of 13-cis-retinoic acid in metastatic breast cancer. *Eur J Cancer Clin Oncol* 18: 925
6. Castaigne C, Chomienne C, Daniel MT, Ballerini P, Berger R, Fenaux P, Degos L (1990) All trans retinoic acid as a differentiation therapy for acute promyelocytic leukemia. I. Clinical results. *Blood* 76: 1704
7. Conley BA, Egorin MJ, Sridhara R, Finley R, Hemady R, Wu S, Tait N, Van Echo DA (1997) Phase I clinical trial of all-trans retinoic acid with correlation of its pharmacokinetics and pharmacodynamics. *Cancer Chemother Pharmacol* 39: 291
8. D'Argenio DZ, Schumitzky A (1979) A program package for simulation and parameter estimation in pharmacokinetic systems. *Comput Methods Prog Biomed* 9: 115
9. Degos L, Chomienne C, Daniel MT, Berger R, Dombret H, Fenaux P, Castaigne S (1990) Treatment of first relapse in acute promyelocytic leukemia with all trans retinoic acid. *Lancet* 336: 1440
10. Fontana J (1987) Interaction of retinoids and tamoxifen on inhibition of human mammary carcinoma cell proliferation. *Exp Cell Biol* 55: 136
11. Halter SA, Fraker LD, Adcock D, Vick S (1988) Effect of retinoids on xenotransplanted human mammary carcinoma cells in athymic mice. *Cancer Res* 48: 3733
12. Hill DL, Struck RF (1983) Pharmacologic disposition of chemopreventive retinoids. *Anticancer Res* 3: 171
13. Hong WK, Endicott J, Itri LM, Doos W, Batsakis JG, Bell R, Fotonoff S, Byers R, Atkinson EN, Vaughan C, Toth BB,

- Kramer A, Dimery IW, Skipper P, Strong S (1986) 13-*cis*-Retinoic acid in the treatment of oral leukoplakia. *N Engl J Med* 315: 1501
14. Huang M, Ye Y, Chen S, Chai J, Lu J-X, Zhao L, Gu L, Wang Z (1988) Use of all-transretinoic acid in the treatment of acute leukemia. *Blood* 72: 567
 15. Jaiyesimi IA, Buzdar AU, Decker DA, Hortobagyi GN (1995) Use of tamoxifen for breast cancer: twenty-eight years later. *J Clin Oncol* 13: 513
 16. Kupfer D, Mani C, Lee CA, Rifkind AB (1994) Induction of tamoxifen-4-hydroxylation by 2,3,7,8-tetraclorodibenzo-p-dioxin (TCDD), beta-naphthoflavone (betaNF), and phenobarbitol (PB) in avian liver: identification of P450 TCDD_{AA} as catalyst of 4-hydroxylation induced by TCDD and betaNF. *Cancer Res* 54: 3140
 17. Lotan R, Nicholson GL (1977) Inhibitory effects of retinoic acid or retinyl acetate on the growth of untransformed, transformed, and tumor cells in vitro. *J Natl Cancer Inst* 59: 1717
 18. McCormick DL, Moon RC (1986) Retinoid-tamoxifen interaction in mammary cancer chemoprevention. *Carcinogenesis (London)* 7: 193
 19. McCormick DL, Sowell ZL, Thompson CA, Moon RC (1983) Inhibition by retinoid and ovariectomy of additional primary malignancies in rats following surgical removal of the first mammary cancer. *Cancer* 51: 594
 20. Muindi JR, Frankel SR, Huselton C, DeGrazia F, Garland WA, Young CW, Warrell RP Jr (1992) Clinical pharmacology of oral all-trans retinoic acid in patients with acute promyelocytic leukemia. *Cancer Res* 52: 2138
 21. Rocci ML, Jusko WJ (1983) LAGRAN program for area and moments in pharmacokinetic analysis. *Comp Prog Biomed* 16: 203
 22. Rubin M, Fenig E, Rosenauer A, Menendez-Botet C, Achkar C, Bentel JM, Yahalom J, Mendelsohn J, Miller WH (1994) 9-*cis*-Retinoic acid inhibits growth of breast cancer cells and down regulates estrogen receptor RNA and protein. *Cancer Res* 54: 6549
 23. Shah JP, Strong EW, DeCrossee JJ, Itri L, Sellers P (1983) Effects of retinoids on oral leukoplakia. *Am J Surg* 146: 466
 24. Sheikh MS, Shao ZM, Li XS, Dawson M, Jetten AM, Wu S, Conley BA, Garcia M, Rochefort H, Fontana JA (1994) Retinoid-resistant estrogen receptor-negative human breast carcinoma cells transfected with retinoic acid receptor alpha acquire sensitivity to growth inhibition by retinoids. *J Biol Chem* 269: 21440
 25. Sheikh MS, Shao ZM, Li X-S, Ordonez JV, Conely BA, Wu S, Dawson ML, Han QX, Chao WR, Quick T, Niles RM, Fontana JA (1995) *N*-(4-Hydroxyphenyl)retinamide (4-HPR)-mediated biological actions involve retinoid receptor-independent pathways in human breast carcinoma. *Carcinogenesis* 16: 2477
 26. Smith MA, Parkinson DR, Cheson BD, Friedman MA (1992) Retinoids in cancer therapy. *J Clin Oncol* 10: 839
 27. Studer UE, Bidermann C, Chollet D, Jenzer S, Kraft R, Thomamichel G, Vonvanck F, Wuscher V (1986) Reduction of superficial bladder tumor recurrences by etretinate. *J Urol* 135: 283A
 28. Toma S, Isnardi L, Raffo P, Dastoli G, DeFrancisci E, Riccardi L, Palumbo R, Bollag W (1997) Effects of all-trans-retinoic acid and 13-*cis*-retinoic acid on breast-cancer cell lines: growth inhibition and apoptosis induction. *Int J Cancer* 70: 619
 29. Warrell RP Jr, Frankel SR, Miller WH Jr, Scheinberg DA, Itri LM, Hittelman WN, Vyas R, Andreeff M, Tafuri A, Jakubowski A, Gabrilove J, Gordon M, Dmitrovsky E (1991) Differentiation therapy of acute promyelocytic leukemia with tretinoin (all-trans retinoic acid). *N Engl J Med* 324: 1385
 30. Wetherell NT, Taylor CM (1986) The effects of retinoid treatment and antiestrogens on the growth of T47D human breast cancer cells. *Eur J Cancer Clin Oncol* 22: 53
 31. Yeh KC, Kwan KC (1978) A comparison of numerical integrating algorithms by trapezoidal, LaGrange and Spline approximation. *J Pharmacokinet Biopharm* 6: 79