



Only FLT3-ITD co-mutation did not have a deleterious effect on acute myeloid leukemia patients with NPM1 mutation, but concomitant with DNMT3A co-mutation or a < 3log reduction of MRD2 predicted poor survival

Wenbing Duan¹ · Jinsong Jia¹ · Jing Wang¹ · Xiaohong Liu¹ · Wenjing Yu¹ · Xiaolu Zhu¹ · Ting Zhao¹ · Qian Jiang¹ · Guorui Ruan¹ · Xiaosu Zhao¹ · Hongxia Shi¹ · Yingjun Chang¹ · Yu Wang¹ · Lanping Xu¹ · Xiaohui Zhang¹ · Xiaojun Huang^{1,2,3} · Hao Jiang¹

Received: 10 June 2024 / Accepted: 7 September 2024

© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2024

Abstract

Co-occurring mutations are frequently observed in acute myeloid leukemia (AML) with *NPM1* mutation, and *NPM1* measurable residual disease (MRD) is an effective prognostic biomarker. This retrospective study investigated the impact of gene co-mutations and *NPM1* MRD on outcomes in these patients. Among 234 patients, 11.5% carried the rare type *NPM1* mutation (*NPM1^{RT}*). The median age was 49 years (IQR 36–58), with a median follow-up of 30.4 months (IQR 12.1–55.7). Nine genes were mutated in > 10%, with *DNMT3A* (53.8%) and *FLT3-ITD* (44.4%) being most prevalent. Univariable analysis in 137 patients showed *FLT3-ITD*, *DNMT3A* co-mutations, and MRD2 < 3 log reduction predicted poorer survival. *FLT3-ITD* and *DNMT3A* co-mutations correlated with the lowest event-free (EFS) and overall survival (OS) (3-year EFS 30.0%; 3-year OS 34.4%; both $p < 0.001$). *FLT3-ITD* alone did not worsen survival compared to patients without *FLT3-ITD*. Multivariable analysis identified *DNMT3A* co-mutation [EFS, HR = 1.9, $p = 0.021$; OS, HR = 2.2, $p = 0.023$] and MRD2 ≥ 3 log reduction (EFS, HR = 0.2; OS, HR = 0.1, both $p < 0.001$) as independent survival predictors. Patients with *FLT3-ITD* and *DNMT3A* co-mutations or a MRD2 < 3 log reduction were identified as high risk, but allogeneic hematopoietic stem cell transplantation (allo-HSCT) improved survival significantly compared to chemotherapy only (3-year EFS, 57.9% vs. 30.0%, $p = 0.012$; 3-year OS, 72.9% vs. 34.4%, $p = 0.001$). In AML patients with *NPM1* mutation, the detrimental impact of *FLT3-ITD* mutation was exacerbated by *DNMT3A* co-mutation. Poor-risk younger patients identified by *FLT3-ITD* and *DNMT3A* co-mutations or MRD2 < 3 log reduction benefit from allo-HSCT.

Keywords *FLT3-ITD* and *DNMT3A* co-mutation · *NPM1* mutation · Measurable residual disease

Introduction

Nucleophosmin 1 (*NPM1*) mutation occurs in 28–35% of acute myeloid leukemia (AML) patients [1–3], and AML with *NPM1* mutation has been recognized as a separate entity in the World Health Organization (WHO) classification of myeloid neoplasms since 2016 [4]. AML with *NPM1* mutation has been shown to be associated with a normal karyotype (NK) at a reported frequency of 48% to 53% [5–7]; these patients are classified into the favorable prognosis group. However, a concurrent mutation in fms-like tyrosine kinase 3 internal tandem duplication (*FLT3-ITD*) diminishes the favorable effect of *NPM1* mutation [8].

✉ Xiaojun Huang
huangxiaojun@bjmu.edu.cn

✉ Hao Jiang
2516735116@qq.com

¹ Peking University People's Hospital, Peking University Institute of Hematology, National Clinical Research Center for Hematology Disease, Beijing, China

² Research Unit of Key Technique for Diagnosis and Treatments of Hematologic Malignancies, Chinese Academy of Medical Sciences, Beijing, China

³ Peking-Tsinghua Center for Life Sciences, Academy for Advanced Interdisciplinary Studies, Peking University, Beijing, China

With the application of next-generation sequencing (NGS), many mutations have been observed to co-occur in AML, especially in AML with *NPM1* mutation. *DNMT3A* mutation, which is associated with preleukemic clones, has been reported in ~50% of AML patients with *NPM1* mutation [9], and the prognostic significance of *DNMT3A* mutation is conflicting. Several studies have shown that *NPM1* measurable residual disease (MRD) is a favorable predictive marker for the survival of AML patients with *NPM1* mutation [10, 11]. However, further investigation is required to determine the prognostic value of the interactions of *FLT3-ITD* and *DNMT3A* or other molecular mutations with *NPM1* MRD in AML patients with *NPM1* mutation.

This study aimed to investigate the value of concurrent mutations and *NPM1* MRD for predicting survival in a retrospective cohort of AML patients with *NPM1* mutation against the background of conventional chemotherapy.

Patients and methods

Patients

From January 2013 to August 2021, treatment-naïve AML patients with *NPM1* mutations who received conventional induction or consolidation chemotherapy at the Peking University Institute of Hematology were included in this retrospective cohort study, and these patients were followed up until January 2024. All patients had an Eastern Cooperative Oncology Group (ECOG) performance status of 2 or less and no serious heart, lung, liver, or kidney dysfunction or severe infection. The study adhered to the principles of the Helsinki Declaration and was approved by the local institutional review boards.

Treatment

Induction therapy

Patients were treated with an anthracycline and cytarabine-based or homoharringtonine and cytarabine-based induction chemotherapy regimen following the Chinese guidelines for the diagnosis and treatment of adult AML (not acute promyelocytic leukemia) (2011) [12] and the 2010 European Leukemia Net recommendations for AML [13]. The chosen induction therapy regimens were IA (idarubicin 10 mg/m²/day, d1-3; cytarabine 100 mg/m²/day, d1-7), GAG (aclacinomycin 20 mg/day, d1-4; cytarabine 10 mg/m² every 12 h for 7 to 14 days; granulocyte colony stimulating factor [G-CSF], 300 µg/day for 7 to 14 days) and HAA (homoharringtonine 2 mg/m²/day, d1-7; aclacinomycin 20 mg/day, d1-7; cytarabine 100 mg/m²/day, d1-7).

Consolidation therapy

Patients who achieved complete remission (CR) or CR with incomplete hematologic recovery (CRi) received four cycles of high-dose cytarabine (2 g/m² every 12 h for 3 days). Refractory or relapsed patients received high-dose cytarabine-based regimens, such as revised CLAG (cladribine 5 mg/m²/day, d1-5; cytarabine 1 g/m²/day, d1-5; G-CSF 300 µg/day, d1-5), FLAG (fludarabine 25 mg/m²/day, d1-5; cytarabine 1 g/m²/day, d1-5; G-CSF 300 µg/day, d1-5), or venetoclax combined with demethylating agents such as azacitidine or decitabine.

Since 2014, the *FLT3* inhibitor (*FLT3i*) sorafenib has been available in our center, and some patients with *FLT3-ITD* mutations received *FLT3i* therapy during induction, consolidation or relapse.

Patients who had *FLT3-ITD* mutation, who underwent morphological/molecular relapse, or who had a log reduction in *NPM1* MRD after consolidation 2 (MRD2) < 3 log were indicated for allogeneic hematopoietic stem cell transplantation (allo-HSCT); the allo-HSCT protocol was performed as previously described [14, 15].

Molecular and cytogenetic analysis

Blood from the bone marrow was analyzed according to the 2009 International System for Human Cytogenetic Nomenclature (ISCN) using the G-banding technique. Types A, B and D of *NPM1* mutation, as well as the rare type, and *NPM1* MRD were detected by real-time quantitative polymerase chain reaction (RT-PCR) [16] at diagnosis and each end of the chemotherapy cycle.

Targeted next-generation sequencing

The DNA was extracted from bone marrow samples obtained at the time of diagnosis. Following extraction and purification of genomic DNA from patient samples, specific regions are captured using hybridization probes or PCR primers designed to target these genes. Utilizing Next-Generation Sequencing (NGS) with targeted capture, we sequenced mutation hotspots or the complete coding regions of 139 genes that are frequently mutated in myeloid neoplasms (refer to Supplementary Table 1). The raw variant results were filtered based on the following criteria: an average effective sequencing depth on target per sample of at least 2,000x, a mapping quality score of at least 30, a base quality score of at least 30, and a variant allele frequency (VAF) of at least 1% for both single-nucleotide variants (SNVs) and small insertions-deletions (InDels). Variants are then annotated based on their potential impact on protein function, using databases like the 1000 Genomes Project, COSMIC, and clinical databases.

Definitions

The criteria for CR were as follows: bone marrow blasts < 5%; absence of circulating blasts and blasts with Auer rods; absence of extramedullary disease; absolute neutrophil count (ANC) $\geq 1.0 \times 10^9/L$; and platelet (PLT) count $\geq 100 \times 10^9/L$. CRi was defined as meeting all CR criteria except for ANC or PLT count. Relapse was defined as bone marrow blasts $\geq 5\%$, the reappearance of blasts in the blood, or the development of extramedullary disease [9]. Molecular relapse was defined according to the European LeukemiaNet (ELN) MRD Working Party [17]. Overall survival (OS) was calculated from the date of diagnosis to the date of death due to any cause. Event-free survival (EFS) was calculated from the date of CR/CRi until the date of morphological relapse, molecular relapse, or death from any cause. MRD2 was defined as the log reduction in the transcript level of NPM1 at the end of the second cycle of consolidation compared with the baseline.

Statistical analysis

Continuous variables are reported as medians and ranges, and categorical variables are expressed as percentages. Competing risk analysis was used to calculate the cumulative incidence of relapse (CIR), and Gray's test was used to test for differences between groups. Maximally selected rank statistics were calculated to determine the cutoff value of *NPM1* MRD for predicting survival. OS and EFS were calculated by the Kaplan–Meier method, and comparisons were made with the log-rank test. A Cox regression model was used for the analysis of prognosis. *P* values < 0.05 were considered to indicate statistical significance. Hazard ratios (HR) were calculated with their 95% confidence intervals (CI). Data analyses were primarily conducted with the Statistical Package for the Social Sciences (SPSS), version 25.0 (SPSS Inc., IBM Corp., Armonk, NY, USA), and R software, version 3.3.1 (R Foundation for Statistical Computing, Vienna, Austria), was used for competing risk analysis.

Results

Initial patient characteristics

A total of 302 patients were diagnosed with AML with *NPM1* mutation, and 234 patients for whom NGS data were available were included in this study (Fig. 1). The median age was 49 years (interquartile range [IQR] 36–58, and the median follow-up was 30.4 months (IQR 12.1–55.7). Forty genes were mutated > 1% of patients, and 9 genes were mutated > 10% of patients (Fig. 2A). *DNMT3A* was the most common co-mutation (53.8%), followed by *FLT3-ITD*

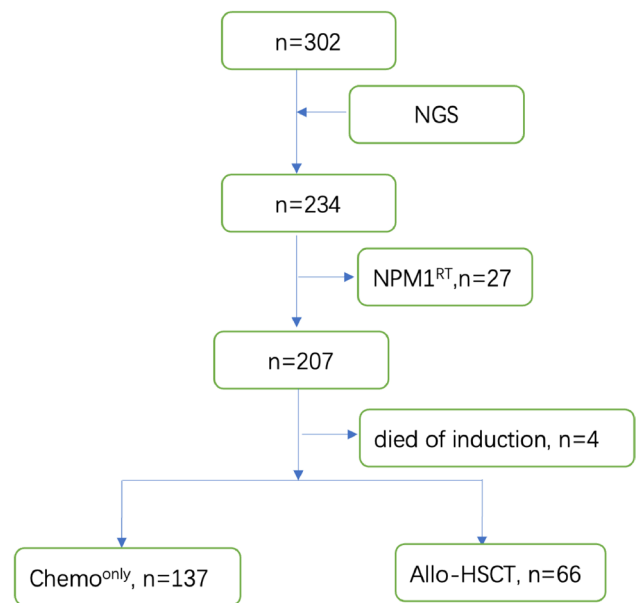


Fig. 1 Overview of patients included in this study. NGS: next-generation sequencing; Chemo^{only}: chemotherapy without allogeneic hematopoietic stem cell transplantation; Allo-HSCT: allogeneic hematopoietic stem cell transplantation; *NPM1*^{RT}: rare type of *NPM1* mutation. In patients with *NPM1*^{RT} mutation, 16 patients received chemotherapy only, and 11 patients received allo-HSCT. However, MRD of *NPM1*^{RT} were not available, these patients were not included except the analysis of baseline

(44.4%). The mutation interactions are shown in Fig. 2B. The pretreatment characteristics of patients are detailed in Table 1.

Treatment

Sorafenib

A total of 44.9% (93/207) of patients had *FLT3-ITD* mutation, and 32.3% (30/93) of those patients received *FLT3i* treatment. A total of 26.7% (8/30) of these patients received sorafenib only during induction chemotherapy, 10.0% (3/30) of patients received sorafenib from induction to consolidation therapy, and 6.7% (2/30) of patients received sorafenib from induction to maintenance therapy. A total of 40.0% (12/30) of patients used sorafenib from induction to allo-HSCT, and 6.7% (2/30) of patients used it from consolidation to allo-HSCT. A total of 10.0% (3/30) of patients received sorafenib when they relapsed.

Allo-HSCT

Among the 207 patients who received induction chemotherapy followed by consolidation chemotherapy, 31.9% (66/207) underwent allo-HSCT. The reasons for allo-HSCT

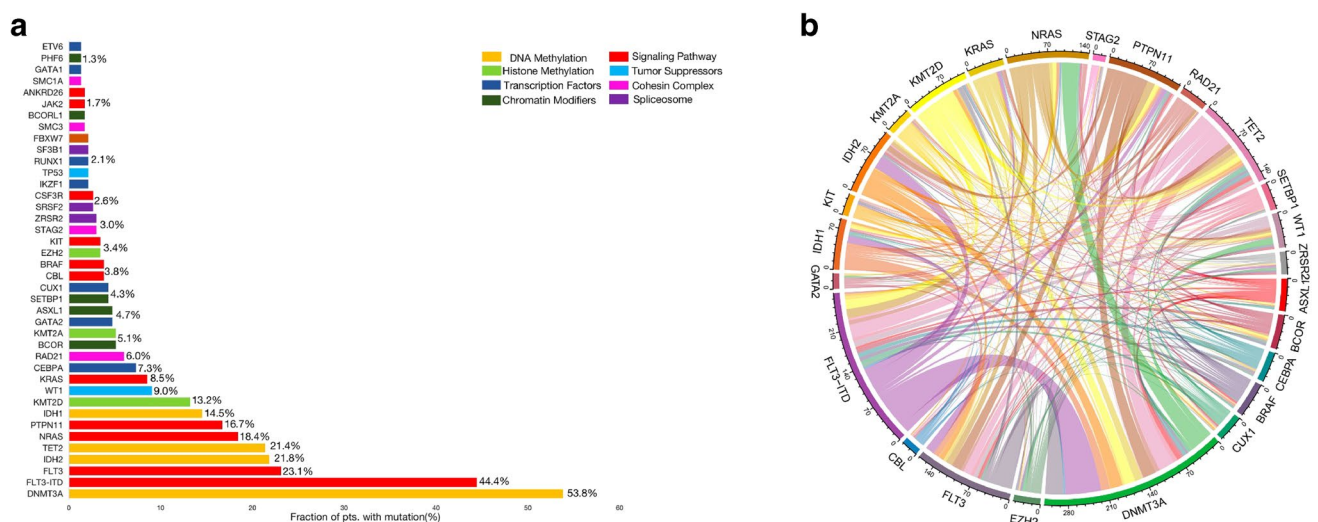


Fig. 2 Overview of gene mutations identified by targeted sequencing in 234 AML patients with *NPM1* mutation. **a**. Histogram showing the frequency of gene mutations detected in > 1% of patients. Bars are

colored according to the functional category assigned to each gene. The FLT3 mutation included 25(49.0%) FLT3-TKD. **b**. Association of gene co-mutations in AML patients with *NPM1* mutation

Table 1 Details about the pretreatment characteristics of patients

| | NGS (<i>n</i> = 234) |
|---|-----------------------|
| Gender, <i>n</i> (%) | |
| Male | 111(47.4%) |
| Female | 123(52.6%) |
| Age, median (IQR), years | 49(36–58) |
| Blast, median (IQR) (%) | 70.0(46–83) |
| WBC, median (IQR) × 10 ⁹ /L | 26.3(7.6–60.8) |
| HGB, median (IQR) g/L | 86(69–109) |
| PLT, median (IQR) × 10 ⁹ /L | 69(36–117) |
| <i>NPM1</i> rare type, <i>n</i> (%) | 27(11.5%) |
| Transcription level of <i>NPM1</i> , median (IQR) % | 27.90(13.78–50.00) |
| Karyotype, <i>n</i> (%) | |
| Normal | 180(76.9%) |
| Complex | 5 (2.1%) |
| Non-complex | 32(13.7%) |
| Unknown | 17(7.3%) |
| HSCT, <i>n</i> (%) | 77(32.9%) |

IQR interquartile range, WBC white blood cell, HGB hemoglobin, PLT platelet

in these 66 patients were *FLT3-ITD* mutation (54.5%, 36/66), molecular or morphological relapse (21.2%, 14/66), continuous positive MRD (16.7%, 11/66), MRD from negativity to positivity (4.5%, 3/66), and CR after 2 courses of induction (3.0%, 2/66). The median time from diagnosis to HSCT was 7.1 (IQR 5.8–10.8) months. The differences in baseline characteristics between the chemo^{only} group (not

including 4 patients who died early) and the allo-HSCT group are shown in Table 2.

Response, relapse and death

Overall, 207 patients received induction therapy, during which 1.9% (4/207) died of infection or hemorrhage; therefore, the CR/CRi rate was available for 203 patients in the cohort. The CR/CRi rate after one induction course was 76.8% (156/203), the cumulative CR/CRi rate after two induction courses was 91.6% (186/203), and the total CR/CRi rate was 91.6% (186/203), with a median age of 50.0 (IQR 35–58) years. The median age of the 17 patients who did not achieve CR/CRi was 55 (IQR 50–61) years.

A total of 44.6% (83/186) and 4.3% (8/186) of patients experienced morphological relapse and molecular relapse, respectively, and the median relapse times were 9.8 (IQR 4.1–18.9) months and 15.7 (IQR 12.3–25.5) months, respectively. Among the 83 patients with morphological relapse, 10.8% (9/83) relapsed after allo-HSCT. Overall, 9.6% (8/83) and 25% (2/8) of patients received a venetoclax-based regimen after morphological relapse and molecular relapse, respectively. The cumulative incidence of morphological relapse (CIRm) at 12 months, 24 months and 36 months was 25.8% (95% CI 19.8–32.3%), 40.5% (95% CI 33.3–47.6%) and 43.8% (95% CI 36.4–51.0%), respectively.

A total of 42.0% (87/207) of patients died, and 1.9% (4/207) of patients died during induction therapy. The survival rates at 12 months, 24 months and 36 months were 79.0% (95% CI 73.5–84.5%), 67.0% (95% CI 60.5–73.5%) and 60.3% (95% CI 53.2–67.3%), respectively.

Table 2 Baseline in chemo^{only} group and Allo-HSCT group

| | Chemo ^{only} (n = 137) | Allo-HSCT (n = 66) | p value |
|--|---------------------------------|--------------------|---------|
| Age, median (IQR), years | 54(46–61) | 39(30–52) | <0.001 |
| Gender, n (%) | | | 0.300 |
| Male | 62(45.3%) | 35(53.0%) | |
| Female | 75(54.7%) | 31(47.0%) | |
| FLT3-ITD, n (%) | 55(40.1%) | 36(54.5%) | 0.054 |
| DNMT3A, n (%) | 79(57.7%) | 38(57.6%) | 0.990 |
| FLT3-ITD and DNMT3A co-mutation, n (%) | 40(29.2%) | 20(30.3%) | - |
| Karyotype, n (%) | | | 0.751 |
| Normal | 105(76.6%) | 50(75.8%) | |
| Complex | 5(3.6%) | 0 | |
| Non-complex | 17(12.4%) | 11(16.7%) | |
| Unknown | 10(7.3%) | 5(7.6%) | |

IQR interquartile range

In the chemotherapy group, 46.0% (63/137) of the patients died due to relapse (63.5%, 40/63), no response to chemotherapy (25.4%, 16/63), infection (7.9%, 5/63), or hemorrhage (3.2%, 2/63). In the allo-HSCT group, 30.3% (20/66) of patients died, 35.0% (7/20) of whom died due to morphological relapse, 10.0% (2/20) due to no response to chemotherapy, and 55% (11/20) due to treatment-related mortality (TRM).

Among the 5 patients with complex karyotypes, 80% (4/5) achieved CR, and 75% (3/4) experienced morphological relapse, with relapse-free survival (RFS) of 20.6, 39.6, and 6.5 months and OS of 81.2, 45.1, and 8.5 months, respectively. One patient who achieved a consistent CR status was still alive at the end of the study, with an OS of 93.9 months. One patient who did not achieve CR died after 6.7 months.

Survival analysis

As *NPM1* MRD data for rare type mutations were not available, survival analysis was conducted for 137 patients who received chemotherapy only, excluding *NPM1*^{RT} patients.

Optimal cutoff value of MRD

MRD was analyzed in 137 chemo^{only} patients, and the optimal cutoff value for MRD2 reduction for both EFS ($p < 0.001$) and OS ($p < 0.001$) was a reduction ≥ 3 log. A reduction in MRD2 ≥ 3 log was significantly associated with increased EFS and OS compared with a reduction in MRD2 < 3 log ($p < 0.001$, $p < 0.001$) (Fig. 3).

Conversion of MRD status from negative to positive

NPM1 MRD in 60% (85/137) of patients converted to a negative status at a median time of 4.1 (IQR 2.8–5.9) months. MRD in 40.0% (34/85) of these patients converted from negative to positive (MRD^{neg to pos}) thereafter, with a median time of 12.5 (IQR 4.6–25.9) months, and the median transcription level of *NPM1* was 0.06% (IQR 0.01–0.57). Patients whose MRD was consistently negative (MRD^{consist neg}) had greater EFS (Fig. 4A) and OS (Fig. 4B) than patients with MRD^{neg to pos} (both $p < 0.001$).

Co-occurring mutations

Genes with a prevalence $\geq 10\%$ and those predicted to be at high risk by the 2022 ELN recommendation in the 137 chemo^{only} patients with NGS data are listed in Table 3. The most prevalent genes were *DNMT3A* (57.7%) and *FLT3-ITD* (40.1%).

Univariable and multivariable analyses of survival were performed for 137 chemo^{only} patients with NGS data. Univariate analysis revealed that age, *FLT3-ITD* co-mutation, *DNMT3A* co-mutation and a decrease in MRD2 < 3 log were associated with poorer EFS and OS, and the white blood cell (WBC) count only was a prognostic factor for poor OS. However, in multivariate analysis, *DNMT3A* co-mutation (EFS, HR = 1.9, $p = 0.021$; OS, HR = 2.2, $p = 0.023$) and MRD2 reduction ≥ 3 log (EFS, HR = 0.1; OS, HR = 0.1, both $p < 0.001$) were found to be independent prognostic factors for survival, and age was associated with lower EFS (HR = 1.3, $p = 0.013$) (Table 4).

The patients were divided into the following 4 groups according to the status of *DNMT3A* co-mutation and *FLT3-ITD* co-mutation: both *FLT3-ITD* and *DNMT3A* negative,

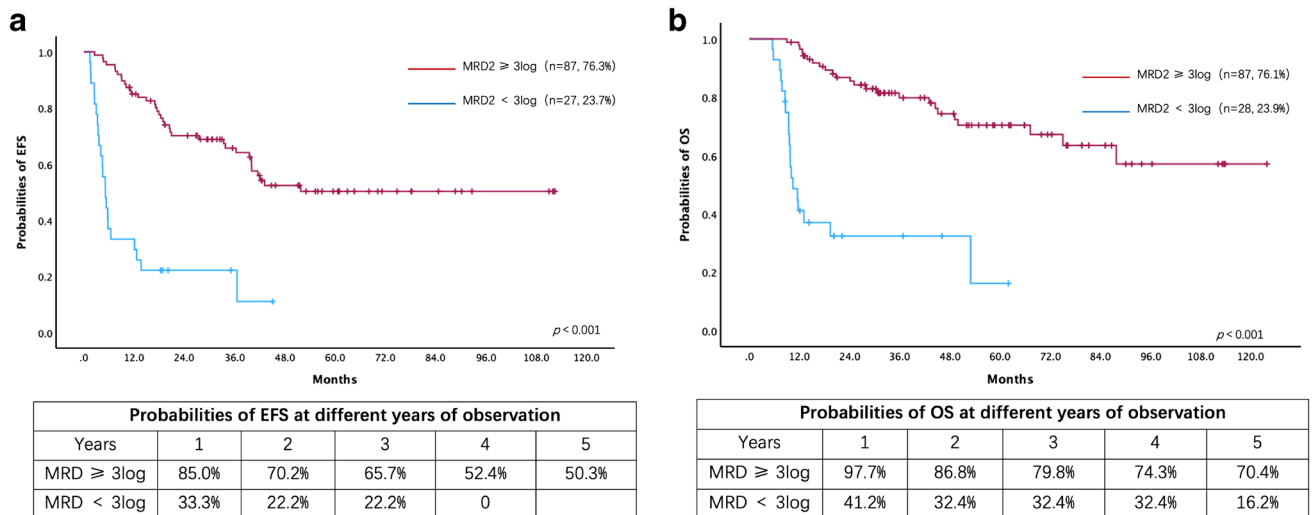


Fig. 3 Maximally selected rank statistics to determine the optimal cutoff values of MRD2 for predicting EFS and OS in 137 patients. **(a)** EFS; **(b)** OS

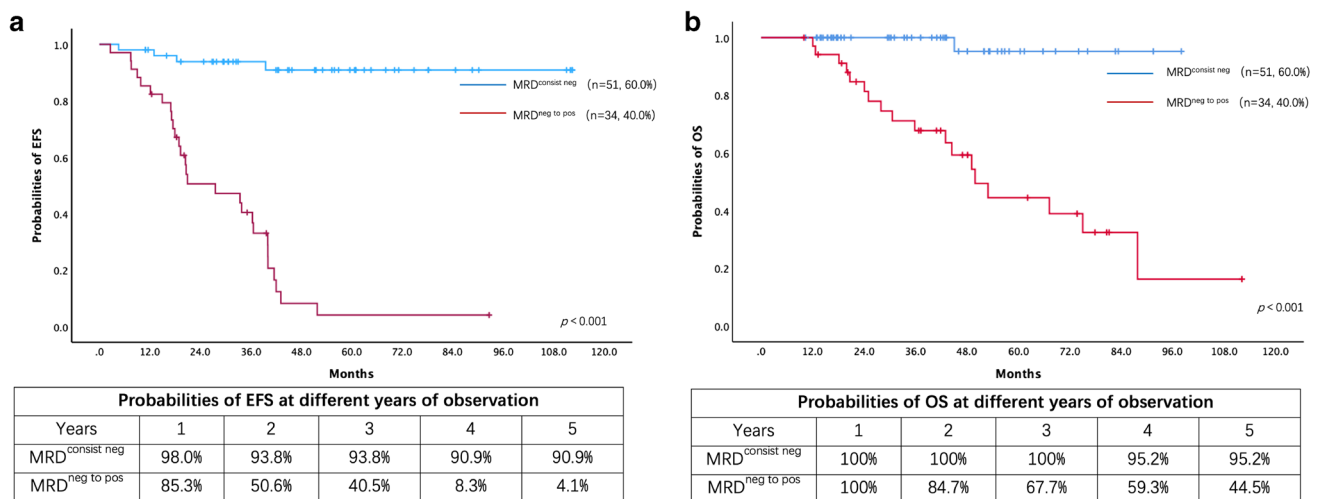


Fig. 4 Whether MRD of *NPM1* could be negative continuously or not predicted survival. **(a)** EFS; **(b)** OS. In 4 patients with complex karyotype who achieved CR, 3 patients got MRD2 negative and then converted to positivity, and 2 patients of them underwent morphology relapse

FLT3-ITD-positive and *DNMT3A*-negative (12.5% [2/16] of patients received sorafenib), *FLT3-ITD*-negative and *DNMT3A*-positive, and both *FLT3-ITD*- and *DNMT3A*-positive (35.0% [14/40] of patients received sorafenib). EFS was calculated in 89.8% (123/137) of patients who achieved CR/CRi. The co-occurrence of *FLT3-ITD* and *DNMT3A* mutations was associated with the lowest EFS and OS (3-year EFS rate 30.0%, $p < 0.001$; 3-year OS rate 34.4%, $p < 0.001$). However, patients with only the *FLT3-ITD* co-mutation did not have poorer survival than patients without the *FLT3-ITD* co-mutation (3-year EFS rate 71.4% vs. 72.8%, $p = 0.998$; 3-year OS rate 75.0% vs. 74.6%, $p = 0.789$) (Fig. 5).

Re-stratifying the risk groups

The favorable, intermediate, and poor-risk groups were re-stratified according to the status of *FLT3-ITD* and *DNMT3A* co-mutations and *NPM1* MRD2 reduction. The poor-risk group comprised patients with a < 3 log reduction in MRD2 or with both *FLT3-ITD* and *DNMT3A* co-mutations achieved the lowest EFS (3-year EFS rate 30.0%) and OS (3-year OS rate 34.4%). The favorable-risk group comprised patients with *FLT3-ITD*^{neg} + *DNMT3A*^{neg} and an MRD2 ≥ 3 log reduction, with a 3-year EFS rate of 80.8% and a 3-year OS rate of 90.3% (Fig. 6).

Table 3 Genes with prevalence $\geq 10\%$ and genes predicting high risk classification

| Mutation detected | Prevalence, n (%) N=137 |
|-------------------|----------------------------|
| <i>DNMT3A</i> | 79(57.7%) |
| <i>FLT3-ITD</i> | 55(40.1%) |
| <i>FLT3*</i> | 37(27.0%) |
| <i>FLT3-TKD</i> | 20(14.6%) |
| <i>TET2</i> | 32(23.4%) |
| <i>IDH2</i> | 31(22.6%) |
| <i>NRAS</i> | 25(18.2%) |
| <i>PTPN11</i> | 22(16.1%) |
| <i>KMT2D</i> | 19(13.9%) |
| <i>IDH1</i> | 18(13.1%) |
| <i>KRAS</i> | 15(10.9%) |
| <i>ASXL1</i> | 8(5.8%) |
| <i>ZRSR2</i> | 5(3.6%) |
| <i>BCOR</i> | 5(3.6%) |
| <i>CSF3R</i> | 4(2.9%) |
| <i>EZH2</i> | 4(2.9%) |
| <i>SRSF2</i> | 4(2.9%) |
| <i>SF3B1</i> | 3(2.2%) |
| <i>STAG2</i> | 3(2.2%) |
| <i>TP53</i> | 2(1.5%) |
| <i>RUNX1</i> | 2(1.5%) |
| <i>U2AF1</i> | 0 |

**FLT3* mutation except *FLT3-ITD* mutation

HSCT improved the survival of poor-risk patients

A total of 29.6% (60/203) of the 203 patients who had NGS data, excluding those with *NPM1*^{RT} or who died early, were classified into the poor-risk group. A total of 66.7% (40/60)

of patients received chemotherapy only, and 33.3% (20/60) of patients underwent allo-HSCT. HSCT improved EFS (3-year EFS rate 57.9% vs. 30.0%, $p=0.012$) compared with patients who received chemotherapy only in 81.7% (49/60) of patients who achieved CR/CRi (Fig. 7a). Additionally, the OS in the allo-HSCT group was greater than that in the chemotherapy group (3-year OS rate 72.0% vs. 34.4%, $p=0.001$) (Fig. 7b).

Discussion

AML with *NPM1* mutation is the most common subtype of adult AML and has relatively strong heterogeneity [9]. Risk stratification of AML patients with *NPM1* mutation according to international guidelines was mainly based on whether an *FLT3-ITD* co-mutation was present, namely, the favorable-risk group included patients without an *FLT3-ITD* co-mutation, and the intermediate-risk group included patients with an *FLT3-ITD* co-mutation [8]. Only approximately 3.4% of patients have chromosomal abnormalities that cause adverse risk [18] and are categorized into the adverse-risk group [8].

NPM1 mutation is an ideal target for MRD monitoring to predict relapse. However, different centers have defined different prognostic thresholds and time points of *NPM1* MRD. In the study by Max et al. of 158 patients from the AMLCG 1999, 2004 and 2008 trials, a reduction of 3 log in *NPM1* MRD after induction or consolidation therapy was the cutoff for predicting relapse ($p=0.001$, $p=0.001$) [19]. In the ALFA-0701 trial, patients with *NPM1* MRD positivity (defined as $>0.1\%$ in the bone marrow) after induction and at the end of treatment also had a greater risk of relapse, but OS did not differ [20]. Balsat et al. also studied the relationship between peripheral blood (PB) *NPM1* MRD and relapse

Table 4 Univariable and multivariable analysis for survival

| Variables | Univariable analysis | | | | Multivariable analysis | | | |
|--------------------------------|----------------------|----------|--------------|----------|------------------------|----------|--------------|----------|
| | EFS | | OS | | EFS | | OS | |
| | HR (95%CI) | P Value | HR (95%CI) | P Value | HR (95%CI) | P Value | HR (95%CI) | P Value |
| Age | 1.3(1.1–1.6) | 0.012 | 1.2(1.0–1.5) | 0.044 | 1.3(1.1–1.7) | 0.013 | | |
| BM blast | 1.0(0.9–1.2) | 0.686 | 1.1(0.9–1.2) | 0.384 | | | | |
| WBC | 1.0(1.0–1.1) | 0.157 | 1.1(1.0–1.1) | 0.012 | | | | |
| HGB | 1.0(0.9–1.1) | 0.999 | 1.0(0.9–1.1) | 0.470 | | | | |
| PLT | 1.3(0.9–1.8) | 0.206 | 1.2(0.8–1.7) | 0.386 | | | | |
| ≥ 3 log reduction in MRD2 | 0.2(0.1–0.3) | <0.001 | 0.1(0.1–0.3) | <0.001 | 0.2(0.1–0.3) | <0.001 | 0.2(0.1–0.3) | <0.001 |
| <i>FLT3-ITD</i> | 1.7(1.0–2.7) | 0.042 | 2.1(1.3–3.5) | 0.003 | | | 1.9(1.0–3.6) | 0.062 |
| <i>FLT3-TKD</i> | 1.6(0.9–3.0) | 0.114 | 1.7(0.9–3.1) | 0.078 | | | 1.7(0.9–3.1) | 0.018 |
| <i>DNMT3A</i> | 2.7(1.6–4.6) | <0.001 | 2.7(1.6–4.8) | <0.001 | 1.9(1.1–3.4) | 0.021 | 2.1(1.1–4.2) | 0.031 |

Univariable analysis of *TET2*, *IDH2*, *NRAS*, *PTPN11*, *KMT2D*, *IDH1* and *KRAS* were showed in Supplementary Table 2

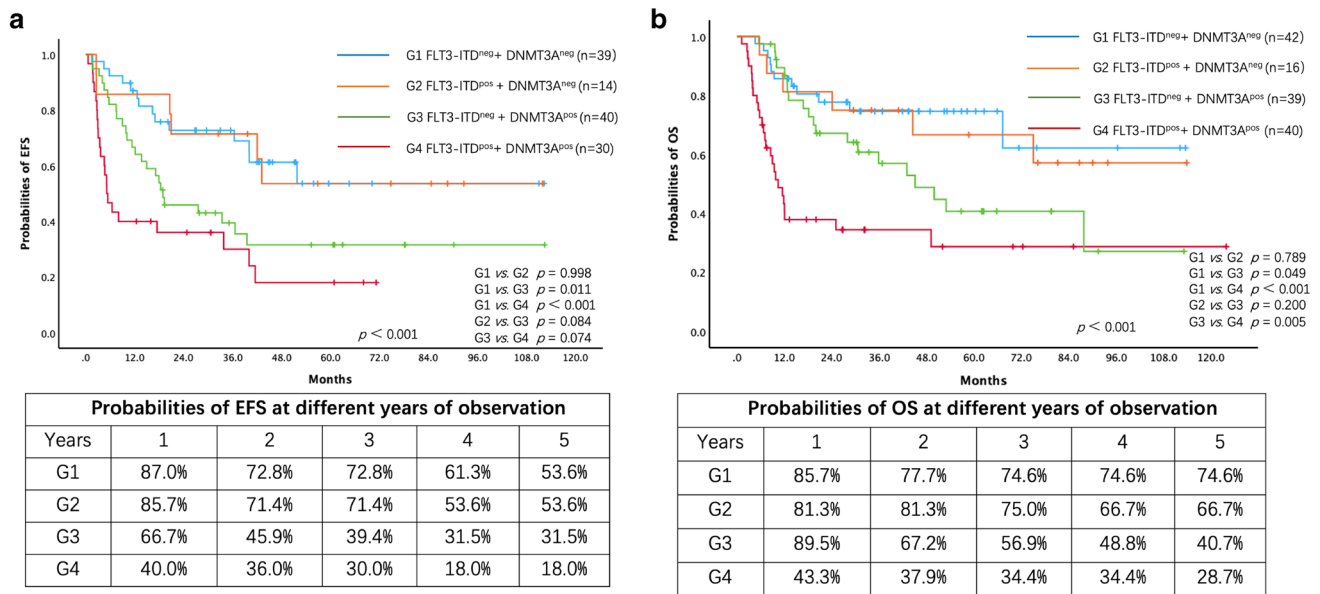


Fig. 5 Survival of co-occurrent mutation status of FLT3-ITD and DNMT3A. **(a)** EFS, **(b)** OS. neg negative, pos positive. 7.1% (1/14) patient and 43.3% (13/30) patients received Sorafenib in G2 and G4

respectively in **(a)**. 12.5% (2/16) patients and 35% (14/40) patients received Sorafenib in G2 and G4 respectively in **(b)**

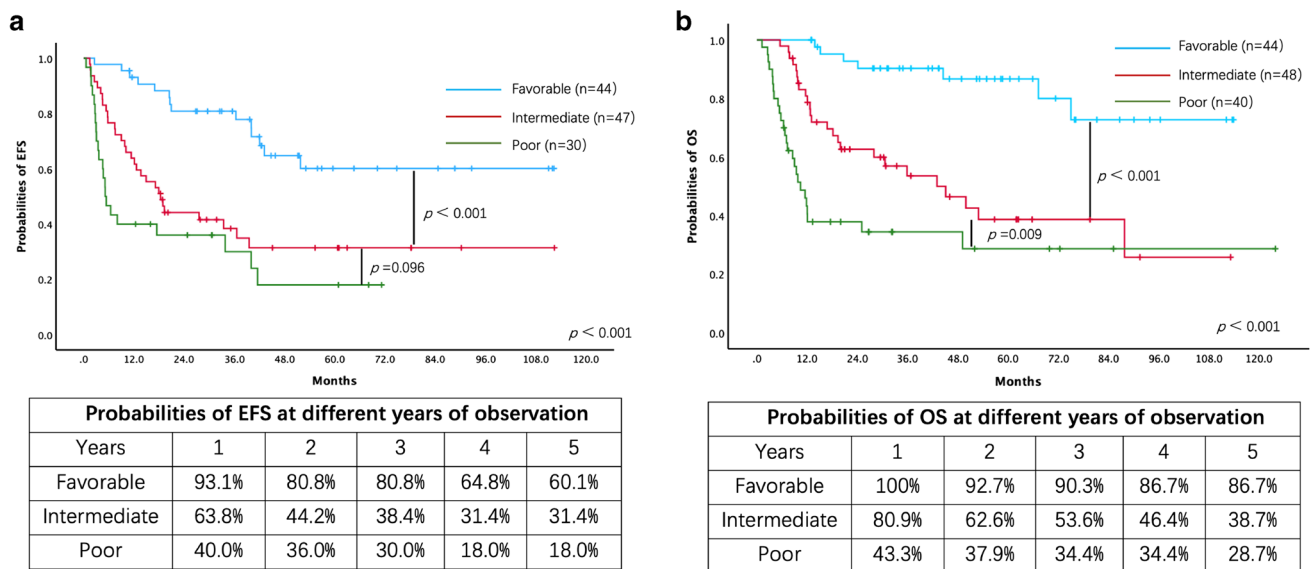


Fig. 6 Survival of patients re-stratified by the status of FLT3-ITD and DNMT3A mutation and NPM1 reduction of MRD2. **(a)** EFS, **(b)** OS. Favorable risk group, patients with FLT3-ITD^{neg} + DNMT3A^{neg} and MRD2 \geq 3log reduction. Poor risk group, FLT3-ITD^{pos} + DNMT3A^{pos}

or MRD2 $<$ 3log reduction. Intermediate risk group, neither favorable nor poor risk group. In **(a)**, the percentage of patients received Sorafenib were 2.3% (1/44), 0(0/47), 43.3% (13/30) in three different risk groups, while 2.3% (1/44), 0(0/48), 35.0% (14/40) in **(b)**

or OS, and a $>$ 4 log reduction in PB MRD after induction therapy was significantly associated with a lower rate of CIR and shorter OS [10]. Our center's results are consistent with our previously published data, in which a reduction in MRD2 \geq 3 log was associated with increased disease-free survival and OS [21]. Moreover, Ivey et al. confirmed

that a positive NPM1 MRD in the PB after the second chemotherapy course was associated with a greater risk of relapse in patients in the AML17 trial (3-year CIR rate 82% vs. 30%, $p < 0.001$) [22]. Fabio Guolo et al. reported that in 19% (8/42) of AML patients with NPM1 mutation who experienced morphological relapse, the recurrence of

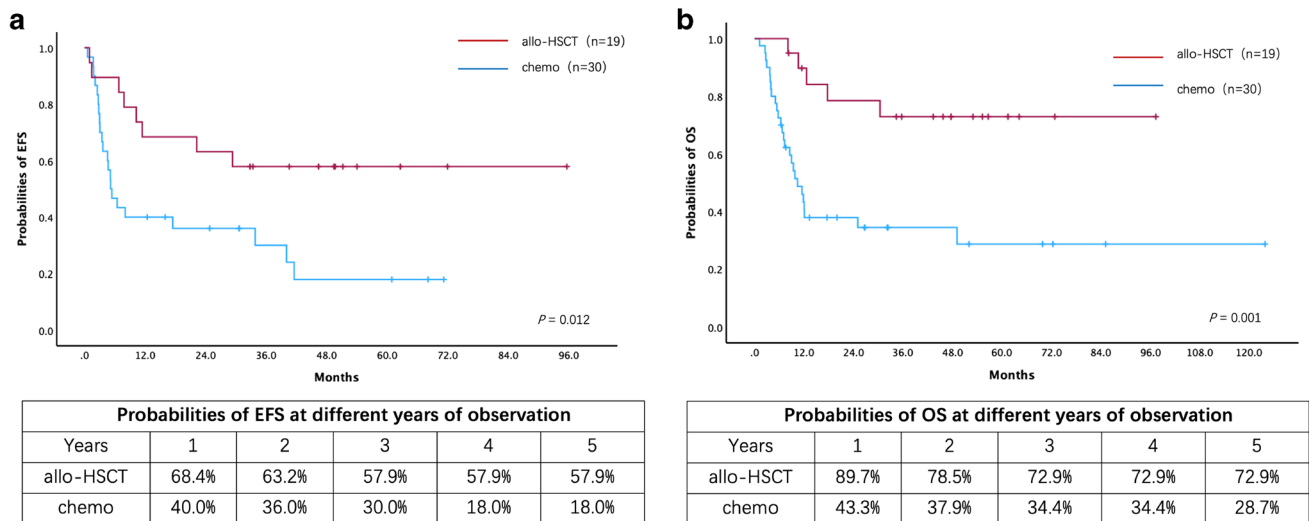


Fig. 7 Survival of determined as poor risk group was improved by HSCT. (a) EFS; (b) OS

NPM1 MRD contributed to relapse, with a median time of 4.5 months [23]. However, our study aimed to explore the significance of the shift from negative to positive MRD, rather than simply comparing negative and positive MRD. Univariable analysis in our study demonstrated that patients with continuously negative *NPM1* MRD had higher EFS and OS than those patients who shifted from negative to positive MRD, but this finding still needs to be confirmed via multivariable analysis.

However, the prognosis of AML patients with *NPM1* mutation could be affected by multiple factors, since the relapse rate was not very low. More co-mutations have been identified in AML patients with *NPM1* mutation than in any other subtype of AML. Accumulating evidence shows that the outcome of AML patients with *NPM1* mutation may vary because of co-mutations other than the *FLT3-ITD* mutation. AML patients with *NPM1* and *FLT3-ITD* co-mutations were classified into the intermediate-risk group [8]; however, the survival of AML patients with *NPM1* and *FLT3-ITD* co-mutations was similar to that of AML patients with only *NPM1* mutations in our study. In our study, *DNMT3A* co-mutations were found in 53.8% of AML patients with *NPM1* mutation, while *FLT3-ITD* co-mutations were found in 44.4%. In the *FLT3-ITD* co-mutation group, 61.5% of patients were *FLT3-ITD*^{POS} + *DNMT3A*^{POS}; therefore, it was hypothesized that these patients contributed to the poor survival rates rather than the patients with *FLT3-ITD* mutation alone. Specifically, only patients with both *FLT3-ITD* and *DNMT3A* co-mutations had a poor prognosis. Elli Papaemmanuil et al. also reported that the deleterious effect of *FLT3-ITD* mutation was most clinically relevant in patients with concomitant *NPM1* and *DNMT3A* co-mutations [24], which

were shown to be associated with a high leukemia stem cell frequency and synergistic upregulation of specific leukemia stem cell regulators [25]. The co-occurrence of *NPM1/FLT3-ITD/DNMT3A* mutations was associated with decreased OS and disease-free survival [25]. Furthermore, a link between the co-occurrence of *NPM1/FLT3-ITD/DNMT3A* mutations and AML resistance to anthracycline has been identified in functional studies [26]. The effect of *FLT3-ITD* mutation on survival was less obvious when it co-occurred with either an *NPM1* or *DNMT3A* mutation alone [24].

Other co-mutations, such as *TET2*, *PTPN11*, *IDH1*, *IDH2* and *NRAS* had no prognostic on OS, which were coincident with the latest report including 1357 patients [27]. However, *FLT3-TKD* was controversial. In the 1357 patients' study and a recent study from China, *FLT3-TKD* did not influence OS [27, 28], other study verified it improved OS [29, 30], and our study found it decreased OS. Maybe different treatment background such as FLT3i may change the outcome.

In addition, we combined MRD and *NPM1* co-mutations to achieve a more accurate risk re-stratification of AML patients with *NPM1* mutation. Maël Heiblig et al. reported that in patients aged older than 60 years (median age 66.1 years), patients with co-occurring *NPM1/FLT3-ITD/DNMT3A* mutations, or with co-occurring *NPM1/DNMT3A* mutations and MRD reduction < 4 log after the first cycle of induction were classified as poor risk, with a median of 7.7 months of leukemia-free survival [11]. Our study verified that the co-occurrence of *NPM1/FLT3-ITD/DNMT3A* mutations or an MRD2 reduction < 3 log could be used to identify the poor-risk group among younger elderly patients with a median age of 54 years. Moreover, the conversion from negative to

positive MRD status was also associated with poor outcomes, but multivariate analysis needs to be performed for verification.

More than half of the patients were diagnosed before FLT3i were widely used; furthermore, owing to the financial limitations of using FLT3i at that time, only a small proportion of patients received sorafenib during treatment—11.7% (16/137) of patients in the chemo^{only} group and 21.2% (14/66) of patients in the HSCT group. However, in our survival analysis, more patients received sorafenib in the *FLT3-ITD*^{pos} + *DNMT3A*^{pos} group than in the *FLT3-ITD*^{pos} + *DNMT3A*^{neg} group (14 patients vs. 2 patients), and the prognosis of patients in the *FLT3-ITD*^{pos} + *DNMT3A*^{pos} group was still poor. In the current era of targeted therapy, the Bcl-2 inhibitor venetoclax is effective for treating AML with *NPM1* mutation [31], and our previous data also suggested that the rate of CR/CRi after one induction cycle was 77.8% (14/18) [32]. Venetoclax-based low-intensity chemotherapy results in 84% of molecular failure.

AML patients with *NPM1* mutation achieved an MRD response, and 71% of these patients became MRD negative [33]. In refractory/relapsed (R/R) AML patients with *NPM1* mutation, venetoclax combination therapy has a greater response rate of CR/CRi (46%) [34]. In our study, a total of 10 patients received venetoclax-based therapy after molecular or morphological relapse, which prolonged survival. Sorafenib plus triple therapy with venetoclax, azacitidine, and homoharringtonine (VAH) was well tolerated and highly effective against R/R AML with *FLT3-ITD* mutation [35], and venetoclax- and FLT3i-based therapy may be more suitable and more effective for treating AML patients with *NPM1* mutation who have *FLT3-ITD* and *DNMT3A* co-mutations.

In conclusion, the deleterious effect of *FLT3-ITD* mutation is more pronounced when concomitant with *DNMT3A* mutation in AML patients with *NPM1* mutation. A < 3 log reduction in MRD2 was also an independent prognostic factor for poor survival, which could be improved by allo-HSCT. In addition, the shift from negative to positive MRD status was also associated with poor EFS and OS according to the univariable analysis.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00277-024-06001-6>.

Acknowledgements We thank medical staff and patients' participants.

Author contributions Hao Jiang and Xiaojun Huang designed the study. Wenbing Duan collected and analyzed the data. Hao Jiang, Wenbing Duan prepared the typescript. All authors approved the final typescript, take responsibility for the content, and agreed to submit for publication.

Funding This work was funded by The Grants from the Beijing Municipal Science and Technology Commission (Z221100007422008), Peking University People's Hospital Research and Development Funds

(RDL2022-13), Peking University Medicine Fund for world's leading discipline or discipline cluster development (No.71003Y3035).

Data availability No datasets were generated or analysed during the current study.

Declarations

This study was approved by the Ethics Committee of Peking University People's Hospital (approval number 2022PHD019-001, 2023PHB164-001) and was conducted according to the principles of the Helsinki Declaration. Informed consent was obtained from all individual participants included in the study. The authors declare no known no potential conflicts of interest.

Conflict of interest The authors declare no competing interests.

References

1. Thiede C et al (2006) Prevalence and prognostic impact of *NPM1* mutations in 1485 adult patients with acute myeloid leukemia (AML). *Blood* 107(10):4011–4020
2. Falini B et al (2005) Cytoplasmic nucleophosmin in acute myelogenous leukemia with a normal karyotype. *N Engl J Med* 352(3):254–266
3. Patel JP et al (2012) Prognostic relevance of integrated genetic profiling in acute myeloid leukemia. *N Engl J Med* 366(12):1079–1089
4. Polyatskin IL, Artemyeva AS, Krivolapov YA (2019) Revised WHO classification of tumors of hematopoietic and lymphoid tissues, 2017 (4th edition):lymphoid tumors. *Arkh Patol* 81(3):59–65
5. Döhner K et al (2005) Mutant nucleophosmin (*NPM1*) predicts favorable prognosis in younger adults with acute myeloid leukemia and normal cytogenetics: interaction with other gene mutations. *Blood* 106(12):3740–3746
6. Schnittger S et al (2005) Nucleophosmin gene mutations are predictors of favorable prognosis in acute myelogenous leukemia with a normal karyotype. *Blood* 106(12):3733–3739
7. Schlenk RF et al (2008) Mutations and treatment outcome in cytogenetically normal acute myeloid leukemia. *N Engl J Med* 358(18):1909–1918
8. Döhner H et al (2022) Diagnosis and management of AML in adults: 2022 ELN recommendations from an international expert panel on behalf of the ELN. *Blood* 140:1345–1377
9. Döhner H et al (2017) Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood* 129(4):424–447
10. Balsat M et al (2017) Postinduction minimal residual disease predicts outcome and benefit from allogeneic stem cell transplantation in acute myeloid leukemia with *NPM1* mutation: a study by the Acute Leukemia French Association Group. *J Clin Oncol* 35(2):185–193
11. Heiblig M et al (2021) The impact of *DNMT3A* Status on *NPM1* MRD predictive value and survival in elderly AML patients treated intensively. *Cancers (Basel)* 13(9):2156
12. 中华医学会血液学分会, 成人急性髓系白血病(非急性早幼粒细胞白血病)中国诊疗指南(2017年版). *Chinese J Hematol* 2011. 32: 804–807
13. Döhner H et al (2010) Diagnosis and management of acute myeloid leukemia in adults: recommendations from an international expert panel, on behalf of the European LeukemiaNet. *Blood* 115(3):453–474
14. Xu L et al (2018) The consensus on indications, conditioning regimen, and donor selection of allogeneic hematopoietic cell

- transplantation for hematological diseases in China—recommendations from the Chinese Society of Hematology. *J Hematol Oncol* 11(1):33
15. Lu DP et al (2006) Conditioning including antithymocyte globulin followed by unmanipulated HLA-mismatched/haploidentical blood and marrow transplantation can achieve comparable outcomes with HLA-identical sibling transplantation. *Blood* 107(8):3065–3073
 16. Ruan GR et al (2009) Nucleophosmin mutations in Chinese adults with acute myelogenous leukemia. *Ann Hematol* 88(2):159–166
 17. Schuurhuis GJ et al (2018) Minimal/measurable residual disease in AML: a consensus document from the European LeukemiaNet MRD Working Party. *Blood* 131(12):1275–1291
 18. Angenendt L et al (2019) Chromosomal abnormalities and prognosis in NPM1-Mutated Acute Myeloid Leukemia: a pooled analysis of individual patient data from nine international cohorts. *J Clin Oncol* 37(29):2632–2642
 19. Hubmann M et al (2014) Molecular response assessment by quantitative real-time polymerase chain reaction after induction therapy in NPM1-mutated patients identifies those at high risk of relapse. *Haematologica* 99(8):1317–1325
 20. Lambert J et al (2014) MRD assessed by WT1 and NPM1 transcript levels identifies distinct outcomes in AML patients and is influenced by gemtuzumab ozogamicin. *Oncotarget* 5(15):6280–6288
 21. Zhao T et al (2017) Prognostic significance of early assessment of minimal residual disease in acute myeloid leukemia with mutated NPM1 patients. *Zhonghua Xue Ye Xue Za Zhi* 38(1):10–16
 22. Ivey A et al (2016) Assessment of minimal residual disease in standard-risk AML. *N Engl J Med* 374(5):422–433
 23. Guolo F et al (2019) Longitudinal minimal residual disease (MRD) evaluation in acute myeloid leukaemia with NPM1 mutation: from definition of molecular relapse to MRD-driven salvage approach. *Br J Haematol* 186(6):e223–e225
 24. Papaemmanuil E et al (2016) Genomic classification and prognosis in Acute Myeloid Leukemia. *N Engl J Med* 374(23):2209–2221
 25. Bezerra MF et al (2020) Co-occurrence of DNMT3A, NPM1, FLT3 mutations identifies a subset of acute myeloid leukemia with adverse prognosis. *Blood* 135(11):870–875
 26. Guryanova OA et al (2016) DNMT3A mutations promote anthracycline resistance in acute myeloid leukemia via impaired nucleosome remodeling. *Nat Med* 22(12):1488–1495
 27. Othman J et al (2024) Molecular, clinical and therapeutic determinants of outcome in NPM1 mutated AML. *Blood* 144:714–728
 28. Yao Y et al (2024) Co-mutation landscape and its prognostic impact on newly diagnosed adult patients with NPM1-mutated de novo acute myeloid leukemia. *Blood Cancer J* 14(1):118
 29. Perry M et al (2018) FLT3-TKD Mutations Associated With NPM1 Mutations Define a Favorable-risk Group in Patients With Acute Myeloid Leukemia. *Clin Lymphoma Myeloma Leuk* 18(12):e545–e550
 30. Boddu P et al (2017) Co-occurrence of FLT3-TKD and NPM1 mutations defines a highly favorable prognostic AML group. *Blood Adv* 1(19):1546–1550
 31. DiNardo CD et al (2020) Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. *N Engl J Med* 383(7):617–629
 32. Yu WJ et al (2022) Short-term efficacy of venetoclax combined with azacitidine in acute myeloid leukemia: a single-institution experience. *Zhonghua Xue Ye Xue Za Zhi* 43(2):134–140
 33. Jimenez-Chillon C et al (2024) Venetoclax-based low intensity therapy in molecular failure of NPM1-mutated AML. *Blood Adv* 8(2):343–352
 34. Stahl M et al (2021) Clinical and molecular predictors of response and survival following venetoclax therapy in relapsed/refractory AML. *Blood Adv* 5(5):1552–1564
 35. Yu S et al (2024) Sorafenib plus triplet therapy with venetoclax, azacitidine and homoharringtonine for refractory/relapsed acute myeloid leukemia with FLT3-ITD: A multicenter phase 2 study. *J Intern Med* 295(2):216–228

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.