## **ORIGINAL ARTICLE**



# **Advertising and receiving from heights increases transmission of vocalizations in semi‑arboreal mice**

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Received: 11 January 2023 / Revised: 14 June 2023 / Accepted: 20 June 2023 / Published online: 5 July 2023 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2023

# **Abstract**

Many animals produce long-distance acoustic signals to mediate a variety of social interactions, and the efficacy of transmission depends in part on environmental attenuation. Vocalizing from positions that optimize transmission is one key solution to minimizing attenuation, though few studies assess the magnitude of this efect in relation to receiver position. In this study, we assessed how transmission of high-frequency vocalizations produced by pinyon mice (*Peromyscus truei*) varied based on the position of senders and receivers. Pinyon mice are semi-arboreal rodents that produce sustained vocalizations to advertise to conspecifcs. Synthesized signals derived from a population-sample-average of fundamental frequency, duration, and amplitude were broadcast and recorded at diferent heights (0, 1, and 2 m) and distances (1, 2, 4, and 8 m) in a full factorial design to mimic hypothetical senders and receivers. We also measured receiver hearing sensitivity using auditory brainstem responses (ABR) to quantify the audible distance (active space) of vocalizations at diferent heights. Vocalizations showed less attenuation when emitted and received from an elevation compared to the ground if the signal was received at least 4 m from the sender. Vocalizations emitted from a 1 m height had an approximately 3 times greater audible distance compared to the ground. Additionally, less attenuation occurred when both senders and receivers were elevated at the same height and when receivers were elevated, regardless of sender height. Our results highlight the importance of considering receiver position in animal communication, especially when senders produce highly directional signals.

## **Signifcance statement**

Vocalizing animals often position themselves in locations that maximize sound transmission. However, the magnitude of this efect is not often quantifed, especially in relation to the position of intended receivers. In this study, we combined acoustic recording, hearing experiments, and modelling of sound attenuation to quantify how sending and receiving vocalizations from trees impacts sound transmission in a semi-arboreal mouse. We found that vocalizations produced from 1 m above the ground could be heard by receivers at 3 times the distance compared to ground level. We also found that no matter the sender position, receivers beneftted from being at elevated positions. Finally, we found that the least attenuation occurred when senders and receivers were elevated at the same height. Our results highlight the importance of considering receiver position in animal communication, especially when senders produce highly directional signals.

**Keywords** Acoustic communication · Environmental attenuation · Active space · Hearing · *Peromyscus* · Pinyon mice

Communicated by: E. Korpimäki.

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# **Introduction**

Many animals produce long-distance acoustic signals to mediate a variety of social interactions (Ryan and Kime  $2003$ ). The efficacy of acoustic signal transmission depends on the environment (Marten and Marler [1977\)](#page-8-1). Atmospheric absorption, ground attenuation, and defection of sound by vegetation contribute to signal attenuation (Marten and

Marler [1977](#page-8-1); Waser and Brown [1986](#page-9-0)). Coping with environmental attenuation is therefore a major factor shaping the evolution of long-distance acoustic signaling (Römer and Lewald [1992](#page-8-2); Boncoraglio and Saino [2007\)](#page-7-0). Two key solutions to minimizing attenuation involve producing signals that are resistant to degradation (i.e. sensory drive or acoustic adaptation and associated receiver adaptations; Obrist et al. [1993](#page-8-3); Ey and Fischer [2009](#page-7-1); Römer [2020\)](#page-8-4), or signaling at times and/or positions that optimize transmission (Waser and Waser [1977](#page-9-1); Barker and Mennill [2009\)](#page-7-2). Although studies on acoustic adaptation are numerous (e.g. Boncoraglio and Saino [2007;](#page-7-0) Goutte et al. [2018](#page-7-3)), comparatively fewer studies have explored the behavioral mechanisms used to minimize attenuation.

Animals can minimize signal degradation by advertising during times when climatic conditions are optimal for sound propagation (Wiley and Richards [1978;](#page-9-2) Hayes and Huntly [2005\)](#page-7-4). For example, the dawn chorus of birds and primates and the evening chorus of insects and frogs are hypothesized to occur due to favorable atmospheric conditions; low temperatures and high relative humidity during these periods are presumed to reduce energy loss via absorption (Marten and Marler [1977](#page-8-1); Wiley and Richards [1978\)](#page-9-2). Similarly, animals may position themselves in a manner that increases signal propagation (Blumenrath and Dabelsteen [2004\)](#page-7-5). Among taxa as diverse as crickets (Arak and Eiriksson [1992\)](#page-7-6), treefrogs (Schwartz et al. [2015\)](#page-8-5), and monkeys (Riondato et al. [2021\)](#page-8-6), senders advertise from elevated positions to minimize attenuation from the ground and understory vegetation. Receivers may also benefit from adjusting their position relative to senders to improve signal detection. Indeed, playback of birdsong indicates that receivers benefit from selection of high perches to optimize signal reception (Dabelsteen et al. [1993;](#page-7-7) Holland et al. [1998;](#page-7-8) Mathevon et al. [2005](#page-8-7)). However, few studies have quantified the magnitude of signal efficacy sender and receiver position in tandem, especially in species that produce directional signals that characterize high-frequency vocalizations of many rodents (Richards and Wiley [1980](#page-8-8); Fernández-Vargas et al. [2022](#page-7-9)).

Deer mice (genus *Peromyscus*) are a widespread and diverse group of rodents that exhibit a wide range of social behaviors, mating systems, and habitat use (Smartt [1978](#page-9-3); Kalcounis-Rueppell et al. [2018a,](#page-8-9) [c](#page-8-10)). Pinyon mice (*P. truei*) of the southwestern United States are among the most specialized species in the genus due to their reliance on pinyon (*Pinus* spp.) and juniper (*Juniperus* spp.) trees for food (Ribble and Samson [1987\)](#page-8-11) and nest sites (Hall and Morrison [1997](#page-7-10)). Of particular importance, pinyon mice possess morphological adaptations thought to be suited for a semi-arboreal lifestyle, including long tails to facilitate balance (Horner [1954;](#page-7-11) Smartt and Lemen [1980;](#page-9-4) Kingsley

et al. [2017](#page-8-12); Hager and Hoekstra [2021](#page-7-12)) and larger brains to navigate spatially complex environments (Lemen [1980](#page-8-13); Camargo et al. [2019](#page-7-13); but see Mace and Eisenberg [1982](#page-8-14)). Similar to other congeners, pinyon mice produce a variety of vocalizations for social communication, including ca. 20 kHz sustained vocalizations (SVs) that function over long distances (> 1 m; Kalcounis-Rueppell et al. [2018b,](#page-8-15) [c\)](#page-8-10). SVs appear to function as general advertisement signals that help maintain territories (e.g. *P. boylii*; Petric and Kalcounis-Rueppell [2013\)](#page-8-16) and/or spatial cohesion between mated pairs (*P. californicus*; Briggs and Kalcounis-Rueppell [2011](#page-7-14)). In pinyon mice, both sexes produce SVs in social isolation to advertise their presence to conspecifcs, but females call more often (Kobrina et al. [2022a\)](#page-8-17). Given that SVs operate over large spatial scales, quantifying how the geometry of senders and receivers infuences signal transmission is fundamental to understanding the ecology and evolution of communication in rodents.

In this study, we investigated if signaling and receiving from trees confers benefts for acoustic communication in pinyon mice. Specifcally, if arboreality confers an advantage for acoustic signal transmission, then SVs produced and received at heights within trees should experience less attenuation than vocalizations produced and received at ground level. We broadcast and recorded synthesized vocalizations in pinyon mouse habitat at diferent heights and distances to test these predictions and contextualized our fndings by incorporating receiver hearing sensitivities at ecologically relevant distances.

# **Materials and methods**

# **Animals**

Twenty pinyon mice (9 females (F), 11 males (M); average mass:  $F = 30.90 \pm 3.92$  g,  $M = 32.85 \pm 3.11$  g) were captured near Deadman Flat, 28 km north of Flagstaf, AZ, using Sherman live traps baited with peanut butter and oats. Mice were transferred to standard mouse cages and maintained in animal facilities at Northern Arizona University, Flagstaf, AZ, USA. Mice were housed individually in the vivarium, maintained on a 14:10 dark:light cycle  $(21 \pm 2 \degree C)$ , and provided rodent chow and water ad libitum. Mice were used for both acoustic recording and hearing experiments (below).

#### **Acoustic recording**

Individually-housed mice were placed in a semi-anechoic sound cubicle lined with acoustic foam and recorded over 3–7 days within their home cage. Calibrated microphones (1/4′′ Type 40BE, G.R.A.S.) connected to preamplifers (Type 26CB, G.R.A.S.) placed 33.3 cm above the center of the cage were used to record mice. Microphone response was flat within  $\pm$  1.5 dB from 10 Hz—50 kHz, and pre-amplifier response was flat within $\pm 0.2$  dB from 2 Hz-200 kHz. Microphones were connected to a National Instruments DAQ (USB 4431) sampling at 102.4 kHz to a desktop computer running MATLAB (v. 2018a).

#### **Acoustic signal generation**

To generate a signal for use in sound transmission experiments, all SV vocalizations (mean  $\pm$  SD = 193 $\pm$ 427, range: 1–1310; *n*=5 F, 3 M; see Kobrina et al. [2022a\)](#page-8-17) were analyzed for average fundamental frequency  $(F_0; kHz)$ , average duration (s), and average amplitude (dB SPL re:  $20 \mu Pa$ ). Frequency and duration measures were extracted using the automated parameter measurements function in Avisoft SASLab Pro (v. 4.2.27, Avisoft Bioacoustics, Germany; 1024-point Fast Fourier Transform, Hann window, 75% frame size, 94 Hz bandwidth, 47 Hz frequency resolution, 1.333 ms with 93.75% overlap temporal resolution). Amplitude measures from calibrated microphones were extracted using the Sound Pressure Level Calculator (Greene [2021\)](#page-7-15) in MATLAB (v. 2018a). To correct amplitude values for the distance between the microphone and calling mouse (33.3 cm), we applied a standard equation that accounts for sound energy dissipation via spherical spreading (A.2 in Brumm and Zollinger [2011\)](#page-7-16) to standardize playback SPL levels at 1 m (72.1 dB) for sound attenuation experiments (see below).

To avoid pseudo-replication (McGregor et al. [1992](#page-8-18); Kroodsma et al. [2001\)](#page-8-19), we calculated a species grand mean (F0 = 19.8  $\pm$  1.5 kHz; duration = 0.14  $\pm$  0.07 s; dB  $SPL = 81.6 \pm 4$  dB) from averaged parameters within individuals to synthesize a signal for use in sound transmission experiments using Avisoft SASLab Pro (48 kHz sampling rate, 16-bit resolution,.wav format). To improve signal to noise ratios, the fle contained 50 replicates of the synthesized signal with 0.5 s silent gaps between each replicate.

#### **Sound attenuation experiments**

We conducted sound transmission experiments at Deadman Flat between March–May 2022 around dusk (18:00–22:00 h), coincident with times that mice become active. Twenty trees  $> 4$  m (min. distance between trees 50 m) in height were selected randomly at the study site where mice were originally captured. At each site, we set a 4 m transect outward from the perimeter of the furthest live branch of the focal tree in a random direction. At 0 m, we placed a microphone (Sennheiser MKH 8060, 50 Hz – 25 kHz fat frequency response) 9 cm above the ground to mimic the height of a hypothetical mouse receiver. The microphone was connected to a feld audio recorder (Sound Devices 702 T) to record stimuli at a sampling rate of 48 kHz and 16-bit resolution. We broadcast synthesized stimuli through a speaker (Ultrasonic Dynamic Speaker, ScanSpeak, Avisoft Bioacoustics, fat frequency response within  $\pm$  1.5 dB from 10–45 kHz) connected to an Avisoft Ultrasound Gate Player 416H connected to a laptop running Avisoft RECORDER (v. 5.2.14). We calibrated the amplitude of the playback fle using a recorded 93.7 dB 1 kHz tone from a sound level calibrator (Brüel and Kjaer Type 4230) and input the resultant recording into Avisoft SASLab Pro to calibrate all subsequent recordings. We modeled the vertical position of senders (ground; 0 m, 1 m, 2 m) and receivers (ground; 0 m, 1 m, 2 m) in a full factorial design and recorded stimuli at 4 horizontal distances (1, 2, 4, and 8 m) per treatment. For example, to mimic a mouse calling from a 2 m height in a tree and a receiver at ground level, we mounted the speaker on a tripod 2 m above the ground and broadcast stimuli to a microphone placed on the ground iteratively at 1, 2, 4, and 8 m.

Prior to the analyses, all recordings were bandpass fltered between 18.73–20.73 kHz (Hamming window, 128 taps) in Avisoft SASLab Pro. The root mean square (rms) amplitude (20 ms averaging time) of each of the 50 replicates was then measured using automated detection in Avisoft SASLab Pro. We randomly selected a silent period between signal replicates to quantify the amplitude of background noise and subtracted noise values from signal amplitude values using a standard equation (equation A1 in Brumm and Zollinger [2011\)](#page-7-16). Researchers were blind to treatment when attenuation data were analyzed to minimize observer bias.

# **Hearing sensitivity**

Auditory brainstem responses (ABRs) (*n*=9 F, 8 M; a subsample of mice from the acoustic recording experiment) were conducted in a semi-anechoic chamber (ETS Lindgren SD-1; internal dimensions 91.4 cm $\times$ 91.4 cm $\times$ 91.4 cm) lined with acoustic foam. We administered ketamine/dexmedetomidine (75/0.5 mg/ kg) intraperitoneally to anesthetize animals. Occasionally, we injected an additional dose of ketamine (<0.01 mL) 15 min after the initial dose to maintain an anesthetic plane. Anesthetized animals were transferred to a gel heating pad  $(32 \pm 5 \degree C)$  to maintain a stable body temperature for the duration of the experiment. Monaural (randomly assigned ear; right *n*=10, left *n*=7) ABR measurements were obtained by placing three subdermal needle electrodes (27 gauge, 12 mm; Rochester Electro-Medical Inc., Lutz, FL, USA) on the mastoid of ear receiving the stimulus (reference), the vertex of the skull (active channel), and in the dorsum close to the base of the tail (ground). Electrodes were connected to a head stage (RA4LI, Tucker Davis Technologies (TDT), Alachua, FL, USA) and a preamplifer (RA4RA, TDT) attached to a processor (RZ6, TDT) via a fber optic cable.

We generated and presented test stimuli, and collected responses using SigGenRZ and BioSigRZ (v. 5.7.0, TDT), a TDT Multi I/O processor RZ6, and a PC. ABR experiments were controlled by a PC Windows computer running an Optibit interface on a TDT driver using BioSigRZ. The stimuli were generated by the RZ6 and played through a speaker (MF1, TDT) located 10 cm away from the pinna of the pinyon mouse at 0-degree azimuth. Digitized data were recorded to the RZ6 processor through the RA4LI preamplifer. Stimuli consisted of clicks (0.1 ms square wave pulse of alternating polarity, obtained from the TDT root click fle) and 5-ms single-channel cosinesquared gated tone bursts at frequencies of 4, 8, 16, 20, 24, 32, and 42 kHz (Cos2 (10%—90%) gating type, obtained from the TDT root tone fle) presented 21 times per second for a total set of 512 repetitions. Test frequencies were selected to examine hearing abilities of both low and high-frequency communication signals previously recorded in *P. truei* (Kobrina et al. [2021\)](#page-8-20) and typical of rodent audiograms (Dent et al. [2018](#page-7-17)). However, in this study, we only use sensitivities derived from the 20 kHz stimulus that was most similar to the F0 of SVs (19.8 kHz; above). The click stimulus has more spectral energy below 10 kHz and usually elicits a more robust ABR response than pure tones. Each stimulus was presented at descending levels starting at 90 dB until a threshold was reached. Step sizes were 10–15 dB at suprathreshold levels, and then decreased to 5 dB bracketing the threshold. The system was calibrated prior to each experiment using a microphone (Brüel and Kjær, Type 2670, Nærum, Denmark) connected to a microphone supply (Brüel and Kjær, Type 5935 L, Nærum, Denmark).

#### **Attenuation statistical analyses**

To assess how SVs were afected by distance and height, we used a generalized linear mixed model (GLMM) with the lme4 package in R studio, v. 3.3.3 (Bates et al. [2015](#page-7-18); R Core Team [2017](#page-8-21)). The GLMM included the log-transformed synthesized stimuli SPL as the response variable and microphone distance  $(1 m, 2 m, 4 m, 8 m)$ , speaker height  $(0 m, 1 m, 2 m)$ , and microphone height (0 m, 1 m, 2 m) as main efects, and all 2- and 3-way interactions. ANOVA and posthoc Tukey models revealed background noise was variable among sites  $(F<sub>1,19</sub>=3.29)$ , *p*<0.0001). Therefore, we included site as a random effect in the GLMM to control for background noise variation. We verifed assumptions of normality and homoscedasticity by visual inspection of q-q and residual plots. Finally, we computed  $\eta^2$  and confdence intervals (CI) as efect sizes for signifcant results in the *efectsize* package for R (Ben-Shachar et al. [2020](#page-7-19)).

#### **ABR statistical analyses**

We used the visual detection method (Green et al. [2019](#page-7-20); Kobrina et al. [2021](#page-8-20), [2022b\)](#page-8-22) to determine the lowest stimulus level (dB) per stimulus that evoked an ABR response.

Thresholds were operationally defned as the dB level halfway (2.5 dB) between the last detectable ABR response and next lowest stimulus level (see Kobrina et al. [2022b](#page-8-22) for details). A two-way repeated-measures mixed ANOVA was used to determine whether hearing sensitivity varied across stimuli and between ears (*lsr* package in R; Navarro [2013](#page-8-23)). Paired Tukey's t-test post hoc analyses were conducted to assess significance. We computed  $\eta^2$  and confidence intervals (CI) as measures of efect sizes for signifcant results (*efectsize* package in R; Ben-Shachar et al. [2020\)](#page-7-19).

## **Audible distance estimation**

Finally, we assessed the audible distance of pinyon mouse SVs produced from the ground (height=0 m) and from a tree  $(height=1 m)$ . To do so, we first calculated sound attenuation via spherical spreading of SV amplitude at 0 m and 1 m heights, using the *attenuation* function in the Seewave package (Sueur et al. [2008\)](#page-9-5). All factors held equal, sound is expected to attenuate 6 dB for every doubling of distance (Brumm and Zollinger [2011\)](#page-7-16). We then modelled excess attenuation for our observed amplitude values at each height by ftting a logarithmic decay model in R at 1 m, 2 m, 4 m, and 8 m distances, and extrapolated amplitude values beyond 8 m using the line of best ft equation. Finally, to estimate the audible distance of pinyon mice SVs, we used the line of best ft equation to calculate the distance at which sound attenuation intersected receiver hearing sensitivity (41.7 dB at 20 kHz; see Results) measured from ABRs.

# **Results**

## **Sound attenuation**

Attenuation of synthesized stimuli were influenced by sender (speaker) height (β=-19.859, df=1, 663, *p*<0.0001,  $\eta^2$  = 0.36; CI [0.31, 0.41]), receiver (microphone) height (β=-24.483, df=1, 663, *p*<0.0001, *η<sup>2</sup>*=0.43; CI [0.38, 0.48]), receiver (microphone) distance ( $\beta$  = -4.230, df = 1, 663, *p*<0.0001; *η<sup>2</sup>*=0.22; CI [0.17, 0.27]), and all 2- and 3-way interactions (Table [1\)](#page-4-0). SVs produced and/or received from the ground were more attenuated than above ground. At all distances beyond 1 m, attenuation was greater when both the sender and receiver were on the ground than at any elevation (Fig. [1\)](#page-4-1). On average, trials where both the sender and receiver were elevated 1 m experienced the least attenuation (Fig. [1\)](#page-4-1). When the sender was on the ground, the 1 m receiver height trials experienced the least attenuation at longer distances (4 m and 8 m; Fig. [1](#page-4-1)). The interaction between speaker and microphone height was the strongest effect in our model ( $β = 20.97$ , df = 1.363, *p* < 0.0001;  $\eta^2$  = 0.50; CI [0.45, 0.54]), indicating amplitude was loudest when the sender and receiver height was matched (Fig. S1).

<span id="page-4-0"></span>**Table 1** The efects of speaker height, microphone height, and microphone distance on attenuation of pinyon mouse (*P. truei*) sustained vocalizations based on full factorial playback experiments. Results are from a generalized linear mixed model

Effect	Estimate	<b>SD</b>		df	P	n2	CI lower	CI upper
Speaker Height (m)	$-19.86$	1.20	$-18.1$	1.363	< 0.0001	0.36	0.31	0.41
Microphone Height (m)	$-24.48$	1.20	$-22.4$	1,363	< 0.0001	0.43	0.38	0.48
Microphone Distance (m)	$-4.23$	0.31	$-13.8$	1,363	< 0.0001	0.22	0.17	0.27
Speaker Height * Microphone Height	20.97	0.85	24.7	1,363	< 0.0001	0.50	0.45	0.54
Speaker Height * Microphone Distance	3.19	0.24	13.4	1,363	< 0.0001	0.23	0.17	0.28
Microphone Height * Microphone Distance	3.62	0.24	15.2	1,363	< 0.0001	0.25	0.20	0.30
Speaker Height * Microphone Height * Micro- phone Distance	$-3.00$	0.18	$-16.3$	1,363	< 0.0001	0.29	0.24	0.35

<span id="page-4-1"></span>

When heights were mismatched, amplitude was optimized if the receiver height was within 1 m of the sender height (Fig. S1). Site, as a random efect, accounted for a relatively large amount of variation ( $\sigma^2$  = 2.83, SD = 1.68).

## **Hearing sensitivity**

In general, pinyon mice were able to detect all stimuli. ANOVA revealed a signifcant main efect of stimulus  $(F_{7.96} = 69.76, p < 0.001, \eta^2 = 0.82$ ; CI [0.75, 0.86]) and a non-significant main effect of ear  $(F_{1,96}=0.07, p=0.79)$ . Paired Tukey's post-hoc analyses indicated that pinyon mice had lower thresholds (i.e. were more sensitive) to click stimuli than to 4, 8, 20, 24, 32, and 42 kHz tones  $(p < 0.004)$ . Pinyon mice were most sensitive to 16 kHz tones than to all other frequencies  $(p < 0.02)$  with the exception of 8 kHz  $(p > 0.05)$ . Mice were least sensitive to 32 and 42 kHz tones than to all other frequencies  $(p < 0.001)$ . Hearing sensitivity for 20 kHz tones were  $41.67 \pm 6.69$  dB and were not significantly different from 4, 8, or 24 kHz tones (*p*>0.05; Fig. [2\)](#page-4-2).

#### **Audible distance estimation**

We assessed the audible distance of SVs on the ground and in a tree by integrating our ABR results with sound attenuation models based on spherical spreading and excess



<span id="page-4-2"></span>**Fig. 2** Audiogram of pinyon mice  $(n=17)$ . Error bars represent  $\pm$  SE

<span id="page-5-0"></span>**Fig. 3** Attenuation curves based on spherical spreading (open circles) and excess attenuation (black circles) of synthesized pinyon mouse stimuli emitted and received on the ground (left panel) vs. emitted and received at 1 m height (right panel). Horizontal dashed lines represent the hearing sensitivity (41.67 dB) of pinyon mice at 20 kHz. Vertical dashed lines and the corresponding value represent the maximum audible distance (m) of vocalizations



attenuation. Attenuation curves indicated stimuli produced from the ground experienced greater excess attenuation relative to spherical spreading (Fig. [3\)](#page-5-0). From this model, the audible distance, or active space, of SVs was 4.35 m (Fig. [3](#page-5-0)); beyond this distance, amplitude fell below the estimated 41.67 dB hearing threshold of pinyon mice. In contrast, stimuli produced from trees at 1 m height was predominantly afected by spherical spreading and had little excess attenuation. Under this scenario, the audible distance of SVs was 3 times farther than the ground (12.4 m; Fig. [3](#page-5-0)).

# **Discussion**

Our fndings indicate both senders and receivers of acoustic signals beneft from positioning themselves at elevated heights. Vocalizations emitted and received at 1 m off the ground had an active space that was approximately 3 times larger compared to on the ground, in part due to ground and vegetation effects that increase acoustic attenuation, especially at high frequencies (Brenowitz [1986;](#page-7-21) Marten and Marler [1977\)](#page-8-1). We discuss our fndings in relation to the ecology and evolution of acoustic communication in animals in general and rodents in particular.

Our results concur with studies in other taxa that exploit elevated heights to minimize attenuation and thereby extend the communication range of acoustic signals (Arak and Eiriksson [1992](#page-7-6); Mathevon et al [2005](#page-8-7); Schwartz et al. [2015](#page-8-5); Riondato et al. [2021\)](#page-8-6). Benefts to elevated heights include increasing the probability of detection, reducing search costs associated with localizing mates and/or competitors, and/ or increasing the number of receivers (Richards and Wiley [1980;](#page-8-8) Ryan and Kime [2003\)](#page-8-0). In some birds, both senders and receivers exploit elevated perches to optimize production and reception (Dabelsteen et al. [1993\)](#page-7-7). For example, in Rufous-and-white wrens (*Thryophilus rufalbus*) and

Blackcaps (*Sylvia atricapilla*), certain perch locations extend communication range, especially if senders and receivers are at the same height (Holland et al. [1998](#page-7-8); Mathevon et al [2005](#page-8-7)). Similarly, our results indicate that both senders and receivers beneft at elevated heights, particularly when each party is on a similar plane. However, benefts wane at higher heights (2 m), likely because most attenuation is caused by ground efects.

Although a growing body of literature exists on acoustic communication in mice (Kalcounis-Rueppell et al. [2018c](#page-8-10); Rieger and Marler [2018;](#page-8-24) Fernández-Vargas et al. [2022](#page-7-9); Kobrina et al. [2022a](#page-8-17), [b](#page-8-22)), few studies are contextualized ecologically. In closely related California mice (*P. californicus*), SVs appear to facilitate spatial contact and territorial defense between monogamous pairs (Briggs and Kalcounis-Rueppell [2011\)](#page-7-14). Our estimate of audible distance of SVs at ground level (4.35 m) concur with values reported in *P. californicus* (3.12 m*;* Timonin et al. [2018\)](#page-9-6) that occupies denser vegetation, adding support to their utility over long distances. In polygynous to promiscuous brush mice (*P. boylii*; Ribble and Stanley [1998;](#page-8-25) Kalcounis-Rueppell and Spoon [2009](#page-8-26)), SVs are implicated in territorial advertisement, especially among females (Petric and Kalcounis-Rueppell [2013](#page-8-16)). Similar to brush mice, pinyon mice are promiscuous (Ribble and Stanley [1998](#page-8-25)) and SVs are produced primarily by females (Kobrina et al. [2022a\)](#page-8-17). Given their large home range sizes (0.4—1.6 ha; Ribble and Stanley [1998](#page-8-25)), female SVs likely facilitate advertisement of their position to neighbors throughout the year and roaming males during the mating season (Emlen and Oring [1977](#page-7-22)). Our fndings suggest that elevated advertisement and reception increases the probability of signal efficacy to mediate intra- and inter-sexual interactions. Notably, varying levels of arboreality are reported in diferent species of *Peromyscus* (e.g. *P. boylii, P. californicus, P. leucopus, P. maniculatus;* Meserve [1977;](#page-8-27) Harney and Dueser [1987;](#page-7-23) Kalcounis-Rüppell and Millar [2002](#page-8-28)) that use

SVs in similar long-distance contexts (Kalcounis-Rueppell et al. [2006](#page-8-29); Petric and Kalcounis-Rueppell [2013](#page-8-16); Rieger and Marler [2018](#page-8-24)). While the origins of arboreal behavior may be due to food availability (Holbrook [1978](#page-7-24)) and/or interspecifc competition (Stah [1980\)](#page-9-7), the communicative benefts realized from the reduction of acoustic attenuation at ground level may help maintain arboreality. Comparative studies that assess the geometry of vocal behavior across this diverse genus (Kalcounis- Rueppell et al. 2018a, c) would provide important insight and resolution.

The tendency for higher frequencies ( $\geq 20$  kHz) that characterize SVs to be highly directional and attenuate rapidly in the environment requires novel adaptations in both senders and receivers to increase signal efficacy (Richards and Wiley [1980](#page-8-8)). In bats, active control (Yovel et al. [2011](#page-9-8)) of high-amplitude echolocation calls (Jakobsen et al. [2013](#page-7-25)) produced by unique laryngeal mechanisms (Metzner and Schuller [2010\)](#page-8-30) are accompanied by morphologically diverse (Leiser-Miller and Santana [2020](#page-8-31)) and positionally dynamic (Müller et al. [2017](#page-8-32)) pinnae to facilitate reception of high frequencies. Peromyscus similarly have unique laryngeal anatomy to produce SVs (Fernández-Vargas et al. [2022](#page-7-9)), but mechanisms of reception are less studied (Dice and Barto [1952;](#page-7-26) Ralls [1967](#page-8-33); Capshaw et al. [2022](#page-7-27)). In pinyon mice, large, conspicuous pinnae are hypothesized to facilitate detection of low-frequency predator cues (Hoffmeister [1951,](#page-7-28) [1981](#page-7-29)), not higher frequency conspecific vocalizations. Indeed, broader and larger pinnae are associated with lower frequency echolocation calls in bats (Obrist et al. [1993;](#page-8-3) Huihua et al. [2003\)](#page-7-30). Furthermore, we did not find secondary peaks in pinyon mouse audiograms as is expected when pinnae morphology is involved in high frequency hearing (Heffner et al. [2001](#page-7-31)). Although more studies are needed on rodent hearing, such patterns further implicate the importance of studying relative positions of senders and receivers.

Our inferences come with minor caveats. Playback of synthesized signals highlight theoretical benefits gained by senders and receivers if they attended to spatial locations when producing or evaluating signals. Although pinyon mice are agile climbers and commonly observed in trees (Bailey [1931;](#page-7-32) Horner [1954](#page-7-11)), assessment of realized benefits will require simultaneous tracking and vocal recording of animals in three-dimensional space. In addition, we focused our study on quantifying signal attenuation of a single note of SVs, which represents only one aspect of signal degradation. Pinyon mouse SVs are typically composed of multiple rapidly repeated notes (Kobrina et al. [2021](#page-8-20)) that likely serve to facilitate detection and recognition (Bradbury and Vehrencamp [2011](#page-7-33)). Studies that integrate other measures of degradation (e.g. distortion of temporal and frequency patterns; Mathevon et al. [2005\)](#page-8-7) can help clarify the robustness of our findings. Finally, our estimates of signal active space are conservative because electrophysiological methods of hearing sensitivity (e.g. ABRs) are typically less sensitive (10–30 dB) than behavioral paradigms (Heffner and Heffner [2003](#page-7-34); Dent et al. [2018](#page-7-17)). Thus, the ecological validity of the spatial extent of acoustic interactions may be greater than reported herein.

From a basic perspective, our study extends the taxonomic scope of positional advertisement and contributes to our understanding of behavioral mechanisms used to facilitate animal communication. From an applied perspective, consideration of the environment where signaling behaviors occur is critical. For example, pinyon mice rely on pinyonjuniper woodlands (Hall and Morrison [1997\)](#page-7-10) that are receding in their southern ranges due to drought (Cliford et al. [2011;](#page-7-35) Redmond et al. [2012;](#page-8-34) Whipple et al. [2019](#page-9-9)). Given the importance of elevated heights for sound propagation, reductions in tree densities may not only negatively impact food (Hall and Morrison [1997](#page-7-10)) and nest sites but indirectly infuence demographic rates that rely on behaviors reliant on acoustic communication. Integrating signaling and sensory ecology will be key to understanding species-specifc resilience (Srivathsa et al. [2019\)](#page-9-10) in response to environmental change.

**Supplementary information** The online version contains supplementary material available at<https://doi.org/10.1007/s00265-023-03352-4>.

**Acknowledgements** We thank E. Roden and D. Hendershott for assistance with trapping and recording mice. We thank two anonymous reviewers for their insightful comments that greatly improved the manuscript.

**Authors' contributions** RB and BP conceived the study, obtained the funding, and wrote the initial draft of the manuscript. RB, BP, and AK conducted the feldwork. AK conducted the hearing experiments. SMM and all authors analyzed the data, and revised, edited, and approved the fnal version of the manuscript.

**Funding** The study was funded by the American Society of Mammalogists Grants-in-Aid (RB), a Jean Schuler Research Mini-Grant from Northern Arizona University (RB), and the National Science Foundation IOS # 1755429 (BP).

**Data availability** All datasets generated and analyzed during this study are included in the supplementary information fles of this published article.

#### **Declarations**

**Ethics approval** All procedures were approved by Northern Arizona University's Institutional Animal Care and Use Committee (19–006). Animals were captured with permits from the Arizona Game and Fish Department (SP403209). All research followed the guidelines for ethical treatment of animals established by the American Society of Mammalogists (Sikes [2016\)](#page-8-35).

**Competing interests** The authors declare no competing interests.

# **References**

- <span id="page-7-6"></span>Arak A, Eiriksson T (1992) Choice of singing sites by male bushcrickets (*Tettigonia viridissima*) in relation to signal propagation. Behav Ecol Sociobiol 30:365–372. [https://doi.org/10.](https://doi.org/10.1007/BF00176170) [1007/BF00176170](https://doi.org/10.1007/BF00176170)
- <span id="page-7-32"></span>Bailey V (1931) Mammals of New Mexico. United States Department of Agriculture, Bureau of Biological Survey, Washington
- <span id="page-7-2"></span>Barker NKS, Mennill DJ (2009) Song perch height in Rufous-and-White Wrens: Does behaviour enhance effective communication in a tropical forest? Ethology 115:897–904. [https://doi.](https://doi.org/10.1111/j.1439-0310.2009.01674.x) [org/10.1111/j.1439-0310.2009.01674.x](https://doi.org/10.1111/j.1439-0310.2009.01674.x)
- <span id="page-7-18"></span>Bates D, Mächler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using lme4. J Stat Softw 67:1–48. <https://doi.org/10.18637/jss.v067.i01>
- <span id="page-7-19"></span>Ben-Shachar MS, Lüdecke D, Makowski D (2020) effectsize: Estimation of effect size indices and standardized parameters. J Open Source Softw 5:2815
- <span id="page-7-5"></span>Blumenrath S, Dabelsteen T (2004) Degradation of Great Tit (*Parus Major*) song before and after foliation: Implications for vocal communication in a deciduous forest. Behaviour 141:935–958.<https://doi.org/10.1163/1568539042360152>
- <span id="page-7-0"></span>Boncoraglio G, Saino N (2007) Habitat structure and the evolution of bird song: A meta-analysis of the evidence for the acoustic adaptation hypothesis. Funct Ecol 21:134–142. [https://doi.org/](https://doi.org/10.1111/j.1365-2435.2006.01207.x) [10.1111/j.1365-2435.2006.01207.x](https://doi.org/10.1111/j.1365-2435.2006.01207.x)
- <span id="page-7-33"></span>Bradbury JW, Vehrencamp SL (2011) Principles of animal communication, 2nd edn. Sinauer Associates, Sunderland, Massachusetts
- <span id="page-7-21"></span>Brenowitz EA (1986) Environmental influences on acoustic and electric animal communication. Brain Behav and Evol 28:32– 42. <https://doi.org/10.1159/000118690>
- <span id="page-7-14"></span>Briggs JR, Kalcounis-Rueppell MC (2011) Similar acoustic structure and behavioural context of vocalizations produced by male and female California mice in the wild. Anim Behav 82:1263–1273.<https://doi.org/10.1016/j.anbehav.2011.09.003>
- <span id="page-7-16"></span>Brumm H, Zollinger SA (2011) The evolution of the Lombard effect: 100 years of psychoacoustic research. Behaviour 148:1173–1198.<https://doi.org/10.1163/000579511X605759>
- <span id="page-7-13"></span>Camargo NF, Machado LF, Mendonça AF, Vieira EM (2019) Cranial shape predicts arboreal activity of Sigmodontinae rodents. J Zool 308:128–138.<https://doi.org/10.1111/jzo.12659>
- <span id="page-7-27"></span>Capshaw G, Vicencio-Jimenez S, Screven LA, Burke K, Weinberg MM, Lauer AM (2022) Physiological evidence for delayed age-related hearing loss in two long-lived rodent species (*Peromyscus leucopus* and *P. californicus*). J Assoc Res Otolaryngol 23:617–631. <https://doi.org/10.1007/s10162-022-00860-4>
- <span id="page-7-35"></span>Clifford MJ, Cobb NS, Buenemann M (2011) Long-Term Tree Cover Dynamics in a Pinyon-Juniper Woodland: Climate-Change-Type Drought Resets Successional Clock. Ecosyst 14:949–962. <https://doi.org/10.1007/s10021-011-9458-2>
- <span id="page-7-7"></span>Dabelsteen T, Pedersen SB, Larsen ON (1993) Habitat-induced degradation of sound signals: quantifying the effects of communication sounds and bird location on blur ratio, excess attenuation and signal-to- noise ratio. J Acoust Soc Am 93:2206–2220. <https://doi.org/10.1121/1.406682>
- <span id="page-7-17"></span>Dent ML, Screven LA, Kobrina A (2018) Hearing in rodents. In: Dent M, Fay R, Popper A (eds) Rodent Bioacoustics. Springer, Cham, pp 71–105. [https://doi.org/10.1007/](https://doi.org/10.1007/978-3-319-92495-3_4) [978-3-319-92495-3\\_4](https://doi.org/10.1007/978-3-319-92495-3_4)
- <span id="page-7-26"></span>Dice LR, Barto E (1952) Ability of mice of the genus *Peromyscus* to hear ultrasonic sounds. Science 116:110–111. [https://doi.](https://doi.org/10.1126/science.116.3005.110) [org/10.1126/science.116.3005.110](https://doi.org/10.1126/science.116.3005.110)
- <span id="page-7-22"></span>Emlen T, Oring LW (1977) Ecology, sexual selection, and the evolution of mating systems. Science 197:215–223. [https://doi.](https://doi.org/10.1126/science.327542) [org/10.1126/science.327542](https://doi.org/10.1126/science.327542)
- <span id="page-7-1"></span>Ey E, Fischer J (2009) The "acoustic adaptation hypothesis"—a review of the evidence from birds, anurans and mammals. Bioacoustics 19:21–48. [https://doi.org/10.1080/09524622.](https://doi.org/10.1080/09524622.2009.9753613) [2009.9753613](https://doi.org/10.1080/09524622.2009.9753613)
- <span id="page-7-9"></span>Fernández-Vargas M, Riede T, Pasch B (2022) Mechanisms and constraints underlying acoustic variation in rodents. Anim Behav 184:135–147. [https://doi.org/10.1016/j.anbehav.2021.](https://doi.org/10.1016/j.anbehav.2021.07.011) [07.011](https://doi.org/10.1016/j.anbehav.2021.07.011)
- <span id="page-7-3"></span>Goutte S, Dubois A, Howard SD, Márquez R, Rowley JJL, Dehling JM, Grandcolas P, Xiong RC, Legendre F (2018) How the environment shapes animal signals: a test of the acoustic adaptation hypothesis in frogs. J Evol Biol 31:148–158. [https://doi.](https://doi.org/10.1111/jeb.13210) [org/10.1111/jeb.13210](https://doi.org/10.1111/jeb.13210)
- <span id="page-7-20"></span>Green DM, Scolman T, Pasch B (2019) A broad filter between call frequency and peripheral auditory sensitivity in northern grasshopper mice (*Onychomys leucogaster*). J Comp Physiol A 205:481–489. <https://doi.org/10.1007/s00359-019-01338-0>
- <span id="page-7-15"></span>Greene C (2021) MATLAB Central File Exchange: Sound Pressure Level Calculator. MathWorks. [https://www.mathworks.com/](https://www.mathworks.com/matlabcentral/fileexchange/35876-sound-pressure-level-calculator) [matlabcentral/fileexchange/35876-sound-pressure-level-calcu](https://www.mathworks.com/matlabcentral/fileexchange/35876-sound-pressure-level-calculator) [lator](https://www.mathworks.com/matlabcentral/fileexchange/35876-sound-pressure-level-calculator)
- <span id="page-7-12"></span>Hager ER, Hoekstra HE (2021) Tail length evolution in deer mice: Linking morphology, behavior, and function. Integr Comp Biol 61:385–397. <https://doi.org/10.1093/icb/icab030>
- <span id="page-7-10"></span>Hall LS, Morrison ML (1997) Den and relocation site characteristics and home ranges of *Peromyscus truei* in the White Mountains of California. Great Basin Nat 57:124–130
- <span id="page-7-23"></span>Harney BA, Dueser RD (1987) Vertical stratification of activity of two *Peromyscus* species: an experimental analysis. Ecology 68:1084–1091. <https://doi.org/10.2307/1938380>
- <span id="page-7-4"></span>Hayes AR, Huntly NJ (2005) Effects of wind on the behavior and call transmission of pikas (*Ochotona princeps*). J Mammal 86:974–981. [https://doi.org/10.1644/1545-1542\(2005\)86\[974:](https://doi.org/10.1644/1545-1542(2005)86[974:EOWOTB]2.0.CO;2) [EOWOTB\]2.0.CO;2](https://doi.org/10.1644/1545-1542(2005)86[974:EOWOTB]2.0.CO;2)
- <span id="page-7-34"></span>Heffner HE, Heffner RS (2003) Audition. In: Davis SF (ed) Handbook of research methods in experimental psychology. Blackwell, Boston, MA, pp 413–440. [https://doi.org/10.1002/97804](https://doi.org/10.1002/9780470756973.ch19) [70756973.ch19](https://doi.org/10.1002/9780470756973.ch19)
- <span id="page-7-31"></span>Heffner RS, Koay G, Heffner HE (2001) Audiograms of five species of rodents: implications for the evolution of hearing and the perception of pitch. Hear Res 157:138–152. [https://doi.](https://doi.org/10.1016/S0378-5955(01)00298-2) [org/10.1016/S0378-5955\(01\)00298-2](https://doi.org/10.1016/S0378-5955(01)00298-2)
- <span id="page-7-28"></span>Hoffmeister DF (1951) A taxonomic and evolutionary study of the piñon mouse, Peromyscus truei. University of Illinois Press, Urbana.<https://doi.org/10.5962/bhl.title.50289>
- <span id="page-7-29"></span>Hoffmeister DF (1981) *Peromyscus truei*. Mamm Species 161:1–5
- <span id="page-7-24"></span>Holbrook S (1978) Habitat relationships and coexistence of four sympatric species of *Peromyscus* in northwestern New Mexico. J Mammal 59:18–26
- <span id="page-7-8"></span>Holland J, Dabelsteen T, Pedersen SB, Larsen ON (1998) Degradation of song in the wren *Troglodytes troglodytes*: Implications for information transfer and ranging. J Acoust Soc Am 103:2154–2166.<https://doi.org/10.1121/1.421361>
- <span id="page-7-11"></span>Horner E (1954) Arboreal adaptations of *Peromyscus*, with special reference to use of the tail. University of Michigan, Ann Arbor, MI
- <span id="page-7-30"></span>Huihua Z, Shuyi Z, Mingxue Z, Jiang Z (2003) Correlations between call frequency and ear length in bats belonging to the families Rhinolophidae and Hipposideridae. J Zool 259:189– 195.<https://doi.org/10.1017/S0952836902003199>
- <span id="page-7-25"></span>Jakobsen L, Brinkløv S, Surlykke A (2013) Intensity and directionality of bat echolocation signals. Front Physiol 4:89. [https://](https://doi.org/10.3389/fphys.2013.00089) [doi.org/10.3389/fphys.2013.00089](https://doi.org/10.3389/fphys.2013.00089)

<span id="page-8-26"></span>Kalcounis-Rueppell MC, Spoon TR (2009) *Peromyscus boylii*. Mamm Species 838:1–14

- <span id="page-8-29"></span>Kalcounis-Rueppell MC, Metheny JD, Vonhof MJ (2006) Production of ultrasonic vocalizations by *Peromyscus* mice in the wild. Front Zool 3:3.<https://doi.org/10.1186/1742-9994-3-3>
- <span id="page-8-9"></span>Kalcounis-Rueppell MC, Petric R, Marler CA (2018) The bold, silent type: predictors of ultrasonic vocalizations in the genus *Peromyscus*. Front Ecol Evol 6:198. [https://doi.org/10.3389/](https://doi.org/10.3389/fevo.2018.00198) [fevo.2018.00198](https://doi.org/10.3389/fevo.2018.00198)
- <span id="page-8-15"></span>Kalcounis-Rueppell MC, Pultorak JD, Blake BH, Marler CA (2018) Ultrasonic vocalizations of young mice in the genus *Peromyscus*. Handb Behav Neurosci 25:149–156. [https://doi.org/10.](https://doi.org/10.1016/B978-0-12-809600-0.00014-7) [1016/B978-0-12-809600-0.00014-7](https://doi.org/10.1016/B978-0-12-809600-0.00014-7)
- <span id="page-8-10"></span>Kalcounis-Rueppell MC, Pultorak JD, Marler CA (2018) Ultrasonic vocalizations of mice in the genus *Peromyscus*. Handb Behav Neurosci 25:227–235. [https://doi.org/10.1016/B978-0-](https://doi.org/10.1016/B978-0-12-809600-0.00014-7) [12-809600-0.00014-7](https://doi.org/10.1016/B978-0-12-809600-0.00014-7)
- <span id="page-8-28"></span>Kalcounis-Rüppell MC, Millar JS (2002) Partitioning of space, food, and time by syntopic *Peromyscus boylii* and *P. californicus*. J Mammal 83:614–625. [https://doi.org/10.1644/15451](https://doi.org/10.1644/15451542(2002)083%3c0614:POSFAT%3e2.0.CO;2) [542\(2002\)083%3c0614:POSFAT%3e2.0.CO;2](https://doi.org/10.1644/15451542(2002)083%3c0614:POSFAT%3e2.0.CO;2)
- <span id="page-8-12"></span>Kingsley EP, Kozak KM, Pfeifer SP, Yang DS, Hoekstra HE (2017) The ultimate and proximate mechanisms driving the evolution of long tails in forest deer mice. Evol 71:261–273. [https://doi.](https://doi.org/10.1111/evo.13150) [org/10.1111/evo.13150](https://doi.org/10.1111/evo.13150)
- <span id="page-8-20"></span>Kobrina A, Hidau MK, Riede T, Guthrie O, Pasch B (2021) Agerelated and noise-induced hearing loss alters grasshopper mouse (Onychomys) vocalizations. Hear Res 404:108210. <https://doi.org/10.1016/j.heares.2021.108210>
- <span id="page-8-17"></span>Kobrina A, Letowt ME, Pasch B (2022) The influence of social context on pinyon mouse (*Peromyscus truei*) vocalizations. J Mammal 103:275–286. [https://doi.org/10.1093/jmammal/](https://doi.org/10.1093/jmammal/gyab127) [gyab127](https://doi.org/10.1093/jmammal/gyab127)
- <span id="page-8-22"></span>Kobrina A, Letowt ME, Pasch B (2022) Vocal repertoire and auditory sensitivity of white-throated woodrats (*Neotoma albigula*). J Comp Psychol 137:116–128. [https://doi.org/10.1037/](https://doi.org/10.1037/com0000330) [com0000330](https://doi.org/10.1037/com0000330)
- <span id="page-8-19"></span>Kroodsma DE, Byers BE, Goodale E, Johnson S, Liu WC (2001) Pseudoreplication in playback experiments, revisited a decade later. Anim Behav 61:1029–1033. [https://doi.org/10.1006/](https://doi.org/10.1006/anbe.2000.1676) [anbe.2000.1676](https://doi.org/10.1006/anbe.2000.1676)
- <span id="page-8-31"></span>Leiser-Miller LB, Santana SE (2020) Morphological diversity in the sensory system of phyllostomid bats: Implications for acoustic and dietary ecology. Funct Ecol 34:1416–1427. <https://doi.org/10.1111/1365-2435.13561>
- <span id="page-8-13"></span>Lemen C (1980) Relationship between relative brain size and climbing ability in *Peromyscus*. J Mammal 61:360–364. <https://doi.org/10.2307/1380068>
- <span id="page-8-14"></span>Mace GM, Eisenberg JF (1982) Competition, niche specialization and the evolution of brain size in the genus *Peromyscus*. Biol J Linn Soc 17:243–257. [https://doi.org/10.1111/j.1095-8312.1982.](https://doi.org/10.1111/j.1095-8312.1982.tb02019.x) [tb02019.x](https://doi.org/10.1111/j.1095-8312.1982.tb02019.x)
- <span id="page-8-1"></span>Marten K, Marler P (1977) Sound transmission and its significance for animal vocalization. Behav Ecol Sociobiol 2:271–290. <https://doi.org/10.1007/BF00299740>
- <span id="page-8-7"></span>Mathevon N, Dabelsteen T, Blumenrath SH (2005) Are high perches in the blackcap *Sylvia atricapilla* song or listening posts? A sound transmission study. J Acoust Soc Am 117:442– 449.<https://doi.org/10.1121/1.1828805>
- <span id="page-8-18"></span>McGregor PK, Catchpole CK, Dabelsteen T et al (1992) Playback and studies of animal communication. Springer, Boston. [https://doi.org/10.1007/978-1-4757-6203-7\\_1](https://doi.org/10.1007/978-1-4757-6203-7_1)
- <span id="page-8-27"></span>Meserve PL (1977) Three-dimensional home ranges of cricetid rodents. J Mammal 58:549–558. [https://doi.org/10.2307/13800](https://doi.org/10.2307/1380003) [03](https://doi.org/10.2307/1380003)
- <span id="page-8-30"></span>Metzner W, Schuller G (2010) Vocal control in echolocating bats. Handb Behav Neurosci 19:403–415. [https://doi.org/10.1016/](https://doi.org/10.1016/B978-0-12-374593-4.00037-1) [B978-0-12-374593-4.00037-1](https://doi.org/10.1016/B978-0-12-374593-4.00037-1)
- <span id="page-8-32"></span>Müller R, Gupta AK, Zhu H, Pannala M, Gillani US, Fu Y, Caspers P, Buck JR (2017) Dynamic substrate for the encoding sensory information in bat biosonar. Phys Rev Lett 118:158102. <https://doi.org/10.1103/PhysRevLett.118.158102>
- <span id="page-8-23"></span>Navarro D (2013) Learning statistics with R. Lulu Press Inc, Morrisville, North Carolina. <https://learningstatisticswithr.com>
- <span id="page-8-3"></span>Obrist MK, Fenton MB, Eger JL, Schlegel PA (1993) What ears do for bats: a comparative study of pinna sound pressure transformation in Chiroptera. J Exp Biol 180:119–152. [https://doi.](https://doi.org/10.1242/jeb.180.1.119) [org/10.1242/jeb.180.1.119](https://doi.org/10.1242/jeb.180.1.119)
- <span id="page-8-16"></span>Petric R, Kalcounis-Rüppell MC (2013) Female and male adult brush mice (*Peromyscus boylii*) use ultrasonic vocalizations in the wild. Behaviour 150:1747–1766. [https://doi.org/10.1163/](https://doi.org/10.1163/1568539X-00003118) [1568539X-00003118](https://doi.org/10.1163/1568539X-00003118)
- <span id="page-8-21"></span>R Core Team (2017) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <http://www.R-project.org>
- <span id="page-8-33"></span>Ralls K (1967) Auditory sensitivity in mice: *Peromyscus* and *Mus musculus*. Anim Behav 15:123–128. [https://doi.org/10.1016/](https://doi.org/10.1016/S0003-3472(67)80022-8) [S0003-3472\(67\)80022-8](https://doi.org/10.1016/S0003-3472(67)80022-8)
- <span id="page-8-34"></span>Redmond M, Forcella F, Barger N (2012) Declines in pinyon pine cone production associated with regional warming. Ecosphere 3:120.<https://doi.org/10.1890/ES12-00306.1>
- <span id="page-8-11"></span>Ribble DO, Samson FB (1987) Microhabitat associations of small mammals in southeastern Colorado, with special emphasis on *Peromyscus* (Rodentia). Southwest Nat 32:291–303. [https://](https://doi.org/10.2307/3671446) [doi.org/10.2307/3671446](https://doi.org/10.2307/3671446)
- <span id="page-8-25"></span>Ribble DO, Stanley S (1998) Home ranges and social organization of syntopic *Peromyscus boylii* and *P. truei*. J Mammal 79:932–941. <https://doi.org/10.2307/1383101>
- <span id="page-8-8"></span>Richards DG, Wiley RH (1980) Reverberations and amplitude fluctuations in the propagation of sound in a forest: implications for animal communication. Am Nat 115:381–399. [https://doi.](https://doi.org/10.1086/283568) [org/10.1086/283568](https://doi.org/10.1086/283568)
- <span id="page-8-24"></span>Rieger NS, Marler CA (2018) The function of ultrasonic vocalizations during territorial defence by pair-bonded male and female California mice. Anim Behav 135:97–108. [https://doi.](https://doi.org/10.1016/j.anbehav.2017.11.008) [org/10.1016/j.anbehav.2017.11.008](https://doi.org/10.1016/j.anbehav.2017.11.008)
- <span id="page-8-6"></span>Riondato I, Gamba M, Tan CL, Niu K, Narins PM, Yang Y, Giacoma C (2021) Allometric escape and acoustic signal features facilitate high-frequency communication in an endemic Chinese primate. J Comp Physiol A 207:327–336. [https://doi.org/](https://doi.org/10.1007/s00359-021-01465-7) [10.1007/s00359-021-01465-7](https://doi.org/10.1007/s00359-021-01465-7)
- <span id="page-8-4"></span>Römer H (2020) Insect acoustic communication: The role of transmission channel and the sensory system and brain of receivers. Funct Ecol 34:310–321.<https://doi.org/10.1111/1365-2435.13321>
- <span id="page-8-2"></span>Römer H, Lewald J (1992) High-frequency sound transmission in natural habitats: implications for the evolution of insect acoustic communication. Behav Ecol Sociobiol 29:437–444. [https://](https://doi.org/10.1007/BF00170174) [doi.org/10.1007/BF00170174](https://doi.org/10.1007/BF00170174)
- <span id="page-8-0"></span>Ryan MJ, Kime NM (2003) Selection on long-distance acoustic signals. In: Simmons AM, Fay RR, Popper AN (eds) Acoustic Communication. Springer, New York, pp 225–274
- <span id="page-8-5"></span>Schwartz JJ, Hunce R, Lentine B, Powers K (2015) Calling site choice and its impact on call degradation and call attractiveness in the gray treefrog, *Hyla versicolor*. Behav Ecol Sociobiol 70:1–19. <https://doi.org/10.1007/s00265-015-2016-8>
- <span id="page-8-35"></span>Sikes RS (2016) Guidelines of the American Society of Mammalogists for the use of wild mammals in research and education. J Mammal 97:663–688. [https://doi.org/10.1093/jmamm](https://doi.org/10.1093/jmammal/gyw078) [al/gyw078](https://doi.org/10.1093/jmammal/gyw078)
- <span id="page-9-3"></span>Smartt RA (1978) A comparison of ecological and morphological overlap in a *Peromyscus* community. Ecology 59:216–220. <https://doi.org/10.2307/1936365>
- <span id="page-9-4"></span>Smartt RA, Lemen C (1980) Intrapopulational morphological variation as a predictor of feeding behavior in deermice. Am Nat 116:891–894
- <span id="page-9-10"></span>Srivathsa A, Tietje W, Rolland V, Polyakov A, Oli MK (2019) Climatic drivers of pinyon mouse *Peromyscus truei* population dynamics in a resource-restricted environment. Popul Ecol 61:122–131. <https://doi.org/10.1002/1438-390X.1006>
- <span id="page-9-7"></span>Stah CD (1980) Vertical nesting distribution of two species of *Peromyscus* under experimental conditions. J Mammal 61:141–143
- <span id="page-9-5"></span>Sueur J, Aubin T, Simonis C (2008) Seewave, a free modular tool for sound analysis and synthesis. Bioacoustics 18:213–226. <https://doi.org/10.1080/09524622.2008.9753600>
- <span id="page-9-6"></span>Timonin M, Kalcounis-Rueppell M, Marler CA (2018) Testosterone pulses at the nest site modify ultrasonic vocalization types in a monogamous and territorial mouse. Ethology 124:804– 815.<https://doi.org/10.1111/eth.12812>
- <span id="page-9-0"></span>Waser PM, Brown CH (1986) Habitat acoustics and primate communication. Am J Primatol 10:135–154. [https://doi.org/10.](https://doi.org/10.1002/ajp.1350100205) [1002/ajp.1350100205](https://doi.org/10.1002/ajp.1350100205)
- <span id="page-9-1"></span>Waser PM, Waser MS (1977) Experimental studies of primate vocalization: Specializations for long-distance propagation. Z Tierpsychol 43:239–263.<https://doi.org/10.1111/j.1439-0310.1977.tb00073.x>
- <span id="page-9-9"></span>Whipple AV, Cobb NS, Gehring CA, Mopper S, Flores-Rentería L, Whitham TG (2019) Long-term studies reveal diferential responses to climate change for trees under soil- or herbivorerelated stress. Front Plant Sci 10:132. [https://doi.org/10.3389/fpls.](https://doi.org/10.3389/fpls.2019.00132) [2019.00132](https://doi.org/10.3389/fpls.2019.00132)
- <span id="page-9-2"></span>Wiley RH, Richards DG (1978) Physical constraints on acoustic communication in the atmosphere: Implications for the evolution of animal vocalizations. Behav Ecol Sociobiol 3:69–94. [https://doi.](https://doi.org/10.1007/BF00300047) [org/10.1007/BF00300047](https://doi.org/10.1007/BF00300047)
- <span id="page-9-8"></span>Yovel Y, Falk B, Moss CF, Ulanovsky N (2011) Active control of acoustic field-of-view in a biosonar system. PLoS Biol 9:e1001150.<https://doi.org/10.1371/journal.pbio.1001150>

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