ORIGINAL ARTICLE

# CXCR5 polymorphisms in non-Hodgkin lymphoma risk and prognosis

Bridget Charbonneau • Alice H. Wang • Matthew J. Maurer • Yan W. Asmann • Clive S. Zent • Brian K. Link • Stephen M. Ansell • George J. Weiner • Nazan Ozsan • Andrew L. Feldman • Thomas E. Witzig • Julie M. Cunningham • Ahmet Dogan • Thomas M. Habermann • Susan L. Slager • Anne J. Novak • James R. Cerhan

Received: 11 March 2013 / Accepted: 13 June 2013 / Published online: 28 June 2013 - Springer-Verlag Berlin Heidelberg 2013

Abstract CXCR5 [chemokine (C-X-C motif) receptor 5; also known as Burkitt lymphoma receptor 1 (BCR1)] is expressed on mature B-cells, subsets of  $CD4^+$  and  $CD8^+$ T-cells, and skin-derived migratory dendritic cells. Together with its ligand, CXCL13, CXCR5 is involved in guiding B-cells into the B-cell zones of secondary lymphoid organs as well as T-cell migration. This study evaluated the role of common germline genetic variation in CXCR5 in the risk and prognosis of non-Hodgkin

Electronic supplementary material The online version of this article (doi:[10.1007/s00262-013-1452-4\)](http://dx.doi.org/10.1007/s00262-013-1452-4) contains supplementary material, which is available to authorized users.

B. Charbonneau · A. H. Wang · M. J. Maurer · Y. W. Asmann  $\cdot$  S. L. Slager  $\cdot$  J. R. Cerhan ( $\boxtimes$ ) Department of Health Sciences Research, College of Medicine, Mayo Clinic, 200 First Street SW, Rochester, MN, USA e-mail: cerhan.james@mayo.edu

C. S. Zent · S. M. Ansell · T. E. Witzig · T. M. Habermann - A. J. Novak Division of Hematology, Department of Internal Medicine, College of Medicine, Mayo Clinic, 200 First Street SW, Rochester, MN, USA

B. K. Link - G. J. Weiner Division of Hematology, Oncology, Blood and Marrow Transplantation, Department of Internal Medicine, College

of Medicine, University of Iowa, Iowa City, IA, USA

N. Ozsan

Department of Pathology, Faculty of Medicine, Ege University, Bornova, İzmir, Turkey

A. L. Feldman - J. M. Cunningham - A. Dogan Department of Laboratory Medicine and Pathology, College of Medicine, Mayo Clinic, 200 First Street SW, Rochester, MN, USA

lymphoma (NHL) using a clinic-based study of 1,521 controls and 2,694 NHL cases including 710 chronic lymphocytic leukemia/small lymphocytic lymphoma, 586 diffuse large B-cell lymphoma (DLBCL), 588 follicular lymphoma (FL), 137 mantle cell lymphoma (MCL), 230 marginal zone lymphoma (MZL), and 158 peripheral T-cell lymphoma (PTCL). Of the ten CXCR5 tag SNPs in our study, five were associated with risk of NHL, with rs1790192 having the strongest association (OR 1.19, 95 % CI 1.08–1.30;  $p = 0.0003$ ). This SNP was most strongly associated with the risk of FL (OR 1.44, 95 % CI 1.25–1.66;  $p = 3.1 \times 10^{-7}$ , with a lower degree of association with DLBCL (OR 1.16, 95 % CI 1.01–1.33;  $p = 0.04$ ) and PTCL (OR 1.29, 95 % CI 1.02-1.64;  $p = 0.04$ ) but no association with the risk of MCL or MZL. For FL patients that were observed as initial disease management, the number of minor alleles of rs1790192 was associated with better event-free survival (HR 0.64; 95 % CI 0.47–0.87;  $p = 0.004$ ). These results provide additional evidence for a role of host genetic variation in CXCR5 in lymphomagenesis, particularly for FL.

Keywords Non-Hodgkin lymphoma · SNPs · Prognosis · Prospective cohort · Case-control

## Introduction

With an estimated 70,130 incident cases of non-Hodgkin lymphoma (NHL) and 18,940 deaths due to disease and an additional 16,060 incident chronic lymphocytic leukemia (CLL) cases and 4,580 death due to disease in the US in 2012 [\[1](#page-7-0)], there is a continuing need to identify factors that influence NHL risk and survival. Lymphomas classified as NHL are distinct diseases with different biology, clinical <span id="page-1-0"></span>courses, and treatment but appear to have some shared risk factors [\[2](#page-7-0)]. Therefore, a better understanding of shared and subtype-specific NHL risk factors should provide insights into the mechanisms of lymphomagenesis.

CXCR5, formerly known as Burkitt lymphoma receptor 1 (BLR1), is the receptor for CXCL13 and plays a role in B-cell and follicular T helper cell migration. Gene knockout studies in mice have also shown this receptor is important for normal development and organization of follicular zones in lymph nodes and Peyer patches [\[3](#page-7-0), [4](#page-7-0)]. Expression of this receptor has been observed in a number of malignancies, including several types of lymphoma [\[5](#page-7-0)– [8](#page-7-0)]. Recently, Song et al. [\[9](#page-7-0)] reported an association of several CXCR5 SNPs with NHL risk in a Chinese population. Additionally, genome-wide association studies (GWAS) have identified polymorphisms in CXCR5 linked to the autoimmune diseases biliary cirrhosis and multiple sclerosis [\[10](#page-7-0), [11](#page-7-0)]. We therefore conducted the first study in a Western population of common germline genetic variation in CXCR5 in relation to risk of NHL and its most common individual diseases (subtypes): follicular lymphoma (FL), diffuse large B-cell lymphoma (DLBCL), marginal zone lymphoma (MZL), mantle cell lymphoma (MCL), peripheral T-cell lymphoma (PTCL), and chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL). We also assessed these variants in relation to prognosis of some of the more common subtypes.

## Materials and methods

# Study population

The Human Subjects Institutional Review Board at the Mayo Clinic and the University of Iowa reviewed and approved the current study. As previously described, the Mayo Clinic Case–Control Study of Lymphoma is a clinicbased study of incident cases and frequency-matched controls (based on age, sex, and residence) [[12\]](#page-8-0). Briefly, cases were lymphoma patients seen at Mayo Clinic Rochester and enrolled within 9 months of diagnosis, who were aged 18 years or older; a resident of Minnesota, Iowa, or Wisconsin; and HIV negative at the time of diagnosis. Diagnoses were confirmed by study hematopathologists and classified according to the recent World Health Organization (WHO) classification of hematopoietic neoplasms [\[13](#page-8-0), [14](#page-8-0)]. For composite or discordant histologies, the lower-grade component was used to classify the subtype for analysis of risk (assumed to have existed first), while the higher-grade component was used to classify the subtype for analysis of outcome (since that would determine treatment strategy).

Controls were ascertained from patients visiting the general medicine clinics at Mayo Clinic Rochester for prescheduled general medical examinations and were frequency matched to the cases on age, sex, and residence. Control patients were excluded if they had prior diagnoses of lymphoma, leukemia, or HIV infection.

We also included newly diagnosed lymphoma patients from the University of Iowa and Mayo Clinic Lymphoma Specialized Program of Research Excellence (SPORE); these patients had the same eligibility requirements as the other cases, except that they could be a resident of any US state and there was no matching control group.

All participants completed questionnaires, and a peripheral blood sample was collected. DNA was extracted from peripheral blood sample using a standard procedure (Gentra Inc., Minneapolis, MN, USA). This analysis included 2,694 NHL (including CLL) cases and 1,522 controls enrolled from 9/1/2002 through 12/31/2009.

For all case patients, clinical, laboratory, and treatment data at the time of diagnosis and initial treatment were abstracted. Cell of origin for germinal center B-cell-like (GCB) and non-GCB subtypes was defined according to the Hans algorithm [\[15](#page-8-0)]. All patients were then actively followed every 6 months for the first 3 years, and then annually thereafter. We verified all reports of disease progression, retreatment, transformation, and death against medical records [\[16](#page-8-0)].

For the transformation analysis, we only included grade I-IIIa FL with no evidence of clinical or pathological transformation at the time of initial diagnosis. Transformation was defined as refractory/recurrent disease with either clinical or pathological diagnosis of lymphoma, as previously described [\[17](#page-8-0)]. Pathology-defined transformation entailed a biopsy-confirmed subtype of FLIIIb, DLBCL, unclassifiable B-cell lymphoma with features intermediate between diffuse large B-cell lymphoma and Burkitt lymphoma, and high-grade B-cell lymphoma (including Burkitt), or evidence of transformation per pathologist report [[18\]](#page-8-0). Clinical transformation was defined using previously published [[19](#page-8-0)] clinical indication of transformation (sudden rise in LDH, rapid discordant localized nodal growth, new involvement of unusual extranodal sites, new B symptoms, or hypercalcemia) or a statement in the medical record that the treating physician was clinically managing the patient as a transformation at the time of recurrence.

# Genotyping

A total of ten CXCR5 tag SNPs were successfully genotyped as part of a larger genotyping project using a custom Illumina Infinium iSelect array (Illumina, San Diego CA). Tag SNPs were selected from Hapmap (Phase 2, release 23a, CEU population) using a standard tagging approach ( $r^2 \ge 0.80$  and minor allele frequency (MAF)  $>0.05$ ) and included SNPs 5 kb upstream and downstream from the gene. Standard genotyping quality control procedures were performed and included duplicate genotyping, dropping samples or SNPs with call rates  $\langle 95 \, % \rangle$ , and testing for Hardy–Weinberg equilibrium (HWE). We found  $>99.99$  % genotyping concordance for 406 SNPs previously genotyped on 3,116 participants using an Illumina GoldenGate platform and for 2,477 SNPs on 386 participants using the Illumina 660 platform. All of the CXCR5 variants had HWE  $p > 0.1$  and SNP call rates  $>99.8$  %.

# Statistical analysis

Main analyses used SAS version 9.2 (SAS Institute Inc., Cary, NC, USA) and R [\(http://www.r-project.org/](http://www.r-project.org/)). Tests for HWE were done using either the Pearson's goodnessof-fit test or the Fisher's exact test, where appropriate. All analyses excluded known non-Caucasians to minimize bias due to population stratification; participants who refused or had missing race/ethnicity were retained in the dataset (presumed Caucasian, given our source population).

For evaluation of risk, we used unconditional logistic regression to estimate odds ratios (OR) and 95 % confidence intervals (CI) for the association of NHL case status (overall NHL and for each common NHL subtype) with each SNP. Analyses were adjusted for age (including its functional form) and gender, and the most common homozygous genotype was treated as the referent category for each of the SNPs. Each SNP was modeled in a logadditive manner in the regression model, and the Wald  $p$  value was used to assess significance. To account for multiple testing of 10 independent SNPs ( $r^2$  < 0.6 for all pairs), we used the Bonferroni-corrected  $p$  value of 0.005 (0.05/10) as our significance threshold.

For evaluation of prognosis, we used Cox proportional hazards regression to estimate hazard ratios (HRs) and 95 % CI. We defined event-free survival (EFS) as the time from diagnosis to disease progression, retreatment, or death due to any cause and overall survival (OS) as time from diagnosis to death due to any cause. Patients without an event were censored at time of last known follow-up. All survival analyses included adjustment for subtype-specific clinical risk score (i.e., IPI [[20](#page-8-0)], FLIPI [\[21](#page-8-0)], MIPI [\[22](#page-8-0)], etc.) as appropriate. For evaluation of FL transformation, we used Cox proportional hazards regression with the inclusion of death without transformation as a competing risk.

#### **Bioinformatics**

All SNPs in LD (at  $r^2 \ge 0.5$ ) with the 10 CXCR5 SNPs in this study were extracted from the 1,000 genomes database. First, all SNPs were annotated using the UCSC tracks downloaded from the UCSC golden path database [\(http://hgdownload.cse.ucsc.edu/goldenpath/hg19/](http://hgdownload.cse.ucsc.edu/goldenpath/hg19/)). Specifically, we looked for SNPs that are located within a predicted conserved element using the PhastCons program, where the predictions are based on a phylogenetic hidden Markov model (compgen.bscb.cornell.edu/phast). The conserved SNPs were then overlapped with other functional tracks including CpG Islands, transcription-factorbinding sites, enhancers, TargetScan-microRNA-binding sites, and Encode epigenetic markers. We also assessed SNPs in CXCR5 for potential location in microRNA (miRNA)-binding sites using the PolymiRTS Database 2.0 [\[23](#page-8-0)] [\(http://compbio.uthsc.edu/miRSNP/](http://compbio.uthsc.edu/miRSNP/)). Additional information on correlated variants and their potential function was obtained from HaploReg v2 [\(http://www.](http://www.broadinstitute.org/mammals/haploreg/haploreg.php) [broadinstitute.org/mammals/haploreg/haploreg.php\)](http://www.broadinstitute.org/mammals/haploreg/haploreg.php) [\[24\]](#page-8-0).

# Results

CXCR5 and risk of NHL and NHL subtypes

The mean age of the 2,694 NHL patients was 61.6 years  $(SD = 13.3)$  and the 1,521 controls was 61.6 years  $(SD = 13.7)$ ; 41 % of cases were female compared to 49 % of the controls. Figure [1](#page-3-0) contains an LD block of SNPs assessed for the CXCR5 gene region based on allele frequencies in our control population. Only two pairs of SNPs were in modest LD with each other: rs11217078 with  $rs523604$   $(r^2 = 0.53)$  and  $rs497916$  with  $rs3922$  $(r^2 = 0.51)$ . All other SNP pairs had  $r^2 < 0.40$ . Total bin coverage for CXCR5 was 91 %.

Of the ten CXCR5 tag SNPs, two were statistically significantly associated with overall NHL risk at the Bonferroni-corrected threshold of  $p < 0.005$ , all with small effect sizes (Table [1\)](#page-4-0): rs1790192 (OR 1.19, 95 % CI 1.08–1.30;  $p = 0.0003$ ) and rs497916 (OR 1.16, 95 % CI 1.05–1.28;  $p = 0.003$ ).

FL- and DLBCL-specific risk associations are presented in Table [2,](#page-4-0) with stronger associations and more SNPs related to risk of FL and the GCB subtype of DLBCL. The most prominent association was observed for rs1790192 with FL risk (OR 1.44, 95 % CI 1.25–1.66;  $p = 3.1 \times 10^{-7}$ ). This SNP was also associated with DLBCL risk (OR 1.16, 95 % CI 1.01–1.33;  $p = 0.04$ ), particularly GCB DLBCL subtype (OR 1.36, 95 % CI 1.05–1.77;  $p = 0.02$ ), although neither association was

<span id="page-3-0"></span>Fig. 1 Chromosome 11q23.3 gene CXCR5 structure and tagSNP mapping.  $Top$  -Log10 p value for trend across tagSNPs for all NHL risk (dark circles) and FL (open circles). Bottom linkage disequilibrium plot of SNPs genotyped in this analysis (darker shading indicates higher  $r^2$  correlation values between SNPs; numbers are |D'| values)



statistically significant after accounting for multiple testing. The SNP rs497916 was also associated with both FL (OR 1.32, 95 % CI 1.14–1.53;  $p = 0.0002$ ) and GCB DLBCL (OR 1.50, 95 % CI 1.15–1.97;  $p = 0.003$ ) risk. The variant rs630923 was only statistically significantly associated with risk of FL (OR 1.30, 95 % CI 1.09–1.55;  $p = 0.003$ ) but not DLBCL, while rs3922 was associated with risk of GCB DLBCL (OR 1.46, 95 % CI 1.12–1.89;  $p = 0.005$ ) but not non-GCB DLBCL or FL.

Associations between several other NHL subtypes and SNPs in *CXCR5* are reported in Table [3](#page-5-0). CLL risk was only statistically significantly associated with rs11217078 (OR 1.23, 95 % CI 1.07–1.40;  $p = 0.003$ ) after accounting for multiple testing; additional associations were observed for CLL risk with rs630923 (OR 1.22, 95 % CI 1.03–1.45;  $p = 0.02$ ), and rs523604 (OR 1.20, 95 % CI 1.05–1.36;  $p = 0.006$ ) but did meet the  $p < 0.005$  criteria. For the other subtypes, rs1623316 (OR 0.80, 95 % CI 0.65–0.99;

<span id="page-4-0"></span>



Bold values indicate  $p < 0.005$ 

 $a$  Ordinal model OR and  $p$  value, adjusted for age and gender

<sup>b</sup> Includes 710 CLL/SLL, 588 FL, 586 DLBCL, 230 MZL, 137 MCL, 158 PTCL, and 285 that did not fit into any of the other major categories

Table 2 Risk of FL and DLBCL for selected SNPs in CXCR5 (11q23.3)

CXCR5 SNP	FL $(N = 588)$		DLBCL $(N = 586)$		DLBCL non-GCB <sup>b</sup> ( $N = 76$ )		DLBCL GCB <sup>c</sup> ( $N = 127$ )	
	OR (95 % $Cla$ )	$\boldsymbol{p}$	OR (95 % $Cla$ )	$\boldsymbol{p}$	OR (95 % $CI)^a$ )	$\boldsymbol{p}$	OR (95 % $CI)^a$	$\boldsymbol{p}$
rs11217078	1.16(1.00, 1.33)	0.044	1.08(0.93, 1.24)	0.31	1.32(0.94, 1.84)	0.11	1.22(0.93, 1.60)	0.15
rs10892306	1.33(1.03, 1.71)	0.030	1.14(0.87, 1.49)	0.34	1.49(0.83, 2.68)	0.18	0.93(0.54, 1.61)	0.79
rs630923	1.30(1.09, 1.55)	0.003	1.09(0.91, 1.31)	0.36	1.23(0.80, 1.89)	0.34	1.29(0.93, 1.81)	0.13
rs10892307	0.90(0.74, 1.10)	0.30	0.97(0.80, 1.17)	0.75	0.71(0.42, 1.19)	0.19	1.03(0.72, 1.48)	0.85
rs523604	1.01(0.88, 1.15)	0.90	1.03(0.90, 1.18)	0.70	1.00(0.72, 1.38)	0.98	1.16(0.89, 1.50)	0.28
rs1790192	1.44(1.25, 1.66)	$3.1E - 07$	1.16(1.01, 1.33)	0.038	1.20(0.86, 1.68)	0.28	1.36(1.05, 1.77)	0.020
rs1623316	0.90(0.78, 1.03)	0.13	0.94(0.81, 1.08)	0.36	1.05(0.75, 1.47)	0.76	0.88(0.68, 1.16)	0.37
rs497916	1.32(1.14, 1.53)	0.0002	1.13(0.98, 1.32)	0.097	1.33(0.94, 1.89)	0.11	1.50(1.15, 1.97)	0.003
rs12363277	1.03(0.79, 1.34)	0.82	0.98(0.75, 1.28)	0.88	1.07(0.57, 2.01)	0.82	1.06(0.64, 1.75)	0.82
rs3922	1.13(0.98, 1.29)	0.085	1.07(0.94, 1.23)	0.30	1.10(0.79, 1.54)	0.57	$1.46$ $(1.12, 1.89)$	0.005

Bold values indicate  $p < 0.005$ 

 $a$  Ordinal model OR and  $p$  value, adjusted for age and gender

<sup>b</sup> non-Germinal center B-cell-like phenotype

<sup>c</sup> Germinal center B-cell-like phenotype

 $p = 0.04$ ) was associated with MZL risk, and PTCL was associated with rs10892306 (OR 1.57, 95 % CI 1.04–2.39;  $p = 0.03$ ), rs1790192 (OR 1.29, 95 % CI 1.02-1.64;  $p = 0.04$ ), and rs12363277 (OR 0.49, 95 % CI 0.27–0.91;  $p = 0.02$ ), although none met the multiple testing threshold.

## CXCR5 and NHL prognosis

Prognosis was evaluated for some of the most common NHL subtypes including DLBCL  $(N = 581)$ , FL  $(N = 548)$ , MCL  $(N = 140)$ , and PTCL  $(N = 109)$ ; numbers were slightly different from the risk analysis based on subtype classification of composite or discordant histologies (see ''[Materials and methods](#page-1-0)'' section) or availability of clinical data. Clinical and treatment characteristics for DLBCL and FL are provided in Supplemental Tables 1 and 2, respectively. We did not assess CLL survival in this paper, due to few deaths for an analysis of overall survival and a lack of an analogous event for EFS endpoint.

Results for DLBCL OS and EFS (all and by GCB/non-GCB subtype) restricted to patients who received R-CHOP treatment are presented in Supplemental Table 3. For DLBCL, no associations were found between CXCR5 SNPs, and EFS, overall or for GCB and non-GCB subtypes. However, in DLBCL patients who received

CXCR5 SNP	CLL $(N = 710)$		$MZL (N = 230)$		MCL $(N = 137)$		PTCL $(N = 158)$	
	OR (95 % CI)*	p	OR (95 % CI)*	p	OR (95 % CI)*	$\boldsymbol{p}$	OR (95 % CI)*	$\boldsymbol{p}$
rs11217078	1.23(1.07, 1.40)	0.003	1.11(0.90, 1.37)	0.31	1.05(0.81, 1.36)	0.73	0.94(0.73, 1.21)	0.63
rs10892306	0.97(0.74, 1.26)	0.82	1.08(0.73, 1.61)	0.70	1.13(0.69, 1.85)	0.63	1.58(1.04, 2.39)	0.03
rs630923	1.22(1.03, 1.45)	0.02	1.00(0.76, 1.32)	0.99	1.02(0.72, 1.44)	0.93	0.99(0.71, 1.36)	0.93
rs10892307	1.05(0.88, 1.26)	0.57	0.97(0.73, 1.27)	0.81	1.19(0.85, 1.66)	0.32	0.89(0.64, 1.25)	0.51
rs523604	1.20(1.05, 1.36)	0.006	1.03(0.84, 1.25)	0.79	0.93(0.72, 1.19)	0.55	0.92(0.73, 1.17)	0.50
rs1790192	1.07(0.94, 1.22)	0.29	1.13(0.92, 1.38)	0.24	1.09(0.84, 1.42)	0.50	1.29(1.02, 1.64)	0.04
rs1623316	0.89(0.78, 1.02)	0.10	0.80(0.65, 0.99)	0.04	1.14(0.89, 1.47)	0.30	0.86(0.67, 1.09)	0.21
rs497916	1.11(0.96, 1.27)	0.17	$1.08$ $(0.87, 1.34)$	0.50	1.20(0.91, 1.58)	0.20	0.94(0.72, 1.23)	0.65
rs12363277	1.10(0.87, 1.41)	0.43	0.83(0.54, 1.27)	0.39	1.09(0.68, 1.75)	0.72	0.49(0.27, 0.91)	0.02
rs3922	1.09(0.96, 1.25)	0.18	1.07(0.88, 1.31)	0.51	1.14(0.88, 1.47)	0.32	0.92(0.72, 1.17)	0.50

<span id="page-5-0"></span>Table 3 Risk of other NHL subtypes for selected SNPs in CXCR5 (11q23.3)

\* Ordinal model OR and p value, adjusted for age and gender

Bold values indicate  $p < 0.005$ 

R-CHOP, there was a suggestive association between OS and rs12363277 (HR 0.54, 95 % CI 0.30–0.98;  $p = 0.04$ ) and rs3922 (HR 0.78, 95 % CI 0.62–0.99;  $p = 0.04$ ), although not statistically significant at the Bonferroni corrected  $p < 0.005$ . When assessing all FL cases, the CXCR5 SNPs were not associated with EFS or OS (Supplemental Table 4). However, when cases were stratified by treatment groups, the number of minor alleles of rs1790192 was associated with improved EFS in patients who were initially observed instead of receiving treatment (HR 0.64, 95 % CI 0.47–0.87;  $p = 0.004$ ) (Supplemental Table 4 and Fig. 2). Additionally, there was an inverse association between the number of rs3922 minor alleles and transformation of FL to DLBCL (HR 0.62, 95 % CI 0.41–0.93;  $p = 0.02$ ), although this did not meet the Bonferroni threshold. None of the CXCR5 variants assessed were associated with EFS or OS in MCL (Supplemental Table 5) or PTCL (Supplemental Table 6).

# Discussion

Polymorphisms in CXCR5 were associated with risk of several NHL subtypes after accounting for multiple testing, with the most notable being an increased risk of FL associated with the minor allele of rs1790192. The number of minor alleles of this SNP was also modestly associated with increased risk of overall DLBCL, and the association was stronger for GCB DLBCL. The latter finding is consistent with the biology of GCB DLBCL and FL, which arise from germinal center B-cells, and a subset of GCB DLBCL share the same t(14;18) translocation as most FLs

Fig. 2 rs1790192-dependent EFS in patients with FL who were observed as initial disease management. Percent event-free survival in observed FL patients  $(N = 172)$  (event defined as disease progression, initiation of treatment, or death due to any cause) according to rs1790192 genotype. Hazard ratio was calculated using Cox proportional hazards regression adjusted for FLIPI



 $[25]$  $[25]$ . On the other hand, the number of minor alleles of rs1790192 was statistically significantly associated with superior EFS in FL, and transformation of FL to DLBCL was modestly inversely associated with the number of minor alleles of rs3922, suggesting biologic differences of this SNP in etiology versus disease progression. We are the first to our knowledge to assess CXCR5 SNPs in association with survival in DLBCL, FL, MCL, and PTCL.

The role of CXCR5 in lymph node development and B-cell recruitment [[26\]](#page-8-0) provides a plausible biological role for this chemokine in lymphoma development and progression. This receptor, normally expressed on T follicular helper cells and B-cells, interacts with chemokine CXCL13 with functions including inducing B-cell migration [\[3](#page-7-0), [4\]](#page-7-0) and augmenting B-cell activation during BCR signaling [\[27](#page-8-0)]. In at least two  $HIV^+$  patient populations, serum CXCL13 levels were higher in HIV-infected patients who developed NHL than those who did not [[28,](#page-8-0) [29](#page-8-0)]. This finding was also confirmed in an immunocompetent population in the Women's Health Initiative Observational Study cohort, where elevated serum CXCL13 was significantly associated with increased risk of B-NHL [\[30](#page-9-0)]. Additionally, Burkle et al. [[31\]](#page-9-0) previously reported that CLL cells expressed high levels of CXCR5, serum CXCL13 was elevated in CLL patients, and CLL B-cells exposed to CXCL13 exhibited prolonged p44/42 MAPK activation, considered to provide a pro-survival signal [\[32](#page-9-0)]. Together, these studies provide support for CXCL13/ CXCR5 involvement in NHL development and progression.

While SNPs in *CXCR5* have not been reported extensively in association with disease states, one recent GWAS found an association of CXCR5 rs6421571 with risk of primary biliary cirrhosis, an autoimmune disease of the liver [\[10](#page-7-0)]. Also of potential interest is the rs630923 SNP, as increased copies of the A allele were found in our study to be modestly associated with increased risk of CLL and FL. The A allele of this SNP was also recently reported to be associated with increased CXCL13 levels in serums of cases and controls in an  $HIV^+$  NHL case–control study [\[29](#page-8-0)]. It should be noted that neither this SNP nor the other CXCR5 tag SNPs in this study were associated with AIDS-NHL; however, the minor allele of CXCL13 tag SNP, rs355689, was associated with both reduced AIDS-NHL risk and reduced CXCL13 serum levels [\[29](#page-8-0)]. The C allele of the rs630923 SNP was also found in a recent GWAS to be associated with increased risk of multiple sclerosis (MS) [\[11](#page-7-0)]. One study, with extremely small numbers, found an association between history of MS or first-degree relative with MS and NHL risk [[33\]](#page-9-0). An increased risk of NHL, particularly FL and DLBCL, was also found for combined autoimmune conditions, which included MS, in a population-based case–control study of women [[34\]](#page-9-0). However, in a large pooled analysis of 12 case–control studies within the InterLymph Consortium, there was no association between MS and NHL risk [[35\]](#page-9-0), and in a large cohort of US veterans, an inverse association was found between MS diagnosis and NHL risk [[36\]](#page-9-0). While the association between MS and NHL is controversial, given the role of B-cells in MS [[37\]](#page-9-0), it would not be surprising that a common CXCR5-related mechanism drives formation of both diseases.

In a recent case–control study in a Chinese population, including 404 NHL cases and 456 age-matched controls, four CXCR5 polymorphisms were assessed for association with NHL [[9\]](#page-7-0). The authors reported an association between rs6421571 and overall NHL risk, Ann Arbor stage, and B-cell subtype. Additionally, they found rs80202369 and the TGG haplotype of rs6421571, rs80202369, and rs78440425 were associated with overall NHL risk [[9\]](#page-7-0). It should be noted these particular SNPs were not included in our CXCR5 tag SNPs, nor were they in LD with any of our tag SNPs at  $r^2 > 0.2$  using information from the 1,000 genomes European population. In this representative Caucasian population, rs80202369 was monomorphic, and rs6421571 and rs78440425 had MAFs of 0.178 and 0.007, respectively. While we did not assess these specific SNPs, the association we found between overall NHL and several SNPs in the CXCR5 gene provides additional support for a potential role of CXCR5 in lymphomagenesis.

The finding that the minor allele of rs1790192 is associated with elevated risk but improved survival suggests that the role of CXCR5 differs between lymphoma initiation and progression. Based on the limited available data, we can only speculate that the major allele (associated with reduced risk and poor prognosis) results in higher expression of CXCR5 compared to the minor allele. Because CXCR5 is associated with immune cell recruitment, it is possible that higher expression of this chemokine in the early stages of lymphomagenesis could enhance immune surveillance and thus provide protection/reduced risk [\[38](#page-9-0)]. However, if premalignant cells manage to escape immune surveillance, proliferate and establish themselves as a tumor, immune cells recruited to the tumor microenvironment may be induced to develop a suppressive phenotype, which could in turn enhance tumor growth and progression accounting for reduced survival in these patients [[38\]](#page-9-0). This hypothesis will need to be tested in future studies.

A general search of the UCSC bioinformatics tools and the PolymiRTS Database 2.0 found little evidence for function of any of the NHL risk or survival associated CXCR5 SNPs or SNPs with which they were in moderateto-high ( $r^2 \ge 0.50$ ) LD in the 1,000 genomes population, with a few exceptions. The rs3922 polymorphism is found in a binding site for three known miRNAs. However, it is only conserved across 3 other species, so there is lower

<span id="page-7-0"></span>likelihood of a functional consequence. This SNP is also in moderate LD with rs598207 ( $r^2 = 0.52$ ) and high LD with rs1704819 ( $r^2 = 0.99$ ), both synonymous SNPs in the coding region of CXCR5. While originally thought to be neutral, synonymous SNPs can alter a number of functions, such as mRNA secondary structures (related to stability), splicing, and normal co-translational folding [\[39](#page-9-0)]. We further examined rs1790192 using HaploReg ([http://www.](http://www.broadinstitute.org/mammals/haploreg/haploreg.php) [broadinstitute.org/mammals/haploreg/haploreg.php\)](http://www.broadinstitute.org/mammals/haploreg/haploreg.php). According to HaploReg, rs1790192 lies in a region of 33 regulatory motifs across a number of different cell types. Based on histone marks, in GM12878 lymphoblastoid cell lines this SNP falls in a poised promoter region and in K562 leukemia cell line this SNP falls in a weak enhancer region. We note that this does not mean that the altered base pair at this location would alter function or expression of the gene product, only that its location in regulatory regions makes this a possibility. It should also be noted that this SNP is not located in an evolutionarily conserved domain. We also note that these tools assess predicted function, and laboratory studies are needed to assess actual consequences of these polymorphisms on gene products. Finally, CXCR5 overlaps the BCL9L gene on chromosome 11, which encodes a protein with high-sequence homology to BCL9 [\[40](#page-9-0)], discovered in lymphoblastic leukemia cells with a  $t(1;14)(q21;q32)$  translocation [[41](#page-9-0)]. BCL9L was shown recently to be related to progression of intestinal tumors [\[42](#page-9-0)], and we cannot rule out a potential role for these SNPs on BCL9L expression.

Strengths of our study include a central pathology review of case phenotype; a representative control group; the large sample size and ability to evaluate common NHL subtypes, as well as GCB and non-GCB DLBCL; assessment of risk and prognosis in the same population; and relatively detailed clinical and outcome data. There were also limitations, including lack of racial and ethnic diversity, which limits generalizability. Additionally, while the bin coverage for *CXCR5* in this study was quite high, we cannot rule out additional associations with SNPs that were not covered by our tagging approach.

In this study, we found that several CXCR5 polymorphisms were related to risk of NHL subtypes and prognosis in FL. The most noteworthy SNP was rs1790192, which was strongly associated with increased risk of FL, while also being associated with superior survival in FL patients who were observed for initial disease management. Future functional assessment of these SNPs could provide useful insights into how CXCR5 might be regulating tumor promotion and progression.

Acknowledgments This study was supported by National Institutes of Health (NIH) Grants R01 CA92153, R01 CA129539, and P50 CA97274; Dr. Charbonneau was supported by R25 CA92049.

Conflict of interest The authors have no known conflict of interests.

#### References

- 1. Siegel R, Naishadham D, Jemal A (2012) Cancer statistics, 2012. CA Cancer J Clin 62(1):10–29. doi:[10.3322/caac.20138](http://dx.doi.org/10.3322/caac.20138)
- 2. Morton LM, Wang SS, Cozen W, Linet MS, Chatterjee N, Davis S, Severson RK, Colt JS, Vasef MA, Rothman N, Blair A, Bernstein L, Cross AJ, De Roos AJ, Engels EA, Hein DW, Hill DA, Kelemen LE, Lim U, Lynch CF, Schenk M, Wacholder S, Ward MH, Hoar Zahm S, Chanock SJ, Cerhan JR, Hartge P (2008) Etiologic heterogeneity among non-Hodgkin lymphoma subtypes. Blood 112(13):5150–5160. doi[:10.1182/blood-](http://dx.doi.org/10.1182/blood-2008-01-133587)[2008-01-133587](http://dx.doi.org/10.1182/blood-2008-01-133587)
- 3. Forster R, Mattis AE, Kremmer E, Wolf E, Brem G, Lipp M (1996) A putative chemokine receptor, BLR1, directs B cell migration to defined lymphoid organs and specific anatomic compartments of the spleen. Cell 87(6):1037–1047
- 4. Ansel KM, Ngo VN, Hyman PL, Luther SA, Forster R, Sedgwick JD, Browning JL, Lipp M, Cyster JG (2000) A chemokine-driven positive feedback loop organizes lymphoid follicles. Nature 406(6793):309–314. doi[:10.1038/35018581](http://dx.doi.org/10.1038/35018581)
- 5. Kurtova AV, Tamayo AT, Ford RJ, Burger JA (2009) Mantle cell lymphoma cells express high levels of CXCR4, CXCR5, and VLA-4 (CD49d): importance for interactions with the stromal microenvironment and specific targeting. Blood 113(19):4604–4613. doi: [10.1182/blood-2008-10-185827](http://dx.doi.org/10.1182/blood-2008-10-185827)
- 6. Krenacs L, Schaerli P, Kis G, Bagdi E (2006) Phenotype of neoplastic cells in angioimmunoblastic T-cell lymphoma is consistent with activated follicular B helper T cells. Blood 108(3):1110–1111. doi:[10.1182/blood-2006-01-0394](http://dx.doi.org/10.1182/blood-2006-01-0394)
- 7. Deutsch AJ, Aigelsreiter A, Steinbauer E, Fruhwirth M, Kerl H, Beham-Schmid C, Schaider H, Neumeister P (2008) Distinct signatures of B-cell homeostatic and activation-dependent chemokine receptors in the development and progression of extragastric MALT lymphomas. J Pathol 215(4):431–444. doi: [10.1002/path.2372](http://dx.doi.org/10.1002/path.2372)
- 8. Machado L, Jarrett R, Morgan S, Murray P, Hunter B, Hamilton E, Crocker J, Thomas W, Steven N, Ismail T, Chapman A, Adams DH, Lee SP (2009) Expression and function of T cell homing molecules in Hodgkin's lymphoma. Cancer Immunol Immunother 58(1):85–94. doi:[10.1007/s00262-008-0528-z](http://dx.doi.org/10.1007/s00262-008-0528-z)
- 9. Song H, Tong D, Cha Z, Bai J (2012) C-X-C chemokine receptor type 5 gene polymorphisms are associated with non-Hodgkin lymphoma. Mol Biol Rep. doi[:10.1007/s11033-012-1717-6](http://dx.doi.org/10.1007/s11033-012-1717-6)
- 10. Mells GF, Floyd JA, Morley KI, Cordell HJ, Franklin CS, Shin SY, Heneghan MA, Neuberger JM, Donaldson PT, Day DB, Ducker SJ, Muriithi AW, Wheater EF, Hammond CJ, Dawwas MF, Jones DE, Peltonen L, Alexander GJ, Sandford RN, Anderson CA (2011) Genome-wide association study identifies 12 new susceptibility loci for primary biliary cirrhosis. Nat Genet 43(4):329–332. doi[:10.1038/ng.789](http://dx.doi.org/10.1038/ng.789)
- 11. Sawcer S, Hellenthal G, Pirinen M, Spencer CC, Patsopoulos NA, Moutsianas L, Dilthey A, Su Z, Freeman C, Hunt SE, Edkins S, Gray E, Booth DR, Potter SC, Goris A, Band G, Oturai AB, Strange A, Saarela J, Bellenguez C, Fontaine B, Gillman M, Hemmer B, Gwilliam R, Zipp F, Jayakumar A, Martin R, Leslie S, Hawkins S, Giannoulatou E, D'Alfonso S, Blackburn H, Martinelli Boneschi F, Liddle J, Harbo HF, Perez ML, Spurkland A, Waller MJ, Mycko MP, Ricketts M, Comabella M, Hammond N, Kockum I, McCann OT, Ban M, Whittaker P, Kemppinen A,

<span id="page-8-0"></span>Weston P, Hawkins C, Widaa S, Zajicek J, Dronov S, Robertson N, Bumpstead SJ, Barcellos LF, Ravindrarajah R, Abraham R, Alfredsson L, Ardlie K, Aubin C, Baker A, Baker K, Baranzini SE, Bergamaschi L, Bergamaschi R, Bernstein A, Berthele A, Boggild M, Bradfield JP, Brassat D, Broadley SA, Buck D, Butzkueven H, Capra R, Carroll WM, Cavalla P, Celius EG, Cepok S, Chiavacci R, Clerget-Darpoux F, Clysters K, Comi G, Cossburn M, Cournu-Rebeix I, Cox MB, Cozen W, Cree BA, Cross AH, Cusi D, Daly MJ, Davis E, de Bakker PI, Debouverie M, D'Hooghe M B, Dixon K, Dobosi R, Dubois B, Ellinghaus D, Elovaara I, Esposito F, Fontenille C, Foote S, Franke A, Galimberti D, Ghezzi A, Glessner J, Gomez R, Gout O, Graham C, Grant SF, Guerini FR, Hakonarson H, Hall P, Hamsten A, Hartung HP, Heard RN, Heath S, Hobart J, Hoshi M, Infante-Duarte C, Ingram G, Ingram W, Islam T, Jagodic M, Kabesch M, Kermode AG, Kilpatrick TJ, Kim C, Klopp N, Koivisto K, Larsson M, Lathrop M, Lechner-Scott JS, Leone MA, Leppa V, Liljedahl U, Bomfim IL, Lincoln RR, Link J, Liu J, Lorentzen AR, Lupoli S, Macciardi F, Mack T, Marriott M, Martinelli V, Mason D, McCauley JL, Mentch F, Mero IL, Mihalova T, Montalban X, Mottershead J, Myhr KM, Naldi P, Ollier W, Page A, Palotie A, Pelletier J, Piccio L, Pickersgill T, Piehl F, Pobywajlo S, Quach HL, Ramsay PP, Reunanen M, Reynolds R, Rioux JD, Rodegher M, Roesner S, Rubio JP, Ruckert IM, Salvetti M, Salvi E, Santaniello A, Schaefer CA, Schreiber S, Schulze C, Scott RJ, Sellebjerg F, Selmaj KW, Sexton D, Shen L, Simms-Acuna B, Skidmore S, Sleiman PM, Smestad C, Sorensen PS, Sondergaard HB, Stankovich J, Strange RC, Sulonen AM, Sundqvist E, Syvanen AC, Taddeo F, Taylor B, Blackwell JM, Tienari P, Bramon E, Tourbah A, Brown MA, Tronczynska E, Casas JP, Tubridy N, Corvin A, Vickery J, Jankowski J, Villoslada P, Markus HS, Wang K, Mathew CG, Wason J, Palmer CN, Wichmann HE, Plomin R, Willoughby E, Rautanen A, Winkelmann J, Wittig M, Trembath RC, Yaouanq J, Viswanathan AC, Zhang H, Wood NW, Zuvich R, Deloukas P, Langford C, Duncanson A, Oksenberg JR, Pericak-Vance MA, Haines JL, Olsson T, Hillert J, Ivinson AJ, De Jager PL, Peltonen L, Stewart GJ, Hafler DA, Hauser SL, McVean G, Donnelly P, Compston A (2011) Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. Nature 476(7359):214–219. doi:[10.1038/nature10251](http://dx.doi.org/10.1038/nature10251)

- 12. Cerhan JR, Fredericksen ZS, Wang AH, Habermann TM, Kay NE, Macon WR, Cunningham JM, Shanafelt TD, Ansell SM, Call TG, Witzig TE, Slager SL, Liebow M (2011) Design and validity of a clinic-based case-control study on the molecular epidemiology of lymphoma. Int J Mol Epidemiol Genet 2(2):95–113
- 13. Jaffe ES, Harris NL, Stein H, Vardiman JW (2001) World Health Organization classification of tumours: pathology and genetics, tumours of haematopoietic and lymphoid tissues. IARC Press, Lyon
- 14. Campo E, Swerdlow SH, Harris NL, Pileri S, Stein H, Jaffe ES (2011) The 2008 WHO classification of lymphoid neoplasms and beyond: evolving concepts and practical applications. Blood 117(19):5019–5032. doi[:10.1182/blood-2011-01-293050](http://dx.doi.org/10.1182/blood-2011-01-293050)
- 15. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G, Muller-Hermelink HK, Campo E, Braziel RM, Jaffe ES, Pan Z, Farinha P, Smith LM, Falini B, Banham AH, Rosenwald A, Staudt LM, Connors JM, Armitage JO, Chan WC (2004) Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. Blood 103(1):275–282. doi[:10.1182/blood-2003-](http://dx.doi.org/10.1182/blood-2003-05-1545) [05-1545](http://dx.doi.org/10.1182/blood-2003-05-1545)
- 16. Drake MT, Maurer MJ, Link BK, Habermann TM, Ansell SM, Micallef IN, Kelly JL, Macon WR, Nowakowski GS, Inwards DJ, Johnston PB, Singh RJ, Allmer C, Slager SL, Weiner GJ, Witzig TE, Cerhan JR (2010) Vitamin D insufficiency and prognosis in

non-Hodgkin's lymphoma. J Clin Oncol 28(27):4191–4198. doi: [10.1200/JCO.2010.28.6674](http://dx.doi.org/10.1200/JCO.2010.28.6674)

- 17. Link BK, Maurer MJ, Nowakowski GS, Ansell SM, Macon WR, Syrbu SI, Slager SL, Thompson CA, Inwards DJ, Johnston PB, Colgan JP, Witzig TE, Habermann TM, Cerhan JR (2013) Rates and outcomes of follicular lymphoma transformation in the immunochemotherapy Era: a report from the University of Iowa/ Mayo Clinic SPORE molecular epidemiology resource. J Clin Oncol (in press)
- 18. Lossos IS, Gascoyne RD (2011) Transformation of follicular lymphoma. Best Pract Res Clin Haematol 24(2):147–163. doi: [10.1016/j.beha.2011.02.006](http://dx.doi.org/10.1016/j.beha.2011.02.006)
- 19. Al-Tourah AJ, Gill KK, Chhanabhai M, Hoskins PJ, Klasa RJ, Savage KJ, Sehn LH, Shenkier TN, Gascoyne RD, Connors JM (2008) Population-based analysis of incidence and outcome of transformed non-Hodgkin's lymphoma. J Clin Oncol 26(32):5165–5169. doi:[10.1200/JCO.2008.16.0283](http://dx.doi.org/10.1200/JCO.2008.16.0283)
- 20. The International Non-Hodgkin's Lymphoma Prognostic Factors Project (1993) A predictive model for aggressive non-Hodgkin's lymphoma. N Engl J Med 329(14):987–994. doi[:10.1056/NEJM1](http://dx.doi.org/10.1056/NEJM199309303291402) [99309303291402](http://dx.doi.org/10.1056/NEJM199309303291402)
- 21. Solal-Celigny P, Roy P, Colombat P, White J, Armitage JO, Arranz-Saez R, Au WY, Bellei M, Brice P, Caballero D, Coiffier B, Conde-Garcia E, Doyen C, Federico M, Fisher RI, Garcia-Conde JF, Guglielmi C, Hagenbeek A, Haioun C, LeBlanc M, Lister AT, Lopez-Guillermo A, McLaughlin P, Milpied N, Morel P, Mounier N, Proctor SJ, Rohatiner A, Smith P, Soubeyran P, Tilly H, Vitolo U, Zinzani PL, Zucca E, Montserrat E (2004) Follicular lymphoma international prognostic index. Blood 104(5):1258–1265. doi:[10.1182/blood-2003-12-4434](http://dx.doi.org/10.1182/blood-2003-12-4434)
- 22. Hoster E, Dreyling M, Klapper W, Gisselbrecht C, van Hoof A, Kluin-Nelemans HC, Pfreundschuh M, Reiser M, Metzner B, Einsele H, Peter N, Jung W, Wormann B, Ludwig WD, Duhrsen U, Eimermacher H, Wandt H, Hasford J, Hiddemann W, Unterhalt M (2008) A new prognostic index (MIPI) for patients with advanced-stage mantle cell lymphoma. Blood 111(2):558–565. doi:[10.1182/blood-2007-06-095331](http://dx.doi.org/10.1182/blood-2007-06-095331)
- 23. Ziebarth JD, Bhattacharya A, Chen A, Cui Y (2012) PolymiRTS Database 2.0: linking polymorphisms in microRNA target sites with human diseases and complex traits. Nucleic Acids Res 40(Database issue):D216–D221. doi:[10.1093/nar/gk](http://dx.doi.org/10.1093/nar/gkr1026) [r1026](http://dx.doi.org/10.1093/nar/gkr1026)
- 24. Ward LD, Kellis M (2012) HaploReg: a resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. Nucleic Acids Res 40(Database issue):D930–D934. doi[:10.1093/nar/gkr917](http://dx.doi.org/10.1093/nar/gkr917)
- 25. Nogai H, Dorken B, Lenz G (2011) Pathogenesis of non-Hodgkin's lymphoma. J Clin Oncol 29(14):1803–1811. doi: [10.1200/JCO.2010.33.3252](http://dx.doi.org/10.1200/JCO.2010.33.3252)
- 26. Muller G, Hopken UE, Lipp M (2003) The impact of CCR7 and CXCR5 on lymphoid organ development and systemic immunity. Immunol Rev 195:117–135
- 27. Saez de Guinoa J, Barrio L, Mellado M, Carrasco YR (2011) CXCL13/CXCR5 signaling enhances BCR-triggered B-cell activation by shaping cell dynamics. Blood 118(6):1560–1569. doi:[10.1182/blood-2011-01-332106](http://dx.doi.org/10.1182/blood-2011-01-332106)
- 28. Widney DP, Gui D, Popoviciu LM, Said JW, Breen EC, Huang X, Kitchen CM, Alcantar JM, Smith JB, Detels R, Martinez-Maza O (2010) Expression and function of the Chemokine, CXCL13, and Its Receptor, CXCR5, in Aids-Associated Non-Hodgkin's Lymphoma. AIDS Res Treat 2010:164586. doi[:10.1155/2010/](http://dx.doi.org/10.1155/2010/164586) [164586](http://dx.doi.org/10.1155/2010/164586)
- 29. Hussain S, Zhu W, Chang SC, Crabb Breen E, Vendrame E, Magpantay L, Widney D, Conn D, Sehl ME, Jacobson L, Bream J, Wolinsky S, Rinaldo C Jr, Ambinder R, Detels R, Zhang ZF, Martinez-Maza O (2012) Serum levels of the chemokine

<span id="page-9-0"></span>CXCL13, genetic variation in CXCL13 and its receptor CXCR5, and HIV-associated non-Hodgkin B cell lymphoma risk. Cancer Epidemiol Biomarkers Prev. doi:[10.1158/1055-9965.EPI-12-1122](http://dx.doi.org/10.1158/1055-9965.EPI-12-1122)

- 30. De Roos AJ, Mirick DK, Edlefsen KL, LaCroix AZ, Kopecky KJ, Madeleine MM, Magpantay L, Martinez-Maza O (2012) Markers of B-cell activation in relation to risk of non-Hodgkin lymphoma.<br>Cancer Res 72(18):4733-4743. doi:10.1158/0008-5472. Cancer Res 72(18):4733–4743. doi[:10.1158/0008-5472.](http://dx.doi.org/10.1158/0008-5472.CAN-12-1639) [CAN-12-1639](http://dx.doi.org/10.1158/0008-5472.CAN-12-1639)
- 31. Burkle A, Niedermeier M, Schmitt-Graff A, Wierda WG, Keating MJ, Burger JA (2007) Overexpression of the CXCR5 chemokine receptor, and its ligand, CXCL13 in B-cell chronic lymphocytic leukemia. Blood 110(9):3316–3325. doi[:10.1182/blood-2007-](http://dx.doi.org/10.1182/blood-2007-05-089409) [05-089409](http://dx.doi.org/10.1182/blood-2007-05-089409)
- 32. Xia Z, Dickens M, Raingeaud J, Davis RJ, Greenberg ME (1995) Opposing effects of ERK and JNK-p38 MAP kinases on apoptosis. Science 270(5240):1326–1331
- 33. Vineis P, Crosignani P, Vigano C, Fontana A, Masala G, Stagnaro E, Miligi L, Costantini AS, Nanni O, Ramazzotti V, Rodella S, Tumino R, Vindigni C (2001) Lymphomas and multiple sclerosis in a multicenter case-control study. Epidemiology 12(1):134–135
- 34. Zhang Y, Holford TR, Leaderer B, Zahm SH, Boyle P, Morton LM, Zhang B, Zou K, Flynn S, Tallini G, Owens PH, Zheng T (2004) Prior medical conditions and medication use and risk of non-Hodgkin lymphoma in Connecticut United States women. Cancer Causes Control: CCC 15(4):419–428. doi[:10.1023/B:CACO.](http://dx.doi.org/10.1023/B:CACO.0000027506.55846.5d) [0000027506.55846.5d](http://dx.doi.org/10.1023/B:CACO.0000027506.55846.5d)
- 35. Ekstrom Smedby K, Vajdic CM, Falster M, Engels EA, Martinez-Maza O, Turner J, Hjalgrim H, Vineis P, Seniori Costantini A, Bracci PM, Holly EA, Willett E, Spinelli JJ, La Vecchia C, Zheng T, Becker N, De Sanjose S, Chiu BC, Dal Maso L, Cocco P, Maynadie

M, Foretova L, Staines A, Brennan P, Davis S, Severson R, Cerhan JR, Breen EC, Birmann B, Grulich AE, Cozen W (2008) Autoimmune disorders and risk of non-Hodgkin lymphoma subtypes: a pooled analysis within the InterLymph Consortium. Blood 111(8):4029–4038. doi[:10.1182/blood-2007-10-119974](http://dx.doi.org/10.1182/blood-2007-10-119974)

- 36. Koshiol J, Lam TK, Gridley G, Check D, Brown LM, Landgren O (2011) Racial differences in chronic immune stimulatory conditions and risk of non-Hodgkin's lymphoma in veterans from the United States. J Clin Oncol 29(4):378–385. doi:[10.1200/JCO.2010.](http://dx.doi.org/10.1200/JCO.2010.30.1515) [30.1515](http://dx.doi.org/10.1200/JCO.2010.30.1515)
- 37. Krumbholz M, Derfuss T, Hohlfeld R, Meinl E (2012) B cells and antibodies in multiple sclerosis pathogenesis and therapy. Nat Rev Neurol 8(11):613–623. doi[:10.1038/nrneurol.2012.203](http://dx.doi.org/10.1038/nrneurol.2012.203)
- 38. Schreiber RD, Old LJ, Smyth MJ (2011) Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. Science 331(6024):1565–1570. doi[:10.1126/science.1203486](http://dx.doi.org/10.1126/science.1203486)
- 39. Sauna ZE, Kimchi-Sarfaty C (2011) Understanding the contribution of synonymous mutations to human disease. Nat Rev Genet 12(10):683–691. doi:[10.1038/nrg3051](http://dx.doi.org/10.1038/nrg3051)
- 40. Katoh M (2003) Identification and characterization of human BCL9L gene and mouse Bcl9 l gene in silico. Int J Mol Med 12(4):643–649
- 41. Willis TG, Zalcberg IR, Coignet LJ, Wlodarska I, Stul M, Jadayel DM, Bastard C, Treleaven JG, Catovsky D, Silva ML, Dyer MJ (1998) Molecular cloning of translocation t(1;14)(q21;q32) defines a novel gene (BCL9) at chromosome 1q21. Blood 91(6):1873–1881
- 42. Brembeck FH, Wiese M, Zatula N, Grigoryan T, Dai Y, Fritzmann J, Birchmeier W (2011) BCL9-2 promotes early stages of intestinal tumor progression. Gastroenterology 141(4):1359–1370, 1370:e1351–e1353. doi[:10.1053/j.gastro.2011.06.039](http://dx.doi.org/10.1053/j.gastro.2011.06.039)