



Prostate cancer aggressive prediction: preponderant diagnostic performances of intravoxel incoherent motion (IVIM) imaging and diffusion kurtosis imaging (DKI) beyond ADC at 3.0 T scanner with gleason score at final pathology

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Abstract

Purpose To explore the preponderant diagnostic performances of IVIM and DKI in predicting the Gleason score (GS) of prostate cancer.

Methods Diffusion-weighted imaging data were postprocessed using monoexponential, IVIM and DK models to quantitate the apparent diffusion coefficient (ADC), molecular diffusion coefficient (D), perfusion-related diffusion coefficient (Dstar), perfusion fraction (F), apparent diffusion for Gaussian distribution (Dapp), and apparent kurtosis coefficient (Kapp). Spearman's rank correlation coefficient was used to explore the relationship between those parameters and the GS, Kruskal–Wallis test, and Mann–Whitney U test were performed to compare the above parameters between the different groups, and a receiver-operating characteristic (ROC) curve was used to analyze the differential diagnosis ability. The interpretation of the results is in view of histopathologic tumor tissue composition.

Results The area under the ROC curves (AUCs) of ADC, F , D , Dapp, and Kapp in differentiating $GS \leq 3 + 4$ and $GS > 3 + 4$ PCa were 0.744 (95% CI 0.581–0.868), 0.726 (95% CI 0.563–0.855), 0.732 (95% CI 0.569–0.860), and 0.752 (95% CI 0.590–0.875), 0.766 (95% CI 0.606–0.885), respectively, and those in differentiating $GS \leq 7$ and $GS > 7$ PCa were 0.755 (95% CI 0.594–0.877), 0.734 (95% CI 0.571–0.861), 0.724 (95% CI 0.560–0.853), and 0.716 (95% CI 0.552–0.847), 0.828 (95% CI 0.676–0.929), respectively. All the P values were less than 0.05. There was no significant difference in the AUC for the detection of different GS groups by using those parameters.

Conclusion Both the IVIM and DKI models are beneficial to predict GS of PCa and indirectly predict its aggressiveness, and they have a comparable diagnostic performance with each other as well as ADC.

Keywords Prostate cancer · Diffusion-weighted imaging · Diffusion kurtosis imaging · Magnetic resonance imaging · Intravoxel incoherent motion · Gleason score

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Introduction

Prostate cancer (PCa) is a significant health issue affecting predominantly elderly men worldwide. Its incidence has been ranked high for many years in the global cancer survey, and its mortality rate is second to lung cancer [1–3]. The Gleason scoring system is the most widely used scoring system for judging the malignant degree of PCa; the higher the GS, the higher the malignancy and the corresponding invasiveness [4]. For low-risk tumors (GS < 7), no immediate treatment is required, that is, either watchful waiting or active surveillance; for intermediate-risk (GS = 7), monotherapy is offered, and for high-risk prostate cancer (GS > 7), combination therapy will be the best treatment option [5]. Recently, much greater attention is given to the intermediate risk group to be subdivided into GS = 3 + 4 and GS = 4 + 3 for subanalysis due to prognostic differences between the two groups [4, 6]. Kamel et al. [7] showed that PCa with GS = 4 + 3 was more prone to metastasis than GS = 3 + 4, the probability was about 2.8%, 0.9%, and the overall survival rate was 23% lower than the latter. Recent studies have shown that GS = 3 + 4 tumors have high biological inertia and good prognosis, and active monitoring is recommended to avoid overtreatment [8]. Transrectal ultrasound-guided prostate biopsy (TRUS-biopsy) can cause side-effects including bleeding, pain, and infection, and it is less sensitive than what we expected [9, 10].

Diffusion-weighted magnetic resonance imaging (DW-MRI) offers a noninvasive visualization approach that reflects the diffusion characteristics of water molecules in biological tissues and indirectly reflects the microscopic changes in tissue structures, which characterize the organization. It is an informative MRI modality in detecting PCa, and it shows moderately high diagnostic accuracy [11]. Routinely, in our clinical work, apparent diffusion coefficient (ADC) values are calculated by means of a monoexponential model via assumption of the diffusion in deference to Gaussian distribution similar to that in pure water. Nevertheless, these movements in biological tissue include molecular diffusion of water and blood microcirculations in a network of capillaries (perfusion). The microcirculation or perfusion of blood can also be considered an incoherent movement due to the pseudorandom tissue of the capillary network at the voxel level. In a significant development, Le Bihan [12] established an *in vivo* biexponential model, which is also known as the intravoxel incoherent motion (IVIM) model. This model correlates the molecular diffusion coefficient and perfusion. A study by Hiroshi Shinmoto showed that the molecular diffusion coefficient and perfusion fraction in prostate cancer were significantly lower than those found in the peripheral zone

(PZ) [13]. Liu et al. found that IVIM could potentially improve the differentiation of prostate cancer in the central gland and offer better accuracy than ADC for differentiating stromal hyperplasia and prostate cancer [14]. Furthermore, some studies concluded that perfusion-free diffusion parameter D performed better in differentiating the GS of PCa [15–18]. When higher b values are added, the perfusion is depressed, and the molecular diffusion was proven to depart from the conventional random diffusion process due to the existence of barriers within cellular complex environments, which is acknowledged as non-Gaussian diffusion behavior. This calls for more advanced modeling of DWI to characterize non-Gaussian behavior—the idea of reflecting organizational heterogeneity and irregularity—detected using high b values. The DKI model allows for the estimation of kurtosis, and higher kurtosis values indicate a more peaked, non-Gaussian distribution of diffusion [19]. Previous studies have shown that the DKI model improves PCa detection and diagnosis [20–24]. Wang et al. reported that the 90th Kapp exhibited better diagnostic performance in differentiating the GS of PCa [23]; Wu's team reported that DKI may help in predicting GS upgrade in biopsy-proven GS 6 prostate cancer [24]. A recent study by Tamada et al. [25] reported that Kapp performed well in differentiating GS $\leq 3 + 3$ and GS $\geq 3 + 4$ tumors, GS $\leq 3 + 4$ and GS $\geq 4 + 3$ tumors, which were similar to the diagnostic performance of ADC; and ADC and Kapp were highly correlated.

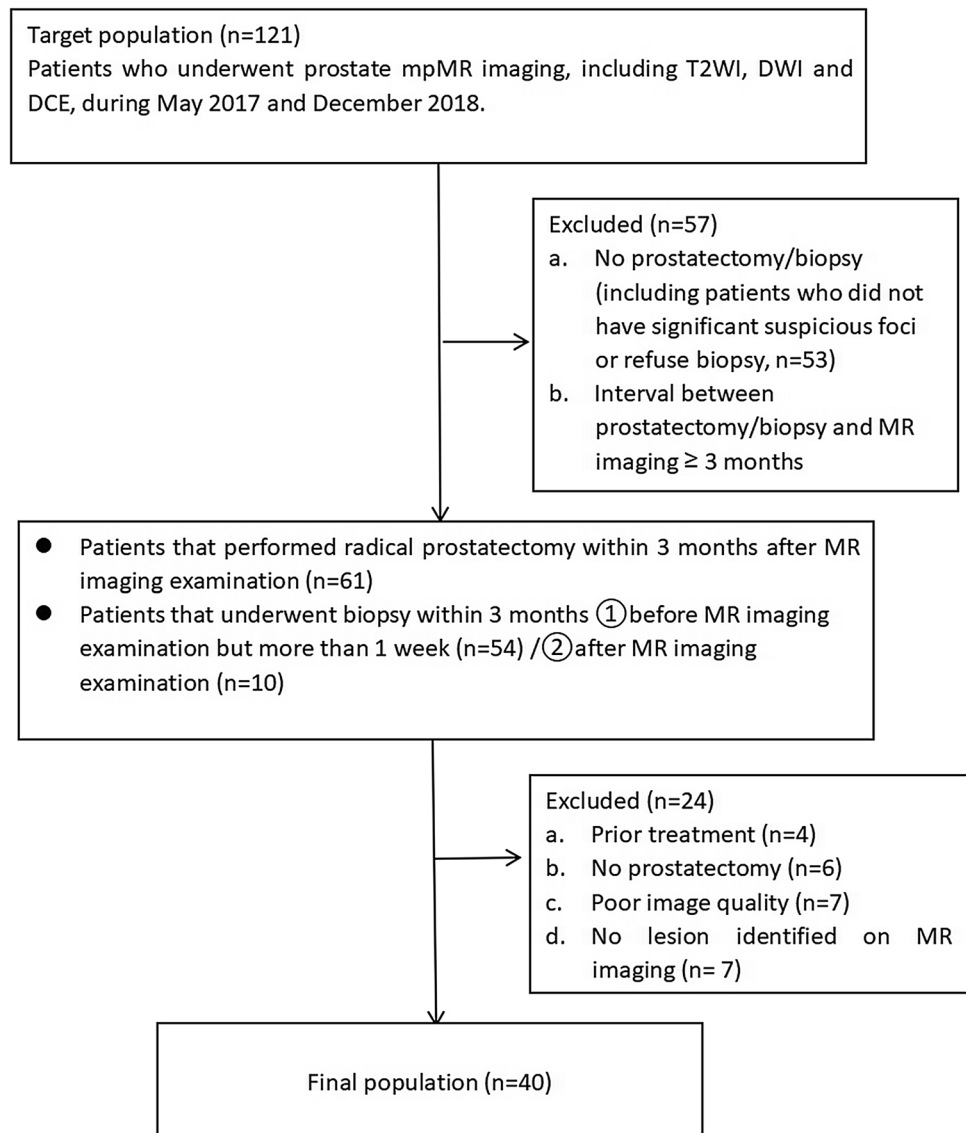
Although much work as we mentioned above had been done, studies on these two models are relatively deficient. And the GS results used in many studies were obtained from biopsies which can be inaccurate due to sampling error considering the fact that the GS is upgraded in every third patient following radical prostatectomy (RP) [26]. The GSs used in this study were obtained from RP. We aimed to explore the preponderant diagnostic performances of these two models in predicting the aggression of PCa; and what their unique parameters added to monoexponential model (ADC).

Materials and methods

Patient population

From May 2017 to December 2018, 121 consecutive patients were enrolled as a part of an ongoing prospective study. All the patients underwent diffusion-weighted MR scanning and gave informed consent. Our target population was people who exhibited remarkable findings in serum prostate-specific antigen (PSA) test and/or digital rectal examination (DRE) and/ultrasonography. Patients who had previously undergone ultrasound-guided transrectal biopsy were also

Fig. 1 Flowchart of patient population. PZ peripheral zone, TZ transitional zone, CZ central zone



included because our primary objective was to detect and characterize clinically significant cancer in the gland [27]. Recently, Jung et al. showed that postbiopsy hemorrhage did not negatively affect the detection of tumors with $GS \geq 3 + 4$ or with volume ≤ 0.5 ml [28]. Through image analysis, we observed that only 11 (28%) cases had hemorrhage, and the signal of hemorrhagic foci was depressed well on high b value (e.g., $b = 2200$) DW images. Eighty-one patients were excluded for the following reasons: (a) those who exhibited neither prostatectomy nor biopsy pathological proof ($n = 53$), including patients who did not have significant suspicious foci on all of mpMRI or refuse biopsy; (b) those in whom the interval between prostatectomy/biopsy was more than 3 months ($n = 4$); (c) those who had prior treatment ($n = 4$), such as endocrine therapy and transurethral resection of carcinoma of the prostate (TURCaP); (d) those with no prostatectomy ($n = 6$); (e) cases with poor image quality ($n = 7$);

and (f) those with no lesion being identified on MR imaging ($n = 7$). Finally, we considered a total number of 43 patients for this study. Figure 1 presents a flowchart of the population. All the GS scores were evaluated using radical prostatectomy gross specimens. The clinical data of the 40 patients are summarized in Table 1. In view of lacking $GS = 6$ and good prognosis of $GS = 3 + 4$, patients in our study were divided into three risk groups as $GS \leq 3 + 4$ (group A, GA), $GS = 4 + 3$ (GB), and $GS > 4 + 3$ (GC).

MR imaging protocol

Multiparametric MR imaging was performed using a 3.0-T MR imager (Discovery MR 750, GE Medical Systems, Milwaukee, WI, USA) and a 32-channel phased-array surface coil without an endorectal coil. The contraindications for enhanced magnetic resonance imaging had been

Table 1 Clinical data of the 40 patients

| Characteristic | Value |
|---------------------------------|--------------------------------|
| Patient age (year) ^a | 70 ± 7 (57–85) |
| PSA lever (ng/ml) ^a | 28.87 ± 26.02 (4.36–102.10) |
| Number of each GS | |
| 3 + 3 | 2 |
| 3 + 4 | 11 |
| 4 + 3 | 11 |
| 3 + 5 | 1 |
| 4 + 4 | 2 |
| 4 + 5 | 9 |
| 5 + 4 | 2 |
| 5 + 5 | 2 |
| Pathologic staging | |
| <T2 | 5 |
| T3a | 24 |
| T3b | 7 |
| T4 | 4 |

^aMean ± standard deviation (minimum–maximum)

excluded, and particular preparations such as gastrointestinal preparation were not highly noted in this study considering that there was no consensus regarding patient preparation issues [27]. Propeller FS T2-weighted MR imaging was used to reduce motion artifact. The inclination angle of the axial-oblique scanning was adjusted according to the inclination degree of the prostate. Echo-planar DW images were acquired in the axial-oblique plane that was

consistent with T2 W imaging using a single-shot spin-echo echo-planar sequence. Eleven b values of 0, 50, 100, 200, 900, 1100, 1400, 1800, 2200, 2500, and 3000 s/mm² (with number of averages of 1, 1, 1, 1, 4, 4, 6, 8, 10, 10, and 12, respectively) were determined. ADC maps were calculated automatically via monoexponential fitting per voxel of the DW images. 3D T1 liver acquisition with volume acceleration flex (LAVA FLEX) sequence was used for DCE-MR imaging. DCE was only used for the facilitation of diagnosis in this study. The detailed parameters of these main acquisition sequences are shown in Table 2.

IVIM and DKI models

IVIM model and its parameters of D , D_{star} , and F are fit for a biexponential equation:

$$S_b/S_0 = (1 - F) \cdot \exp(-b \cdot D) + F \cdot \exp(-b \cdot D_{\text{star}}), \quad (1)$$

where D characterizes extravascular diffusion of water, while D_{star} represents signal changes attributing to the intravascular movement of water. F is the perfusion fraction. S_b is the DWI signal intensity at a specified b value, and S_0 is the baseline signal at $b = 0$.

The DKI model is based on the following equation:

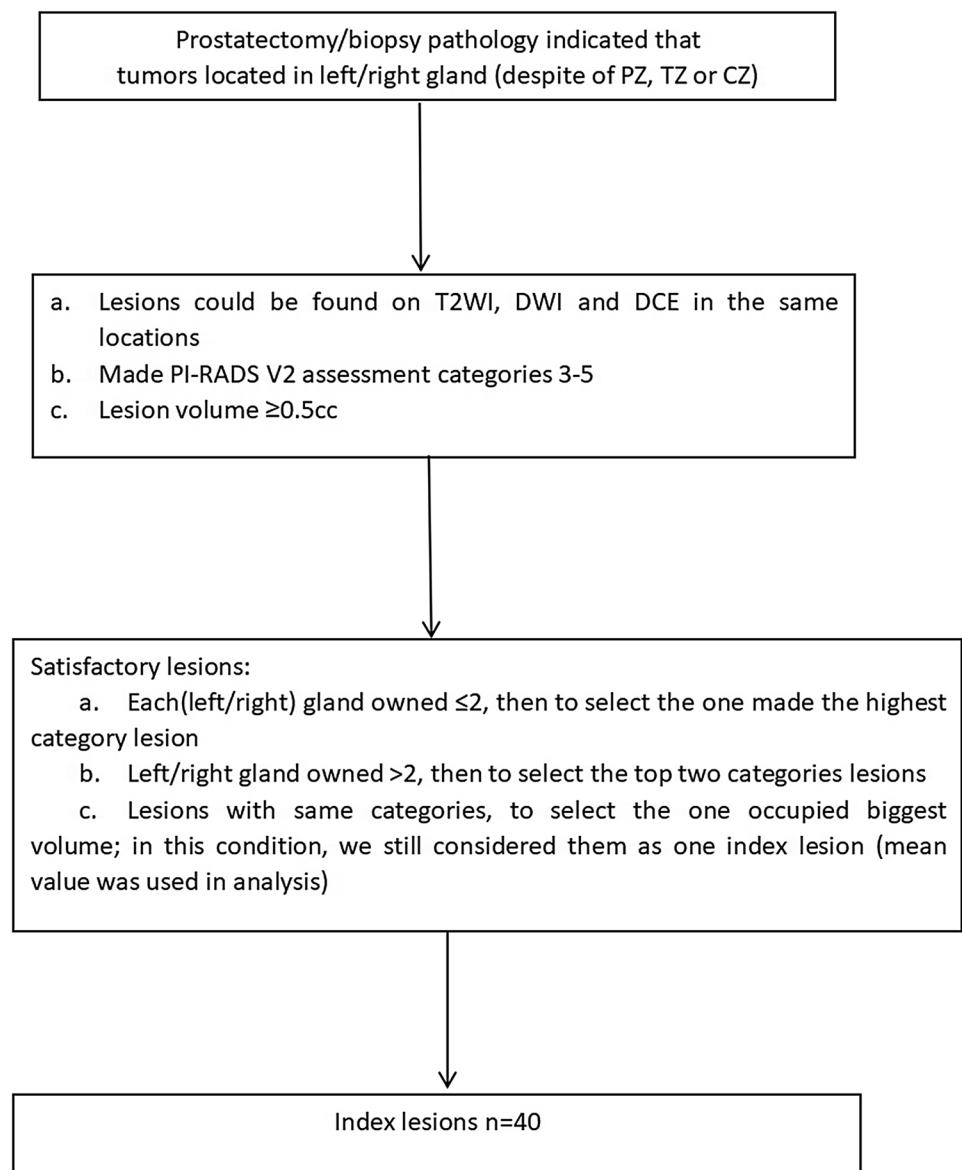
$$S_b/S_0 = \exp(-b \cdot D_{\text{app}} + b^2 \cdot D_{\text{app}} \cdot K_{\text{app}}/6). \quad (2)$$

In Eq. [2], S_b and S_0 have the same meaning as in Eq. [1]. When S_0 is known, D_{app} and K_{app} are obtained. The parameter K_{app} represents the apparent diffusional kurtosis (unitless), and D_{app} is the diffusion coefficient that is corrected to account for the observed non-Gaussian behavior [29].

Table 2 Acquisition parameter values of major Sequences

| Parameter | Multiplanar propeller fs T2WI | | | DWI | DCE (3D) 50 phase |
|-------------------------------|-------------------------------|------------------|------------------|---------------------|--------------------|
| | Axial | Sagittal | Coronal | | |
| Repetition time (ms) | 7972 | 7486 | 6432 | 2000 | 3.6 |
| Echo time (ms) | 93 | 106 | 95 | Minimum | Min full |
| Matrix (frequency × phase) | 320 × 320 | 320 × 320 | 320 × 320 | 128 × 160 | 160 × 128 |
| Echo train length | 28 | 32 | 24 | / | / |
| Band width (KHz) | 62.5 | 62.5 | 62.5 | 250 | 166.67 |
| No. of acquired signals | 3.0 | 2.5 | 2.0 | b value-dependent | 1 |
| Field of view (cm) | 20 × 20 | 22 × 22 | 22 × 22 | 28 × 25.2 | 28 × 28 |
| Section thickness (mm) | 3.0 | 3.0 | 3.0 | 3.0 | 4.0 (ZIP2) |
| Gap (mm) | 0.5 | 0.5 | 0.5 | 0.5 | 0 |
| Acquisition voxel size | 0.6 × 0.6 × 3 mm | 0.7 × 0.7 × 3 mm | 0.7 × 0.7 × 3 mm | 2.2 × 1.6 × 3 mm | 1.8 × 2.2 × 4 mm |
| Reconstruction voxel size | / | / | / | / | 0.55 × 0.55 × 2 mm |
| Orientation of phase encoding | A/P | A/P | R/L | A/P | A/P |
| Acquisition time | 03:54 | 3:30 | 03:34 | 05:48 | 04:43 |

fs fat suppression

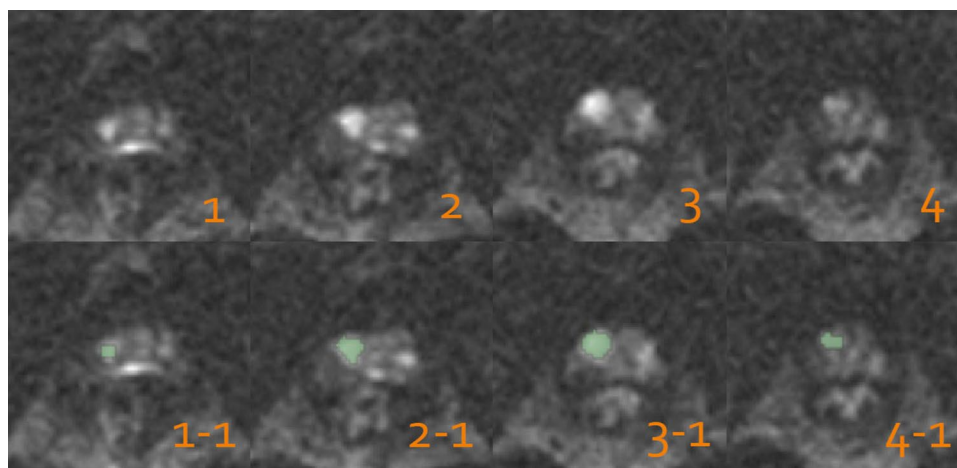
Fig. 2 Flowchart of index lesion

ROI analysis

Two experienced radiologists (Professor A with 3 years of experience in prostate MRI, and Professor B with 4 years of experience in prostate MRI) identified suspicious tumors in consensus according to the criteria in Prostate Imaging-Reporting and Data System, Version 2 [27]. These radiologists had not been previously informed of the pathological results. Usually, a patient having more than one suspicious focus as well as the prostatic tumor had usually multiple foci separated by noncancerous tissue. Index lesion of each patient was evaluated in this study. An index lesion is one that locates in the zone which is depicted in prostatectomy/biopsy pathologic result and can be found on MRI. The method for index lesion definition is presented in Fig. 2. The two radiologists depicted every

region of interest (ROI) separately on high b ($b = 2200$ s/mm²) DWI with reference to the ADC imaging which was generated automatically after scanning, using the IMAge/enGINE MR_Diffusion software (V2.0.3, Vusion Tech, Hefei, China, <http://www.vusion.com.cn>) to perform each DW-MR imaging, obtaining parameters of the IVIM (F , D , and D_{star}) and DKI models (Dapp, Kapp) [30]. Their mean values were used for data analysis. The three-dimensional ROI data measurement capability of this version offered more convenient measurement and more comprehensive use of the diffusion information of lesions. The placement of 3D-ROIs was in accordance with the index lesion, avoiding the urethral and ejaculatory ducts, as well as hemorrhage. Figure 3 shows an example of manual ROI placement.

Fig. 3 ROIs being signed as green by postprocessing software on DWI when $b=2200$



Statistical analysis

Data analysis was conducted using the SPSS software (version 20.0; SPSS, Chicago, USA) and the MedCalc Statistical Software (version 15.8; MedCalc Software bvba, Ostend, Belgium; <https://www.medcalc.org>; 2015). The interobserver agreement for each parameter measurement was assessed by calculating the interclass correlation coefficient (<0.40, poor; 0.40–0.59, fair; 0.60–0.74, good; and 0.751–1.00, excellent) [31]. The mean values of those parameters measured by the two radiologists were used in the following data analysis. Shapiro–Wilk test of normality was performed to assess the normality of each parameter at P value >0.05. Spearman’s rank correlation coefficient (0.0–0.2, very weak to negligible; 0.2–0.4, weak; 0.4–0.7, moderate; 0.7–0.9, strong; 0.9–1.0, very strong) [32] was used to crystallize the correlation between each parameter and GS. Correlations of ADC and the unique parameters of IVIM and DKI models were also computed. The Kruskal–Wallis one-way analysis of variance (ANOVA) (k samples) and Mann–Whitney U test were used to analyze the differences of each parameter between different groups. The ROC curves were employed to analyze the diagnostic performance for predicting GS of PCa. Areas under the curves (AUCs) were compared using the DeLong method [33]; and 95% confidence intervals (CIs), optimal cutoff values, and the corresponding sensitivity and specificity values were calculated. A two-sided significance level of 0.05 was set for the above statistical tests.

Results

These 40 index lesions consisted of 4 PI-RADS category 3, 24 PI-RADS category 4, and 23 PI-RADS category 5 foci. The agreements for these metrics between the two readers were excellent for ADC (interclass correlation

coefficient (ICC): 0.95; 95% CI 0.90–0.97), D (ICC: 0.96; 95% CI 0.92–0.98), D_{star} (ICC: 0.94; 95% CI 0.90–0.97), F (ICC: 0.97; 95% CI 0.95–0.99), D_{app} (ICC: 0.97; 95% CI 0.94–0.98), and K_{app} (ICC: 0.94; 95% CI 0.89–0.97).

GS was moderately inversely correlated with ADC ($\rho = -0.487$, $P < 0.01$), F ($\rho = -0.473$, $P < 0.01$), D ($\rho = -0.432$, $P < 0.01$) and D_{app} ($\rho = -0.436$, $P < 0.01$), and positively associated with K_{app} ($\rho = 0.611$, $P < 0.01$); GS showed no significant correlation with D_{star} ($\rho = 0.255$, $P = 0.11$). The differences in ADC, F , D , D_{app} , and K_{app} values between GC and GA, GC and GA + GB, GA and GB + GC were all significant ($P < 0.05$) and were all not significant between GA and GB, and GB and GC. Details are presented in Table 3. The distribution of each parameter’s values according to different GS groups are shown in Fig. 4. ADC exhibited a strong positive correlation with F ($\rho = 0.785$; $P < 0.001$), and a strong negative association with K_{app} ($\rho = -0.849$, $P < 0.001$).

Figure 5 and Table 4 display the results of the ROC curve analysis of the diffusion metrics for distinguishing different GS PCa values. The AUCs of ADC, F , D , D_{app} , and K_{app} in differentiating $GS \leq 3 + 4$ and $GS > 3 + 4$ PCa were 0.744

Table 3 Nonparametric tests results of diffusion parameters between different GS group

| Sample 1–Sample 2 | ADC | F | D | D_{app} | K_{app} |
|-------------------------|-------|-------|-------|------------------|------------------|
| GC–GB ^a | 0.364 | 0.452 | 0.620 | 0.918 | 0.054 |
| GC–GA ^a | 0.012 | 0.021 | 0.023 | 0.018 | 0.002 |
| GB–GA ^a | 0.751 | 0.844 | 0.663 | 0.381 | 1.000 |
| GC–GA + GB ^b | 0.008 | 0.013 | 0.018 | 0.022 | 0.001 |
| GA–GB + GC | 0.014 | 0.022 | 0.019 | 0.011 | 0.007 |

GA: $GS \leq 3 + 4$, GB: $GS = 4 + 3$, GC: $GS > 7$, GA + GB: $GS \leq 7$, GB + GC: $GS \geq 4 + 3$

^aData shown are adjusted significance of each parameter in pairwise comparisons (k samples)

^b P values assessed by Mann–Whitney test

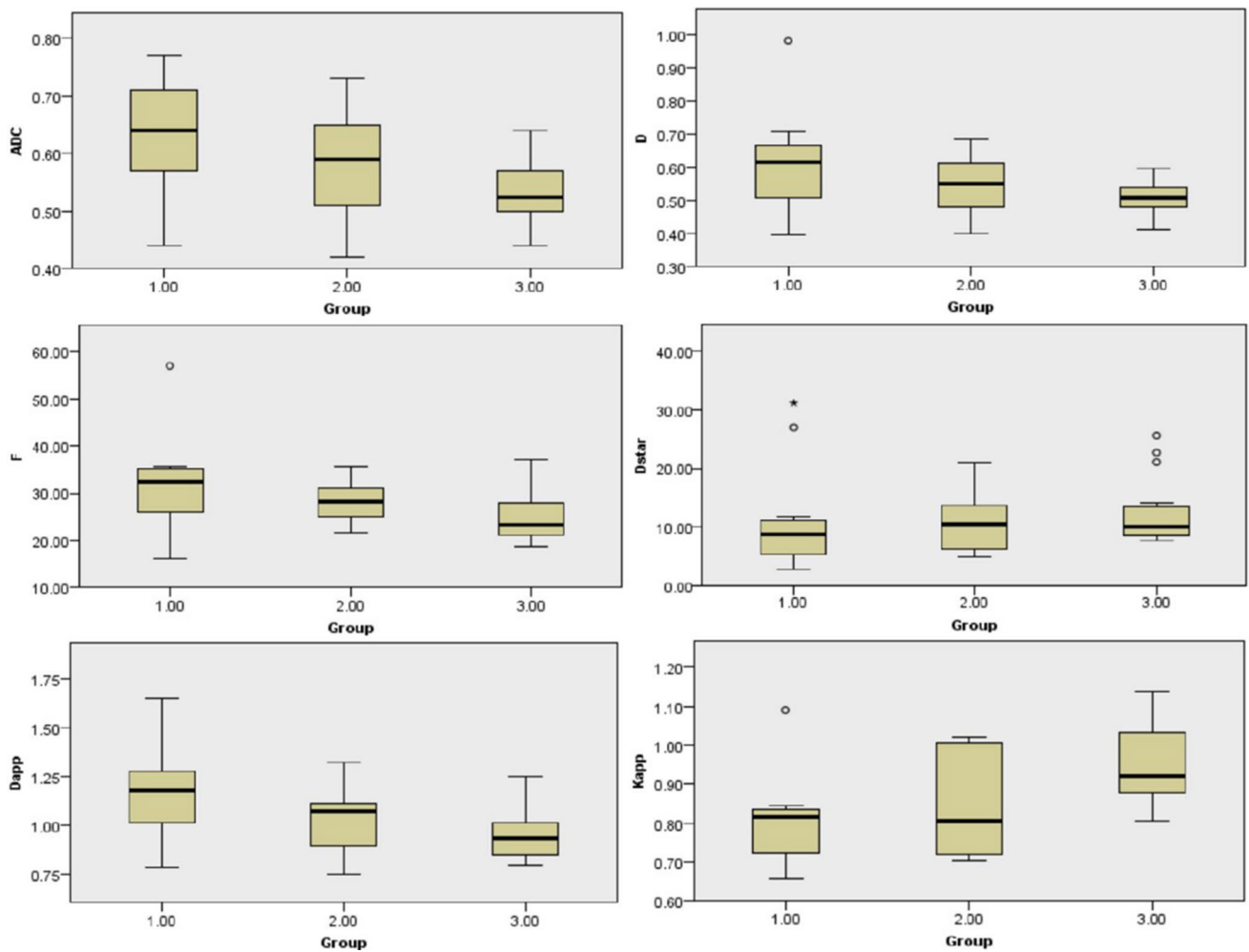


Fig. 4 Boxplots above showing the results of Kruskal–Wallis test of parameters for independent samples among group1 ($GS \leq 3+4$), group2 ($GS = 4+3$), and group3 ($GS > 7$). Center line indicates median, top of box indicates the 75th percentile, bottom of box indicates the 25th percentile, whiskers indicate the 10th and 90th per-

centiles, asterisk indicates extreme values (more than 3 interquartile ranges), and circles indicate outliers (between 1.5 and 3 interquartile ranges). ADC, *F*, *D*, and *Dapp* display a decreasing trend with GS, while *Dstar* and *Kapp* display an increasing trend with GS

Fig. 5 Graph showing utility of ROC curves of ADC, *F*, *D*, *Dapp*, and *Kapp* to differentiate $GS \leq 3+4$ and $GS > 3+4$ Pca. Graph b shows utility of ROC curve of those parameters to differentiate $GS \leq 7$ and $GS > 7$. Gray line = chance diagonal

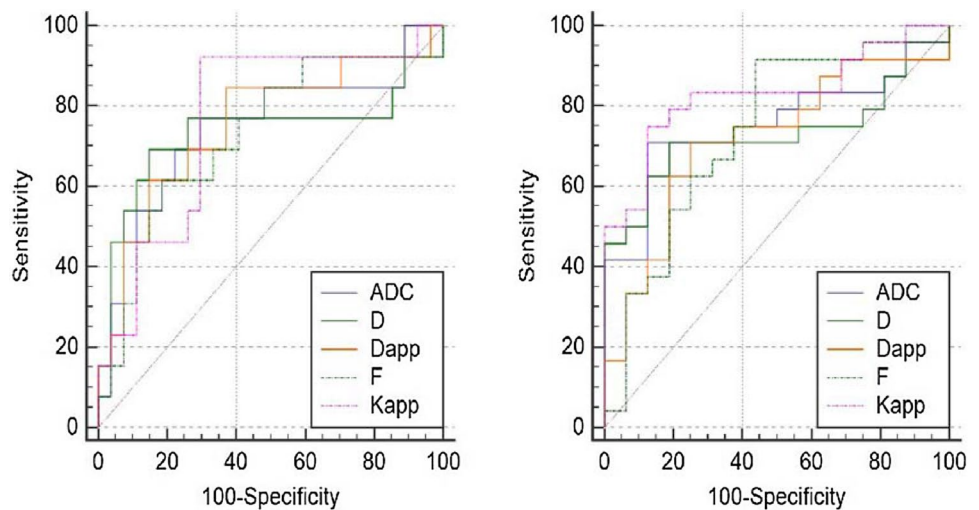


Table 4 Diagnostic test characteristics of diffusion parameters for the diagnosis of GS

| Measurement | AUC (95% CI) | <i>P</i> | Sensitivity (%) | Specificity (%) | Cutoff value | Youden index <i>J</i> |
|-----------------|---------------------|-----------|-----------------|-----------------|---|-----------------------|
| ADC | | | | | | |
| GA–GC+GB | 0.744 (0.581–0.868) | 0.010 | 76.92 | 70.37 | $> 0.59 \times 10^{-3} \text{mm}^2/\text{s}$ | 0.47 |
| GC–GA+GB | 0.755 (0.594–0.877) | 0.001 | 70.83 | 87.50 | $\leq 0.59 \times 10^{-3} \text{mm}^2/\text{s}$ | 0.58 |
| <i>F</i> | | | | | | |
| GA–GC+GB | 0.726 (0.563–0.855) | 0.014 | 61.54 | 81.48 | $> 31.04\%$ | 0.43 |
| GC–GA+GB | 0.734 (0.571–0.861) | 0.006 | 91.67 | 56.25 | $\leq 23.51\%$ | 0.48 |
| <i>D</i> | | | | | | |
| GA–GC+GB | 0.732 (0.569–0.860) | 0.032 | 69.23 | 85.19 | $> 0.59 \times 10^{-3} \text{mm}^2/\text{s}$ | 0.54 |
| GC–GA+GB | 0.724 (0.560–0.853) | 0.007 | 70.83 | 81.25 | $\leq 0.54 \times 10^{-3} \text{mm}^2/\text{s}$ | 0.52 |
| Dapp | | | | | | |
| GA–GC+GB | 0.752 (0.590–0.875) | 0.006 | 84.62 | 62.96 | $> 0.98 \times 10^{-3} \text{mm}^2/\text{s}$ | 0.48 |
| GC–GA+GB | 0.716 (0.552–0.847) | 0.010 | 70.83 | 75.00 | $\leq 0.98 \times 10^{-3} \text{mm}^2/\text{s}$ | 0.46 |
| Kapp | | | | | | |
| GA–GC+GB | 0.766 (0.606–0.885) | 0.002 | 92.31 | 70.37 | ≤ 0.84 | 0.63 |
| GC–GA+GB | 0.828 (0.676–0.929) | < 0.001 | 75.00 | 87.50 | > 0.84 | 0.63 |

GA: GS $\leq 3+4$, GB: GS = 4+3, GC: GS > 7 , GA+GB: GS ≤ 7 , GB+GC: GS $\geq 4+3$

AUC Area under the curve, 95% CI 95% confidence interval

(95% CI 0.581–0.868), 0.726 (95% CI 0.563–0.855), 0.732 (95% CI 0.569–0.860), and 0.752 (95% CI 0.590–0.875), 0.766 (95% CI 0.606–0.885), respectively, and those in differentiating GS ≤ 7 and GS > 7 PCa were 0.755 (95% CI 0.594–0.877), 0.734 (95% CI 0.571–0.861), 0.724 (95% CI 0.560–0.853), and 0.716 (95% CI 0.552–0.847), 0.828 (95% CI 0.676–0.929), respectively, with all the *P* values less than 0.05. For pairwise comparisons of ROC curves, there were no significant differences among ADC, *F*, *D*, Dapp, and Kapp in differentiating different GS group ($P=0.0501$ –0.9414). Figures 6 and 7 display representative patients and diffusion parameter maps.

Discussion

Our study findings demonstrated that altered IVIM (*F* and *D*) and DKI parameters (Dapp and Kapp) in different GS PCa, revealed good diagnostic performance in differentiating GS $\leq 3+4$ and GS $> 3+4$ PCa, GS ≤ 7 and GS > 7 PCa. We could interpret our findings in view of histopathologic tumor tissue composition. The increasing Gleason pattern is attributed to the increased heterogeneity of prostate histological compartments which consist of vascular (i.e., capillaries), fibromuscular stroma, epithelium, and glandular lumen, correlating with tumor aggressiveness [34, 35]. Recently, Chatterjee et al. [36] found that Gleason patterns exhibited a strong positive correlation with the epithelium and a negative correlation with the stroma and lumen space, but no remarkable correlation with cellularity metrics. But no parameter was able to differentiate GS $\leq 3+4$ and

GS = 4+3, and this might indicate that these two GS tumors' microstructures had no significant differences, and it also could be attributed to small samples. Similar to conventional ADC, *D*, and Dapp are the adjusted diffusion coefficients, respectively, for IVIM and DKI. A number of studies have described the relationship between ADC and GS [37–41], and they have almost consistently reported a negative correlation. An increase of diffusion-restricting ingredients (i.e., vascular, epithelial fractions) associated with loss of diffusion-promoting components (i.e., stromal, luminal space) in tumors [42], leads to the decline of values of these diffusion parameters.

Although previous studies have proven the influence of tissue perfusion on ADC [43], the nature of the biexponential model has not yet been well explained. A report of Kuru et al. in 2014 [15] also indicated that perfusion-free diffusion constant *D* might hold potential for improved image-based tumor grading, which was consistent with our findings. It has been reported that the *D*star was at least one magnitude greater than *D*, and perfusion may be only palpable at very low *b* values [44]. Low *b* values were proposed in precluding high *b* values for IVIM to avoid the interference by high *b* values, where the contribution due to non-Gaussian diffusion was appreciable. However, in our study and other studies with a high *b* value, Le Bihan [45] suggested that the slow diffusion component may represent water that is associated with cell membranes and with cytoskeleton structures, while the fast diffusion component represents the remaining, less-restricted water, which is found in both intra- and extracellular spaces. A study with a larger patient population (50 patients) concluded that *b* value distribution influences

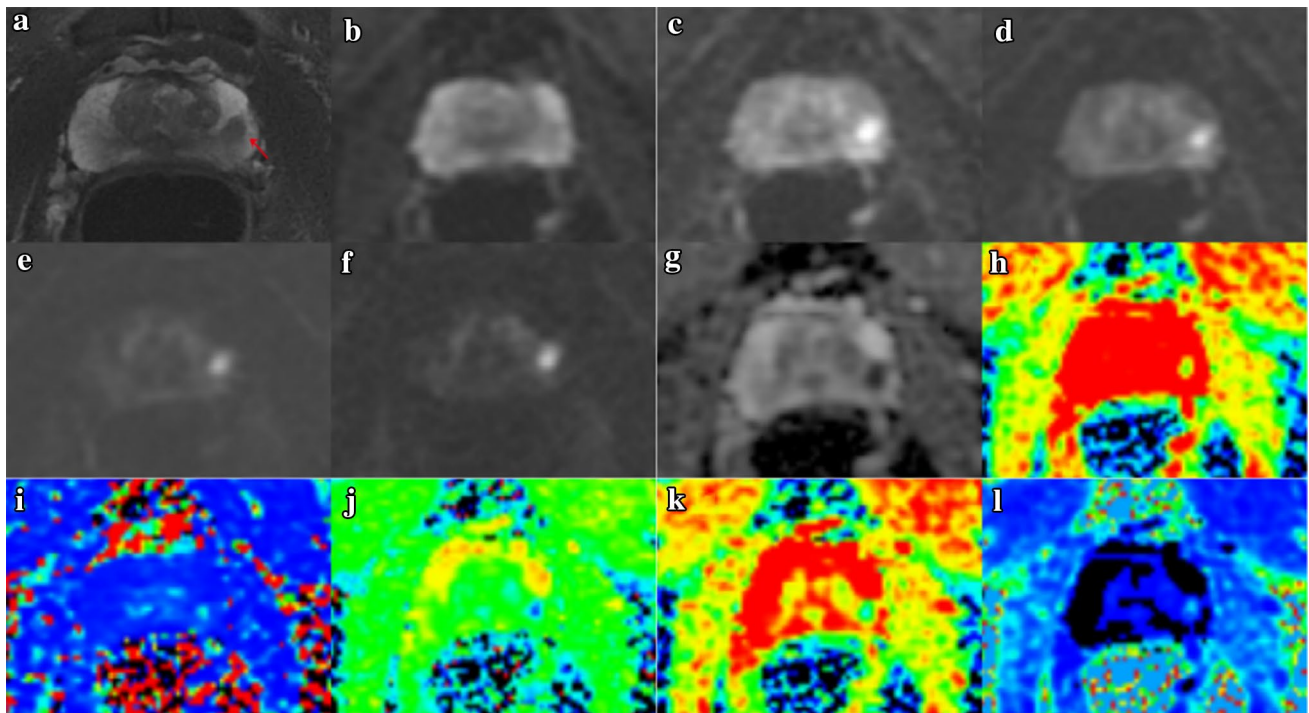


Fig. 6 72-year-old man with prostate cancer (GS 3+4=7, lesions in left lobe of prostate, <T2, PSA 4.4 ng/ml). Pictures above show the index lesion (0.7 cm) in left PZ, PI-RADS V2 category 4. **a** Lesion is indicated by an arrow on T2WI; **b–f** images obtained with *b* values of 200, 900, 1100, 2200, and 3000 s/mm²; as the *b* value

increases, the high signal of the normal tissue is gradually suppressed, whereas the tumors become more and more obvious; **g** ADC map processed by monoexponential model; **h, i** pseudo color maps of *D* ($=0.67 \times 10^{-3} \text{mm}^2/\text{s}$), *F* ($=35.76\%$), *Dstar* ($=5.97 \times 10^{-3} \text{mm}^2/\text{s}$), *Dapp* ($=1.30 \times 10^{-3} \text{mm}^2/\text{s}$), *Kapp* ($=0.72$)

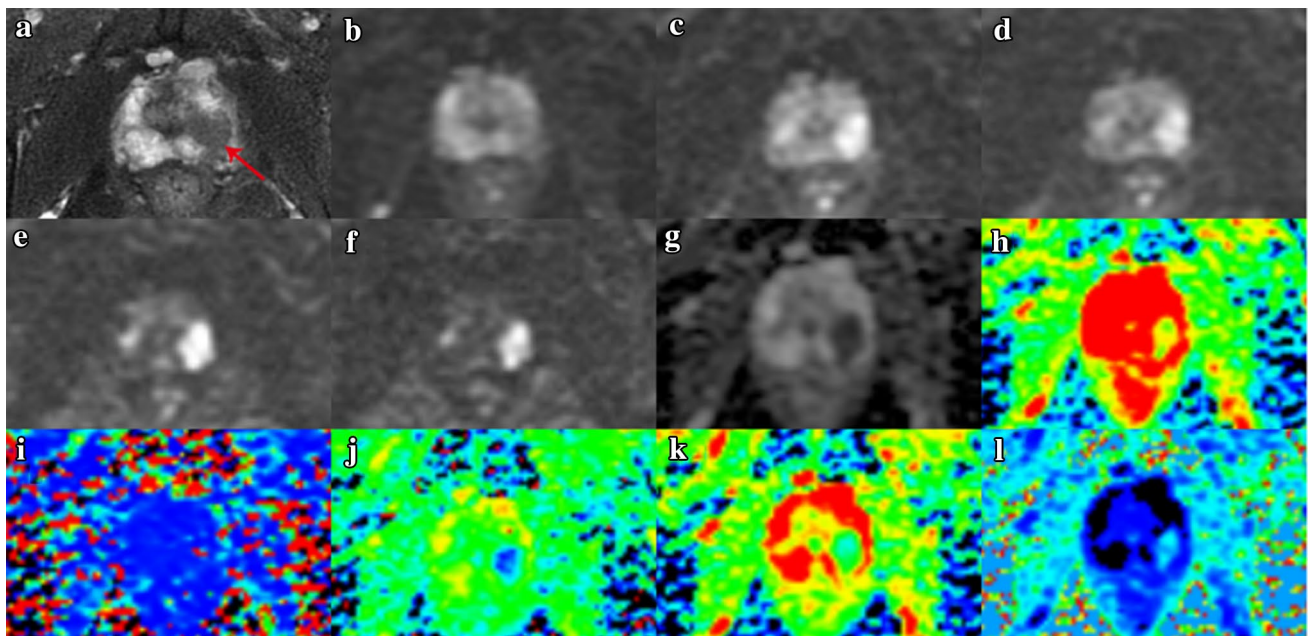


Fig. 7 70-year-old man with prostate cancer (GS 4+5=9, lesions in both lobes of prostate, T3a, PSA 7.9 ng/ml). Pictures above show the index lesion (1.8 cm) in left PZ, PI-RADS V2 category 5. **a** Lesion is indicated by an arrow on T2WI; **b–f** images obtained with *b* values of 200, 900, 1100, 2200, and 3000 s/mm²; as the *b* value

increases, the high signal of the normal tissue is gradually suppressed, whereas the tumors become more and more obvious; **g** ADC map processed by monoexponential model; **h, i** pseudo color maps of *D* ($=0.50 \times 10^{-3} \text{mm}^2/\text{s}$), *F* ($=25.45\%$), *Dstar* ($=8.40 \times 10^{-3} \text{mm}^2/\text{s}$), *Dapp* ($=0.94 \times 10^{-3} \text{mm}^2/\text{s}$), *Kapp* ($=0.94$)

mainly the repeatability of DWI-derived parameters (including IVIM and DKI parameters) rather than the diagnostic performance [46]. In the present study, the measurement of the relevant parameter *D*star indicated remarkably large standard deviations of most cancer lesions, which was similar to previous studies [47, 48], and we all found negative result of *D*star in predicting GS; but what differentiated from them to our study was that *D* and *F* performed well in differentiating different GS groups. A reason might be their different group ($GS = 6$ and $GS \geq 7$). The *F* value can be calculated by assuming the random direction of the capillary segment at the voxel level [12]. A relatively purer IVIM parameter investigation by Pang et al. [44], which used different combinations of five *b* values (0, 188, 375, 563, and 750 s/mm²), reported a significant increase in *F* in tumors compared to benign tissues with *b* values below 750 s/mm², and when high *b* values were employed, *F* might become lower or indistinguishable. However, even for low *b* values, they did not observe a significant difference in *F* among different GS tumors. Some previous studies reported that *F* was significantly smaller in PCa than in healthy PZ [13, 48], and in our study *F* was found to negatively correlated with GS. This may be interpreted by a theory of bulky phenomenon [49], where *F* is not only specific to perfusion but also may be sensitive to glandular secretion and fluid flow in the prostatic ducts, which corresponded to the results obtained by Le Bihan, as stated above.

Previous studies showed that kurtosis had significant correlations with histopathologic parameters (cytoplasmic, cellular, and stromal fractions) [50, 51]. ADCs obtained with *b* values less than 1000 s/mm² were thought to mainly reflect diffusion of water in the extracellular space; when the *b* value increases to more than 1000 s/mm², the intracellular interaction promotes non-Gaussian diffusion behavior and increases kurtosis, and the kurtosis parameter was supposed to reflect the interaction of water molecules with cell membranes and intracellular components [50, 52]. Therefore, Kapp has an excellent diagnostic ability for high GS lesions, which is proven by our results (AUC = 0.828, $P < 0.001$). Similar results had been concluded in a recent study [53]. A recent study by Lawrence et al. [51] showed that Dapp exhibited a significant positive correlation with luminal space and a negative correlation with cellularity, which assisted in differentiating cancerous lesions from normal tissue. However, they found that only the median Kapp was significantly different between groups with $GS \geq 4 + 3$ and $\leq 3 + 4$ ($P < 0.05$). Being different from them, in our study, mean values were used for analysis, and we found Dapp could also assist to differentiate $GS \geq 4 + 3$ and $\leq 3 + 4$. In another recent study, Wu et al. [24] reported that both Kapp and Dapp helped in the prediction of GS upgrade in biopsy-proven GS 6 prostate cancer.

Although there was no significant difference in the AUCs among ADC, *F*, *D*, Dapp, and Kapp for differentiating $GS \leq 3 + 4$ and $GS > 3 + 4$ PCa, $GS \leq 7$ and $GS > 7$ PCa, Kapp always had the biggest one in our every periodical (when the number of cases was 20/34/40) analysis. A previous study with big sample size ($n = 121$) report that Kapp exhibited significantly greater sensitivity for differentiating low- and high-grade PCa than ADC or *D* (68.6% vs 51.0% and 49.0%, respectively; $P < 0.004$) [54]. That in this present study was 92.31% with the Youden index of 0.63. These might suggest a potential clinical advantage for incorporating the DKI model into prostate MRI protocols. From another aspect, strong correlations were observed between ADC and Kapp, *F*, which may suggest that these metrics individually provide similar information in PCa. The similar correlation between ADC and Kapp had been reported before [32].

The amount of $GS = 3 + 3$ PCa involved was deficient in this study; actually, the original number of $GS = 3 + 3$ patients proved by biopsy was 13, and they all underwent mpMRI examination, but 7 (54%) of them upgraded to $GS = 3 + 4$ at final pathology through prostatectomy, and 4 (31%) of them did not find a defined lesion on mpMRI. As the method to define prostatic foci was based on the PI-RADS V2 which was incomprehensive, it gave the definition of clinically significant PCa as $GS \geq 7$ (including $3 + 4$ with prominent but not predominant Gleason 4 component), and/or volume ≥ 0.5 cc, and/or extraprostatic extension (EPE) [27]. PI-RADS score of ≥ 3 might rarely yield PCa of $GS \leq 6$. In our study and clinic, there could be cases in which mpMRI missed the diagnosis of $GS \leq 6$ PCa, but fortunately, this group was with low risk or harmless disease which is not likely to cause problems in a man's lifetime, and they are increasingly being managed with active surveillance [55]. And we recommend that those aged more than 50 years old without significant findings on mpMRI should follow-up (every 3 months) with PSA or ultrasound, etc.

There were some limitations to this study. First, the number of cases included in this study is limited, which may lead to errors due to sampling bias. In addition, the geographical source of our patients is relatively limited. These are common problems faced in other single-center studies. Second, the influence of image signal-to-noise ratio and the one-to-one correspondence between the lesion location on the gross specimen and the lesion location in the image were not solved in this study; therefore, there are some data measurement errors. Regarding the extent of misregistration, it is hoped that in future research, the quality of the image can be further improved, and the layer-by-layer slice pathology can be used as a reference. Third, the cancerous sample analysis did not consider differences in the central gland and peripheral lesions, because many cases had cancerous lesions in both regions. In addition, IVIM imaging and DK imaging

were scanned in a series of b values simultaneously, so IVIM measurements might be biased to some degree, as mentioned above. A large-sized sample study is warranted for further discussion and for regulating and refining the above results.

In conclusion, both the IVIM and DKI models are beneficial to predict GS of PCa and indirectly predict its aggressiveness. However, we found no significant additional performance to ADC in the present study. Nonetheless, work remains to be performed to fully understand the mechanisms underlying these two models, as well as the manner in which b values generate differences.

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References

- Parkin DM, Bray F, Ferlay J, Pisani P.(2005) Global cancer statistics, 2002. *CA Cancer J Clin*;55:74-108.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D.(2011) Global cancer statistics. *CA: A Cancer Journal for Clinicians*;61:69-90.
- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A.(2015) Global cancer statistics, 2012. *CA Cancer J Clin*;65:87-108.
- Epstein JI, Egevad L, Amin MB, Delahunt B, Srigley JR, Humphrey PA.(2016) The 2014 International Society of Urological Pathology (ISUP) Consensus Conference on Gleason Grading of Prostatic Carcinoma: Definition of Grading Patterns and Proposal for a New Grading System. *Am J Surg Pathol*;40:244-252.
- Anwar SSM, Anwar Khan Z, Shoab Hamid R, Haroon F, Sayani R, Beg M, Khattak YJ.(2014) Assessment of Apparent Diffusion Coefficient Values as Predictor of Aggressiveness in Peripheral Zone Prostate Cancer: Comparison with Gleason Score. *ISRN Radiology*;2014:1-7.
- Boesen L, Chabanova E, Løgager V, Balslev I, Thomsen HS.(2015) Apparent diffusion coefficient ratio correlates significantly with prostate cancer gleason score at final pathology. *J Magn Reson Imaging*;42:446-453.
- Kamel MH, Khalil MI, Alobuia WM, Su J, Davis R.(2018) Incidence of metastasis and prostate-specific antigen levels at diagnosis in Gleason 3 + 4 versus 4 + 3 prostate cancer. *Urol Ann*;10:203-208.
- Morash C, Tey R, Agbassi C, Klotz L, McGowan T, Srigley J, Evans A.(2015) Active surveillance for the management of localized prostate cancer: Guideline recommendations. *Can Urol Assoc J*;9:171-178.
- Ahmed HU, El-Shater BA, Brown LC, Gabe R, Kaplan R, Parmar MK, Collaco-Moraes Y, et al.(2017) Diagnostic accuracy of multiparametric MRI and TRUS biopsy in prostate cancer (PROMIS): a paired validating confirmatory study. *Lancet*;389:815-822.
- Rosenkrantz AB, Taneja SS.(2015) Prostate MRI Can Reduce Overdiagnosis and Overtreatment of Prostate Cancer. *Acad Radiol*;22:1000-1006.
- Jie C, Rongbo L, Ping T.(2014) The value of diffusion-weighted imaging in the detection of prostate cancer: a meta-analysis. *Eur Radiol*;24:1929-1941.
- Le Bihan D, Breton E, Lallemand D, Aubin ML, Vignaud J, Lavaljeantet M.(1988) Separation of diffusion and perfusion in intravoxel incoherent motion MR imaging. *Radiology*;168:497-505.
- Shinmoto H, Tamura C, Soga S, Shiomi E, Yoshihara N, Kaji T, Mulkern RV.(2012) An intravoxel incoherent motion diffusion-weighted imaging study of prostate cancer. *AJR Am J Roentgenol*;199:W496-W500.
- Liu X, Zhou L, Peng W, Wang C, Wang H.(2013) Differentiation of central gland prostate cancer from benign prostatic hyperplasia using monoexponential and biexponential diffusion-weighted imaging. *Magn Reson Imaging*;31:1318-1324.
- Kuru TH, Roethke MC, Stieltjes B, Maier-Hein K, Schlemmer HP, Hadaschik BA, Fenchel M.(2014) Intravoxel incoherent motion (IVIM) diffusion imaging in prostate cancer - what does it add? *J Comput Assist Tomogr*;38:558-564.
- Zhang Y, Wang Q, Wu C, Wang X, Zhang J, Liu H, Liu X, et al. (2015) The Histogram Analysis of Diffusion-Weighted Intravoxel Incoherent Motion (IVIM) Imaging for Differentiating the Gleason grade of Prostate Cancer. *Eur Radiol*;25:994-1004.
- Valerio M, Zini C, Fierro D, Giura F, Colarieti A, Giuliani A, Laghi A, et al.(2016) 3T multiparametric MRI of the prostate: Does intravoxel incoherent motion diffusion imaging have a role in the detection and stratification of prostate cancer in the peripheral zone? *Eur J Radiol*;85:790-794.
- Yang DM, Kim HC, Kim SW, Jahng GH, Won KY, Lim SJ, Oh JH.(2016) Prostate cancer: correlation of intravoxel incoherent motion MR parameters with Gleason score. *Clin Imaging*;40:445-450.
- Jensen JH, Helpert JA, Ramani A, Lu H, Kaczynski K.(2005) Diffusional kurtosis imaging: The quantification of non-gaussian water diffusion by means of magnetic resonance imaging. *Magn Reson Med*;53:1432-1440.
- Suo S, Chen X, Wu L, Zhang X, Yao Q, Fan Y, Wang H, et al. (2014) Non-Gaussian water diffusion kurtosis imaging of prostate cancer. *Magn Reson Imaging*;32:421-427.
- Tamura C, Shinmoto H, Soga S, Okamura T, Sato H, Okuaki T, Pang Y, et al.(2014) Diffusion kurtosis imaging study of prostate cancer: Preliminary findings. *J Magn Reson Imaging*;40:723-729.
- Quentin M, Pentang G, Schimmöller L, Kott O, Müller-Lutz A, Blondin D, Arsov C, et al.(2014) Feasibility of diffusional kurtosis tensor imaging in prostate MRI for the assessment of prostate cancer: Preliminary results. *Magn Reson Imaging*;32:880-885.
- Wang Q, Li H, Yan X, Wu C, Liu X, Shi H, Zhang Y.(2015) Histogram analysis of diffusion kurtosis magnetic resonance imaging in differentiation of pathologic Gleason grade of prostate cancer. *Urologic Oncology: Seminars and Original Investigations*;33:315-337.
- Wu CJ, Zhang YD, Bao ML, Li H, Wang XN, Liu XS, Shi HB.(2017) Diffusion Kurtosis Imaging Helps to Predict Upgrading in Biopsy-Proven Prostate Cancer With a Gleason Score of 6. *AJR Am J Roentgenol*;209:1081-1087.
- Tamada T, Prabhu V, Li J, Babb JS, Taneja SS, Rosenkrantz AB.(2017) Prostate Cancer: Diffusion-weighted MR Imaging for Detection and Assessment of Aggressiveness-Comparison between Conventional and Kurtosis Models. *Radiology*;284:100-108.
- Epstein JI, Feng Z, Trock BJ, Pierorazio PM.(2012) Upgrading and Downgrading of Prostate Cancer from Biopsy to Radical Prostatectomy: Incidence and Predictive Factors Using the Modified Gleason Grading System and Factoring in Tertiary Grades. *Eur Urol*;61:1019-1024.
- Weinreb JC, Barentsz JO, Choyke PL, Cornud F, Haider MA, Macura KJ, Margolis D, et al.(2016) PI-RADS Prostate Imaging – Reporting and Data System: 2015, Version 2. *Eur Urol*;69:16-40.
- Jung SI, Jeon HJ, Park HS, Yu MH, Kim YJ, Lee SE, Lim SD.(2018) Multiparametric MR imaging of peripheral zone prostate cancer: effect of postbiopsy hemorrhage on cancer detection

- according to Gleason score and tumour volume. *The British journal of radiology*;91:20180001.
29. Rosenkrantz AB, Padhani AR, Chenevert TL, Koh D, De Keyzer F, Taouli B, Le Bihan D.(2015) Body diffusion kurtosis imaging: Basic principles, applications, and considerations for clinical practice. *J Magn Reson Imaging*;42:1190-1202.
 30. Yang M, Yan Y, Wang H.(2019) IMAge/enGINE: a freely available software for rapid computation of high-dimensional quantification. *Quant Imag Med Surg*;9:210-218.
 31. Cicchetti DV.(1994) Guidelines, Criteria, and Rules of Thumb for Evaluating Normed and Standardized Assessment Instruments in Psychology. *Psychol Assessment*;6:284-290.
 32. Yang L, Rao S, Wang W, Chen C, Ding Y, Yang C, Grimm R, et al. (2018) Staging liver fibrosis with DWI: is there an added value for diffusion kurtosis imaging? *Eur Radiol*;28:3041-3049.
 33. DeLong ER, DeLong DM, Clarke-Pearson DL.(1988) Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics*;44:837-845.
 34. Humphrey PA.(2004) Gleason grading and prognostic factors in carcinoma of the prostate. *Modern Pathol*;17:292-306.
 35. Epstein JI, Allsbrook WC, Amin MB, Egevad LL, Bastacky S, Beltran AL, Berner A, et al.(2005) The 2005 International Society of Urological Pathology (ISUP) Consensus Conference on Gleason Grading of Prostatic Carcinoma. *Am J Surg Pathol*;29:1228-1242.
 36. Chatterjee A, Watson G, Myint E, Sved P, McEntee M, Bourne R.(2015) Changes in Epithelium, Stroma, and Lumen Space Correlate More Strongly with Gleason Pattern and Are Stronger Predictors of Prostate ADC Changes than Cellularity Metrics. *Radiology*;277:751-762.
 37. Boesen L, Chabanova E, Løgager V, Balslev I, Thomsen HS.(2015) Apparent diffusion coefficient ratio correlates significantly with prostate cancer gleason score at final pathology. *J Magn Reson Imaging*;42:446-453.
 38. De Cobelli F, Ravelli S, Esposito A, Giganti F, Gallina A, Montorsi F, Del Maschio A.(2015) Apparent Diffusion Coefficient Value and Ratio as Noninvasive Potential Biomarkers to Predict Prostate Cancer Grading: Comparison With Prostate Biopsy and Radical Prostatectomy Specimen. *Am J Roentgenol*;204:550-557.
 39. Doo KW, Sung DJ, Park BJ, Kim MJ, Cho SB, Oh YW, Ko YH, et al.(2012) Detectability of low and intermediate or high risk prostate cancer with combined T2-weighted and diffusion-weighted MRI. *Eur Radiol*;22:1812-1819.
 40. Hambroek T, Somford DM, Huisman HJ, van Oort IM, Witjes JA, Hulsbergen-van DKC, Scheenen T, et al.(2011) Relationship between apparent diffusion coefficients at 3.0-T MR imaging and Gleason grade in peripheral zone prostate cancer. *Radiology*;259:453-461.
 41. Anwar SS, Anwar KZ, Shoaib HR, Haroon F, Sayani R, Beg M, Khattak YJ.(2014) Assessment of apparent diffusion coefficient values as predictor of aggressiveness in peripheral zone prostate cancer: comparison with Gleason score. *ISRN Radiol*;2014:263417.
 42. Gilani N, Malcolm P, Johnson G.(2017) A model describing diffusion in prostate cancer. *Magn Reson Med*;78:316-326.
 43. Koh D, Collins DJ, Orton MR.(2011) Intravoxel incoherent motion in body diffusion-weighted MRI: reality and challenges. *AJR. American journal of roentgenology*;196:1351.
 44. Pang Y, Turkbey B, Bernardo M, Kruecker J, Kadoury S, Merino MJ, Wood BJ, et al.(2013) Intravoxel incoherent motion MR imaging for prostate cancer: An evaluation of perfusion fraction and diffusion coefficient derived from different b-value combinations. *Magn Reson Med*;69:553-562.
 45. Le Bihan D.(2007) The ‘wet mind’: water and functional neuroimaging. *Phys Med Biol*;52:R57-R90.
 46. Merisaari H, Toivonen J, Pesola M, Taimen P, Boström PJ, Pahikala T, Aronen HJ, et al.(2015) Diffusion-weighted imaging of prostate cancer: effect of b-value distribution on repeatability and cancer characterization. *Magn Reson Imaging*;33:1212-1218.
 47. Barbieri S, Brönnimann M, Boxler S, Vermathen P, Thoeny HC.(2017) Differentiation of prostate cancer lesions with high and with low Gleason score by diffusion-weighted MRI. *Eur Radiol*;27:1547-1555.
 48. Pesapane F, Patella F, Fumarola EM, Panella S, Ierardi AM, Pompili GG, Franceschelli G, et al.(2017) Intravoxel Incoherent Motion (IVIM) Diffusion Weighted Imaging (DWI) in the Peripheral Prostate Cancer Detection and Stratification. *Med Oncol*;34:35.
 49. Patel J, Sigmund EE, Rusinek H, Oei M, Babb JS, Taouli B.(2010) Diagnosis of cirrhosis with intravoxel incoherent motion diffusion MRI and dynamic contrast-enhanced MRI alone and in combination: Preliminary experience. *J Magn Reson Imaging*;31:589-600.
 50. Hectors SJ, Semaan S, Song C, Lewis S, Haines GK, Tewari A, Rastinehad AR, et al.(2018) Advanced Diffusion-weighted Imaging Modeling for Prostate Cancer Characterization: Correlation with Quantitative Histopathologic Tumor Tissue Composition-A Hypothesis-generating Study. *Radiology*;286:938-948.
 51. Lawrence EM, Warren AY, Priest AN, Barrett T, Goldman DA, Gill AB, Gnanapragasam VJ, et al.(2016) Evaluating Prostate Cancer Using Fractional Tissue Composition of Radical Prostatectomy Specimens and Pre-Operative Diffusional Kurtosis Magnetic Resonance Imaging. *Plos One*;11:e159652.
 52. Le Bihan D.(2013) Apparent diffusion coefficient and beyond: what diffusion MR imaging can tell us about tissue structure. *Radiology*;268:318-322.
 53. Li C, Chen M, Wan B, Yu J, Liu M, Zhang W, Wang J.(2018) A comparative study of Gaussian and non-Gaussian diffusion models for differential diagnosis of prostate cancer with in-bore transrectal MR-guided biopsy as a pathological reference. *Acta Radiol*;59:1395-1402.
 54. Rosenkrantz AB, Sigmund EE, Johnson G, Babb JS, Mussi TC, Melamed J, Taneja SS, et al.(2012) Prostate Cancer: Feasibility and Preliminary Experience of a Diffusional Kurtosis Model for Detection and Assessment of Aggressiveness of Peripheral Zone Cancer. *Radiology*;264:126-135.
 55. Purysko AS, Rosenkrantz AB, Barentsz JO, Weinreb JC, Macura KJ.(2016) PI-RADS Version 2: A Pictorial Update. *Radiographics*;36:1354-1372.

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