**ORIGINAL ARTICLE**

# **Why bother with alpha particles?**

**A. Paden King1,2 · Frank I. Lin1 · Freddy E. Escorcia1,[2](http://orcid.org/0000-0002-0727-3242)**

Received: 3 April 2021 / Accepted: 24 May 2021 / Published online: 27 June 2021 © This is a U.S. government work and not under copyright protection in the U.S.; foreign copyright protection may apply 2021

#### **Abstract**

The approval of  $^{223}$ RaCl<sub>2</sub> for cancer therapy in 2013 has heralded a resurgence of interest in the development of α-particle emitting radiopharmaceuticals. In the last decade, over a dozen α-emitting radiopharmaceuticals have entered clinical trials, spawned by strong preclinical studies. In this article, we explore the potential role of  $\alpha$ -particle therapy in cancer treatment. We begin by providing a background for the basic principles of therapy with  $\alpha$ -emitters, and we explore recent breakthroughs in therapy with  $\alpha$ -emitting radionuclides, including conjugates with small molecules and antibodies. Finally, we discuss some outstanding challenges to the clinical adoption of  $\alpha$ -therapies and potential strategies to address them.

### **Introduction**

There has been a resurgence of interest in radiopharmaceutical therapy (RPT) for cancer treatment, with three RPTs—  $[$ <sup>177</sup>Lu]Lu-DOTATATE (NCT01578239),  $[$ <sup>131</sup>I]Iobenguane (NCT00874614), and  $^{223}$ RaCl<sub>2</sub> (NCT00699751)—recently demonstrating overall survival beneft and receiving FDA approval  $[1-3]$  $[1-3]$  $[1-3]$ . Treament with  $[177$ Lu]Lu-PSMA-617 was compared to standard of care in the phase III VISION trial in men with metastatic castration-resistance prostate cancer (mCRPC) and demonstrated an overall survival beneft [\[4](#page-7-2)]. Over a dozen more agents are currently in clinical trials, including radioconjugates of small molecules, antibodies, and nanoparticles [[5\]](#page-7-3). These conjugates have high tumor specifcity and potency, unlike many traditional chemotherapeutics. Recent advances, built upon decades of preclinical studies, have demonstrated the therapeutic efficacy of  $\alpha$ -emitting radionuclides, leading to the frst FDA approval of an α-particle therapeutic,  $^{223}$ RaCl<sub>2</sub>, for bone-limited mCRPC in 2013. The success of  $^{223}$ RaCl<sub>2</sub> has reinspired further research into

This article is part of the Topical Collection on Editorial.

 $\boxtimes$  Freddy E. Escorcia freddy.escorcia@nih.gov

- <sup>1</sup> Molecular Imaging Branch, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD 20817, USA
- Radiation Oncology Branch, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD 20817, USA

α-particle-emitting radioconjugates using radionuclides such as  $^{225}$ Ac,  $^{212}$ Pb,  $^{227}$ Th,  $^{213}$ Bi, and  $^{211}$ At. Herein, we introduce the premise of molecular radiotherapy, detailing the features that make α-emitting nuclides ideal for this application.

The basic principle of using targeted radiation for cancer treatment is simple: deliver cytotoxic ionizing radiation that causes single-strand and double-strand breaks in the DNA of a cancer cell, and subsequent cell death, while limiting damage to healthy tissues  $[6]$  $[6]$ . However, this concept has proven difficult in application, and even clinically approved radiopharmaceutical therapies have faced an uphill battle for widespread clinical implementation. Radiotherapy has been employed successfully for over a century with the frst external beam radiation therapy being developed shortly after the discovery of the X-ray [[7\]](#page-7-5). While efective at improving outcomes for patients with various types of cancers at early and oligometastatic stages, external radiation cannot defnitively treat widely disseminated disease. In contrast, parenterally administered radionuclides can be selectively targeted to cancerous cells, delivering therapeutic radiation doses to numerous sites of metastases. Molecular radiotherapy can be reduced to two main components that must be considered: the choice of biomolecule used as the tumor-targeting vector, and the emission: typically, α-particle, β−-particle, or Auger electrons. By carefully selecting and optimizing these components, researchers can develop RPT to improve the outcomes of cancer patients. In this article, we focus on the development of α-particle radiopharmaceuticals, description of their properties, suitability for incorporation into radioconjugates, and key advances in the field of  $\alpha$ -RPT.



### **Physical properties and anticancer mechanisms of radiotherapeutics**

The decay characteristics of  $\alpha$ -emitting radionuclides, namely their impressive cytotoxic potential and no known mechanism of cellular resistance, make them compelling for RPT. It is estimated that several hundred  $\beta^-$  traversals through the nucleus are required to induce cell death, while less than ten  $\alpha$ -decays can achieve the same result [[8\]](#page-7-6). This incredible efficacy arises from the higher linear energy transfer (LET) of α-particles compared to β− -particles. Auger emitters have high LET and shorter range than either  $\alpha$ - or β−-particles, requiring internalization and localization to the nucleus to have a therapeutic effect [\[9](#page-7-7)]. α-particles deposit all their energy over a distance of a few cell diameters, whereas  $\beta^-$ -particles may pass completely through cells without interacting with cellular components and without depositing any energy within a cell, as shown in Fig. [1.](#page-1-0) The anticancer mechanism of  $\alpha$ -particles also supports their use for RPT. DNA damage by α-particles is multi-faceted and includes indirect damage via reactive oxygen species (ROS) or direct damage with resulting double-strand DNA breaks, DNA cross-linking, and complex chromosomal rearrangement [[10\]](#page-7-8). The propensity of α-particles to cause catastrophic double-strand DNA breaks is a well-documented signature therapeutic effect [\[11](#page-7-9), [12\]](#page-7-10). Conversely,  $\beta^-$ -particles cause mostly single-strand DNA breaks. Unsurprisingly, double-strand breaks are more challenging to repair than single-strand breaks, and the more induced, the higher the probability of cell kill [[13](#page-7-11)]. Additionally, recent studies show that the cytotoxic effects of α-particles can activate antitumor immune responses, further increasing their potency beyond the initial cytotoxic insult  $[14]$  $[14]$ . Finally,  $\alpha$ -particles have a penetration depth through tissue of only 40–100  $\mu$ m, much shorter than the up to 12-mm penetration depth of β−-particles. If one successfully localizes an α-emitter to the tumor via a targeting vector, the shorter range can theoretically result in less collateral damage to healthy tissues, increasing the therapeutic index [[15](#page-7-13)]. However, this shorter range limits the "cross-fire" effect of α-particles, which is advantageous in β−-particle therapies.

The cytotoxicity of  $\alpha$ -emitters renders them less susceptible to the development of resistance than traditional chemotherapeutics or even β−-emitters [\[16,](#page-7-14) [17\]](#page-7-15). Recent clinical studies have demonstrated that  $\alpha$ -emitting radioconjugates can be used successfully as a salvage therapy after failure of β−-emitting RPTs. For instance, in patients with neuroendocrine cancers refractory to treatment with the β−-emitting [ 177Lu]Lu-DOTATATE, subsequent treatment with  $[^{225}Ac]Ac$ -DOTATATE induced an antitumor response in some patients [\[18](#page-7-16)]. Similar results were observed when prostate cancer patients refractory to treatment with  $[177$ Lu] Lu-PSMA-617 were treated with  $[^{225}Ac]Ac$ -PSMA-617 [[19,](#page-7-17) [20\]](#page-7-18). In both the PSMA and DOTATATE examples, simply switching to the  $\alpha$ -emitter, while keeping the targeting ligand constant, overcame resistance and improved response in some patients. Although randomized controlled trials are needed to defnitively conclude a beneft, these studies suggest that  $\alpha$ -particle therapy represents at least a promising salvage treatment following  $\beta^-$ -therapy.

In addition to the observed lack of resistance to  $\alpha$  therapies, the anticancer mechanisms of  $\alpha$ -emitters have also been investigated in vitro [[17](#page-7-15), [21\]](#page-7-19). Although these studies are complicated by the hazards associated with longlived α-emitters and the challenges of accurately estimating absorbed radiation doses on cellular and subcellular levels, several key insights have been gained into  $\alpha$ -particle toxicity and resistance. One recent preclinical study showed that the  $\alpha$ -emitter <sup>223</sup>RaCl<sub>2</sub> exhibited 10 × more potency than the γ-emitter 137Cs, which primarily induces cell death via the generation of ROS [[17](#page-7-15)]. Further experimentation demonstrated that  $\alpha$ -therapy was not cross-resistant with  $^{137}Cs$ . Cells with very high resistance to γ-radiation remained



<span id="page-1-0"></span>**Fig. 1** Relative LET and path lengths of the three classes of therapeutic radionuclides

sensitive to  $\alpha$ -emissions. Mutations in cellular antioxidant response proteins Nrf2 and KEAP1 greatly increased the effectiveness of  $\alpha$ -therapy indicating their presence may represent biomarkers for α-RPT. The same gain-of-function Nrf2 mutations conferred resistance to γ-radiation. Another study explored the effectiveness of the  $\alpha$ -emitter <sup>223</sup>Ra in cell lines with varying mutations that afect DNA repair pathways [[22\]](#page-7-20). This study found that cell lines with mutations in *BRCA2* or *ATM* were especially sensitive to  $\alpha$ -radiation, and suggests that inhibition of DNA-damage repair pathways could augment treatment response. In addition to causing double-strand DNA breaks, α-emitters have been shown to induce cell-cycle arrest and inhibit DNA synthesis in vivo [[23\]](#page-7-21). A follow-up study demonstrated that the combination of the same  $\alpha$ -therapy with gemcitabine could prevent G2-arrest and increase cell death [\[24](#page-8-0)].

A unique advantage of most radioconjugates, including  $\alpha$ -emitters, is their natural pairing with complementary diagnostic imaging. One direct beneft of this "theranostic" approach resides in optimal patient selection and subsequent monitoring of patients following RPT. Patients may be first injected with a diagnostic molecule, such as [<sup>68</sup> Ga] Ga-DOTATATE, to assess disease status prior to treatment with an  $\alpha$ -emitter DOTATATE conjugate, for example. In some cases, the same element can be used for both diagnosis and therapy, for example the  $\beta^-$ -emitter <sup>212</sup>Pb (in vivo generator of  $\alpha$ -emitter <sup>212</sup>Bi) is used for therapy and its matched element 203Pb is useful for diagnostic purposes via SPECT imaging  $[25]$  $[25]$  $[25]$ . This approach allows for monitoring of the radioconjugates' biodistribution as well as real-time tracking of the patients' treatment response. Finally, some α-emitting nuclides (or their daughters) may be imaged directly. Several recent studies have focused on SPECT imaging of X-ray and  $\gamma$ -emissions of <sup>225</sup>Ac, <sup>227</sup>Th, <sup>223</sup>Ra, and their daughters [\[26–](#page-8-2)[28\]](#page-8-3). The broader application of this approach remains uncertain due to the inherently low sensitivity of SPECT and the low activities needed for RPT. The use of partner  $\beta^+$ -emitting isotopes having biodistribution patterns at least

<span id="page-2-0"></span>**Table 1** α-emitting nuclides and their physical properties

partially similar to  $\alpha$ -emitters (e.g., <sup>18</sup>F and <sup>223</sup>Ra) may be helpful and warrant study [\[29](#page-8-4), [30](#page-8-5)].

Hundreds of  $\alpha$ -emitting radionuclides are theoretically accessible, but only a few have suitable half-lives and realistic production to make them potentially useful for clinical RPT. These radionuclides are  $^{227}$ Th,  $^{223}$ Ra,  $^{225}$ Ac,  $^{212}$ Pb,  $^{211}$ At,  $^{149}$  Tb, and  $^{213}$ Bi. All have unique properties that make them advantageous for combination with diferent targeting moieties. Detailed information on the physical properties and relevant information for these commonly used  $\alpha$ -emitting nuclides are presented in Table [1](#page-2-0). Broadly, these radionuclides may be categorized according to half-life, with  $^{227}$ Th,  $^{225}$ Ac, and  $^{223}$ Ra having half-lives > 10 days, while <sup>212</sup>Pb, <sup>211</sup>At <sup>149</sup> Tb, and <sup>213</sup>Bi all have half-lives < 12 h.

Conventionally and optimally, the half-life of the isotope is matched to the half-life of the biomolecule to which they are coupled. Accordingly, the short half-life radioisotopes are considered better partners for small molecules, and longlived isotopes are considered better partners for antibodies, which have long biological half-lives (Table [1](#page-2-0)). However, preclinical and clinical studies have shown that <sup>225</sup>Ac radioconjugates with small molecules are also therapeutically effective, if inefficient.

## <sup>223</sup>RaCl<sub>2</sub>: first-in-class α-particle RPT

Unconjugated radionuclides, such as  $131$ I, are some of the oldest and most successful RPTs. These therapies rely on the radionuclide's intrinsic localization to specifc cancerous cells. The only "free" α-emitter to have clinical or preclinical success is  $223$ Ra. Although naturally occurring radium was first used therapeutically over a century ago  $[32]$  $[32]$ ,  $^{223}$ RaCl<sub>2</sub> only gained FDA approval in 2013  $[3, 33, 34]$  $[3, 33, 34]$  $[3, 33, 34]$  $[3, 33, 34]$  $[3, 33, 34]$  $[3, 33, 34]$ . <sup>223</sup>Ra localizes naturally to newly forming bone, including sites of metastases, due to its chemical similarity to  $Ca^{2+}$ . <sup>223</sup>Ra improved overall survival in patients with bone-limited mCRPC with an acceptable toxicity profle. Clinical trials



are ongoing to explore additional applications of  $^{223}RaCl<sub>2</sub>$ as a cancer therapeutic.

Despite its clinical approval and current use, several questions must be answered to optimize the efficacy of  $^{223}$ RaCl<sub>2</sub>, and to understand and mitigate associated adverse events sufficiently. Several studies focusing on  $^{223}$ RaCl<sub>2</sub> are summa-rized in Table [2](#page-3-0). Exploiting  $\gamma$ -emissions of <sup>223</sup>Ra for imaging using SPECT is possible, though at the expense of long scan times. Treatment combinations of  $^{223}$ Ra with standard-ofcare chemotherapy are being explored in a phase I/IIa trial. This trial looks to define the safety and efficacy combining 223Ra and docetaxel (NCT03574571) in patients with mCRPC [\[35](#page-8-10)]. The phase II RAVENS trial (NCT04037358) is evaluating whether  $^{223}$ RaCl<sub>2</sub> might improve progressionfree survival in men with oligometastic prostate cancer receiving stereotactic ablative radiotherapy [[36\]](#page-8-11). Such studies are critical in determining which α-emitter-drug combinations are benefcial, or just as importantly, harmful [\[37\]](#page-8-12). For example, a recent combination study investigating the addition of  $2^{23}$ Ra to abiraterone acetate and prednisone therapy in mCRPC patients unexpectedly reported more fractures in the patients treated with the investigational arm compared with the placebo (NCT02043678) [\[38](#page-8-13)]. Long-term sequelae of  $^{223}$ RaCl<sub>2</sub> treatment are incompletely

understood. An observational clinical study (NCT04256993) is investigating long-term efficacy and side effects in a large cohort of patients, focusing on the incidence of bone fractures in patients treated with  $^{223}$ RaCl<sub>2</sub>. Collectively, results from these trials can help guide  $^{223}$ RaCl<sub>2</sub> treatment, improve patient selection, and mitigate untoward effects. Despite  $223$ Ra's effectiveness as a therapeutic drug, the potential for expanding its use in targeted radiotherapy conjugates is limited due to a lack of available in vivo compatible chelators. In a recent study, Abou et al. reported on stable  $^{223}$ Ra chelation in vivo, which may facilitate the development of new  $223$ Ra-based conjugates using tumor-targeted vectors [\[39\]](#page-8-14).

### **α‑Emitter conjugates to small molecules and antibodies**

Several properties outlined in the previous sections make α-emitters promising for development of conjugated RPT agents. The natural pairing of radioactive groups with cancer-targeting molecules has long been recognized. Much of the groundwork for therapeutic radioconjugates may be established using radiodiagnostics [[40](#page-8-15)[–44\]](#page-8-16). Unlike conventional chemotherapeutics, which must interact directly

<span id="page-3-0"></span>**Table 2** Categories of alpha-RPT, their properties, and relevant clinical studies

Radiotherapeutic Category	<b>Advantages</b>	<b>Disadvantages</b>	<b>Notable Clinical Studies</b>	<b>Cancer Type</b>
Unconjugated Radionuclides $^{223}\mbox{RaCl}_2$	Simple to produce	Little $\bullet$ modification possible Limited number of potential nuclides	RaCl <sub>2</sub> FDA-approved in 2013 [26]	mCRPC $\bullet$
	and test Intrinsic $\bullet$		RaCl <sub>2</sub> biodistribution (Phase 1, NCT04521361)	mCRPC
	localization to cancerous tissue		RaC <sub>b</sub> side effects (NCT04256993) $\bullet$	mCRPC $\bullet$
	Proven clinical success		RaCl <sub>2</sub> combination with docetaxel (Phase 3, $\bullet$ NCT03574571)	mCRPC $\bullet$
			RaCl <sub>2</sub> treatment of metastatic breast cancer (Phase 2, NCT02366130, NCT04090398)	<b>Breast Cancer with Bone</b> Metastases
			RaCl <sub>2</sub> combination with abiraterone (Phase 3, NCT02043678)	mCRPC $\bullet$
Small Molecule Radioconjugates $M^{n+} = {}^{225}Ac$ , ${}^{213}Bi$ , ${}^{212}Pb$ , ${}^{211}At$	Short half-life of small molecules	Only moderate $\bullet$ specificity Short lived $\bullet$	$[{}^{212}Pb]Pb$ -DOTAMTATE (Phase 1, NCT03466216) $\bullet$	SSTR <sup>+</sup> NET $\bullet$
	facilitates		$[^{225}Ac]Ac$ -PSMA-617 (Phase 1, NCT04597411) $\bullet$	mCRPC $\bullet$
	radionuclide clearance Concept established clinically with $\beta$ emitters	conjugates must be labeled onsite	$[^{225}Ac]Ac$ -PSMA ligand (Phase 1, NCT04225910)	mCRPC $\bullet$
Antibody Radioconjugates $M^{n+} = {}^{227}Th$ , ${}^{225}Ac$ , ${}^{212}Pb$ , ${}^{211}At$ , $^{213}$ Bi	Excellent binding $\bullet$ affinity for targets	Long biological half lives require excellent chelators Long lived $\bullet$ daughters may redistribute Radiolabeling $\bullet$ often difficult	$[^{225}Ac]Ac$ -aCD33 (Phase 1, NCT02998047) $\bullet$	Multiple Myeloma
	Complements $\bullet$		[225 Ac] Ac-IGF-1R (Phase 1, NCT03746431) $\bullet$	IGF-1R <sup>+</sup> Solid Tumors
	unlabeled antibody therapies		[225Ac]Ac-J591 (Phase 1, NCT03276572, NCT04506567)	mCRPC $\bullet$
	High potency pairs well with low-		$[^{227}Th]Th-aMSLN$ (Phase 1, NCT03507452)	<b>MSLN<sup>+</sup></b> Tumors $\bullet$
	expression targets		[227Th]Th-aCD22 (Phase 1, NCT02581878) $\bullet$	Non-Hodgkin's Lymphoma $\bullet$
			$[^{227}Th]Th-aPSMA$ (Phase 1, NCT03724747) $\bullet$	mCRPC $\bullet$
			[ <sup>212</sup> Pb]Pb-aHER-2 (Phase 1, NCT01384253) $\bullet$	HER2 <sup>+</sup> Tumors $\bullet$
			$[{}^{213}Bi]Bi$ aCD33 (Phase 1, NCT03466216)	SSTR <sup>+</sup> NET $\bullet$
			$[{}^{211}At]At$ -aCD45 (Phase 1/2, NCT03670966)	Leukemia, Myelodysplasia
			$[^{211}At]At aCD38$ (Phase 1, NCT04466475)	Multiple Myeloma

*mCRCP*, metastatic castration-resistant prostate cancer; *SSTR*, somatostatin receptor; *NET*, neuroendocrine tumor; *MSLN*, mesothelin; *IGF-1R*, insulin-like growth-factor 1 receptor

with a target biomolecule (e.g., DNA, microtubules), RPT agents can induce cell death from outside the cell. This property is advantageous for target-based therapies. Cleavable linkers or cellular uptake and degradation are not necessary to induce cell death, unlike traditional smallmolecule and antibody-based treatments. However, cellular uptake and retention is highly advantageous for RPTs. Cell uptake prevents daughter redistribution, and increased proximity to the cell nucleus can lead to higher cytotoxicity. Another potential advantage of radioconjugates, relative to many targeted drugs, is that they are cytocidal. This feature can transform an unlabeled targeted agent that does not have much of a therapeutic efect to a radiolabeled one that does  $[45]$  $[45]$  $[45]$ . Furthermore, efficient tumor cell kill can help mitigate development of resistance via compensatory changes (e.g., decreased availability or expression of target molecules, selection for clones with resistant mutations) following therapy.

Thus far, small-molecule radioconjugates have enjoyed the greatest clinical success, and the small-molecule conjugates that are currently clinically approved use  $\beta^-$ emitting nuclides, likely because these conjugates are easier to develop and more widely available than their α-emitting counterparts. However, as discussed in the previous section, α-particle-based therapies are also being explored. Somatostatin receptor (SSTR)–targeting ligands have been conjugated to DOTA or DOTA-derivative chelators to treat SSTR-expressing neuroendocrine tumors [[46,](#page-8-18) [47](#page-8-19)]. Several  $\alpha$ -emitting radionuclides have been investigated preclinically in SSTR-targeting conjugates, including  $^{225}$ Ac,  $^{213}$ Bi, and  $^{212}$ Pb. Preclinical investigation of  $[^{225}Ac]Ac$ -DOTATOC showed that this conjugate has marked antitumor activity in vivo [\[48\]](#page-8-20). The related conjugate  $[^{225}Ac]Ac$ -DOTATATE has been investigated clinically in patients who progressed or had stable disease following treatment with  $\left[ {}^{177}$ Lu]Lu-DOTATATE  $\left[ 18, 49 \right]$  $\left[ 18, 49 \right]$  $\left[ 18, 49 \right]$  $\left[ 18, 49 \right]$  $\left[ 18, 49 \right]$ . Short-term results from this phase I trial indicate that this conjugate is well-tolerated. Early efficacy data reported 1 patient of 53 in the study with complete remission, 41 patients with partial remission, and 9 patients showing stable disease. A retrospective clinical evaluation of  $[^{213}B1]$ Bi-DOTATOC in 8 patients showed that this conjugate is very well-tolerated and exhibited a promising antitumor response [[50](#page-8-22)]. Finally, the related compound  $[^{212}Pb]Pb$ -DOTAMTATE has shown preclinical efficacy for  $SSTR<sup>+</sup>$ tumor treatment [[51](#page-8-23)], and a phase I trial of this conjugate is currently underway (NCT03466216). Notably, unlike the other studies mentioned, this trial focuses on patients naive to therapy with  $177$ Lu- or  $90$ Y-labeled DOTATATE. Preliminary results indicate that the conjugate is welltolerated and has a promising efficacy signal with two patients exhibiting complete response and one with a partial response [[52](#page-8-24)].

In addition to SSTR-targeting conjugates, several studies have focused on PSMA-targeting  $\alpha$ -particle RPTs [\[53](#page-9-0)]. Chelates of 213Bi, 212Pb, and 225Ac have been conjugated to PSMA-targeting ligands. [<sup>225</sup>Ac]Ac-PSMA-617 has demonstrated promising efectiveness with complete remission for 2 out of 2 patients with late-stage mCRPC as reported in a small retrospective frst-in-human study [\[19](#page-7-17)]. Importantly, a prospective phase 1 trial of this conjugate is currently underway (NCT04597411). One prospective cohort study compared 28 mCRPC patients of whom half were refractory to  $\left[ {}^{177}$ Lu]Lu-PSMA-617 treatment, whereas the other half were RPT-naïve [[20\]](#page-7-18). Patients in both groups showed objective response to treatment. The dose was well-tolerated, suggesting that  $[^{225}Ac]Ac-PSMA-617$  may be a good candidate for both front-line and second-line treatment of mCRPC.  $^{212}Pb$  and  $^{213}Bi$  have also been investigated with PSMA-617 and other PSMA-targeting molecules. Several studies have shown promising preclinical results in vivo using 212Pb, but no clinical data has yet been collected [[54\]](#page-9-1). A case report of one patient with mCRPC who exhibited cancer progression demonstrated remarkable radiographic and biochemical response after treatment with  $[^{213}Bi]Bi-PSMA-617[55]$  $[^{213}Bi]Bi-PSMA-617[55]$  $[^{213}Bi]Bi-PSMA-617[55]$ . These reports are helpful in providing bioplausibility and feasibility of more systematic clinical evaluation of such agents, but the short half-life and difficulty in distribution of 213Bi represent signifcant hurdles to further clinical development.

Antibody-based radioconjugates have been explored to augment the inherent limited therapeutic efect of tumor target-specifc antibodies. Many antibody therapies currently in the clinic are unconjugated relying upon antibody-dependent cellular cytotoxicity (ADCC) to induce cell death [\[56](#page-9-3)]. However, ADCC alone may be insufficient for tumor eradication. This limitation has spurred a renewed excitement for antibody–drug and antibody-radionuclide conjugates. Currently, two antibody-based RPTs are FDA-approved. Both therapeutics use β−-emitters conjugated to antibodies targeting the B-cell lymphoma biomarker CD20. However, the frst approved radioconjugate, [ 131I]I-Tositumomab (Bexxar®) is no longer in production, and the second conjugate,  $[{}^{90}Y]$ Y-Ibritumomab (Zevalin®), has seen decreased year-overyear utilization [[57](#page-9-4)[–60\]](#page-9-5). Despite demonstrating durable complete responses in patients with B-cell malignancies, these agents were not broadly adopted clinically. This outcome refects broader structural barriers to RPTs within the practice of medicine, particularly in the USA. In addition, the non-radioactive CD20-specifc antibody, rituximab, has the same indication as the radioconjugates and fts more neatly with existing clinical workfows.

Two  $\alpha$ -emitting antibody conjugates targeting the leukemia biomarker CD33 have been clinically investigated. The first paired <sup>213</sup>Bi with anti-CD33. The clinical studies showed that the conjugate induced remission in most

patients [[61](#page-9-6), [62\]](#page-9-7). Furthermore, in vitro studies of  $[^{213}Bi]$ Bi-anti-CD33 showed this conjugate was not cross-resistant with cells resistant to  $\beta^-$ - and γ-radiation [\[63\]](#page-9-8). However, the short half-life of this isotope  $(t_{1/2}=46 \text{ min})$  and difficulties in production and cost led to a shift to  $[^{225}Ac]Ac$ -anti-CD33 [\[64\]](#page-9-9). This second-generation CD33-targeting radioconjugate is currently in clinical trials (NCT02575963), and preliminary fndings indicate it is well-tolerated and exhibits anti-leukemic activity [[65,](#page-9-10) [66](#page-9-11)]. Despite the logistic and economic difficulties of implementing the  $[^{213}Bi]Bi$ -anti-CD33 conjugate, other antibody conjugates of 213Bi have entered clinical trials, and preliminary results indicate that these are well-tolerated in patients. In fact, the conjugate  $[^{213}Bi]$ Bi-cDTPA-9.2.27, which targets chondroitin sulfate proteoglycan 4 (a.k.a NG2) overexpressed in melanomas, has proven to be tolerable at doses of up to 450 mCi (16.7 GBq) and shows activity in patients with melanoma [[67\]](#page-9-12). Another 213Bi-antibody conjugate targeting EGFR has recently been investigated in a pilot feasibility prospective clinical study for the treatment of high grade carcinoma in situ of the bladder [\[68](#page-9-13)]. The RPT induced complete remission in three of 12 patients with no observed adverse efects and warrants further study.

Several clinical studies have also investigated <sup>211</sup>Atlabeled antibodies for RPT [\[69\]](#page-9-14). Notably, an older study of a 211At-labeled antitenascin antibody administered intracranially to patients bearing brain tumors demonstrated a favorable treatment response with minimal toxicity [[70](#page-9-15)]. More recently, a 12-year follow-up of a dose-fnding study in ovarian cancer patients treated with  $211$ At-labeled antibody fragment  $(F(ab')_2)$  targeting SLC34A2, a cell surface glycoprotein, was reported [\[71](#page-9-16)]. The conjugate exhibited low toxicity and demonstrated a therapeutic response in some patients. Other clinical trials evaluating  $211$ At-labeled antibodies focusing on multiple myeloma, recurrent, refractory acute lymphocytic leukemia, and myelodysplastic disorders are currently in progress (NCT04466475, NCT03670966).

The long-lived  $\alpha$ -emitter <sup>227</sup>Th has emerged as another option to pair with antibodies due to its ease of chelation and relatively high availability [\[72\]](#page-9-17). Preclinical studies of PSMA-targeting antibody conjugates with  $227$ Th have been promising, and a phase I clinical trial of one conjugate is ongoing (NCT03724747) [[73](#page-9-18)]. Preclinical investigation using this conjugate showed increased tumor uptake relative to healthy tissue, as well as dose-dependent tumor inhibition in both subcutaneous tumors and a metastatic model. Notably, the study showed potential synergy between the PSMA-targeted <sup>227</sup>Th RPT in conjunction with androgen receptor antagonists. <sup>227</sup>Th-antibody conjugates targeting mesothelin (NCT03507452) and CD22 (NCT02581878) are also in clinical trials [[73](#page-9-18), [74](#page-9-19)]. A known challenge of intact antibody-based radioimmunoconjugates is hematologic toxicity in already heavily pretreated patients, which may limit their translational potential. While true for both α- and β−- emitters, it may be more prominent in β−-emitting antibody radioconjugates due to the longer path length. Efforts to overcome these challenges include pre-targeting strategies and dose fractionation [[75\]](#page-9-20).

The growing body of research indicates that  $\alpha$ -emitters can make safe, efective therapeutics when conjugated to appropriate targeting molecules. Although no α-emitting radioconjugates have received clinical approval, their high potency and tunable molecular footprint make them excellent partners for targeting antibodies or small molecules. Based on convention,  $t^{213}$ Bi and <sup>212</sup>Pb may pair best with rapidly cleared smaller molecules, whereas  $^{225}$ Ac and  $^{227}$ Th may be better suited for antibodies, which clear much more slowly. Despite its relatively long half-life, 225Ac has proven efective in both small-molecule and antibody conjugates, but its high toxicity due to its long half-life and daughter nuclide redistribution remains a concern.

### **A clinical perspective: potential and challenges for α‑based RPT**

Currently, there is no consensus on the proper dose, timing, administration route, or ideal chemotherapeutic combinations with RPTs—and the best approach will likely be different for each RPT and disease presentation. Any of these variables can have a profound impact on the ultimate success of the therapy. Guidelines should be established, if possible, and commonalities may emerge across the various RPT agents as a class.

Studies have shown that administering several smaller doses of the RPT over an extended period of time (fractionation) can increase the treatments' efectiveness and decrease the off-target radiation toxicity  $[45, 76, 77]$  $[45, 76, 77]$  $[45, 76, 77]$  $[45, 76, 77]$  $[45, 76, 77]$  $[45, 76, 77]$ . This approach is intuitive, as most anticancer therapeutics are administered in multiple cycles to increase tolerability and minimize side effects while maintaining efficacy. External beam radiation is similarly administered. Potential challenges, however, include the logistics inherent to a multi-dose regimen. Many radionuclides, and  $\alpha$ -emitters in particular, are relatively scarce and are typically produced on a strict schedule. Thus, receiving frequent, regular shipments of radionuclides for patient treatment on a clinical use schedule can be challenging, especially for those with shorter half-lives. While having a six-cycle regimen is feasible for long-lived agents, such as  $^{223}$ RaCl<sub>2</sub>, which has already been FDA-approved and enjoys commercial/clinical success, it is critical that radionuclide supply is able to meet the demand for clinical trials of new α-emitting RPTs.

The administration route of RPTs can also have a profound impact on the treatment. While intravenous (IV) administration is common, it may not be ideal in all cases, especially for tumors within enclosed compartments. For example, injection of radioconjugates intraperitoneally (IP) or intracranially for tumors located in these regions may increase the tumor-absorbed dose relative to the off-target dose. In one notable, though small, clinical study, patients were treated with a  $[{}^{212}Pb]Pb$ -trastuzumab conjugate via IP administration [\[78](#page-9-23), [79\]](#page-10-0). The radioimmunoconjugate showed confnement to the peritoneal cavity, with only 6% of the conjugate observed elsewhere. Another study showed that a 213Bi-labeled radioconjugate targeting the NK-1 receptor was well-tolerated and efective in treating glioblastoma when administered intracranially [\[80,](#page-10-1) [81\]](#page-10-2).

Further considerations for  $\alpha$ -emitter-based RPTs include the potential for synergistic pairings with traditional chemotherapy or other targeted therapies. Because clinical studies of α-based RPTs are still few, ideal partners have not yet been established, although several promising combinations are being tested [[82,](#page-10-3) [83\]](#page-10-4). In theory, RPTs should synergize with any agents that increase their uptake or the tumor's susceptibility to DNA damage, such as platinum-based drugs and taxanes. Preclinical studies have shown that these agents may augment response to  $\alpha$ -therapies [[84,](#page-10-5) [85](#page-10-6)]. Pairing RPTs with novel radiosensitizers, such as PARP inhibitors, can greatly improve their efectiveness [[86](#page-10-7)]. Others have also shown that RPTs combine well with anti-angiogenic drugs and therapeutic antibodies [\[84\]](#page-10-5). Further analysis and careful experimentation are needed to identify ideal treatment partners, which may vary widely depending on the type, target expression profle, and stage of the cancer to be treated. In addition, high-quality, randomized controlled studies are critical to identify specifc patient populations to achieve the greatest beneft from RPT treatment and provide clear treatment indications.

### **The future of α‑therapy**

α-particle-based RPT represents a compelling therapeutic modality for disseminated cancers. Combination therapies and innovative dosing strategies have emerged recently that may greatly improve upon the prior iterations of α-RPTs. Also, as  $\alpha$ -emitting radionuclide production and distribution capacity increases, investigators can correspondingly increase our collective knowledge of this currently underexplored feld.

As mentioned earlier, one of the greatest potential clinical uses of  $\alpha$ -based RPT is as a salvage or second-line therapy in patients with disease refractory to other treatment. α-therapy has demonstrated ability to overcome cancer resistance. Its distinct mechanism of cellular toxicity and indiference to hypoxia or drug-transporter overexpression make it ideal for use in cancers with resistance to traditional treatments. Furthermore,  $\alpha$ -emitting conjugates can also be used in combination with  $β$ <sup>-</sup>-emitters partnered with the same targeting motif or as salvage therapy following treatment failure with the β<sup>-</sup>-emitting conjugate. Additionally,  $α$ -therapy could be combined with external beam radiation to doseescalate therapy to gross disease, while molecular RPT can address disseminated macroscopic or microscopic disease [\[87](#page-10-8), [88](#page-10-9)]. This approach is particularly compelling in the setting of oligometastatic disease [\[36\]](#page-8-11). Finally, combination with standard-of-care anticancer treatments (e.g., chemotherapy, anti-angiogenic therapy, kinase inhibitors, immune checkpoint blockade) or exploratory agents (e.g., DNA repair pathway inhibitors) must be properly investigated for use in conjunction with  $\alpha$ -RPT in order to find synergistic pairings, while remaining mindful of overlapping toxicities.

 Pre-targeting strategies may also enhance the RPT paradigm and preclinical results have shown effective  $^{225}$ Ac therapy using this approach [\[89\]](#page-10-10). Typically, the tumor-targeting moiety is administered, allowed to bind to the target with the excess cleared from the body prior to administration of the fast-clearing radioactive component, which can bind to the tumor-targeting moiety in vivo using several established methods [[75](#page-9-20), [90\]](#page-10-11). Pre-targeting has been evaluated in humans and has been shown to reduce the overall non-target radiation load on the patient while maintaining the high tumor radiation dose of traditional antibody therapy [[91](#page-10-12)].

Another potential avenue for expansion of the current α-RPT is the development of imaging methods that allow for visualization of the treatment.  $225$ Ac and  $223$ Ra both have characteristic photon emissions that may be imaged via SPECT during their decays, which can faciliate accurate absorbed dose estimates. While several recent studies have focused on imaging these nuclides in vivo, the relatively low administered activities of the nuclides and the long imaging times required for SPECT may ultimately make clinical imaging difficult. Perhaps an alternative strategy is to partner α-emitting nuclides with congeners or other isotopes for imaging. For instance,  $^{211}$ At may be combined with  $^{124}$ I (PET) or  $^{123}$ I (SPECT) for diagnostics imaging, and  $^{212}$ Pb may be partnered with <sup>203</sup>Pb for SPECT. The α-emitting  $149$  Tb might offer another interesting option since it can be imaged by PET [[92\]](#page-10-13). The difficult production and long-lived daughters of this nuclide have hindered clinical translation to date; however, improved production methods may increase the availability of  $149$  Tb in the near future [[31](#page-8-9)].

In addition to the importance of overcoming technical challenges, it is absolutely critical that we test the most favorable agents in prospective, randomized controlled trials for defnitive evaluation. Anecdotal and small singleinstitution series are important to demonstrate feasibility and bioplausibility; however, such studies are insufficient to move the feld forward. Fortunately, as level I evidence from RCTs accumulate for  $\beta$ <sup>-</sup>-emitting RPTs, we can use these as templates for testing  $\alpha$ -emitting RPTs. In addition, while it is intuitive that personalized dosimetry in patients treated with RPTs might prove benefcial for patient safety and therapeutic efficacy, as with external beam radiotherapy, prospective studies in patients are needed to test this hypothesis [\[93](#page-10-14)].

The improving quality and quantity of radionuclide production, combined with increasingly available tools to identify tumor-selective targets using next-generation sequencing, enable us to engineer diverse molecules specific to cancer targets of interest. These efforts have been further facilitated by the development of new radiochelates for alpha-emitting nuclides, which may provide safer, more easily manufactured conjugates. In addition, the marked ability of targeted  $\alpha$ -emitting agents to effectively kill cells, which may be enhanced with strategic drug combinations, coupled with no known mechanisms of resistance all suggest a promising future of  $\alpha$ -particle RPT. Now that these advances are in place, it is up to our community of multidisciplinary investigators to do the hard work to realize the full potential of this modality and expand the cancer therapy armamentarium for patients.

**Acknowledgements** Authors would like to thank Dr. Martin Brechbiel for critical comments and review of early versions of this manuscript.

**Funding** This work was supported by the Intramural Research Program of the NIH, NCI, Center for Cancer Research (ZIA BC 011800; ZIA BC 010891).

### **References**

- <span id="page-7-0"></span>1. Strosberg J, El-Haddad G, Wolin E, Hendifar A, Yao J, Chasen B, et al. Phase 3 trial of 177Lu-dotatate for midgut neuroendocrine tumors. N Engl J Med. 2017;376:125–35. [https://doi.org/10.1056/](https://doi.org/10.1056/NEJMoa1607427) [NEJMoa1607427](https://doi.org/10.1056/NEJMoa1607427).
- 2. Pryma DA, Chin BB, Noto RB, Dillon JS, Perkins S, Solnes L, et al. Efficacy and safety of high-specific-activity <sup>131</sup>I-MIBG therapy in patients with advanced pheochromocytoma or paraganglioma. J Nucl Med. 2019;60:623–30. [https://doi.org/10.2967/](https://doi.org/10.2967/jnumed.118.217463) [jnumed.118.217463.](https://doi.org/10.2967/jnumed.118.217463)
- <span id="page-7-1"></span>3. Parker C, Nilsson S, Heinrich D, Helle SI, O'Sullivan JM, Fossa SD, et al. Alpha emitter radium-223 and survival in metastatic prostate cancer. N Engl J Med. 2013;369:213–23. [https://doi.org/](https://doi.org/10.1056/NEJMoa1213755) [10.1056/NEJMoa1213755.](https://doi.org/10.1056/NEJMoa1213755)
- <span id="page-7-2"></span>4. Sartor O, de Bono J, Chi KN, Fizazi K, Herrmann K, Rahbar K, et al. Lutetium-177–PSMA-617 for Metastatic Castration-Resistant Prostate Cancer. N Engl J Med. 2021;NEJMoa2107322.
- <span id="page-7-3"></span>5. Sgouros G, Bodei L, McDevitt MR, Nedrow JR. Radiopharmaceutical therapy in cancer: clinical advances and challenges. Nat Rev Drug Discov. 2020;19:589–608. [https://doi.org/10.1038/](https://doi.org/10.1038/s41573-020-0073-9) [s41573-020-0073-9.](https://doi.org/10.1038/s41573-020-0073-9)
- <span id="page-7-4"></span>6. Baskar R, Dai J, Wenlong N, Yeo R, Yeoh KW. Biological response of cancer cells to radiation treatment. Front Mol Biosci. 2014;1:24. [https://doi.org/10.3389/fmolb.2014.00024.](https://doi.org/10.3389/fmolb.2014.00024)
- <span id="page-7-5"></span>7. Connell PP, Hellman S. Advances in radiotherapy and implications for the next century: a historical perspective. Cancer Res. 2009;69:383–92. [https://doi.org/10.1158/0008-5472.](https://doi.org/10.1158/0008-5472.CAN-07-6871) [CAN-07-6871](https://doi.org/10.1158/0008-5472.CAN-07-6871).
- <span id="page-7-6"></span>8. Walicka MA, Vaidyanathan G, Zalutsky MR, Adelstein SJ, Kassis AI. Survival and DNA damage in Chinese hamster V79 cells exposed to alpha particles emitted by DNA-incorporated astatine-211. Radiat Res. 1998;150:263–8.
- <span id="page-7-7"></span>9. Ku A, Facca VJ, Cai Z, Reilly RM. Auger electrons for cancer therapy - a review. EJNMMI Radiopharm Chem. 2019;4:27. <https://doi.org/10.1186/s41181-019-0075-2>.
- <span id="page-7-8"></span>10. Baidoo KE, Yong K, Brechbiel MW. Molecular pathways: targeted alpha-particle radiation therapy. Clin Cancer Res. 2013;19:530–7. [https://doi.org/10.1158/1078-0432.CCR-12-0298.](https://doi.org/10.1158/1078-0432.CCR-12-0298)
- <span id="page-7-9"></span>11. Goodhead DT. Initial events in the cellular efects of ionizing radiations: clustered damage in DNA. Int J Radiat Biol. 1994;65:7–17. <https://doi.org/10.1080/09553009414550021>.
- <span id="page-7-10"></span>12. Sutherland BM, Bennett PV, Sidorkina O, Laval J. Clustered damages and total lesions induced in DNA by ionizing radiation: oxidized bases and strand breaks. Biochemistry. 2000;39:8026–31. <https://doi.org/10.1021/bi9927989>.
- <span id="page-7-11"></span>13. Willers H, Azzoli CG, Santivasi WL, Xia F. Basic mechanisms of therapeutic resistance to radiation and chemotherapy in lung cancer. Cancer J. 2013;19:200–7. [https://doi.org/10.1097/PPO.](https://doi.org/10.1097/PPO.0b013e318292e4e3) [0b013e318292e4e3](https://doi.org/10.1097/PPO.0b013e318292e4e3).
- <span id="page-7-12"></span>14. Malamas AS, Gameiro SR, Knudson KM, Hodge JW. Sublethal exposure to alpha radiation  $(^{223}Ra$  dichloride) enhances various carcinomas' sensitivity to lysis by antigen-specifc cytotoxic T lymphocytes through calreticulin-mediated immunogenic modulation. Oncotarget. 2016;7:86937–47. [https://doi.org/10.18632/](https://doi.org/10.18632/oncotarget.13520) [oncotarget.13520.](https://doi.org/10.18632/oncotarget.13520)
- <span id="page-7-13"></span>15. Marcu L, Bezak E, Allen BJ. Global comparison of targeted alpha vs targeted beta therapy for cancer: in vitro, in vivo and clinical trials. Crit Rev Oncol Hematol. 2018;123:7–20. [https://doi.org/](https://doi.org/10.1016/j.critrevonc.2018.01.001) [10.1016/j.critrevonc.2018.01.001.](https://doi.org/10.1016/j.critrevonc.2018.01.001)
- <span id="page-7-14"></span>16. Haro KJ, Scott AC, Scheinberg DA. Mechanisms of resistance to high and low linear energy transfer radiation in myeloid leukemia cells. Blood. 2012;120:2087–97. [https://doi.org/10.1182/](https://doi.org/10.1182/blood-2012-01-404509) [blood-2012-01-404509.](https://doi.org/10.1182/blood-2012-01-404509)
- <span id="page-7-15"></span>17. Yard BD, Gopal P, Bannik K, Siemeister G, Hagemann UB, Abazeed ME. Cellular and genetic determinants of the sensitivity of cancer to alpha-particle irradiation. Cancer Res. 2019;79:5640– 51. [https://doi.org/10.1158/0008-5472.CAN-19-0859.](https://doi.org/10.1158/0008-5472.CAN-19-0859)
- <span id="page-7-16"></span>18. Ballal S, Yadav MP, Bal C, Sahoo RK, Tripathi M. Broadening horizons with 225Ac-DOTATATE targeted alpha therapy for gastroenteropancreatic neuroendocrine tumour patients stable or refractory to <sup>177</sup>Lu-DOTATATE PRRT: first clinical experience on the efficacy and safety. Eur J Nucl Med Mol Imaging. 2020;47:934–46. [https://doi.org/10.1007/s00259-019-04567-2.](https://doi.org/10.1007/s00259-019-04567-2)
- <span id="page-7-17"></span>19. Kratochwil C, Bruchertseifer F, Giesel FL, Weis M, Verburg FA, Mottaghy F, et al. <sup>225</sup>Ac-PSMA-617 for PSMA-targeted α-radiation therapy of metastatic castration-resistant prostate cancer. J Nucl Med. 2016;57:1941–4. [https://doi.org/10.2967/jnumed.](https://doi.org/10.2967/jnumed.116.178673) [116.178673.](https://doi.org/10.2967/jnumed.116.178673)
- <span id="page-7-18"></span>20. Yadav MP, Ballal S, Sahoo RK, Tripathi M, Seth A, Bal C. Efficacy and safety of 225Ac-PSMA-617 targeted alpha therapy in metastatic castration-resistant prostate cancer patients. Theranostics. 2020;10:9364–77.<https://doi.org/10.7150/thno.48107>.
- <span id="page-7-19"></span>21. Thompson LH. Recognition, signaling, and repair of DNA double-strand breaks produced by ionizing radiation in mammalian cells: the molecular choreography. Mutat Res. 2012;751:158–246. [https://doi.org/10.1016/j.mrrev.2012.06.002.](https://doi.org/10.1016/j.mrrev.2012.06.002)
- <span id="page-7-20"></span>22. Bannik K, Madas B, Jarzombek M, Sutter A, Siemeister G, Mumberg D, et al. Radiobiological effects of the alpha emitter Ra-223 on tumor cells. Sci Rep. 2019;9:18489. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-019-54884-7) [s41598-019-54884-7](https://doi.org/10.1038/s41598-019-54884-7).
- <span id="page-7-21"></span>23. Yong KJ, Milenic DE, Baidoo KE, Brechbiel MW. 212Pb-radioimmunotherapy induces G(2) cell-cycle arrest and delays DNA damage repair in tumor xenografts in a model for disseminated

intraperitoneal disease. Mol Cancer Ther. 2012;11:639–48. <https://doi.org/10.1158/1535-7163.MCT-11-0671>.

- <span id="page-8-0"></span>24. Yong KJ, Milenic DE, Baidoo KE, Brechbiel MW. Sensitization of tumor to 212Pb radioimmunotherapy by gemcitabine involves initial abrogation of G2 arrest and blocked DNA damage repair by interference with Rad51. Int J Radiat Oncol Biol Phys. 2013;85:1119–26. [https://doi.org/10.1016/j.ijrobp.2012.09.015.](https://doi.org/10.1016/j.ijrobp.2012.09.015)
- <span id="page-8-1"></span>25. Li M, Sagastume EA, Lee D, McAlister D, DeGrafenreid AJ, Olewine KR, et al. <sup>203/212</sup>Pb theranostic radiopharmaceuticals for image-guided radionuclide therapy for cancer. Curr Med Chem. 2020;27:7003–31. [https://doi.org/10.2174/092986732799920](https://doi.org/10.2174/0929867327999200727190423) [0727190423.](https://doi.org/10.2174/0929867327999200727190423)
- <span id="page-8-2"></span>26. Larsson E, Brolin G, Cleton A, Ohlsson T, Linden O, Hindorf C. Feasibility of thorium-227/radium-223 gamma-camera imaging during radionuclide therapy. Cancer Biother Radiopharm. 2020;35:540–8. [https://doi.org/10.1089/cbr.2019.3516.](https://doi.org/10.1089/cbr.2019.3516)
- 27. Usmani S, Rasheed R, Al Kandari F, Marafi F, Naqvi SAR. <sup>225</sup>Ac Prostate-specifc membrane antigen posttherapy alpha imaging: comparing 2 and 3 photopeaks. Clin Nucl Med. 2019;44:401–3. [https://doi.org/10.1097/RLU.0000000000002525.](https://doi.org/10.1097/RLU.0000000000002525)
- <span id="page-8-3"></span>28. Abou DS, Rittenbach A, Tomlinson RE, Finley PA, Tsui B, Simons BW, et al. Preclinical single photon emission computed tomography of alpha particle-emitting radium-223. Cancer Biother Radiopharm. 2020;35:520–9. [https://doi.org/10.1089/cbr.](https://doi.org/10.1089/cbr.2019.3308) [2019.3308.](https://doi.org/10.1089/cbr.2019.3308)
- <span id="page-8-4"></span>29. Phelps TE, Roy J, Green MV, Seidel J, Baidoo KE, Adler S, et al. Sodium fuoride-18 and radium-223 dichloride uptake colocalize in osteoblastic mouse xenograft tumors. Cancer Biother Radiopharm. 2021;36:133-42. <https://doi.org/10.1089/cbr.2020.4068>.
- <span id="page-8-5"></span>30. Murray I, Chittenden SJ, Denis-Bacelar AM, Hindorf C, Parker CC, Chua S, et al. The potential of  $223$ Ra and  $18$ F-fluoride imaging to predict bone lesion response to treatment with  $^{223}$ Radichloride in castration-resistant prostate cancer. Eur J Nucl Med Mol Imaging. 2017;44:1832–44. [https://doi.org/10.1007/](https://doi.org/10.1007/s00259-017-3744-y) [s00259-017-3744-y](https://doi.org/10.1007/s00259-017-3744-y).
- <span id="page-8-9"></span>31. Moiseeva AN, Aliev RA, Unezhev VN, Zagryadskiy VA, Latushkin ST, Aksenov NV, et al. Cross section measurements of 151Eu((3)He,5n) reaction: new opportunities for medical alpha emitter 149Tb production. Sci Rep. 2020;10:508. [https://doi.org/](https://doi.org/10.1038/s41598-020-57436-6) [10.1038/s41598-020-57436-6](https://doi.org/10.1038/s41598-020-57436-6).
- <span id="page-8-6"></span>32. Dronsfeld A, Ellis P. Radium – a key element in early cancer treatment. Educ Chem. 2011:56–9. [https://www.rsc.org/images/](https://www.rsc.org/images/Dronsfield_tcm18-200827.pdf) [Dronsfeld\\_tcm18-200827.pdf](https://www.rsc.org/images/Dronsfield_tcm18-200827.pdf).
- <span id="page-8-7"></span>33. Wissing MD, van Leeuwen FW, van der Pluijm G, Gelderblom H. Radium-223 chloride: Extending life in prostate cancer patients by treating bone metastases. Clin Cancer Res. 2013;19:5822–7. [https://doi.org/10.1158/1078-0432.CCR-13-1896.](https://doi.org/10.1158/1078-0432.CCR-13-1896)
- <span id="page-8-8"></span>34. Hoskin P, Sartor O, O'Sullivan JM, Johannessen DC, Helle SI, Logue J, et al. Efficacy and safety of radium-223 dichloride in patients with castration-resistant prostate cancer and symptomatic bone metastases, with or without previous docetaxel use: a prespecifed subgroup analysis from the randomised, double-blind, phase 3 ALSYMPCA trial. Lancet Oncol. 2014;15:1397–406. [https://doi.org/10.1016/S1470-2045\(14\)70474-7](https://doi.org/10.1016/S1470-2045(14)70474-7).
- <span id="page-8-10"></span>35. Morris MJ, Loriot Y, Sweeney CJ, Fizazi K, Ryan CJ, Shevrin DH, et al. Radium-223 in combination with docetaxel in patients with castration-resistant prostate cancer and bone metastases: a phase 1 dose escalation/randomised phase 2a trial. Eur J Cancer. 2019;114:107–16. [https://doi.org/10.1016/j.ejca.2019.04.007.](https://doi.org/10.1016/j.ejca.2019.04.007)
- <span id="page-8-11"></span>36. Hasan H, Deek MP, Phillips R, Hobbs RF, Malek R, Radwan N, et al. A phase II randomized trial of radium-223 dichloride and SABR versus SABR for oligometastatic prostate cancers (RAVENS). BMC Cancer. 2020;20:492. [https://doi.org/10.1186/](https://doi.org/10.1186/s12885-020-07000-2) [s12885-020-07000-2](https://doi.org/10.1186/s12885-020-07000-2).
- <span id="page-8-12"></span>37. Morris MJ, Corey E, Guise TA, Gulley JL, Kevin Kelly W, Quinn DI, et al. Radium-223 mechanism of action: implications for use

in treatment combinations. Nat Rev Urol. 2019;16:745–56. [https://](https://doi.org/10.1038/s41585-019-0251-x) [doi.org/10.1038/s41585-019-0251-x](https://doi.org/10.1038/s41585-019-0251-x).

- <span id="page-8-13"></span>38. Smith M, Parker C, Saad F, Miller K, Tombal B, Ng QS, et al. Addition of radium-223 to abiraterone acetate and prednisone or prednisolone in patients with castration-resistant prostate cancer and bone metastases (ERA 223): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet Oncol. 2019;20:408–19. [https://doi.org/10.1016/S1470-2045\(18\)30860-X](https://doi.org/10.1016/S1470-2045(18)30860-X).
- <span id="page-8-14"></span>39. Abou DS, Thiele NA, Gutsche NT, Villmer A, Zhang H, Woods JJ, Baidoo KE, Escorcia FE, Wilson JJ, Thorek DL. Towards the stable chelation of radium for biomedical applications with an 18-membered macrocyclic ligand. Chem Sci. 2021;12:3733–42. [https://doi.org/10.1039/D0SC06867E.](https://doi.org/10.1039/D0SC06867E)
- <span id="page-8-15"></span>40. Scheinberg DA, McDevitt MR. Actinium-225 in targeted alphaparticle therapeutic applications. Curr Radiopharm. 2011;4:306– 20. [https://doi.org/10.2174/1874471011104040306.](https://doi.org/10.2174/1874471011104040306)
- 41. Kim YS, Brechbiel MW. An overview of targeted alpha therapy. Tumour Biol. 2012;33:573–90. [https://doi.org/10.1007/](https://doi.org/10.1007/s13277-011-0286-y) [s13277-011-0286-y](https://doi.org/10.1007/s13277-011-0286-y).
- 42. Guerra Liberal FDC, O'Sullivan JM, McMahon SJ, Prise KM. Targeted alpha therapy: current clinical applications. Cancer Biother Radiopharm. 2020;35:404–17. [https://doi.org/10.1089/cbr.](https://doi.org/10.1089/cbr.2020.3576.t) [2020.3576.t](https://doi.org/10.1089/cbr.2020.3576.t).
- 43. McDevitt MR, Sgouros G, Sofou S. Targeted and nontargeted alpha-particle therapies. Annu Rev Biomed Eng. 2018;20:73–93. <https://doi.org/10.1146/annurev-bioeng-062117-120931>.
- <span id="page-8-16"></span>44. Mulford DA, Scheinberg DA, Jurcic JG. The promise of targeted {alpha}-particle therapy. J Nucl Med. 2005;46(Suppl 1):199S-204S.
- <span id="page-8-17"></span>45. Escorcia FE, Houghton JL, Abdel-Atti D, Pereira PR, Cho A, Gutsche NT, et al. ImmunoPET predicts response to met-targeted radioligand therapy in models of pancreatic cancer resistant to Met kinase inhibitors. Theranostics. 2020;10:151–65. [https://doi.](https://doi.org/10.7150/thno.37098) [org/10.7150/thno.37098](https://doi.org/10.7150/thno.37098).
- <span id="page-8-18"></span>46. Navalkissoor S, Grossman A. Targeted alpha particle therapy for neuroendocrine tumours: the next generation of peptide receptor radionuclide therapy. Neuroendocrinology. 2019;108:256–64. [https://doi.org/10.1159/000494760.](https://doi.org/10.1159/000494760)
- <span id="page-8-19"></span>47. Kunikowska J, Krolicki L. Targeted alpha-emitter therapy of neuroendocrine tumors. Semin Nucl Med. 2020;50:171–6. [https://doi.](https://doi.org/10.1053/j.semnuclmed.2019.11.003) [org/10.1053/j.semnuclmed.2019.11.003](https://doi.org/10.1053/j.semnuclmed.2019.11.003).
- <span id="page-8-20"></span>48. Miederer M, Henriksen G, Alke A, Mossbrugger I, Quintanilla-Martinez L, Senekowitsch-Schmidtke R, et al. Preclinical evaluation of the alpha-particle generator nuclide  $^{225}$ Ac for somatostatin receptor radiotherapy of neuroendocrine tumors. Clin Cancer Res. 2008;14:3555–61. [https://doi.org/10.1158/1078-0432.](https://doi.org/10.1158/1078-0432.CCR-07-4647) [CCR-07-4647](https://doi.org/10.1158/1078-0432.CCR-07-4647).
- <span id="page-8-21"></span>49. Bal C, Yadav M, Ballal S, Tripathi M. Safety and therapeutic efficacy of  $225$  Ac-DOTATATE targeted alpha therapy in metastatic gastroenteropancreatic neuroendocrine tumors stable or refractory to 177Lu-DOTATATE PRRT. J Nucl Med. 2020;61:416.
- <span id="page-8-22"></span>50. Kratochwil C, Giesel FL, Bruchertseifer F, Mier W, Apostolidis C, Boll R, et al. 213Bi-DOTATOC receptor-targeted alpha-radionuclide therapy induces remission in neuroendocrine tumours refractory to beta radiation: a frst-in-human experience. Eur J Nucl Med Mol Imaging. 2014;41:2106–19. [https://doi.org/10.](https://doi.org/10.1007/s00259-014-2857-9) [1007/s00259-014-2857-9](https://doi.org/10.1007/s00259-014-2857-9).
- <span id="page-8-23"></span>51. Stallons TAR, Saidi A, Tworowska I, Delpassand ES, Torgue JJ. Preclinical Investigation of 212Pb-DOTAMTATE for peptide receptor radionuclide therapy in a neuroendocrine tumor model. Mol Cancer Ther. 2019;18:1012–21. [https://doi.org/10.1158/](https://doi.org/10.1158/1535-7163.MCT-18-1103) [1535-7163.MCT-18-1103](https://doi.org/10.1158/1535-7163.MCT-18-1103).
- <span id="page-8-24"></span>52. Delpassand E, Tworowska I, Shanoon F, Nunez R, Flores L II, Muzammil A, et al. First clinical experience using targeted alpha-emitter therapy with <sup>212</sup>Pb-DOTAMTATE (AlphaMedix<sup>TM</sup>

in patients with SSTR(+) neuroendocrine tumors. J Nucl Med. 2019;60:559.

- <span id="page-9-0"></span>53. Filippi L, Chiaravalloti A, Schillaci O, Bagni O. The potential of PSMA-targeted alpha therapy in the management of prostate cancer. Expert Rev Anticancer Ther. 2020;20:823–9. [https://doi.](https://doi.org/10.1080/14737140.2020.1814151) [org/10.1080/14737140.2020.1814151.](https://doi.org/10.1080/14737140.2020.1814151)
- <span id="page-9-1"></span>54. Dos Santos JC, Schafer M, Bauder-Wust U, Lehnert W, Leotta K, Morgenstern A, et al. Development and dosimetry of <sup>203</sup>Pb/<sup>212</sup>Pblabelled PSMA ligands: bringing "the lead" into PSMA-targeted alpha therapy? Eur J Nucl Med Mol Imaging. 2019;46:1081–91. <https://doi.org/10.1007/s00259-018-4220-z>.
- <span id="page-9-2"></span>55. Sathekge M, Knoesen O, Meckel M, Modiselle M, Vorster M, Marx S. 213Bi-PSMA-617 targeted alpha-radionuclide therapy in metastatic castration-resistant prostate cancer. Eur J Nucl Med Mol Imaging. 2017;44:1099–100. [https://doi.org/10.1007/](https://doi.org/10.1007/s00259-017-3657-9) [s00259-017-3657-9.](https://doi.org/10.1007/s00259-017-3657-9)
- <span id="page-9-3"></span>56. Lo Nigro C, Macagno M, Sangiolo D, Bertolaccini L, Aglietta M, Merlano MC. NK-mediated antibody-dependent cell-mediated cytotoxicity in solid tumors: biological evidence and clinical perspectives. Ann Transl Med. 2019;7:105. [https://doi.org/10.21037/](https://doi.org/10.21037/atm.2019.01.42) [atm.2019.01.42.](https://doi.org/10.21037/atm.2019.01.42)
- <span id="page-9-4"></span>57. Rizzieri D. Zevalin((R)) (ibritumomab tiuxetan): after more than a decade of treatment experience, what have we learned? Crit Rev Oncol Hematol. 2016;105:5–17. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.critrevonc.2016.07.008) [critrevonc.2016.07.008.](https://doi.org/10.1016/j.critrevonc.2016.07.008)
- Schaefer NG, Huang P, Buchanan JW, Wahl RL. Radioimmunotherapy in non-Hodgkin lymphoma: opinions of nuclear medicine physicians and radiation oncologists. J Nucl Med. 2011;52:830–8. [https://doi.org/10.2967/jnumed.110.085589.](https://doi.org/10.2967/jnumed.110.085589)
- 59. Kaminski MS, Zelenetz AD, Press OW, Saleh M, Leonard J, Fehrenbacher L, et al. Pivotal study of iodine I 131 tositumomab for chemotherapy-refractory low-grade or transformed low-grade B-cell non-Hodgkin's lymphomas. J Clin Oncol. 2001;19:3918– 28. <https://doi.org/10.1200/JCO.2001.19.19.3918>.
- <span id="page-9-5"></span>60. Morschhauser F, Radford J, Van Hoof A, Vitolo U, Soubeyran P, Tilly H, et al. Phase III trial of consolidation therapy with yttrium-90-ibritumomab tiuxetan compared with no additional therapy after frst remission in advanced follicular lymphoma. J Clin Oncol. 2008;26:5156–64. [https://doi.org/10.1200/JCO.2008.](https://doi.org/10.1200/JCO.2008.17.2015) [17.2015.](https://doi.org/10.1200/JCO.2008.17.2015)
- <span id="page-9-6"></span>61. Sgouros G, Ballangrud AM, Jurcic JG, McDevitt MR, Humm JL, Erdi YE, et al. Pharmacokinetics and dosimetry of an alphaparticle emitter labeled antibody: <sup>213</sup>Bi-HuM195 (anti-CD33) in patients with leukemia. J Nucl Med. 1999;40:1935–46.
- <span id="page-9-7"></span>62. Jurcic JG, Larson SM, Sgouros G, McDevitt MR, Finn RD, Divgi CR, et al. Targeted alpha particle immunotherapy for myeloid leukemia. Blood. 2002;100:1233–9.
- <span id="page-9-8"></span>63. Friesen C, Roscher M, Hormann I, Leib O, Marx S, Moreno J, et al. Anti-CD33-antibodies labelled with the alpha-emitter Bismuth-213 kill CD33-positive acute myeloid leukaemia cells specifcally by activation of caspases and break radio- and chemoresistance by inhibition of the anti-apoptotic proteins X-linked inhibitor of apoptosis protein and B-cell lymphoma-extra large. Eur J Cancer. 2013;49:2542–54. [https://doi.org/10.1016/j.ejca.](https://doi.org/10.1016/j.ejca.2013.04.008) [2013.04.008.](https://doi.org/10.1016/j.ejca.2013.04.008)
- <span id="page-9-9"></span>64. Jurcic JG. Clinical studies with bismuth-213 and actinium-225 for hematologic malignancies. Curr Radiopharm. 2018;11:192– 9. <https://doi.org/10.2174/1874471011666180525102814>.
- <span id="page-9-10"></span>65. Rosenblat TL, McDevitt MR, Pandit-Taskar N, Carrasquillo JA, Chanel S, Frattini MG, et al. Phase I trial of the targeted alphaparticle nano-generator actinium-225 (<sup>225</sup>Ac)-HuM195 (Anti-CD33) in acute myeloid leukemia (AML). Blood. 2007;110:910. <https://doi.org/10.1182/blood.V110.11.910.910>.
- <span id="page-9-11"></span>66. Jurcic JG, Rosenblat TL, McDevitt MR, Pandit-Taskar N, Carrasquillo JA, Chanel SM, Zikaras K, Frattini MG, Maslak PM, Cicic D, Larson SM, Scheinberg DA. Targeted alpha-particle

 $\circled{2}$  Springer

nano-generator actinium-225 ( $^{225}$ Ac)-lintuzumab (anti-CD33) in acute myeloid leukemia (AML), clinical lymphoma myeloma and leukemia. 2013;13:S379-S80. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.clml.2013.07.088) [clml.2013.07.088](https://doi.org/10.1016/j.clml.2013.07.088).

- <span id="page-9-12"></span>67. Allen BJ, Singla AA, Rizvi SM, Graham P, Bruchertseifer F, Apostolidis C, et al. Analysis of patient survival in a Phase I trial of systemic targeted alpha-therapy for metastatic melanoma. Immunotherapy. 2011;3:1041–50. [https://doi.org/10.](https://doi.org/10.2217/imt.11.97) [2217/imt.11.97](https://doi.org/10.2217/imt.11.97).
- <span id="page-9-13"></span>68. Autenrieth ME, Seidl C, Bruchertseifer F, Horn T, Kurtz F, Feuerecker B, et al. Treatment of carcinoma in situ of the urinary bladder with an alpha-emitter immunoconjugate targeting the epidermal growth factor receptor: a pilot study. Eur J Nucl Med Mol Imaging. 2018;45:1364–71. [https://doi.org/10.1007/](https://doi.org/10.1007/s00259-018-4003-6) [s00259-018-4003-6.](https://doi.org/10.1007/s00259-018-4003-6)
- <span id="page-9-14"></span>69. Lindegren S, Albertsson P, Back T, Jensen H, Palm S, Aneheim E. Realizing clinical trials with astatine-211: the chemistry infrastructure. Cancer Biother Radiopharm. 2020;35:425–36. [https://doi.org/10.1089/cbr.2019.3055.](https://doi.org/10.1089/cbr.2019.3055)
- <span id="page-9-15"></span>70. Zalutsky MR, Reardon DA, Akabani G, Coleman RE, Friedman AH, Friedman HS, et al. Clinical experience with alpha-particle emitting <sup>211</sup>At: treatment of recurrent brain tumor patients with <sup>211</sup>At-labeled chimeric antitenascin monoclonal antibody 81C6. J Nucl Med. 2008;49:30–8. [https://doi.org/10.2967/jnumed.107.](https://doi.org/10.2967/jnumed.107.046938) [046938.](https://doi.org/10.2967/jnumed.107.046938)
- <span id="page-9-16"></span>71. Hallqvist A, Bergmark K, Back T, Andersson H, Dahm-Kahler P, Johansson M, et al. Intraperitoneal alpha-emitting radioimmunotherapy with 211At in relapsed ovarian cancer: long-term follow-up with individual absorbed dose estimations. J Nucl Med. 2019;60:1073–9. [https://doi.org/10.2967/jnumed.118.](https://doi.org/10.2967/jnumed.118.220384) [220384.](https://doi.org/10.2967/jnumed.118.220384)
- <span id="page-9-17"></span>72. Frantellizzi V, Cosma L, Brunotti G, Pani A, Spanu A, Nuvoli S, et al. Targeted alpha therapy with thorium-227. Cancer Biother Radiopharm. 2020;35:437–45. [https://doi.org/10.1089/cbr.2019.](https://doi.org/10.1089/cbr.2019.3105) [3105](https://doi.org/10.1089/cbr.2019.3105).
- <span id="page-9-18"></span>73. Hammer S, Hagemann UB, Zitzmann-Kolbe S, Larsen A, Ellingsen C, Geraudie S, et al. Preclinical efficacy of a PSMA-targeted thorium-227 conjugate (PSMA-TTC), a targeted alpha therapy for prostate cancer. Clin Cancer Res. 2020;26:1985–96. [https://doi.](https://doi.org/10.1158/1078-0432.CCR-19-2268) [org/10.1158/1078-0432.CCR-19-2268.](https://doi.org/10.1158/1078-0432.CCR-19-2268)
- <span id="page-9-19"></span>74. Hagemann UB, Ellingsen C, Schuhmacher J, Kristian A, Mobergslien A, Cruciani V, et al. Mesothelin-Targeted Thorium-227 Conjugate (MSLN-TTC): Preclinical evaluation of a new targeted alpha therapy for mesothelin-positive cancers. Clin Cancer Res. 2019;25:4723–34. [https://doi.org/10.1158/1078-0432.](https://doi.org/10.1158/1078-0432.CCR-18-3476) [CCR-18-3476](https://doi.org/10.1158/1078-0432.CCR-18-3476).
- <span id="page-9-20"></span>75. Verhoeven M, Seimbille Y, Dalm SU. Therapeutic applications of pretargeting. Pharmaceutics. 2019;11. [https://doi.org/10.3390/](https://doi.org/10.3390/pharmaceutics11090434) [pharmaceutics11090434](https://doi.org/10.3390/pharmaceutics11090434).
- <span id="page-9-21"></span>76. Tagawa ST, Vallabhajosula S, Christos PJ, Jhanwar YS, Batra JS, Lam L, et al. Phase 1/2 study of fractionated dose lutetium-177-labeled anti-prostate-specifc membrane antigen monoclonal antibody J591 (<sup>177</sup>Lu-J591) for metastatic castration-resistant prostate cancer. Cancer. 2019;125:2561–9. [https://doi.org/10.1002/cncr.](https://doi.org/10.1002/cncr.32072) [32072.](https://doi.org/10.1002/cncr.32072)
- <span id="page-9-22"></span>77. Milenic DE, Garmestani K, Brady ED, Albert PS, Ma D, Abdulla A, et al. Alpha-particle radioimmunotherapy of disseminated peritoneal disease using a <sup>212</sup>Pb-labeled radioimmunoconjugate targeting HER2. Cancer Biother Radiopharm. 2005;20:557–68. [https://](https://doi.org/10.1089/cbr.2005.20.557) [doi.org/10.1089/cbr.2005.20.557](https://doi.org/10.1089/cbr.2005.20.557).
- <span id="page-9-23"></span>78. Meredith RF, Torgue J, Azure MT, Shen S, Saddekni S, Banaga E, et al. Pharmacokinetics and imaging of 212Pb-TCMC-trastuzumab after intraperitoneal administration in ovarian cancer patients. Cancer Biother Radiopharm. 2014;29:12–7. [https://doi.org/10.](https://doi.org/10.1089/cbr.2013.1531) [1089/cbr.2013.1531.](https://doi.org/10.1089/cbr.2013.1531)
- <span id="page-10-0"></span>79. Meredith RF, Torgue JJ, Rozgaja TA, Banaga EP, Bunch PW, Alvarez RD, et al. Safety and outcome measures of frst-in-human intraperitoneal alpha radioimmunotherapy with 212Pb-TCMC-trastuzumab. Am J Clin Oncol. 2018;41:716–21. [https://doi.org/10.](https://doi.org/10.1097/COC.0000000000000353) [1097/COC.0000000000000353.](https://doi.org/10.1097/COC.0000000000000353)
- <span id="page-10-1"></span>80. Krolicki L, Bruchertseifer F, Kunikowska J, Koziara H, Krolicki B, Jakucinski M, et al. Safety and efficacy of targeted alpha therapy with 213Bi-DOTA-substance P in recurrent glioblastoma. Eur J Nucl Med Mol Imaging. 2019;46:614–22. [https://doi.org/10.1007/](https://doi.org/10.1007/s00259-018-4225-7) [s00259-018-4225-7.](https://doi.org/10.1007/s00259-018-4225-7)
- <span id="page-10-2"></span>81. Krolicki L, Bruchertseifer F, Kunikowska J, Koziara H, Krolicki B, Jakucinski M, et al. Prolonged survival in secondary glioblastoma following local injection of targeted alpha therapy with <sup>213</sup>Bi-substance P analogue. Eur J Nucl Med Mol Imaging. 2018;45:1636–44. [https://doi.org/10.1007/s00259-018-4015-2.](https://doi.org/10.1007/s00259-018-4015-2)
- <span id="page-10-3"></span>82. Zaheer J, Kim H, Lee YJ, Kim JS, Lim SM. Combination radioimmunotherapy strategies for solid tumors. Int J Mol Sci. 2019;20. <https://doi.org/10.3390/ijms20225579>.
- <span id="page-10-4"></span>83. Ahmad SS, Crittenden MR, Tran PT, Kluetz PG, Blumenthal GM, Bulbeck H, et al. Clinical development of novel drug-radiotherapy combinations. Clin Cancer Res. 2019;25:1455–61. [https://doi.org/](https://doi.org/10.1158/1078-0432.CCR-18-2466) [10.1158/1078-0432.CCR-18-2466](https://doi.org/10.1158/1078-0432.CCR-18-2466).
- <span id="page-10-5"></span>84. Yong KJ, Milenic DE, Baidoo KE, Brechbiel MW. 212Pb-radioimmunotherapy potentiates paclitaxel-induced cell killing efficacy by perturbing the mitotic spindle checkpoint. Br J Cancer. 2013;108:2013–20.<https://doi.org/10.1038/bjc.2013.189>.
- <span id="page-10-6"></span>85. Milenic DE, Baidoo KE, Shih JH, Wong KJ, Brechbiel MW. Evaluation of platinum chemotherapy in combination with HER2 targeted alpha-particle radiation. Cancer Biother Radiopharm. 2013;28:441–9. [https://doi.org/10.1089/cbr.2012.1423.](https://doi.org/10.1089/cbr.2012.1423)
- <span id="page-10-7"></span>86. Cesaire M, Thariat J, Candeias SM, Stefan D, Saintigny Y, Chevalier F. Combining PARP inhibition, radiation, and immunotherapy: a possible strategy to improve the treatment of cancer? Int J Mol Sci. 2018;19.<https://doi.org/10.3390/ijms19123793>.
- <span id="page-10-8"></span>87. Tafreshi NK, Doligalski ML, Tichacek CJ, Pandya DN, Budzevich MM, El-Haddad G, et al. Development of targeted alpha particle therapy for solid tumors. Molecules. 2019;24. [https://doi.org/10.](https://doi.org/10.3390/molecules24234314) [3390/molecules24234314.](https://doi.org/10.3390/molecules24234314)
- <span id="page-10-9"></span>88. Anderson PM, Scott J, Parsai S, Zahler S, Worley S, Shrikanthan S, et al. 223-Radium for metastatic osteosarcoma: combination therapy with other agents and external beam radiotherapy. ESMO Open. 2020;5. [https://doi.org/10.1136/esmoopen-2019-000635.](https://doi.org/10.1136/esmoopen-2019-000635)
- <span id="page-10-10"></span>89. Poty S, Carter LM, Mandleywala K, Membreno R, Abdel-Atti D, Ragupathi A, et al. Leveraging bioorthogonal click chemistry to improve 225Ac-radioimmunotherapy of pancreatic ductal adenocarcinoma. Clin Cancer Res. 2019;25:868–80. [https://doi.org/10.](https://doi.org/10.1158/1078-0432.CCR-18-1650) [1158/1078-0432.CCR-18-1650](https://doi.org/10.1158/1078-0432.CCR-18-1650).
- <span id="page-10-11"></span>90. Patra M, Zarschler K, Pietzsch HJ, Stephan H, Gasser G. New insights into the pretargeting approach to image and treat tumours. Chem Soc Rev. 2016;45:6415–31. [https://doi.org/10.1039/c5cs0](https://doi.org/10.1039/c5cs00784d) [0784d.](https://doi.org/10.1039/c5cs00784d)
- <span id="page-10-12"></span>91. Forero A, Weiden PL, Vose JM, Knox SJ, LoBuglio AF, Hankins J, et al. Phase 1 trial of a novel anti-CD20 fusion protein in pretargeted radioimmunotherapy for B-cell non-Hodgkin lymphoma. Blood. 2004;104:227–36. [https://doi.org/10.1182/](https://doi.org/10.1182/blood-2003-09-3284) [blood-2003-09-3284](https://doi.org/10.1182/blood-2003-09-3284).
- <span id="page-10-13"></span>92. Muller C, Vermeulen C, Koster U, Johnston K, Turler A, Schibli R, et al. Alpha-PET with terbium-149: evidence and perspectives for radiotheragnostics. EJNMMI Radiopharm Chem. 2017;1:5. <https://doi.org/10.1186/s41181-016-0008-2>.
- <span id="page-10-14"></span>93. St James S, Bednarz B, Benedict S, Buchsbaum JC, Dewaraja Y, Frey E, et al. Current status of radiopharmaceutical therapy. Int J Radiat Oncol Biol Phys. 2021;109:891–901. [https://doi.org/10.](https://doi.org/10.1016/j.ijrobp.2020.08.035) [1016/j.ijrobp.2020.08.035.](https://doi.org/10.1016/j.ijrobp.2020.08.035)

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.