ORIGINAL ARTICLE



Quantitative [¹⁸F]florbetapir PET/CT may identify lung involvement in patients with systemic AL amyloidosis

Yiu Ming Khor¹ · Sarah Cuddy² · Hendrik J. Harms¹ · Marie F. Kijewski¹ · Mi-Ae Park¹ · Matthew Robertson¹ · Hyewon Hyun¹ · Marcelo F. Di Carli¹ · Giada Bianchi³ · Heather Landau⁴ · Andrew Yee⁵ · Vaishali Sanchorawala⁶ · Frederick L. Ruberg⁶ · Ronglih Liao⁷ · John Berk⁶ · Rodney H. Falk² · Sharmila Dorbala^{1,2}

Received: 4 June 2019 / Accepted: 18 November 2019 / Published online: 5 December 2019 \odot Springer-Verlag GmbH Germany, part of Springer Nature 2019

Abstract

Purpose The clinical diagnosis of pulmonary involvement in individuals with systemic AL amyloidosis remains challenging. $[^{18}F]$ florbetapir imaging has previously identified AL amyloid deposits in the heart and extra-cardiac organs. The aim of this study is to determine quantitative $[^{18}F]$ florbetapir pulmonary kinetics to identify pulmonary involvement in individuals with systemic AL amyloidosis.

Methods We prospectively enrolled 58 subjects with biopsy-proven AL amyloidosis and 9 control subjects (5 without amyloidosis and 4 with ATTR cardiac amyloidosis). Pulmonary [¹⁸F]florbetapir uptake was evaluated visually and quantified as distribution volume of specific binding (Vs) derived from compartmental analysis and simpler semiquantitative metrics of maximum standardized uptake values (SUVmax), retention index (RI), and target-to-blood ratio (TBR).

Results On visual analysis, pulmonary tracer uptake was absent in most AL subjects (40/58, 69%); 12% (7/58) of AL subjects demonstrated intense bilateral homogeneous tracer uptake. In this group, compared to the control group, Vs (median Vs 30-fold higher, 9.79 vs. 0.26, p < 0.001), TBR (median TBR 12.0 vs. 1.71, p < 0.001), and RI (median RI 0.310 vs. 0.033, p < 0.001) were substantially higher. Notably, the AL group without visually apparent pulmonary [¹⁸F]florbetapir uptake also demonstrated a > 3-fold higher Vs compared to the control group (median 0.99 vs. 0.26, p < 0.001). Vs was independently related to left ventricular SUVmax, a marker of cardiac AL deposition, but not to ejection fraction, a marker of cardiac dysfunction. Also, intense [¹⁸F]florbetapir lung uptake was not related to [¹¹C]acetate lung uptake, suggesting that intense [¹⁸F]florbetapir lung uptake represents AL amyloidosis rather than heart failure.

Conclusions [¹⁸F]florbetapir PET/CT offers the potential to noninvasively identify pulmonary AL amyloidosis, and its clinical relevance warrants further study.

Keywords Systemic light chain amyloidosis · [18F]florbetapir · PET/CT · Lung · AL · Quantitative

This article is part of the Topical Collection on Miscellanea

Electronic supplementary material The online version of this article (https://doi.org/10.1007/s00259-019-04627-7) contains supplementary material, which is available to authorized users.

Sharmila Dorbala sdorbala@bwh.harvard.edu

Yiu Ming Khor khor.yiu.ming@singhealth.com.sg

- ¹ Division of Nuclear Medicine, Department of Radiology, Brigham and Women's Hospital, Boston, MA, USA
- ² Cardiac Amyloidosis Program, Division of Cardiology, Department of Medicine, Brigham and Women's Hospital, Boston, MA, USA
- ³ Division of Medical Oncology, Dana Farber Cancer Institute, Boston, MA, USA
- 🖄 Springer

- ⁴ Division of Medical Oncology, Memorial Sloan Kettering Medical Center, New York, NY, USA
- ⁵ Division of Hematology and Oncology, Department of Medicine, Massachusetts General Hospital, Boston, MA, USA
- ⁶ Amyloidosis Center, Boston University School of Medicine, Boston, MA, USA
- ⁷ Stanford University Cardiovascular Institute and Cardiovascular Medicine, Stanford Amyloid Center, Stanford, CA, USA

Introduction

Amyloidosis is a heterogeneous group of disorders that result from misfolding and aggregation of a family of proteins into insoluble β -pleated-sheet structures. In systemic light chain amyloidosis (AL), misfolding of immunoglobulin light chain proteins from a plasma cell dyscrasia results in extensive extracellular deposition of amyloid fibrils, which can lead to dysfunction of the affected organ [1].

Involvement of the lung is common in systemic AL amyloidosis; two postmortem series reported pulmonary involvement in 90–92% of patients [2, 3]. However, the diagnosis of pulmonary amyloidosis is challenging. Clinical symptoms such as dyspnea, exercise intolerance, and cough are nonspecific and could be attributed to heart failure. Laboratory testing, pulmonary function testing, diffusion capacity of lungs for carbon monoxide (DLCO) studies, or computed tomography of the lungs, though sometimes suggestive, cannot specifically confirm the diagnosis of pulmonary amyloidosis [4, 5]. While histology remains the gold standard, biopsy is generally avoided due to concerns of recurrent pleural effusions and bleeding risk [6]. Consequently, diagnosis of pulmonary AL amyloidosis during life is rare.

Targeted imaging of systemic amyloid deposits using positron emitting radiotracers may address this unmet clinical need. Several amyloid radiotracers which are wellestablished for imaging beta amyloid in Alzheimer's disease are emerging as promising tools to evaluate other forms of systemic amyloid deposition [7, 8]. Our group and others have shown that [¹⁸F]florbetapir positron emission tomography computed tomography (PET/CT) can identify amyloid deposits in the heart [9] and extra-cardiac organs [10].

Increased lung uptake of [¹⁸F]florbetapir could represent specific binding to pulmonary AL amyloid deposits. However, in heart failure, elevated left atrial pressures and slower transit time through the pulmonary vascular bed are known to result in pulmonary accumulation of myocardial perfusion tracers (thallium-201, technetium-99 m sestamibi and tetrofosmin) [11–13], especially on early images [14, 15]. Whether this finding is unique to perfusion tracers or may occur with [¹⁸F]florbetapir as well is unclear.

The primary aim of this study was to test our hypothesis that the distribution volume of [¹⁸F]florbetapir specific binding in the lungs, evaluated by compartmental modelling, reflects pulmonary AL amyloid deposition in subjects with systemic AL amyloidosis. Because kinetic modelling is impractical outside of research setting, we assessed several simple, clinically relevant semiquantitative [¹⁸F]florbetapir metrics for identifying pulmonary AL amyloidosis. To distinguish pulmonary AL amyloidosis from heart failure-related increased lung tracer uptake, we evaluated the clinical correlates of pulmonary [¹⁸F]florbetapir uptake and compared lung uptake of [¹⁸F]florbetapir to lung uptake of [¹¹C]acetate in a subgroup of subjects.

Methods

Subject eligibility and study design

This analysis included 67 subjects: 58 subjects in the AL amyloidosis group and 9 subjects in the control group. Fifty-eight subjects with biopsy-proven systemic AL amyloidosis were prospectively recruited for this study, "Molecular Imaging of Primary Amyloid Cardiomyopathy" (ClinicalTrials.gov Identifier: NCT02641145), per predefined inclusion and exclusion criteria (Supplemental Table 1). The control group included 5 subjects without amyloidosis and 4 subjects with cardiac transthyretin (ATTR) amyloidosis enrolled in a prior pilot study of [¹⁸F]florbetapir cardiac PET [9]. The study was approved by the institutional Human Research Committee, and all subjects provided written informed consent.

Study protocol

All 58 subjects underwent research [¹⁸F]florbetapir PET/CT, cardiac magnetic resonance imaging, and clinical echocardiography within a median of 1–2 days of each other. Eighteen of the 58 subjects underwent [¹¹C]acetate cardiac PET/CT scan within a median of 1 day of the [¹⁸F]florbetapir PET/CT scan.

[¹⁸F]florbetapir PET/CT

Acquisition

Subjects underwent PET/CT using a GE Discovery DRX imaging system (GE Healthcare, Chicago, IL, USA). This PET/ CT scanner was accredited by Intersocietal Accreditation Commission Nuclear/PET as per American College of Radiology (ACR) guideline and manufacturer requirements to maintain good image quality. System calibration, wellcounter cross-calibration, and SUV accuracy (within 5%) were done quarterly.

Following a low-dose CT scan of the heart (120–140 kV, 80 mA), 358 ± 45 MBq of [¹⁸F]florbetapir (PETNET, Woburn, MA) was injected, and a 60-min dynamic PET scan of the heart was acquired in three-dimensional list mode. A partial-body PET scan was then obtained from skull base to below the kidneys for 30 min with 5 min per PET bed position. CT images of the same region were acquired for

attenuation correction using a non-contrast technique (10 mA tube current, 140 kVP tube voltage, free tidal breathing). Dynamic cardiac image series were generated by reconstruction of the list-mode data into 47 dynamic frames (20×5 s, 8×10 s, 6×30 s, 4×60 s, 8×300 s, 1×600 s). Static images were reconstructed from 4 to 30 min for review of myocardial tracer uptake. Images were reconstructed onto a 128×128 matrix ($4.09 \times 4.09 \times 3.27$ mm³ voxel size) by ordered subset expectation maximization (OSEM) with two iterations, 21 subsets, and 4.48 mm full-width at half-maximum Gaussian post-filter. All scans were corrected for decay, scatter, random coincidences and photon attenuation.

Analysis

Lung Visually, [¹⁸F]florbetapir uptake was assessed from the static partial-body images, and static cardiac images, and categorized as no uptake, mild diffuse uptake, or intense diffuse uptake. Subjects with minimal uptake limited to the dependent portions of the lungs, typically in the posterior bases of the lungs, and otherwise no tracer uptake in the rest of the lungs were categorized as no uptake.

Kinetic modelling For kinetic modelling, compartmental analyses were performed using PMOD version 3.805 (PMOD Technologies LLC., Zurich, Switzerland). The lung was manually segmented, in a slice-by-slice manner from the apex to base. Care was taken to exclude the major airways, pulmonary vessels, and the liver, which typically had intense tracer uptake as the radiotracer was excreted via the hepatobiliary system. Tracer activity concentration was corrected for density using CT data, i.e., activity concentration in kBq/ml was converted to kBq/g. This was done to remove the confounding effect of any physiological or pathological process that might increase the amount of lung tissue in a given volume, such as atelectasis or collapse, which would lead to higher tracer activity in the given volume not attributable to actual tracer uptake by the tissue. A spherical VOI of 3 mm radius in the pulmonary artery below the bifurcation was defined as the blood pool and used as the input function. Several models were tested on a subset of subjects (i.e., 3 subjects from each group). Models included 1-tissue-2-kinetic-parameter compartment model (1T2k), 2T4k, 3T6k, all with fitted blood volume fraction, and Logan plot. The 2T4K model was selected as the best model based on the lowest Akaike information criteria (AIC) scores and was then used for compartmental analysis for all subjects on the dynamic PET data using PMOD. The outcome measure for the model was distribution volume of specific binding (Vs = $K_1/k_2 * k_3/k_4$). Normal kinetic parameters were derived from the control cohort (n = 9).

Semiquantitative metrics We also evaluated pulmonary [¹⁸F]florbetapir uptake semiquantitatively using maximum

standardized uptake values (SUVmax), retention index (RI), and target-to-blood ratio (TBR). TBR and RI were corrected for lung density, while SUVmax was not. From the dynamic images, we obtained the RI of the whole lungs from 10 to 30 min after radiotracer injection using PMOD. The same VOIs of the whole lungs and blood pool were also used to obtain TBR from the reconstructed dynamic images from 50 to 60 min after radiotracer injection.

We determined the average SUVmax of the lungs from the static partial-body images by averaging the SUVmax within four spherical volumes of 1 cm radius over the lungs on the delayed partial-body PET/CT images, two at the level of the aortic arch to represent the upper lobes in each lung and two at the level of the left atrium to represent the lower lobes. An additional spherical volume of interest (VOI) was drawn over a pleural effusion if present. This analysis could not be performed in the control cohort as static images at a comparable time point (60 to 90 min) were not available. For the purpose of comparison to [¹¹C]acetate PET/CT, the average SUVmax values of the lungs were derived from images acquired from 2 to 6 min post radiotracer injection.

Normalized [¹⁸F]florbetapir time–activity curves were generated from the dynamic images by standardizing the measured activity in MBq/cc to an injected dose of 300 MBq (unit, MBq/cc/300 MBq).

Heart We evaluated [¹⁸F]florbetapir uptake in the left ventricle using SUVmax within the left ventricular myocardium and global RI from 10 to 30 min with Carimas 2.9 (Turku PET Centre, Finland) as described previously [16]. Image-derived input function was defined by the blood pool within the left atrium. The right ventricle was assessed visually for presence or absence of tracer uptake.

Other organs Images were assessed for tracer uptake in the following organs: parotid, tongue, thyroid, pancreas, spleen, kidney, muscle, subcutaneous fat, and humeral head bone marrow as described before. The organ was deemed to be involved if its SUVmax exceeded 2.5 [10].

CT Methods

All CT images were evaluated for pleural effusion and CT features suggestive of pulmonary amyloidosis. The whole lung was manually segmented as described above. The average density was approximated by [17]:

Density $\left[g/cm^3 \right] = 1 + HU/1000$

The diameter of the main pulmonary artery (PA) and ascending aorta was measured on a transaxial slice at the level of the PA bifurcation, and the PA to ascending aorta ratio was calculated.

[¹¹C]acetate PET/CT

Following a CT scout and a low-dose CT scan of the heart (120–140 kV, 80 mA), 609 \pm 146 MBq of [¹¹C]acetate was injected intravenously, and a 30-min dynamic PET scan of the heart was acquired in three-dimensional list mode. Images were reconstructed into static myocardial perfusion images (2–6 min), and SUVmax was estimated as described in the above section on lung/semiquantitative metrics/average SUVmax.

Echocardiography methods

2D echocardiography with spectral and color Doppler imaging was performed in all subjects according to standard American Society of Echocardiography recommendations [18]. Tricuspid regurgitation pressure and peak velocity were measured to estimate the pulmonary artery systolic pressure (PASP) using the modified Bernoulli equation. Elevated PASP was defined as > 36 mmHg [19].

Cardiac magnetic resonance imaging (CMR) methods

All CMR images were acquired on a 3.0-T system (Tim Trio, Siemens, Erlangen, Germany), with electrocardiographic gating and breath holding. The protocol consisted of steady-state free precession cine imaging for assessing ventricular function and morphology. Gadolinium contrast was used. A commercial software package (Medis Suite 3.0 Medical Imaging Systems, Leiden, the Netherlands) was used to post-process and quantify left and right ventricular end diastolic volume (EDV), end systolic volume (ESV), stroke volume (SV), ejection fraction (EF), and right ventricular wall thickness. Abnormal left ventricular (LV) EF was defined as described previously: < 56% for males and < 58% for females [20]. Abnormal right ventricular (RV) EF was defined as described previously: < 48% for males and < 50% for females [21]. Abnormal RV wall thickness was defined as > 5 mm [22].

Six-minute walk test

Subjects performed a 6-min walk test using a standardized protocol adapted from the American Thoracic Society guidelines; the subject walked on a level surface at a self-governed pace for 6 min, and the total distance travelled was measured.

Statistical analysis

Statistical analyses were performed using SPSS version 20.0.0 (IBM Corp., Armonk, NY). To investigate demographic and

clinical data, χ^2 tests for discrete variables and Student's ttests for continuous data were used. Results of non-normally distributed variables were reported as medians with interguartile range (IOR). Differences were compared using the Mann-Whitney U test for 2 groups of continuous variables and the Kruskal-Wallis test for 3 groups of continuous variables. Spearman's rank correlation was used to determine the association between two non-normally distributed continuous variables. We evaluated several categorical parameters that could affect pulmonary [¹⁸F]florbetapir tracer kinetics including pulmonary parenchymal changes and pleural effusions, circulating light chain pathology (active vs. remission status of AL disease), left and right ventricular function, amyloid burden in both ventricles, and systemic organ involvement. Univariable correlates of Vs were determined. Significant univariable correlates were included in a multivariable linear regression model to determine the independent predictors of Vs. A 2-tailed p value of less than 0.05 was considered significant.

Results

Baseline characteristics

Demographics and clinical characteristics of the 58 subjects included in this analysis are listed in Table 1. Forty-two subjects had active AL amyloidosis and abnormal serum free light chain levels (32 with and 10 without cardiac involvement). Sixteen subjects had AL amyloidosis with cardiac involvement in hematologic remission with normal serum free light chain levels for at least 1-year post treatment (remission-AL).

Pulmonary parenchymal changes

Most of the lung findings in this study (visual, quantitative, and semiquantitative) were bilateral, without appreciable unilateral differences.

Visual evaluation

On visual evaluation, we noted intense diffuse lung uptake of tracer in 12% (n = 7), while most of the remaining subjects showed mild diffuse uptake (n = 11, 19%) or no significant uptake (n = 40, 69%), including two subjects with minimal uptake in the dependent areas of the lungs (Fig. 1a–c). None of the subjects showed any specific lung CT findings suggestive of amyloid deposits. Nonspecific patchy or diffuse ground glass opacity was seen in 9/18 (50%) on CT, but the average lung densities did not differ among the three patterns of [¹⁸F]florbetapir uptake in the lungs as well as the control and ATTR subjects (Fig. 1d).

In the subjects with active AL amyloidosis, the majority (61.9%, 26/42) showed no lung uptake of [¹⁸F]florbetapir; the

| Characteristics | 18 F-florbetapir lung uptake present (n = 18) | 18 F-florbetapir lung uptake absent (n = 40) | p value |
|---------------------------------------|--|---|---------|
| Age (years) ^a | 62 ± 7 | 61 ± 7 | 0.973 |
| Gender | | | |
| Male | 67% | 60% | 0.772 |
| Monoclonal light chain isotyp | e | | |
| к (mg/L) | 15.2 (1.9–1716.0) | 15.1 (2.2–704.6) | 0.593 |
| λ (mg/L) | 196.2 (3.8–1640.0) | 44.3 (1.3–921.6) | 0.101 |
| κ/λ ratio | 0.12 (0.01–228.80) | 0.40 (0.01–56.65) | 0.112 |
| NT-proBNP (pg/mL) | 4350 (146–15,116) | 1248 (44–48,412) | 0.022 |
| Troponin (ng/mL) | 0.09 (0-248.0) | 0.01 (0-247) | 0.022 |
| PA diameter (cm) ^a | 2.9 ± 0.3 | 2.8 ± 0.3 | 0.167 |
| PA/aorta ratio ^a | 0.90 ± 0.13 | 0.85 ± 0.09 | 0.064 |
| 6-min walk test (meters) ^a | 427 ± 101 | 411 ± 96 | 0.604 |

Table 1 Baseline characteristics

Values given as median (range) unless otherwise stated; ^a Values given as mean \pm standard deviation; *NT-proBNP* N-terminal pro b-type natriuretic peptide, *PA* pulmonary artery

remaining demonstrated mild (19%, 11/42) or intense (8.6%, 5/42) lung uptake of [¹⁸F]florbetapir. In the remission-AL group, while the majority (84%, 14/16) showed no lung uptake, a small percentage of subjects showed intense uptake (12%, 2/16).

Quantitative evaluation

Mean time–activity curves of normalized [¹⁸F]florbetapir activity in the lungs differed significantly among the three patterns of lung uptake and was lowest in the control group (Fig.







parenchyma was unremarkable in these subjects. CT average lung densities did not differ among subjects with no significant uptake, mild diffuse uptake, intense homogeneous uptake, and the control group (bottom, d). The mean and standard deviation are shown. Each marker corresponds with a single subject



Fig. 2 [18 F]Florbetapir time-activity curves in the lungs. The rate of [18 F]florbetapir tracer washout was slowest in subjects with intense uptake and fastest in the control subjects. Shaded area represents standard deviation

2). On compartmental modelling, [¹⁸F]florbetapir pulmonary Vs was highest in the subjects with intense [¹⁸F]florbetapir lung uptake (median 9.788, IQR 4.796–14.500, 30-fold



higher than control group Vs) and lowest in the control group (Fig. 3a). Notably, even among AL amyloidosis subjects without pulmonary [¹⁸F]florbetapir uptake, median [¹⁸F]florbetapir pulmonary Vs was more than threefold higher than in the control group (median 0.994, IQR 0.483–1.693 vs. 0.262, IQR 0.098–0.380, p < 0.001). Among these 9 control subjects, LVEF was < 50% in three subjects, > 50% in three subjects, and unavailable in the remaining three subjects.

Semiquantitative evaluation

Estimation of lung Vs requires dynamic imaging and compartmental modelling and is therefore not suitable for routine clinical use. For this reason, we evaluated three simpler metrics of $[^{18}F]$ florbetapir lung uptake—SUVmax, TBR, and RI. Measurement of SUVmax could not be performed on the control and ATTR subjects as static images at a comparable time point (60 to 90 min) were not available. All three simplified quantitative metrics of $[^{18}F]$ florbetapir lung uptake were highest in subjects with intense lung uptake, followed



Fig. 3 Quantitative metrics of [¹⁸F]florbetapir stratified by visual lung uptake. Median Vs of subjects from the control group (5 subjects without amyloidosis and 4 subjects with ATTR amyloidosis) was more than threefold lower than AL amyloidosis subjects (a). RI (b), TBR (c), and SUVmax (d) were the highest in subjects with intense tracer uptake,

followed by subjects with mild and no uptake. The control and ATTR subjects demonstrated the lowest TBR and RI. A SUVmax value of 1.5 distinguished subjects with and without lung uptake. The median and interquartile range are shown. Each marker corresponds with a single subject

by those with mild uptake and then no uptake. The control and ATTR subjects demonstrated the lowest TBR and RI (Fig. 3b to d). A SUVmax threshold value of 1.5 perfectly classified subjects with and without [¹⁸F]florbetapir lung uptake. Vs was highly correlated with SUVmax, RI, and TBR, with the strongest relationship to RI (Fig. 4a to c).

Pleural effusions

Forty percent of the study subjects (n = 23) had pleural effusion on at least one side, ranging in size from minimal to large. Clinical and laboratory differences in subjects with and without pleural effusion are shown in Supplemental Table 2. Notably, the effusions showed no significant tracer uptake with mean SUVmax of 0.84 ± 0.43 . No significant tracer uptake or tracer-avid nodularity was detected along the pleura in any subjects. Pleural effusion was only seen in subjects with cardiac AL amyloidosis (48% vs. 0%, p = 0.004). Visual ¹⁸F]florbetapir lung uptake was more prevalent in subjects with compared to without pleural effusion (52% vs. 23%, p= 0.027) (Fig. 5a, b). However, Vs, SUVmax, TBR, and RI of [¹⁸F]florbetapir in the lungs were not significantly different between subjects with and without pleural effusion. Moderate to intense tracer uptake adjacent to the pleural effusion was seen in 6 subjects, likely attributable to accumulation of tracer uptake within the atelectatic or collapsed lung (Fig. 5c).

Distinguishing pulmonary AL amyloidosis from heart failure-related lung tracer uptake

Univariable predictors of Vs, the distribution volume of specific binding, of [¹⁸F]florbetapir in the lungs

We evaluated the distribution volume of specific binding of [¹⁸F]florbetapir in the lungs based on the subjects' LV and RV structure and function, LV and RV amyloid content, systemic organ involvement, and circulating light chain pathology. Certain parameters of amyloid load, such as LV SUVmax (r = 0.517, p < 0.001), RV uptake (0.321, p = 0.025), and log troponin T (0.279, p = 0.045), correlated with Vs. However, parameters of cardiac function (LVEF, RVEF, RV wall thickness, estimated PASP) or wall stress (NTproBNP) did not correlate with Vs. Serum free light chain levels were not also related to Vs. Median Vs was similar between subjects with normal and abnormal values of LVEF, RVEF, estimated PASP, and RV wall thickness, in the active AL vs. remission group, and between subjects with AL amyloid involvement of ≥ 3 or < 3 extra-thoracic organs (Table 2).



Fig. 4 Simplified quantitative metrics of [¹⁸F]florbetapir in the lungs and their correlation with outcome of kinetic modelling. SUVmax, target-to-blood ratio, and RI were strongly correlated with Vs

Independent predictors of Vs, the distribution volume of specific binding of [¹⁸F]florbetapir in the lungs

We performed multiple linear regression analysis to determine independent determinants of Vs. The model included

 Table 2
 Summary of univariable predictors of distribution volume of specific binding (Vs)

| Variable | Distribution volume of specific binding (Vs) | p value |
|--|--|---------|
| LVEF | | |
| Normal Abnormal RVEF | 1.12 (0.44–2.43) 1.76 (1.04–3.55) | 0.103 |
| Normal Abnormal PASP | 1.12 (0.53–2.59) 1.75 (0.91–3.55) | 0.199 |
| Normal Abnormal RV wall thickne | 1.26 (0.56–2.34) 2.00 (0.27–6.72) | >0.99 |
| Normal Abnormal | 1.24 (0.58–2.72) 1.41 (0.46–3.80) | 0.732 |
| In remission Active | 1.46 (0.61–2.27) 1.30 (0.56–3.40) | 0.857 |
| Extra-thoracic of Less than $3 \ge 3$ | 1.29 (0.57–2.43) 1.43 (0.64–3.80) | 0.327 |

Values given as median (range). *LV* left ventricle, *EF* ejection fraction, *RV* right ventricle, *PASP* pulmonary artery systolic pressure, *AL* light chain amyloidosis

significant univariable parameters, log LV SUVmax, RV uptake, log troponin T, as well as the parameter of interest, namely, LVEF. This model significantly predicted Vs (F = 3.978, r = 0.570, p = 0.010). However, log LV SUVmax was the only significant predictor of lung Vs (Table 3).

Comparing lung uptake of [¹¹C]acetate to [¹⁸F]florbetapir

To further discriminate lung AL amyloidosis from heart failure (high filling pressures and/or slow pulmonary transit of radiotracer), we analyzed a subset of 18 subjects who underwent [¹¹C]acetate and [¹⁸F]florbetapir scans (56% same day studies, median 1 day apart). Visually, none of the 18 subjects had intense [¹¹C]acetate lung uptake, despite four of them showing intense [¹⁸F]florbetapir lung uptake suggesting lung amyloidosis (Fig. 6). SUVmax of [¹¹C]acetate lung uptake in these four subjects was indistinguishable from the subjects with no [¹⁸F]florbetapir lung uptake (1.1, IQR 0.8–1.7, vs. 1.4, IQR 0.8–1.6, p = 0.959, Fig. 7). The discordant finding of minimal [¹¹C]acetate lung uptake in subjects with intense [¹⁸F]florbetapir lung uptake further supports the notion that intense lung uptake of [¹⁸F]florbetapir likely represents lung amyloidosis.

Discussion

To our knowledge, our study is the first study to date to prospectively evaluate for pulmonary AL amyloidosis using ¹⁸F]florbetapir PET/CT. In this study of 58 subjects with systemic AL amyloidosis and 10 control subjects, our findings show that pulmonary [¹⁸F]florbetapir Vs, the distribution volume of [¹⁸F]florbetapir specific binding in the lungs, derived from compartmental modelling, is substantially higher (3-30 fold) in subjects with AL amyloidosis compared to the control subjects, suggesting specific binding to AL lung deposits. Vs was strongly correlated with lung SUVmax and TBR, which were measured late (50-90 min post injection of ¹⁸F]florbetapir). Notably, Vs was independently related to LV SUVmax, a marker of cardiac AL deposition, and not to LVEF, a marker of cardiac dysfunction. Furthermore, on early phase images, increased lung uptake of [¹¹C]acetate might have suggested heart failure; but none of the subjects with intense [¹⁸F]florbetapir lung uptake showed increased [¹¹C]acetate lung uptake. Together, these findings support our hypothesis that intense [¹⁸F]florbetapir lung uptake reflects binding to pulmonary amyloid deposits rather than heart failurerelated lung tracer uptake. Importantly, our results show that simpler metrics of SUVmax, TBR, and RI are reliable measures for assessment of AL lung involvement in clinical practice.



Fig. 5 [¹⁸**F**]**florbetapir imaging in AL amyloidosis and pleural effusion.** Axial fused [¹⁸**F**]**f**lorbetapir PET/CT images from three subjects with right pleural effusion showed no significant pulmonary parenchymal

uptake (a), homogeneous intense uptake in the lungs (b), and intense tracer uptake in the collapsed lung anterior to the pleural effusion on the background of diffuse intense tracer uptake in the lung parenchyma (c)

 Table 3
 Summary of multiple linear regression analysis for variables predicting Vs

| Beta | 95% CI Lower bound | p value |
|--------|--|---|
| 0.191 | -0.048-0.197 | 0.226 |
| -0.148 | -0.269-0.133 | 0.495 |
| -0.078 | -0.022-0.014 | 0.635 |
| 0.582 | 0.243-1.564 | 0.009 |
| | Beta 0.191 -0.148 -0.078 0.582 | Beta 95% CI Lower bound 0.191 -0.048-0.197 -0.148 -0.269-0.133 -0.078 -0.022-0.014 0.582 0.243-1.564 |

Model R = 0.570, F = 3.978, p = 0.010. β standardized beta coefficient, RV right ventricular, EF ejection fraction, LV left ventricular, RI retention index, SUV standardized uptake value, NT-proBNP N-terminal pro b-type natriuretic peptide

Three forms of amyloidosis can be found in the lung: nodular pulmonary amyloidosis, diffuse alveolar-septal amyloidosis, and tracheobronchial amyloidosis. The form that is most commonly associated with systemic AL amyloidosis is diffuse alveolar-septal amyloidosis, which typically involves all lobes and is characterized by the presence of amyloid deposits in the interstitium within the alveolar septa and vessel walls [23]. The PET/CT appearance of homogeneous [¹⁸F]florbetapir uptake in the lung parenchyma in our subjects most closely reflects this histopathology. None of the subjects demonstrated tracer-avid nodules in the lung parenchyma or airways which, if present, would have suggested the nodular and tracheobronchial forms of amyloidosis.

We found that 21% of subjects with AL cardiac amyloidosis (either active or in remission) showed diffuse mild [¹⁸F]florbetapir tracer uptake in the lungs and 15% showed diffuse intense tracer uptake. These findings are consistent with those of an autopsy series from Johns Hopkins Hospital, in which 24% of patients with cardiac amyloidosis were found to have moderate amyloid involvement of the alveolar septa and/or pulmonary vessels and 13% had severe amyloid involvement [24]. Together, these findings imply that presence of visually apparent pulmonary [¹⁸F]florbetapir uptake in



Fig. 6 Discordant uptake of [¹¹C]acetate and [¹⁸F]florbetapir in the lungs. Early (2–6 min after injection of radiotracer) axial and coronal images in one subject showed minimal lung uptake of [11 C]acetate (Fig. 6a) despite intense lung uptake of [18 F]florbetapir (Fig. 6b). We propose

that this discordant finding combined with high $[^{18}F]$ florbetapir Vs, the distribution volume of $[^{18}F]$ florbetapir specific binding in the lungs derived from compartmental modelling, suggests pulmonary AL amyloidosis



Fig. 7 SUVmax values of [¹¹**C**]acetate **PET/CT and** [¹⁸**F**]florbetapir **PET/CT.** On the early images (2–6 min after injection of radiotracer), subjects with intense visual lung uptake of [¹⁸**F**]florbetapir had significantly higher [¹⁸**F**]florbetapir SUVmax values than subjects with no uptake (1.5, IQR 1.1–3.2, vs 12.8, IQR 9.4–18.0, p < 0.001), but the median [¹¹C]acetate SUVmax of the lungs were similar in all subjects (1.1, IQR 0.8–1.7, vs. 1.4, IQR 0.8–1.6, p = 0.959), suggesting pulmonary AL amyloidosis. The median is shown. Each marker corresponds with a single subject

subjects with systemic AL amyloidosis indicates moderate to severe amyloid involvement in the lungs.

The prevalence of pleural effusions in our study subjects (40%) is higher than previously reported by Berk et al. (6–18%) [25]. This difference can be explained by the selective enrolment of subjects with cardiac involvement in our study compared to that cohort. We did not detect tracer-avid pleural nodularity in any subject with effusion, either because none of the subjects had amyloid infiltration of the pleura or because the pleural amyloid nodules were too small for detection by PET.

Until now pulmonary amyloidosis could not be diagnosed pre-mortem; hence, its clinical significance has not been established. Based on several studies [2, 26-28], it is generally believed to be rarely symptomatic and of little clinical consequence; however, some studies have implied the opposite. In a 20-year review at Boston Medical Centre of 76 patients with biopsyproven pulmonary amyloidosis, 71% had abnormal pulmonary function test results, and approximately half demonstrated restrictive pattern [29]. A study by Mayo Clinic with complete survival data of patients with biopsy-proven pulmonary amyloidosis reported poor long-term outcome, with a median survival after diagnosis of 16 months [30]. These findings imply a wide spectrum of disease manifestation, ranging from asymptomatic to life-threatening. Importantly, failure to recognize a coexisting lung pathology in patients with cardiac amyloidosis may lead to futile hearttargeted therapy, such as over-diuresis, which can be dangerous. Noninvasive detection of pulmonary amyloidosis, now available for the first time using [¹⁸F]florbetapir PET/CT, will likely advance our understanding of this disease and improve management.

All the metrics used to evaluate pulmonary uptake of [¹⁸F]florbetapir performed well. We find that measurement of SUVmax of the lungs is a fast, simple, and reliable method of quantifying pulmonary uptake of [¹⁸F]florbetapir. It is comparable to the more complex and time-consuming metric of Vs which requires long scanning time in dynamic mode and knowledge of kinetic modelling.

Other PET radiotracers had also been evaluated for diagnosis of systemic amyloidosis; for example, 2-deoxy-2-[¹⁸F]fluoro-D-glucose ([¹⁸F]FDG) is widely available and is shown to be helpful in distinguishing between localized and systemic amyloidosis [31]. [¹⁸F]FDG is, however, limited by relatively low sensitivity [32] as well as specificity [33] due to tracer accumulation in other common conditions such as neoplasm, infection, and inflammation. [¹⁸F]florbetapir has the advantage of highly specific binding to amyloid fibrils [34, 35] and does not require an on-site cyclotron unlike [¹¹C]-labeled Pittsburgh compound B (PiB).

A limitation of the study is the lack of histological confirmation as gold standard to evaluate the diagnostic performance of [18F]florbetapir PET/CT to identify pulmonary amyloidosis. Lung/pleural biopsies are not commonly performed clinically in AL amyloidosis, and a correlative study with pulmonary histology is likely not feasible. However, we were able to demonstrate a 3 to 30-fold higher Vs in the AL cohort compared to control cohort, strongly supporting the notion of tracer binding to AL. AL lung activity can show increased [¹⁸F]FDG uptake. However, unfortunately, ¹⁸F]FDG PET was not performed for the study subjects in this protocol. Our study used the 6-min walk test as a measure of subjects' functional capacity, which was not significantly different between subjects with and without pulmonary uptake of [¹⁸F]florbetapir. As a composite measure of cardiopulmonary fitness, the test is not able to distinguish between a cardiac and respiratory cause of subjects' functional limitation. Future studies with pulmonary function tests with DLCO and a prospective study of a select group of patients with proven lung involvement/diffuse interstitial lung pattern using ¹⁸F]florbetapir PET will be critical to expand these pilot results.

Conclusion

[¹⁸F]florbetapir PET/CT appears to be a promising noninvasive imaging approach to diagnose pulmonary involvement in individuals with systemic AL amyloidosis. The clinical and prognostic implications of pulmonary AL amyloidosis diagnosed by [¹⁸F]florbetapir warrant further study. **Acknowledgments** We are extremely grateful to the study subjects and the study sites for their participation.

Authors' contributions YMK, SC, HJH, MFK, MAP, MR, HH, and SD were responsible for the study design and performed primary data collection, analysis, and manuscript preparation. FLR, VS, HL, AJY, GB, RL, JB, RHF, and MFD assisted with subject recruitment and/or manuscript editing.

Funding information SD and RF are supported by NIH RO1 grant (RO1 HL 130563); SD and RL are supported by American Heart Association Grant (AHA 16 CSA 2888 0004). HL receives support from NIH/NCI Cancer Center Support Grant P30 CA008748. MDC is supported by Spectrum Dynamics and Gilead.

Compliance with ethical standards

Conflict of interest HH is a working owner of MedTrace Pharma. MDC has received consulting fees from Sanofi and GE Healthcare. HL has received consulting fees from Celgene, Takeda, Janssen, Prothena, Pfizer, and Juno and research support from Amgen, Spectrum, and Takeda. VS has received research support from Takeda, Celgene, Janssen, and Prothena and is on the scientific advisory board for Caleum Biosciences. FLR has received consulting fees from Pfizer, GlaxoSmithKline, and Caleum Biosciences and research support from Eidos Therapeutics. RHF has received consulting fees from Ionis Pharmaceuticals and Alnylam Pharmaceuticals and research funding from GlaxoSmithKline. SD has received consulting fees from Pfizer, GE Healthcare, and AAA and research grants from Pfizer. YMK declares that she has no conflict of interest. SC declares that she has no conflict of interest. MFK declares that she has no conflict of interest. MAP declares that she has no conflict of interest. MR declares that he has no conflict of interest. HH declares that she has no conflict of interest. GB declares that she has no conflict of interest. AJY declares that he has no conflict of interest. RL declares that she has no conflict of interest. JB declares that he has no conflict of interest.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the Partners Human Research Committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

Informed consent Statement of informed consent was obtained from all individual participants included in the study.

Abbreviations AL, light chain amyloidosis; ATTR, transthyretin amyloidosis; FLC, free light chains; PET/CT, positron emission tomography computed tomography; RI, retention index; SUVmax, maximum standardized uptake value; TBR, target-to-blood ratio; Vs, distribution volume of specific binding; HU, Hounsfield unit; PA, pulmonary artery; PASP, pulmonary artery systolic pressure; CMR, cardiac magnetic resonance imaging; EDV, end diastolic volume; ESV, end systolic volume; SV, stroke volume; EF, ejection fraction; LV, left ventricle; RV, right ventricle; 6mwt, 6-min walk test; IQR, interquartile range; NT-proBNP, N-terminal pro b-type natriuretic peptide

References

- Merlini G, Bellotti V. Molecular mechanisms of amyloidosis. N Engl J Med. 2003;349(6):583–96.
- Celli BR, Rubinow A, Cohen AS, Brody JS. Patterns of pulmonary involvement in systemic amyloidosis. Chest. 1978;74(5):543–7.

- Browning MJ, Banks RA, Tribe CR, et al. Ten years' experience of an amyloid clinic–a clinicopathological survey. Q J Med. 1985;54(215):213–27.
- Czeyda-Pommersheim F, Hwang M, Chen SS, Strollo D, Fuhrman C, Bhalla S. Amyloidosis: modern cross-sectional imaging. Radiographics. 2015;35(5):1381–92.
- Pickford HA, Swensen SJ, Utz JP. Thoracic cross-sectional imaging of amyloidosis. AJR Am J Roentgenol. 1997;168(2):351–5.
- Yood RA, Skinner M, Rubinow A, Talarico L, Cohen AS. Bleeding manifestations in 100 patients with amyloidosis. JAMA. 1983;249(10):1322–4.
- Ezawa N, Katoh N, Oguchi K, Yoshinaga T, Yazaki M, Sekijima Y. Visualization of multiple organ amyloid involvement in systemic amyloidosis using11C-PiB PET imaging. Eur J Nucl Med Mol Imaging. 2018;45(3):452–61.
- Wagner T, Page J, Burniston M, et al. Extracardiac 18F-florbetapir imaging in patients with systemic amyloidosis: more than hearts and minds. Eur J Nucl Med Mol Imaging. 2018;45(7):1129–38.
- 9. Dorbala S, Vangala D, Semer J, et al. Imaging cardiac amyloidosis: a pilot study using 18F-florbetapir positron emission tomography. Eur J Nucl Med Mol Imaging. 2014;41(9):1652–62.
- Ehman EC, El-Sady MS, Kijewski MF, et al. Early detection of multiorgan light chain (AL) amyloidosis by whole body (18)Fflorbetapir PET/CT. J Nucl Med. 2019;60(9):1234–9.
- 11. Patel GM, Hauser TH, Parker JA, et al. Quantitative relationship of stress Tc-99 m sestamibi lung uptake with resting Tl-201 lung uptake and with indices of left ventricular dysfunction and coronary artery disease. J Nucl Cardiol. 2004;11(4):408–13.
- Sanders GP, Pinto DS, Parker JA, Koutkia P, Aepfelbacher FC, Danias PG. Increased resting Tl-201 lung-to-heart ratio is associated with invasively determined measures of left ventricular dysfunction, extent of coronary artery disease, and resting myocardial perfusion abnormalities. J Nucl Cardiol. 2003;10(2):140–7.
- Georgoulias P, Demakopoulos N, Kontos A, Xaplanteris P, Xydis K, Fezoylidis I. Early post-stress pulmonary uptake of 99mTc tetrofosmin during exercise (SPECT) myocardial perfusion imaging: correlation with haemodynamic, perfusion and function parameters. Nucl Med Commun. 2006;27(2):119–26.
- Martinez EE, Horowitz SF, Castello HJ, et al. Lung and myocardial thallium-201 kinetics in resting patients with congestive heart failure: correlation with pulmonary capillary wedge pressure. Am Heart J. 1992;123(2):427–32.
- Marcassa C, Galli M, Baroffio C, Eleuteri E, Campini R, Giannuzzi P. Independent and incremental prognostic value of 201Tl lung uptake at rest in patients with severe postischemic left ventricular dysfunction. Circulation. 2000;102(15):1795–801.
- Nesterov SV, Han C, Maki M, et al. Myocardial perfusion quantitation with 15O-labelled water PET: high reproducibility of the new cardiac analysis software (Carimas). Eur J Nucl Med Mol Imaging. 2009;36(10):1594–602.
- Rosenblum LJ, Mauceri RA, Wellenstein DE, et al. Density patterns in the normal lung as determined by computed tomography. Radiology. 1980;137:409–16.
- Mitchell C, Rahko PS, Blauwet LA, et al. Guidelines for performing a comprehensive transhoracic echocardiographic examination in adults: recommendations from the American Society of Echocardiography. J Am Soc Echocardiogr. 2019;32(1):1–64.
- Bossone E, D'Andrea A, D'Alto M, et al. Echocardiography in pulmonary arterial hypertension: from diagnosis to prognosis. J Am Soc Echocardiogr. 2013;26(1):1–14.
- Yeon SB, Salton CJ, Gona P, et al. Impact of age, sex and indexation method on MR left ventricular reference values in the Framingham Heart Study Offspring cohort. J Magn Reson Imaging. 2015;41(4): 1038–45.
- Alfakih K, Plein S, Thiele H, Jones T, Ridgway JP, Sivananthan MU. Normal human left and right ventricular dimensions for MRI

as assessed by turbo gradient echo and steady-state free precession imaging sequences. J Magn Reson Imaging. 2003;17(3):323–9.

- Maceira AM, Prasad SK, Khan M, Pennell DJ. Reference right ventricular systolic and diastolic function normalized to age, gender and body surface area from steady-state free precession cardiovascular magnetic resonance. Eur Heart J. 2006 Dec;27(23):2879–88.
- Khoor A, Colby TV. Amyloidosis of the lung. Arch Pathol Lab Med. 2017;141(2):247–54.
- Smith RRL, Hutchins GM, Moore GW, Humphrey RL. Type and distribution of pulmonary parenchymal and vascular amyloid. Am J Med. 1979;66(1):96–104.
- Berk JL. Pleural effusions in systemic amyloidosis. Curr Opin Pulm Med. 2005;11(4):324–8.
- Cordier J-F. Pulmonary amyloidosis in hematological disorders. Semin Respir Crit Care Med. 2005;26(5):502–13.
- Milani P, Basset M, Russo F, Foli A, Palladini G, Merlini G. The lung in amyloidosis. Eur Respir Rev. 2017;26(145):170046.
- Berk JL, Keane J, Seldin DC, et al. Persistent pleural effusions in primary systemic amyloidosis: etiology and prognosis. Chest. 2003;124(3):969–77.
- Govender P, Keyes CM, Hankinson EA, O'Hara CJ, Sanchorawala V, Berk JL. Transbronchial biopsies safely diagnose amyloid lung disease. Amyloid. 2017 Mar;24(1):37–41.

- Utz SG. Pulmonary amyloidosis. Ann Intern Med. 1996;28(1):75– 83.
- Glaudemans AWJM, Slart RHJA, Noordzij W, Dierckx RAJO, Hazenberg BPC. Utility of 18F-FDG PET/CT in patients with systemic and localized amyloidosis. Eur J Nucl Med Mol Imaging. 2013;40(7):1095–101.
- Lee JH, Lee GY, Kim SJ, et al. Imaging findings and literature review of 18F-FDG PET/CT in primary systemic AL amyloidosis. Nucl Med Mol Imaging. 2010;49(3):182–90.
- Baqir M, Lowe V, Yi ES, Ryu JH. 18F-FDG PET scanning in pulmonary amyloidosis. J Nucl Med. 2014;55(4):565–8.
- Park MA, Padera RF, Belanger A, et al. 18F-Florbetapir binds specifically to myocardial light chain and transthyretin amyloid deposits: an autoradiography study. Circ Cardiovasc Imaging. 2016;59(3):157–61.
- 35. Choi SR, Schneider JA, Bennett DA, et al. Correlation of amyloid PET ligand florbetapir F 18 binding with Aβ aggregation and neuritic plaque deposition in postmortem brain tissue. Alzheimer Dis Assoc Disord. 2012;26(1):8–16.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.