ORIGINAL ARTICLE

Standardized added metabolic activity (SAM) IN ¹⁸F-FDG PET assessment of treatment response in colorectal liver metastases

Jeroen Mertens • S. De Bruyne • N. Van Damme • P. Smeets • W. Ceelen • R. Troisi • S. Laurent • K. Geboes • M. Peeters • I. Goethals • C. Van de Wiele

Received: 22 November 2012 / Accepted: 1 April 2013 / Published online: 1 May 2013 © Springer-Verlag Berlin Heidelberg 2013

Abstract

Purpose Standardized added metabolic activity (SAM) is a PET parameter for assessing the total metabolic load of malignant processes, avoiding partial volume effects and lesion segmentation. The potential role of this parameter in the assessment of response to chemotherapy and bevacizumab was tested in patients with metastatic colorectal cancer with potentially resectable liver metastases (mCRC).

Methods ¹⁸F-FDG PET/CT was performed in 18 mCRC patients with liver metastases before treatment and after

J. Mertens (⊠) · I. Goethals · C. Van de Wiele Department of Nuclear Medicine, Ghent University Hospital, De Pintelaan 185, 9000 Ghent, Belgium e-mail: jeroen.mertens@ugent.be

S. De Bruyne Department of Respiratory Medicine, Ghent University Hospital, Ghent, Belgium

N. Van Damme · W. Ceelen Department of Surgery, Ghent University Hospital, Ghent, Belgium

P. Smeets Department of Radiology, Ghent University Hospital, Ghent, Belgium

R. Troisi Department of General & Hepato-Biliary Surgery, Liver Transplantation Service, Ghent University Hospital, Ghent, Belgium

S. Laurent · K. Geboes · M. Peeters Department of Gastroenterology, Ghent University Hospital, Ghent, Belgium

M. Peeters Department of Oncology, Antwerp University Hospital, Edegem, Belgium five cycles of FOLFOX/FOLFIRI and bevacizumab. Of the 18 patients, 16 subsequently underwent resection of liver metastases. Baseline and follow-up SUV_{max}, and SAM as well as reduction in SUV_{max} (Δ SUV_{max}) and SAM (Δ SAM) of all liver metastases were correlated with morphological response, and progression-free and overall survival (PFS and OS).

Results A significant reduction in metabolic activity of the liver metastases was seen after chemotherapy with a median ΔSUV_{max} of 25.3 % and ΔSAM of 94.5 % (p=0.033 and 0.003). Median baseline SUVmax and SAM values were significantly different between morphological responders and nonresponders (3.8 vs. 7.2, p=0.021; and 34 vs. 211, p=0.002, respectively), but neither baseline PET parameters nor morphological response was correlated with PFS or OS. Follow-up SUV_{max} and SAM as well as Δ SAM were found to be prognostic factors. The median PFS and OS in the patient group with a high follow-up SUV_{max} were 10.4 months and 32 months, compared to a median PFS of 14.7 months and a median OS which had not been reached in the group with a low follow-up SUV_{max} (p=0.01 and 0.003, respectively). The patient group with a high follow-up SAM and a low Δ SAM had a median PFS and OS of 9.4 months and 32 months, whereas the other group had a median PFS of 14.7 months and a median OS which had not been reached (p=0.002 for both PFS and OS).

Conclusion ¹⁸F-FDG PET imaging is a useful tool to assess treatment response and predict clinical outcome in patients with mCRC who undergo chemotherapy before liver metastasectomy. Follow-up SUV_{max}, follow-up SAM and Δ SAM were found to be significant prognostic factors for PFS and OS.

Keywords Standardized added metabolic activity (SAM) \cdot ¹⁸F-FDG PET \cdot Treatment monitoring \cdot Colorectal cancer \cdot Liver metastases \cdot Bevacizumab

Introduction

Approximately 50 % of patients diagnosed with a primary colorectal carcinoma eventually develop liver metastases (mCRC) during the course of the disease [1]. Resection of liver metastases in patients without extrahepatic disease can improve overall survival (OS) rate with 5-year survival rates ranging from 30 % to 47 %, which is comparable to that of patients with stage III disease [2-5]. The use of neoadjuvant chemotherapy regimens has led to an increase in the proportion of patients eligible for surgery by downsizing the liver metastases and rendering unresectable lesions resectable [6, 7]. The advent of targeted therapies against VEGF and EGFR signalling has further raised response rates, progression-free survival (PFS) and OS in mCRC, when combined with conventional chemotherapy regimens [8, 9]. As response rates have improved, the proportion of patients eligible for curative resection of liver metastases has also increased [6].

The recombinant humanized monoclonal immunoglobulin G antibody bevacizumab (Avastin) inhibits angiogenesis by neutralizing all isoforms of human VEGF and blocking their binding to VEGF receptors [10]. It is the antiangiogenic agent with the largest body of evidence in the treatment of mCRC [11]. The treatment response to bevacizumab is incompletely captured by the conventional size-based RECIST criteria (Response Evaluation Criteria in Solid Tumors) [12, 13]. These criteria were developed for assessing volume reduction of tumours following cytotoxic therapy, whereas these new targeted therapies are rather cytostatic. As such, new imaging modalities for rapid and effective identification of treatment (non)response seem necessary [14–16].

¹⁸F-FDG PET imaging has an established role in the staging and restaging of CRC and several studies have also suggested a potential role in the assessment of treatment response [17–19]. Reliable quantification of this ¹⁸F-FDG PET-derived treatment response remains a topic of ongoing debate [20]. The maximum standardized uptake value (SUV_{max}) , which is the activity concentration in the hottest voxel normalized to administered activity and patient weight, is widely used and has proven its usefulness in various settings of response assessment [21]. However, there are several issues concerning its use, particularly its lack of robustness in terms of image noise, reconstruction methods, scanner sensitivity and partial volume effects (PVE) [22]. Another attractive and more comprehensive parameter is total lesion glycolysis (TLG), which is the product of the mean SUV and the volume of a lesion [23]. Though less susceptible to noise, significant variability in TLG values can be introduced by tumour delineation methods as well as PVE, which plays an important role in response assessment because of variations in lesion size [20].

Recently our group introduced the concept of standardized added metabolic activity (SAM) [24]. This parameter is a measure of the total excess tumoral SUV above the tumour background whilst excluding PVE. It is calculated by drawing a volume of interest (VOI1) around the tumour and a larger VOI (VOI2) around VOI1. Subtracting the background activity in [VOI2-VOI1] from that in VOI1 yields SAM. These VOIs have to be placed around the tumoral lesions at a sufficient distance from the actual tumour border to avoid PVE. As such, the method also avoids the variability which can be introduced by manual tumour delineation or lesion segmentation algorithms [24]. As correctly stated by Fleming et al. in their letter to the editor [25], the equation used to derive SAM is essentially the same as the specific uptake size index (SUSI) method, which was introduced and tested on planar and SPECT scintigraphy [26].

In this hypothesis-generating study, the potential role of ¹⁸F-FDG PET imaging in response assessment was evaluated in patients with mCRC and potentially resectable liver metastases, who underwent a neoadjuvant bevacizumab/ chemotherapy treatment. We investigated the value of morphological as well as metabolic parameters (including SAM) in predicting therapy outcome.

Materials and methods

Patient selection, treatment and follow-up

This study was approved by the Ethics Committee of Ghent University Hospital. Included in the study were 19 patients with a diagnosis of mCRC and potentially resectable liver metastases. All patients provided written informed consent prior to inclusion. Before surgical resection of the liver metastases, patients were treated with either FOLFOX6 (oxaliplatin 100 mg/m²/2 h, leucovorin 400 mg/m²/2 h, 5-FU bolus 400 mg/m²/10 min and 5-FU continuous infusion 2,400 mg/m²/46 h) or FOLFIRI (irinotecan 180 mg/m²/90 min, leucovorin 400 mg/m²/2 h, 5-FU bolus 400 mg/m²/10 min and 5-FU continuous infusion 2,400 mg/m²/46 h). Both regimens were combined with bevacizumab (Avastin, 5 mg/kg). Altogether five cycles of chemotherapy with a cycle duration of 14 days were administered. The fifth cycle was administered without bevacizumab in order to avoid adverse surgical events such as impaired wound healing or bleeding complications.

Liver resection was performed at a median of 48 days (interquartile range, IQR, 38–55 days) after the last cycle of chemotherapy in 16 patients. Surgical eligibility was assessed by a multidisciplinary team, whereas the type and the extent of liver resection was decided by the hepatobiliary surgeon. Postoperatively, seven additional cycles of adjuvant chemotherapy (without bevacizumab) were administered. Two patients were not eligible for liver surgery and were directed to other treatments. As they did not receive the same treatment, they were excluded from the survival analysis.

The PFS was measured from the start date of treatment until the date of progressive or recurrent disease. The OS was measured from the start date of chemotherapy until death. The close-out date of the study was set at 30 September 2012. The patients who did not reach the specific end-point (progression/death) by the close-out date as well as the patients who were lost to follow-up, were censored at the date of their last visit.

¹⁸F-FDG PET/CT image acquisition and reconstruction

¹⁸F-FDG PET/CT scans were performed before the start of neoadjuvant chemotherapy and 15 days after the last cycle. All examinations were performed according to the standard FDG PET/CT protocol at the Department of Nuclear Medicine of Ghent University Hospital. The patients fasted for at least 6 h prior to the investigation and their blood glucose level was tested to ensure normoglycaemia. The patients then received 3.7-5.5 MBq/kg (0.10-0.15 mCi/kg) of 18 F-FDG intravenously followed by 250 ml of sodium chloride and 20 mg of furosemide. Image acquisition was started 60 min after injection using an integrated PET/CT scanner (Philips Gemini PET/CT, Philips Medical Systems) which consists of a gadolinium oxyorthosilicate full-ring PET scanner with 5-mm spatial resolution and a 16-slice helical CT scanner. First a "whole-body" CT scan was performed (from the top of the skull to the proximal third of the femora). PET scanning was done immediately after acquisition of the CT images without moving the patient. The patients were instructed to breathe normally during the acquisition of CT and PET images. The standard software recommended by the manufacturer was used for acquisition and processing. The PET images were reconstructed using a three-dimensional row action maximum likelihood algorithm (3D-RAMLA) as provided by the manufacturer. The images were corrected for decay, scatter, random events and attenuation.

Image analysis

The morphological treatment response of the liver metastases was assessed by an experienced radiologist by applying the revised RECIST, version 1.1 [27]. According to these criteria, treatment responses were categorized into four categories: complete response (CR), partial response (PR), stable disease (SD) and progressive disease (PD). The "metabolic" treatment response was evaluated by quantifying the FDG uptake of the liver metastases on both the baseline as well as the follow-up PET images. These image analyses were performed by an experienced nuclear medicine physician using PMOD software (version 3.1; PMOD Technologies Ltd). VOIs were drawn around all liver metastases using an isocontour threshold method with the threshold set at 2 standard deviations above the mean SUV in healthy liver parenchyma. The SUV_{max} of the liver metastases was determined in the voxel with the highest measured activity concentration (becquerels per millilitre) and was calculated by dividing this activity concentration by the injected dose normalized to the body weight (becquerels per gram). If the liver lesions had disappeared on the follow-up images, the VOI of the most metabolically active lesion was copied from the baseline image and placed carefully in the same location. The SUV_{max} in this VOI was used as the follow-up value.

The rationale behind calculating SAM values has been described elsewhere [24] and was also used previously by Fleming et al. for planar and SPECT scintigraphy, as mentioned above [26]. A first VOI (VOI1) was drawn around all metastatic liver lesions. A second VOI (VOI2) was delineated around VOI1 directed to a small zone of homogeneous background. The borders of VOI1 were set at a reasonable distance from the tumour lesions in order to avoid PVE and to ensure that no spill-over from the metastatic lesion to VOI2 occurred. An example is shown in Fig. 1. Subsequently, the mean background activity (BG) was derived for all lesions using the following formula: mean BG=(total SUV VOI2 - total SUV VOI1)/(volume VOI2 - volume VOI1) in which total SUV is the product of the mean SUV and the respective volume. SAM was then calculated using the following formula: SAM = total SUV VOI1 – (mean BG \times volume VOI1). The obtained lesional SAMs were summed to produce one SAM value per scan, which represented the total metabolic burden of all liver metastases. These cumulative SAM values were used for further analysis.



Fig. 1 ¹⁸F-FDG PET scan in a patient with mCRC. A volume of interest (VOI1 *yellow*) is set at a reasonable distance from the liver lesion in order to avoid PVE. A second VOI (VOI2 *blue*) is drawn around VOI1. SAM is calculated by subtracting the background activity in [VOI 2-VOI1] from the total activity in VOI1

Statistical analysis

Statistical analysis was performed using SPSS version 19.0. Because of the sample size and the associated non-normality of the data, nonparametric tests were used. An independent Mann-Whitney U test and Wilcoxon's rank sum test were used to evaluate the differences in SUV_{max} and SAM values between RECIST responders and nonresponders. A paired Wilcoxon signed ranks test was used to evaluate the differences in PET parameters between baseline and follow-up scans. PFS and OS estimates were calculated using Kaplan-Meier survival analysis and the log-rank test was used to evaluate the significance of the differences between the acquired survival curves. A p value <0.05 was deemed significant for all hypothesis tests.

Results

Patient characteristics

Patient characteristics are presented in Table 1. The mean age of the patients included was 61 years and 63 % were men. In spite of initial suspicion, one patient appeared to have only extrahepatic metastases and was excluded from all further data analysis. The remaining 18 patients had a median of 3.5 liver metastases (IQR 1-9.25). Of these 18 patients, 12 received the FOLFOX6 and bevacizumab regimen before liver surgery, and 6 were treated with FOLFIRI and bevacizumab. Despite this treatment, two patients were still not suitable candidates for liver surgery and were directed to other treatments. As they did not receive the same treatment, they were excluded from the survival analysis. The median PFS and OS in the remaining population of 16 patients were 10.9 months and 48 months, respectively. One patient was still disease-free at the close-out date, five patients had relapsed but were still alive and one patient was lost to follow-up. These patients were censored at their last visit (for PFS and/or OS, if applicable). The median follow-up time was 53 months.

FDG PET and RECIST-defined treatment response

According to the revised RECIST criteria, eight patients showed a PR (44.4 %), nine SD (50 %) and one PD (5.6 %). When dichotomized into responders (PR) and non-responders (SD and PD), the median number of liver metastases was 3.5 in both groups (p=0.57). In contrast, both baseline SUV_{max} and baseline SAM values proved to be significantly different between morphological responders and nonresponders. The median baseline SUV_{max} value in

Table 1 Patient and tumour characteristics of the 19 patients with mCRC

Characteristic	Value
Gender, <i>n</i> (%)	
Men	12 (63)
Women	7 (37)
Age (years), median	61
Primary tumour, n (%)	
Colon	16 (84)
Rectum	3 (16)
Synchronicity of liver metastases, n (%)	
Synchronous	7 (37)
Metachronous	11 (58)
No liver metastasis ^a	1 (5)
Previous chemotherapy for metastatic disease, n (%)	
FOLFIRI	1 (5)
FOLFOX	3 (16)
Xeloda	1 (5)
FOLFOX + panitumumab	1 (5)
None	13 (68)
Chemotherapy before liver resection, n (%)	
FOLFOX + bevacizumab	12 (63)
FOLFIRI + bevacizumab	6 (32)
Postoperative chemotherapy for liver metastases, n (%)	
FOLFOX	7 (37)
FOLFIRI	4 (21)
None	5 (26)
No liver resection	3 (16)
Number of liver metastases, median	3.5

^a Screening failure; this patient had only extrahepatic metastases and was excluded from further analysis.

responders was 3.8 (IQR 2.9–5.6) and 7.2 (IQR 4.7–8.7) in nonresponders (p=0.021). The median baseline SAM value in responders was 34 (IQR 15–59) and 211 (IQR 68–1139) in nonresponders (p=0.002). The follow-up SUV_{max} and SAM values as well as the difference between the baseline and follow-up values (Δ SUV_{max} and Δ SAM) were not significantly correlated with the RECIST response.

RECIST-defined treatment response and survival

The Kaplan-Meier estimates for the median PFS were almost identical for patients with and without a morphological response: 10.9 and 10.8 months, respectively (p=0.93). The median OS in patients with a morphological response was 51 months and in nonresponders was 39 months, but this difference was not statistically significant (p=0.24). The survival curves are shown in Fig. 2.



Fig. 2 Kaplan-Meier survival curves for PFS and OS in morphological responders and nonresponders (according to the RECIST criteria)

FDG PET-defined treatment response and survival

The median SUV_{max} and SAM values at baseline were 5.2 (IQR 3.6–8.4) and 67 (IQR 37–469). Decreases in SUV_{max} and SAM were seen after treatment (median decreases 25.3 % and 94.5 %; p=0.033 and 0.003, respectively). The median and mean values as well as the distribution of the various PET parameters are presented in Table 2.

The median PFS and OS were similar in patients with SUV_{max} above and below the median of 5.2: 11.4 vs. 10.4 months (p=0.81) and 47 vs. 48 months (p=0.88), respectively. In contrast, a low follow-up SUV_{max} had a beneficial effect on both PFS and OS. Patients with a follow-up SUV_{max} higher than 2.85 had a median PFS of 10.4 months compared with 14.7 months in those with a low follow-up SUV_{max}. Patients with a high follow-up SUV_{max} had a median OS of 32 months, whereas the median OS had not been reached in those with a low follow-up SUV_{max}. This survival benefit was significant for both PFS (p=0.01) and OS (p=0.003). Survival curves stratified by follow-up SUV_{max} are shown in Fig. 3. On the other hand, the difference between baseline and follow-up SUV_{max} values

(Δ SUV_{max}) appeared to have no prognostic value. The median PFS and OS in patients with Δ SUV_{max} above and below the median of 26.9 % were 10.2 vs. 10.9 months (p=0.39) and 47 vs. 51 months (p=0.51), respectively. Recently, Heijmen et al. found a coefficient of repeatability of SUV_{max} of 33.3 % in untreated liver metastases [28]. Using this value as a cut-off, similar results were found: the median PFS and OS in patients with Δ SUV_{max} above and below 33.3 % were 11.4 vs. 10.9 months (p=0.73) and 48 vs. 51 months (p=0.47), respectively.

The baseline SAM was not significantly correlated with PFS (p=0.38) or OS (p=0.88), as the median PFS and OS estimates were almost identical between those with a baseline SAM above and below 65 (11.4 vs. 10.4 months for PFS and 47 vs. 48 months for OS, respectively). In contrast, both follow-up SAM and Δ SAM values appeared to be significantly correlated with PFS and OS. When dichotomized according to their median values, the patients were categorized into identical groups. The group with a high follow-up SAM and a low Δ SAM had a median PFS of 9.4 months, and the other group had a median PFS of 14.7 months (p=0.002). The median OS in the former group

Table 2 Parameter values and	-
distributions for the entire]
patient population ($n=18$; two	-
patients were eventually]
excluded from survival]
analysis as they did not	,
receive the same treatment)	2

Parameter	Median	First quartile	Third quartile	Mean	Standard deviation
Baseline SUV _{max}	5.2	3.6	8.4	5.8	2.4
Follow-up SUV _{max}	3.0	2.4	5.9	4.1	2.6
ΔSUV_{max} (%)	25.3	6.2	60.9	16.7	59.2
Baseline SAM	67.5	36.5	468.8	376.9	605.2
Follow-up SAM	4.6	0	54	118	264.2
ΔSAM (%)	94.5	32.4	100	47	86.4



Fig. 3 Kaplan-Meier survival curves for PFS and OS in patients with a low follow-up SUV_{max} versus patients with a high follow-up SUV_{max}

was 32 months, whereas the median OS had not been reached in the latter group (p=0.002). The corresponding survival curves are shown in Fig. 4.

Discussion

Lower baseline SAM and SUV_{max} were associated with a potential morphological response (defined according to RECIST criteria). A correlation between baseline SUV_{max} and morphological response was also seen by Byström et al. in a similar setting [29]. On the other hand, baseline metabolic parameters as well as morphological response did not predict PFS or OS. The finding that baseline PET imaging had no prognostic value may appear to contradict previous

results of Riedl et al. [30], who found a strong correlation between baseline FDG uptake values in liver metastases before metastasectomy and survival. This may most likely be explained by the fact that the patients in the present study received several cycles of chemotherapy and bevacizumab before liver resection, whereas the patients in the study by Riedl et al. were immediately directed to surgery.

The weak prognostic value of RECIST-defined morphological response in this study can possibly be explained by the presence of bevacizumab as an antiangiogenic agent in our treatment regimen. Despite the proven survival benefit of addition of bevacizumab to classic chemotherapy regimens [8], no increase in RECIST-defined response could be demonstrated by adding bevacizumab [13]. Although the RECIST criteria are a well-validated and widely used



Fig. 4 Kaplan-Meier survival curves for PFS and OS in patients with a low follow-up SAM and a high Δ SAM versus patients with a high follow-up SAM and a low Δ SAM. When the patients were



dichotomized according to the median values of follow-up SAM and Δ SAM, identical patient groups were formed

instrument for assessment of the response of solid tumours to classic cytotoxic chemotherapy, new antiangiogenic and other targeted therapies are putting pressure on these conventional size-dependent criteria. The effects of these new therapeutic modalities are considered predominantly cytostatic rather than cytotoxic and beneficial patient outcomes can be seen without an initial change in lesion size [14, 15, 31].

Follow-up SUV_{max} and SAM values were not correlated with morphological response, but proved to be significant prognostic factors for PFS as well as OS. This finding adds to the existing evidence about the prognostic power of ¹⁸F-FDG PET imaging in assessing treatment response of mCRC [18, 19, 32–34] and suggests a potential role for a PET-guided treatment algorithm in the management of these patients. Further research is warranted in this field.

While high follow-up SUV_{max} proved to be an adverse prognostic factor, the ΔSUV_{max} between baseline and follow-up studies did not have any prognostic value. In contrast, the reduction in total metabolic tumour burden as assessed by Δ SAM proved to be significantly related to PFS and OS. The reason for this discrepancy is probably twofold. On the one hand, SUV_{max} is more prone to variability caused by several factors such as image noise and resolution, reconstruction methods, scanner sensitivity and PVE. It is determined in the hottest voxel, which, by definition, means that the value will be an outlier [22]. In a recent study, Heijmen et al. demonstrated a coefficient of repeatability of SUV_{max} of 33.3 % in untreated liver metastases [28], suggesting that ΔSUV_{max} should at least be of this magnitude to represent a true treatment response. However, dichotomizing the present patient population using 33.3 % as a cut-off ΔSUV_{max} yielded comparable results: no significant difference in PFS or OS was found between the high and low ΔSUV_{max} group.

On the other hand, SUV_{max} does not entirely capture the treatment response to therapy as it only reflects the metabolic activity per gram of tissue in one voxel and does not take into account the total tumour metabolic load [35]. SAM is a marker derived from TLG which takes into account metabolic activity as well as lesion volume [23]. In contrast to TLG, SAM uses a background subtraction, assuming that tumoral tissue develops on top of normal tissue. As such, the SAM method also avoids the problems of lesion segmentation. Moreover, when the necessary VOIs are placed at a sufficient distance from the actual tumour border, PVE are avoided, which makes accurate response assessment in small lesions possible [24].

Some limitations of this study should be addressed. First, the study population was rather small. Nevertheless, the significant p values in this hypothesis-generating study justify further evaluation in a larger patient population. Second, the follow-up PET scans were performed after completion of

preoperative chemotherapy. Therefore, no conclusions can be made about the potential of PET imaging in picking up an early (non)response to treatment. Early identification of a nonresponder could reduce the side effects and costs of futile treatment as well as allow selection of patients who would be better directed immediately to surgery or secondline treatment. Further research is needed to clarify the potential value of "interim" PET in the treatment of mCRC.

In the survival analysis of this small patient population, SAM and SUV_{max} values were dichotomized according to the median value in order to obtain two patient groups of comparable size. Further research in a larger patient population could identify the optimal cut-off value to separate the patients in terms of prognosis.

Regarding the calculation of SAM, we have to consider potential drawbacks of the presented methodology. As SAM includes background correction, the values obtained might be less reliable in the presence of an irregular background. However, the drawback of an irregular background did not apply to liver lesions in the setting of this study as diffuse liver involvement (and the associated heterogeneous FDG uptake) would have obviated the need for neoadjuvant chemotherapy and subsequent resection of liver metastases. Nevertheless, further research is needed to identify the repeatability of the SAM method in liver metastases as has been performed recently for metabolic volume and TLG [28].

Finally, SAM values were compared with SUV_{max} values as the latter is the most widely used parameter in the assessment of treatment response by PET imaging. It cannot be excluded that metabolic volume and SUV_{mean} , provided they are estimated using robust and repeatable lesion segmentation techniques [36], may provide similar or additional prognostic information to that obtained by SAM.

Conclusion

¹⁸F-FDG PET imaging could be a useful tool to assess the treatment response and predict the clinical outcome in patients with mCRC who undergo preoperative chemotherapy before liver metastasectomy. Both follow-up SUV_{max} and SAM as well as Δ SAM were found to be significant prognostic factors for PFS and OS in this hypothesis-generating study. Further research is warranted to confirm these findings in a larger population as well as to explore to what extent ¹⁸F-FDG PET imaging might guide treatment in patients with mCRC.

Acknowledgments This work was supported by an unrestricted educational grant from Roche and by the Belgian National Cancer Plan (NKP29/026).

Conflicts of interest M.P. received an educational grant from Roche. The other authors disclose no conflicts.

References

- Scheele J, Stangl R, Altendorf-Hofmann A. Hepatic metastases from colorectal carcinoma: impact of surgical resection on the natural history. Br J Surg. 1990;77:1241–6.
- Morris EJ, Forman D, Thomas JD, Quirke P, Taylor EF, Fairley L, et al. Surgical management and outcomes of colorectal cancer liver metastases. Br J Surg. 2010;97:1110–8. doi:10.1002/bjs.7032.
- Simmonds PC, Primrose JN, Colquitt JL, Garden OJ, Poston GJ, Rees M. Surgical resection of hepatic metastases from colorectal cancer: a systematic review of published studies. Br J Cancer. 2006;94:982–99. doi:10.1038/sj.bjc.6603033.
- Wei AC, Greig PD, Grant D, Taylor B, Langer B, Gallinger S. Survival after hepatic resection for colorectal metastases: a 10year experience. Ann Surg Oncol. 2006;13:668–76. doi:10.1245/ ASO.2006.05.039.
- Cummings LC, Payes JD, Cooper GS. Survival after hepatic resection in metastatic colorectal cancer: a population-based study. Cancer. 2007;109:718–26. doi:10.1002/cncr.22448.
- Nordlinger B, Van Cutsem E, Rougier P, Kohne CH, Ychou M, Sobrero A, et al. Does chemotherapy prior to liver resection increase the potential for cure in patients with metastatic colorectal cancer? A report from the European Colorectal Metastases Treatment Group. Eur J Cancer. 2007;43:2037–45. doi:10.1016/ j.ejca.2007.07.017.
- Chaudhury P, Hassanain M, Bouganim N, Salman A, Kavan P, Metrakos P. Perioperative chemotherapy with bevacizumab and liver resection for colorectal cancer liver metastasis. HPB (Oxford). 2010;12:37–42. doi:10.1111/j.1477-2574.2009.00119.x.
- Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med. 2004;350:2335–42. doi:10.1056/NEJMoa032691.
- Van Cutsem E, Kohne CH, Lang I, Folprecht G, Nowacki MP, Cascinu S, et al. Cetuximab plus irinotecan, fluorouracil, and leucovorin as first-line treatment for metastatic colorectal cancer: updated analysis of overall survival according to tumor KRAS and BRAF mutation status. J Clin Oncol. 2011;29:2011–9. doi:10.1200/ JCO.2010.33.5091.
- Shih T, Lindley C. Bevacizumab: an angiogenesis inhibitor for the treatment of solid malignancies. Clin Ther. 2006;28:1779–802. doi:10.1016/j.clinthera.2006.11.015.
- Edwards MS, Chadda SD, Zhao Z, Barber BL, Sykes DP. A systematic review of treatment guidelines for metastatic colorectal cancer. Colorectal Dis. 2012;14:e31–47. doi:10.1111/j.1463-1318.2011.02765.x.
- Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). Eur J Cancer. 2009;45: 228–47.
- Klinger M, Eipeldauer S, Hacker S, Herberger B, Tamandl D, Dorfmeister M, et al. Bevacizumab protects against sinusoidal obstruction syndrome and does not increase response rate in neoadjuvant XELOX/FOLFOX therapy of colorectal cancer liver metastases. Eur J Surg Oncol. 2009;35:515–20. doi:10.1016/ j.ejso.2008.12.013.
- Desar IM, van Herpen CM, van Laarhoven HW, Barentsz JO, Oyen WJ, van der Graaf WT. Beyond RECIST: molecular and functional imaging techniques for evaluation of response to targeted therapy. Cancer Treat Rev. 2009;35:309–21. doi:10.1016/ j.ctrv.2008.12.001.
- Sharma MR, Maitland ML, Ratain MJ. RECIST: no longer the sharpest tool in the oncology clinical trials toolbox – point. Cancer Res. 2012;72:5145–9. doi:10.1158/0008-5472.CAN-12-0058.

- 16. de Langen AJ, van den Boogaart V, Lubberink M, Backes WH, Marcus JT, van Tinteren H, et al. Monitoring response to antiangiogenic therapy in non-small cell lung cancer using imaging markers derived from PET and dynamic contrast-enhanced MRI. J Nucl Med. 2011;52:48–55. doi:10.2967/jnumed.110.078261.
- de Geus-Oei LF, Vriens D, van Laarhoven HW, van der Graaf WT, Oyen WJ. Monitoring and predicting response to therapy with 18F-FDG PET in colorectal cancer: a systematic review. J Nucl Med. 2009;50 Suppl 1:43S–54S. doi:10.2967/jnumed.108.057224.
- Vriens D, van Laarhoven HW, van Asten JJ, Krabbe PF, Visser EP, Heerschap A, et al. Chemotherapy response monitoring of colorectal liver metastases by dynamic Gd-DTPA-enhanced MRI perfusion parameters and 18F-FDG PET metabolic rate. J Nucl Med. 2009;50:1777–84. doi:10.2967/jnumed.109.064790.
- Hendlisz A, Golfinopoulos V, Garcia C, Covas A, Emonts P, Ameye L, et al. Serial FDG-PET/CT for early outcome prediction in patients with metastatic colorectal cancer undergoing chemotherapy. Ann Oncol. 2012;23:1687–93. doi:10.1093/annonc/ mdr554.
- Visvikis D, Hatt M, Tixier F, Cheze Le Rest C. The age of reason for FDG PET image-derived indices. Eur J Nucl Med Mol Imaging. 2012;39:1670–2. doi:10.1007/s00259-012-2239-0.
- Wahl RL, Jacene H, Kasamon Y, Lodge MA. From RECIST to PERCIST: evolving considerations for PET response criteria in solid tumors. J Nucl Med. 2009;50 Suppl 1:122S–50S.
- Boellaard R, Krak NC, Hoekstra OS, Lammertsma AA. Effects of noise, image resolution, and ROI definition on the accuracy of standard uptake values: a simulation study. J Nucl Med. 2004;45:1519–27.
- 23. Larson SM, Erdi Y, Akhurst T, Mazumdar M, Macapinlac HA, Finn RD, et al. Tumor treatment response based on visual and quantitative changes in global tumor glycolysis using PET-FDG imaging. The visual response score and the change in total lesion glycolysis. Clin Positron Imaging. 1999;2:159–71.
- 24. Mertens J, Dobbeleir A, Ham H, D'Asseler Y, Goethals I, Van de Wiele C. Standardized added metabolic activity (SAM): a partial volume independent marker of total lesion glycolysis in liver metastases. Eur J Nucl Med Mol Imaging. 2012;39:1441–8. doi:10.1007/s00259-012-2166-0.
- 25. Fleming JS, Tossici-Bolt L, Guy M, Kemp P. Comment on Mertens et al.: standardized added metabolic activity (SAM): a partial volume independent marker of total lesion glycolysis in liver metastases. Eur J Nucl Med Mol Imaging. 2013;40:788–9. doi:10.1007/s00259-013-2364-4.
- Fleming JS, Bolt L, Stratford JS, Kemp PM. The specific uptake size index for quantifying radiopharmaceutical uptake. Phys Med Biol. 2004;49:N227–34.
- Bogaerts J, Ford R, Sargent D, Schwartz LH, Rubinstein L, Lacombe D, et al. Individual patient data analysis to assess modifications to the RECIST criteria. Eur J Cancer. 2009;45:248–60.
- Heijmen L, de Geus-Oei LF, de Wilt JH, Visvikis D, Hatt M, Visser EP, et al. Reproducibility of functional volume and activity concentration in 18F-FDG PET/CT of liver metastases in colorectal cancer. Eur J Nucl Med Mol Imaging. 2012;39:1858–67. doi:10.1007/s00259-012-2233-6.
- 29. Bystrom P, Berglund A, Garske U, Jacobsson H, Sundin A, Nygren P, et al. Early prediction of response to first-line chemotherapy by sequential [18F]-2-fluoro-2-deoxy-D-glucose positron emission tomography in patients with advanced colorectal cancer. Ann Oncol. 2009;20:1057–61. doi:10.1093/annonc/mdn744.
- Riedl CC, Akhurst T, Larson S, Stanziale SF, Tuorto S, Bhargava A, et al. 18F-FDG PET scanning correlates with tissue markers of poor prognosis and predicts mortality for patients after liver resection for colorectal metastases. J Nucl Med. 2007;48:771–5. doi:10.2967/jnumed.106.037291.

- 31. Avril NE, Weber WA. Monitoring response to treatment in patients utilizing PET. Radiol Clin North Am. 2005;43:189–204.
- 32. Dimitrakopoulou-Strauss A, Strauss LG, Burger C, Ruhl A, Irngartinger G, Stremmel W, et al. Prognostic aspects of 18F-FDG PET kinetics in patients with metastatic colorectal carcinoma receiving FOLFOX chemotherapy. J Nucl Med. 2004;45:1480–7.
- 33. de Geus-Oei LF, van Laarhoven HW, Visser EP, Hermsen R, van Hoorn BA, Kamm YJ, et al. Chemotherapy response evaluation with FDG-PET in patients with colorectal cancer. Ann Oncol. 2008;19:348–52.
- Dimitrakopoulou-Strauss A, Strauss LG, Rudi J. PET-FDG as predictor of therapy response in patients with colorectal carcinoma. Q J Nucl Med. 2003;47:8–13.
- Weber WA, Figlin R. Monitoring cancer treatment with PET/ CT: does it make a difference? J Nucl Med. 2007;48 Suppl 1:36S-44S.
- Hatt M, Cheze Le Rest C, Albarghach N, Pradier O, Visvikis D. PET functional volume delineation: a robustness and repeatability study. Eur J Nucl Med Mol Imaging. 2011;38:663–72. doi:10.1007/ s00259-010-1688-6.