

First preclinical evaluation of mono-[¹²³I]iodohypericin as a necrosis-avid tracer agent

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Abstract. *Purpose:* We have labelled hypericin, a polyphenolic polycyclic quinone found in St. John's wort (*Hypericum perforatum*), with ¹²³I and evaluated mono-[¹²³I]iodohypericin (MIH) as a potential necrosis-avid diagnostic tracer agent.

Methods: MIH was prepared by an electrophilic radioiodination method. The new tracer agent was evaluated in animal models of liver infarction in the rat and heart infarction in the rabbit using single-photon emission computed tomography (SPECT), triphenyltetrazolium chloride (TTC) histochemical staining, serial sectional autoradiography and microscopy, and radioactivity counting techniques.

Results: Using in vivo SPECT imaging, hepatic and cardiac infarctions were persistently visualised as well-defined hot spots over 48 h. Preferential uptake of the tracer agent in necrotic tissue was confirmed by perfect match of images from post-mortem TTC staining, autoradiography (ARX) and histology. Radioactivity concentration in infarcted tissues was over 10 times (liver; 3.51% ID/g in necrotic tissue vs 0.38% ID/g in normal tissue at 60 h p.i.) and over 6 times (myocardium; 0.36% ID/g in necrotic tissue vs 0.054% ID/g in normal tissue; ratios up to 18 for selected parts on ARX images) higher than in normal tissues.

Conclusion: The results suggest that hypericin derivatives may serve as powerful necrosis-avid diagnostic agents for assessment of tissue viability.

Keywords: Hypericin – SPECT imaging – Autoradiography – Radioiodine – Necrosis

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Introduction

Non-invasive “hot spot imaging” and localisation of necrotic tissue may be helpful for the diagnosis of different disorders. This is especially true in the case of ischaemic myocardial injury, where imaging using necrosis-avid agents could help in measuring infarct size. This could allow the rapid use of interventions for myocardial salvage through early identification of the acute event, thereby improving the clinical outcome. Also, other cardiovascular disorders associated with cardiac cell death could be visualised and diagnosed, such as cardiomyopathies, myocarditis and heart transplant rejection [1].

In nuclear medicine, only a limited number of hot spot imaging agents have been used or proposed for diagnosis of necrosis. ^{99m}Tc-pyrophosphate was the first of them and is supposed to bind to necrotic myocardium by targeting calcium phosphate deposited in the mitochondria of infarcted or severely injured myocardial tissue [2]. However, ^{99m}Tc-pyrophosphate scintigraphy has never gained widespread use because of its limited diagnostic accuracy and the relatively poor stability of the tracer agent, which may lead to the in vivo formation of pertechnetate and so result in imaging of the cardiac chambers. More recently, ¹¹¹In-labelled murine monoclonal antimyosin Fab antibody fragments were introduced for infarct-avid scintigraphy [3, 4]. This so-called ¹¹¹In-antimyosin has a selective affinity for the intracellular heavy chain of cardiac myosin, which is exposed when the integrity of the sarcolemma is lost as a result of cell damage [5, 6]. It is incorporated in the necrotic myocardium in an inverse relationship to regional flow, with maximum uptake in areas with severe flow impairment, although its uptake is more intense in myocardial

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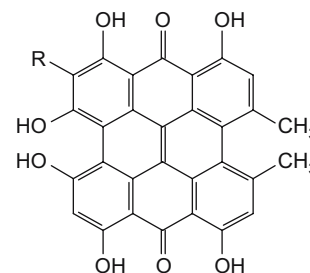
infarcts with reperfusion than in those with persistent coronary occlusion [7]. It was also found useful for detection of acute or chronic diffuse myocardial damage in allograft rejection in cardiac transplantation [8, 9], doxorubicin-induced cardiotoxicity [10], acute myocarditis [11, 12] and various cardiomyopathies [13, 14]. Although ^{111}In -antimyosin has been approved by the FDA for clinical use, the limitation of approved indications to ischaemic heart disease has resulted in cessation of commercial production.

A few years ago, it was observed that the complex of $^{99\text{m}}\text{Tc}$ with glucaric acid, a simple dicarboxylic acid sugar, localised in canine reperfused myocardial infarction soon after injection [15]. The tracer agent is supposed to bind to positively charged histones within disintegrated nuclei and reduced subcellular organelle proteins in necrotic myocytes [16]. It has the advantages of a rapid blood pool clearance, a good target to background ratio, and lack of toxicity and antigenicity, and is claimed to have a good sensitivity and specificity for detection of early irreversible myocyte injury. On the other hand, its uptake is limited to the first 9 h after the onset of acute myocardial infarction because of the relatively rapid disintegration of the positively charged histones. Further clinical trials will be required to reveal its real clinical usefulness.

In the field of magnetic resonance imaging (MRI), it has recently been reported that some porphyrin derivatives that were originally developed as tumour-seeking contrast agents [17] have avidity only for non-viable tissues (typically necrosis) instead of viable tumour cells and could therefore be re-categorised as necrosis-avid contrast agents [18–20]. The extraordinary targetability of these agents for necrosis has elicited novel applications for MRI visualisation of acute myocardial infarction [21–32] and therapeutic tissue ablation [33]. Some porphyrin derivatives are also useful as photosensitisers and tumour localisers for photodynamic therapy (PDT) [34]. PDT is a relatively recent modality for the treatment of cancer, which requires combination of a chemical photosensitiser that is preferentially taken up and retained by tumour cells and light of an appropriate wavelength to generate singlet oxygen free radicals out of the photosensitiser [34]. While photofrin-2 represents the commercially available porphyrin derivative that is most frequently used, hypericin (Fig. 1), a polyphenolic polycyclic quinone found in St. John's wort (*Hypericum perforatum*) is regarded as a powerful naturally occurring photosensitiser. It is being applied in PDT under the same principle as that for porphyrin derivatives [35].

Considering the role of hypericin and photofrin-2 in PDT and the recently recognised necrosis avidity of certain porphyrin-derived contrast agents, we hypothesised that hypericin may share the similar properties of preferential uptake and retention in necrotic tissues. For this reason, we have synthesised mono- ^{123}I iodohypericin (MIH), a ^{123}I -labelled derivative of hypericin (Fig. 1), and evaluated it in an imaging and histomorphological study designed to explore its potential utility as a necrosis-avid tracer agent.

Fig. 1. Chemical structure of hypericin (R=H) and mono- ^{123}I iodohypericin (MIH) (R= ^{123}I)



Materials and methods

Synthesis and preparation of mono- ^{123}I iodohypericin

MIH was synthesised in no-carrier-added form using a standard electrophilic radioiodination method in the presence of peracetic acid and was purified with high-pressure liquid chromatography (HPLC) as described previously [36]. After purification the MIH solution was evaporated to dryness at 60°C under a flow of nitrogen and the tracer agent was redissolved in water/polyethylene glycol (PEG) 400 (8:2, V/V) immediately before injection.

Animal models of necrosis

All animal experiments were conducted with the approval of the institutional ethical committee.

Reperfused hepatic infarction

Three adult Wistar rats weighing 400–450 g were anaesthetised with intraperitoneal injection of pentobarbital (Nembutal; Sanofi Santé Animale, Brussels, Belgium) at a dose of 40 mg/kg. Under laparotomy, reperfused hepatic infarction was induced by temporarily clamping the hilum of the right liver lobes for 3 h. After reperfusion by declamping hepatic inflow, the abdominal cavity was closed with two-layer sutures and the rats were allowed to recover for 8–24 h after the surgery [19, 37]. One rat underwent a sham operation and served as a non-infarcted control.

Occlusive myocardial infarction

A New Zealand adult rabbit weighing about 5 kg was sedated with intramuscular injection of a mixed solution of ketamine hydrochloride (Ketalar; Pfizer, Brussels, Belgium) at 15 mg/kg and xylazine hydrochloride (Rompun; Bayer, Leverkusen, Germany) at 2.5 mg/kg. The animal was then anaesthetised with an intravenous bolus injection of Nembutal at 15 mg/kg followed by i.v. infusion of Nembutal at 0.1 mg/kg per minute through an ear vein. Under a cold-light laryngoscope with no. 1 paediatric blade (Riester; Jungingen, Germany), the animal was intubated with a 3.5-mm endotracheal tube, which was then connected to an artificial ventilator (Mark 7 respirator; Bird Corporation, Palm Springs, CA, USA). Open chest surgery (left posterolateral thoracotomy) was performed. The pericardium was opened to expose the left circumflex coronary artery, which was then permanently ligated with a 3-0 silk suture. The chest was closed after evacuation of the pneumothorax.

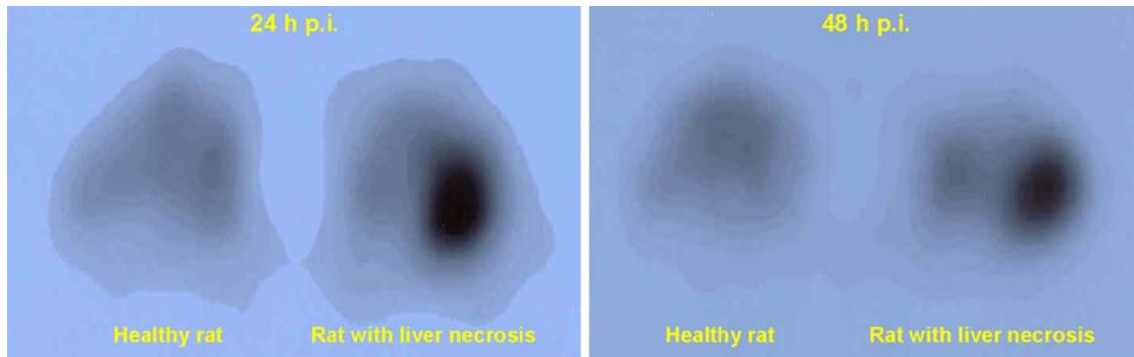


Fig. 2. SPECT transverse section images obtained in normal rats (*left*) and rats with reperused hepatic infarction (RHI, *right*) at 24 h and 48 h post injection of MIH. The persistent hyperactivity is found only in the infarcted right liver lobe of the rat RHI model, and not in the intact liver lobe of the rat RHI model or in the liver of the sham-operated rat (results not shown)

Tracer administration

Five hours after reperfusion of the hepatic infarction, the rats were intravenously injected with 18.5 MBq of MIH through a tail vein. Five hours after induction of the myocardial infarction, the rabbit was injected with 130 MBq MIH via an ear vein.

SPECT imaging

All imaging measurements were performed using a two-detector gamma camera (E-cam; Siemens Medical Systems, Erlangen, Germany). Each animal was secured to the head-holder of the patient bed with the organ of interest in the centre of the field of view. SPECT projections were collected using the following acquisition parameters: 120 projections over 360° per detector head (step and shoot mode), 25 s per projection, a matrix size of 256×256, a pixel size of 2.4 mm and a non-circular orbit. The images of the rats were reconstructed with filtered back projection, using a ramp filter and cut-off at the Nyquist frequency. The images of the rabbit were reconstructed with maximum-likelihood expectation-maximisation, applying 47 iterations.

In vivo SPECT was conducted under the same anaesthetic regimens as were used for surgery in both the rats and the rabbit. The rats were imaged consecutively during the first 2–3 h, and then 24 and 48 h after injection of MIH. One rat with liver lobe infarction and one sham-operated control rat were placed parallel supine and imaged simultaneously. The rabbit was imaged in a similar fashion but only 24 and 48 h after injection of MIH.

Animal sacrifice and tissue sampling

Forty-eight to 60 h post injection, the animals were sacrificed with an intravenous overdose of Nembutal. The rat liver and rabbit heart were excised. The partially necrotic part of the liver was cut into blocks at a thickness of about 1 cm, which were quickly frozen in isopentane cooled over dry ice to -40°C . The heart was cut perpendicular to the long axis into 5-mm-thick slices starting from the apex.

Histochemical staining

Representative blocks of liver tissue and the slices of heart were immersed in a buffered triphenyltetrazolium chloride (TTC) solution for staining during 10 min at 37°C . After this staining, normal liver and myocardial tissues appeared brick red and the necrotic areas were

unstained [22, 24, 27–32]. The TTC-stained specimens were digitally photographed and stored for later imaging–histochemical match.

Ex vivo autoradiography

The frozen liver and heart tissues, adjacent to the tissue sections for TTC staining, were cut at -20°C into 20- to 50- μm serial sections and exposed overnight to a high-performance storage phosphor screen (Super resolution screen; Canberra-Packard, Ontario, Canada). The screen was read using a Phosphor Imager scanner (Cyclone; Canberra-Packard). The images were analysed using Optiquant software (Canberra-Packard). Afterwards, the same sections were stained with haematoxylin-eosin (H&E) using the conventional procedure. These H&E-stained sections were digitally scanned and the obtained images were corrected for brightness and contrast.

Tissue radioactivity counting

Guided by TTC staining, the liver lobes of the rat and heart sections of the rabbit were sampled for normal and necrotic parts, weighed separately and counted for radioactivity using a 3-in. NaI(Tl)

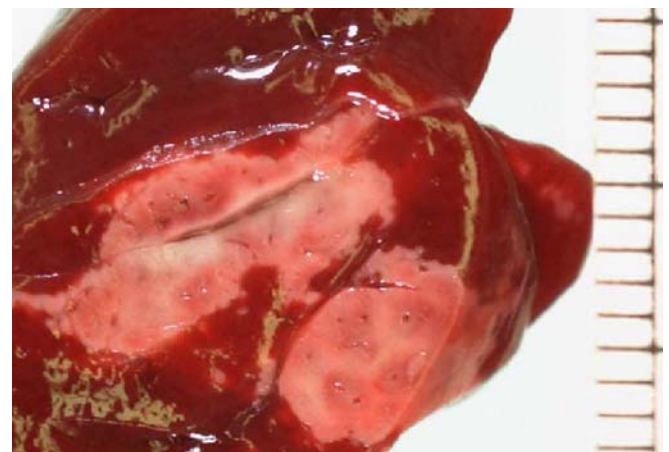


Fig. 3. TTC staining of the liver lobe with focal reperused infarction. From the 5-mm tissue block adjacent to the slice for autoradiography and histology (Fig. 4), the normal liver tissue (red) and necrotic tissue (pale) were sampled for radioactivity counting



Fig. 4. Comparison of autoradiography (*left*) and H&E staining without (*middle*) and with (*right*) enhanced contrast and brightness of the same 50- μ m frozen liver slice containing reperfused infarct from the rat

scintillation detector mounted in a sample changer (Wallac Wizard, Turku, Finland). Corrections were made for background radiation and physical decay during counting. The activity in the normal and infarcted parts of the liver and heart was then expressed as counts per gram tissue.

Results

Liver lobe infarction in rats

SPECT imaging During the first couple of hours after i.v. administration of MIH in both infarcted and normal control rats, a systemic distribution pattern appeared. Higher activity uptake was seen especially in the liver and kidneys as well as in the intestines and urinary bladder, indicating the major elimination pathways of the compound. In comparison with a complete outline of the liver in the normal rat, there was a filling defect at the lower right quadrant of the liver, corresponding to the induced liver infarction. At 24 h and 48 h post injection, on the transverse SPECT image, the uptake of radioactivity in the infarcted liver lobe was apparently much higher than that in the non-infarcted liver lobe and that in the liver of the

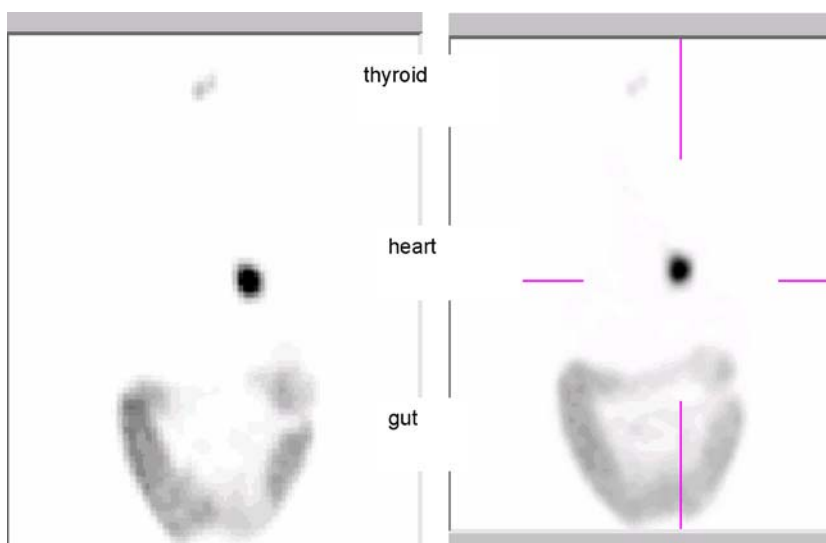
sham-operated rat (Fig. 2). Residual radioactivity was also shown in the regions of the thyroid and intestines.

Tissue radioactivity counting Sixty hours post injection of the tracer agent, the results of radioactivity quantification of dissected tissues showed high uptake in the necrotic liver lobe of 3.51% ID/g, which was almost 10 times the radioactivity concentration found in normal liver lobes (0.38% ID/g). Furthermore, the amount in the liver lobes of the control rat was comparable to the amount found in the untreated liver lobes of the rat with infarction (0.58% ID/g).

Histochemical staining Figure 3 shows a picture of a 1-cm-thick block of the partially necrotic liver of the rat. After staining with TTC, the reddish normal liver and adjacent pale necrotic tissue could be easily identified, as was confirmed by H&E staining of the 50- μ m serial sections taken from the tissue 5 mm next to this slice (Fig. 4).

Ex vivo autoradiography Figure 4 compares the images of autoradiography and H&E staining of the same 50- μ m sections. High tracer uptake appears only in the eosino-

Fig. 5. SPECT images (coronal section) obtained at 24 h (*left*) and 48 h (*right*) post injection of MIH in a rabbit with myocardial infarction. The radioactivity can be persistently found in the heart region and to a lesser extent in the thyroid and gut



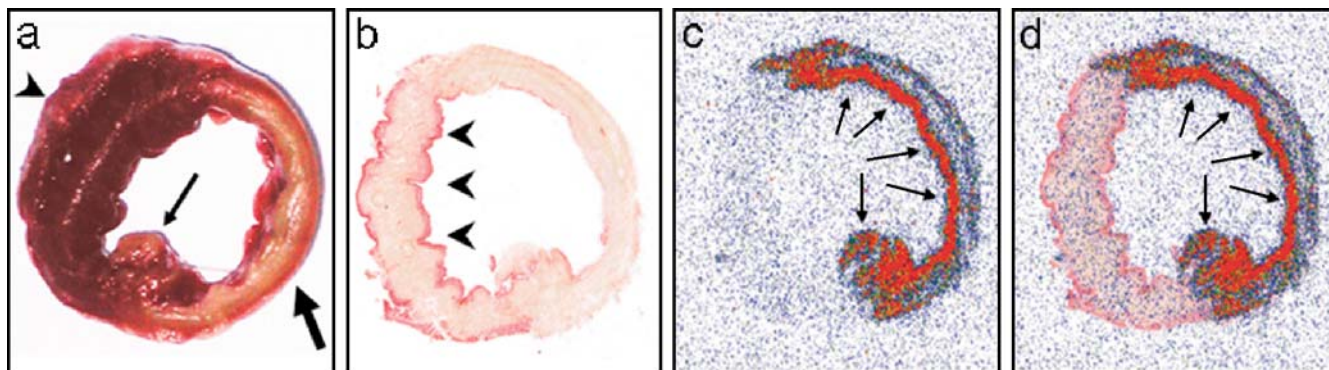


Fig. 6. Ex vivo images of rabbit heart. After TTC staining of a 5-mm-thick slice (**a**), a 50- μ m frozen slice (**b**, TTC stained) was obtained. The autoradiographic image (**c**) of this 50- μ m slice matched perfectly when overlapped with the image of TTC staining (**d**). **a** On the TTC-stained specimen with right ventricular wall attached (*arrowhead*), the transmural infarction (pale or negatively stained with TTC) involves the entire anterior and lateral wall of the left ventricle (*thick arrow*), including the posterior papillary muscle (*thin arrow*). **b** On the frozen slice with right ventricular wall

philic necrotic parts (pink coloured on H&E images), whereas low tracer uptake is found in the (haematoxylinophilic) healthy parts of the liver (purple coloured after H&E staining). The topographic distribution of the radioactivity matches perfectly with that of the necrotic tissue components, suggesting that MIH specifically accumulates in necrotic tissue.

Myocardial infarction in the rabbit

In vivo SPECT imaging Twenty-four and 48 h post injection of MIH in the rabbit with occlusive myocardial infarction, a coronal section view after iterative reconstruction of whole-body SPECT persistently displayed only one hot spot in the anatomical area of the heart. Radioactivity to a much lesser extent could also be seen in the regions of the thyroid and intestines (Fig. 5).

Tissue radioactivity counting TTC staining enabled selective sampling of reddish normal myocardium and whitish infarcted myocardium for radioactivity quantification. At 48 h post injection of the tracer agent, radioactivity concentration in necrotic heart tissue was 0.36% ID/g, which was about 6.6 times the radioactivity concentration found in normal myocardium (0.054% ID/g). The ratio would be even much higher if tissue sampling were to be guided by the outcome of autoradiography in this model of occlusive myocardial infarction (Fig. 6). On the basis of the autoradiographic images, an 18-fold higher radiotracer concentration was found in selected infarcted tissue than in healthy tissue.

Ex vivo histochemical staining and autoradiography Figure 6 compares TTC staining of a 5-mm-thick slice and a 50- μ m frozen slice with the corresponding autoradiographic image of this frozen slice and a super-

removed, non-infarcted myocardium (*arrowheads*) at the posterior wall and interventricular septum is stained brick red only superficially with TTC (by the nature of this macroscopic staining technique). **c, d** On autoradiography, a “doughnut” pattern of radioactivity uptake appears only in the infarcted region. However, the highest activity (in red) is found mainly subendocardially and near the lateral border zone, where the tracer is able to diffuse from the blood circulation into this occlusive myocardial infarct (*arrows*)

position of the last two images. On the TTC-stained specimen with right ventricular wall attached (Fig. 6a), the transmural infarction involves the entire anterior and lateral wall of the left ventricle, including the posterior papillary muscle (negative staining with TTC). On the frozen slice with right ventricular wall removed (Fig. 6b), the normal myocardium at the posterior wall and interventricular septum are stained only superficially with TTC (by the nature of this macroscopic staining technique). On autoradiography (Fig. 6c,d), a “doughnut” pattern of radioactivity uptake appears only in the infarcted region. However, the highest activity (in red) is found mainly subendocardially and near the lateral border zone, a sign characteristic for occlusive myocardial infarct [38, 39].

Discussion

As hypericin is a polyphenolic polycyclic quinone, it can be labelled efficiently with radioiodine by electrophilic substitution in the *ortho* position of a phenol group [36]. Structural analysis of the radioiodinated derivative has shown that in this way one ^{123}I radionuclide is reproducibly introduced on carbon atom 2, in the *ortho* position of the phenolic group with the most acidic characteristics (Fig. 1). The so-formed mono- ^{123}I iodohypericin (MIH) could be efficiently separated from the starting material hypericin by reversed phase HPLC and was obtained with more than 99% purity in non-carrier-added form in a yield of 70–97% relative to starting ^{123}I activity.

For a preliminary evaluation of the necrosis-avid characteristics of MIH, only a limited number of animals were included in the present study owing to an increasing appeal for animal protection. However, the meticulous and accurate methodologies applied allow us to conclude that MIH is a highly sensitive necrosis-avid agent which may have future clinical applications. After intravenous injec-

tion in rats, gamma camera images showed MIH to be rapidly distributed over the whole body, with early visualisation of blood pool, liver and kidneys, followed by intestines and to a lesser degree urinary bladder. At 24 h and 48 h after injection, there was clear accumulation of MIH in a well-defined area of the induced liver infarction; by contrast, in the untreated rat, an even distribution throughout the liver region was seen at these time points (Fig. 2). As hypericin and MIH are very lipophilic compounds, it is normal that their excretion via liver and intestines is quite high [36, 40]. For this reason, the clear visualisation of a hot spot in the liver is even more remarkable, indicating a high target to non-target activity ratio. Confirmation that this preferential activity uptake was in the infarcted tissue was obtained by comparison of the images generated with TTC staining, H&E staining and autoradiography of tissue blocks from excised liver lobes after sacrifice of the animal 48 h post injection. There was a perfect match between the distribution of radioactivity seen on the autoradiographic image of infarcted liver tissue and the areas stained with H&E, the latter confirming the presence of necrotic tissue (Fig. 4). The same holds true for the images obtained after staining with TTC (Fig. 3). Comparison of the activity per gram tissue in infarcted liver tissue over that in normal liver tissue reveals a 10-fold higher radioactivity concentration in the infarcted area. This is a clear indication that the new tracer agent MIH indeed has a high affinity for necrotic tissue.

The suitability of MIH for visualisation of necrotic myocardium was confirmed using a rabbit model of myocardial infarction. Also in the rabbit study a remarkable hot spot was visible in the region of the myocardial infarction, both at 24 h and at 48 h post injection (Figs. 5, 6). As in the rat study, a perfect match was found between the areas stained by TTC and the areas with higher uptake of radioactivity. As the heart normally has a low uptake of hypericin and MIH [36, 40], it was a logical finding that the target (necrotic myocardium) to non-target (normal myocardium) radiotracer concentration ratio in selected tissue areas was even higher than in the case of the rat liver, namely by a factor of 18. This ratio could have been even higher had more accurate tissue sampling been performed at the subendocardial region, which is the location of higher activity in this model of occlusive myocardial infarction (Fig. 6).

The *in vivo* SPECT images obtained from the small animals considered in this study, i.e. rat and rabbit, suffer from limited spatial resolution. The use of micro-SPECT would eliminate this shortcoming, but this technique was not available in our department at the time of this study. However, *ex vivo* autoradiography in combination with histochemical staining techniques convincingly confirmed the accumulation of MIH in infarcted liver and myocardium. These results suggest that in patients, MIH in combination with SPECT will probably be able to display detailed uptake patterns such as the “doughnut sign”. This implies that MIH-SPECT will be of value in differential diagnosis between occlusive and reperfused myocardial infarction, which is crucial for making clinical decisions

regarding prompt interventions for coronary revascularisation [38, 39].

Regarding the mechanisms behind the observed necrosis avidity of MIH and other compounds mentioned in this paper, there exist more hypotheses than solid experimental proofs [1–33, 37–39]. Despite a great diversity in chemical structures and physicochemical properties among the various necrosis-avid tracer agents for nuclear medicine and necrosis-avid contrast agents for MRI, common physicochemical interactions should be shared by at least some of these agents. A careful analysis of the relationship between structure and uptake/retention of the known necrosis-avid agents in necrotic tissue may provide deeper insights into the unique mechanisms of the phenomenon and yield useful information regarding the structural requirements for further optimisation of this type of agent. Moreover, studies are in progress to identify the exact site of binding of MIH in necrotic tissue and to elucidate the influence of structural modifications on its necrosis avidity.

The present preliminary study has surprisingly shown that MIH has pronounced necrosis-avid properties and merits further evaluation as a potentially useful tracer agent for tissue viability assessment and possibly also as a targeting carrier for tissue revival therapies. It is clear, however, that more investigations are needed, especially with respect to the dynamics of uptake, the earliest time point when necrosis becomes visible and the mechanism of uptake in comparison with that of previously reported necrosis-avid tracer agents such as ^{99m}Tc -pyrophosphate and ^{99m}Tc -glucarate. In addition, further optimisation appears to be indicated, e.g. with respect to rate of excretion, solubility of the tracer agent and nature of the radionuclide. In view of the optimal characteristics of ^{99m}Tc , labelling with ^{99m}Tc would be more appropriate and will be included in further studies.

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