MINI-REVIEW

The potential role of plant secondary metabolites on antifungal and immunomodulatory efect

Xue Zhou1 · Meng Zeng¹ · Fujiao Huang1 · Gang Qin2 · Zhangyong Song1,3 · Fangyan Liu[1](http://orcid.org/0009-0002-5596-3317)

Received: 21 March 2023 / Revised: 13 May 2023 / Accepted: 17 May 2023 / Published online: 5 June 2023 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2023

Abstract

With the widespread use of antibiotic drugs worldwide and the global increase in the number of immunodeficient patients, fungal infections have become a serious threat to global public health security. Moreover, the evolution of fungal resistance to existing antifungal drugs is on the rise. To address these issues, the development of new antifungal drugs or fungal inhibitors needs to be targeted urgently. Plant secondary metabolites are characterized by a wide variety of chemical structures, low price, high availability, high antimicrobial activity, and few side effects. Therefore, plant secondary metabolites may be important resources for the identifcation and development of novel antifungal drugs. However, there are few studies to summarize those contents. In this review, the antifungal modes of action of plant secondary metabolites toward diferent types of fungi and fungal infections are covered, as well as highlighting immunomodulatory efects on the human body. This review of the literature should lay the foundation for research into new antifungal drugs and the discovery of new targets.

Key points

- *Immunocompromised patients who are infected the drug-resistant fungi are increasing.*
- *Plant secondary metabolites toward various fungal targets are covered.*
- *Plant secondary metabolites with immunomodulatory efect are verifed in vivo.*

Keywords Plant secondary metabolites · Fungal infection · Immunoregulation effect · Antifungal mechanism

- \boxtimes Zhangyong Song szy83529@163.com
- \boxtimes Fangyan Liu liufangyan1989@163.com

Xue Zhou zxlovebb0720@163.com

Meng Zeng zengmeng19881026@163.com

Fujiao Huang huang111209@163.com

Gang Qin qin-lzm@163.com

- School of Basic Medical Sciences, Southwest Medical University, Luzhou 646000, People's Republic of China
- ² Department of Otolaryngology Head and Neck Surgery, The Affiliated Hospital of Southwest Medical University, Luzhou 646000, China
- ³ Molecular Biotechnology Platform, Public Center of Experimental Technology, Southwest Medical University, Luzhou 646000, People's Republic of China

Introduction

Fungal infections are characterized by high morbidity and mortality. Every year, at least 1.5 million people are killed, and the lives of more than one billion people are afected by fungal infections (Bongomin et al. [2017](#page-13-0)). In recent years, the frequency of fungal infections has been increasing rapidly. Fungal infections can be divided into superficial and deep infections. Deep fungal infections can cause invasive mycosis, which is an infectious disease caused by fungal invasion of subcutaneous tissues, mucous membranes, and internal organs. About 6 in 100,000 people are infected with invasive fungi each year, although only one-half of fatal cases are diagnosed prior to death (Dignani [2014\)](#page-15-0). Therefore, the lack of timely diagnosis, and treatment, is one of the causes of the high mortality of invasive fungal infections (von Lilienfeld-Toal et al. [2019\)](#page-20-0). At present, invasive fungal infections are mainly caused by species of *Aspergillus*, *Candida*, and *Cryptococcus* (Pathakumari et al. [2020](#page-18-0)), and echinomycin, fucytosine, polyenes, and triazoles are the drugs used most commonly for their treatment. Superficial fungal infections can lead to cutaneous mycosis and superfcial mycosis, which are caused by pathogenic fungi parasitizing keratin tissues, such as hair, nails, and skin. Cutaneous mycosis is one of the most common superfcial fungal infections, with an incidence of up to 25% (Havlickova et al. [2008](#page-15-1)). Cutaneous mycosis is also known as "ringworm." Ringworm infections are caused mainly by species of *Epidermophyton*, *Microsporum*, and *Trichophyton*, with the common skin ringworm infection in human being caused by *Trichophyton* rubrum. At present, topical drugs used to treat superficial fungal infections include allylamines, azoles, and griseofulvin (Ademe [2020](#page-13-1); Khurana et al. [2019\)](#page-16-0).

Because of the increased incidence of fungal infections, the limited range of antifungal drugs available in clinics, and the emergence of drug-resistant or multi-drug-resistant fungal strains, the successful treatment of patients with a fungal infection is a challenging issue. For example, multi-drugresistant *Candida auris* has become one of the major threats to global public health security (Du et al. [2020](#page-15-2)). At the same time, with the emergence of azole-resistant *Aspergillus* strains, *Aspergillus* infections have gradually become one of the global public health concerns (Zhang et al. [2021b](#page-21-0)). During the coronavirus disease 2019 (COVID-19) pandemic, it was found that the number of patients with COVID-19 complicated by *Aspergillus* infection was as high as 23.3% (Lai and Yu [2021\)](#page-16-1). In addition, drug-resistant fungal strains are not limited to deep fungal infections, but are also rapidly increasing in association with superficial fungal infections (Saunte et al. [2019](#page-18-1)). It had found that the incidence of terbinafne-resistant superfcial fungal strains ranged from 16 to 77% in India (Singh et al. [2020](#page-19-0)). Therefore, in order to solve these problems, efforts should be made to develop new antifungal drugs or fungal inhibitors.

In Chinese literature, from Shen Nung, who tested hundreds of grasses, to Li Shizhen's "Compendium of Materia Medica," plants and their derivatives have been successfully used in practice in medicine. For instance, the development of the antimalarial drugs quinine and artemisinin from plants are well-established examples (Achan et al. [2011](#page-13-2); Ma et al. [2020](#page-17-0)). Prescriptions of traditional herbal remedies are still used today, using various active compounds in preparations extracted from plants to treat a range of diseases. Plant secondary metabolites represent an enormous range of small molecular organic compounds developed during the long-term evolution of plants, which are generally not directly involved in plant survival, but which fulfll secondary roles, such as defense chemicals, attractant pigments and fragrances, and plant hormones or growth regulators (Erb and Kliebenstein [2020](#page-15-3)). At present, research on plant metabolites (primary and secondary) is increasing, although research on the antifungal properties of plant metabolites is still mainly focused on plant secondary metabolites (Mickymaray [2019\)](#page-17-1). More than 200,000 diferent plant secondary

metabolites have been isolated and identifed. According to the biosynthetic pathways involved, plant secondary metabolites include nitrogenous organic substances (such as alkaloids, cyanogenic glucosides, and non-protein amino acids), phenolics and terpenoids, as well as other secondary metabolites such as essential oils (Loi et al. [2020](#page-17-2)). The successful development of drugs derived from plant secondary metabolites has resulted in better treatment of cardiovascular diseases, malignant tumors, and neurodegenerative diseases (Table [1\)](#page-2-0).

Invasive pathogens often occur in people with compromised immunity (Xie et al. [2022\)](#page-20-1). Therefore, antifungal agents could act by either having a direct antifungal (fungicidal or fungistatic) efect on pathogenic fungi or playing a role in upregulating a patient's immunity (Arastehfar et al. [2020;](#page-13-3) Lei et al. [2023,](#page-16-2) [2022\)](#page-16-3). There is an urgent need to develop new antifungal drugs, drugs that help to increase the activity of existing antifungal drugs, or drugs with immunomodulatory effects on the host. Additionally, many plant secondary metabolites had been confrmed to show antifungal activity or greatly increase the antifungal action of existing antifungal drugs by synergistic action (Ganesan and Xu [2017;](#page-15-4) Loi et al. [2020\)](#page-17-2). However, compared with investigations on bacterial infectious diseases of humans, there has been a few review about the role of plant extracts or their components in the treatment of fungal infections. Therefore, more attention needs to be paid to the treatment of fungal infections with plant preparations or individual plant secondary metabolites.

Antifungal mechanisms and immunomodulatory efects of existing antifungal drugs

Currently, antifungal targets have been identifed for drugs that treat superfcial or systemic fungal infections. Acrylamide targets the fungal enzyme squalene cyclooxylate and then blocks the synthesis of ergosterol, so that squalene accumulates on the cell membrane, increases the cell membrane brittleness, and leads to the rupture and death of fungal cells (Thapa et al. [2015\)](#page-19-1). Although triazole drugs are the most widely used antifungal drugs, with low hepatorenal toxicity, there is widespread triazole resistance among pathogenic fungi. Triazole drugs mainly bind to 14-α-lanosterol demethylase, a key enzyme in ergosterol biosynthesis in fungal cell membranes, with binding causing inhibition of cell membrane synthesis, cell rupture, and death (Chen et al. [2022a](#page-14-0)). Polyenes, such as amphotericin B (AmB), were among the earliest antifungal drugs; even today, it is still the "gold standard" for the treatment of systemic fungal infections (Mahor et al. [2022](#page-17-3)). By acting on sterols of the phospholipid bilayer of fungal cell membranes, polyenes can

change the permeability of the cell membrane and generate water-soluble pores, thus leading to the loss of intracellular contents and resulting in fungal cell death. However, due to their potential liver and kidney toxicity, polyenes are used in clinical circumstances with caution (Carolus et al. [2020](#page-14-1)). Although the price of echinocandin drugs is high, the use of echinocandin drugs in clinical treatment is increasing due to their broad spectrum, low toxicity, and high efficiency. The echinocandin drugs (large lipoprotein molecules) target and non-competitively inhibit activity of the catalytic subunit of β-1,3-D-glucan synthase, encoded by *fks* genes, causing interference with the fungal cell β-1,3-D-glucan synthesis, and resulting in fungal cell wall permeability changes, leading to cell lysis and death (Campoy and Adrio [2017](#page-14-2)). The drug 5-fuorocytosine exhibits antifungal activity by disrupting fungal DNA and protein synthesis. As mammalian cells lack cytosine deaminase in the antimicrobial pathway, this drug does not have a direct toxic efect on mammals (Delma et al. [2021\)](#page-14-3). Unfortunately, because of the widespread drug-resistant in pathogenic fungi, the use of 5-fucytosine has been greatly reduced. It is only recommended for the treatment of lower urinary tract infections caused by *Candida* (Pappas et al. [2016](#page-18-2)). However, it has become one of the most reliable treatments for cryptococcal meningitis and complex *Candida* infections in combination with AmB (Perfect et al. [2010](#page-18-3)).

Following fungal infection, the efficacy of a drug treatment in vivo is infuenced by the interaction between the body's autoimmune response and antifungal activity of the drug. In addition to their direct antifungal efects, some antifungal agents also have immunomodulatory efects and thus play a therapeutic role in both pathogen killing and host immune system regulation. Azoles are the most costefective drugs for antifungal prophylaxis for immunocompromised patients. For example, itraconazole exerted an anti-infammatory efect by inhibiting *p*-glycoprotein activity in an acute fungal sinusitis with immune dysregulation in Th2-related epithelial cells. The *p*-glycoprotein activity is positively related to infammation in primary human sinus epithelial cells. Therefore, itraconazole treatment has a good anti-infammatory efect and contributes to reducing the incidence of reoperation in patients with acute fungal sinusitis (Lam et al. [2015\)](#page-16-10). Furthermore, in the mouse model of keratoplasty, the infltration of infammatory cells in the corneal tissue of the itraconazole treatment group was signifcantly improved, compared with the phosphate-bufered saline negative control group, and the expression levels of the tumor necrosis factor-alpha (*TNF-α*) and interleukin-6 (*IL-6*) genes were also signifcantly decreased (Cho et al. [2017](#page-14-8)). In addition, posaconazole is efective at preventing invasive fungal infection in patients with severe graft-versus-host disease reaction (Furuno et al. [2018](#page-15-12)). Furthermore, fuconazole (FLC) can activate macrophages through the Toll-like receptor 2 (TLR2)/nuclear factor kappaB (NF-κB) pathway. It could promote the release of TNF- α , interferongamma (IFN-γ), and IL-1β and facilitate the achievement of antifungal effects. The production of TNF- α in THP-1 cells, stimulated by *Aspergillus fumigatus* conidia, decreased after voriconazole treatment (Choi et al. [2010](#page-14-9)). In the situation with polyene treatment, it had been found that AmB treatment increased the expression of the genes encoding IL-1β and IL-20 and decreased the expression of IL-10, IL-2, and IL-3 in human monocytes during response to *A. fumigatus* (Simitsopoulou et al. [2011\)](#page-19-7). In addition, AmB rapidly activates platelets to against *Aspergillus* or *Mucor* infection (Carolus et al. [2020\)](#page-14-1). Further investigation found that AmB treatment induced the secretion of IL-1β and IL-18 and the expression of infammasome pathway genes (André et al. [2020\)](#page-13-6). It has also been confrmed that AmB could activate immune cells through action on the TLR1/TLR2 and CD14 proteins and, at the same time, form an infammatory signaling cascade through myeloid diferentiation factor 88 (Myd88) and the NF-κB signaling pathway. This activity could promote the release of a number of pro-infammatory cytokines (Razonable et al. [2005](#page-18-13)). In addition, intravenous administration of AmB could enhance the phagocytic activity of macrophages and antifungal ability to *A. fumigatus* infection (Nivoix et al. [2020\)](#page-17-13). Among echinocandin drugs, micafungin has also been reported as a fungal prophylactic for bone marrow transplantation, HIV/AIDS, autologous hematopoietic stem cell transplantation, and neutropenia patients (Kim et al. [2022](#page-16-11)). For example, caspofungin signifcantly inhibited the release of pro-infammatory cytokines, such as TNF- α in THP-1 cells (Itoh et al. [2021](#page-15-13)). At the same time, micafungin may condition THP-1 cells toward an infammatory response by TLR2/TLR4 recruitment (Simitsopoulou et al. [2018\)](#page-19-8); what is more, micafungin exerts an anti-inflammatory effect by reducing $TNF-\alpha$ concentrations and increasing IL-10 levels in aspergillosis (Moretti et al. [2014](#page-17-14)). Further investigation confrmed that inhibiting spleen tyrosine kinase (Syk) and its downstream signaling molecules, NF-κB and p38 protein kinase (p38), lead to the decreased secretion of pro-infammatory factors and chemokines in THP-1 cells (Itoh et al. [2021\)](#page-15-13). Whether antifungal drugs promote or suppress infammation is still controversial, but there is no doubt that good antifungal drugs are often not limited to suppressing or killing fungi, but also play a role in regulating host immune system function.

Antifungal mechanism and immunomodulatory efects of terpenoids

Terpenoids, also known as isoprenoid compounds, are synthesized via the mevalonate pathway in the cytoplasm and the deoxyxylulose-5-phosphate pathway in plastids. More than 80,000 terpenoids are known to exist across various plant species (Rudolf et al. [2021\)](#page-18-14). Terpenoids include compounds with the 5-C isoprene unit as the basic structural unit and their derivatives. The molecular formula is $(C_5H_8)_{\text{n}}$, with *n* being the number of connected isoprene units. Based on the number of carbon atoms on the skeleton, terpenoids can be divided into monoterpenoids, sesquiterpenoids, diterpenoids, triterpenoids, tetraterpenoids, and polyterpenoids (Yazaki et al. [2017](#page-21-10)). A wide range of biological functions, such as antibacterial, antifungal, anticancer, anti-infammatory, analgesic, and food preservative, have been reported (Balahbib et al. [2021;](#page-13-7) Pandey et al. [2016\)](#page-18-15).

With respect to antifungal activity, investigations have found that carvacrol, from plants of the *Lamiaceae* family, has an antifungal action similar to FLC, which inhibits fungal ergosterol synthesis, destroys the fungal cell membrane integrity, and then promotes the dissolution and death of fungal cells (Ahmad et al. [2011](#page-13-8)). Kaurene, isolated from members of the *Asteraceae* family, has good anti-cluster induction activity against *C. albicans* (Dal Piaz et al. [2018](#page-14-10)). The upregulated expression of the terpene synthase gene *OsTPS19* in rice can promote the production of limonene and enhance its defense against *Aspergillus oryzae* infection (Chen et al. [2018\)](#page-14-11). Meanwhile, limonene can induce *C. albicans* apoptosis via induction of fungal cell wall/membrane destruction, oxidative and nucleolar stress, DNA damage, and the metacaspase-dependent pathway (Thakre et al. [2018](#page-19-9)). In addition, limonene showed signifcant antifungal activity against *A. fumigatus*, *Cryptococcus neoformans*, *T. rubrum*, and *T. mentagrophytes* (Pinto et al. [2017](#page-18-16)). Citronellal can inhibit the growth of *C. albicans* by destroying the cell membrane integrity, inhibiting bioflm formation, and blocking the fungal cell cycle (Trindade et al. [2022](#page-19-10); Zore et al. [2011\)](#page-21-11). What is more, citronellal can downregulate expression of the *erg3* gene to inhibit the transformation of lanosterol into ergosterol, inhibiting the growth of *Penicillium* (OuYang et al. [2021](#page-18-17)). Moreover, the combination of carvone and AmB greatly increased the inhibition of the growth rate of *C. albicans* (O'Brien et al. [2019](#page-17-15)). In addition, geraniol, widely distributed in geranium, rose, bee-sweet mint, and other plant species, could inhibit *C. albicans* from forming pseudomycelia and chlamydia spores and increased fungal cell membrane permeability, leading to intracellular potassium outfow (Leite et al. [2015](#page-16-12)). At the same time, it was also found that geraniol has a marked antifungal efect on *A. niger* (Kamatou et al. [2013](#page-16-13)). Thymol, a monoterpenoid phenolic derivative isolated from *Syringa*, among other species, had a signifcant antifungal efect on *C. albicans* and *C. tropicalis*; moreover, the minimum inhibitory concentration (MIC) value of nystatin was decreased by 87.4% after combination with thymol (de Castro et al. [2015\)](#page-14-12). Moreover, it was also found in in vivo models, where carveol has been successfully used in the treatment of vulvovaginal *Candida* infection in mice; the antifungal target of carveol results in inhibition of the production of yeast plasma membrane H+-ATPase (Cacciatore et al. [2015](#page-14-13); Menon et al. [2021](#page-17-16)). In the mouse vaginal *Candida* model, limonene treatment signifcantly reduced the fungal load (Munoz et al. [2020](#page-17-17)).

In recent years, further investigations have found that terpenoids regulate the immune function of the human body and improve various symptoms caused by infammation. Soybean saponins played an anti-infammatory role by inhibiting the degradation of an inhibitor of NF-κBα (IκBα) and activating NF-κB to downregulate the production of cyclooxygenase-2 (COX-2), nitric oxide synthase (iNOS), human macrophage chemoattractant protein-1 (MCP-1), and TNF- α in lipopolysaccharide (LPS)-stimulated RAW264.7 cells (Kang et al. [2005](#page-16-14)). Meanwhile, quinoa saponins, with a dose-dependent action, could downregulate the secretion of TNF-α, nitric oxide (NO), and IL-6 in LPS-activated RAW264.7 cells (Yao et al. [2014\)](#page-21-12), and geniposide, the main bioactive component of *Gardenia jasminoides* fruits, could inhibit the activation of NF-κB in a concentration-dependent manner, downregulating gene expression of *IFN-γ* and *iNOS* in LPS-activated RAW264.7 cells (Koo et al. [2004](#page-16-15)). In addition, laurene can inhibit the activation of NF-κB, c-Jun N-terminal kinase (JNK), and p38 in human chondrocytes induced by IL-1β. Furthermore, compared with IL-1β-induced inflammatory cells, laurene has an antiinflammatory effect by reducing the activity of extracellular regulated protein kinases (ERK1/2) (Rufno et al. [2015](#page-18-18)). β-patchoulene signifcantly altered sepsis-induced neuroinfammation and microglia activation and improved the peripheral immune function (Tian et al. [2023\)](#page-19-11). In a mouse model of acute inflammation, β-patchoulene alleviated edema and infammatory cell infltration. β-patchoulene, in a dose-dependent manner, inhibited the secretion of IL-6, TNF- α , and IL-1 β in a mouse model of inflammation and could signifcantly downregulate the expression of genes *iNOS* and *COX-2* (Zhang et al. [2016b\)](#page-21-13). Furthermore, ganoderic acid, a triterpenoid compound extracted from the Chinese medicinal mushroom *Ganoderma lucidum*, was found to signifcantly inhibit phorbol ester-induced infammation in mice (Akihisa et al. [2007\)](#page-13-9). Therefore, terpenoids have an efect on the growth, inhibition, and killing of common human-pathogenic fungi and phytopathogenic fungi. At the same time, some terpenoids can also help the host to regulate and balance the infammatory state of the body in the face of fungal infection by regulating the secretion of related cytokine.

Antifungal targets and immunomodulatory efects of phenolic compounds

Phenolic compounds are one of the most common classes of plant secondary metabolites; more than 8000 phenolic compounds with known structures are found in plants. Most phenolic compounds in plants are synthesized by the shikimic acid (the major pathway) and malonic acid pathways. These compounds are formed by an aromatic ring or rings combined with one or multiple hydroxyl groups. Phenolics are mainly composed of polymeric or monomeric structures, which can exist in the form of glycosides, aglycones, substrates, or free-binding compounds (Alara et al. [2021](#page-13-10)). Phenolic compounds include favonoids, phenolic acids, and polyphenols and tannins and can be found in fruits, vegetables, legumes, and tea. In addition, biological functions of phenolic compounds include antioxidant, antibacterial, anti-infammatory, analgesic, antipyretic, and anti-tumor, among others (Alara et al. [2021](#page-13-10)), as well as in the prevention of Alzheimer's disease, cardiovascular disease, obesity, and other diseases (de Araújo et al. [2021](#page-14-14)). In recent years, more and more natural plant phenolic compounds have been recognized, and they have become an important research target for the development of novel antifungal agents (Chtioui et al. [2022](#page-14-15)).

Flavonoids are hydroxylated phenolic molecules with a C_6 - C_3 - C_6 carbon skeleton structure, connecting two aromatic rings with a heterocyclic ring containing three

carbon atoms. Since the frst discovery of favonoids in orange peel in 1930, more than 4000 flavonoids have been identifed in plants. Therefore, favonoids represent one of the most important classes of the phenolic family and account for one-half of the total number of phenolic compounds. For the human body, favonoids in fruits and vegetables are directly related to the health status of the human diet, contributing to anticancer activity and prevention of cardiovascular diseases (Middleton [1998;](#page-17-18) Tungmunnithum et al. [2018](#page-19-12)). In addition, investigations have found that natural favonoids have the ability to act directly as antifungal molecules and to interact synergistically with other antifungals (Jin [2019\)](#page-16-16). It has also been reported that some favonoids can signifcantly reduce the spore germination of plant-pathogenic fungi, such as *Botryosphaeria* (Ma et al. [2022b\)](#page-17-19). Therefore, favonoids are expected to become one of the important research targets from which to develop antifungal drugs. It has been confrmed that certain favonoid compounds have antifungal activities against human pathogens such as *Aspergillus* (Wang et al. [2021a](#page-20-16)), *Candida* (da Fonseca et al. [2022](#page-14-16)), *Cryptococcus* (Fowler et al. [2011](#page-15-14)), and *Malassezia* spp*.* (Alves et al. [2017\)](#page-13-11). For example, the favonoids 4′,5-dihydroxy-7-methoxy-6-(3-methyl-[2-butenyl])-(2S)-favanone, isolated from the shrub *Eysenhardtia texana* (Wächter et al. [1999\)](#page-20-17), and 7-hydroxy-3′,4′-(methylenedioxy)-favan, isolated from the tree *Terminalia bellirica*, have antifungal activity toward *C. albicans* (Valsaraj et al. [1997\)](#page-20-18). The reason why the propolis has an inhibitory efect on *Aspergillus niger* and *C. albicans* is that it contains a high concentration of favonoids (Vică et al. [2022\)](#page-20-19). In addition, curcumin from *Curcuma longa* L. has a fungistatic efect on *A. favus* (Temba et al. [2019\)](#page-19-13). Additionally, the favonoid baicalin, the main bioactive component of the traditional Chinese medicinal plant *Scutellaria baicalensis*, exerts a concentration-dependent antifungal efect by inhibition of *C. albicans* bioflm formation and increasing the apoptosis rate of this human pathogen (Cao et al. [2008](#page-14-17)). Meanwhile, further studies have found that the bioflm inhibition mechanism of baicalin is caused by the upregulated expression of many redox-related genes (such as *CAP1*, *SOD2*, and *TRR1*), increasing the Ca^{2+} concentration in the cytosol, damaging the cell ultrastructure, and increasing the *C. albicans* apoptosis rate (Cao et al. [2008;](#page-14-17) Yang et al. [2014\)](#page-20-20). In addition to *C. albicans*, baicalin also has antifungal efects on *A. fumigatus*, *T. rubens*, and *T. trichophyton* (Da et al. [2019](#page-14-18)). Moreover, quercetin, extracted from *Morus alba* L., showed antifungal activity against *A. fumigatus*, with the fungal load in the corneal tissue of mice treated with quercetin being signifcantly lower than in control mice (Yin et al. [2021\)](#page-21-14).

In addition to favonoids, the polyphenol chlorogenic acid, widely found in apple, coffee, potato, and tomato, has a synergistic antifungal efect (in combination with FLC) on *C. albicans* and *Malassezia* spp. (Rhimi et al. [2020](#page-18-19)). In addition, chlorogenic acid was found to have antifungal efects against phytopathogenic fungi such as *Fusarium* spp. and *Verticillium dahliae* through a reactive oxygen species (ROS)-dependent mode of action and alteration of fungal cell membrane permeability (Kai et al. [2021](#page-16-17)). In addition, gallic acid, another class (phenolic acid) of phenolic compound, also had antifungal efects on *C. albicans* (Teodoro et al. [2018](#page-19-14)). In addition to efects on *A. fumigatus* and *A. niger*, resveratrol (another polyphenol) not only inhibits bioflm formation of *C. albicans*, but also has an inhibitory effect on the growth of superficial skin fungi, especially *Epidermophyton floccosum*, *Microsporum gypseum*, *T. mentagrophytes*, *T. tonsurans*, and *T. rubrum* (Jediyi et al. [2020;](#page-16-18) Okamoto-Shibayama et al. [2021](#page-17-20)). Pterostilbene (a demethylated derivative of resveratrol), extracted from grape leaves, is one of the more active antifungal compounds against *C. albicans*, with the antifungal mechanism acting on the Ras/cAMP pathway of *C. albicans*, downregulating the expression of genes involved in ergosterol biosynthesis (Li et al. [2014\)](#page-16-19). In addition, in the bioflm model of rat central venous catheter, defects in *C. albicans* bioflm formation were observed in the lumen at a treatment concentration of 16 μg/mL pterostilbene, whereas no bioflm formation was observed and no fungal cells were even detected in the lumen of central venous catheter in the treatment groups at concentrations of 32 μg/mL and 64 μg/mL (Kolouchova et al. [2018](#page-16-20); Li et al. [2014](#page-16-19)). Interestingly, the antifungal effect of pterostilbene against *C*. *albicans* exceeded that of resveratrol. Recent investigations found that the licochalcone A in the ethanol extract from the licorice roots of *Glycyrrhiza* species signifcantly inhibited the formation of the *C. albicans* bioflm, and the fungal load in tongues of mice treated with these isoprene acylated ketones was signifcantly lower in the oral *Candida* mouse model than that in the control mice (Seleem et al. [2016](#page-19-15)).

Many phenolic compounds have anti-inflammatory and antioxidant efects (Dominguez-Avila et al. [2021\)](#page-15-15). For example, gingerol extracted from ginger rhizomes has high antioxidant activity. The potential antioxidant mechanism of gingerol in human intestinal epithelial cells prevents the degradation of Keap1-Nrf2 protease, promotes the translocation of Nrf2 into the nucleus, increases the expression of *Nrf2* target genes, and increases the level of the antioxidant reduced glutathione, decreasing the concentration of ROS. Moreover, gingerol also reduces the production of NO and prostaglandin E2 (PGE2) in RAW264.7 cells (Mao et al. [2019;](#page-17-21) Zhang et al. [2013](#page-21-15)). Quercetin acts on immune cells and then targets intracellular signaling kinases, phosphatases, and membrane proteins to regulate cell-specific functions. Therefore, there is a hypothesis that quercetin is an immunomodulatory molecule. In LPS-activated RAW264.7 cells, quercetin inhibits phosphatidylinositol kinase (PI3K) phosphorylation and subsequent TLR4/ MyD88/PI3K complex formation via nonreceptor tyrosine kinase c (Src) and Syk (Domínguez-Avila et al. [2022\)](#page-15-16). Kaempferol, a favonol, which is a type of favonoid, also exerts anti-infammatory efects by inhibiting the activity of hyaluronidase and reducing the level of ROS produced during cell stimulation (Yang et al. [2020](#page-20-21)). Chlorogenic acid, gallic acid, and kaempferol showed anti-infammatory activities by inhibiting the expression of *iNOS*, secretion of pro-infammatory factors, and production of ROS in LPS-induced RAW264.7 cells (Yang et al. [2020\)](#page-20-21). Ferulic acid, a phenolic acid compound, has been shown to play an anti-infammatory role in bovine endometrial epithelial cells by inhibiting IκB degradation, the phosphorylation of NF-κB (p65) and mitogen-activated protein kinase, and reducing the production of the pro-inflammatory cytokines IL-1β, IL-6, IL-8, and TNF-α (Yin et al. [2019\)](#page-21-16). Quercetin and resveratrol exert anti-infammatory efects by downregulating expression at the mRNA and protein levels of NO, iNOS, TNF-α, IL-1β, IL-6, and the granulocyte–macrophage colony-stimulating factor (GM-CSF) (Endale et al. [2013](#page-15-17)). Moreover, quercetin reduced IL-8 production in LPS-activated lung A549 cells (Geraets et al. [2007\)](#page-15-18). It was also reported that quercetin and resveratrol could signifcantly reduce TNF-α production in LPS-activated microglia, and resveratrol also could inhibit IL-1 production (Bureau et al. [2008\)](#page-14-19), whereas quercetin was also found to have immunomodulatory efects in mast cells, T lymphocytes, and peripheral blood mononuclear cells (Li et al. [2016;](#page-16-21) Yang et al. [2015](#page-20-22)). After feeding quercetin to healthy broiler chickens for 6 weeks, the concentration of serum immunoglobulin, IL-4, as well as the spleen index, thymus index, and bursa of Fabricius index, were all increased. These data indicated that quercetin can enhance the immune ability of animals by stimulating the development of immune organs and the subsequent amplifcation of humoral immunity (Yang et al. [2020](#page-20-21)). A small number of phenolic compounds such as quercetin and resveratrol with antifungal efects also have immunomodulatory efects on the body, which is one of the promising new drug research and development targets.

Antifungal mechanism and immunomodulatory efects of nitrogenous secondary compounds

Nitrogenous secondary compounds are another type of secondary metabolites in plants, most of which are synthesized from amino acids. Their metabolism is complicated and can be affected by plant hormones and environmental stress (Cho et al. [2008](#page-14-20)). Nitrogen-containing secondary metabolites include alkaloids, cyanogenic glycosides (CNGs), and nonprotein amino acids. CNGs, glycosides with α-hydroxynitrile, are composed of an aglycone with a sugar group attached (Bolarinwa et al. [2015](#page-13-12)). CNGs are potentially highly toxic substance, releasing hydrogen cyanide when hydrolyzed, although the compounds themselves are not toxic, and may lead to acute cyanide poisoning (Cressey et al. [2013\)](#page-14-21). Another class of plant nitrogenous compounds is non-protein amino acids, which are important stores of nitrogen in plants. In addition to their antibacterial, antifungal, and anticancer effects, non-protein amino acids primarily help plants to resist harmful insects (Huang et al. [2011\)](#page-15-19). Alkaloids are found in over 20% of plant species and are generally found in high concentrations in plant families such as the *Fabaceae*, *Apocynaceae*, *Polygonaceae*, *Papaveraceae*, *Ranunculaceae*, *Rutaceae*, and *Solanaceae*. They are compounds that contain at least one nitrogen atom in a naturally occurring heterocyclic ring. According to the chemical structure classification, alkaloids can be divided into pyridine, isoquinoline, indole, scopolamine, and organic amine alkaloids, among others. Alkaloids have a range of biological activities and are used in treatment of bacterial infections, cancer, dementia, and pain and are an important source of many drugs (Adamski et al. [2020\)](#page-13-13). Here, the antifungal efects of alkaloids will be discussed. Alkaloids extracted from sea buckthorn (*Hippophae rhamnoides*), including acridone, fuoroquinolone, and 4-quinolone, play an inhibitory role against fungi by downregulating the expression of the *ICL1* gene in *C. albicans* (Kamal et al. [2021\)](#page-16-22). Magnoforine, present in *Acorus calamus*, *Tinospora cordifolia*, and *Celastrus paniculatus*, exerts antifungal effects by inhibiting α -glucosidase activity and reducing bioflm formation in *C. albicans* (Kim et al. [2018](#page-16-23)). Meanwhile, magnoforine can also damage the *T. rubrum* cell membrane, increasing the leakage of nucleic acids from fungal cells, reducing the activities of squalene epoxidase and 14-α-lanosterol demethylase, and reducing the concentration of ergosterol in mycelia (Luo et al. [2021](#page-17-22)). In addition, graveoline, from rue (*Ruta graveolens*), showed signifcant antifungal activity against *C. albicans*, *Fusarium oxysporum*, and *T. rubrum* (Cantrell et al. [2005](#page-14-22); Duraipandiyan and Ignacimuthu [2009\)](#page-15-20). Pteleine, one of the furanoquinoline alkaloids, exhibits antifungal activity against *C. albicans* (Shang et al. [2018](#page-19-16)). Recent investigations also found that 8-acetylnorchylerythrine and 8-methoxydictamnine, from *Zanthoxylum* (*Toddalia*) *asiatica*, exhibit antifungal activity against *C. albicans*, *Candida glabrata*, and *Candida tropicalis* (Hu et al. [2014](#page-15-21)). The half-maximal inhibitory concentration (IC_{50}) values of 2,3-dihydro-1H-indolizinium chloride, an indoleazine alkaloid extracted from the fern *Dryopteris enneaphylla*, against *A. fumigatus* and *C. neoformans* were 0.3 μg/mL and 4.0 μg/mL, respectively (Samoylenko et al. [2009;](#page-18-20) Thawabteh et al. [2019\)](#page-19-17). Berberine, isolated from *Coptis chinensis* and *Phellodendron*

chinense, exerted an antifungal effect on *C. albicans* by upregulating the expression of core genes (*sln1*, *ssk2*, *hog1*, and *pbs2*) and inducing ROS accumulation and inhibiting the expression of the chitin synthase gene (*chs3*) and the β-(1,3) glucan synthase gene (*gsc1*) to damage the integrity of the cytoplasm, inhibit the formation of germ tubes and hyphae, and destroy the integrity of cell wall (Huang et al. [2021](#page-15-22)). Berberine also exerted synergistic antifungal spread ability in vivo when combined with AmB or FLC; when berberine was combined with AmB, the survival time of mice with disseminated infection of pathogenic *C. albicans* was increased from 14 to 36 d (Huang et al. [2022;](#page-15-23) Quan et al. [2006\)](#page-18-21).

To regulate the immune response, steroidal alkaloids, from the bulbs of *Fritillaria* spp. of the *Liliaceae*, blocked LPS-induced phosphorylation and degradation of both IκBα and JNK and signifcantly inhibited the production of NO, IL-6, and TNF- α in RAW264.7 cells (Wang et al. [2021d](#page-20-23)). In addition, oral administration of sinomenine could reduce the activities of iNOS and COX-2 in rats (Zhu et al. [2019](#page-21-17)). Furthermore, aconitine can improve LPS-induced acute lung injury in rats by inhibiting the activation of NF-κB and reducing the concentrations of TNF- α , IL-6, and IL-1 β (Wang et al. [2019c\)](#page-20-24). Moreover, berberine and matrine (from Sophora sp.) are more effective against inflammation, with berberine reducing the concentration of COX-2 and inhibiting the synthesis of PGE2 to achieve the anti-swelling efect in the formalin-induced foot swelling mice model (Yao et al. [2019](#page-21-18)). Berberine also had an inhibitory efect on the secretion of inflammatory factors, including TNF- α , IL-8, IL-6, and MCP-1, in dinitrofuorobenzene-induced delayedtype hypersensitivity rat model. Meanwhile, to alleviate chronic colitis, berberine could regulate the production of INF- γ and IL-17A in CD4⁺ T cells through activation of adenosine 5′-monophosphate-activated protein kinase (AMPK) (Takahara et al. [2019;](#page-19-18) Yu et al. [2019\)](#page-21-19). Recent studies also found that matrine, isolated from the roots of *Sophora favescens*, has a strong negative regulatory efect on the secretion of TNF- α , IL-8, and IL-1 α in THP-1 cells (Zhou et al. [2019](#page-21-20)). It also exerted anti-infammatory efects by downregulating the expression of the genes encoding lipopolysaccharide recognition receptor, lipopolysaccharidebinding protein, CD14 and TLR4, and the transcription of the nuclear factors *c-Jun* and *c-fos* in the mouse model of foot swelling (Li et al. [2020b\)](#page-16-24). In addition, matrine inhibited the NF-κB signaling pathway in mouse airway epithelial cells, reduced the expression of suppressors of cytokine signaling 3, and reduced the production of ROS and infammatory factors in alveolar macrophages to inhibit airway inflammation in a mouse model of asthma (Li et al. [2019\)](#page-16-25). Therefore, alkaloids mainly improve infammation by inhibiting related infammatory factors and signaling pathways and then play a role in immune regulation in the body.

Antifungal mechanism and immunomodulatory efects of other secondary metabolites

In addition to terpenoids, phenolics, and nitrogencontaining secondary compounds, plant-derived essential oils, also known as volatile oils, have attracted much attention for their antifungal activities. Essential oils are a mixture of secondary metabolites. Most are volatile aromatic oil-like liquids, containing aliphatic compounds, aromatic compounds, sulfur and nitrogen compounds, and terpenes and their oxygen derivatives. Since 1987, more than 500 articles have reported on the antifungal properties of essential oils (Kalemba and Kunicka [2003\)](#page-16-26). Essential oils have been listed as the most widely used special plant metabolites due to their anti-infective properties (Plant et al. [2019\)](#page-18-22). Investigations have shown that essential oils have signifcant antifungal efects, not only on deep fungal infections caused by *Aspergillus* and *Candida*, but also on superficial fungal infections caused by *Microsporum canis*, *M. gypseum*, *T. rubrum*, and *T. mentagrophytes*, and even on phytopathogenic fungi in crops (Bakkali et al. [2008](#page-13-14); Danielli et al. [2018](#page-14-23)). The antifungal targets of essential oils mainly involve inhibition of fungal cell growth and mycotoxin synthesis, for example, by disruption of cell membrane permeability and intracellular electron transport chains, resulting in intracellular metabolic disorders (Mirza Alizadeh et al. [2022](#page-17-23)). Investigations have found that rosemary (*Rosmarinus officinalis*) essential oil not only targets mycelial ergosterol synthesis to inhibit toxin biosynthesis in *A. favus*, but also inhibits mycelium growth to play an antifungal efect on the phytopathogen *F. oxysporum* (da Silva Bomfm et al. [2020;](#page-14-24) Hussein et al. [2020\)](#page-15-24). Furthermore, *Rhododendron tomentosum* essential oils not only have an inhibitory efect on *Candida parapsilosis*, but also play an antifungal role in afecting the permeability of cell membranes in the yeast, *Saccharomyces cerevisiae* (Judzentiene et al. [2020\)](#page-16-27). The concentrations of thyme essential oil and ginger essential oil were 11.25 μg/mL and 364 μg/mL, respectively, which exerted a marked antifungal effect on *Fusarium* and an inhibitory efect on the synthesis of *Fusarium* mycotoxins such as deoxypyrimethamine and zearalenone (Romoli et al. [2022](#page-18-23)). In addition to extensive antibacterial activity, *Bupleurum rigidum* essential oil can also play an antifungal role by changing the ultrastructure of *C. albicans*, *C. neoformans*, and *T. rubrum* (Zuzarte et al. [2021\)](#page-21-21). *Melaleuca alternifolia* essential oil, which is mainly pinene-4-alcohol, showed antifungal activity in vitro against *A. niger* and both azole-sensitive and azole-resistant *C. albicans* (Hammer et al. [2002](#page-15-25); Kumar [2020](#page-16-28)); in the rat vaginal *C. albicans* infection model, this essential oil also

had a great impact on eliminating *C. albicans* infection and enhancing the anti-infection ability (Mondello et al. [2006\)](#page-17-24). Oregano oil at a concentration of 0.25 mg/mL completely inhibited the growth of *C. albicans* in vitro. In addition, it also inhibited spore germination and mycelium growth in a concentration-dependent manner (Manohar et al. [2001](#page-17-25)). In vivo, the survival rate of the mice was as high as 80% after continuous treatment with oregano oil for 30 days in a mouse systemic candidiasis model, whereas the fungal load in mouse kidney tissue was close to zero (Manohar et al. [2001\)](#page-17-25). Sodium houttuyfonate (SH), a volatile oil from *Houttuynia cordata*, exerted an antifungal effect on *C. albicans* by afecting gene expression in the Ras1-cAMP-Efg1 pathway and decreasing bioflm formation and the production of cAMP. Compared with the infected group, the survival rate of the SH-treated group was signifcantly higher in the experimental model of *Galleria mellonella* caterpillars (Wu et al. [2020](#page-20-25)). In addition, our previous investigation also found that sodium new houttuyfonate (SNH), which were modified compounds of SH, had a

Fig. 1 Mechanism of antifungal action of existing antifungal drugs and plant secondary metabolites. **A** Antifungal targets had been confrmed in the existing antifungal drug. Existing antifungal agents include allylamines, azoles, polyalenes, echinocins, and fucytosine. **B** Antifungal targets had been confrmed in the plant secondary metabolites. The antifungal pathway of plant secondary metabolites include (1) prevention of fungal bioflm formation and destruction of the established fungal bioflm; (2) alteration of cellular DNA replication and disruption of the cell cycle; (3) inhibition of fungal mycotoxin synthesis; (4) prevention of ergosterol synthesis, disruption of cell membrane permeability, and promotion of cell wall destruction and lysis; and (5) decrease in the number of spores and the growth of hyphae

Classification	Name of active compounds/ secondary metabolites	Experimental model	Mechanism of action	Reference
Alkaloid	Aconitine	Rat	Inhibit the activation of NF- κ B Wang et al. (2019c) and the production of TNF- α , IL-6, and IL-1 β	
	Berberine	Mice	Reduce the synthesis of COX-2 Yao et al. (2019) and PGE2	
		Rat	Inhibit the secretion of TNF- α , Yu et al. (2019) IL-8, IL-6, and MCP-1	
		$CD4+$ T cells	Activate AMPK signal pathway	Takahara et al. (2019)
	Matrine	Mice	downregulate the expression of CD14 and TLR4	Li et al. $(2020b)$
		Mouse airway epithelial cells Inhibit the production of	SOCS3 and ROS by NF-KB signal pathway	Li et al. (2019)
		THP-1 cells	Negative regulate the secretion Zhou et al. (2019) of TNF- α , IL-8, and IL-1 α	
	Sinomenine	Rat	Inhibit the activities of iNOS and COX-2	Zhu et al. (2019)
Phenolics	Gingerol		Human intestinal epithelial cell Increase the expression of $Nrf2$ Mao et al. (2019) and the level of the antioxi- dant reduced glutathione and decrease the concentration of ROS	
		RAW264.7 cell	Inhibit the production of NO and PGE2	Zhang et al. (2013)
	Quercetin	RAW264.7 cell	and TLR4/MyD88/PI3K complex formation via Src and Syk	Inhibit PI3K phosphorylation Domínguez-Avila et al. (2022)
		Lung A549 cell	Inhibit the production of IL-8 Geraets et al. (2007)	
		Microglia	Inhibit the production of TNF- α	Bureau et al. (2008)
		Broiler chickens	Activate the NF- _{KB} signal pathway	Yang et al. (2020b)
	Resveratrol	Microglia	Inhibit the production of TNF- α and IL-1	Bureau et al. (2008)
	Ferulic acid	Bovine uterine epithelial endometrial cell	Inhibit the production of IL-1 β , IL-6, IL-8, and TNF- α by IKB/NF-KB/ MAPK signal pathway	Yin et al. (2019)
Terpenes	Geniposide	RAW264.7 cell	Inhibit the activation of NF-kB and expression of $IFN-\gamma$ and $iNOS$	Koo et al. (2004)
	Laurene	Human chondrocytes	Inhibit the activation of NF-KB, JNK, ERK1/2, and p38	Rufino et al. (2015)
	Quinoa saponin	RAW264.7 cell	Downregulate the expression of $TNF-\alpha$, IL-6 and iNOS	Yao et al. (2014)
	Soybean saponins	RAW264.7 cell	Inhibit the degradation of Iκ $B\alpha$ and activation of NF - κ B, and the production of COX-2, iNOS, MCP-1, and TNF- α	Kang et al. (2005)
	β -patchoulene	Mice	Inhibit the production of IL-6, TNF- α , and IL-1 β , and the expression of iNOS and COX-2	Zhang et al. $(2016b)$

Table 2 Regulatory immune mechanisms of plant secondary metabolites in vitro and in vivo

AMPK adenosine 5′-monophosphate (AMP)-activated protein kinase, *Caspase-1* cysteinyl aspartate-specifc proteinase, *COX-2* cyclooxygenase-2, *ERK* extracellular regulated protein kinases, *GM-CSF* granulocyte–macrophage colony stimulating factor, *GSH* gonad-stimulating hormone, *iNOS* inducible nitric oxide synthase, *IL-1* interleukin-1, *IL-6* interleukin-6, *IL-8* interleukin-8, *IL-17A* interleukin-17A, *INF-γ* interferongamma, *IκB* inhibitor of NF-κB, *JNK* c-Jun N-terminal kinase, *JAK* janus kinase, *MAPK* mitogen-activated protein kinase, *MCP-1* monocyte chemoattractant protein-1, *MPO* myeloperoxidase, *Myd88* myeloid diferentiation factor 88, *NF-κB* nuclear factor kappa-B, *NO* nitric oxide, *Nrf2* nuclear factor erythroid 2, *NLRP3* nucleotide-binding oligomerization domain, leucine-rich repeat and pyrin domain-containing 3, *PGE2* prostaglandin E2, *PI3K* phosphatidylinositol-3-kinase, *p38* phosphorylated p38 mitogen-activated protein kinase, *ROS* reactive oxygen species, *SOCS3* recombinant suppressors of cytokine signaling 3, *STAT* signal transducer and activator of transcription, *Src* nonreceptor tyrosine kinase c, *Syk* spleen tyrosine kinase, *TNF* tumor necrosis factor; *TLR4* toll-like receptor 4

marked antifungal efect on *A. fumigatus*. SNH achieves antifungal efects by inhibiting the synthesis of ergosterol in the cell membrane of *A. fumigatus*. In addition, in a mouse model of systemic *A. fumigatus* infection, SNH treatment signifcantly reduced the fungal load in the tissues (Zhang et al. [2022b\)](#page-21-22). Studies have also found that essential oils can synergistically improve the antifungal effect when combined with existing antifungal drugs. For example, the combination of oregano essential oil and winter savory essential oil with the synthetic antifungal drug clotrimazole signifcantly reduced the metabolic activity of *C. glabrata*. At the same time, low concentrations of winter savory essential oil combined with clotrimazole caused organellar disorder in this fungus, with autophagic vacuoles, whereas high concentrations of winter savory essential oil combined with clotrimazole caused complete destruction of *C. glabrata* organelles (Massa et al. [2018](#page-17-26)).

Recent investigations have also reported that peppermint essential oil could alleviate the excessive infammation

exhibited by LPS-induced RAW264.7 cells by inhibiting the ERK/NF-κB pathway and the gene expression of *COX-2*, *iNOS*, *IL-6*, and *IL-1β* (Kim et al. [2021](#page-16-29)). It also found that the essential oil from *Citrus* fower blocked the MAPK signaling pathway by inhibiting the phosphorylation of p38 and JNK and downregulating the gene expression of *IL-6*, *IL-1β*, and *TNF-* α in RAW264.7 cells (Shen et al. [2017](#page-19-19)). In addition, *Lanxangia tsaoko* (black cardamom, formerly *Amomum tsao-ko*) essential oil could reduce the activation of the NLRP3 infammasome by inhibiting the production of Caspase-1 and downregulate the expression of infammatory mediators iNOS and COX-2 by inhibiting the activation of JAK/STAT and the processing of IL-1β and pyroptosis in THP-1 cells (Chen et al. [2017,](#page-14-25) [2021\)](#page-14-26). Like dexamethasone, 1-h early injection of lavender essential oil had a therapeutic efect in the rat model of kappa-carrageenan-induced pleurisy. Further research found that the volume and total protein concentration of the exudate collected from the rats were both signifcantly reduced, while the total numbers

Fig. 2 Summary of plant secondary metabolites with both antifungal and immunomodulatory efect. These compounds are derived from alkaloids, phenols, terpenoids, and other plant secondary metabolites, respectively

of leukocytes and polymorphocytic leukocytes migrating into the pleural cavity were also reduced (Silva et al. [2015](#page-19-20)). This phenomenon also occurred in an animal model of carrageenan-induced pleurisy treated with rosemary essential oil. Investigations found that rosemary essential oil could induce leukocyte migration in vivo as well as induce chemotaxis in vitro. Recent studies found that dietary supplementation of rosemary essential oil signifcantly reduced MPO activity and IL-6 level in a 2,3,6-trinitrobenzenesulfonic acid (TNBS)-induced colitis mouse model (Borges et al. [2019\)](#page-13-15). The efect of *Citrus bergamia* (bergamot orange) essential oil on acne vulgaris was explored in the *Mesocricetus auratus* (golden hamster) model. It was found that the serum levels of IL-1 α and TNF- α decreased in response to the oil in a dose-dependent manner after treatment (Sun et al. [2020](#page-19-21)). Recently, nutmeg (*Myristica fragrans*) essential oil extract was found to have anti-infammatory activity in rodents, associated with reduced release of COX-2 and P-substances in the blood, as well as reduction of joint swelling induced by Freund's adjuvant injection in rats (Ashokkumar et al. [2022](#page-13-16); Zhang et al. [2016a](#page-21-23)). Moreover, intragastric administration of SNH could signifcantly reduce fungal load in tissues and exerted anti-infammatory efects through downregulating the production of infammatory cytokines IL-6 and IL-17A in a mouse model of systemic *A. fumigatus* infection (Zhang et al. [2022b](#page-21-22)). Peppermint essential oil has marked anti-infammatory efect, not only inhibiting NO and PGE2 production in LPS-activated RAW264.7 cells, but also inhibiting the production of NO and PGE2 on croton oil-induced ear edema in mice (Sun et al. [2014\)](#page-19-22). Because of the antifungal potential of some essential oils when used alone or in combination, and of the efficacy of some essential oils in regulating inflammation, plant-derived essential oils also have great development potential in antifungal applications.

Conclusion

This review frst briefy summarizes the current situation of clinical antifungal treatments of common superfcial and deep fungal infections, the antifungal mechanisms of existing antifungal drugs (Fig. [1A](#page-8-0)), and their immunomodulatory effects on the host. At present, although there are few types of antifungal drugs and their targets, antifungal drugs can better help treat fungal infections by regulating the secretion of pro-infammatory or antiinfammatory factors to regulate the immune function of the host. So, can plant secondary metabolites cause similar efects? The mode of action of diferent antifungal plant secondary metabolites against various pathogenic fungi and the immunomodulatory efects of plant metabolites on the host were further reviewed. The antifungal mechanisms of plant secondary metabolites primarily include (1) inhibition of fungal mycotoxin synthesis; (2) prevention of fungal bioflm formation and destruction of the established fungal biofilm; (3) decrease in the number of spores and the growth of hyphae; (4) prevention of ergosterol synthesis, disruption of cell membrane permeability, and promotion of cell wall destruction and lysis; and (5) alteration of cellular DNA replication and disruption of the cell cycle (Fig. [1B](#page-8-0)). At the same time, it was found that many plant secondary metabolites also reduced tissue infammation and played an immunoregulatory function by reducing the release of pro-infammatory factors (Table [2](#page-9-0)). In summary, many plant secondary metabolites have both antifungal and immunomodulatory effect (Fig. [2\)](#page-11-0). And most of them have been used in clinical therapy independently or combined with existing antifungal drugs to better exert antifungal efects (Table [3](#page-12-0)). Therefore, plant secondary metabolites have broad prospects for the development of novel antifungal drugs.

AmB amphotericin B, CAS caspofungin, FLC fluconazole, ITR itraconazole, KET ketoconazole, MCZ miconazole, VRC voriconazole, "-", no study were mentioned in the corresponding refer-
ences; "+", synergistic or additive actio AmB amphotericin B, CAS caspofungin, FLC fluconazole, ITR itraconazole, KET ketoconazole, MCZ miconazole, WRC voriconazole, "-", no study were mentioned in the corresponding references; "+", synergistic or additive action

[2021c\)](#page-20-28). Generally, plant extracts are complex mixtures, and their active ingredients vary according to plant species, chemical types, and extraction methods. Each ingredient may have multiple targets in the body. Therefore, it is still necessary to explore the specifc mechanisms of individual antifungal plant metabolites against fungi or for immunoregulation in vivo. Additionally, the development of antifungal and immunomodulatory drugs from plant secondary metabolites is still at the preliminary stage. The available clinical references are extremely scarce, and such drugs are still a long way from being used in the clinic. Therefore, what techniques should be used to extract plant secondary metabolites, which secondary metabolites have antifungal or broad-spectrum antifungal efects, how do they afect the immune status of the host body, and can they be used as antifungal or immunomodulatory drugs in clinical? All these issues need to be addressed urgently. It is up to more funding institutions and researchers to invest more human and material resources into exploring the vast "plant kingdom."

Author contribution ZYS had the idea for the article; MZ, FJH, and GQ performed the literature search and data analysis; and XZ and FYL drafted and/or critically revised the work. All authors read and approved the manuscript.

Funding This research was supported fnancially by the Sichuan Science and Technology Program (2022NSFSC1539, 2023NSFSC1698, 2023NSFSC0529, and 2022YFS0629) and the Foundation of Southwest Medical University (2022QN042, 2022QN085, 2022QN102, and 2022QN118).

Declarations

Conflict of interest The authors declare no competing interests.

References

- Achan J, Talisuna AO, Erhart A, Yeka A, Tibenderana JK, Baliraine FN, Rosenthal PJ, D'Alessandro U (2011) Quinine, an old antimalarial drug in a modern world: role in the treatment of malaria. Malar J 10:144.<https://doi.org/10.1186/1475-2875-10-144>
- Adamski Z, Blythe LL, Milella L, Bufo SA (2020) Biological activities of alkaloids: from toxicology to pharmacology. Toxins (basel) 12(4):210. <https://doi.org/10.3390/toxins12040210>
- Ademe M (2020) Immunomodulation for the treatment of fungal infections: opportunities and challenges. Front Cell Infect Microbiol 10:469. <https://doi.org/10.3389/fcimb.2020.00469>
- Ahmad A, Khan A, Akhtar F, Yousuf S, Xess I, Khan LA, Manzoor N (2011) Fungicidal activity of thymol and carvacrol by disrupting ergosterol biosynthesis and membrane integrity against Candida. Eur J Clin Microbiol Infect Dis 30(1):41–50. <https://doi.org/10.1007/s10096-010-1050-8>
- Akihisa T, Nakamura Y, Tagata M, Tokuda H, Yasukawa K, Uchiyama E, Suzuki T, Kimura Y (2007) Anti-infammatory and antitumor-promoting efects of triterpene acids and sterols from the fungus Ganoderma lucidum. Chem Biodivers 4(2):224–231. <https://doi.org/10.1002/cbdv.200790027>
- Alara OR, Abdurahman NH, Ukaegbu CI (2021) Extraction of phenolic compounds: a review. Curr Res Food Sci 4:200–214. [https://doi.](https://doi.org/10.1016/j.crfs.2021.03.011) [org/10.1016/j.crfs.2021.03.011](https://doi.org/10.1016/j.crfs.2021.03.011)
- Ali AM, Kunugi H (2021) Propolis, bee honey, and their components protect against coronavirus disease 2019 (COVID-19): a review of in silico, in vitro, and clinical studies. Molecules 26(5):1232. <https://doi.org/10.3390/molecules26051232>
- Alves DR, Maia de Morais S, Tomiotto-Pellissier F, Miranda-Sapla MM, Vasconcelos FR, da Silva ING, Araujo de Sousa H, Assolini JP, Conchon-Costa I, Pavanelli WR, Freire F (2017) Flavonoid composition and biological activities of ethanol extracts of Caryocar coriaceum Wittm., a native plant from Caatinga Biome. Evid Based Complement Alternat Med 2017:6834218. [https://](https://doi.org/10.1155/2017/6834218) doi.org/10.1155/2017/6834218
- Anand V, Govila V, Gulati M, Anand B, Jhingaran R, Rastogi P (2012) Chlorhexidine-thymol varnish as an adjunct to scaling and root planing: a clinical observation. J Oral Biol Craniofac Res 2(2):83–89.<https://doi.org/10.1016/j.jobcr.2012.05.006>
- André S, Rodrigues V, Pemberton S, Laforge M, Fortier Y, Cordeiroda-Silva A, MacDougall J, Estaquier J (2020) Antileishmanial drugs modulate IL-12 expression and infammasome activation in primary human cells. J Immunol 204(7):1869-1880. [https://](https://doi.org/10.4049/jimmunol.1900590) doi.org/10.4049/jimmunol.1900590
- Annamalai G, Suresh K (2018) [6]-Shogaol attenuates infammation, cell proliferation via modulate NF-kappaB and AP-1 oncogenic signaling in 7,12-dimethylbenz[a]anthracene induced oral carcinogenesis. Biomed Pharmacother 98:484–490. [https://doi.org/](https://doi.org/10.1016/j.biopha.2017.12.009) [10.1016/j.biopha.2017.12.009](https://doi.org/10.1016/j.biopha.2017.12.009)
- Anwar S, Bhandari U, Panda BP, Dubey K, Khan W, Ahmad S (2018) Trigonelline inhibits intestinal microbial metabolism of choline and its associated cardiovascular risk. J Pharm Biomed Anal 159:100–112.<https://doi.org/10.1016/j.jpba.2018.06.027>
- Arastehfar A, Gabaldon T, Garcia-Rubio R, Jenks JD, Hoenigl M, Salzer HJF, Ilkit M, Lass-Florl C, Perlin DS (2020) Drug-resistant fungi: an emerging challenge threatening our limited antifungal armamentarium. Antibiotics (basel) 9(12):877. [https://doi.](https://doi.org/10.3390/antibiotics9120877) [org/10.3390/antibiotics9120877](https://doi.org/10.3390/antibiotics9120877)
- Ashokkumar K, Simal-Gandara J, Murugan M, Dhanya MK, Pandian A (2022) Nutmeg (Myristica fragrans Houtt.) essential oil: a review on its composition, biological, and pharmacological activities. Phytother Res 36(7):2839–2851. <https://doi.org/10.1002/ptr.7491>
- Bakkali F, Averbeck S, Averbeck D, Idaomar M (2008) Biological efects of essential oils–a review. Food Chem Toxicol 46(2):446– 475. <https://doi.org/10.1016/j.fct.2007.09.106>
- Balahbib A, El Omari N, Hachlaf NE, Lakhdar F, El Menyiy N, Salhi N, Mrabti HN, Bakrim S, Zengin G, Bouyahya A (2021) Health benefcial and pharmacological properties of p-cymene. Food Chem Toxicol 153:112259. <https://doi.org/10.1016/j.fct.2021.112259>
- Bolarinwa IF, Orfla C, Morgan MR (2015) Determination of amygdalin in apple seeds, fresh apples and processed apple juices. Food Chem 170:437–442. [https://doi.org/10.1016/j.foodchem.](https://doi.org/10.1016/j.foodchem.2014.08.083) [2014.08.083](https://doi.org/10.1016/j.foodchem.2014.08.083)
- Bongomin F, Gago S, Oladele RO, Denning DW (2017) Global and multi-national prevalence of fungal diseases-estimate precision. J Fungi (basel) 3(4):57.<https://doi.org/10.3390/jof3040057>
- Borges RS, Ortiz BLS, Pereira ACM, Keita H, Carvalho JCT (2019) Rosmarinus officinalis essential oil: a review of its phytochemistry, anti-inflammatory activity, and mechanisms of action involved. J Ethnopharmacol 229:29–45. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jep.2018.09.038) [jep.2018.09.038](https://doi.org/10.1016/j.jep.2018.09.038)
- Bureau G, Longpre F, Martinoli MG (2008) Resveratrol and quercetin, two natural polyphenols, reduce apoptotic neuronal cell death induced by neuroinfammation. J Neurosci Res 86(2):403–410. <https://doi.org/10.1002/jnr.21503>
- Burgos RA, Alarcon P, Quiroga J, Manosalva C, Hancke J (2020) Andrographolide, an anti-infammatory multitarget drug: all roads lead to cellular metabolism. Molecules 26(1):5. [https://](https://doi.org/10.3390/molecules26010005) doi.org/10.3390/molecules26010005
- Cacciatore I, Di Giulio M, Fornasari E, Di Stefano A, Cerasa LS, Marinelli L, Turkez H, Di Campli E, Di Bartolomeo S, Robufo I, Cellini L (2015) Carvacrol codrugs: a new approach in the antimicrobial plan. PLoS One 10(4):e0120937. [https://doi.org/](https://doi.org/10.1371/journal.pone.0120937) [10.1371/journal.pone.0120937](https://doi.org/10.1371/journal.pone.0120937)
- Campoy S, Adrio JL (2017) Antifungals. Biochem Pharmacol 133:86– 96.<https://doi.org/10.1016/j.bcp.2016.11.019>
- Cantrell CL, Schrader KK, Mamonov LK, Sitpaeva GT, Kustova TS, Dunbar C, Wedge DE (2005) Isolation and identifcation of antifungal and antialgal alkaloids from Haplophyllum sieversii. J Agric Food Chem 53(20):7741–7748. [https://doi.org/10.1021/](https://doi.org/10.1021/jf051478v) [jf051478v](https://doi.org/10.1021/jf051478v)
- Cao Y, Dai B, Wang Y, Huang S, Xu Y, Cao Y, Gao P, Zhu Z, Jiang Y (2008) In vitro activity of baicalein against Candida albicans bioflms. Int J Antimicrob Agents 32(1):73–77. [https://doi.org/](https://doi.org/10.1016/j.ijantimicag.2008.01.026) [10.1016/j.ijantimicag.2008.01.026](https://doi.org/10.1016/j.ijantimicag.2008.01.026)
- Cao W, Hu C, Wu L, Xu L, Jiang W (2016) Rosmarinic acid inhibits infammation and angiogenesis of hepatocellular carcinoma by suppression of NF-kappaB signaling in H22 tumor-bearing mice. J Pharmacol Sci 132(2):131–137. [https://doi.org/10.1016/j.jphs.](https://doi.org/10.1016/j.jphs.2016.09.003) [2016.09.003](https://doi.org/10.1016/j.jphs.2016.09.003)
- Carolus H, Pierson S, Lagrou K, Van Dijck P (2020) Amphotericin B and other polyenes-discovery, clinical use, mode of action and drug resistance. J Fungi (basel) 6(4):321. [https://doi.org/10.3390/](https://doi.org/10.3390/jof6040321) [jof6040321](https://doi.org/10.3390/jof6040321)
- Chen LL, Zhang HJ, Chao J, Liu JF (2017) Essential oil of Artemisia argyi suppresses infammatory responses by inhibiting JAK/ STATs activation. J Ethnopharmacol 204:107–117. [https://doi.](https://doi.org/10.1016/j.jep.2017.04.017) [org/10.1016/j.jep.2017.04.017](https://doi.org/10.1016/j.jep.2017.04.017)
- Chen X, Chen H, Yuan JS, Kollner TG, Chen Y, Guo Y, Zhuang X, Chen X, Zhang YJ, Fu J, Nebenfuhr A, Guo Z, Chen F (2018) The rice terpene synthase gene OsTPS19 functions as an (S)-limonene synthase in planta, and its overexpression leads to enhanced resistance to the blast fungus Magnaporthe oryzae. Plant Biotechnol J 16(10):1778–1787.<https://doi.org/10.1111/pbi.12914>
- Chen P, Bai Q, Wu Y, Zeng Q, Song X, Guo Y, Zhou P, Wang Y, Liao X, Wang Q, Ren Z, Wang Y (2021) The essential oil of Artemisia argyi H.Lev. and Vaniot attenuates NLRP3 infammasome activation in THP-1 cells. Front Pharmacol 12:712907. [https://](https://doi.org/10.3389/fphar.2021.712907) doi.org/10.3389/fphar.2021.712907
- Chen L, Li L, Chen W (2022a) Use of modeling and simulation to predict the infuence of triazole antifungal agents on the pharmacokinetics of crizotinib. Clin Pharmacol Drug Dev 11(6):724–733. <https://doi.org/10.1002/cpdd.1049>
- Chen M, Cheng T, Xu C, Pan M, Wu J, Wang T, Wu D, Yan G, Wang C, Shao J (2022) Sodium houttuyfonate enhances the monotherapy of fuconazole on oropharyngeal candidiasis (OPC) through HIF-1 α /IL-17 axis by inhibiting cAMP mediated filamentation in Candida albicans-Candida glabrata dual bioflms. Virulence 13(1):428–443. [https://doi.org/10.1080/21505594.](https://doi.org/10.1080/21505594.2022.2035066) [2022.2035066](https://doi.org/10.1080/21505594.2022.2035066)
- Cheng H, Lu C, Tang R, Pan Y, Bao S, Qiu Y, Xie M (2017) Ellagic acid inhibits the proliferation of human pancreatic carcinoma PANC-1 cells in vitro and in vivo. Oncotarget 8(7):12301–12310. <https://doi.org/10.18632/oncotarget.14811>
- Cho HY, Son SY, Rhee HS, Yoon SY, Lee-Parsons CW, Park JM (2008) Synergistic effects of sequential treatment with methyl jasmonate, salicylic acid and yeast extract on benzophenanthridine

alkaloid accumulation and protein expression in Eschscholtzia californica suspension cultures. J Biotechnol 135(1):117–122. <https://doi.org/10.1016/j.jbiotec.2008.02.020>

- Cho YK, Shin EY, Uehara H, Ambati B (2017) Efect of itraconazole on the cornea in a murine suture model and penetrating keratoplasty model. Int J Ophthalmol 10(11):1647–1654. [https://doi.](https://doi.org/10.18240/ijo.2017.11.03) [org/10.18240/ijo.2017.11.03](https://doi.org/10.18240/ijo.2017.11.03)
- Choi JH, Kwon EY, Park CM, Choi SM, Lee DG, Yoo JH, Shin WS, Stevens DA (2010) Immunomodulatory effects of antifungal agents on the response of human monocytic cells to Aspergillus fumigatus conidia. Med Mycol 48(5):704–709. [https://doi.org/](https://doi.org/10.3109/13693780903471784) [10.3109/13693780903471784](https://doi.org/10.3109/13693780903471784)
- Chtioui W, Balmas V, Delogu G, Migheli Q, Oufensou S (2022) Bioprospecting phenols as inhibitors of trichothecene-producing Fusarium: sustainable approaches to the management of wheat pathogens. Toxins (basel) 14(2):72. [https://doi.org/10.3390/toxin](https://doi.org/10.3390/toxins14020072) [s14020072](https://doi.org/10.3390/toxins14020072)
- Cressey P, Saunders D, Goodman J (2013) Cyanogenic glycosides in plant-based foods available in New Zealand. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 30(11):1946–53. <https://doi.org/10.1080/19440049.2013.825819>
- Cui Z, Zhao X, Amevor FK, Du X, Wang Y, Li D, Shu G, Tian Y, Zhao X (2022) Therapeutic application of quercetin in agingrelated diseases: SIRT1 as a potential mechanism. Front Immunol 13:943321. [https://doi.org/10.3389/fmmu.2022.943321](https://doi.org/10.3389/fimmu.2022.943321)
- Da X, Nishiyama Y, Tie D, Hein KZ, Yamamoto O, Morita E (2019) Antifungal activity and mechanism of action of Ou-gon (Scutellaria root extract) components against pathogenic fungi. Sci Rep 9(1):1683. <https://doi.org/10.1038/s41598-019-38916-w>
- da Fonseca STD, Teixeira TR, Ferreira JMS, Lima L, Luyten W, Castro AHF (2022) Flavonoid-rich fractions of Bauhinia holophylla leaves inhibit Candida albicans bioflm formation and hyphae growth. Plants (basel) 11(14):1796. [https://doi.org/10.3390/plant](https://doi.org/10.3390/plants11141796) [s11141796](https://doi.org/10.3390/plants11141796)
- da Silva Bomfm N, Kohiyama CY, Nakasugi LP, Nerilo SB, Mossini SAG, Romoli JCZ, Graton Mikcha JM, Abreu Filho BA, Machinski M Jr (2020) Antifungal and antiafatoxigenic activity of rosemary essential oil (Rosmarinus officinalis L.) against Aspergillus favus. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 37(1):153–161. [https://doi.org/10.1080/19440049.](https://doi.org/10.1080/19440049.2019.1678771) [2019.1678771](https://doi.org/10.1080/19440049.2019.1678771)
- Dal Piaz F, Bader A, Malafronte N, D'Ambola M, Petrone AM, Porta A, Ben Hadda T, De Tommasi N, Bisio A, Severino L (2018) Phytochemistry of compounds isolated from the leaf-surface extract of Psiadia punctulata (DC.) Vatke growing in Saudi Arabia. Phytochemistry 155:191–202. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.phytochem.2018.08.003) [phytochem.2018.08.003](https://doi.org/10.1016/j.phytochem.2018.08.003)
- Danielli LJ, Pippi B, Duarte JA, Maciel AJ, Lopes W, Machado MM, Oliveira LFS, Vainstein MH, Teixeira ML, Bordignon SAL, Fuentefria AM, Apel MA (2018) Antifungal mechanism of action of Schinus lentiscifolius Marchand essential oil and its synergistic efect in vitro with terbinafne and ciclopirox against dermatophytes. J Pharm Pharmacol 70(9):1216–1227. [https://](https://doi.org/10.1111/jphp.12949) doi.org/10.1111/jphp.12949
- de Araújo FF, de Paulo Farias D, Neri-Numa IA, Pastore GM (2021) Polyphenols and their applications: an approach in food chemistry and innovation potential. Food Chem 338:127535. [https://](https://doi.org/10.1016/j.foodchem.2020.127535) doi.org/10.1016/j.foodchem.2020.127535
- de Castro RD, de Souza TM, Bezerra LM, Ferreira GL, Costa EM, Cavalcanti AL (2015) Antifungal activity and mode of action of thymol and its synergism with nystatin against Candida species involved with infections in the oral cavity: an in vitro study. BMC Complement Altern Med 15:417. [https://doi.org/10.1186/](https://doi.org/10.1186/s12906-015-0947-2) [s12906-015-0947-2](https://doi.org/10.1186/s12906-015-0947-2)
- Delma FZ, Al-Hatmi AMS, Bruggemann RJM, Melchers WJG, de Hoog S, Verweij PE, Buil JB (2021) Molecular mechanisms of

5-Fluorocytosine resistance in yeasts and flamentous fungi. J Fungi (basel) 7(11):909.<https://doi.org/10.3390/jof7110909>

- Dignani MC (2014) Epidemiology of invasive fungal diseases on the basis of autopsy reports. F1000Prime Rep 6:81. [https://doi.org/](https://doi.org/10.12703/p6-81) [10.12703/p6-81](https://doi.org/10.12703/p6-81)
- Dominguez-Avila JA, Villa-Rodriguez JA, Montiel-Herrera M, Pacheco-Ordaz R, Roopchand DE, Venema K, Gonzalez-Aguilar GA (2021) Phenolic compounds promote diversity of gut microbiota and maintain colonic health. Dig Dis Sci 66(10):3270– 3289. <https://doi.org/10.1007/s10620-020-06676-7>
- Domínguez-Avila JA, Salazar-López NJ, Montiel-Herrera M, Martínez-Martínez A, Villegas-Ochoa MA, González-Aguilar GA (2022) Phenolic compounds can induce systemic and central immunomodulation, which result in a neuroprotective efect. J Food Biochem 46(12):e14260. [https://doi.org/10.1111/jfbc.](https://doi.org/10.1111/jfbc.14260) [14260](https://doi.org/10.1111/jfbc.14260)
- Du ZA, Sun MN, Hu ZS (2018) Saikosaponin a ameliorates LPS-Induced acute lung injury in mice. Infammation 41(1):193–198. <https://doi.org/10.1007/s10753-017-0677-3>
- Du H, Bing J, Hu T, Ennis CL, Nobile CJ, Huang G (2020) Candida auris: epidemiology, biology, antifungal resistance, and virulence. PLoS Pathog 16(10):e1008921. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.ppat.1008921) [journal.ppat.1008921](https://doi.org/10.1371/journal.ppat.1008921)
- Duraipandiyan V, Ignacimuthu S (2009) Antibacterial and antifungal activity of findersine isolated from the traditional medicinal plant, Toddalia asiatica (L.) Lam. J Ethnopharmacol 123(3):494– 8.<https://doi.org/10.1016/j.jep.2009.02.020>
- Endale M, Park SC, Kim S, Kim SH, Yang Y, Cho JY, Rhee MH (2013) Quercetin disrupts tyrosine-phosphorylated phosphatidylinositol 3-kinase and myeloid diferentiation factor-88 association, and inhibits MAPK/AP-1 and IKK/NF-kappaB-induced infammatory mediators production in RAW 264.7 cells. Immunobiology 218(12):1452–67.<https://doi.org/10.1016/j.imbio.2013.04.019>
- Erb M, Kliebenstein DJ (2020) Plant secondary metabolites as defenses, regulators, and primary metabolites: the blurred functional trichotomy. Plant Physiol 184(1):39–52. [https://doi.org/](https://doi.org/10.1104/pp.20.00433) [10.1104/pp.20.00433](https://doi.org/10.1104/pp.20.00433)
- Fernandez LF, Palomino OM, Frutos G (2014) Effectiveness of Rosmarinus officinalis essential oil as antihypotensive agent in primary hypotensive patients and its infuence on health-related quality of life. J Ethnopharmacol 151(1):509-516. [https://doi.](https://doi.org/10.1016/j.jep.2013.11.006) [org/10.1016/j.jep.2013.11.006](https://doi.org/10.1016/j.jep.2013.11.006)
- Fowler ZL, Shah K, Panepinto JC, Jacobs A, Koffas MA (2011) Development of non-natural favanones as antimicrobial agents. PLoS One 6(10):e25681. <https://doi.org/10.1371/journal.pone.0025681>
- Freires IA, Queiroz V, Furletti VF, Ikegaki M, de Alencar SM, Duarte MCT, Rosalen PL (2016) Chemical composition and antifungal potential of Brazilian propolis against Candida spp. J Mycol Med 26(2):122–132. <https://doi.org/10.1016/j.mycmed.2016.01.003>
- Furuno JP, Tallman GB, Noble BN, Bubalo JS, Forrest GN, Lewis JS 2nd, Bienvenida AF, Holmes CA, Weber BR, McGregor JC (2018) Clinical outcomes of oral suspension versus delayedrelease tablet formulations of posaconazole for prophylaxis of invasive fungal infections. Antimicrob Agents Chemother 62(10):e00893-e918. <https://doi.org/10.1128/aac.00893-18>
- Ganesan K, Xu B (2017) Polyphenol-rich lentils and their health promoting efects. Int J Mol Sci 18(11):2390. [https://doi.org/10.](https://doi.org/10.3390/ijms18112390) [3390/ijms18112390](https://doi.org/10.3390/ijms18112390)
- Geraets L, Moonen HJ, Brauers K, Wouters EF, Bast A, Hageman GJ (2007) Dietary flavones and flavonoles are inhibitors of poly(ADP-ribose)polymerase-1 in pulmonary epithelial cells. J Nutr 137(10):2190–2195. <https://doi.org/10.1093/jn/137.10.2190>
- Gu L, Deng WS, Liu Y, Jiang CH, Sun LC, Sun XF, Xu Q, Zhou H (2014) Ellagic acid protects Lipopolysaccharide/D-galactosamine-induced acute hepatic injury in mice. Int Immunopharmacol 22(2):341–345.<https://doi.org/10.1016/j.intimp.2014.07.005>
- Guo S, Yan T, Shi L, Liu A, Zhang T, Xu Y, Jiang W, Yang Q, Yang L, Liu L, Zhao R, Zhang S (2021) Matrine, as a CaSR agonist promotes intestinal GLP-1 secretion and improves insulin resistance in diabetes mellitus. Phytomedicine 84:153507. [https://doi.](https://doi.org/10.1016/j.phymed.2021.153507) [org/10.1016/j.phymed.2021.153507](https://doi.org/10.1016/j.phymed.2021.153507)
- Hammer KA, Carson CF, Riley TV (2002) In vitro activity of Melaleuca alternifolia (tea tree) oil against dermatophytes and other flamentous fungi. J Antimicrob Chemother 50(2):195–199. <https://doi.org/10.1093/jac/dkf112>
- Han Y, Lee JH (2005) Berberine synergy with amphotericin B against disseminated candidiasis in mice. Biol Pharm Bull 28(3):541– 544. <https://doi.org/10.1248/bpb.28.541>
- Han Q, Yuan Q, Meng X, Huo J, Bao Y, Xie G (2017) 6-Shogaol attenuates LPS-induced infammation in BV2 microglia cells by activating PPAR-gamma. Oncotarget 8(26):42001–42006. <https://doi.org/10.18632/oncotarget.16719>
- Han X, Xu T, Fang Q, Zhang H, Yue L, Hu G, Sun L (2021) Quercetin hinders microglial activation to alleviate neurotoxicity via the interplay between NLRP3 infammasome and mitophagy. Redox Biol 44:102010.<https://doi.org/10.1016/j.redox.2021.102010>
- Havlickova B, Czaika VA, Friedrich M (2008) Epidemiological trends in skin mycoses worldwide. Mycoses 51(Suppl 4):2–15. [https://](https://doi.org/10.1111/j.1439-0507.2008.01606.x) doi.org/10.1111/j.1439-0507.2008.01606.x
- Hu J, Shi X, Chen J, Mao X, Zhu L, Yu L, Shi J (2014) Alkaloids from Toddalia asiatica and their cytotoxic, antimicrobial and antifungal activities. Food Chem 148:437–444. [https://doi.org/](https://doi.org/10.1016/j.foodchem.2012.12.058) [10.1016/j.foodchem.2012.12.058](https://doi.org/10.1016/j.foodchem.2012.12.058)
- Huang T, Jander G, de Vos M (2011) Non-protein amino acids in plant defense against insect herbivores: representative cases and opportunities for further functional analysis. Phytochemistry 72(13):1531–1537. [https://doi.org/10.1016/j.phytochem.2011.](https://doi.org/10.1016/j.phytochem.2011.03.019) [03.019](https://doi.org/10.1016/j.phytochem.2011.03.019)
- Huang X, Yi Y, Yong J, Sun J, Song Z, Li D, Li Y (2021) Inhibitory effect of berberine hydrochloride against Candida albicans and the role of the HOG-MAPK pathway. J Antibiot (tokyo) 74(11):807–816.<https://doi.org/10.1038/s41429-021-00463-w>
- Huang X, Zheng D, Yong J, Li Y (2022) Antifungal activity and potential mechanism of berberine hydrochloride against fuconazoleresistant Candida albicans. J Med Microbiol 71(6):10. [https://](https://doi.org/10.1099/jmm.0.001542) doi.org/10.1099/jmm.0.001542
- Hugo Infante V, Maria Maia Campos P, Darvin M, Lohan S, Schleusener J, Schanzer S, Lademann J, Meinke M (2023) Cosmetic formulations with Melaleuca alternifolia essential oil for the improvement of photoaged skin: a double-blind, randomized, placebo-controlled clinical study. Photochem Photobiol 99(1):176–183.<https://doi.org/10.1111/php.13660>
- Hussein KA, Lee YD, Joo JH (2020) Efect of rosemary essential oil and Trichoderma koningiopsis VOCs on pathogenic fungi responsible for ginseng root-rot disease. J Microbiol Biotechnol 30(7):1018–1026. <https://doi.org/10.4014/jmb.2002.02013>
- Imenshahidi M, Hosseinzadeh H (2019) Berberine and barberry (Berberis vulgaris): a clinical review. Phytother Res 33(3):504–523. <https://doi.org/10.1002/ptr.6252>
- Itoh K, Shigemi H, Chihara K, Sada K, Yamauchi T, Iwasaki H (2021) Caspofungin suppresses zymosan-induced cytokine and chemokine release in THP-1 cells: possible involvement of the spleen tyrosine kinase pathway. Transl Res 227:53–63. [https://](https://doi.org/10.1016/j.trsl.2020.07.005) doi.org/10.1016/j.trsl.2020.07.005
- Jafri H, Ahmad I (2020) Thymus vulgaris essential oil and thymol inhibit bioflms and interact synergistically with antifungal drugs against drug resistant strains of Candida albicans and Candida tropicalis. J Mycol Med 30(1):100911. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.mycmed.2019.100911) [mycmed.2019.100911](https://doi.org/10.1016/j.mycmed.2019.100911)
- Jahanbakhshi F, Maleki Dana P, Badehnoosh B, Yousef B, Mansournia MA, Jahanshahi M, Asemi Z, Halajzadeh J (2021) Curcumin anti-tumor effects on endometrial cancer with focus on its
- Jayachandran M, Chandrasekaran B, Namasivayam N (2015) Geraniol attenuates fbrosis and exerts anti-infammatory efects on diet induced atherogenesis by NF-kappaB signaling pathway. Eur J Pharmacol 762:102–111. [https://doi.org/10.1016/j.ejphar.2015.](https://doi.org/10.1016/j.ejphar.2015.05.039) [05.039](https://doi.org/10.1016/j.ejphar.2015.05.039)
- Jediyi H, Naamani K, Elkoch AA, Dihazi A, Lemjiber N (2020) A comparative study of phenols composition, antioxidant, and antifungal potency of leaves extract from five Moroccan Vitis vinifera L. varieties. J Food Saf 40:e12756. [https://doi.org/10.](https://doi.org/10.1111/jfs.12756) [1111/jfs.12756](https://doi.org/10.1111/jfs.12756)
- Jiang J, Meng Y, Hu S, Botchway BOA, Zhang Y, Liu X (2020) Saikosaponin D: a potential therapeutic drug for osteoarthritis. J Tissue Eng Regen Med 14(8):1175–1184. [https://doi.org/10.](https://doi.org/10.1002/term.3090) [1002/term.3090](https://doi.org/10.1002/term.3090)
- Jin YS (2019) Recent advances in natural antifungal favonoids and their derivatives. Bioorg Med Chem Lett 29(19):126589. [https://](https://doi.org/10.1016/j.bmcl.2019.07.048) doi.org/10.1016/j.bmcl.2019.07.048
- Judzentiene A, Budiene J, Svediene J, Garjonyte R (2020) Toxic, radical scavenging, and antifungal activity of Rhododendron tomentosum H. essential oils. Molecules 25(7):1676. [https://doi.org/10.](https://doi.org/10.3390/molecules25071676) [3390/molecules25071676](https://doi.org/10.3390/molecules25071676)
- Kai K, Wang R, Bi W, Ma Z, Shi W, Ye Y, Zhang D (2021) Chlorogenic acid induces ROS-dependent apoptosis in Fusarium fujikuroi and decreases the postharvest rot of cherry tomato. World J Microbiol Biotechnol 37(6):93. [https://doi.org/10.1007/](https://doi.org/10.1007/s11274-021-03062-x) [s11274-021-03062-x](https://doi.org/10.1007/s11274-021-03062-x)
- Kalemba D, Kunicka A (2003) Antibacterial and antifungal properties of essential oils. Curr Med Chem 10(10):813–829. [https://doi.](https://doi.org/10.2174/0929867033457719) [org/10.2174/0929867033457719](https://doi.org/10.2174/0929867033457719)
- Kamal LZM, Adam MAA, Shahpudin SNM, Shuib AN, Sandai R, Hassan NM, Tabana Y, Basri DF, Than LTL, Sandai D (2021) Identifcation of alkaloid compounds arborinine and graveoline from Ruta angustifolia (L.) Pers for their antifungal potential against isocitrate lyase (ICL1) gene of Candida albicans. Mycopathologia 186(2):221–236. [https://doi.org/10.1007/](https://doi.org/10.1007/s11046-020-00523-z) [s11046-020-00523-z](https://doi.org/10.1007/s11046-020-00523-z)
- Kamatou GP, Vermaak I, Viljoen AM, Lawrence BM (2013) Menthol: a simple monoterpene with remarkable biological properties. Phytochemistry 96:15–25. [https://doi.org/10.1016/j.phyto](https://doi.org/10.1016/j.phytochem.2013.08.005) [chem.2013.08.005](https://doi.org/10.1016/j.phytochem.2013.08.005)
- Kang JH, Sung MK, Kawada T, Yoo H, Kim YK, Kim JS, Yu R (2005) Soybean saponins suppress the release of proinfammatory mediators by LPS-stimulated peritoneal macrophages. Cancer Lett 230(2):219–227.<https://doi.org/10.1016/j.canlet.2004.12.041>
- Khurana A, Sardana K, Chowdhary A (2019) Antifungal resistance in dermatophytes: recent trends and therapeutic implications. Fungal Genet Biol 132:103255. [https://doi.org/10.1016/j.fgb.](https://doi.org/10.1016/j.fgb.2019.103255) [2019.103255](https://doi.org/10.1016/j.fgb.2019.103255)
- Kim J, Ha Quang Bao T, Shin YK, Kim KY (2018) Antifungal activity of magnoforine against Candida strains. World J Microbiol Biotechnol 34(11):167. <https://doi.org/10.1007/s11274-018-2549-x>
- Kim SY, Han SD, Kim M, Mony TJ, Lee ES, Kim KM, Choi SH, Hong SH, Choi JW, Park SJ (2021) Mentha arvensis essential oil exerts anti-infammatory in LPS-stimulated infammatory responses via inhibition of ERK/NF-kappaB signaling pathway and antiatopic dermatitis-like efects in 2,4-dinitrochlorobezene-induced BALB/c mice. Antioxidants (basel) 10(12):1941. [https://doi.org/](https://doi.org/10.3390/antiox10121941) [10.3390/antiox10121941](https://doi.org/10.3390/antiox10121941)
- Kim BK, Choi JY, Hong KT, An HY, Shin HY, Kang HJ (2022) Prospective study on prophylactic micafungin sodium against invasive fungal disease during neutropenia in pediatric & adolescent patients undergoing autologous hematopoietic stem cell transplantation. Children (basel) 9(3):372. [https://doi.org/10.3390/](https://doi.org/10.3390/children9030372) [children9030372](https://doi.org/10.3390/children9030372)
- Kolouchova I, Matatkova O, Paldrychova M, Kodes Z, Kvasnickova E, Sigler K, Cejkova A, Smidrkal J, Demnerova K, Masak J (2018) Resveratrol, pterostilbene, and baicalein: plant-derived antibioflm agents. Folia Microbiol (praha) 63(3):261–272. [https://](https://doi.org/10.1007/s12223-017-0549-0) doi.org/10.1007/s12223-017-0549-0
- Kong Y, Peng Q, Lv N, Yuan J, Deng Z, Liang X, Chen S, Wang L (2020) Paeoniforin exerts neuroprotective efects in a transgenic mouse model of Alzheimer's disease via activation of adenosine A(1) receptor. Neurosci Lett 730:135016. [https://doi.org/10.](https://doi.org/10.1016/j.neulet.2020.135016) [1016/j.neulet.2020.135016](https://doi.org/10.1016/j.neulet.2020.135016)
- Koo HJ, Song YS, Kim HJ, Lee YH, Hong SM, Kim SJ, Kim BC, Jin C, Lim CJ, Park EH (2004) Antiinfammatory efects of genipin, an active principle of gardenia. Eur J Pharmacol 495(2–3):201– 208. <https://doi.org/10.1016/j.ejphar.2004.05.031>
- Krawisz AK, Raja A, Secemsky EA (2021) Femoral-popliteal peripheral artery disease: from symptom presentation to management and treatment controversies. Prog Cardiovasc Dis 65:15–22. <https://doi.org/10.1016/j.pcad.2021.02.004>
- Kumar PS (2020) The infuence of Azadirachta indica, Melaleuca alternifolia, and Cocos nucifera on Candida albicans strain in tissue conditioner at varying time intervals. J Indian Prosthodont Soc 20(2):171–179. https://doi.org/10.4103/jips.jips_366_19
- Kumar G, Dange P, Kailaje V, Vaidya MM, Ramchandani AG, Maru GB (2012) Polymeric black tea polyphenols modulate the localization and activity of 12-O-tetradecanoylphorbol-13-acetate-mediated kinases in mouse skin: mechanisms of their anti-tumor-promoting action. Free Radic Biol Med 53(6):1358– 1370. <https://doi.org/10.1016/j.freeradbiomed.2012.07.017>
- Lai CC, Yu WL (2021) COVID-19 associated with pulmonary aspergillosis: a literature review. J Microbiol Immunol Infect 54(1):46– 53.<https://doi.org/10.1016/j.jmii.2020.09.004>
- Lam A, Hoang JD, Singleton A, Han X, Bleier BS (2015) Itraconazole and clarithromycin inhibit P-glycoprotein activity in primary human sinonasal epithelial cells. Int Forum Allergy Rhinol 5(6):477–480. <https://doi.org/10.1002/alr.21454>
- Lei J, Xiao W, Zhang J, Liu F, Xin C, Zhou B, Chen W, Song Z (2022) Antifungal activity of vitamin $D_{(3)}$ against Candida albicans in vitro and in vivo. Microbiol Res 265:127200. [https://doi.org/](https://doi.org/10.1016/j.micres.2022.127200) [10.1016/j.micres.2022.127200](https://doi.org/10.1016/j.micres.2022.127200)
- Lei J, Huang J, Xin C, Liu F, Zhang J, Xie Y, Mao Y, Chen W, Song Z (2023) Ribofavin targets the cellular metabolic and ribosomal pathways of Candida albicans in vitro and exhibits efficacy against oropharyngeal candidiasis. Microbiol Spectr 11(1):e0380122. <https://doi.org/10.1128/spectrum.03801-22>
- Leite MC, de Brito Bezerra AP, de Sousa JP, de Oliveira LE (2015) Investigating the antifungal activity and mechanism(s) of geraniol against Candida albicans strains. Med Mycol 53(3):275–284. <https://doi.org/10.1093/mmy/myu078>
- Li DD, Zhao LX, Mylonakis E, Hu GH, Zou Y, Huang TK, Yan L, Wang Y, Jiang YY (2014) In vitro and in vivo activities of pterostilbene against Candida albicans bioflms. Antimicrob Agents Chemother 58(4):2344–2355.<https://doi.org/10.1128/aac.01583-13>
- Li Y, Yao J, Han C, Yang J, Chaudhry MT, Wang S, Liu H, Yin Y (2016) Quercetin, inflammation and immunity. Nutrients 8(3):167.<https://doi.org/10.3390/nu8030167>
- Li WW, Wang TY, Cao B, Liu B, Rong YM, Wang JJ, Wei F, Wei LQ, Chen H, Liu YX (2019) Synergistic protection of matrine and lycopene against lipopolysaccharide-induced acute lung injury in mice. Mol Med Rep 20(1):455–462. [https://doi.org/10.3892/](https://doi.org/10.3892/mmr.2019.10278) [mmr.2019.10278](https://doi.org/10.3892/mmr.2019.10278)
- Li J, Li F, Tang F, Zhang J, Li R, Sheng D, Lee SM, Zhou GC, Leung GP (2020a) AGS-30, an andrographolide derivative, suppresses tumor angiogenesis and growth in vitro and in vivo. Biochem Pharmacol 171:113694.<https://doi.org/10.1016/j.bcp.2019.113694>
- Li S, Liu X, Chen X, Bi L (2020b) Research progress on anti-infammatory efects and mechanisms of alkaloids from Chinese medical

herbs. Evid Based Complement Alternat Med 2020:1303524. <https://doi.org/10.1155/2020/1303524>

- Lim SH, Nam KH, Kim K, Yi SA, Lee J, Han JW (2020) Rosmarinic acid methyl ester regulates ovarian cancer cell migration and reverses cisplatin resistance by inhibiting the expression of forkhead box M1. Pharmaceuticals (basel) 13(10):302. [https://doi.](https://doi.org/10.3390/ph13100302) [org/10.3390/ph13100302](https://doi.org/10.3390/ph13100302)
- Liu Q, Peng YB, Zhou P, Qi LW, Zhang M, Gao N, Liu EH, Li P (2013) 6-Shogaol induces apoptosis in human leukemia cells through a process involving caspase-mediated cleavage of eIF2alpha. Mol Cancer 12(1):135.<https://doi.org/10.1186/1476-4598-12-135>
- Liu C, Dunkin D, Lai J, Song Y, Ceballos C, Benkov K, Li XM (2015) Anti-infammatory efects of Ganoderma lucidum triterpenoid in human Crohn's disease associated with downregulation of NFkappaB signaling. Infamm Bowel Dis 21(8):1918–1925. [https://](https://doi.org/10.1097/MIB.0000000000000439) doi.org/10.1097/MIB.0000000000000439
- Liu Z, Lv Y, Zhang Y, Liu F, Zhu L, Pan S, Qiu C, Guo Y, Yang T, Wang J (2017) Matrine-type alkaloids inhibit advanced glycation end products induced reactive oxygen species-mediated apoptosis of aortic endothelial cells in vivo and in vitro by targeting MKK3 and p38MAPK signaling. J Am Heart Assoc 6(12):e007441.<https://doi.org/10.1161/JAHA.117.007441>
- Liu X, Qiu Y, Liu Y, Huang N, Hua C, Wang Q, Wu Z, Lu J, Song P, Xu J, Li P, Yin Y (2021) Citronellal ameliorates doxorubicininduced hepatotoxicity via antioxidative stress, antiapoptosis, and proangiogenesis in rats. J Biochem Mol Toxicol 35(2):e22639. <https://doi.org/10.1002/jbt.22639>
- Loi M, Paciolla C, Logrieco AF, Mule G (2020) Plant bioactive compounds in pre- and postharvest management for afatoxins reduction. Front Microbiol 11:243.<https://doi.org/10.3389/fmicb.2020.00243>
- Lu H, Wu L, Liu L, Ruan Q, Zhang X, Hong W, Wu S, Jin G, Bai Y (2018) Quercetin ameliorates kidney injury and fbrosis by modulating M1/M2 macrophage polarization. Biochem Pharmacol 154:203–212.<https://doi.org/10.1016/j.bcp.2018.05.007>
- Luo Y, Lu S, Gao Y, Yang K, Wu D, Xu X, Sun G, Sun X (2020) Araloside C attenuates atherosclerosis by modulating macrophage polarization via Sirt1-mediated autophagy. Aging (Albany NY) 12(2):1704–1724.<https://doi.org/10.18632/aging.102708>
- Luo N, Jin L, Yang C, Zhu Y, Ye X, Li X, Zhang B (2021) Antifungal activity and potential mechanism of magnoforine against Trichophyton rubrum. J Antibiot (tokyo) 74(3):206–214. [https://doi.](https://doi.org/10.1038/s41429-020-00380-4) [org/10.1038/s41429-020-00380-4](https://doi.org/10.1038/s41429-020-00380-4)
- Ma N, Zhang Z, Liao F, Jiang T, Tu Y (2020) The birth of artemisinin. Pharmacol Ther 216:107658. [https://doi.org/10.1016/j.pharm](https://doi.org/10.1016/j.pharmthera.2020.107658) [thera.2020.107658](https://doi.org/10.1016/j.pharmthera.2020.107658)
- Ma SR, Tong Q, Lin Y, Pan LB, Fu J, Peng R, Zhang XF, Zhao ZX, Li Y, Yu JB, Cong L, Han P, Zhang ZW, Yu H, Wang Y, Jiang JD (2022a) Berberine treats atherosclerosis via a vitamine-like efect down-regulating Choline-TMA-TMAO production pathway in gut microbiota. Signal Transduct Target Ther 7(1):207. [https://](https://doi.org/10.1038/s41392-022-01027-6) doi.org/10.1038/s41392-022-01027-6
- Ma Y, Wang L, Lu A, Xue W (2022b) Synthesis and biological activity of novel oxazinyl favonoids as antiviral and anti-phytopathogenic fungus agents. Molecules 27(20):6875. [https://doi.org/10.](https://doi.org/10.3390/molecules27206875) [3390/molecules27206875](https://doi.org/10.3390/molecules27206875)
- Mahor A, Sawant DM, Goyal AK (2022) Chemical and physical approaches for improved biopharmaceutical activity of amphotericin B: current and future prospective. Curr Top Med Chem 22(19):1571–1592. [https://doi.org/10.2174/156802662266622](https://doi.org/10.2174/1568026622666220610141243) [0610141243](https://doi.org/10.2174/1568026622666220610141243)
- Manohar V, Ingram C, Gray J, Talpur NA, Echard BW, Bagchi D, Preuss HG (2001) Antifungal activities of origanum oil against Candida albicans. Mol Cell Biochem 228(1–2):111–117. [https://](https://doi.org/10.1023/a:1013311632207) doi.org/10.1023/a:1013311632207
- Mao QQ, Xu XY, Cao SY, Gan RY, Corke H, Beta T, Li HB (2019) Bioactive compounds and bioactivities of ginger (Zingiber

officinale Roscoe). Foods 8(6) [https://doi.org/10.3390/foods](https://doi.org/10.3390/foods8060185) [8060185](https://doi.org/10.3390/foods8060185)

- Massa N, Cantamessa S, Novello G, Ranzato E, Martinotti S, Pavan M, Rocchetti A, Berta G, Gamalero E, Bona E (2018) Antifungal activity of essential oils against azole-resistant and azolesusceptible vaginal Candida glabrata strains. Can J Microbiol 64(10):647–663.<https://doi.org/10.1139/cjm-2018-0082>
- Menon S, Liang X, Vartak R, Patel K, Di Stefano A, Cacciatore I, Marinelli L, Billack B (2021) Antifungal activity of novel formulations based on terpenoid prodrugs against C. albicans in a mouse model. Pharmaceutics 13(5):633. [https://doi.org/10.3390/](https://doi.org/10.3390/pharmaceutics13050633) [pharmaceutics13050633](https://doi.org/10.3390/pharmaceutics13050633)
- Mickymaray S (2019) Efficacy and mechanism of traditional medicinal plants and bioactive compounds against clinically important pathogens. Antibiotics (basel) 8(4):257. [https://doi.org/10.3390/](https://doi.org/10.3390/antibiotics8040257) [antibiotics8040257](https://doi.org/10.3390/antibiotics8040257)
- Middleton E Jr (1998) Efect of plant favonoids on immune and infammatory cell function. Adv Exp Med Biol 439:175–182. https://doi.org/10.1007/978-1-4615-5335-9_13
- Mirza Alizadeh A, Golzan SA, Mahdavi A, Dakhili S, Torki Z, Hosseini H (2022) Recent advances on the efficacy of essential oils on mycotoxin secretion and their mode of action. Crit Rev Food Sci Nutr 62(17):4726–4751. [https://doi.org/10.1080/10408398.](https://doi.org/10.1080/10408398.2021.1878102) [2021.1878102](https://doi.org/10.1080/10408398.2021.1878102)
- Mondello F, De Bernardis F, Girolamo A, Cassone A, Salvatore G (2006) In vivo activity of terpinen-4-ol, the main bioactive component of Melaleuca alternifolia Cheel (tea tree) oil against azolesusceptible and -resistant human pathogenic Candida species. BMC Infect Dis 6:158. <https://doi.org/10.1186/1471-2334-6-158>
- Moradi-Marjaneh R, Hassanian SM, Rahmani F, Aghaee-Bakhtiari SH, Avan A, Khazaei M (2018) Phytosomal curcumin elicits anti-tumor properties through suppression of angiogenesis, cell proliferation and induction of oxidative stress in colorectal cancer. Curr Pharm Des 24(39):4626–4638. [https://doi.org/10.2174/](https://doi.org/10.2174/1381612825666190110145151) [1381612825666190110145151](https://doi.org/10.2174/1381612825666190110145151)
- Moretti S, Bozza S, Massi-Benedetti C, Prezioso L, Rossetti E, Romani L, Aversa F, Pitzurra L (2014) An immunomodulatory activity of micafungin in preclinical aspergillosis. J Antimicrob Chemother 69(4):1065–1074. <https://doi.org/10.1093/jac/dkt457>
- Munoz JE, Rossi DCP, Jabes DL, Barbosa DA, Cunha FFM, Nunes LR, Arruda DC, Pelleschi Taborda C (2020) In vitro and in vivo inhibitory activity of limonene against diferent isolates of Candida spp. J Fungi (basel) 6(3):183.<https://doi.org/10.3390/jof6030183>
- Nidhi P, Rolta R, Kumar V, Dev K, Sourirajan A (2020) Synergistic potential of Citrus aurantium L. essential oil with antibiotics against Candida albicans. J Ethnopharmacol 262:113135. [https://](https://doi.org/10.1016/j.jep.2020.113135) doi.org/10.1016/j.jep.2020.113135
- Nikolić I, Chua EG, Tay ACY, Kostrešević A, Pavlović B, Jončić Savić K (2023) Savory, oregano and thyme essential oil mixture (HerbELICO(®)) counteracts Helicobacter pylori. Molecules 28(5):2138.<https://doi.org/10.3390/molecules28052138>
- Nivoix Y, Ledoux MP, Herbrecht R (2020) Antifungal therapy: new and evolving therapies. Semin Respir Crit Care Med 41(1):158– 174. <https://doi.org/10.1055/s-0039-3400291>
- O'Brien DM, Vallieres C, Alexander C, Howdle SM, Stockman RA, Avery SV (2019) Epoxy-amine oligomers from terpenes with applications in synergistic antifungal treatments. J Mater Chem B 7(34):5222–5229.<https://doi.org/10.1039/c9tb00878k>
- Okamoto-Shibayama K, Yoshida A, Ishihara K (2021) Inhibitory efect of resveratrol on Candida albicans bioflm formation. Bull Tokyo Dent Coll 62(1):1–6. [https://doi.org/10.2209/tdcpublica](https://doi.org/10.2209/tdcpublication.2020-0023) [tion.2020-0023](https://doi.org/10.2209/tdcpublication.2020-0023)
- Ostovan M, Fazljou SMB, Khazraei H, Araj Khodaei M, Torbati M (2020) The anti-infammatory efect of Pistacia Lentiscus in a rat model of colitis. J Infamm Res 13:369–376. [https://doi.org/](https://doi.org/10.2147/JIR.S259035) [10.2147/JIR.S259035](https://doi.org/10.2147/JIR.S259035)
- OuYang Q, Liu Y, Oketch OR, Zhang M, Shao X, Tao N (2021) Citronellal exerts its antifungal activity by targeting ergosterol biosynthesis in Penicillium digitatum. J Fungi (basel) 7(6):432. [https://](https://doi.org/10.3390/jof7060432) doi.org/10.3390/jof7060432
- Pandey AK, Kumar P, Singh P, Tripathi NN, Bajpai VK (2016) Essential oils: sources of antimicrobials and food preservatives. Front Microbiol 7:2161.<https://doi.org/10.3389/fmicb.2016.02161>
- Pappas PG, Kaufman CA, Andes DR, Clancy CJ, Marr KA, Ostrosky-Zeichner L, Reboli AC, Schuster MG, Vazquez JA, Walsh TJ, Zaoutis TE, Sobel JD (2016) Clinical practice guideline for the management of candidiasis: 2016 update by the infectious diseases society of America. Clin Infect Dis 62(4):e1-50. [https://](https://doi.org/10.1093/cid/civ933) doi.org/10.1093/cid/civ933
- Pathakumari B, Liang G, Liu W (2020) Immune defence to invasive fungal infections: a comprehensive review. Biomed Pharmacother 130:110550.<https://doi.org/10.1016/j.biopha.2020.110550>
- Paul S, Roy D, Pati S, Sa G (2021) The adroitness of andrographolide as a natural weapon against colorectal cancer. Front Pharmacol 12:731492.<https://doi.org/10.3389/fphar.2021.731492>
- Pei XD, He ZL, Yao HL, Xiao JS, Li L, Gu JZ, Shi PZ, Wang JH, Jiang LH (2021) 6-Shogaol from ginger shows anti-tumor efect in cervical carcinoma via PI3K/Akt/mTOR pathway. Eur J Nutr 60(5):2781–2793.<https://doi.org/10.1007/s00394-020-02440-9>
- Peng Y, Wang Y, Tang N, Sun D, Lan Y, Yu Z, Zhao X, Feng L, Zhang B, Jin L, Yu F, Ma X, Lv C (2018) Andrographolide inhibits breast cancer through suppressing COX-2 expression and angiogenesis via inactivation of p300 signaling and VEGF pathway. J Exp Clin Cancer Res 37(1):248. [https://doi.org/10.](https://doi.org/10.1186/s13046-018-0926-9) [1186/s13046-018-0926-9](https://doi.org/10.1186/s13046-018-0926-9)
- Perfect JR, Dismukes WE, Dromer F, Goldman DL, Graybill JR, Hamill RJ, Harrison TS, Larsen RA, Lortholary O, Nguyen MH, Pappas PG, Powderly WG, Singh N, Sobel JD, Sorrell TC (2010) Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the infectious diseases society of America. Clin Infect Dis 50(3):291–322. [https://doi.org/10.](https://doi.org/10.1086/649858) [1086/649858](https://doi.org/10.1086/649858)
- Pinto E, Goncalves MJ, Cavaleiro C, Salgueiro L (2017) Antifungal activity of Thapsia villosa essential oil against Candida, Cryptococcus, Malassezia. Aspergillus and Dermatophyte Species Molecules 22(10):1595. <https://doi.org/10.3390/molecules22101595>
- Pivari F, Mingione A, Brasacchio C, Soldati L (2019) Curcumin and type 2 diabetes mellitus: prevention and treatment. Nutrients 11(8):1837.<https://doi.org/10.3390/nu11081837>
- Plant RM, Dinh L, Argo S, Shah M (2019) The essentials of essential oils. Adv Pediatr 66:111–122. [https://doi.org/10.1016/j.yapd.](https://doi.org/10.1016/j.yapd.2019.03.005) [2019.03.005](https://doi.org/10.1016/j.yapd.2019.03.005)
- Polo MP, de Bravo MG (2006) Effect of geraniol on fatty-acid and mevalonate metabolism in the human hepatoma cell line Hep G2. Biochem Cell Biol 84(1):102–111. [https://doi.org/10.1139/](https://doi.org/10.1139/o05-160) [o05-160](https://doi.org/10.1139/o05-160)
- Qu SY, Li XY, Heng X, Qi YY, Ge PY, Ni SJ, Yao ZY, Guo R, Yang NY, Cao Y, Zhang QC, Zhu HX (2021) Analysis of antidepressant activity of Huang-lian Jie-du decoction through network pharmacology and metabolomics. Front Pharmacol 12:619288. <https://doi.org/10.3389/fphar.2021.619288>
- Quan H, Cao YY, Xu Z, Zhao JX, Gao PH, Qin XF, Jiang YY (2006) Potent in vitro synergism of fuconazole and berberine chloride against clinical isolates of Candida albicans resistant to fuconazole. Antimicrob Agents Chemother 50(3):1096–1099. [https://](https://doi.org/10.1128/AAC.50.3.1096-1099.2006) doi.org/10.1128/AAC.50.3.1096-1099.2006
- Quezada-Fernandez P, Trujillo-Quiros J, Pascoe-Gonzalez S, Trujillo-Rangel WA, Cardona-Muller D, Ramos-Becerra CG, Barocio-Pantoja M, Rodriguez-de la Cerda M, Nerida Sanchez-Rodriguez E, Cardona-Munoz EG, Garcia-Benavides L, Grover-Paez F (2019) Efect of green tea extract on arterial stifness, lipid profle and sRAGE in patients with type 2 diabetes mellitus: a

randomised, double-blind, placebo-controlled trial. Int J Food Sci Nutr 70(8):977–985. [https://doi.org/10.1080/09637486.2019.](https://doi.org/10.1080/09637486.2019.1589430) [1589430](https://doi.org/10.1080/09637486.2019.1589430)

- Razonable RR, Henault M, Lee LN, Laethem C, Johnston PA, Watson HL, Paya CV (2005) Secretion of proinfammatory cytokines and chemokines during amphotericin B exposure is mediated by coactivation of toll-like receptors 1 and 2. Antimicrob Agents Chemother 49(4):1617–1621. [https://doi.org/10.1128/AAC.49.4.](https://doi.org/10.1128/AAC.49.4.1617-1621.2005) [1617-1621.2005](https://doi.org/10.1128/AAC.49.4.1617-1621.2005)
- Ren S, Cai Y, Hu S, Liu J, Zhao Y, Ding M, Chen X, Zhan L, Zhou X, Wang X (2021) Berberine exerts anti-tumor activity in diffuse large B-cell lymphoma by modulating c-myc/CD47 axis. Biochem Pharmacol 188:114576 [https://doi.org/10.1016/j.bcp.](https://doi.org/10.1016/j.bcp.2021.114576) [2021.114576](https://doi.org/10.1016/j.bcp.2021.114576)
- Rhimi W, Aneke CI, Annoscia G, Otranto D, Boekhout T, Cafarchia C (2020) Efect of chlorogenic and gallic acids combined with azoles on antifungal susceptibility and virulence of multidrugresistant Candida spp. and Malassezia furfur isolates. Med Mycol 58(8):1091–1101. <https://doi.org/10.1093/mmy/myaa010>
- Ricci C, Rizzello F, Valerii MC, Spisni E, Gionchetti P, Turroni S, Candela M, D'Amico F, Spigarelli R, Bellocchio I, Marasco G, Barbara G (2022) Geraniol treatment for irritable bowel syndrome: a double-blind randomized clinical trial. Nutrients 14(19):4208. <https://doi.org/10.3390/nu14194208>
- Rocha OB, do Carmo Silva L, de Carvalho Júnior MAB, de Oliveira AA, de Almeida Soares CM, Pereira M (2021) In vitro and in silico analysis reveals antifungal activity and potential targets of curcumin on Paracoccidioides spp. Braz J Microbiol 52(4):1897– 1911.<https://doi.org/10.1007/s42770-021-00548-6>
- Romoli JCZ, Silva MV, Pante GC, Hoeltgebaum D, Castro JC, Oliveira da Rocha GH, Capoci IRG, Nerilo SB, Mossini SAG, Micotti da Gloria E, Svidzinski TIE, Graton Mikcha JM, Machinski MJ (2022) Anti-mycotoxigenic and antifungal activity of ginger, turmeric, thyme and rosemary essential oils in deoxynivalenol (DON) and zearalenone (ZEA) producing Fusarium graminearum. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 39(2):362–372. [https://doi.org/10.1080/19440049.2021.](https://doi.org/10.1080/19440049.2021.1996636) [1996636](https://doi.org/10.1080/19440049.2021.1996636)
- Rudolf JD, Alsup TA, Xu B, Li Z (2021) Bacterial terpenome. Nat Prod Rep 38(5):905–980.<https://doi.org/10.1039/d0np00066c>
- Rufno AT, Ribeiro M, Sousa C, Judas F, Salgueiro L, Cavaleiro C, Mendes AF (2015) Evaluation of the anti-infammatory, anticatabolic and pro-anabolic efects of E-caryophyllene, myrcene and limonene in a cell model of osteoarthritis. Eur J Pharmacol 750:141–150.<https://doi.org/10.1016/j.ejphar.2015.01.018>
- Samoylenko V, Ashfaq MK, Jacob MR, Tekwani BL, Khan SI, Manly SP, Joshi VC, Walker LA, Muhammad I (2009) Indolizidine, antiinfective and antiparasitic compounds from Prosopis glandulosa var. glandulosa. J Nat Prod 72(1):92–8. [https://doi.org/](https://doi.org/10.1021/np800653z) [10.1021/np800653z](https://doi.org/10.1021/np800653z)
- Sasaki K, Ferdousi F, Fukumitsu S, Kuwata H, Isoda H (2021) Antidepressant- and anxiolytic-like activities of Rosmarinus officinalis extract in rodent models: involvement of oxytocinergic system. Biomed Pharmacother 144:112291. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biopha.2021.112291) [biopha.2021.112291](https://doi.org/10.1016/j.biopha.2021.112291)
- Saunte DML, Hare RK, Jorgensen KM, Jorgensen R, Deleuran M, Zachariae CO, Thomsen SF, Bjornskov-Halkier L, Kofoed K, Arendrup MC (2019) Emerging terbinafne resistance in Trichophyton: clinical characteristics, squalene epoxidase gene mutations, and a reliable EUCAST method for detection. Antimicrob Agents Chemother 63(10):e01126-e1219. [https://doi.org/10.](https://doi.org/10.1128/AAC.01126-19) [1128/AAC.01126-19](https://doi.org/10.1128/AAC.01126-19)
- Scribano CM, Wan J, Esbona K, Tucker JB, Lasek A, Zhou AS, Zasadil LM, Molini R, Fitzgerald J, Lager AM, Laffin JJ, Correia-Staudt K, Wisinski KB, Tevaarwerk AJ, O'Regan R, McGregor SM, Fowler AM, Chappell RJ, Bugni TS, Burkard ME, Weaver BA

(2021) Chromosomal instability sensitizes patient breast tumors to multipolar divisions induced by paclitaxel. Sci Transl Med 13(610):eabd4811.<https://doi.org/10.1126/scitranslmed.abd4811>

- Scuteri D, Rombola L, Morrone LA, Bagetta G, Sakurada S, Sakurada T, Tonin P, Corasaniti MT (2019) Neuropharmacology of the neuropsychiatric symptoms of dementia and role of pain: essential oil of bergamot as a novel therapeutic approach. Int J Mol Sci 20(13):3327.<https://doi.org/10.3390/ijms20133327>
- Seleem D, Benso B, Noguti J, Pardi V, Murata RM (2016) In vitro and in vivo antifungal activity of Lichochalcone-A against Candida albicans bioflms. PLoS One 11(6):e0157188. [https://doi.org/10.](https://doi.org/10.1371/journal.pone.0157188) [1371/journal.pone.0157188](https://doi.org/10.1371/journal.pone.0157188)
- Serif I, Tzima E, Bardouki H, Lampri E, Papamarcaki T (2019) Efects of the essential oil from Pistacia lentiscus Var. chia on the lateral line system and the gene expression profle of zebrafsh (Danio rerio). Molecules 24(21):3919. [https://doi.org/10.3390/molec](https://doi.org/10.3390/molecules24213919) [ules24213919](https://doi.org/10.3390/molecules24213919)
- Shang XF, Morris-Natschke SL, Liu YQ, Guo X, Xu XS, Goto M, Li JC, Yang GZ, Lee KH (2018) Biologically active quinoline and quinazoline alkaloids part I. Med Res Rev 38(3):775–828. <https://doi.org/10.1002/med.21466>
- Shen CY, Jiang JG, Zhu W, Ou-Yang Q (2017) Anti-infammatory efect of essential oil from Citrus aurantium L. var. amara Engl. J Agric Food Chem 65(39):8586–8594. [https://doi.org/](https://doi.org/10.1021/acs.jafc.7b02586) [10.1021/acs.jafc.7b02586](https://doi.org/10.1021/acs.jafc.7b02586)
- Shin S (2003) Anti-Aspergillus activities of plant essential oils and their combination efects with ketoconazole or amphotericin B. Arch Pharm Res 26(5):389–393. [https://doi.org/10.1007/](https://doi.org/10.1007/bf02976696) [bf02976696](https://doi.org/10.1007/bf02976696)
- Silva GL, Luft C, Lunardelli A, Amaral RH, Melo DA, Donadio MV, Nunes FB, de Azambuja MS, Santana JC, Moraes CM, Mello RO, Cassel E, Pereira MA, de Oliveira JR (2015) Antioxidant, analgesic and anti-infammatory efects of lavender essential oil. An Acad Bras Cienc 87(2 Suppl):1397–1408. [https://doi.](https://doi.org/10.1590/0001-3765201520150056) [org/10.1590/0001-3765201520150056](https://doi.org/10.1590/0001-3765201520150056)
- Simitsopoulou M, Roilides E, Georgiadou E, Paliogianni F, Walsh TJ (2011) Diferential transcriptional profles induced by amphotericin B formulations on human monocytes during response to hyphae of Aspergillus fumigatus. Med Mycol 49(2):176–85. <https://doi.org/10.3109/13693786.2010.510539>
- Simitsopoulou M, Chlichlia K, Kyrpitzi D, Walsh TJ, Roilides E (2018) Pharmacodynamic and immunomodulatory efects of micafungin on host responses against bioflms of Candida parapsilosis in comparison to those of Candida albicans. Antimicrob Agents Chemother 62(8):e00478-e518. [https://doi.org/10.](https://doi.org/10.1128/aac.00478-18) [1128/aac.00478-18](https://doi.org/10.1128/aac.00478-18)
- Singh S, Chandra U, Anchan VN, Verma P, Tilak R (2020) Limited efectiveness of four oral antifungal drugs (fuconazole, griseofulvin, itraconazole and terbinafne) in the current epidemic of altered dermatophytosis in India: results of a randomized pragmatic trial. Br J Dermatol 183(5):840–846. [https://doi.org/](https://doi.org/10.1111/bjd.19146) [10.1111/bjd.19146](https://doi.org/10.1111/bjd.19146)
- Sliva D, Loganathan J, Jiang J, Jedinak A, Lamb JG, Terry C, Baldridge LA, Adamec J, Sandusky GE, Dudhgaonkar S (2012) Mushroom Ganoderma lucidum prevents colitis-associated carcinogenesis in mice. PLoS One 7(10):e47873. [https://](https://doi.org/10.1371/journal.pone.0047873) doi.org/10.1371/journal.pone.0047873
- Spyridopoulou K, Tiptiri-Kourpeti A, Lampri E, Fitsiou E, Vasileiadis S, Vamvakias M, Bardouki H, Goussia A, Malamou-Mitsi V, Panayiotidis MI, Galanis A, Pappa A, Chlichlia K (2017) Dietary mastic oil extracted from Pistacia lentiscus var. chia suppresses tumor growth in experimental colon cancer models. Sci Rep 7(1):3782. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-017-03971-8) [s41598-017-03971-8](https://doi.org/10.1038/s41598-017-03971-8)
- Sun J (2007) D-Limonene: safety and clinical applications. Altern Med Rev 12(3):259–264
- Sun Z, Wang H, Wang J, Zhou L, Yang P (2014) Chemical composition and anti-infammatory, cytotoxic and antioxidant activities of essential oil from leaves of Mentha piperita grown in China. PLoS One 9(12):e114767. [https://doi.org/10.1371/journal.pone.](https://doi.org/10.1371/journal.pone.0114767) [0114767](https://doi.org/10.1371/journal.pone.0114767)
- Sun P, Zhao L, Zhang N, Wang C, Wu W, Mehmood A, Zhang L, Ji B, Zhou F (2020) Essential oil and juice from bergamot and sweet orange improve acne vulgaris caused by excessive androgen secretion. Mediators Infamm 2020:8868107. [https://doi.org/10.](https://doi.org/10.1155/2020/8868107) [1155/2020/8868107](https://doi.org/10.1155/2020/8868107)
- Takahara M, Takaki A, Hiraoka S, Adachi T, Shimomura Y, Matsushita H, Nguyen TTT, Koike K, Ikeda A, Takashima S, Yamasaki Y, Inokuchi T, Kinugasa H, Sugihara Y, Harada K, Eikawa S, Morita H, Udono H, Okada H (2019) Berberine improved experimental chronic colitis by regulating interferon-gammaand IL-17A-producing lamina propria CD4(+) T cells through AMPK activation. Sci Rep 9(1):11934. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-019-48331-w) [s41598-019-48331-w](https://doi.org/10.1038/s41598-019-48331-w)
- Tang J, Diao P, Shu X, Li L, Xiong L (2019) Quercetin and quercitrin attenuates the infammatory response and oxidative stress in LPS-induced RAW264.7 cells: in vitro assessment and a theoretical model. Biomed Res Int 2019:7039802. [https://doi.](https://doi.org/10.1155/2019/7039802) [org/10.1155/2019/7039802](https://doi.org/10.1155/2019/7039802)
- Temba BA, Fletcher MT, Fox GP, Harvey J, Okoth SA, Sultanbawa Y (2019) Curcumin-based photosensitization inactivates Aspergillus favus and reduces afatoxin B1 in maize kernels. Food Microbiol 82:82–88.<https://doi.org/10.1016/j.fm.2018.12.013>
- Teodoro GR, Gontijo AVL, Salvador MJ, Tanaka MH, Brighenti FL, Delbem ACB, Delbem ÁCB, Koga-Ito CY (2018) Efects of acetone fraction from buchenavia tomentosa aqueous extract and gallic acid on Candida albicans bioflms and virulence factors. Front Microbiol 9:647. [https://doi.org/10.3389/fmicb.](https://doi.org/10.3389/fmicb.2018.00647) [2018.00647](https://doi.org/10.3389/fmicb.2018.00647)
- Thakre A, Zore G, Kodgire S, Kazi R, Mulange S, Patil R, Shelar A, Santhakumari B, Kulkarni M, Kharat K, Karuppayil SM (2018) Limonene inhibits Candida albicans growth by inducing apoptosis. Med Mycol 56(5):565–578. [https://doi.org/10.](https://doi.org/10.1093/mmy/myx074) [1093/mmy/myx074](https://doi.org/10.1093/mmy/myx074)
- Thapa RK, Han SD, Park HG, Son M, Jun JH, Kim JO (2015) DA 5505: a novel topical formulation of terbinafne that enhances skin penetration and retention. Chem Pharm Bull (tokyo) 63(7):525–530. <https://doi.org/10.1248/cpb.c15-00108>
- Thawabteh A, Juma S, Bader M, Karaman D, Scrano L, Bufo SA, Karaman R (2019) The biological activity of natural alkaloids against herbivores, cancerous cells and pathogens. Toxins (basel) 11(11):656. <https://doi.org/10.3390/toxins11110656>
- Tian Y, Wang L, Fan X, Zhang H, Dong Z, Tao T (2023) β-patchoulene alleviates cognitive dysfunction in a mouse model of sepsis associated encephalopathy by inhibition of microglia activation through Sirt1/Nrf2 signaling pathway. PLoS One 18(1):e0279964. [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.pone.0279964) [pone.0279964](https://doi.org/10.1371/journal.pone.0279964)
- Trindade LA, Cordeiro LV, de Figuerêdo Silva D, Figueiredo PTR, de Pontes MLC, de Oliveira Lima E, de Albuquerque Tavares Carvalho A (2022) The antifungal and antibioflm activity of Cymbopogon nardus essential oil and citronellal on clinical strains of Candida albicans. Braz J Microbiol 53(3):1231–1240. [https://doi.](https://doi.org/10.1007/s42770-022-00740-2) [org/10.1007/s42770-022-00740-2](https://doi.org/10.1007/s42770-022-00740-2)
- Tungmunnithum D, Thongboonyou A, Pholboon A, Yangsabai A (2018) Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: an overview. Medicines (Basel) 5(3):93. [https://doi.org/10.3390/medicines5](https://doi.org/10.3390/medicines5030093) [030093](https://doi.org/10.3390/medicines5030093)
- Vagedes J, Henes J, Deckers B, Vagedes K, Kuderer S, Helmert E, von Schoen-Angerer T (2022) Topical Rosmarinus officinalis L. in systemic sclerosis-related Raynaud's phenomenon: an open-label

pilot study. Complement Med Res 29(3):242–248. [https://doi.](https://doi.org/10.1159/000522507) [org/10.1159/000522507](https://doi.org/10.1159/000522507)

- Valsaraj R, Pushpangadan P, Smitt UW, Adsersen A, Christensen SB, Sittie A, Nyman U, Nielsen C, Olsen CE (1997) New anti-HIV-1, antimalarial, and antifungal compounds from Terminalia bellerica. J Nat Prod 60(7):739–742. [https://doi.org/10.1021/np970](https://doi.org/10.1021/np970010m) [010m](https://doi.org/10.1021/np970010m)
- Verma H, Shivavedi N, Tej G, Kumar M, Nayak PK (2022) Prophylactic administration of rosmarinic acid ameliorates depressionassociated cardiac abnormalities in Wistar rats: evidence of serotonergic, oxidative, and infammatory pathways. J Biochem Mol Toxicol 36(10):e23160.<https://doi.org/10.1002/jbt.23160>
- Vică ML, Glevitzky M, Dumitrel GA, Bostan R, Matei HV, Kartalska Y, Popa M (2022) Qualitative characterization and antifungal activity of Romanian honey and propolis. Antibiotics (basel) 11(11):1552.<https://doi.org/10.3390/antibiotics11111552>
- von Lilienfeld-Toal M, Wagener J, Einsele H, Cornely OA, Kurzai O (2019) Invasive Fungal Infection. Dtsch Arztebl Int 116(16):271– 278. <https://doi.org/10.3238/arztebl.2019.0271>
- Wächter GA, Hofmann JJ, Furbacher T, Blake ME, Timmermann BN (1999) Antibacterial and antifungal favanones from Eysenhardtia texana. Phytochemistry 52(8):1469–1471. [https://doi.org/10.](https://doi.org/10.1016/s0031-9422(99)00221-6) [1016/s0031-9422\(99\)00221-6](https://doi.org/10.1016/s0031-9422(99)00221-6)
- Wang T, Shi G, Shao J, Wu D, Yan Y, Zhang M, Cui Y, Wang C (2015) In vitro antifungal activity of baicalin against Candida albicans biofilms via apoptotic induction. Microb Pathog 87:21–29. <https://doi.org/10.1016/j.micpath.2015.07.006>
- Wang X, Zhao S, Su M, Sun L, Zhang S, Wang D, Liu Z, Yuan Y, Liu Y, Li Y (2016) Geraniol improves endothelial function by inhibiting NOX-2 derived oxidative stress in high fat diet fed mice. Biochem Biophys Res Commun 474(1):182–187. [https://doi.org/](https://doi.org/10.1016/j.bbrc.2016.04.097) [10.1016/j.bbrc.2016.04.097](https://doi.org/10.1016/j.bbrc.2016.04.097)
- Wang W, Zhang H, Wang X, Patterson J, Winter P, Graham K, Ghosh S, Lee JC, Katsetos CD, Mackey JR, Tuszynski JA, Wong GK, Luduena RF (2017) Novel mutations involving betaI-, betaIIA-, or betaIVB-tubulin isotypes with functional resemblance to betaIII-tubulin in breast cancer. Protoplasma 254(3):1163–1173. <https://doi.org/10.1007/s00709-016-1060-1>
- Wang J, Pan Y, Hu J, Ma Q, Xu Y, Zhang Y, Zhang F, Liu Y (2018) Tea polyphenols induce S phase arrest and apoptosis in gallbladder cancer cells. Braz J Med Biol Res 51(4):e6891. [https://doi.org/](https://doi.org/10.1590/1414-431X20176891) [10.1590/1414-431X20176891](https://doi.org/10.1590/1414-431X20176891)
- Wang C, Liu X, Lian C, Ke J, Liu J (2019a) Triterpenes and aromatic meroterpenoids with antioxidant activity and neuroprotective effects from Ganoderma lucidum. Molecules 24(23):4353. <https://doi.org/10.3390/molecules24234353>
- Wang L, Wang C, Tao Z, Zhao L, Zhu Z, Wu W, He Y, Chen H, Zheng B, Huang X, Yu Y, Yang L, Liang G, Cui R, Chen T (2019b) Curcumin derivative WZ35 inhibits tumor cell growth via ROS-YAP-JNK signaling pathway in breast cancer. J Exp Clin Cancer Res 38(1):460.<https://doi.org/10.1186/s13046-019-1424-4>
- Wang Y, Shan Y, Wang Y, Fang Y, Huang T, Wang S, Zhu Q, Li X, Ge RS (2019) Aconitine inhibits androgen synthesis enzymes by rat immature Leydig cells via down-regulating androgen synthetic enzyme expression in vitro. Chem Biol Interact 312:108817. <https://doi.org/10.1016/j.cbi.2019.108817>
- Wang F, Chen L, Chen S, Chen H, Liu Y (2021a) Microbial biotransformation of Pericarpium Citri Reticulatae (PCR) by Aspergillus niger and effects on antioxidant activity. Food Sci Nutr 9(2):855– 865. <https://doi.org/10.1002/fsn3.2049>
- Wang G, Ji C, Wang C, Liu Z, Qu A, Wang H (2021b) Matrine ameliorates the infammatory response and lipid metabolism in vascular smooth muscle cells through the NF-kappaB pathway. Exp Ther Med 22(5):1309. <https://doi.org/10.3892/etm.2021.10744>
- Wang K, Chen Q, Shao Y, Yin S, Liu C, Liu Y, Wang R, Wang T, Qiu Y, Yu H (2021) Anticancer activities of TCM and their active

components against tumor metastasis. Biomed Pharmacother 133:111044. <https://doi.org/10.1016/j.biopha.2020.111044>

- Wang L, Jiang Y, Yaseen A, Li F, Chen B, Shen XF, Zheng C, Zhang GL, Wang MK (2021) Steroidal alkaloids from the bulbs of Fritillaria pallidifora Schrenk and their anti-infammatory activity. Bioorg Chem 112:104845. [https://doi.org/10.1016/j.bioorg.2021.](https://doi.org/10.1016/j.bioorg.2021.104845) [104845](https://doi.org/10.1016/j.bioorg.2021.104845)
- Weaver BA (2014) How taxol/paclitaxel kills cancer cells. Mol Biol Cell 25(18):2677–2681. [https://doi.org/10.1091/mbc.](https://doi.org/10.1091/mbc.E14-04-0916) [E14-04-0916](https://doi.org/10.1091/mbc.E14-04-0916)
- Wen J, Xu B, Sun Y, Lian M, Li Y, Lin Y, Chen D, Diao Y, Almoiliqy M, Wang L (2019) Paeoniforin protects against intestinal ischemia/reperfusion by activating LKB1/AMPK and promoting autophagy. Pharmacol Res 146:104308. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.phrs.2019.104308) [phrs.2019.104308](https://doi.org/10.1016/j.phrs.2019.104308)
- Wen Y, Han C, Liu T, Wang R, Cai W, Yang J, Liang G, Yao L, Shi N, Fu X, Deng L, Sutton R, Windsor JA, Hong J, Phillips AR, Du D, Huang W, Xia Q (2020) Chaiqin chengqi decoction alleviates severity of acute pancreatitis via inhibition of TLR4 and NLRP3 infammasome: identifcation of bioactive ingredients via pharmacological sub-network analysis and experimental validation. Phytomedicine 79:153328. [https://doi.org/](https://doi.org/10.1016/j.phymed.2020.153328) [10.1016/j.phymed.2020.153328](https://doi.org/10.1016/j.phymed.2020.153328)
- Wicha P, Onsa-Ard A, Chaichompoo W, Suksamrarn A, Tocharus C (2020) Vasorelaxant and antihypertensive efects of neferine in rats: an in vitro and in vivo study. Planta Med 86(7):496–504. <https://doi.org/10.1055/a-1123-7852>
- Wu H, Chen S, Yu J, Li Y, Zhang XY, Yang L, Zhang H, Hou Q, Jiang M, Brunicardi FC, Wang C, Wu S (2018) Single-cell transcriptome analyses reveal molecular signals to intrinsic and acquired paclitaxel resistance in esophageal squamous cancer cells. Cancer Lett 420:156–167. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.canlet.2018.01.059) [canlet.2018.01.059](https://doi.org/10.1016/j.canlet.2018.01.059)
- Wu L, Li J, Liu T, Li S, Feng J, Yu Q, Zhang J, Chen J, Zhou Y, Ji J, Chen K, Mao Y, Wang F, Dai W, Fan X, Wu J, Guo C (2019) Quercetin shows anti-tumor efect in hepatocellular carcinoma LM3 cells by abrogating JAK2/STAT3 signaling pathway. Cancer Med 8(10):4806–4820. <https://doi.org/10.1002/cam4.2388>
- Wu J, Wu D, Zhao Y, Si Y, Mei L, Shao J, Wang T, Yan G, Wang C (2020) Sodium new houttuyfonate inhibits Candida albicans bioflm formation by inhibiting the Ras1-cAMP-Efg1 pathway revealed by RNA-seq. Front Microbiol 11:2075. [https://doi.](https://doi.org/10.3389/fmicb.2020.02075) [org/10.3389/fmicb.2020.02075](https://doi.org/10.3389/fmicb.2020.02075)
- Xiao X, Luo F, Fu M, Jiang Y, Liu S, Liu B (2022) Evaluating the therapeutic role of selected active compounds in Plumula Nelumbinis on pulmonary hypertension via network pharmacology and experimental analysis. Front Pharmacol 13:977921. <https://doi.org/10.3389/fphar.2022.977921>
- Xie Y, Zhou X, Zhang J, Yu H, Song Z (2022) Immunomodulatory responses of diferentially polarized macrophages to fungal infections. Int Immunopharmacol 111:109089. [https://doi.org/](https://doi.org/10.1016/j.intimp.2022.109089) [10.1016/j.intimp.2022.109089](https://doi.org/10.1016/j.intimp.2022.109089)
- Xu YY, Wang WW, Huang J, Zhu WG (2020) Ellagic acid induces esophageal squamous cell carcinoma cell apoptosis by modulating SHP-1/STAT3 signaling. Kaohsiung J Med Sci 36(9):699–704. <https://doi.org/10.1002/kjm2.12224>
- Yang S, Fu Y, Wu X, Zhou Z, Xu J, Zeng X, Kuang N, Zeng Y (2014) Baicalin prevents Candida albicans infections via increasing its apoptosis rate. Biochem Biophys Res Commun 451(1):36–41. <https://doi.org/10.1016/j.bbrc.2014.07.040>
- Yang D, Liu X, Liu M, Chi H, Liu J, Han H (2015) Protective efects of quercetin and taraxasterol against $H(2)O(2)$ -induced human umbilical vein endothelial cell injury in vitro. Exp Ther Med 10(4):1253–1260. <https://doi.org/10.3892/etm.2015.2713>
- Yang JX, Maria TC, Zhou B, Xiao FL, Wang M, Mao YJ, Li Y (2020) Quercetin improves immune function in Arbor Acre

broilers through activation of NF-κB signaling pathway. Poult Sci 99(4):2305. <https://doi.org/10.1016/j.psj.2020.03.003>

- Yao Y, Yang X, Shi Z, Ren G (2014) Anti-infammatory activity of saponins from quinoa (Chenopodium quinoa Willd.) seeds in lipopolysaccharide-stimulated RAW 264.7 macrophages cells. J Food Sci 79(5):H1018-23. [https://doi.org/10.1111/](https://doi.org/10.1111/1750-3841.12425) [1750-3841.12425](https://doi.org/10.1111/1750-3841.12425)
- Yao M, Fan X, Yuan B, Takagi N, Liu S, Han X, Ren J, Liu J (2019) Berberine inhibits NLRP3 infammasome pathway in human triple-negative breast cancer MDA-MB-231 cell. BMC Complement Altern Med 19(1):216. [https://doi.org/10.1186/](https://doi.org/10.1186/s12906-019-2615-4) [s12906-019-2615-4](https://doi.org/10.1186/s12906-019-2615-4)
- Yazaki K, Arimura GI, Ohnishi T (2017) 'Hidden' terpenoids in plants: their biosynthesis, localization and ecological roles. Plant Cell Physiol 58(10):1615–1621. [https://doi.org/10.1093/](https://doi.org/10.1093/pcp/pcx123) [pcp/pcx123](https://doi.org/10.1093/pcp/pcx123)
- Yi H, Peng H, Wu X, Xu X, Kuang T, Zhang J, Du L, Fan G (2021) The therapeutic efects and mechanisms of quercetin on metabolic diseases: pharmacological data and clinical evidence. Oxid Med Cell Longev 2021:6678662. [https://doi.org/10.1155/2021/](https://doi.org/10.1155/2021/6678662) [6678662](https://doi.org/10.1155/2021/6678662)
- Yin P, Zhang Z, Li J, Shi Y, Jin N, Zou W, Gao Q, Wang W, Liu F (2019) Ferulic acid inhibits bovine endometrial epithelial cells against LPS-induced infammation via suppressing NK-kappaB and MAPK pathway. Res Vet Sci 126:164–169. [https://doi.org/](https://doi.org/10.1016/j.rvsc.2019.08.018) [10.1016/j.rvsc.2019.08.018](https://doi.org/10.1016/j.rvsc.2019.08.018)
- Yin J, Peng X, Lin J, Zhang Y, Zhang J, Gao H, Tian X, Zhang R, Zhao G (2021) Quercetin ameliorates Aspergillus fumigatus keratitis by inhibiting fungal growth, toll-like receptors and infammatory cytokines. Int Immunopharmacol 93:107435. [https://doi.org/10.](https://doi.org/10.1016/j.intimp.2021.107435) [1016/j.intimp.2021.107435](https://doi.org/10.1016/j.intimp.2021.107435)
- Yu ZC, Cen YX, Wu BH, Wei C, Xiong F, Li DF, Liu TT, Luo MH, Guo LL, Li YX, Wang LS, Wang JY, Yao J (2019) Berberine prevents stress-induced gut infammation and visceral hypersensitivity and reduces intestinal motility in rats. World J Gastroenterol 25(29):3956–3971. <https://doi.org/10.3748/wjg.v25.i29.3956>
- Zhang L, Wei W (2020) Anti-infammatory and immunoregulatory efects of paeoniforin and total glucosides of paeony. Pharmacol Ther 207:107452. [https://doi.org/10.1016/j.pharmthera.2019.](https://doi.org/10.1016/j.pharmthera.2019.107452) [107452](https://doi.org/10.1016/j.pharmthera.2019.107452)
- Zhang G, Nitteranon V, Chan LY, Parkin KL (2013) Glutathione conjugation attenuates biological activities of 6-dehydroshogaol from ginger. Food Chem 140(1–2):1–8. [https://doi.org/10.1016/j.foodc](https://doi.org/10.1016/j.foodchem.2013.02.073) [hem.2013.02.073](https://doi.org/10.1016/j.foodchem.2013.02.073)
- Zhang WK, Tao SS, Li TT, Li YS, Li XJ, Tang HB, Cong RH, Ma FL, Wan CJ (2016a) Nutmeg oil alleviates chronic infammatory pain through inhibition of COX-2 expression and substance P release in vivo. Food Nutr Res 60:30849. [https://doi.org/10.](https://doi.org/10.3402/fnr.v60.30849) [3402/fnr.v60.30849](https://doi.org/10.3402/fnr.v60.30849)
- Zhang Z, Chen X, Chen H, Wang L, Liang J, Luo D, Liu Y, Yang H, Li Y, Xie J, Su Z (2016b) Anti-infammatory activity of betapatchoulene isolated from patchouli oil in mice. Eur J Pharmacol 781:229–238.<https://doi.org/10.1016/j.ejphar.2016.04.028>
- Zhang C, Deng J, Liu D, Tuo X, Xiao L, Lai B, Yao Q, Liu J, Yang H, Wang N (2018) Nuciferine ameliorates hepatic steatosis in highfat diet/streptozocin-induced diabetic mice through a PPARalpha/PPARgamma coactivator-1alpha pathway. Br J Pharmacol 175(22):4218–4228.<https://doi.org/10.1111/bph.14482>
- Zhang J, Zheng Y, Luo Y, Du Y, Zhang X, Fu J (2019) Curcumin inhibits LPS-induced neuroinfammation by promoting microglial M2 polarization via TREM2/ TLR4/ NF-kappaB pathways in BV2 cells. Mol Immunol 116:29–37. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.molimm.2019.09.020) [molimm.2019.09.020](https://doi.org/10.1016/j.molimm.2019.09.020)
- Zhang Q, Wang X, Cao S, Sun Y, He X, Jiang B, Yu Y, Duan J, Qiu F, Kang N (2020) Berberine represses human gastric cancer cell growth in vitro and in vivo by inducing cytostatic autophagy

via inhibition of MAPK/mTOR/p70S6K and Akt signaling pathways. Biomed Pharmacother 128:110245. [https://doi.org/](https://doi.org/10.1016/j.biopha.2020.110245) [10.1016/j.biopha.2020.110245](https://doi.org/10.1016/j.biopha.2020.110245)

- Zhang JH, Yang HZ, Su H, Song J, Bai Y, Deng L, Feng CP, Guo HX, Wang Y, Gao X, Gu Y, Zhen Z, Lu Y (2021a) Berberine and ginsenoside Rb1 ameliorate depression-like behavior in diabetic rats. Am J Chin Med 49(5):1195–1213. [https://doi.org/10.1142/](https://doi.org/10.1142/S0192415X21500579) [S0192415X21500579](https://doi.org/10.1142/S0192415X21500579)
- Zhang Q, Liu F, Zeng M, Mao Y, Song Z (2021b) Drug repurposing strategies in the development of potential antifungal agents. Appl Microbiol Biotechnol 105(13):5259–5279. [https://doi.org/](https://doi.org/10.1007/s00253-021-11407-7) [10.1007/s00253-021-11407-7](https://doi.org/10.1007/s00253-021-11407-7)
- Zhang PP, Zhang F, Zhu K, Zhu JF, Yuan Y, Yang YL, Liu L, Wang M, Li JJ (2022a) Matrine exerted an anti-tumor effect on acute myeloid leukemia via the lncRNA LINC01116/miR-592-mediated JAK/STAT pathway inactivation. Neoplasma 69(1):123–135. https://doi.org/10.4149/neo_210802N1083
- Zhang Q, Liu F, Zeng M, Zhang J, Liu Y, Xin C, Mao Y, Song Z (2022) Antifungal activity of sodium new houttuyfonate against Aspergillus fumigatus in vitro and in vivo. Front Microbiol 13:856272. <https://doi.org/10.3389/fmicb.2022.856272>
- Zhao S, Lo CS, Miyata KN, Ghosh A, Zhao XP, Chenier I, Cailhier JF, Ethier J, Lattouf JB, Filep JG, Ingelfnger JR, Zhang SL, Chan JSD (2021) Overexpression of Nrf2 in renal proximal tubular cells stimulates sodium-glucose cotransporter 2 expression and exacerbates dysglycemia and kidney injury in diabetic mice. Diabetes 70(6):1388–1403. <https://doi.org/10.2337/db20-1126>
- Zhou J, Ma W, Wang X, Liu H, Miao Y, Wang J, Du P, Chen Y, Zhang Y, Liu Z (2019) Matrine suppresses reactive oxygen species (ROS)-mediated MKKs/p38-induced infammation in oxidized low-density lipoprotein (ox-LDL)-stimulated macrophages. Med Sci Monit 25:4130–4136. <https://doi.org/10.12659/msm.917151>
- Zhou N, Li S, Zhang F, Chen C, Li Y (2022) Matrine combined with mammalian target of rapamycin inhibitor enhances anti-tumor efficacy of dendritic cell vaccines in hepatocellular carcinoma. Bioengineered 13(4):9274–9283. [https://doi.org/10.1080/21655](https://doi.org/10.1080/21655979.2022.2037855) [979.2022.2037855](https://doi.org/10.1080/21655979.2022.2037855)
- Zhu RL, Zhi YK, Yi L, Luo JF, Li J, Bai SS, Liu L, Wang PX, Zhou H, Dong Y (2019) Sinomenine regulates CD14/TLR4, JAK2/ STAT3 pathway and calcium signal via α 7nAChR to inhibit infammation in LPS-stimulated macrophages. Immunopharmacol Immunotoxicol 41(1):172–177. [https://doi.org/10.1080/](https://doi.org/10.1080/08923973.2019.1568451) [08923973.2019.1568451](https://doi.org/10.1080/08923973.2019.1568451)
- Zhu J, Qiu J, Chen K, Wang W, Zheng S (2021) Tea polyphenols and Levofloxacin alleviate the lung injury of hepatopulmonary syndrome in common bile duct ligation rats through Endotoxin -TNF signaling. Biomed Pharmacother 137:111263. [https://doi.org/10.](https://doi.org/10.1016/j.biopha.2021.111263) [1016/j.biopha.2021.111263](https://doi.org/10.1016/j.biopha.2021.111263)
- Zore GB, Thakre AD, Jadhav S, Karuppayil SM (2011) Terpenoids inhibit Candida albicans growth by afecting membrane integrity and arrest of cell cycle. Phytomedicine 18(13):1181–1190. <https://doi.org/10.1016/j.phymed.2011.03.008>
- Zuzarte M, Correia PMP, Alves-Silva JM, Goncalves MJ, Cavaleiro C, Cruz T, Salgueiro L (2021) Antifungal and anti-infammatory potential of Bupleurum rigidum subsp. paniculatum (Brot.) H. Wolff essential oil. Antibiotics (Basel) 10(5):592. [https://doi.](https://doi.org/10.3390/antibiotics10050592) [org/10.3390/antibiotics10050592](https://doi.org/10.3390/antibiotics10050592)

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.