MINI-REVIEW

Gut microbiota: a new avenue to reveal pathological mechanisms of constipation

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Abstract

Constipation is very pervasive all over the world. It is a common multifactorial gastrointestinal disease, and its etiology and pathomechanism are not completely clear. Now, increasing evidence shows that intestinal fora is closely related to constipation. Intestinal fora is the largest microbiota in the human body and has powerful metabolic functions. Intestinal fora can produce a variety of metabolites, such as bile acids, short-chain fatty acids, tryptophan metabolites, and methane, which have important efects on intestinal motility and secretion. The host can also monitor the intestinal fora and regulate gut dysbacteriosis in constipation. To explore the relationship between intestinal fora and host, the combination of multiomics technology has become the powerful and efective method. Furthermore, the homeostasis restoration of intestinal fora also provides a new strategy for the treatment of constipation. This review aims to explore the interaction between intestinal fora and host in constipation, which contributes to disclose the pathogenesis of constipation and the development of novel drugs for the treatment of constipation from the perspective of intestinal fora.

Key points

• *This review highlights the regulation of gut microbiota on the intestinal motility and secretion of host.*

• *The current review gives an insight into the role of the host on the recognition and regulation of intestinal ecology under constipation.*

• *The article also introduces some novel methods of current gut microbiota research and gut microbiota-based constipation therapies.*

Keywords Constipation · Gut microbiota · Metabolites · Gut immunity

Introduction

Constipation is common worldwide and a frequent clinical symptom. According to statistics, the global incidence of constipation ranges from 2 to 35% (Andromanakos et al. [2006](#page-9-0); Mugie et al. [2011](#page-11-0)). Constipation can be induced by many factors, including lifestyle, diet, psychosocial factors, colonic propulsive or rectal emptying disorders, and the use

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of some drugs (such as anticholinergics, opioids, antihistamines, antipsychotics) (Vriesman et al. [2020\)](#page-13-0). According to the duration of constipation, it is divided into acute constipation (usually lasting no more than a week) and chronic constipation (usually lasting more than 4 weeks or 3 months) (Camilleri et al. [2017\)](#page-9-1). The main symptoms of constipation are dry and hard feces, difficult defecation and reduced defecation times. Currently, intestinal dysfunction is considered to be the main mechanism of the pathogenesis of constipation, including intestinal fuid transport, intestinal motility, mucus secretion and intestinal nerve conduction disorder (Zhao et al. [2021](#page-13-1)). However, a large number of recent studies showed that gut microbiota imbalance was also an important factor in the occurrence and development of constipation.

Recently, with the growing interest in the gut microbiome, our understanding of gut health and disease has been greatly advanced. The gut microbiota has been implicated in various diseases, such as type 2 diabetes, ulcerative colitis,

and chronic kidney disease (Zhang et al. [2021a](#page-13-2), [b;](#page-13-3) Xiao et al. [2020](#page-13-4); Zou et al. [2020](#page-14-0)). There are numerous bacteria living in the gut whose number is 10 times that of our human cells, and they even control 100 times more genetic information than ours (Fujimura et al. [2010\)](#page-10-0). They play an important role on assisting host metabolism, maintaining the function of intestinal barrier and promoting the development of immune system (Obata and Pachnis [2016;](#page-11-1) Schoeler and Caesar [2019](#page-12-0)). Many studies found that the intestinal fora was disordered in constipation patients and animal models (Fan et al. [2022;](#page-10-1) Liang et al. [2019\)](#page-11-2). It was shown that the benefcial bacteria were reduced and the potential pathogenic bacteria were increased in the gut of constipation (Huang et al. [2022;](#page-10-2) Mancabelli et al. [2017;](#page-11-3) Zhu et al. [2014\)](#page-13-5). These phenomena suggest that the intestinal fora is involved in the occurrence and development of constipation. Although the intestinal fora imbalance is prevalent in constipation, the exact mechanism of how dysbiosis is involved in constipation needs to be fully explored.

This review focuses on clarifying the potential mechanisms by which the gut microbiota is involved in the occurrence and progress of constipation and introduces the research progress and technical replacement. Furthermore, the regulation of host on the disordered intestinal fora in the state of constipation is also introduced. Currently, the modulation of intestinal fora has become a clinical therapy for improving constipation. However, the mechanism of intestinal fora in the process of constipation is not completely clear. We hope that this review can give readers a new understanding on the relationship between intestinal fora and constipation and provide novel ideas for the research and clinical treatment of constipation.

Constipation and imbalance of intestinal flora

Increasing evidence suggests that the shift of intestinal fora is closely associated with constipation (Table [1](#page-2-0)). In the past, with culture-based methods, researchers found that constipation was usually accompanied by the disorder of gut microbiota. Zoppi et al. frstly indicated the imbalance of intestinal fora in constipation by using the culture-based microbiological method (Zoppi et al. [1998](#page-13-6)). Subsequently, Khalif et al. also used this method to conduct a similar study on adult patients. Results showed that the abundance of *Bifdobacterium* and *Lactobacillus* in patients with constipation was lower than that in the control groups, while the levels of *Enterobacteriaceae* (such as *Escherichia coli*), *Staphylococcus aureus*, and fungi increased (Khalif et al. 2005). However, this conventional detection technique underestimates about half of the colonic microbes, which limits our further understanding of gut microbiota in constipation (Quigley [2011](#page-12-1)). There are several reasons: many species are not cultured, strictly anaerobic bacteria die in aerobic conditions, and in vitro culture changes the original structure of the microflora. But, the culture-based microbiological approach made us initially understand the relationship between constipation and gut microbes.

With the maturity of sequencing technology, gut microbiome sequencing technology enables us to have deeper understanding of gut microbiota and it has been widely used to reveal the efect of gut microbiota on constipation. Zhu et al. used 16S rRNA gene pyrosequencing to show that the abundance of phylum *Bacteroidetes* in children with functional constipation (FC) was signifcantly reduced, while the level of several species in the phylum *Firmicutes* was markedly increased (Zhu et al. [2014\)](#page-13-5). Subsequently, Kim et al. also reported that the species of *Bifdobacteria* and *Bacteroides* were notably decreased in the feces of adult patients with FC by real-time quantitative polymerase chain reaction (Kim et al. [2015](#page-11-4)). Conversely, Tian et al. found that *Bifdobacterium* was more abundance in slow transit constipation patients than in healthy controls (Tian et al. [2021](#page-12-2)). Mancabelli et al. employed 16S rRNA sequencing and whole genome sequencing to detect the intestinal microbial composition of FC patients (Mancabelli et al. [2017\)](#page-11-3). Their data showed that short-chain fatty acids-producing bacteria (such as *Bacteroides*, *Roseburia*, and *Coprococcus 3*) were signifcantly reduced in the gut of FC patients, and the genes associated with hydrogen production, methanogenesis, and glycerol degradation had high abundance. In addition to fecal samples, some studies also examined mucosal microflora with mucosal biopsy samples. Durbán et al. found that the levels of *Bacteroides* and *Enterobacteriaceae* were elevated in the intestinal mucosa of constipated patients by 16S rRNA metagenomic analysis (V1-V2) (Durbán et al. [2012\)](#page-10-3). Using this technique, Parthasarathy et al. reported that *Bacteroides* were remarkably enriched in the intestinal mucosa of constipated patients (Parthasarathy et al. [2016](#page-12-3)). Compared with the ancient culture methods, intestinal microbial sequencing can more comprehensively and accurately refect the changes of the original structure and function of gut microbes.

Collectively, gut microbiota in constipated patients is mainly characterized by the relative reduction of benefcial bacteria (such as *Lactobacillus* and *Bifdobacterium*), the relative increase of potential pathogens, and the decrease of species richness (Ohkusa et al. [2019\)](#page-11-5). However, as mentioned above, there are opposite situation that constipation patients have more beneficial bacteria in their guts than healthy controls. For example, Du et al. observed that beneficial bacteria (such as *Bifdobacterium* and *Lactobacillales*) were the predominant bacteria in the gut of Parkinson's patients with constipation (Du et al. [2022\)](#page-10-4). Diferences in the type of constipation may be the main reason for this situation. The environment is crucial to the shaping of the microbiome, and the

FC functional constipation, *IBS-C* irritable bowel syndrome with constipation, *ND* no description, *↑* increased abundance of bacteria, vs healthy control, *↓* decreased abundance of bacteria, vs healthy control

intestinal environment is also diferent for diferent types of constipation, resulting that the intestinal fora may be diferent (Coyte and Rakoff-Nahoum 2019). In addition, the relationship between gut microbiota and constipation may be bidirectional. Prolonged colonic transit during constipation may promote the expansion and colonization of slow-growing species, leading to profound changes in the structure and function of the whole microbiome. On the other hand, external factors (such as diet, drugs, and exercise) can cause the disorder of the intestinal fora and its metabolism, which contributes to intestinal dysfunction and promote the development of constipation.

Microbial signals in intestinal motility and secretion

The normal motility and secretion of the intestine can ensure the normal operation of the intestinal contents, and at the same time, it can maintain the homeostasis of the intestinal environment and provide a suitable habitat for the intestinal fora. In the past, it was believed that the motility and secretory functions of the gut were regulated by the host. For example, the loss of enteric nerve subsets and interstitial cells of Cajal, malfunction of smooth

Fig. 1 Regulation of gut microbiota on host intestinal motility and secretion-related functions. **A** Stimulated by bacterial metabolites, intestinal epithelial cells can secrete water and ions into the intestinal cavity through ion pump, transporter, and exchanger. **B** Bacterial metabolites can induce goblet cells to secrete mucin and form a natural physical barrier in the intestine. In addition, intestinal microfora

muscle, and changes in the immune cells were regarded as the basis of internal motility obstacle (Mazzone and Farrugia [2007\)](#page-11-6). However, a recent series of studies have shown that gut microbial signals can also infuence gut motility and secretion (Bhattarai et al. [2020](#page-9-2); Fukui et al. [2018;](#page-10-6) Vicentini et al. [2021\)](#page-12-4). The efect of gut microbiota on gut motility and secretory function is shown in Fig. [1.](#page-3-0)

Motility

Abnormal intestinal motility is an important factor in the occurrence of constipation. Previous studies showed that the metabolites and cellular constituents can directly stimulate intestinal neurons (**C**) or regulate intestinal movement through intestinal endocrine cells (**D**). 5-HT, 5-hydroxytryptamine; 5-HTR, 5-HT receptor; AhR, aryl hydrocarbon receptor; BA, bile acid; TGR5, Takeda G protein-coupled receptor 5; TLR, Toll-like receptor; TRPA1, transient receptor potential ankyrin subtype 1 protein

intestinal nervous system could strictly control the movement of the intestine. There are abundant receptors in the intestinal cells, which can sense physical and chemical stimuli and transmit excitation to intestinal smooth muscle through enteric nerves (Joshi et al. [2021;](#page-10-7) Steensels and Depoortere [2018](#page-12-5)). It has been found that gut microbiotaderived metabolites such as bile acids, short-chain fatty acids, and tryptophan metabolites, are also able to modulate gut motility by stimulating gut chemoreceptors (Dalziel et al. [2021](#page-10-8)).

Bile acids (BAs) are an important regulator of intestinal function, which are produced by the host and metabolized by the intestinal fora. About 5% of bile salts in bile entered the colon, where they were uncoupled or metabolized into secondary bile acids by gut fora (Appleby and Walters [2014](#page-9-3); Wahlström et al. [2016\)](#page-13-7). BAs could activate G-protein coupled BA receptor of enterochromaffin cells and endogenous primary aferent neurons to stimulate the release of serotonin 5-hydroxytryptamine (5-HT) and calcitonin gene-related peptide, resulting in intestinal peristaltic refex (Begley et al. [2006;](#page-9-4) Bunnett [2014](#page-9-5)). Several studies showed that the inhibitor of the ileal BA transporter (Elobixibat) could increase the levels of colonic BAs and improve the colonic transport (Chedid et al. [2018;](#page-9-6) Nakajima et al. [2018](#page-11-7), [2022](#page-11-8)). Microbial transplantation study in mice revealed that the altered microfora could infuence the gastrointestinal transport by afecting the uncoupling of bile salts (Dey et al. [2015](#page-10-9); Li et al. [2021a](#page-11-9), [b\)](#page-11-10). Of note, the efect of bile acids on intestinal motility requires the participation of intestinal fora, but its mechanism remains to be clarifed.

Tryptamine metabolized from tryptophan by *Clostridium sporogenes* and *Ruminococcus gnavus* could reduce the colonic transport time through 5-HT receptor 4 (5-HTR4) in colonic epithelial cells (Bhattarai et al. [2018](#page-9-7); Williams et al. [2014\)](#page-13-8). Additionally, tryptamine also was a ligand for aryl hydrocarbon receptor (AhR) in intestinal cells (Vyhlídalová et al. [2020\)](#page-13-9). Interestingly, Obata and his colleagues recently concluded that intestinal fora could induce the expression of AhR in intestinal neurons and make them respond to AhR ligands derived from microfora, which was benefcial to their excitation and intestinal peristalsis (Obata et al. [2020](#page-11-11)). Indeed, compared with germ-free mice, mice colonized by *Ruminococcus gnavus* showed the induction of several genes involved in tryptophan metabolism (Hofmann et al. [2016\)](#page-10-10). *Ruminococcus gnavus* was found in about 90% of adults and infants, and their ability to produce tryptamines could afect human health (Qin et al. [2010;](#page-12-6) Sagheddu et al. [2016\)](#page-12-7). Ye et al. explored a new mystery that the transient receptor latent ankyrin A1 (TRPA1) could sense microbial tryptophan metabolites (indole, indoleacetic acid) to stimulate enteroendocrine cells to secrete 5-HT, and then transmit bacterial signals to the gut and vagus nerve (Ye et al. [2021](#page-13-10)). Thus, the perturbation of tryptophan metabolism, which were closely associated with the composition, diversity, and metabolism of gut microbiota, might afect intestinal transit time (Roager et al. [2016](#page-12-8); Vandeputte et al. [2016\)](#page-12-9).

Short-chain fatty acids (SCFAs) produced by intestinal bacteria from carbohydrate fermentation were the energy source of intestinal tissue and the regulator of colon motility (Vonk and Reckman [2017\)](#page-13-11), among them, acetic acid, propionic acid, and butyric acid are the most common short-chain fatty acids in the human body (Li et al. [2021a,](#page-11-9) [b](#page-11-10)). The stimulating efect of SCFAs on intestinal motility is hormonedependent (Martin-Gallausiaux et al. [2021\)](#page-11-12). SCFAs could regulate some gastrointestinal hormones, such as glucagon like peptide-1 (GLP-1) and peptide YY (PYY), which could regulate gastrointestinal motility, were released in response to SCFAs (Gribble and Reimann [2019\)](#page-10-11). Recently, a study showed that depletion of gut microbiota led to a decrease in enteric neurons and induced intestinal hypomotility, whereas the supplementation with SCFAs could promote the recovery of enteric neurons (Vicentini et al. [2021](#page-12-4)). Butyrate was found to restore gut motility in germ-free mice in the presence of 5-HT (Vincent et al. [2018](#page-13-12)). Furthermore, propionic acid (enteral) could slow colonic movement by CO-activating free fatty acid receptors 2 and 3 (FFA2 and FFA3) through PYY and enteric nervous system pathways, respectively (Tough et al. [2018\)](#page-12-10). *Lactobacillus* and *Bifdobacterium* in the intestine were proved to produce γ-aminobutyric acid (GABA), which was a derivative of SCFAs (Cui et al. [2020\)](#page-10-12). GABA receptors were also expressed in intestinal neurons and could modulate the gastrointestinal motility, which was reviewed by Auteri et al. (Auteri et al. [2015](#page-9-8)). These evidences suggest that gastrointestinal hormones and enteric nerves are the pathways of SCFAs regulating intestinal motility.

The gas yielded by intestinal flora also had an important impact on intestinal motility. It was believed that methanogens could slow down intestinal movement and cause constipation (Sahakian et al. [2010\)](#page-12-11). Clinical evidence showed that a signifcant increase in the abundance of methanogens was observed in constipation patients (Ojetti et al. [2017](#page-11-13); Ghoshal et al. [2018](#page-10-13)). Additionally, in patients with irritable bowel syndrome dominated by constipation, antibiotic treatment could reduce methanogenic bacteria in the intestinal microflora, which led to the improvement of pathological symptoms (Low et al. [2010](#page-11-14)). *Methanobrevibacter smithii* is the most common methanogenic bacterium in human intestine (Dridi et al. [2009](#page-10-14)). A clinical study indicated that *Methanobrevibacter smithii* was overgrown in the intestines of patients with constipation and accompanied by elevated levels of methane (Takakura et al. [2022\)](#page-12-12). Furthermore, symbiotic microorganisms could produce NO with nitrate or nitrite in intestinal lumen as substrate (Koch et al. [2017](#page-11-15)). NO was proved to be an inhibitory neurotransmitter, which was essential for the relaxation of gastrointestinal smooth muscle and intestinal motility (Groneberg et al. [2016](#page-10-15)).

Toll-like receptors (TLRs), a kind of pattern recognition receptors, which can recognize intestinal bacteria to initiate intracellular signaling afecting the gastrointestinal motility. For instance, TLR2 and TLR4 were located on the membrane of intestinal nerve cells, muscle and glial cells to regulate movement. TLR4 could recognize bacterial lipopolysaccharide (LPS), while TLR2 could recognize lipopeptides and peptidoglycans (Akira et al. [2006](#page-9-9)). In addition, bacterial microbubbles could stimulate TLR2 (Al-Nedawi et al. [2015](#page-9-10)). Reduced fecal production and longer defecation time were found in mice with TLR4 defciency or impaired LPS response, which confrmed that bacterial components could afect the intestinal motility (Anitha et al. [2012](#page-9-11); Caputi et al. [2017\)](#page-9-12). Recently, Yarandi et al. indicated that intestinal bacteria maintained the gastrointestinal motility via TLR2 induced intestinal neurogenesis in mice (Yarandi et al. [2020](#page-13-13)). Their results illustrated that the number of colonic myenteric neurons were markedly increased after the administration of TLR2 agonists. Emerging evidence suggested that resident intestinal fora was essential for regulating intestinal neurons and intestinal endocrine cell populations, as well as neurogenic colonic activity. The gut resident microbe *Bacteroides thetaiotaomicron* could normalize the changes in the expression of nitric oxide synthase and choline acetyltransferase in the myenteric plexus of germ-free mice (Aktar et al. [2020](#page-9-13)).

In conclusion, intestinal microbial signals can transmit information to the host to affect the gastrointestinal motility. These signals may be microbial metabolites or cellular constituents. Therefore, the microbial signal disorder caused by the imbalance of intestinal fora in the process of constipation is an important factor for the slowdown of the gastrointestinal transport. Although the dynamic balance of intestinal microfora is important for the healthy colonic transport, there is no clearly defned microfora for the optimal colonic transport. So, it is necessary to further explore the microflora under the optimal colonic transport.

Secretion

Intestinal secretion includes water, ions, and mucin, which play an important role in the smooth operation of feces in the intestinal lumen. Gut flora are also able to influence intestinal secretion through their metabolites. BAs not only afect intestinal motility and also stimulate the secretion in the colon (Keely et al. [2022](#page-11-16)). It was reported that BAs could promote chloride secretion and inhibit sodium absorption in colonic epithelial cells by regulating colonic ion pump, exchanger, and transporter to induce lumen fuid accumulation (Keely and Walters [2016](#page-11-17)). Brianna et al. found that tryptamine caused the change of short-circuit current by using Ussing chamber with the proximal mouse colon, which confrmed that tryptamine could afect the ion secretion of colonic epithelial cells (Williams et al. [2014](#page-13-8)). Additionally, tryptamine was found to act on 5-HTR4 to increase cyclic adenosine monophosphate (cAMP) to stimulate colonic anion and fuid secretion (Bhattarai et al. [2018](#page-9-7)). Several studies showed that intestinal hormone secretion was also afected by intestinal fora. Zhuang et al. reported that SCFAs produced by intestinal fora could stimulate the secretion of intestinal peptide YY in constipated rats (Zhuang et al. [2019\)](#page-13-14). Wichmann et al. found that germ-free mice did not produce SCFA, resulting in a signifcant increase in the level of plasma glucagon like peptide-1 (GLP-1), which could inhibit intestinal peristalsis (Wichmann et al. [2013](#page-13-15)). Through fecal microbiota transplantation (FMT) in patients with irritable bowel syndrome (IBS), it was found that FMT changed the density of enteroendocrine cells in intestine in patients with IBS (Mazzawi et al. [2021\)](#page-11-18). Enteroendocrine cells, as the largest hormone secreting population in the gut, expressed a diverse array of G protein coupled receptors as well as Toll- like receptors (Yu et al. [2020\)](#page-13-16). Therefore, enteroendocrine cells might be the direct or indirect target of the gut microbiota infuencing gut hormone secretion.

Mucin in the gut is mainly secreted by intestinal goblet cells, which is the main component of intestinal mucus layer. The intestinal mucus layer can provide the habitat for intestinal fora and lubricate the contents. A large number of studies verifed that intestinal symbiotic bacteria played an important role in promoting intestinal mucus secretion (Hayes et al. [2018](#page-10-16); Sicard et al. [2017](#page-12-13); Bergstrom et al. [2020](#page-9-14)). For example, compared with normal mice, the number of intestinal goblet cells in sterile mice were reduced and the maturation of mucus system was slow (Johansson et al. [2015](#page-10-17)). Intestinal meprin β enzymes could cleave mucin to release mucus, but the cleavage process required the induction of intestinal fora (Schütte et al. [2014\)](#page-12-14). In the intestine, some commensal bacteria such as *Bacteroides thetaiotaomicron* and *Faecalibacterium prausnitzii* could increase the diferentiation of goblet cell and the expression of mucus related genes (Wrzosek et al. [2013\)](#page-13-17). In fact, the outer mucous layer of the colon was inhabited by symbiotic bacteria that could degrade mucin to obtain energy, such as *Bacteroides acidifaciens* (in mice), *Bacteroides fragilis*, *Bifdobacteriaceae*, *Lactobacillus* spp., and *Akkermansia muciniphila* (in mice and humans) (Donaldson et al. [2016\)](#page-10-18). The degradation of glycoprotein of outer mucous layer by symbiotic bacteria can promote the natural replacement of intestinal mucus. Although these phenomena show that intestinal fora can afect the secretion of intestinal mucus, a large number of studies are still needed to clarify its mechanism.

Regulation of dysregulated intestinal fora in patients with constipation

The microenvironment of the intestinal lumen is created by intestinal fora and intestinal cells, so the intestine also affects the microflora. Increasing evidence showed that intestinal innate immunity could shape the microbiota (Kurilshikov et al. [2017](#page-11-19)). A recent study indicated that there was an infammatory response and fora imbalance in constipated mice, and the transplantation of the dysfunctional fora into the intestines of healthy mice could also induce the intestinal infammation (Lin et al. [2021\)](#page-11-20). Previously, colonic histopathological studies proved that there was the microscopic infammation in the colonic tissue of patients with constipated irritable bowel syndrome, which

was mainly characterized by the increase of mast cells, intraepithelial lymphocytes, and $CD3⁺$ T cells in the colonic mucosa (Barbara et al. [2004](#page-9-15); Chadwick et al. [2002\)](#page-9-16). Intestinal cells could express pattern recognition receptors (PRRs), which were used to sense microbial associated molecular patterns (MAMPs), and then promote immune response to resist pathogens (Potrykus et al. [2021](#page-12-15)). PRRs are divided into five families: Toll-like receptors (TLRs), C-type lectin-like receptors (CLRs), nucleotide binding oligomerization domain (NOD)-like receptors (NLRs), retinoic acidinduced gene-I (RIG-I)-like receptors (RLR), and recently identifed absent in melanoma (AIM)-like receptors (ALR) (Agier et al. [2018;](#page-9-17) Kurilshikov et al. [2017\)](#page-11-19). After PRRs sense microbial antigens, they could regulate the function of other intestinal cells against pathogens through activating intestinal immunity, and restore the intestinal ecological balance (Bonder et al. [2016](#page-9-18); Hoving et al. [2014](#page-10-19); Xiao et al. [2019](#page-13-18)). Therefore, in constipation, the host may activate the immune response through these PRRs to restore the balance of intestinal microbiota.

In the colon, the mucus layer, an important physical barrier, is further divided into two layers: an internal frm layer for isolating microorganisms and an external loose layer that provides the habitat for microorganisms (Zhao and Maynard [2022;](#page-13-19) Li et al. [2015](#page-11-21); Lin et al. [2021](#page-11-20)). Intestinal mucus is mainly composed of mucin. A report suggested that the deletion of mucin 2 (MUC2) gene in mice could alter the intestinal microbiota (Wu et al. [2018](#page-13-20)). The levels of *Desulfovibrio*, *Escherichia*, *Akkermansia*, *Turicibacter*, *Erysipelotrichaceae*, and *Ruminococcaceae* increased and contents of *Lactobacilli* and *Lachnospiraceae* decreased in MUC2 deficient mice. Mucin were often modified by complex polysaccharides, and some bacterial species used glycosidases to degrade mucin to obtain polysaccharides as a carbon source (Crouch et al. [2020\)](#page-10-20). It was found that O-glycans on MUC2 from the proximal colon regulate the structure and function of the microfora (Bergstrom et al. [2020\)](#page-9-14). When constipation occurs, dry and hard feces stay in the colon for a long time, which is easy to take away the external loose mucus layer, creating the opportunity for bacteria to invade the internal mucus layer and cause infammation. Excitingly, it was reported that the mediators produced by the immune system could drive the proliferation of goblet cells and increase mucus secretion (Khan and Collins [2004;](#page-11-22) Oeser et al. [2015](#page-11-23)). The increase of mucus secretion of goblet cells could help to wash away the bacteria in the inner mucus layer (Birchenough et al. [2016](#page-9-19)). However, the replenishment rate of mucus is slower than the consumption rate and the constipation cannot be relieved, which is regarded as the host-autonomous regulation of dysbiosis in constipation.

On the other hand, there are a lot of antibacterial substances (such as immunoglobulin A (IgA) and antimicrobial peptides) in the mucus layer inside the intestine, which play

an important role on maintaining the balance of intestinal fora. IgA produced by intestinal lamina propria plasma cells can maintain the intestinal homeostasis through binding and interacting with mucin and intestinal bacteria in the outer mucus layer (Rogier et al. [2014\)](#page-12-16). IgA not only help the host to remove pathogens but also anchor symbiotic bacteria in mucus. It has proved that symbiotic *Bacteroides fragilis* can specifcally recognize IgA to promote mucosal colonization (Donaldson et al. [2018\)](#page-10-21). Inatomi et al. found that the concentration of IgA was notably increased in the feces of constipated rats after the treatment with probiotics (Inatomi and Honma [2021\)](#page-10-22). IgA was proved to enhance the adhesion of *Escherichia coli*, *Bifdobacterium lactis*, and *Lactobacillus rhamnosus* to epithelial cells, revealing that microbes could also beneft from IgA to establish mucosal microbial communities (Bollinger et al. [2003;](#page-9-20) Mathias et al. [2010](#page-11-24)). Furthermore, antimicrobial peptides produced by Paneth cells also play an important role on improving the intestinal dysbiosis (Suchodolski and Jergens [2016\)](#page-12-17). It was found that intestinal symbiotic bacteria had stronger antimicrobial peptide resistance compare with pathogens, which was also the key for antimicrobial peptides to maintain the homeostasis of host intestinal fora (Cullen et al. [2015\)](#page-10-23). Antimicrobial peptides and IgA in mucus layer are very important to maintain the balance of intestinal microfora, and their expression is regulated by intestinal immune system. Therefore, in the process of constipation, low-grade intestinal infammation is a means for the host to perceive and maintain intestinal eubiosis (Fig. [2\)](#page-7-0).

Constipation therapy and research strategy based on intestinal flora

In recent years, some new constipation therapies, such as prebiotics, synbiotics, inhibitors of ileal bile acid transporters, antibiotic treatment for patients with methanogenesis, and 5-HT4 receptor agonists, have been clinically available (Choi and Chang [2015;](#page-9-21) Ford et al. [2014](#page-10-24); Prichard and Bharucha [2018;](#page-12-18) Triantafyllou et al. [2014\)](#page-12-19). Notably, microbial related agents could not only improve pathological symptoms of constipation but also regulate the host immune system (Yeşilyurt et al. [2021\)](#page-13-21). For example, probiotics, especially *Lactobacillus* and *Bifdobacterium*, could stimulate immune cells (such as Th1, Th2, Th17, T regulatory cells, and B cells) and increase the production of SIgA and antimicrobial substances, which resisted pathogens and toxins in the intestine and maintained the integrity of the intestinal barrier (Dargahi et al. [2019;](#page-10-25) Shi et al. [2017\)](#page-12-20). Nevertheless, studies indicated that probiotics might be a potential risk for the treatment of constipation. Especially in elderly constipated patients with impaired intestinal mucosal barrier or immunosuppressive state, probiotic therapy carried

Fig. 2 Host supervision and regulation of gut microbiota in constipation. Constipation is caused by many factors, such as diet, drugs, lack of exercise, and genetic factors. Constipation is accompanied by the imbalance of intestinal fora and the consumption of intestinal mucus. The mucus layer in the intestine is divided into outer loose layer and inner solid layer. The outer loose layer provides the habitat for symbiotic bacteria. Constipation leads to the depletion of outer loose layer in the intestine, which can cause the run off of symbiotic bacteria and the invasion of pathogens and symbiotic bacteria into inner solid

risks such as microbial translocation, opportunistic pathogen infection, D-lactic acidosis, and loss of bioactivity of antimicrobial or antifungal agents (Camilleri [2019;](#page-9-22) Rao et al. [2018](#page-12-21); Suez et al. [2019](#page-12-22)). Fortunately, evidence indicated that the use of inactivated probiotics or probiotic metabolites could eliminate the risk of probiotic treatment of constipation (De Marco et al. [2018;](#page-10-26) Żółkiewicz et al. [2020](#page-13-22)). Because they are not living microorganisms, bacterial translocation and detrimental metabolic activities do not occur (Sotoudegan et al.

layer. The inner solid layer contains a large number of antibacterial substances (such as IgA and antibacterial peptides), and the entry of bacteria and their antigens into this layer will induce the immune response. The intestinal tract can recognize the pathogens and symbionts invading the internal solid layer through pattern recognition receptors to trigger the immune response, so as to regulate the synthesis of mucin, IgA, and antimicrobial peptides to resist invasion and regulate the imbalance of intestinal fora in the state of constipation

[2019;](#page-12-23) Rossi et al. [2022](#page-12-24)). Additionally, based on the efect of gut microbiota on gut function, fecal microbiota transplantation (FMT) therapy for constipation has also received widespread attention (Liu et al. [2021\)](#page-11-25). Researchers have carried out some clinical trials on FMT for the treatment of constipation, which obtained certain clinical efects (Johnsen et al. [2018](#page-10-27); Zhang et al. [2021a](#page-13-2), [b;](#page-13-3) Kuai et al. [2021\)](#page-11-26). However, the microbial species of FMT are very complex, including bacteria, fungi, viruses, and other components, and it is difficult to determine its risk source, which hinders its clinical application (Blaser [2019](#page-9-23)). Therefore, a large number of studies are still needed to prove the safety of FMT to promote the promotion of FMT in the clinical treatment of constipation.

In Southeast Asia, plant laxatives or herbs including traditional Chinese medicines (TCMs) are usually used to relieve constipation. Anthraquinones (such as senna, aloe, rhubarb, frangula, and cascara) are the most commonly used plant laxatives, which can improve constipation mainly by stimulating fuid secretion and intestinal motility, especially for the short-term treatment of tension constipation, acute constipation, and before lower gastrointestinal endoscopy (Cirillo and Capasso [2015;](#page-10-28) Wang and Yin [2015](#page-13-23)). Interestingly, TCM formulas are proved to treat diseases (including constipation) through restoring the normal composition and function of gut microflora in clinical or animal studies (Tong et al. [2018;](#page-12-25) Xu et al. [2015](#page-13-24)). Zengye decoction (ZYD) composed of Sophorae Flavescentis Radix, Ophiopogonis Radix, and Rehmanniae Radix Praeparata notably decreased the abundance of harmful microbes (such as *Desulfovibrio*, *Prevotella*, and *Ruminococcus*), whereas it markedly increased the abundance of *Oxalobacter*, *Clostridium*, and *Roseburia* in aged rats with constipation (Liu et al. [2019](#page-11-27)). Additionally, the latest fndings in our lab illustrated that rhubarb could markedly improve various pathological symptoms in constipated rats by restoring the homeostasis of intestinal fora and ameliorating the disorder of its metabolism (Yang et al. [2022](#page-13-25)). However, TCM formulas usually exerted their effects by multitargets and multipathways, while their mechanisms related to gut microflora have not been adequately investigated.

During the past two decades, with the implementation of the human genome project and the development of a new generation of sequencing technology, researches on intestinal microorganisms have received increasing attention and achieved fruitful results. In fact, these microbial detection technologies tend to refect diferences in composition and cannot obtain accurate results of the functional changes of gut microfora. However, the combination of multiomics and alternative medical technology complement the study of intestinal microfora. As an example, intestinal microbiota sequencing could identify new functional genes, microbial pathways, and dysfunctions of intestinal microbiome and determine the interaction and coevolution between micro-flora and host (Wang and Yin [2015\)](#page-13-23). However, due to different primers and GC contents, each gene may not be amplifed with the same efficiency during the PCR reaction, resulting in sequencing bias (Wensel et al. [2022](#page-13-26)). Furthermore, taxonomy accuracy relies on the integrity of the reference database, and the quality and quantity of databases determine the accuracy and resolution of taxonomy classifcations (Jo et al. [2016](#page-10-29)). Metabolomics provided important help for us to understand the metabolic diferences of intestinal fora in the disease state (including constipation), identify new metabolic markers, and uncover the functional changes of intestinal flora (Bauermeister et al. [2022](#page-9-24)). Furthermore, environmental transcriptomics and proteomics also provided an important supplement from the gene and protein levels for the study of intestinal microbial function (Bashiardes et al. [2016](#page-9-25); Verberkmoes et al. [2009\)](#page-12-26). Simultaneously, the newly developed intestinal chip and microfuidic technology enable us to have a clearer understanding of the interaction between intestinal fora and host, because these in vitro techniques can help us eliminate some interference factors and can more easily achieve variable control (Puschhof et al. [2021](#page-12-27)). Therefore, we can explore the effect of gut microflora on composition by regulating intestinal motility and secretion function based on integrated technologies and approaches, so as to clarify the intestinal fora related pathogenesis of constipation and develop new therapeutic drugs.

Conclusion

In the process of constipation, the imbalance of intestinal fora has been adequately confirmed. The intestinal flora is in the closed environment of the host intestine and is afected by the external environment and the host. In terms of external factors, constipation inducements such as diet, drugs, living habits, and social pressure will trigger the disorder of intestinal fora, while the damage of intestinal mucus caused by constipation and the disorder of intestinal hormones can also destroy the habitat and environment of intestinal symbiotic bacteria. In addition, intestinal fora can afect intestinal motility and secretion. There are abundant chemoreceptors in the intestine, which can sense the metabolites and cellular components from intestinal bacteria. In the state of constipation, the disorder of intestinal fora and its metabolism can result in the abnormality of gut motility and secretion. At the same time, the host has a regulatory role on the intestinal fora by some pattern recognition receptors. When the fora is disordered, the host activates the intestinal immune system and mucus to resist the pathogen and assist the colonization of probiotics. Therefore, the study of constipation based on intestinal fora can give us a comprehensive understanding of the pathogenesis of constipation and contribute to the development of new therapies. However, due to the structural complexity and functional diversity of intestinal fora, it is an appropriate strategy to uncover the relationship between constipation and intestinal fora through multiomics approaches.

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Declarations

Ethical approval This article does not contain any studies with animals performed by any of the authors.

Conflict of interest The authors declare no competing interests.

References

- Agier J, Pastwińska J, Brzezińska-Błaszczyk E (2018) An overview of mast cell pattern recognition receptors. Infamm Res 67:737–746. <https://doi.org/10.1007/s00011-018-1164-5>
- Akira S, Uematsu S, Takeuchi O (2006) Pathogen recognition and innate immunity. Cell 124:783–801. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cell.2006.02.015) [cell.2006.02.015](https://doi.org/10.1016/j.cell.2006.02.015)
- Aktar R, Parkar N, Stentz R, Baumard L, Parker A, Goldson A, Brion A, Carding S, Blackshaw A, Peiris M (2020) Human resident gut microbe *Bacteroides thetaiotaomicron* regulates colonic neuronal innervation and neurogenic function. Gut Microbes 11:1745–1757. <https://doi.org/10.1080/19490976.2020.1766936>
- Al-Nedawi K, Mian MF, Hossain N, Karimi K, Mao Y, Forsythe P, Min KK, Stanisz AM, Kunze WA, Bienenstock J (2015) Gut commensal microvesicles reproduce parent bacterial signals to host immune and enteric nervous systems. FASEB J 29:684–695. [https://doi.org/10.1096/f.14-259721](https://doi.org/10.1096/fj.14-259721)
- Andromanakos N, Skandalakis P, Troupis T, Filippou D (2006) Constipation of anorectal outlet obstruction: pathophysiology, evaluation and management. J Gastroenterol Hepatol 21:638–646. <https://doi.org/10.1111/j.1440-1746.2006.04333.x>
- Anitha M, Vijay-Kumar M, Sitaraman SV, Gewirtz AT, Srinivasan S (2012) Gut microbial products regulate murine gastrointestinal motility via Toll-like receptor 4 signaling. Gastroenterology 143:1006–1016. <https://doi.org/10.1053/j.gastro.2012.06.034>
- Appleby RN, Walters JRF (2014) The role of bile acids in functional GI disorders. Neurogastroenterol Motil 26:1057–1069. [https://](https://doi.org/10.1111/nmo.12370) doi.org/10.1111/nmo.12370
- Auteri M, Zizzo MG, Serio R (2015) GABA and GABA receptors in the gastrointestinal tract: from motility to infammation. Pharmacol Res 93:11–21.<https://doi.org/10.1016/j.phrs.2014.12.001>
- Barbara G, Stanghellini V, De Giorgio R, Cremon C, Cottrell GS, Santini D, Pasquinelli G, Morselli-Labate AM, Grady EF, Bunnett NW, Collins SM, Corinaldesi R (2004) Activated mast cells in proximity to colonic nerves correlate with abdominal pain in irritable bowel syndrome. Gastroenterology 126:693–702. [https://](https://doi.org/10.1053/j.gastro.2003.11.055) doi.org/10.1053/j.gastro.2003.11.055
- Bashiardes S, Zilberman-Schapira G, Elinav E (2016) Use of metatranscriptomics in microbiome research. Bioinform Biol Insights 10:19–25.<https://doi.org/10.4137/BBI.S34610>
- Bauermeister A, Mannochio-Russo H, Costa-Lotufo LV, Jarmusch AK, Dorrestein PC (2022) Mass spectrometry-based metabolomics in microbiome investigations. Nat Rev Microbiol 20:143–160. <https://doi.org/10.1038/s41579-021-00621-9>
- Begley M, Hill C, Gahan CGM (2006) Bile salt hydrolase activity in probiotics. Appl Environ Microbiol 72:1729–1738. [https://doi.](https://doi.org/10.1128/AEM.72.3.1729-1738.2006) [org/10.1128/AEM.72.3.1729-1738.2006](https://doi.org/10.1128/AEM.72.3.1729-1738.2006)
- Bergstrom K, Shan X, Casero D, Batushansky A, Lagishetty V, Jacobs JP, Hoover C, Kondo Y, Shao B, Gao L, Zandberg W, Noyovitz B, McDaniel JM, Gibson DL, Pakpour S, Kazemian N, McGee S, Houchen CW, Rao CV, Griffin TM, Sonnenburg JL, McEver RP, Braun J, Xia L (2020) Proximal colon-derived O-glycosylated mucus encapsulates and modulates the microbiota. Science 370:467–472.<https://doi.org/10.1126/science.aay7367>
- Bhattarai Y, Williams BB, Battaglioli EJ, Whitaker WR, Till L, Grover M, Linden DR, Akiba Y, Kandimalla KK, Zachos NC, Kaunitz JD, Sonnenburg JL, Fischbach MA, Farrugia G, Kashyap PC (2018) Gut microbiota-produced tryptamine activates an epithelial G-orotein-coupled receptor to increase colonic secretion. Cell Host Microbe 23:775–785. [https://doi.org/10.1016/j.chom.](https://doi.org/10.1016/j.chom.2018.05.004) [2018.05.004](https://doi.org/10.1016/j.chom.2018.05.004)
- Bhattarai Y, Jie S, Linden DR, Ghatak S, Mars RAT, Williams BB, Pu M, Sonnenburg JL, Fischbach MA, Farrugia G, Sha L, Kashyap PC (2020) Bacterially derived tryptamine increases mucus release by activating a host receptor in a mouse model of infammatory bowel disease. iScience 23: 101798. [https://doi.org/10.](https://doi.org/10.1016/j.isci.2020.101798) [1016/j.isci.2020.101798](https://doi.org/10.1016/j.isci.2020.101798)
- Birchenough GMH, Nyström EEL, Johansson MEV, Hansson GC (2016) A sentinel goblet cell guards the colonic crypt by triggering Nlrp6-dependent Muc2 secretion. Science 352:1535–1542. <https://doi.org/10.1126/science.aaf7419>
- Blaser MJ (2019) Fecal microbiota transplantation for dysbiosis - predictable risks. N Engl J Med 381:2064–2066. [https://doi.org/10.](https://doi.org/10.1056/NEJMe1913807) [1056/NEJMe1913807](https://doi.org/10.1056/NEJMe1913807)
- Bollinger RR, Everett ML, Palestrant D, Love SD, Lin SS, Parker W (2003) Human secretory immunoglobulin A may contribute to bioflm formation in the gut. Immunology 109:580–587. [https://](https://doi.org/10.1046/j.1365-2567.2003.01700.x) doi.org/10.1046/j.1365-2567.2003.01700.x
- Bonder MJ, Kurilshikov A, Tigchelaar EF, Mujagic Z, Imhann F, Vila AV, Deelen P, Vatanen T, Schirmer M, Smeekens SP, Zhernakova DV, Jankipersadsing SA, Jaeger M, Oosting M, Cenit MC, Masclee AAM, Swertz MA, Li Y, Kumar V, Joosten L, Harmsen H, Weersma RK, Franke L, Hofker MH, Xavier RJ, Jonkers D, Netea MG, Wijmenga C, Fu J, Zhernakova A (2016) The efect of host genetics on the gut microbiome. Nat Genet 48:1407–1412. <https://doi.org/10.1038/ng.3663>
- Bunnett NW (2014) Neuro-humoral signalling by bile acids and the TGR5 receptor in the gastrointestinal tract. J Physiol 592:2943– 2950.<https://doi.org/10.1113/jphysiol.2014.271155>
- Camilleri M (2019) Leaky gut: mechanisms, measurement and clinical implications in humans. Gut 68:1516–1526. [https://doi.org/10.](https://doi.org/10.1136/gutjnl-2019-318427) [1136/gutjnl-2019-318427](https://doi.org/10.1136/gutjnl-2019-318427)
- Camilleri M, Ford AC, Mawe GM, Dinning PG, Rao SS, Chey WD, Simrén M, Lembo A, Young-Fadok TM, Chang L (2017) Chronic Constipation Nat Rev Dis Primers 3:17095. [https://doi.](https://doi.org/10.1038/nrdp.2017.95) [org/10.1038/nrdp.2017.95](https://doi.org/10.1038/nrdp.2017.95)
- Caputi V, Marsilio I, Cerantola S, Roozfarakh M, Lante I, Galuppini F, Rugge M, Napoli E, Giulivi C, Orso G, Giron MC (2017) Tolllike receptor 4 modulates small intestine neuromuscular function through nitrergic and purinergic pathways. Front Pharmacol 8:350.<https://doi.org/10.3389/fphar.2017.00350>
- Chadwick VS, Chen W, Shu D, Paulus B, Bethwaite P, Tie A, Wilson I (2002) Activation of the mucosal immune system in irritable bowel syndrome. Gastroenterology 122:1778–1783. [https://doi.](https://doi.org/10.1053/gast.2002.33579) [org/10.1053/gast.2002.33579](https://doi.org/10.1053/gast.2002.33579)
- Chedid V, Vijayvargiya P, Camilleri M (2018) Elobixibat for the treatment of constipation. Expert Rev Gastroenterol Hepatol 12:951– 960. <https://doi.org/10.1080/17474124.2018.1522248>
- Choi CH, Chang SK (2015) Alteration of gut microbiota and efficacy of probiotics in functional constipation. J Neurogastroenterol Motil 21:4–7. <https://doi.org/10.5056/jnm14142>
- Cirillo C, Capasso R (2015) Constipation and botanical medicines: an overview. Phytother Res 29:1488–1493. [https://doi.org/10.](https://doi.org/10.1002/ptr.5410) [1002/ptr.5410](https://doi.org/10.1002/ptr.5410)
- Coyte KZ, Rakof-Nahoum S (2019) Understanding competition and cooperation within the mammalian gut microbiome. Curr Biol 29:R538–R544.<https://doi.org/10.1016/j.cub.2019.04.017>
- Crouch LI, Liberato MV, Urbanowicz PA, Baslé A, Lamb CA, Stewart CJ, Cooke K, Doona M, Needham S, Brady RR, Berrington JE, Madunic K, Wuhrer M, Chater P, Pearson JP, Glowacki R, Martens EC, Zhang F, Linhardt RJ, Spencer DIR, Bolam DN (2020) Prominent members of the human gut microbiota express endoacting O-glycanases to initiate mucin breakdown. Nat Commun 11:4017. <https://doi.org/10.1038/s41467-020-17847-5>
- Cui Y, Miao K, Niyaphorn S, Qu X (2020) Production of gammaaminobutyric acid from lactic acid bacteria: a systematic review. Int J Mol Sci 21:995.<https://doi.org/10.3390/ijms21030995>
- Cullen TW, Schofeld WB, Barry NA, Putnam EE, Rundell EA, Trent MS, Degnan PH, Booth CJ, Yu H, Goodman AL (2015) Antimicrobial peptide resistance mediates resilience of prominent gut commensals during infammation. Science 347:170–175. [https://](https://doi.org/10.1126/science.1260580) doi.org/10.1126/science.1260580
- Dalziel JE, Spencer NJ, Young W (2021) Microbial signalling in colonic motility. Int J Biochem Cell Biol 134:105963. [https://](https://doi.org/10.1016/j.biocel.2021.105963) doi.org/10.1016/j.biocel.2021.105963
- Dargahi N, Johnson J, Donkor O, Vasiljevic T, Apostolopoulos V (2019) Immunomodulatory efects of probiotics: can they be used to treat allergies and autoimmune diseases? Maturitas 119:25–38. <https://doi.org/10.1016/j.maturitas.2018.11.002>
- De Marco S, Sichetti M, Muradyan D, Piccioni M, Traina G, Pagiotti R, Pietrella D (2018) Probiotic cell-free supernatants exhibited anti-infammatory and antioxidant activity on human gut epithelial cells and macrophages stimulated with LPS. Evid Based Complement Alternat Med 2018:1756308. [https://doi.org/10.](https://doi.org/10.1155/2018/1756308) [1155/2018/1756308](https://doi.org/10.1155/2018/1756308)
- Dey N, Wagner VE, Blanton LV, Cheng J, Fontana L, Haque R, Ahmed T, Gordon JI (2015) Regulators of gut motility revealed by a gnotobiotic model of diet-microbiome interactions related to travel. Cell 163:95–107.<https://doi.org/10.1016/j.cell.2015.08.059>
- Donaldson GP, Lee SM, Mazmanian SK (2016) Gut biogeography of the bacterial microbiota. Nat Rev Microbiol 14:20–32. [https://](https://doi.org/10.1038/nrmicro3552) doi.org/10.1038/nrmicro3552
- Donaldson GP, Ladinsky MS, Yu KB, Sanders JG, Yoo BB, Chou W, Conner ME, Earl AM, Knight R, Bjorkman PJ, Mazmanian SK (2018) Gut microbiota utilize immunoglobulin A for mucosal colonization. Science 360:795–800. [https://doi.org/10.1126/scien](https://doi.org/10.1126/science.aaq0926) [ce.aaq0926](https://doi.org/10.1126/science.aaq0926)
- Dridi B, Henry M, El Khéchine A, Raoult D, Drancourt M (2009) High prevalence of *Methanobrevibacter smithii* and *Methanosphaera stadtmanae* detected in the human gut using an improved DNA detection protocol. PLoS ONE 4:e7063. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pone.0007063) [journal.pone.0007063](https://doi.org/10.1371/journal.pone.0007063)
- Du Y, Li Y, Xu X, Li R, Zhang M, Cui Y, Zhang L, Wei Z, Wang S, Tuo H (2022) Probiotics for constipation and gut microbiota in Parkinson's disease. Parkinsonism Relat Disord 103:92–97. <https://doi.org/10.1016/j.parkreldis.2022.08.022>
- Durbán A, Abellán JJ, Jiménez-Hernández N, Salgado P, Ponce M, Ponce J, Garrigues V, Latorre A, Moya A (2012) Structural alterations of faecal and mucosa-associated bacterial communities in irritable bowel syndrome. Environ Microbiol Rep 4:242–247. <https://doi.org/10.1111/j.1758-2229.2012.00327.x>
- Fan Y, Xu C, Xie L, Wang Y, Zhu S, An J, Li Y, Tian Z, Yan Y, Yu S, Liu H, Jia B, Wang Y, Wang L, Yang L, Bian Y (2022) Abnormal bile acid metabolism is an important feature of gut microbiota and fecal metabolites in patients with slow transit constipation. Front Cell Infect Microbiol 12:956528. [https://doi.org/10.3389/](https://doi.org/10.3389/fcimb.2022.956528) [fcimb.2022.956528](https://doi.org/10.3389/fcimb.2022.956528)
- Ford AC, Quigley EMM, Lacy BE, Lembo AJ, Saito YA, Schiller LR, Soffer EE, Spiegel BMR, Moayyedi P (2014) Efficacy of prebiotics, probiotics, and synbiotics in irritable bowel syndrome and chronic idiopathic constipation: systematic review and metaanalysis. Am J Gastroenterol 109:1547–1562. [https://doi.org/](https://doi.org/10.1038/ajg.2014.202) [10.1038/ajg.2014.202](https://doi.org/10.1038/ajg.2014.202)
- Fujimura KE, Slusher NA, Cabana MD, Lynch SV (2010) Role of the gut microbiota in defning human health. Expert Rev Anti Infect Ther 8:435–454. <https://doi.org/10.1586/eri.10.14>
- Fukui H, Xu X, Miwa H (2018) Role of gut microbiota-gut hormone axis in the pathophysiology of functional gastrointestinal disorders. J Neurogastroenterol Motil 24:367–386. [https://doi.org/10.](https://doi.org/10.5056/jnm18071) [5056/jnm18071](https://doi.org/10.5056/jnm18071)
- Ghoshal UC, Srivastava D, Misra A (2018) A randomized double-blind placebo-controlled trial showing rifaximin to improve constipation by reducing methane production and accelerating colon transit: a pilot study. Indian J Gastroenterol 37:416–423. [https://](https://doi.org/10.1007/s12664-018-0901-6) doi.org/10.1007/s12664-018-0901-6
- Gribble FM, Reimann F (2019) Function and mechanisms of enteroendocrine cells and gut hormones in metabolism. Nat Rev Endocrinol 15:226–237.<https://doi.org/10.1038/s41574-019-0168-8>
- Groneberg D, Voussen B, Friebe A (2016) Integrative control of gastrointestinal motility by nitric oxide. Curr Med Chem 23:2715– 2735.<https://doi.org/10.2174/0929867323666160812150907>
- Hayes CL, Dong J, Galipeau HJ, Jury J, McCarville J, Huang X, Wang XY, Naidoo A, Anbazhagan AN, Libertucci J, Sheridan C, Dudeja PK, Bowdish DME, Surette MG, Verdu EF (2018) Commensal microbiota induces colonic barrier structure and functions that contribute to homeostasis. Sci Rep 8:14184
- Hofmann TW, Pham H, Bridonneau C, Aubry C, Lamas B, Martin-Gallausiaux C, Moroldo M, Rainteau D, Lapaque N, Six A, Richard ML, Fargier E, Le Guern M, Langella P, Sokol H (2016) Microorganisms linked to infammatory bowel disease-associated dysbiosis diferentially impact host physiology in gnotobiotic mice. ISME J 10:460–477.<https://doi.org/10.1038/ismej.2015.127>
- Hoving JC, Wilson GJ, Brown GD (2014) Signalling C-type lectin receptors, microbial recognition and immunity. Cell Microbiol 16:185–194. <https://doi.org/10.1111/cmi.12249>
- Huang J, Lin B, Zhang Y, Xie Z, Zheng Y, Wang Q, Xiao H (2022) Bamboo shavings derived O-acetylated xylan alleviates loperamide-induced constipation in mice. Carbohydr Polym 276:118761. <https://doi.org/10.1016/j.carbpol.2021.118761>
- Inatomi T, Honma M (2021) Efects of probiotics on loperamideinduced constipation in rats. Sci Rep 11:24098. [https://doi.org/](https://doi.org/10.1038/s41598-021-02931-7) [10.1038/s41598-021-02931-7](https://doi.org/10.1038/s41598-021-02931-7)
- Jo JH, Kennedy EA, Kong HH (2016) Research techniques made simple: bacterial 16S ribosomal RNA gene sequencing in cutaneous pesearch. J Invest Dermatol 136:e23–e27. [https://doi.org/](https://doi.org/10.1016/j.jid.2016.01.005) [10.1016/j.jid.2016.01.005](https://doi.org/10.1016/j.jid.2016.01.005)
- Johansson MEV, Sjövall H, Hansson GC (2013) The gastrointestinal mucus system in health and disease. Nat Rev Gastroenterol Hepatol 10:352–361.<https://doi.org/10.1038/nrgastro.2013.35>
- Johansson MEV, Jakobsson HE, Holmén-Larsson J, Schütte A, Ermund A, Rodríguez-Piñeiro AM, Arike L, Wising C, Svensson F, Bäckhed F, Hansson GC (2015) Normalization of host intestinal mucus layers requires long-term microbial colonization. Cell Host Microbe 18:582–592. [https://doi.org/10.1016/j.chom.](https://doi.org/10.1016/j.chom.2015.10.007) [2015.10.007](https://doi.org/10.1016/j.chom.2015.10.007)
- Johnsen PH, Hilpüsch F, Cavanagh JP, Leikanger IS, Kolstad C, Valle PC, Goll R (2018) Faecal microbiota transplantation versus placebo for moderate-to-severe irritable bowel syndrome: a doubleblind, randomised, placebo-controlled, parallel-group, singlecentre trial. Lancet Gastroenterol Hepatol 3:17–24. [https://doi.](https://doi.org/10.1016/S2468-1253(17)30338-2) [org/10.1016/S2468-1253\(17\)30338-2](https://doi.org/10.1016/S2468-1253(17)30338-2)
- Joshi V, Strege PR, Farrugia G, Beyder A (2021) Mechanotransduction in gastrointestinal smooth muscle cells: role of mechanosensitive

ion channels. Am J Physiol Gastrointest Liver Physiol 320:G897– G906. <https://doi.org/10.1152/ajpgi.00481.2020>

- Keely SJ, Walters JRF (2016) The farnesoid X receptor: good for bad. Cell Mol Gastroenterol Hepatol 2:725–732. [https://doi.org/10.](https://doi.org/10.1016/j.jcmgh.2016.08.004) [1016/j.jcmgh.2016.08.004](https://doi.org/10.1016/j.jcmgh.2016.08.004)
- Keely SJ, Urso A, Ilyaskin AV, Korbmacher C, Bunnett NW, Poole DP, Carbone SE (2022) Contributions of bile acids to gastrointestinal physiology as receptor agonists and modifers of ion channels. Am J Physiol Gastrointest Liver Physiol 322:G201–G222. <https://doi.org/10.1152/ajpgi.00125.2021>
- Khan WI, Collins SM (2004) Immune-mediated alteration in gut physiology and its role in host defence in nematode infection. Parasite Immunol 26:319–326. [https://doi.org/10.1111/j.0141-9838.2004.](https://doi.org/10.1111/j.0141-9838.2004.00715.x) [00715.x](https://doi.org/10.1111/j.0141-9838.2004.00715.x)
- Kim S, Choi SC, Park KS, Park MI, Shin JE, Lee TH, Jung KW, Koo HS, Myung S (2015) Change of fecal fora and efectiveness of the short-term VSL#3 probiotic treatment in patients with functional constipation. J Neurogastroenterol Motil 21:111–120. <https://doi.org/10.5056/jnm14048>
- Koch CD, Gladwin MT, Freeman BA, Lundberg JO, Weitzberg E, Morris A (2017) Enterosalivary nitrate metabolism and the microbiome: intersection of microbial metabolism, nitric oxide and diet in cardiac and pulmonary vascular health. Free Radic Biol Med 105:48–67.<https://doi.org/10.1016/j.freeradbiomed.2016.12.015>
- Kuai XY, Yao XH, Xu LJ, Zhou YQ, Zhang LP, Liu Y, Pei SF, Zhou CL (2021) Evaluation of fecal microbiota transplantation in Parkinson's disease patients with constipation. Microb Cell Fact 20:98.<https://doi.org/10.1186/s12934-021-01589-0>
- Kurilshikov A, Wijmenga C, Fu J, Zhernakova A (2017) Host genetics and gut microbiome: challenges and perspectives. Trends Immunol 38:633–647. <https://doi.org/10.1016/j.it.2017.06.003>
- Lee K, Paik C, Chung WC, Yang J, Choi M (2013) Breath methane positivity is more common and higher in patients with objectively proven delayed transit constipation. Eur J Gastroenterol Hepatol 25:726–732. [https://doi.org/10.1097/MEG.0b013e3283](https://doi.org/10.1097/MEG.0b013e32835eb916) [5eb916](https://doi.org/10.1097/MEG.0b013e32835eb916)
- Li H, Limenitakis JP, Fuhrer T, Geuking MB, Lawson MA, Wyss M, Brugiroux S, Keller I, Macpherson JA, Rupp S, Stolp B, Stein JV, Stecher B, Sauer U, McCoy KD, Macpherson AJ (2015) The outer mucus layer hosts a distinct intestinal microbial niche. Nat Commun 6:8292. <https://doi.org/10.1038/ncomms9292>
- Li Y, Zhang Y, Wei K, He J, Ding N, Hua J, Zhou T, Niu F, Zhou G, Shi T, Zhang L, Liu Y (2021b) Review: efect of gut microbiota and its metabolite SCFAs on radiation-induced intestinal injury. Front Cell Infect Microbiol 11:577236. [https://doi.org/10.3389/](https://doi.org/10.3389/fcimb.2021.577236) [fcimb.2021.577236](https://doi.org/10.3389/fcimb.2021.577236)
- Li N, Koester ST, Lachance DM, Dutta M, Cui JY, Dey N (2021a) Microbiome-encoded bile acid metabolism modulates colonic transit times. iScience 24: 102508. [https://doi.org/10.1016/j.isci.](https://doi.org/10.1016/j.isci.2021a.102508) [2021a.102508](https://doi.org/10.1016/j.isci.2021a.102508)
- Liang Y, Wen P, Wang Y, OuYang D, Wang D, Chen Y, Song Y, Deng J, Sun Y, Wang H (2019) The constipation-relieving property of d-tagatose by modulating the composition of gut microbiota. Int J Mol Sci 20:5721. <https://doi.org/10.3390/ijms20225721>
- Lin X, Liu Y, Ma L, Ma X, Shen L, Ma X, Chen Z, Chen H, Li D, Su Z, Chen X (2021) Constipation induced gut microbiota dysbiosis exacerbates experimental autoimmune encephalomyelitis in C57BL/6 mice. J Transl Med 19:317. [https://doi.org/10.1186/](https://doi.org/10.1186/s12967-021-02995-z) [s12967-021-02995-z](https://doi.org/10.1186/s12967-021-02995-z)
- Liu D, Lin L, Lin Y, Zhong Y, Zhang S, Liu W, Zou B, Liao Q, Xie Z (2019) Zengye decoction induces alterations to metabolically active gut microbiota in aged constipated rats. Biomed Pharmacother 109:1361–1371. [https://doi.org/10.1016/j.biopha.](https://doi.org/10.1016/j.biopha.2018.11.013) [2018.11.013](https://doi.org/10.1016/j.biopha.2018.11.013)
- Liu J, Gu L, Zhang M, Zhang S, Wang M, Long Y, Zhang X (2021) The fecal microbiota transplantation: a remarkable clinical

therapy for slow transit constipation in future. Front Cell Infect Microbiol 11:732474. [https://doi.org/10.3389/fcimb.](https://doi.org/10.3389/fcimb.2021.732474) [2021.732474](https://doi.org/10.3389/fcimb.2021.732474)

- Low K, Hwang L, Hua J, Zhu A, Morales W, Pimentel M (2010) A combination of rifaximin and neomycin is most efective in treating irritable bowel syndrome patients with methane on lactulose breath test. J Clin Gastroenterol 44:547–550. [https://](https://doi.org/10.1097/MCG.0b013e3181c64c90) doi.org/10.1097/MCG.0b013e3181c64c90
- Mancabelli L, Milani C, Lugli GA, Turroni F, Mangifesta M, Viappiani A, Ticinesi A, Nouvenne A, Meschi T, van Sinderen D, Ventura M (2017) Unveiling the gut microbiota composition and functionality associated with constipation through metagenomic analyses. Sci Rep 7:9879. <https://doi.org/10.1038/s41598-017-10663-w>
- Martin-Gallausiaux C, Marinelli L, Blottière HM, Larraufe P, Lapaque N (2021) SCFA: mechanisms and functional importance in the gut. Proc Nutr Soc 80:37–49. [https://doi.org/10.1017/S0029](https://doi.org/10.1017/S0029665120006916) [665120006916](https://doi.org/10.1017/S0029665120006916)
- Mathias A, Duc M, Favre L, Benyacoub J, Blum S, Corthésy B (2010) Potentiation of polarized intestinal Caco-2 cell responsiveness to probiotics complexed with secretory IgA. J Biol Chem 285:33906–33913.<https://doi.org/10.1074/jbc.M110.135111>
- Mazzawi T, El-Salhy M, Lied GA, Hausken T (2021) The efects of fecal microbiota transplantation on the symptoms and the duodenal neurogenin 3, musashi 1, and enteroendocrine cells in patients with diarrhea-predominant irritable bowel syndrome. Front Cell Infect Microbiol 11:524851. [https://doi.org/10.3389/](https://doi.org/10.3389/fcimb.2021.524851) [fcimb.2021.524851](https://doi.org/10.3389/fcimb.2021.524851)
- Mazzone A, Farrugia G (2007) Evolving concepts in the cellular control of gastrointestinal motility: neurogastroenterology and enteric sciences. Gastroenterol Clin North Am 36:499–513. <https://doi.org/10.1016/j.gtc.2007.07.003>
- Mugie SM, Benninga MA, Di Lorenzo C (2011) Epidemiology of constipation in children and adults: a systematic review. Best Pract Res Clin Gastroenterol 25:3–18. [https://doi.org/10.1016/j.bpg.](https://doi.org/10.1016/j.bpg.2010.12.010) [2010.12.010](https://doi.org/10.1016/j.bpg.2010.12.010)
- Nakajima A, Seki M, Taniguchi S, Ohta A, Gillberg P, Mattsson JP, Camilleri M (2018) Safety and efficacy of elobixibat for chronic constipation: results from a randomised, double-blind, placebocontrolled, phase 3 trial and an open-label, single-arm, phase 3 trial. Lancet Gastroenterol Hepatol 3:537–547. [https://doi.org/](https://doi.org/10.1016/S2468-1253(18)30123-7) [10.1016/S2468-1253\(18\)30123-7](https://doi.org/10.1016/S2468-1253(18)30123-7)
- Nakajima A, Ishizaki S, Matsuda K, Kurosu S, Taniguchi S, Gillberg PG, Mattsson JP, Hasunuma T, Camilleri M (2022) Impact of elobixibat on serum and fecal bile acid levels and constipation symptoms in patients with chronic constipation. J Gastroenterol Hepatol 37:883–890.<https://doi.org/10.1111/jgh.15800>
- Obata Y, Pachnis V (2016) The effect of microbiota and the immune system on the development and organization of the enteric nervous system. Gastroenterology 151:836–844. [https://doi.org/10.](https://doi.org/10.1053/j.gastro.2016.07.044) [1053/j.gastro.2016.07.044](https://doi.org/10.1053/j.gastro.2016.07.044)
- Obata Y, Castaño Á, Boeing S, Bon-Frauches AC, Fung C, Fallesen T, de Agüero MG, Yilmaz B, Lopes R, Huseynova A, Horswell S, Maradana MR, Boesmans W, Vanden Berghe P, Murray AJ, Stockinger B, Macpherson AJ, Pachnis V (2020) Neuronal programming by microbiota regulates intestinal physiology. Nature 578:284–289.<https://doi.org/10.1038/s41586-020-1975-8>
- Oeser K, Schwartz C, Voehringer D (2015) Conditional IL-4/IL-13-defcient mice reveal a critical role of innate immune cells for protective immunity against gastrointestinal helminths. Mucosal Immunol 8:672–682. [https://doi.org/10.1038/mi.](https://doi.org/10.1038/mi.2014.101) [2014.101](https://doi.org/10.1038/mi.2014.101)
- Ohkusa T, Koido S, Nishikawa Y, Sato N (2019) Gut microbiota and chronic constipation: a review and update. Front Med 6:19. <https://doi.org/10.3389/fmed.2019.00019>
- Ojetti V, Petruzziello C, Migneco A, Gnarra M, Gasbarrini A, Franceschi F (2017) Efect of *Lactobacillus reuteri* (DSM 17938)

on methane production in patients afected by functional constipation: a retrospective study. Eur Rev Med Pharmacol Sci 21:1702–1708

- Ostaff MJ, Stange EF, Wehkamp J (2013) Antimicrobial peptides and gut microbiota in homeostasis and pathology. EMBO Mol Med 5:1465–1483.<https://doi.org/10.1002/emmm.201201773>
- Parthasarathy G, Chen J, Chen X, Chia N, O'Connor HM, Wolf PG, Gaskins HR, Bharucha AE (2016) Relationship between microbiota of the colonic mucosa vs feces and symptoms, colonic transit, and methane production in female patients with chronic constipation. Gastroenterology 150:367–379. [https://doi.org/](https://doi.org/10.1053/j.gastro.2015.10.005) [10.1053/j.gastro.2015.10.005](https://doi.org/10.1053/j.gastro.2015.10.005)
- Potrykus M, Czaja-Stolc S, Stankiewicz M, Kaska Ł, Małgorzewicz S (2021) Intestinal microbiota as a contributor to chronic inflammation and its potential modifications. Nutrients 13:3839. <https://doi.org/10.3390/nu13113839>
- Prichard DO, Bharucha AE (2018) Recent advances in understanding and managing chronic constipation. F1000Res 7: F1000 Faculty Rev-1640. [https://doi.org/10.12688/f1000research.](https://doi.org/10.12688/f1000research.15900.1) [15900.1](https://doi.org/10.12688/f1000research.15900.1)
- Puschhof J, Pleguezuelos-Manzano C, Clevers H (2021) Organoids and organs-on-chips: insights into human gut-microbe interactions. Cell Host Microbe 29:867–878. [https://doi.org/10.](https://doi.org/10.1016/j.chom.2021.04.002) [1016/j.chom.2021.04.002](https://doi.org/10.1016/j.chom.2021.04.002)
- Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto J, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Li S, Jian M, Zhou Y, Li Y, Zhang X, Li S, Qin N, Yang H, Wang J, Brunak S, Doré J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J, Bork P, Ehrlich SD, Wang J (2010) A human gut microbial gene catalogue established by metagenomic sequencing. Nature 464:59–65. <https://doi.org/10.1038/nature08821>
- Quigley EMM (2011) The enteric microbiota in the pathogenesis and management of constipation. Best Pract Res Clin Gastroenterol 25:119–126.<https://doi.org/10.1016/j.bpg.2011.01.003>
- Rao SSC, Rehman A, Yu S, Andino NMD (2018) Brain fogginess, gas and bloating: a link between SIBO, probiotics and metabolic acidosis. Clin Transl Gastroenterol 9:162. [https://doi.org/](https://doi.org/10.1038/s41424-018-0030-7) [10.1038/s41424-018-0030-7](https://doi.org/10.1038/s41424-018-0030-7)
- Roager HM, Hansen LBS, Bahl MI, Frandsen HL, Carvalho V, Gøbel RJ, Dalgaard MD, Plichta DR, Sparholt MH, Vestergaard H, Hansen T, Sicheritz-Pontén T, Nielsen HB, Pedersen O, Lauritzen L, Kristensen M, Gupta R, Licht TR (2016) Colonic transit time is related to bacterial metabolism and mucosal turnover in the gut. Nat Microbiol 1:16093. [https://doi.org/10.](https://doi.org/10.1038/nmicrobiol.2016.93) [1038/nmicrobiol.2016.93](https://doi.org/10.1038/nmicrobiol.2016.93)
- Rogier EW, Frantz AL, Bruno MEC, Kaetzel CS (2014) Secretory IgA is concentrated in the outer layer of colonic mucus along with gut bacteria. Pathogens 3:390–403. [https://doi.org/10.](https://doi.org/10.3390/pathogens3020390) [3390/pathogens3020390](https://doi.org/10.3390/pathogens3020390)
- Rossi F, Amadoro C, Gasperi M, Colavita G (2022) *Lactobacilli* infection case reports in the last three years and safety implications. Nutrients 14:1178. <https://doi.org/10.3390/nu14061178>
- Sagheddu V, Patrone V, Miragoli F, Puglisi E, Morelli L (2016) Infant early gut colonization by *Lachnospiraceae*: high frequency of *Ruminococcus gnavus*. Front Pediatr 4:57. [https://doi.org/10.](https://doi.org/10.3389/fped.2016.00057) [3389/fped.2016.00057](https://doi.org/10.3389/fped.2016.00057)
- Sahakian AB, Jee S, Pimentel M (2010) Methane and the gastrointestinal tract. Dig Dis Sci 55:2135–2143. [https://doi.org/10.1007/](https://doi.org/10.1007/s10620-009-1012-0) [s10620-009-1012-0](https://doi.org/10.1007/s10620-009-1012-0)
- Schoeler M, Caesar R (2019) Dietary lipids, gut microbiota and lipid metabolism. Rev Endocr Metab Disord 20:461–472. [https://doi.](https://doi.org/10.1007/s11154-019-09512-0) [org/10.1007/s11154-019-09512-0](https://doi.org/10.1007/s11154-019-09512-0)
- Schütte A, Ermund A, Becker-Pauly C, Johansson MEV, Rodriguez-Pineiro AM, Bäckhed F, Müller S, Lottaz D, Bond JS, Hansson GC (2014) Microbial-induced meprin β cleavage in MUC2 mucin and a functional CFTR channel are required to release anchored small intestinal mucus. Proc Natl Acad Sci U S A 111:12396–12401.<https://doi.org/10.1073/pnas.1407597111>
- Shi N, Li N, Duan X, Niu H (2017) Interaction between the gut microbiome and mucosal immune system. Mil Med Res 4:14. [https://](https://doi.org/10.1186/s40779-017-0122-9) doi.org/10.1186/s40779-017-0122-9
- Sicard JF, Le Bihan G, Vogeleer P, Jacques M, Harel J (2017) Interactions of intestinal bacteria with components of the intestinal mucus. Front Cell Infect Microbiol 7:387. [https://doi.org/10.](https://doi.org/10.3389/fcimb.2017.00387) [3389/fcimb.2017.00387](https://doi.org/10.3389/fcimb.2017.00387)
- Sotoudegan F, Daniali M, Hassani S, Nikfar S, Abdollahi M (2019) Reappraisal of probiotics' safety in human. Food Chem Toxicol 129:22–29. <https://doi.org/10.1016/j.fct.2019.04.032>
- Steensels S, Depoortere I (2018) Chemoreceptors in the gut. Annu Rev Physiol 80:117–141. [https://doi.org/10.1146/annurev-physi](https://doi.org/10.1146/annurev-physiol-021317-121332) [ol-021317-121332](https://doi.org/10.1146/annurev-physiol-021317-121332)
- Suchodolski JS, Jergens AE (2016) Recent advances and understanding of using probiotic-based interventions to restore homeostasis of the microbiome for the prevention/therapy of bacterial diseases. Microbiol Spectr 4. [https://doi.org/10.1128/microbiolspec.](https://doi.org/10.1128/microbiolspec.VMBF-0025-2015) [VMBF-0025-2015](https://doi.org/10.1128/microbiolspec.VMBF-0025-2015)
- Suez J, Zmora N, Segal E, Elinav E (2019) The pros, cons, and many unknowns of probiotics. Nat Med 25:716–729. [https://doi.org/](https://doi.org/10.1038/s41591-019-0439-x) [10.1038/s41591-019-0439-x](https://doi.org/10.1038/s41591-019-0439-x)
- Takakura W, Pimentel M, Rao S, Villanueva-Millan MJ, Chang C, Morales W, Sanchez M, Torosyan J, Rashid M, Hosseini A, Wang J, Leite G, Kowalewski E, Mathur R, Rezaie A (2022) A single fasting exhaled methane level correlates with fecal methanogen load, clinical symptoms and accurately detects intestinal methanogen overgrowth. Am J Gastroenterol 117:470–477. <https://doi.org/10.14309/ajg.0000000000001607>
- Tian H, Chen Q, Yang B, Qin H, Li N (2021) Analysis of gut microbiome and metabolite characteristics in patients with slow transit constipation. Dig Dis Sci 66:3026–3035. [https://doi.org/10.1007/](https://doi.org/10.1007/s10620-020-06500-2) [s10620-020-06500-2](https://doi.org/10.1007/s10620-020-06500-2)
- Tong X, Xu J, Lian F, Yu X, Zhao Y, Xu L, Zhang M, Zhao X, Shen J, Wu S, Pang X, Tian J, Zhang C, Zhou Q, Wang L, Pang B, Chen F, Peng Z, Wang J, Zhen Z, Fang C, Li M, Chen L, Zhao L (2018) Structural alteration of gut microbiota during the amelioration of human type 2 diabetes with hyperlipidemia by metformin and a traditional chinese herbal formula: a multicenter, randomized, open label clinical trial. mBio 9: e02392–17. [https://](https://doi.org/10.1128/mBio.02392-17) doi.org/10.1128/mBio.02392-17
- Tough IR, Forbes S, Cox HM (2018) Signaling of free fatty acid receptors 2 and 3 difers in colonic mucosa following selective agonism or coagonism by luminal propionate. Neurogastroenterol Motil 30:e13454.<https://doi.org/10.1111/nmo.13454>
- Triantafyllou K, Chang C, Pimentel M (2014) Methanogens, methane and gastrointestinal motility. J Neurogastroenterol Motil 20:31– 40.<https://doi.org/10.5056/jnm.2014.20.1.31>
- Vandeputte D, Falony G, Vieira-Silva S, Tito RY, Joossens M, Raes J (2016) Stool consistency is strongly associated with gut microbiota richness and composition, enterotypes and bacterial growth rates. Gut 65:57–62. [https://doi.org/10.1136/](https://doi.org/10.1136/gutjnl-2015-309618) [gutjnl-2015-309618](https://doi.org/10.1136/gutjnl-2015-309618)
- Verberkmoes NC, Russell AL, Shah M, Godzik A, Rosenquist M, Halfvarson J, Lefsrud MG, Apajalahti J, Tysk C, Hettich RL, Jansson JK (2009) Shotgun metaproteomics of the human distal gut microbiota. ISME J 3:179–189. [https://doi.org/10.1038/](https://doi.org/10.1038/ismej.2008.108) [ismej.2008.108](https://doi.org/10.1038/ismej.2008.108)
- Vicentini FA, Keenan CM, Wallace LE, Woods C, Cavin J, Flockton AR, Macklin WB, Belkind-Gerson J, Hirota SA, Sharkey KA (2021) Intestinal microbiota shapes gut physiology and

regulates enteric neurons and glia. Microbiome 9:210. [https://](https://doi.org/10.1186/s40168-021-01165-z) doi.org/10.1186/s40168-021-01165-z

- Vincent AD, Wang XY, Parsons SP, Khan WI, Huizinga JD (2018) Abnormal absorptive colonic motor activity in germ-free mice is rectified by butyrate, an effect possibly mediated by mucosal serotonin. Am J Physiol Gastrointest Liver Physiol 315:G896– G907. <https://doi.org/10.1152/ajpgi.00237.2017>
- Vonk RJ, Reckman G (2017) Progress in the biology and analysis of short chain fatty acids. J Physiol 595:419–420. [https://doi.](https://doi.org/10.1113/JP273260) [org/10.1113/JP273260](https://doi.org/10.1113/JP273260)
- Vriesman MH, Koppen IJN, Camilleri M, Di Lorenzo C, Benninga MA (2020) Management of functional constipation in children and adults. Nat Rev Gastroenterol Hepatol 17:21–39. [https://](https://doi.org/10.1038/s41575-019-0222-y) doi.org/10.1038/s41575-019-0222-y
- Vyhlídalová B, Krasulová K, Pečinková P, Marcalíková A, Vrzal R, Zemánková L, Vančo J, Trávníček Z, Vondráček J, Karasová M, Mani S, Dvořák Z (2020) Gut microbial catabolites of tryptophan are ligands and agonists of the aryl hydrocarbon receptor: a detailed characterization. Int J Mol Sci 21:2614. <https://doi.org/10.3390/ijms21072614>
- Wahlström A, Sayin SI, Marschall H, Bäckhed F (2016) Intestinal crosstalk between bile acids and microbiota and its impact on host metabolism. Cell Metab 24:41–50. [https://doi.org/10.](https://doi.org/10.1016/j.cmet.2016.05.005) [1016/j.cmet.2016.05.005](https://doi.org/10.1016/j.cmet.2016.05.005)
- Wang X, Yin J (2015) Complementary and alternative therapies for chronic constipation. Evid Based Complement Alternat Med 2015:396396. <https://doi.org/10.1155/2015/396396>
- Wells JM, Rossi O, Meijerink M, van Baarlen P (2011) Epithelial crosstalk at the microbiota-mucosal interface. Proc Natl Acad Sci U S A 108(Suppl 1):4607–4614. [https://doi.org/10.1073/](https://doi.org/10.1073/pnas.1000092107) [pnas.1000092107](https://doi.org/10.1073/pnas.1000092107)
- Wensel CR, Pluznick JL, Salzberg SL, Sears CL (2022) Next-generation sequencing: insights to advance clinical investigations of the microbiome. J Clin Invest 132:e154944. [https://doi.org/](https://doi.org/10.1172/JCI154944) [10.1172/JCI154944](https://doi.org/10.1172/JCI154944)
- Wichmann A, Allahyar A, Greiner TU, Plovier H, Lundén GÖ, Larsson T, Drucker DJ, Delzenne NM, Cani PD, Bäckhed F (2013) Microbial modulation of energy availability in the colon regulates intestinal transit. Cell Host Microbe 14:582–590. [https://](https://doi.org/10.1016/j.chom.2013.09.012) doi.org/10.1016/j.chom.2013.09.012
- Williams BB, Van Benschoten AH, Cimermancic P, Donia MS, Zimmermann M, Taketani M, Ishihara A, Kashyap PC, Fraser JS, Fischbach MA (2014) Discovery and characterization of gut microbiota decarboxylases that can produce the neurotransmitter tryptamine. Cell Host Microbe 16:495–503. [https://doi.org/](https://doi.org/10.1016/j.chom.2014.09.001) [10.1016/j.chom.2014.09.001](https://doi.org/10.1016/j.chom.2014.09.001)
- Wrzosek L, Miquel S, Noordine M, Bouet S, Joncquel Chevalier-Curt M, Robert V, Philippe C, Bridonneau C, Cherbuy C, Robbe-Masselot C, Langella P, Thomas M (2013) *Bacteroides thetaiotaomicron* and *Faecalibacterium prausnitzii* infuence the production of mucus glycans and the development of goblet cells in the colonic epithelium of a gnotobiotic model rodent. BMC Biol 11:61.<https://doi.org/10.1186/1741-7007-11-61>
- Wu M, Wu Y, Li J, Bao Y, Guo Y, Yang W (2018) The dynamic changes of gut microbiota in *Muc2* defcient mice. Int J Mol Sci 19:2809. <https://doi.org/10.3390/ijms19092809>
- Xiao L, Chen B, Feng D, Yang T, Li T, Chen J (2019) TLR4 may be involved in the regulation of colonic mucosal microbiota by vitamin A. Front Microbiol 10:268. [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2019.00268) [fmicb.2019.00268](https://doi.org/10.3389/fmicb.2019.00268)
- Xiao SW, Liu C, Chen MJ, Zou JF, Zhang ZM, Cui X, Jiang S, Shang EX, Qian DW, Duan JA (2020) Scutellariae radix and coptidis rhizoma ameliorate glycolipid metabolism of type 2 diabetic rats by modulating gut microbiota and its metabolites. Appl

Microbiol Biotechnol 104:303–317. [https://doi.org/10.1007/](https://doi.org/10.1007/s00253-019-10174-w) [s00253-019-10174-w](https://doi.org/10.1007/s00253-019-10174-w)

- Xu J, Lian F, Zhao L, Zhao Y, Chen X, Zhang X, Guo Y, Zhang C, Zhou Q, Xue Z, Pang X, Zhao L, Tong X (2015) Structural modulation of gut microbiota during alleviation of type 2 diabetes with a Chinese herbal formula. ISME J 9:552–562. <https://doi.org/10.1038/ismej.2014.177>
- Yang L, Wan Y, Li WW, Liu C, Li HF, Dong ZL, Zhu K, Jiang S, Shang EX, Qian DW, Duan JA (2022) Targeting intestinal fora and its metabolism to explore the laxative efects of rhubarb. Appl Microbiol Biotechnol 106:1615–1631. [https://doi.org/10.](https://doi.org/10.1007/s00253-022-11813-5) [1007/s00253-022-11813-5](https://doi.org/10.1007/s00253-022-11813-5)
- Yarandi SS, Kulkarni S, Saha M, Sylvia KE, Sears CL, Pasricha PJ (2020) Intestinal bacteria maintain adult enteric nervous system and nitrergic neurons via Toll-like receptor 2-induced neurogenesis in mice. Gastroenterology 159:200–213. [https://](https://doi.org/10.1053/j.gastro.2020.03.050) doi.org/10.1053/j.gastro.2020.03.050
- Ye L, Bae M, Cassilly CD, Jabba SV, Thorpe DW, Martin AM, Lu H, Wang J, Thompson JD, Lickwar CR, Poss KD, Keating DJ, Jordt S, Clardy J, Liddle RA, Rawls JF (2021) Enteroendocrine cells sense bacterial tryptophan catabolites to activate enteric and vagal neuronal pathways. Cell Host Microbe 29:179–196. <https://doi.org/10.1016/j.chom.2020.11.011>
- Yeşilyurt N, Yılmaz B, Ağagündüz D, Capasso R (2021) Involvement of probiotics and postbiotics in the immune system modulation. Biologics 1:89–110. [https://doi.org/10.3390/biologics1](https://doi.org/10.3390/biologics1020006) [020006](https://doi.org/10.3390/biologics1020006)
- Yu Y, Yang W, Li Y, Cong Y (2020) Enteroendocrine cells: sensing gut microbiota and regulating infammatory bowel diseases. Infamm Bowel Dis 26:11–20. [https://doi.org/10.1093/ibd/](https://doi.org/10.1093/ibd/izz217) [izz217](https://doi.org/10.1093/ibd/izz217)
- Zhang X, Li N, Chen Q, Qin H (2021a) Fecal microbiota transplantation modulates the gut fora favoring patients with functional constipation. Front Microbiol 12:700718. [https://doi.org/10.](https://doi.org/10.3389/fmicb.2021.700718) [3389/fmicb.2021.700718](https://doi.org/10.3389/fmicb.2021.700718)
- Zhang ZM, Yang L, Wan Y, Liu C, Jiang S, Shang EX, Duan JA (2021b) Integrated gut microbiota and fecal metabolomics reveal the renoprotective efect of Rehmanniae Radix Preparata and Corni Fructus on adenine-induced CKD rats. J Chromatogr B 1174:122728. [https://doi.org/10.1016/j.jchromb.2021.](https://doi.org/10.1016/j.jchromb.2021.122728) [122728](https://doi.org/10.1016/j.jchromb.2021.122728)
- Zhao Q, Maynard CL (2022) Mucus, commensals, and the immune system. Gut Microbes 14:2041342. [https://doi.org/10.1080/](https://doi.org/10.1080/19490976.2022.2041342) [19490976.2022.2041342](https://doi.org/10.1080/19490976.2022.2041342)
- Zhao Q, Chen YY, Xu DQ, Yue SJ, Fu RJ, Yang J, Xing LM, Tang YP (2021) Action mode of gut motility, fuid and electrolyte transport in chronic constipation. Front Pharmacol 12:630249. <https://doi.org/10.3389/fphar.2021.630249>
- Zhu L, Liu W, Alkhouri R, Baker RD, Bard JE, Quigley EM, Baker SS (2014) Structural changes in the gut microbiome of constipated patients. Physiol Genomics 46:679–686. [https://doi.org/](https://doi.org/10.1152/physiolgenomics.00082.2014) [10.1152/physiolgenomics.00082.2014](https://doi.org/10.1152/physiolgenomics.00082.2014)
- Zhuang M, Shang W, Ma Q, Strappe P, Zhou Z (2019) Abundance of probiotics and butyrate-production microbiome manages constipation via short-chain fatty acids production and hormones secretion. Mol Nutr Food Res 63:e1801187. [https://doi.org/10.](https://doi.org/10.1002/mnfr.201801187) [1002/mnfr.201801187](https://doi.org/10.1002/mnfr.201801187)
- Żółkiewicz J, Marzec A, Ruszczyński M, Feleszko W (2020) Postbiotics-A step beyond pre- and probiotics. Nutrients 12:2189. <https://doi.org/10.3390/nu12082189>
- Zoppi G, Cinquetti M, Luciano A, Benini A, Muner A, Bertazzoni Minelli E (1998) The intestinal ecosystem in chronic functional constipation. Acta Paediatr 87:836–841. [https://doi.org/10.1080/](https://doi.org/10.1080/080352598750013590) [080352598750013590](https://doi.org/10.1080/080352598750013590)

Zou JF, Shen YM, Chen MJ, Zhang ZM, Xiao SW, Liu C, Wan Y, Yang L, Jiang S, Shang EX, Qian DW, Duan JA (2020) Lizhong decoction ameliorates ulcerative colitis in mice via modulating gut microbiota and its metabolites. Appl Microbiol Biotechnol 104:5999–6012. <https://doi.org/10.1007/s00253-020-10665-1>

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