



Antimicrobial peptides used as growth promoters in livestock production

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Abstract

Antibiotic growth promoters (AGPs) have been administered in livestock for decades to improve food digestion in growing animals, while also contributing to the control of microbial pathogens. The long-term and indiscriminate use of AGPs has generated genetic modifications in bacteria, leading to antimicrobial resistance (AMR), which can be disseminated to commensal and pathogenic bacteria. Thus, antimicrobial peptides (AMPs) are used to replaced AGPs. AMPs are found in all domains of life, and their cationic characteristics can establish electrostatic interactions with the bacterial membrane. These molecules used as growth promoters can present benefits for nutrient digestibility, intestinal microbiota, intestinal morphology, and immune function activities. Therefore, this review focuses on the application of AMPs with growth promoting potential in livestock, as an alternative to conventional antibiotic growth promoters, in an attempt to control AMR.

Key points

- *The long-term and indiscriminate use of AGPs in animal food can cause AMR.*
- *AMPs can be used as substitute of antibiotics in animal food supplementation.*
- *Animal food supplemented with AMPs can provided economic efficiency and sustainable livestock production.*

Keywords Antibiotics · Antimicrobial resistance · Public health · Livestock farming · Food production

Introduction

Antibiotics have been of vast importance to human health and are also employed in farm animal health to control disease and as growth promoters. Antibiotic growth promoters (AGPs) have been administered in sub-therapeutic doses with the role of eradicating or inhibiting pathogenic bacteria (Hugues and Heritage 2004). AGPs are administered in livestock to improve the animals' digestion, so that they get the highest benefit from foodstuffs and grow into strong and healthy individuals (NOAH 2001; U.S. Food and Drug Administration 2015). Although the

AGP mechanism of action is unclear, it is supposed that AGPs inhibit the sensitive populations of bacteria in the intestines, and decrease energy loss with fermentative processes (Jensen 1998). AGPs also act in reducing the frequency and severity of subclinical infections (George et al. 1982; Brennan et al. 2003); they decrease microbial use of nutrients, and boost nutrient absorption, due to intestinal wall thinning (Snyder and Wostmann 1987; Feighner and Dashkevich 1987; Knarreborg et al. 2004; Huyghebaert et al. 2011). Thus, by regulating the microbial population and controlling microbial nutrients, energy is transformed into animal growth (Hugues and Heritage, 2004).

AGPs have been employed to improve the development of farm animals since the 1950s (Jones and Ricke 2003; Brown et al. 2017; Ronquillo and Hernandez 2017). Over the years, several antibiotics have been administered in livestock as GPs (Table 1). This use helped to produce meat on an industrial scale (Van Boeckel et al. 2015). However, the indiscriminate use of AGPs for decades caused genetic modifications in bacteria and has led to antimicrobial resistance (AMR), which can be disseminated to commensal and pathogenic bacteria (Aslam et al. 2018; Founou et al. 2016; Innes et al. 2020; Li

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Table 1 Antibiotics employed as growth promoters in livestock

Class	Antibiotic	Spectrum	Growth promotion
Aminoglycosides	Neomycin	Narrow spectrum	Cattle
	Gentamicin	Broad spectrum	Cattle, Swine
	Spectinomycin		Sheep
	Streptomycin		Chickens, Swine, Sheep, Cattle
Penicillins	Penicillin G potassium	Broad spectrum	Chickens
	Penicillin G procaine		Chickens, swine
Ionophores	Lasalocid sodium	Broad spectrum	Cattle
	Salinomycin		Cattle, Swine
	Narasin		Swine
	Monesin		Cattle
Macrolides	Erythromycin	Broad spectrum	Chickens
	Tylosin	Broad spectrum	Swine
	Tilmicosin	Narrow spectrum	Chickens
Streptogramins	Virginiamycin	Broad spectrum	Chickens, swine
Tetracyclines	Oxytetracycline	Broad spectrum	Chickens, swine, sheep, cattle
	Chlortetracycline		
B-lactam	Amoxicillin	Narrow spectrum	Chickens, swine, sheep, cattle
	Ampicillin	Broad spectrum	
	Penicillin V	Narrow spectrum	Swine
Bacitracin	Polypeptides	Narrow spectrum	Bovine

Adapted of Brown et al., 2017; Ronquillo and Hernandez, 2017

et al. 2018). AGPs are therefore the subject of controversy associated with their risks and advantages.

Thus, antimicrobial peptides (AMPs) are being used to replace AGPs (Jenssen et al. 2006; Cheng et al. 2014; Zhao et al. 2016). AMPs are found in all domains of life, present chemical diversity and structure, and usually present cationic and amphipathic properties (Cardoso et al. 2020; Gomes et al. 2018; Spohn et al. 2019; Brogden 2005; Jenssen et al. 2006). The cationic characteristics of AMPs can establish electrostatic interactions with the bacterial membrane, which is commonly composed of negatively charged phospholipids (Hancock and Chapple 1999; Shai 2002). AMPs can interact with the outer membrane, disturbing its physical integrity, and may also be translocated across the membrane and act on internal targets (Hancock and Sahl 2006). AMPs exhibit activity against bacteria, fungi, viruses, and cancer (Cardoso et al. 2020; Hwang et al. 2011; Oshiro et al. 2019; Rodrigues et al. 2019; Saito-Sakanaka et al. 2004). In addition, these peptides can act indirectly by stimulating the host's immune system (Ageitos et al. 2017; Hancock 2001; Ward et al. 2013; Wang et al. 2016; WHO 2014). Therefore, this review summarized the application of AMPs as growth promoters with potential for livestock, as an alternative to traditional antibiotics, in an effort to control AMR.

Concern about growth promoters in livestock

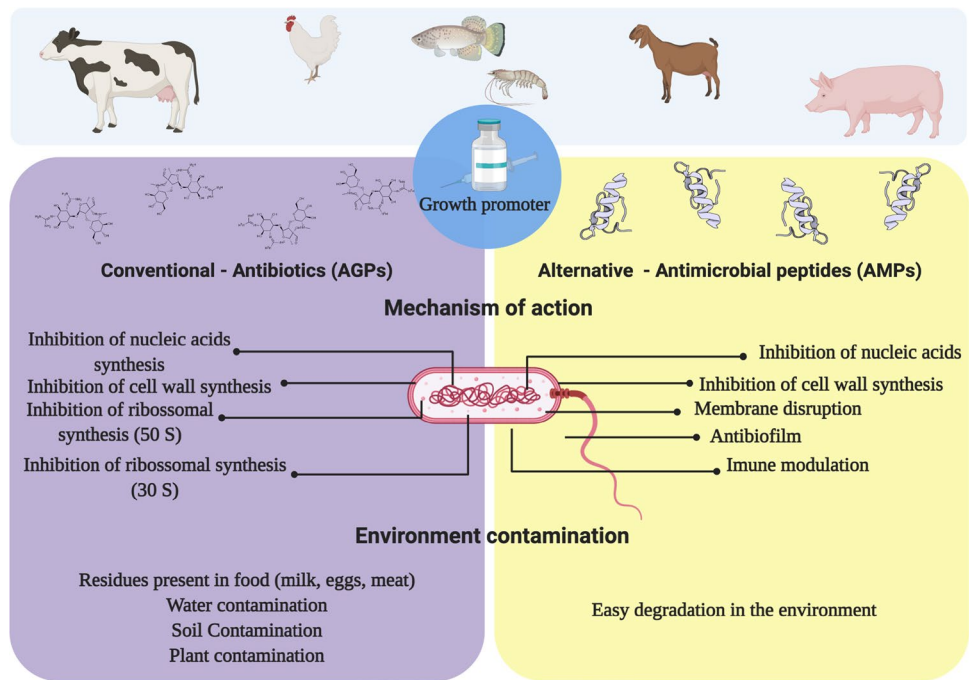
As mentioned, the most common growth promoters applied in livestock are antibiotics (AGPs). However, the excessive

use of antimicrobials has already contributed to the emergence of global public health problems, such as antimicrobial resistance, hypersensitivity responses, and damage to normal bacterial biota (Ronquillo and Hernandez 2017; Bacanlı and Başaran 2019). Thus, concern about the risks associated with AGPs in livestock and the consequences for human health has been increasing (Hughes and Heritage 2004; Marquardt and Li 2018; Tona 2018; Ma et al. 2020).

AMR development generally occurs through mutations (vertical AMR acquisition) and gene horizontal transfer (horizontal AMR acquisition) (Nadeem et al. 2020; Vidovic and Vidovic 2020). Occurrence of vertical AMR acquisition may be related with the exposure of bacterial populations to antibiotics, even in low concentrations. In this situation, any mutation that confers partial or full resistance against these antibiotics can be positively selected and transferred to subsequent generations, and this resistance may be against a specific antibiotic or a whole class (Tenover 2006; Davies and Davies 2010; Vidovic and Vidovic 2020). Horizontal AMR acquisition may occur by transference of resistant genetic elements such as plasmids or transposons, by horizontal transfer such as conjugation, transduction, or other mechanisms. Such processes can provide resistance against several antibiotic classes (Fig. 1) (Tenover 2006; Medina et al. 2020; Nadeem et al. 2020; Vidovic and Vidovic 2020).

Some studies estimated that approximately 90% of antibiotics applied in farm animals can be excreted by urine and/or feces. Furthermore, a number of antibiotics applied

Fig. 1 Mechanism of action of antibiotics and antimicrobial peptides used as growth promoters



in animals can be stored in tissues such as muscle, milk, eggs, and fat. These residues can be dispersed by water and waste-water systems, or by fertilizers that employ manure, and cause the contamination of soil and consequently affect the soil microbiota, water, and plants (Ronquillo and Hernandez 2017; Iwu et al. 2020). Humans can be contaminated by AGPs in different ways, by ingestion in food, including milk, eggs, and meat, or by consuming water with residues of AGPs (Ben et al. 2019; Kraemer et al. 2019).

In this context, regulatory agencies, such as the European Commission and the US Food and Drug Administration (FDA), have established limits for antibiotic residues in animal foods (European Commission 1998; FDA 2016; FAO 2018). In 2006, the use of AGPs was completely banned in the European Union countries, and the use of antibiotics is permitted only for veterinary purposes (ten Doeschate and Raine 2006). In recent years, countries such as Canada, India, China, and Malaysia have restricted the use of AGPs in livestock, but many countries do not present any formal restrictions on AGPs (Brown et al. 2017; Ronquillo and Hernandez 2017; Salim et al. 2018; Bacanlı and Başaran 2019; Ben et al. 2019).

Thus, in recent years, the use of AGPs has been reduced and gradually replaced by effective dietary supplements, such as probiotics and/or prebiotics, enzymes, and novel antimicrobial peptides. In addition, the application of antibiotics in livestock requires control and prudence (Ma et al. 2020; Magnusson 2020; Patel et al. 2020).

Antimicrobial peptide growth promoters

Antimicrobial peptides (AMPs) display a broad spectrum of activity against bacteria, fungi, viruses, and cancer, and these characteristics have already been widely discussed (Cardoso et al. 2020; Hwang et al. 2011; Rodrigues et al. 2019; Saïdo-Sakanaka et al. 2004). Normally, the activity of AMPs can be related to bacterial membrane interaction. This interaction can occur associated with ion channel/pore formation and/or detergent-like effect, indicating the molecular basis of their attraction to membranes (Brogden 2005; Nguyen et al. 2011). Furthermore, AMPs display different modes of action, like membrane disruption, increased membrane permeability, and/or disturbances in key cellular processes by interacting with intracellular targets (Yeaman and Yount 2003; Nguyen et al. 2011; Wimley and Hristova 2011; Sani and Separovic 2016). Besides, the use of AMPs as a growth promoter has demonstrated beneficial effects on nutrient digestibility, the intestinal microbiota, intestinal morphology, and immune function activities (Fig. 1) (Liu et al. 2008; Xiao et al. 2015; Gadde et al. 2017).

In this regard, studies using lactoferricin-lactoferrin (LF-chimera) were used to supplement piglet feed. The results demonstrated an increase in body weight and in the average daily gain (ADG) of 13.3 and 29.3%, respectively, compared with pigs fed control diets (Tang et al. 2012). Other studies tested the growth and digestive capacity after administering AMPs in poultry and pigs (Wang et al. 2016). The use of synthetic AMPs, such as AMP-A3 and AMP-P5 (both derived from the amino acid substitution of *Helicobacter pylori*-HP and cecropin-magainin2 fusion,

Table 2 Antimicrobial peptides as growth promoters

AMP	Source	Target bacteria	Animal	Reference
Microcin J25	<i>Escherichia coli</i>	<i>E. coli</i> , <i>Salmonella sp.</i>	Broilers	Wang et al. 2020
Pediocin A	<i>Pediococcus pentosaceus</i>	<i>Clostridium perfringens</i>	Broilers	Grilli et al. 2009
Plectasin	<i>Pseudoplectania nigrella</i>	<i>E. coli</i> , <i>Salmonella sp.</i>	Broilers	Ma et al. 2019
RSRP	<i>Oryctolagus cuniculus-sacculus rotundus</i>	<i>E. coli</i>	Broilers	Aguirre et al. 2015
Lactoferrin (bLf)	<i>Bos taurus</i>	<i>E. coli</i> , <i>Salmonella sp.</i>	Broilers	Cao et al. 2007; Tang et al. 2012; Messaoudi et al. 2012
SMXD51	<i>Lactobacillus salivarius</i>	<i>Campylobacter jejuni</i>	Poultry	Ceotto-Vigoder et al. 2016; Kogut et al. 2013
BT	<i>Brevibacillus texasporus</i>	<i>Salmonella enterica serovar Enteritidis</i>	Neonatal poultry	Kogut et al. 2012

respectively), increased the efficiency of gain of weaning pigs and broilers, with additional benefits concerning nutrient uptake and intestinal morphology. The maximal AMP concentrations tested were 90 and 60 mg kg⁻¹ for AMP-A3 and AMP-P5, respectively. The results showed the effect for both body weight increases and intestinal injury reduction (Choi et al. 2013; Yoon et al. 2012, 2013, 2014). Another study evaluated the response of pig anti-bacterial peptides (PABP) in growth performance and small intestine mucosal immune responses in broilers. The authors reported that this PABP added to drinking water (20 and 30 mg/L) or supplemented in feed (150 and 200 mg/kg) can enhance growth performance, raise the intestinal ability to absorb nutrients, and improve the mucosal immunity of the intestine (Bao et al. 2009).

A different group used the AMP *Epinephelus lanceolatus piscidin* (EP), heterologously expressed and used as a dietary supplement for *Gallus gallus domesticus*. Treatment groups included control, and EP supplemented the diet at different doses (0.75, 1.5, 3.0, 6.0, and 12%). The results indicated that EP supplementation increased *G. domesticus* weight gain, feed efficiency, IL-10, and IFN- γ production, when compared to control (Tai et al. 2020). The pediocin A from *Pediococcus pentosaceus* FBB61 was tested by Grilli et al. (2009) as feed supplementation, and also tested against the *Clostridium perfringens* proliferation in broilers. The authors used a control group and another group where 80 AU.g⁻¹ of pediocin A was added to the feed. The broilers were fed for 21 days, and they were challenged with culture of *C. perfringens* type A, which was administered by mouth on days 14, 15, and 16, twice daily (106 cfu/broiler). According to the authors, supplementation with pediocin A increased broiler growth performance during the challenge with *C. perfringens* (Grilli et al. 2009).

AMPs can also be used as AGPs in aquaculture, as described by Gyan et al. (2020). The application of AMPs can enhance the innate immune system, and boosts growth performance and disease resistance in Pacific white leg shrimp. In this study, different concentrations of AMP were tested in feed supplementation (0% until 1%). The results demonstrated the optimum concentration of AMP is 0.4% (400 g/kg). Researchers also observed that excess AMP in supplementation negatively affected the growth performance and immune system of the shrimp. Other studies which demonstrate efficient results using AMPs as growth promoters in livestock are shown in Table 2.

Concluding remarks and prospects

In the last 70 years, AGPs have been synonymous with productivity in livestock farming. However, the extensive use of these growth promoters has contributed to the development of bacterial strains with antimicrobial resistance. Antimicrobial resistance represents a worldwide problem and is treated with concern by the WHO. Hence, the European Union and the USA have limited the use of antibiotics in animal production. In an attempt to maintain livestock production, studies using AMPs as growth promoters have taken place, showing effective results in animal weight gains, and in some cases improving host immunity.

AMPs can therefore be an excellent way to substitute antibiotics due to characteristics such as a lower risk of inducing antimicrobial resistance, good inhibitory effects, and ease of degradation. Further studies using AMPs will allow a better understanding of the effects on the gastrointestinal ecosystem, and this will enable the best use of antimicrobial peptides for economic efficiency and sustainable livestock production.

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Declarations

Ethical statement This article does not contain any studies with human participants or animals performed by any of the authors.

Conflict of interest The authors declare no competing interests.

References

- Ageitos JM, Sánchez-Pérez A, Calo-Mata P, Villa TG (2017) Antimicrobial peptides (AMPs): ancient compounds that represent novel weapons in the fight against bacteria. *Biochem Pharmacol* 133:117–138. <https://doi.org/10.1016/j.bcp.2016.09.018>
- Aguirre ATA, Acda SP, Angeles AA, Oliveros MCR, Merca FE, Cruz FA (2015) Effect of bovine lactoferrin on growth performance and intestinal histologic features of broiler. *Philipp J Vet Anim Sci* 41(1)
- Aslam B, Wang W, Arshad MI, Khurshid M, Muzammil S, Rasool MH, Nisar MA, Alvi RF, Aslam MA, Qamar MU, Salamat MKF, Baloch Z (2018) Antibiotic resistance: a rundown of a global crisis. *Infect Drug Resist* 11:1645. <https://doi.org/10.2147/IDR.S173867>
- Bacanlı M, Başaran N (2019) Importance of antibiotic residues in animal food. *Food Chem Toxicol* 125:462–466. <https://doi.org/10.1016/j.fct.2019.01.033>
- Bao H, She R, Liu T, Zhang Y, Peng KS, Luo D, Yue Z, Ding Y, Hu Y, Liu W, Zhai L (2009) Effects of pig antibacterial peptides on growth performance and intestine mucosal immune of broiler chickens. *Poult Sci J* 88(2):291–297. <https://doi.org/10.3382/ps.2008-00330>
- Ben Y, Fu C, Hu M, Liu L, Wong MH, Zheng C (2019) Human health risk assessment of antibiotic resistance associated with antibiotic residues in the environment: a review. *Environ Res* 169:483–493. <https://doi.org/10.1016/j.envres.2018.11.040>
- Brennan J, Skinner J, Barnum DA, Wilson J (2003) The efficacy of bacitracin methylene disalicylate when fed in combination with narasin in the management of necrotic enteritis in broiler chickens. *Poult Sci J* 82(3):360–363. <https://doi.org/10.1093/ps/82.3.360>
- Brogden KA (2005) Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nat Rev Microbiol* 3:238–250. <https://doi.org/10.1038/nrmicro1098>
- Brown K, Uwiera RR, Kalmokoff ML, Brooks SP, Inglis GD (2017) Antimicrobial growth promoter use in livestock: a requirement to understand their modes of action to develop effective alternatives. *Int J Antimicrob Agents* 49(1):12–24. <https://doi.org/10.1016/j.ijantimicag.2016.08.006>
- Cao LT, Wu JQ, Xie F, Hu SH, Mo Y (2007) Efficacy of nisin in treatment of clinical mastitis in lactating dairy cows. *J Dairy Sci* 90(8):3980–3985. <https://doi.org/10.3168/jds.2007-0153>
- Cardoso MH, Orozco RQ, Rezende SB, Rodrigues G, Oshiro KG, Cândido ES, Franco OL (2020) Computer-aided design of antimicrobial peptides: are we generating effective drug candidates? *Front Microbiol* 10:3097. <https://doi.org/10.3389/fmicb.2019.03097>
- Ceotto-Vigoder H, Marques SLS, Santos INS, Alves MDB, Barrias ES, Potter A, Alviano DS, Bastos MCF (2016) Nisin and lysostaphin activity against preformed biofilm of *Staphylococcus aureus* involved in bovine mastitis. *J Appl Microbiol* 121(1):101–114. <https://doi.org/10.1111/jam.13136>
- Cheng G, Hao H, Xie S, Wang X, Dai M, Huang L, Yuan Z (2014) Antibiotic alternatives: the substitution of antibiotics in animal husbandry. *Front Microbiol* 5:69–83. <https://doi.org/10.3389/fmicb.2014.00217>
- Choi SC, Ingale SL, Kim JS, Park YK, Kwon IK, Chae BJ (2013) Effects of dietary supplementation with an antimicrobial peptide-P5 on growth performance, nutrient retention, excreta and intestinal microflora and intestinal morphology of broilers. *Anim Feed Sci Technol* 185(1–2):78–84. <https://doi.org/10.1016/j.anifeeds.2013.07.005>
- Davies J, Davies D (2010) Origins and evolution of antibiotic resistance. *Microbiol Mol Biol Rev* 74(3):417–433. <https://doi.org/10.1128/MMBR.00016-10>
- European Commission (1998) Agricultural situation and prospects in the central and eastern countries – Poland, directorate general for agriculture brussels
- FAO (2018) World Livestock: Transforming the livestock sector through the sustainable development goals. Rome. 222 pp. Licence: CC BY-NC-SA 3.0 IGO
- FDA U (2016) NARMS Integrated Report: 2014-The National Antimicrobial Resistance Monitoring System: Enteric Bacteria
- Feighner SD, Dashkevich MP (1987) Subtherapeutic levels of antibiotics in poultry feeds and their effects on weight gain, feed efficiency, and bacterial cholytaurine hydrolase activity. *Appl Environ Microbiol* 53(2):331–336
- Founou LL, Founou RC, Essack SY (2016) Antibiotic resistance in the food chain: a developing country-perspective. *Front Microbiol* 7:1881. <https://doi.org/10.3389/fmicb.2016.01881>
- Gadde U, Kim WH, Oh ST, Lillehoj HS (2017) Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry: a review. *Anim Health Res Rev* 18(1):26–45. <https://doi.org/10.1017/S1466252316000207>
- Gyan WR, Yang Q, Tan B, Jan SS, Jiang L, Chi S, Dong X, Liu H, Shuang Z (2020) Effects of antimicrobial peptides on growth, feed utilization, serum biochemical indices and disease resistance of juvenile shrimp, *Litopenaeus vannamei*. *Aquac Res* 51(3):1222–1231. <https://doi.org/10.1111/are.14473>
- Gomes B, Augusto MT, Felício MR, Hollmann A, Franco OL, Gonçalves S, Santos NC (2018) Designing improved active peptides for therapeutic approaches against infectious diseases. *Biotechnol Adv* 36(2):415–429. <https://doi.org/10.1016/j.biotechadv.2018.01.004>
- George BA, Quarle CL, Fagerberg DJ (1982) Virginiamycin effects on controlling necrotic enteritis infections in chickens. *Poult Sci J* 61:447–450. <https://doi.org/10.3382/ps.0610447>
- Grilli E, Messina MR, Catelli E, Morlacchini M, Piva A (2009) Pediocin A improves growth performance of broilers challenged with *Clostridium perfringens*. *Poult Sci J* 88(10):2152–2158. <https://doi.org/10.3382/ps.2009-00160>
- Hancock RE, Sahl HG (2006) Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies. *Nat Biotechnol* 24(12):1551–1557. <https://doi.org/10.1038/nbt1267>
- Hancock RE (2001) Cationic peptides: effectors in innate immunity and novel antimicrobials. *The Lancet Infect Dis* 1(3):156–164. [https://doi.org/10.1016/S1473-3099\(01\)00092-5](https://doi.org/10.1016/S1473-3099(01)00092-5)

- Hancock R, Chapple D (1999) MINIREVIEW Peptide antibiotics. *Antimicrob Agents Chemother* 1317–1323. <https://doi.org/10.1128/AAC.43.6.1317>
- Hugues P, Heritage J (2004) Antibiotic growth-promoters in food animals. *FAO Animal Production and Health Paper* 129–152
- Huyghebaert G, Ducatelle R, Van Immerseel F (2011) An update on alternatives to antimicrobial growth promoters for broilers. *Vet J* 187(2):182–188. <https://doi.org/10.1016/j.tvjl.2010.03.003>
- Hwang B, Hwang JS, Lee J, Lee DG (2011) The antimicrobial peptide, psacothasin induces reactive oxygen species and triggers apoptosis in *Candida albicans*. *Biochem Biophys Res Commun* 405:267–271. <https://doi.org/10.1016/j.bbrc.2011.01.026>
- Innes GK, Randad R, Korinek A, Davis MF, Price LB, So AD, Heaney CD (2020) External societal costs of antimicrobial resistance in humans attributable to antimicrobial use in livestock. *Annu Rev Public Health* 41. <https://doi.org/10.1146/annurev-publhealth-040218-043954>. Accessed in 10 January 2021
- Iwu CD, Korsten L, Okoh AI (2020) The incidence of antibiotic resistance within and beyond the agricultural ecosystem: a concern for public health. *Microbiol Open* 9(9):e1035. <https://doi.org/10.1002/mbo3.1035>
- Jensen BB (1998) The impact of feed additives on the microbial ecology of the gut in young pigs. *J Anim Feed Sci* 7:45–64. <https://doi.org/10.22358/jafs/69955/1998>
- Jenssen H, Hamill P, Hancock RE (2006) Peptide antimicrobial agents. *Clin Microbiol Rev* 19(3):491–511. <https://doi.org/10.1128/CMR.00056-05>
- Jones FT, Ricke SC (2003) Observations on the history of the development of antimicrobials and their use in poultry feeds. *Poult Sci J* 82(4):613–617. <https://doi.org/10.1093/ps/82.4.613>
- Knarreborg A, Lauridsen C, Engberg RM, Jensen SK (2004) Dietary antibiotic growth promoters enhance the bioavailability of alpha-tocopheryl acetate in broilers by altering lipid absorption. *J Nutr* 134:1487–1492. <https://doi.org/10.1093/jn/134.6.1487>
- Kogut MH, Genovese KJ, He H, Swaggerty CL, Jiang YW (2012) BT cationic peptides: small peptides that modulate innate immune responses of chicken heterophils and monocytes. *Vet Immunol Immunopathol* 145(1–2):151–158. <https://doi.org/10.1016/j.vetimm.2011.10.023>
- Kogut MH, Genovese KJ, He H, Swaggerty CL, Jiang Y (2013) Modulation of chicken intestinal immune gene expression by small cationic peptides as feed additives during the first week posthatch. *Clin Vaccine Immunol* 20(9):1440–1448. <https://doi.org/10.1128/CVI.00322-13>
- Kraemer SA, Ramachandran A, Perron GG (2019) Antibiotic pollution in the environment: from microbial ecology to public policy. *Microorganisms* 7(6):180. <https://doi.org/10.3390/microorganisms7060180>
- Li Z, Hu Y, Yang Y, Lu Z, Wang Y (2018) Antimicrobial resistance in livestock: antimicrobial peptides provide a new solution for a growing challenge. *Anim Front* 8(2):21–29. <https://doi.org/10.1093/af/vfy005>
- Liu T, She R, Wang K, Bao H, Zhang Y, Luo D, Hu Y, Ding Y, Wang D, Peng K (2008) Effects of rabbit sacculus rotundus antimicrobial peptides on the intestinal mucosal immunity in chickens. *Poult Sci J* 87(2):250–254. <https://doi.org/10.3382/ps.2007-00353>
- Ma JL, Zhao LH, Sun DD, Zhang J, Guo YP, Zhang ZQ, Ma QG, Ji C, Zhao LH (2020) Effects of dietary supplementation of recombinant plectasin on growth performance, intestinal health and innate immunity response in broilers. *Probiotics Antimicrob Proteins* 1–10. <https://doi.org/10.1007/s12602-019-9515-2>
- Magnusson U (2020) Prudent and effective antimicrobial use in a diverse livestock and consumer's world. *J Anim Sci* 98:S4–S8. <https://doi.org/10.1093/jas/skaa148>
- Marquardt RR, Li S (2018) Antimicrobial resistance in livestock: advances and alternatives to antibiotics. *v* 8(2):30–37. <https://doi.org/10.1093/af/vfy001>
- Medina MJ, Legido-Quigley H, Hsu LY (2020) Antimicrobial resistance in one health. In: *Global health security*. Springer, pp 209–229. https://doi.org/10.1007/978-3-030-23491-1_10
- Messaoudi S, Madi A, Prévost H, Feuilloley M, Manai M, Dousset X, Connil N (2012) *In vitro* evaluation of the probiotic potential of *Lactobacillus salivarius* SMXD51. *Anaerobe* 18(6):584–589. <https://doi.org/10.1016/j.anaerobe.2012.10.004>
- Nadeem SF, Gohar UF, Tahir SF, Mukhtar H, Pornpukdeewattana S, Nukthamna P, Moula Ali AM, Bavisetty SCB, Massa S (2020) Antimicrobial resistance: more than 70 years of war between humans and bacteria. *Crit Rev Microbiol* 46(5):578–599. <https://doi.org/10.1080/1040841X.2020.1813687>
- National Office of Animal Health (NOAH) (2001) Antibiotics for animals. <http://www.noah.co.uk/issues/antibiotics.htm>. Accessed 18 Jan 2021
- Nguyen LT, Haney EF, Vogel HJ (2011) The expanding scope of antimicrobial peptide structures and their modes of action. *Trend Biotechnol* 29(9):464–472. <https://doi.org/10.1016/j.tibtech.2011.05.001>
- Oshiro KG, Rodrigues G, Monges BED, Cardoso MH, Franco OL (2019) Bioactive peptides against fungal biofilms. *Front Microbiol*. <https://doi.org/10.3389/fmicb.2019.02169>
- Patel SJ, Wellington M, Shah RM, Ferreira MJ (2020) Antibiotic stewardship in food-producing animals: challenges, progress, and opportunities. *Clin Ther*. <https://doi.org/10.1016/j.clinthera.2020.07.004>
- Ronquillo MG, Hernandez JCA (2017) Antibiotic and synthetic growth promoters in animal diets: review of impact and analytical methods. *Food Control* 72:255–267. <https://doi.org/10.1016/j.foodcont.2016.03.001>
- Rodrigues G, Silva GGO, Buccini DF, Duque HM, Dias SC, Franco OL (2019) Bacterial proteinaceous compounds with multiple activities toward cancers and microbial infection. *Front Microbiol* 10. <https://doi.org/10.3389/fmicb.2019.01690>
- Saido-Sakanaka H, Ishibashi J, Momotani E, Amano F, Yamakawa M (2004) *In vitro* and *in vivo* activity of antimicrobial peptides synthesized based on the insect defensin. *Peptides* 25:19–27. <https://doi.org/10.1016/j.peptides.2003.12.009>
- Salim HM, Huque KS, Kamaruddin KM, Haque Beg A (2018) Global restriction of using antibiotic growth promoters and alternative strategies in poultry production. *Sci Prog* 101(1):52–75. <https://doi.org/10.3184/003685018X15173975498947>
- Sani MA, Separovic F (2016) How membrane-active peptides get into lipid membranes. *Acc Chem Res* 49(6):1130–1138. <https://doi.org/10.1021/acs.accounts.6b00074>
- Shai Y (2002) Mode of action of membrane active antimicrobial peptides. *Pep Sci* 66:236–248. <https://doi.org/10.1002/bip.10260>
- Snyder DL, Wostmann BS (1987) Growth rate of male germfree Wistar rats fed *ad libitum* or restricted natural ingredient diet. *Lab Anim Sci* 37:320–325
- Spohn R, Daruka L, Lázár V, Martins A, Vidovics F, Grézal G, Méhi O, Kintsés B, Számel M, Jangir PK, Csörgő B, Györkei Á, Bódi Z, Faragó A, Bodai L, Földesi I, Kata D, Maróti G, Pap B, Wirth R, Papp B, Pál C (2019) Integrated evolutionary analysis reveals antimicrobial peptides with limited resistance. *Nat Commun* 10(1):1–13. <https://doi.org/10.1038/s41467-019-12364-6>
- Tai HM, Huang HN, Tsai TY, You MF, Wu HY, Rajanbabu V, Chang HY, Pan CY, Chen JY (2020) Dietary supplementation of recombinant antimicrobial peptide *Epinephelus lanceolatus piscidin* improves growth performance and immune response in *Gallus gallus domesticus*. *PLoS One* 15(3):e0230021. <https://doi.org/10.1371/journal.pone.0230021>

- Tang X, Fatufe AA, Yin Y, Tang Z, Wang S, Liu Z, Li TJ (2012) Dietary supplementation with recombinant lactoferrampin-lactoferricin improves growth performance and affects serum parameters in piglets. *J Anim Vet Adv* 11:2548–2555
- ten Doeschate RAHM, Raine H (2006) History and current use of feed additives in the European Union: legislative and practical aspects. *Avian Gut Funct Health Dis* 28:1
- Tenover FC (2006) Mechanisms of antimicrobial resistance in bacteria. *Am J Med* 119(6):S3–S10
- Tona GO (2018) Current and future improvements in livestock nutrition and feed resources. *Anim Husbandry Nutr* 147–169. <https://doi.org/10.5772/intechopen.73088>
- U.S. Food and Drug Administration. 2015. Veterinary feed directive. Available at: <https://www.fda.gov/AnimalVeterinary/ucm071807.htm>. Accessed 15 Jan 2021.
- Van Boeckel TP, Brower C, Gilbert M, Grenfell BT, Levin SA, Robinson TP, Teillant A, Laxminarayan R (2015) Global trends in antimicrobial use in food animals. *Proc Natl Acad Sci USA* 112(18):5649–5654. <https://doi.org/10.1073/pnas.1503141112>
- Vidovic N, Vidovic S (2020) Antimicrobial resistance and food animals: Influence of livestock environment on the emergence and dissemination of antimicrobial resistance. *Antibiotics* 9(2):52. <https://doi.org/10.3390/antibiotics9020052>
- Xiao H, Shao F, Wu M, Ren W, Xiong X, Tan B, Yin Y (2015) The application of antimicrobial peptides as growth and health promoters for swine. *J Anim Sci Biotechnol* 6(1):1–6. <https://doi.org/10.1186/s40104-015-0018-z>
- Ward BP, Ottaway NL, Perez-Tilve D, Ma D, Gelfanov VM, Tschöp MH, DiMarchi RD (2013) Peptide lipidation stabilizes structure to enhance biological function. *Mol Metabolism* 2(4):468–479. <https://doi.org/10.1016/j.molmet.2013.08.008>
- Wang G, Song Q, Huang S, Wang Y, Cai S, Yu H, Ding X, Zeng X, Zhang J (2020) Effect of antimicrobial peptide microcin J25 on growth performance, immune regulation, and intestinal microbiota in broiler chickens challenged with *Escherichia coli* and *Salmonella sp.* *Animals (Basel)* 10(2). <https://doi.org/10.3390/ani10020345>
- Wang S, Zeng X, Yang Q, Qiao S (2016) Antimicrobial peptides as potential alternatives to antibiotics in food animal industry. *Int J Mol Sci* 17(5):603. <https://doi.org/10.3390/ijms17050603>
- Wimley WC, Hristova K (2011) Antimicrobial peptides: successes, challenges and unanswered questions. *J Membr Biol* 239(1):27–34. <https://doi.org/10.1007/s00232-011-9343-0>
- World Health Organization (2014) WHO's first global report on antibiotic resistance Reveals serious, worldwide threat to public health. Available from: <http://www.who.int/mediacentre/news/releases/2014/amr-report/en/>. Accessed 10 Jan 2021
- Zhao Y, Zhang M, Qiu S, Wang J, Peng J, Zhao P, Zhu R, Wang H, Li Y, Wang K, Yan W, Wang R (2016) Antimicrobial activity and stability of the D-amino acid substituted derivatives of antimicrobial peptide polybia-MPI. *AMB Express* 6:122. <https://doi.org/10.1186/s13568-016-0295-8>
- Yeaman MR, Yount NY (2003) Mechanisms of antimicrobial peptide action and resistance. *Pharmacol Rev* 55(1):27–55. <https://doi.org/10.1124/pr.55.1.2>
- Yoon JH, Ingale SL, Kim JS, Kim KH, Lee SH, Park YK, Lee SC, Kwona IK, Chae BJ (2014) Effects of dietary supplementation of synthetic antimicrobial peptide-A3 and P5 on growth performance, apparent total tract digestibility of nutrients, fecal and intestinal microflora and intestinal morphology in weanling pigs. *Livest Sci* 159:53–60. <https://doi.org/10.1016/j.livsci.2013.10.025>
- Yoon JH, Ingale SL, Kim JS, Kim KH, Lee SH, Park YK, Kwona IK, Chae BJ (2012) Effects of dietary supplementation of antimicrobial peptide-A3 on growth performance, nutrient digestibility, intestinal and fecal microflora and intestinal morphology in weanling pigs. *Anim Feed Sci Technol* 177(1–2):98–107. <https://doi.org/10.1016/j.anifeedsci.2012.06.009>
- Yoon JH, Ingale SL, Kim JS, Kim KH, Lohakare J, Park YK, Park JC, Kwon IK, Chae BJ (2013) Effects of dietary supplementation with antimicrobial peptide-P5 on growth performance, apparent total tract digestibility, faecal and intestinal microflora and intestinal morphology of weanling pigs. *J Sci Food Agric* 93(3):587–592. <https://doi.org/10.1002/jsfa.5840>

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