MINI-REVIEW



Recent advances and new strategies on leishmaniasis treatment

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Abstract

Leishmaniasis is one of the most important tropical neglected diseases according to the World Health Organization. Even after more than a century, we still have few drugs for the disease therapy and their great toxicity and side effects put in check the treatment control program around the world. Moreover, the emergence of strains resistant to conventional drugs, co-infections such as HIV/*Leishmania* spp., the small therapeutic arsenal (pentavalent antimonials, amphotericin B and formulations, and miltefosine), and the low investment for the discovery/development of new drugs force researchers and world health agencies to seek new strategies to combat and control this important neglected disease. In this context, the aim of this review is to summarize new advances and new strategies used on leishmaniasis therapy addressing alternative and innovative treatment paths such as physical and local/topical therapies, combination or multi-drug uses, immunomodulation, drug repurposing, and the nanotechnology-based drug delivery systems.

Key points

- The treatment of leishmaniasis is a challenge for global health agencies.
- Toxicity, side effects, reduced therapeutic arsenal, and drug resistance are the main problems.
- New strategies and recent advances on leishmaniasis treatment are urgent.
- Immunomodulators, nanotechnology, and drug repurposing are the future of leishmaniasis treatment.

Keywords Leishmaniasis \cdot Conventional chemotherapy \cdot Multi-drug therapy \cdot Immunomodulators \cdot Drug repurposing \cdot Nanotechnology

Introduction

Leishmaniasis constitutes a group of human and animal diseases caused by *Leishmania*, a protozoan parasite from the *Trypanosomatidae* family. More than 20 *Leishmania* species,

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around the world, are known to be transmitted to humans by the bite of infected phlebotomine sandflies during their blood meal. Distinct species of *Leishmania* spp. cause different clinical manifestations of the disease, which can be characterized by at least three syndromes: visceral leishmaniasis (VL, also known as kala-azar); cutaneous leishmaniasis (CL); and mucocutaneous leishmaniasis (MCL) (Burza et al. 2018). Postkala-azar dermal leishmaniasis (PKDL) constitutes a complication from visceral leishmaniasis after treatment, with macular, maculopapular, and nodular rash as clinical manifestations. These diseases mainly affect the poorest regions where health services are the most precarious (Alvar et al. 2006).

Since the report of the first cases of leishmaniasis associated with infection by the human immunodeficiency virus (HIV) in Europe, Mediterranean, Brazil, and East African countries, an increasing number of co-infection cases have been described (Desjeux 1995). There are regions in Ethiopia where approximately 40% of leishmaniasis patients are co-infected with HIV (Alvar et al. 2008). Facing this scenario, the WHO Department of Neglected Tropical Diseases

Table 1 Conventional strategies for leishmaniasis treatment recommended by the World Health Organization

Visceral leishmaniasis treatment

Anthroponotic visceral leishmaniasis caused by L. donovani in the Indian subcontinent

Liposomal amphotericin B: dose of 3-5 mg/kg daily by infusion given over 3-5 days period (total dose of 15 mg/kg) or 10 mg/kg as a single dose by infusion

Visceral leishmaniasis caused by L. donovani in East Africa

Pentavalent antimonials (20 mg Sb^v/kg/day intramuscularly or intravenously) + paromomycin (15 mg (11 mg base)/kg/day intramuscularly), both for 17 days

Visceral leishmaniasis caused by L. infantum: Mediterranean Basin, Middle East, Central Asia, South America

Pentavalent antimonials (20 mg Sb^v/kg/day intramuscularly or intravenously) for 28 days

Cutaneous leishmaniasis treatment*

Old World leishmaniasis

L. major, L. tropica, L. aethiopica**, and L. infantum**

Local therapy: 15% paromomycin/12% methylbenzethonium chloride ointment twice daily for 20 days or intralesional antimonials (1–5 ml per session) every 3–7 days (1–5 sessions)

Systemic therapy: pentavalent antimonials (as above) for 10-20 days

L. aethiopica

Systemic therapy: pentavalent antimonials (as above) for 10–20 days + paromomycin (as above) for 60 days or longer to treat diffuse cutaneous leishmaniasis (C)

New World Leishmaniasis

L. mexicana, L. guyanensis, L. panamensis, L. braziliensis, L. amazonensis, L. peruviana, L. venezuelensis Local therapy: 15% paromomycin/12% methylbenzethonium chloride ointment twice daily for 20 days Systemic therapy: pentavalent antimonials (as above) for 20 days

Mucocutaneous leishmaniasis

All species***

Pentavalent antimonials (as above) for 30 days

*The most used treatment strategy for CL. However, the treatment of CL varies from observation, locality, local, or systemic therapy depending on gravity of lesions, etiological species, and its potential to develop into mucosal leishmaniasis

**Few data are available on therapy for CL caused by L. infantum and L. aethiopica

***Few data are available for the therapy of MCL due to *L. aethiopica*

brought together a group of experts aiming to develop evidence-based guidelines for the treatment of co-infection HIV-VL in East Africa and Southeast Asia, since treatment of leishmaniasis in HIV-infected patients is a special condition with reduced therapeutic options (WHO 2020).

In CL, despite being less severe than VL, in untreated patients, the lesions may worsen due to secondary infections and ulcerations. Another complication is the disfiguring scars and psychosocial suffering due to the stigma and social isolation caused by the disease (Kassi et al. 2008; Bailey et al. 2017). The lesions in CL are usually self-healing; however, they can lead to mucosal involvement or diffuse form, depending on the species of parasite, immunological status, and genetically determined responses of patients (Reithinger et al. 2007).

Since the beginning of the treatment of VL and CL with pentavalent antimonials, resistance of the population to the treatment has been observed, due to its great toxicity and side effects. These factors impair the patient's adherence to treatment and in turn select resistant strains of the parasite. The improvement of amphotericin B deoxycholate in liposomal form reduced the side effects; however, the financial impact in poor regions is worrying. Other drugs have been developed, among them miltefosine; however, there are resistant strains, and in the tegumentary form of the disease, the response varies according to the species of the parasite. In this sense, it is important to develop new treatment strategies for the affected population. Thus, the present review addresses important points in conventional therapies and therapeutic advances in the present and future of leishmaniasis treatment.

Conventional antileishmanial therapy

Pentavalent antimonials

Pentavalent antimonial (Sb^v) compounds have been available for almost seven decades and constitute the first-line treatment against leishmaniasis. The two main formulations are meglumine antimoniate (MA) and sodium stibogluconate (SSG). Sb^v would behave as a prodrug that is reduced within the organism into active Sb^{III} form, a borderline metal ion with high affinity towards nitrogen- and sulfhydryl-containing

ligands. The antileishmanial mechanisms of Sb^{III} are probably related to its interaction with sulfhydryl-containing biomolecules, including thiols, peptides, proteins, and enzymes (Frézard et al. 2009). Although effective, the use of these compounds is many times limited by the need for daily parenteral administration, severe toxicity and side effects, and treatment failures. With emerging resistance to this drug in Bihar, alternative treatment strategy has been adopted for these areas. In the eastern Africa, the combination of SSG and paromomycin has been recommended as first-line drug for VL treatment (Table 1) (Musa et al. 2012; Kimutai et al. 2017). In cutaneous forms, Sb^{v} has shown varied responses in different etiological species. Despite that, they are the mainstays of treatment for CL in several regions of the world. Currently, in Old World cutaneous leishmaniasis (OWCL), most patients are treated with intralesional Sb^v infiltration. Systemic use can be considered for only mucocutaneous lesions (Table 1). In New World cutaneous leishmaniasis (NWCL), most patients are treated with systemic MA (20 mg of $Sb^{v}/kg/day$ for 20 days), even though it has already been recommended the use of intralesional MA has already been recommended by the Pan American Health Organization (PAHO). Side effects of Sb^v include local irritation, cardiac and hepatic alterations, anorexia, nausea, and vomiting (Sundar and Chakravarty 2015). Further, HIV/leishmaniasis co-infected-treated patients present pronounced side effects, high failure and relapse rates, and increased mortality during Sb^v therapy (Cota et al. 2013).

Amphotericin B deoxycholate and liposomal amphotericin B

Amphotericin B deoxycholate (AmB-D) is a broad-spectrum antifungal and active against protozoan Leishmania species. Following the loss of effectiveness of antimonial drugs in India, AmB-D was elected as the first-line drug of choice for VL. This drug binds to membrane sterols with particular affinity towards ergosterol, forming complexes that arrange into ion channels and increase membrane permeability (Gray et al. 2012). Despite its efficacy, the treated patients present frequent adverse effects such as fever, hypokalemia, myocarditis, and mainly nephrotoxicity, which demand hospital monitoring (Sundar et al. 2007a, b). Many lipid formulations of amphotericin were developed in recent years to minimize the side effects of AmB-D. But the liposomal amphotericin B (AmB-L—AmBisome®) is the only one approved by US Food and Drug Administration (FDA). The total dose requirements of lipid formulations for treatment of VL vary by region (Sundar et al. 2007a, b) (Table 1). Although it has expensive cost, the high effectiveness of AmB-L ensures that it is the first treatment choice for HIV co-infection patients, pregnant, and transplanted individuals in most countries (Sundar and Chakravarty 2013). In cutaneous forms, AmB-D treatment shows at least equivalent results to Sb^{v} treatment. Therefore, it is not considered an alternative treatment due to considerably serious side effects. Thus, there are only a few studies with AmB-D, as well as the liposomal form in CL and MCL patients, with absent randomized clinical trials (Wortmann et al. 2010; Balasegaram et al. 2012).

Miltefosine

The antileishmanial activity of miltefosine, an antineoplastic drug, was first identified in the 1980s and until today is the only oral drug approved for leishmaniasis treatment (Croft et al. 1987; Sundar and Chakravarty 2015). This drug is able to kill parasites in vitro and in vivo by changing signaling pathways and cell membrane synthesis, thus leading to apoptosis (Verma and Dey 2004). Following a phase III clinical trial, in which 50-100 mg/day dose for 28 days resulted in a cure rate of 94%, miltefosine was first registered for VL treatment in India (Sundar et al. 2002; Sundar et al. 2008a, b). Unfortunately, after a decade of use, there was an increase in resistance with reduced effectiveness doubling the relapse rate (Sundar et al. 2012; Dorlo et al. 2014; Srivastava et al. 2017). This caused the exchange of miltefosine by single-dose AmB-L as therapeutic strategy of VL elimination program in the India subcontinent (Table 1). Currently, its use has been recommended only in combinations with other drugs in this region (Chakravarty and Sundar 2019; van Griensven and Diro 2019). In other countries, the studies with miltefosine have shown varied performance demonstrating the need of higher drug doses (Ritmeijer et al. 2006; Wasunna et al. 2016). For cutaneous leishmaniasis, the recommended dose is 2.5 mg/kg/ day orally for 28 days. However, miltefosine has varied species-dependent effectiveness. A clinical trial demonstrated that miltefosine is useful against CL caused by Leishmania panamensis in Colombia but not against CL caused by Leishmania braziliensis in Guatemala (Soto et al. 2004). On the other hand, Machado et al. (2010) observed that miltefosine therapy was more effective and safer than standard Sb^v for the treatment of CL caused by L. braziliensis in Bahia, Brazil. Adverse effects of miltefosine include gastrointestinal alterations and less often hepatotoxicity. Moreover, it presents teratogenic effects, so women of child-bearing age are advised contraception during the treatment regimen (Sundar and Chakravarty 2015).

Paromomycin

Paromomycin, an aminoglycoside antibiotic that interferes in protein synthesis and modifies mitochondrial membrane fluidity inhibiting respiration, has also been shown to be an effective antileishmanial agent. This drug is used in topical and systemic treatment for CL and as a systemic drug for VL in some regions of the Indian continent (Sundar and Chakravarty 2008; Jhingran et al. 2009). Paromomycin was licensed by the Indian government for VL treatment in 2006, and a phase IV trial demonstrated cure rates of 94% (Sinha et al. 2011). However, paromomycin has limited effectiveness as monotherapy in East Africa (Hailu et al. 2010; Musa et al. 2012). Adverse effects of systemic therapy are uncommon and include pain and burning on application site, low hepatotoxicity, and ototoxicity (Sundar et al. 2007b). Paromomycin has been evaluated in topical formulations with variable clinical results in OW and NW Leishmania species. In general, the efficacy of topical paromomycin did not differ from that of intralesional Sb^v in the OWCL, whereas its efficacy was inferior to parenteral Sb^v in the NWCL (Kim et al. 2009). For CL cases, the decision to use local or systemic drugs usually depends on the lesion characteristic, etiological species, and imminent risk of progressing to mucocutaneous disease (PAHO 2018) (Table 1).

Other antileishmanial therapies

Pentamidine

Since it was first synthesized in the 1940s, the pentamidine has been studied for its anti-Leishmania action, mainly after the cases of antimonial resistance in India subcontinent (Jha et al. 1991). However, its use was abandoned due to toxicity, such as pancreatitis leading to diabetes mellitus, hypoglycemia, hypotension, cardiac alterations, and hyperkalemia (Sundar and Chakravarty 2015). Recently, the monthly use of 4 mg/kg dose for 12 months has been recommended as a secondary prophylaxis HIV-VL co-infected patients in Ethiopia, if the T CD4 lymphocyte count is below 200 cells/ µl (Diro et al. 2015; Diro et al. 2019). Despite scarce studies demonstrating the use of pentamidine in OWCL, its use has been demonstrated in NWCL. However, the effectiveness of these studies varies according the Leishmania species, duration and composition of therapy, and geographic region (Soto et al. 1994; Lai et al. 2002; Paula et al. 2003; Roussel et al. 2006; Amato et al. 2009; Soto et al. 2016; Christen et al. 2018; Gadelha et al. 2018).

Azoles

Azole antifungal agents have been evaluated as therapeutic strategies, mainly for cutaneous forms. In OWCL, the oral fluconazole therapy presented cure rates of approximately 60%, with substantial increase in effectiveness when the drug concentration was doubled. However, this treatment was associated with side effects, such as hepatotoxicity and cardiotoxicity (Alrajhi et al. 2002; Emad et al. 2011). High-dose oral fluconazole therapy in *L. braziliensis*-infected patients demonstrated cure rate of 75–100% (Sousa et al. 2011).

However, a randomized clinical trial evaluated the efficacy of oral fluconazole, in a similar therapeutic scheme, for the CL treatment caused by *L. braziliensis*, evidencing cure rates of only 22% (Prates et al. 2017). Itraconazole demonstrated low effectiveness for the MCL treatment in Brazil, and the complete resolution of MCL lesions was observed in only three (23%) high-dose itraconazole-treated patients in Ecuador (Amato et al. 2000; Calvopina et al. 2004). A recent systematic meta-analysis identified that none of the azoles is effective for the treatment of cutaneous forms of leishmaniasis, and its use should be associated only with other drugs (Galvão et al. 2017).

New strategies and recent advances on leishmaniasis treatment

Emergence of resistant strains to conventional drugs (Sundar 2000; Mueller et al. 2007), high toxicity, co-infections such as HIV/*Leishmania* spp., the small therapeutic arsenal available for treatment of the disease, and the low investment for the discovery/development of new drugs force researchers and world health agencies to seek new strategies to combat and control this important neglected disease. In this sense, the following section is a brief summary of recent advances and new strategies used to treat leishmaniasis (Table 2).

Cutaneous and mucocutaneous leishmaniasis

Physical and local therapies

 $\ensuremath{\text{CO}}_2$ laser administration and thermotherapy Based on the principle to directly destroy the Leishmania parasites, the CO₂ laser and thermotherapy are a simple way to deliver external heat on infected tissues, causing damage to specific areas with parasitism (Asilian et al. 2004; Valencia et al. 2013). The direct application of heat can accelerate the cure of the cutaneous lesions (Navin et al. 1990). In OWCL, some studies have demonstrated that thermotherapy showed better results in relation to cure rate compared with intralesional treatment with antimonials with similar or reduced side effects (Sadeghian et al. 2007; Aronson et al. 2010). An improvement was described using a CO₂ laser thermotherapy technology, demonstrating in a clinical trial 93.7% of cure rate compared with combined therapy using intralesional antimonials (78% of cure rate) (Shamsi et al. 2011). For CL, these strategies have been used with relative success, both for old and new world infections.

Cryotherapy Cryotherapy also known as cryosurgery was firstly evaluated in Saudi Arabia patients infected with *Leishmania major* and obtained 100% of cure using a CO_2 cryomachine (Bassiouny et al. 1992). Nowadays, cryotherapy

Advance/strategy	Disease	Benefits compare to conventional therapy	Treatment efficacy	References
Thermotherapy and CO ₂ laser administration	Cutaneous and mucocutaneous leishmaniasis	Reduction of side effects and accelerated cure of the cutaneous lesions	Marked	Sadeghian et al. (2007); Aronson et al. (2010); Asilian et al. (2004); Valencia et al. (2013); Shamsi et al. (2011)
Cryotherapy and electrotherapy	Cutaneous and mucocutaneous leishmaniasis	Reduction of side effects and accelerated cure of the cutaneous lesions	Moderate	Bassiouny et al. (1992); Mosleh et al. (2008); Negera et al. (2012); Chakravarty and Sundar (2019)
Topical drug therapy using nitric oxide derivates	Cutaneous and mucocutaneous leishmaniasis	Accelerated heal of the cutaneous lesions and reduction of side effects	Moderate/low	Lopez-Jaramillo et al. (1998); Lopez-Jaramillo et al. (2010)
Intralesional drug administration	Cutaneous and mucocutaneous leishmaniasis	Reduction of side effects and relapses, accelerated heal of the cutaneous lesions, high percentage of cure, and low cost	Marked	Uzun et al. (2004); Bashir et al. (2019); Ramalho et al. (2018); Arboleda et al. (2019); Masmoudi et al. (2006)
Multi-drug or combination therapy	Cutaneous, mucocutaneous, and visceral leishmaniasis	Reduction of side effects and relapses, high percentage of cure, low risk of developing resistant parasites, and low cost	Marked	Sundar et al. (2008b); Melaku et al. (2007); Mahajan et al. (2015)
Immunomodulators	Cutaneous, mucocutaneous, and visceral leishmaniasis	Reduction of side effects and relapses, high percentage of cure, and low risk of developing resistant parasites	Marked	Sundar and Murray (1995); Squires et al. (1993); Convit et al. (2003); Badaro et al. (1999); Sundar et al. (1994); Mastroianni (2004); Monjour et al. (1994); Mayrink et al. (2006)
Nanotechnology	Cutaneous, mucocutaneous, and visceral leishmaniasis	Reduction of side effects and relapses, high percentage of cure, and low risk of developing resistant parasites	In vitro and in vivo testing	Sazgarnia et al. (2013); Jebali and Kazemi (2013); Costa Lima et al. (2012); De Mattos et al. (2015); Italia et al. (2012); Kumar et al. (2019)
Drug repurposing	Cutaneous, mucocutaneous, and visceral leishmaniasis	Reduction of side effects and relapses, high percentage of cure, and low risk of developing resistant parasites	In silico, in vitro, and in vivo testing	Bustamante et al. (2019); Braga (2019)

uses liquid nitrogen (at -195 °C), and applied once or twice weekly in *Leishmania* lesions, it can achieve an efficiency rate over 95% (Mosleh et al. 2008; Negera et al. 2012). The mechanism of killing parasites was described by the formation of ice intracellularly causing the disruption of cells leading to localized ischemic necrosis. The secondary side effects were mainly associated with edema and erythema at the site, hyperor hypopigmentation (Chakravarty and Sundar 2019).

Electrotherapy Electric field stimulation, a non-drug treatment, has been described as a potential tool to control microbial infection. Previous test using mice infected with *Pseudomonas aeruginosa* resulted in significantly inhibiting bacteria in lung infections (Giladi et al. 2010). Authors have shown that exposure to electrical currents could lead to healing the skin lesions and intractable ulcers and the new skin produced has better tensile properties compared with that skin produced naturally (Wolcott et al. 1969). In leishmaniasis, the use of therapeutic electricity applied on infected mice with *L. major* showed important death of parasites at the lesion sites (Hejazi et al. 2004). Recent advances have demonstrated that electric fields affected *Leishmania tarentolae* promastigote motility, clumping, and viability in vitro (Dorsey et al. 2018). However, these studies are in an initial phase requiring more data related to what currents, potentials, numbers of stimulations, and durations are safe but effective for clinical use against CL.

Topical drug therapies

Nitric oxide derivates This is a promise strategy used against some *Leishmania* species from Americas (*L. braziliensis* and *L. panamensis*). A *S*-nitroso-*N*-acetylpenicillamine (SNAP) cream, compound that generates NO, was firstly tested for 10 days showing, after 30 days of treatment, the healing of all lesions and the formation of new skin in patients presenting CL (Lopez-Jaramillo et al. 1998). Moreover, a most recent study using a topical nanofiber nitric oxide (NO)-releasing patch administered for 12 h a day for 20 days showed ineffectiveness, with only 37.1% of cure rates in CL Colombian patients (Lopez-Jaramillo et al. 2010). The authors suggest

that therapeutic failure can be reversed by increasing drug concentration or treatment time.

Intralesional drug administration Thinking of reducing the adverse effects while maintaining efficacy and safety of the form of conventional use (intramuscular or intravenous infusion), this new way to treat CL using pentavalent antimonials has gained prominence. Since 2013, the use of intralesional antimonial therapy has been recommended by the PAHO guidelines (OPS 2013) when systemic treatment is not indicated or if local treatment for CL is required. Intralesional pentavalent antimonials achieved cure rates over 90% against Asia and Mediterranean species of the parasite (L. major and Leishmania tropica) (Uzun et al. 2004; Bashir et al. 2019). For new world Leishmania species, open-label phase II clinical trials showed elevated cure rates (87-91.6%) after 180 days of treatment (Ramalho et al. 2018; Arboleda et al. 2019). The most common side effects observed were bacterial secondary infection, erythema, local itching, and pain during administration, which tend to disappear few days after the end of the treatment (Masmoudi et al. 2006; Ramalho et al. 2018).

Visceral leishmaniasis

Multi-drug or combination therapy

It is well known that untreated symptomatic VL is almost always fatal. Moreover, it is observed a large range on the variability in the effectiveness of antileishmanial drugs associated with the region where the *Leishmania* infection occurred and the host immune status (van Griensven and Diro 2019). In this sense, combined therapy has the following objectives: shortening the treatment duration (reducing side effects and improved adherence to the regimen by the patient), controlling the development of parasite resistance, lowering the costs, and encouraging a cure, especially in complicated cases of VL (Monge-Maillo and López-Vélez 2013).

The combined therapy is mainly recommended for patients who had not responded to monotherapy with Sb^v. Most of the studies were conducted in India especially in Leishmania donovani-infected patients. Using AmB-L at 5 mg/kg/day in a single dose followed by different regimens of miltefosine, the authors demonstrated greater efficacy with combinations compared with AmB-L at 5 mg/kg/day in a single dose (98% versus 91%) (Sundar et al. 2008b). In Eastern Africa, some studies have demonstrated that the combination of paromomycin with SSG increased the cure rate response in comparison with SSG as monotherapy (Melaku et al. 2007). A recent study in VL/HIV co-infection demonstrated reduced rates of mortality and VL relapse when AmB-L (AmBisome) and miltefosine (Impavido) were combined and administered. Moreover, the authors concluded that combination therapy appeared to be well tolerated, safe, and

effective and may be considered as an important option for treatment of VL in HIV co-infected patients (Mahajan et al. 2015).

Immunomodulators

Leishmania parasites have stated systematic resistance against the immune system manipulating different mechanisms to survive into the host. In this way, treatments with substances that promote the restoration of the immune response against the parasite are an alternative approach to combat the infection (Musa et al. 2010; Roatt et al. 2014; Taslimi et al. 2018).

The IFN- γ is well recognized as a cytokine capable of inducing macrophages to kill *Leishmania* parasites. In VL patients, the use of IFN- γ as immunotherapy promoted accelerated parasitological control (Squires et al. 1993; Sundar and Murray 1995) and enhanced the clinical efficacy of conventional Sb^v therapy, promoting more than 80% cure rate (Badaro et al. 1999; Sundar et al. 1994; Sundar and Murray 1995). More recently, in a case report on HIV/VL co-infection in Italy, the combination treatment using rHuGM-CSF (recombinant human granulocyte macrophage colony-stimulating factor) showed to be effective leading to a reduction on the spleen size, disappearance of symptoms, and clinical cure of the patient (Mastroianni 2004).

Taking into account the use of immunomodulators, the most used strategy in leishmaniasis is therapeutic vaccines. In literature, many studies around the world describe important results obtained using vaccines as immunotherapeutic tool. More than 5000 patients were treated against CL with heat-killed Leishmania parasites plus BCG in Venezuela with an incredible 95.7% of cure rate achieved (Convit et al. 2003). In the same way, patients infected with L. braziliensis were treated with a therapeutic vaccine composed of parasitederived antigen Fraction 2 (LbbF2) that promoted secretion of key cytokines by T cells leading to clinical cure of the infected patients (Monjour et al. 1994). In Brazil, more than 500 patients with CL were treated either with pentavalent antimony, killed Leishmania vaccine plus BCG, BCG, or a combination. The cure rates in therapeutic vaccine or pentavalent antimony chemotherapy were the same, but with fewer adverse effects and shorter recovery time (Mayrink et al. 2006). As observed in these studies, activation of the immune system through immunotherapy associated with application of antileishmanial drugs can solve the complicated cases of the disease mainly in patients with drug refractory Leishmania infections.

Nanotechnology

Recently, nanotechnology-based drug delivery systems have advanced to efficiently deliver different types of drugs to specific tissues and cells. In leishmaniasis, the combination of nanocarrier system with antileishmanial drugs enables targeted and efficient delivery improving the on-target effect, bioavailability, reducing toxicity, and side effects (Jebali and Kazemi 2013; Wagner et al. 2019). In this horizon, new advances in development of nanostructured lipid carriers, liposomes, nanoemulsions, solid-lipid nanoparticles, niosomes, nanocapsules, nanoparticles, metallic nanosphere, polymeric nanoparticles, and nanostructured layered films have been done to trigger efficient drug delivery for leishmaniasis treatment (Costa Lima et al. 2012; Italia et al. 2012; De Mattos et al. 2015; Kumar et al. 2019; Wagner et al. 2019; Saleem et al. 2019).

Various nanocarrier strategies have been used for leishmaniasis treatment demonstrating their own advantages and disadvantages. As it is known, the main cell targets in leishmaniasis are macrophages. Thence, the most employed nanoparticles in the disease are polymeric nanoparticles and liposomes because of the easy and fast way of internalization by the infected cells. Liposomes are nanocarriers that have unique properties to load and deliver hydrophobic and/or hydrophilic molecules by surface activation. Moreover, because they are positively charged, liposomes are promptly internalized by the macrophages. Due to these properties, they are the most commonly nanocarriers employed in leishmaniasis (Saleem et al. 2019).

Nanoemulsions are considered one of the best drug delivery systems due to their simple preparation, their ability to solubilize hydrophobic drugs, and their physicochemical stability over several years, and they can be easily scaled-up (Bilia et al. 2014). Similarly, polymeric nanoparticles are also widely used, besides having small size, capacity to deliver more than one drug, and present low toxicity, and they are cost effective. The main characteristics of nanoparticles are the possibility of functionalizing their surface and the use of components with excellent biodegradation capacity and biocompatibility (poly lactide-co-glycolide (PLGA)) (Saleem et al. 2019). Metal-based nanoparticle systems also offer significant advantages as drug delivery against leishmaniasis including enlargement of the biological action of the carried molecule and decrease in side effects. Moreover, nanostructure metal compounds show enhanced selectivity for parasite biomolecules, like cysteine protease that is a promising family of Leishmania enzymes. It is important to note that many nanotherapeutic agents have been approved by the FDA and will be available for clinical use as soon as possible (Eifler and Thaxton 2011). Indeed, for leishmaniasis treatment, most of the nanostructured drugs are still undergoing in vitro and in vivo testing. Only few nanostructures of amphotericin B are available for human use until now.

A recent study of in vitro and preclinical evaluation of amine functionalized carbon-based nanoparticle loaded with AmB (f-Comp-AmB) showed a remarkable antileishmanial efficacy in comparison to AmB-D. In golden Syrian hamsters' model, the authors observed 98% of suppression of the parasite replication in the spleen in comparison with the untreated animals (Gedda et al. 2020). In CL, the development of an effective topical drug formulation is desirable mainly by its non-invasive nature, which may potentially enhance the treatment accessibility and patient adherence. In this sense, the use of nanotechnology has brought great advances. In a preclinical study, a combination of amphotericin B (AmB) and miltefosine (MTF) co-loaded in ultra-deformable liposomes (SGUDLs) was evaluated. An important antileishmanial activity of co-loaded SGUDLs was observed against amastigotes of Leishmania mexicana and the in vivo study demonstrated a significant reduction in the parasitic burden in BALB/c model of CL (Dar et al. 2020). The authors suggest that the great efficacy of the co-loaded SGUDLs is directly associated with synergistic interaction between AmB and MTF and concluded that AmB-MTF co-loaded SGUDLs could be an effective topical treatment option against CL (Dar et al. 2020).

Drug repurposing

Drug repurposing is regarded as one of the most important strategy for the rational use of drugs, especially against neglected diseases. Called as drug reprofiling or repositioning is a modern strategy to identify and develop new uses for existing drugs (Ashburn and Thor 2004). In addition, repositioning has main advantages, including lower risk of drug failure, reduced time frame for development/application of the drug, and reduced costs, and can reveal new pathways and targets that may be further explored (Pushpakom et al. 2019).

Focusing on leishmaniasis drug repurposing, computational approaches are the main strategies that have been applied with relative success among the diseases. These computational techniques involve systematic analysis of any data type such as bioinformatics targeting gene expression, chemical structure, and genotype or proteomic data. In this sense, molecular docking is a computational strategy to predict binding sites between the ligands (drugs, for example) and the target (a receptor) (Bustamante et al. 2019). In conventional docking, one receptor/protein target is chosen so that multiple drugs could be tested against that target. In this case, the knowledge about the target/protein or the drug class may favor the choice of a possible ideal drug with a greater chance of success in subsequent trials (Pushpakom et al. 2019). Another important tool is called signature matching that is based on the comparison between the characteristic (signature) of a drug or molecule compared with that of another drug or molecule (Keiser et al. 2009). The signature analyses are derived from some general data such as metabolomic, proteomic, transcriptomic, or chemical structures. These two drug-repositioning tools are the most used strategies against leishmaniasis currently.



In a recent study, Bustamante et al. (2019) used bioinformatic predictions to detect some repurposing drugs for leishmaniasis treatment. In this study, the authors performed some simulations to identify and predict these drugs with in vitro validations and pharmacokinetic simulations. As strategy, the bioinformatic predictions were used to detect potential homologs between targeted proteins by approved drugs and other proteins of the *Leishmania* spp. parasites. In this study, 33 drugs were identified with potential target prediction with in vitro action (rifabutin and perphenazine) (Bustamante et al. 2019).

Metallodrugs have been identified with important antitumor, anti-inflammatory, and antimicrobial actions. Auranofin (Ridaura), a gold(I) triethylphosphine thiosugar drug, has been described as having antileishmanial activity with the ability to inhibit trypanothione reductase (TR). In this sense, a recent study performed a preclinical evaluation of gold(I) triphenylphosphine- and triethylphosphine-based complexes showing their activity against *Leishmania infantum* and *L. braziliensis* intracellular amastigotes (Tunes et al. 2020). Using bioimaging, the authors observed reduced lesion size and parasite burden in BALB/c mice infected with luciferaseexpressing *L. braziliensis* or *Leishmania amazonensis* and orally treated with gold(I) complexes. According to the authors, the gold(I) complexes are promising antileishmanial agents, with a potential for therapeutic use (Tunes et al. 2020).

An interesting review discussing drug repurposing has recently been published and describes multi-target drugs active against leishmaniasis (Braga 2019). In this review, it is shown that the azoles presented growth inhibitory activity against both fungi and *Leishmania*. Some compounds such as posaconazole, fluconazole, and itraconazole act against the same target, lanosterol 14-a-demethylase enzyme. Similarly, it is shown that both amphotericin B and miltefosine act on small molecules, proteins, genes, and even organelles showing their profile of multi-target agents as known. The authors concluded that some steps towards drug repurposing for multi-target strategy will be the future in the search for leishmanicidal drug candidates (Braga 2019) (Table 2).

Conclusions and prospects

There are few drugs for the leishmaniasis treatment, and the great toxicity and side effects put in check the international treatment control. Moreover, the emergence of resistant strains to conventional drugs, co-infections such as HIV/Leishmania spp., the small therapeutic arsenal, and the low investment for the discovery/development of new drugs force researchers and world health agencies to seek new strategies to combat and control this important neglected disease. In this context, new strategies with important advances in physical and local therapies including thermotherapy and CO₂ laser administration and topical drug therapies using NO compounds and intralesional drug administration have given a better perspective of cure in patients with CL. Moreover, the use of combination therapy or multi-drug therapy and activation of immune system using immunomodulators have helped to solve problems in relation to parasitic resistance and serious cases of HIV/Leishmania spp. infection. Finally, being considered as the future of leishmaniasis treatment, the drug repurposing and the nanotechnology-based drug delivery systems bring the opportunity to use computational tools for the identification of existing drugs which are used in the treatment of the disease with less time, cost, and using nanotechnology that promotes an efficient delivery of different types of drugs to specific tissues and cells infected by the Leishmania parasites (Fig. 1). Thus, efforts need to be directed for the rational investment in new therapies and treatment strategies against the disease, in order to seek therapies with less side effects, lower costs, and better efficacy against these parasites.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethical statement This article does not contain any studies with human participants or animals performed by any of the authors.

References

- Alvar J, Yactayo S, Caryn B (2006) Leishmaniasis and poverty. Trends Parasitol 22(12):552–557. https://doi.org/10.1016/j.pt.2006.09.004
- Alvar J, Aparicio P, Aseffa A, Den Boer M (2008) The relationship between leishmaniasis and AIDS: the second 10 years. Clin Microbiol Rev 21(2):334–359. https://doi.org/10.1128/CMR. 00061-07
- Alrajhi AA, Ibrahim EA, De Vol EB, Kharait M, Faris RM, Maguires JH (2002) Fluconazole for the treatment of cutaneous leishmaniasis caused by *Leishmania major*. N Engl J Med 346(12):891–895. https://doi.org/10.1056/NEJMoa011882
- Amato VS, Padilha ARS, Nicodemo AC, Duarte MIS, Valentini M, Uip DE, Boulos M, Neto VA (2000) Use of itraconazole in the treatment of mucocutaneous leishmaniasis: a pilot study. Int J Infect Dis 4(3): 153–157. https://doi.org/10.1016/s1201-9712(00)90077-8
- Amato VS, Tuon FF, Imamura R, Camargo RA, Duarte MI, Neto VA (2009) Mucosal leishmaniasis: description of case management approaches and analysis of risk factors for treatment failure in a cohort of 140 patients in Brazil. J Eur Acad Dermatol Venereol 23(9): 1026–1034. https://doi.org/10.1111/j.1468-3083.2009.03238.x
- Arboleda M, Barrantes S, Úsuga LY, Robledo SM (2019) Successful treatment of cutaneous leishmaniasis with intralesional meglumine antimoniate: a case series. Rev Soc Bras Med Trop 16(52): e20180211. https://doi.org/10.1590/0037-8682-0211-2018
- Aronson NE, Wortmann GW, Byrne WR, Howard RS, Bernstein WB, Marovich MA, Polhemus ME, Yoon IK, Hummer KA, Gasser RA Jr, Oster CH, Benson PM (2010) A randomized controlled trial of local heat therapy versus intravenous sodium stibogluconate for the treatment of cutaneous *Leishmania major* infection. PLoS Negl Trop Dis 4:e628. https://doi.org/10.1371/journal.pntd.0000628
- Ashburn TT, Thor KB (2004) Drug repositioning: identifying and developing new uses for existing drugs. Nat Rev Drug Discov 3(8):673– 683. https://doi.org/10.1038/nrd1468
- Asilian A, Sharif A, Faghihi G, Enshaeieh S, Shariati F, Siadat AH (2004) Evaluation of CO laser efficacy in the treatment of cutaneous leishmaniasis. Int J Dermatol 43:736–738. https://doi.org/10.1111/j. 1365-4632.2004.02349.x
- Badaro R, Falcoff E, Badaro FS, Carvalho EM, Pedral-Sampaio D, Barral A, Carvalho JS, Barral-Neto M, Brandely M, Silva L, Bina JC, Teixeira R, Falcoff R, Rocha H, Ho JL, Johnson WD (1999) Treatment of visceral leishmaniasis with pentavalent antimony and interferon gamma. N Engl J Med 322(1):16–21. https://doi.org/10. 1056/NEJM199001043220104

- Bailey F, Mondragon-Shem K, Hotez P, Ruiz-Postigo JA, Al-Salem W, Acosta-Serrano A, Molyneux DH (2017) A new perspective on cutaneous leishmaniasis—implications for global prevalence and burden of disease estimates. PLoS Neg Trop Dis 11:e0005739. https://doi.org/10.1371/journal.pntd.0005739
- Balasegaram M, Ritmeijer K, Lima MA, Burza S, Genovese GO, Milani B, Gaspani S, Potet J, Chappuis F (2012) Liposomal amphotericin B as a treatment for human leishmaniasis. Expert Opin Emerg Drugs 17(4):493–510. https://doi.org/10.1517/14728214.2012.748036
- Bashir U, Tahir M, Anwar MI, Manzoor F (2019) Comparison of intralesional meglumine antimoniate along with oral itraconazole to intralesional meglumine antimoniate in the treatment of cutaneous leishmaniasis. Pak J Med Sci 35(6):1669–1673. https://doi.org/10. 12669/pjms.35.6.363
- Bassiouny A, El Meshad M, Talaat M, Kutty K, Metawaa B (1992) Cryosurgery in cutaneous leishmaniasis. Br J Dermatol 107:467– 474. https://doi.org/10.1111/j.1365-2133.1982.tb00390.x
- Bilia AR, Guccione C, Isacchi B, Righeschi C, Firenzuoli F, Bergonzi MC (2014) Essential oils loaded in nanosystems: a developing strategy for a successful therapeutic approache. Evid Based Complement Alternat Med:651593. https://doi.org/10.1155/2014/651593
- Braga SS (2019) Multi-target drugs active against leishmaniasis: a paradigm of drug repurposing. Eur J Med Chem 1(183):111660. https:// doi.org/10.1016/j.ejmech.2019.111660
- Burza S, Croft SL, Boelaert M (2018) Leishmaniasis. Lancet. 392(10151):951–970. https://doi.org/10.1016/S0140-6736(18) 31204-2
- Bustamante C, Ochoa R, Asela C, Muskus C (2019) Repurposing of known drugs for leishmaniasis treatment using bioinformatic predictions, *in vitro* validations and pharmacokinetic simulations. J Comput Aided Mol Des 33(9):845–854. https://doi.org/10.1007/ s10822-019-00230-y
- Calvopina M, Guevara AG, Armijos RX, Hashiguchi Y, Davidson RN, Cooper PJ (2004) Itraconazole in the treatment of New World mucocutaneous leishmaniasis. Int J Dermatol 43(9):659–663. https:// doi.org/10.1111/j.1365-4632.2004.02183.x
- Chakravarty J, Sundar S (2019) Current and emerging medications for the treatment of leishmaniasis. Expert Opin Pharmacother 20(10):1251– 1265. https://doi.org/10.1080/14656566.2019.1609940
- Christen JR, Bourreau E, Demar M, Lightburn E, Couppié P, Ginouvés M, Prévot G, Gangneux JP, Savini H, de Laval F, Santi VP, Briolant S (2018) Use of the intramuscular route to administer pentamidine isethionate in *Leishmania guyanensis* cutaneous leishmaniasis increases the risk of treatment failure. Travel Med Infect Dis 24:31– 36. https://doi.org/10.1016/j.tmaid.2018.02.010
- Convit J, Ulrich M, Zerpa O, Borges R, Aranzazu N, Valera M, Villarroel H, Zapata Z, Tomedes I (2003) Immunotherapy of American cutaneous leishmaniasis in Venezuela during the period 1990-99. Trans R Soc Trop Med Hyg 4:469–472. https://doi.org/10.1016/S0035-9203(03)90093-9
- Costa Lima AS, Resende M, Silvestre R, Tavares J, Ouaissi A, Lin PKT, Cordeiro-da-Silva A (2012) Characterization and evaluation of BNIPDaoct-loaded PLGA nanoparticles for visceral leishmaniasis: *in vitro* and *in vivo* studies. Nanomedicine 7(12):1839–1849. https:// doi.org/10.2217/nnm.12.74
- Cota GF, de Sousa MR, Fereguetti TO, Rabello A (2013) Efficacy of antileishmania therapy in visceral leishmaniasis among HIV infected patients: a systematic review with indirect comparison. PLoS Negl Trop Dis 7(5):e2195. https://doi.org/10.1371/journal.pntd.0002195
- Croft SL, Neal RA, Pendergast W, Chan JH (1987) The activity of alkyl phosphorylcholines and related derivatives against *Leishmania donovani*. Biochem Pharmacol 36:2633–2636. https://doi.org/10. 1016/0006-2952(87)90543-0
- Dar MJ, Khalid S, McElroy CA, Satoskar AR, Khan GM (2020) Topical treatment of cutaneous leishmaniasis with novel amphotericin Bmiltefosine co-incorporated second generation ultra-deformable

liposomes. Int J Pharm 5(573):118900. https://doi.org/10.1016/j. jpharm.2019.118900

- De Mattos CB, Argenta DF, de Lima Melchiades G, Cordeiro MNS, Tonini ML, Moraes MH, Weber TN, Roman SS, Nunes RJ, Teixeira HF, Steindel M, Koester LS (2015) Nanoemulsions containing a synthetic chalcone as an alternative for treating cutaneous leishmaniasis: optimization using a full factorial design. Int J Nanomedicine 10:5529–5542. https://doi.org/10.2147/IJN.S83929
- Desjeux P (1995) Leishmania / HIV co-infections. Afr Health 18(1):20–22
- Diro E, Ritmeijer K, Boelaert M, Alves F, Mohammed R, Abongomera C, Ravinetto R, de Crop M, Fikre H, Adera C, Colebunders R, van Loen H, Menten J, Lynen L, Hailu A, Griensven v (2015) Use of pentamidine as secondary prophylaxis to prevent visceral leishmaniasis relapse in HIV infected patients, the first twelve months of a prospective cohort study. PLoS Negl Trop Dis 9(10):e0004087. https://doi.org/10.1371/journal.pntd.0004087
- Diro E, Edwards T, Ritmeijer K, Fikre H, Abongomera C, Kibret A, Bardonneau C, Soipei P, Mutinda B, Omollo R, van Griensen J, Zijlstra EE, Wassuna M, Alves F, Alvar J, Hailu A, Alexander N, Blesson S (2019) Long term outcomes and prognostics of visceral leishmaniasis in HIV infected patients with use of pentamidine as secondary prophylaxis based on CD4 level: a prospective cohort study in Ethiopia. PLoS Negl Trop Dis 13(2):e0007132. https:// doi.org/10.1371/journal.pntd.0007132
- Dorlo TP, Rijal S, Ostyn B, Vries PJ, Singh R, Bhattarai N, Uranw S, Dujardim JC, Boelaert M, Beihnen JH, Huitema ADR (2014) Failure of miltefosine in visceral leishmaniasis is associated with low drug exposure. J Infect Dis 210(1):146–153. https://doi.org/ 10.1093/infdis/jiu039
- Dorsey BM, Cass CL, Cedeño DL, Vallejo R, Jones MA (2018) Effects of specific electric field stimulation on the release and activity of secreted acid phosphatases from *Leishmania tarentolae* and implications for therapy. Pathogens 7(4):77. https://doi.org/10.3390/ pathogens7040077
- Eifler AC, Thaxton CS (2011) Nanoparticle therapeutics: FDA approval, clinical trials, regulatory pathways, and case study. Biomed Nanotechnol Methods Mol Biol 726:325–338. https://doi.org/10. 1007/978-1-61779-052-221
- Emad M, Hayati F, Fallahzadeh MK, Namazi MR (2011) Superior efficacy of oral fluconazole 400 mg daily versus oral fluconazole 200 mg daily in the treatment of cutaneous *Leishmania major* infection: a randomized clinical trial. J Am Acad Dermatol 64(3):606– 608. https://doi.org/10.1016/j.jaad.2010.04.014
- Frézard F, Demicheli C, Ribeiro RR (2009) Pentavalent antimonials: new perspectives for old drugs. Molecules 14(7):2317–2336. https://doi. org/10.3390/molecules14072317
- Gadelha EPN, Ramasawmy R, Oliveira BC, Rocha NM, Guerra JAO, Silva GAR, Mesquita TGR, Cortez CCT, Talhari AC (2018) An open label randomized clinical trial comparing the safety and effectiveness of one, two or three weekly pentamidine isethionate doses (seven milligrams per kilogram) in the treatment of cutaneous leishmaniasis in the Amazon Region. PLoS Negl Trop Dis 12(10): e0006850. https://doi.org/10.1371/journal.pntd.0006850
- Galvão EL, Rabello A, Cota GF (2017) Efficacy of azole therapy for tegumentary leishmaniasis: a systematic review and meta-analysis. PLoS One 12(10):e0186117. https://doi.org/10.1371/journal.pone. 0186117
- Gedda MR, Madhukar P, Vishwakarma AK, Verma V, Kushwaha AK, Yadagiri G, Mudavath SL, Singh OP, Srivastava ON, Sundar S (2020) Evaluation of safety and antileishmanial efficacy of amine functionalized carbon-based composite nanoparticle appended with amphotericin B: an *in vitro* and preclinical study. Front Chem 8(510). https://doi.org/10.3389/fchem.2020.00510
- Giladi M, Porta Y, Blatt A, Shmueli E, Wasserman Y, Kirson ED (2010) Microbial growth inhibition by alternating electric fields in mice

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with *Pseudomonas aeruginosa* lung infection. Antimicrob Agents Chemother 54(8):3212–3218. https://doi.org/10.1128/AAC.01841-09

- Gray KC, Palacios DS, Dailey I, Endo MM, Uno BE, Wilcock BC, Burle MD (2012) Amphotericin primarily kills yeast by simply binding ergosterol. Proc Natl Acad Sci U S A 109(7):2234–2239. https://doi. org/10.1073/pnas.1117280109
- Hailu A, Musa A, Wasunna M, Balasegaram M, Yifu S, Mengistu G, Hurissa Z, Hailu W, Weldegebreal T, Tesfaye S, Makonnen E, Khalil E, Ahmed O, Fadlalla A, El-Hassan A, Raheem M, Mueller M, Koummuki Y, Rashid J, Mbui J, Mucee G, Njoroge S, Manduku V, Musibi A, Matuma G, Kirui F, Lodenyo H, Mutea D, Kirigi G, Edwards T, Smith P, Muthami L, Royce C, Ellis S, Alobo M, Omollo R, Kesusu J, Owiti R, Kinuthia J (2010) Geographical variation in the response of visceral leishmaniasis to paromomycin in East Africa: a multicentre, open-label, randomized trial. PLoS Negl Trop Dis 4(10):e709. https://doi.org/10.1371/journal.pntd.0000709
- Hejazi H, Eslami G, Dalimi A (2004) The parasiticidal effect of electricity on *Leishmania major*, both *in vitro* and *in vivo*. Ann Trop Med Parasitol 98(1):37-42. https://doi.org/10.1179/ 136485913X13789813917661
- Italia JL, Ravi Kumar M, Carter K (2012) Evaluating the potential of polyester nanoparticles for per oral delivery of amphotericin B in treating visceral leishmaniasis. J Biomed Nanotechnol 8(4):695– 702. https://doi.org/10.1166/jbn.2012.1414
- Jebali A, Kazemi B (2013) Nano-based antileishmanial agents: a toxicological study on nanoparticles for future treatment of cutaneous leishmaniasis. Toxicol in Vitro 27:1896–1904. https://doi.org/10. 1016/j.tiv.2013.06.002
- Jha SN, Singh NK, Jha TK (1991) Changing response to diamidine compounds in cases of kala-azar unresponsive to antimonial. J Assoc Physicians India 39(4):314–316
- Jhingran A, Chawla B, Saxena S, Barrett MP, Madhubala R (2009) Paromomycin: uptake and resistance in *Leishmania donovani*. Mol Biochem Parasitol 164(2):111–117. https://doi.org/10.1016/j. molbiopara.2008.12.007
- Kassi M, Kassi M, Afghan AK, Rehman R, Kasi PM (2008) Marring leishmaniasis: the stigmatization and the impact of cutaneous leishmaniasis in Pakistan and Afghanistan. PLoS Neg Trop Dis 2:e259. https://doi.org/10.1371/journal.pntd.0000259
- Keiser MJ, Setola V, Irwin JJ (2009) Predicting new molecular targets for known drugs. Nature 462:175–181p. https://doi.org/10.1038/ nature08506
- Kimutai R, Musa AM, Njoroge S, Omollo R, Alves F, Hailu A, Khalil EAG, Diro E, Soipei P, Musa B, Salman K, Ritmeijer K, Chappuis F, Rashid J, Mohammed R, Jameneh A, Makonnen E, Olobo J, Okello L, Sagaki P, Strub N, Ellis S, Alvar J, Balasegaram M, Alirol E, Wasunna M (2017) Safety and effectiveness of sodium stibogluconate and paromomycin combination for the treatment of visceral leishmaniasis in Eastern Africa: results from a pharmacovigilance programme. Clin Drug Investig 37(3):259– 272. https://doi.org/10.1007/s40261-016-0481-0
- Kim DH, Chung HJ, Bleys J, Ghohestani RF (2009) Is paromomycin an effective and safe treatment against cutaneous leishmaniasis? A meta-analysis of 14 randomized controlled trials. PLoS Negl Trop Dis (2):e381. https://doi.org/10.1371/journal.pntd.0000381
- Kumar P, Shivam P, Mandal S, Prasanna P, Kumar S, Prasad SR, Kumar A, Das P, Ali V, Singh SK, Mandal D (2019) Synthesis, characterization, and mechanistic studies of a gold nanoparticle-amphotericin B covalent conjugate with enhanced antileishmanial efficacy and reduced cytotoxicity. Int J Nanomedicine 14:6073–6101. https:// doi.org/10.2147/IJN.S196421
- Lai A Fat EJ, Vrede MA, Soetosenojo RM, Fat RFMLA (2002) Pentamidine, the drug of choice for the treatment of cutaneous leishmaniasis in Suriname. Int J Dermatol 41(11):796–800. https://doi. org/10.1046/j.1365-4362.2002.01633.x

- Lopez-Jaramillo P, Ruano C, Rivera J, Terán E, Salazar-Irigoyen R, Esplugues JV, Moncada S (1998) Treatment of cutaneous leishmaniasis with nitric-oxide donor. Lancet. 351:1176–1177. https://doi. org/10.1016/s0140-6736(05)79119-4
- Lopez-Jaramillo P, Rincon MY, Garcia RG, Silva SY, Smith E, Kampeerapappun P, García C, Smith DJ, López M, Vélez ID (2010) A controlled, randomized-blinded clinical trial to assess the efficacy of a nitric oxide releasing patch in the treatment of cutaneous leishmaniasis by *Leishmania* (V.) panamensis. Am J Trop Med Hyg 83:97–10. https://doi.org/10.4269/ajtmh.2010.09-0287
- Machado PR, Ampuero J, Guimarães LH (2010) Miltefosine in the treatment of cutaneous leishmaniasis caused by *Leishmania braziliensis* in Brazil: a randomized and controlled trial. PLoS Negl Trop Dis 4(12):e912. https://doi.org/10.1371/journal.pntd.0000912
- Mahajan R, Das P, Isaakidis P, Sunyoto T, Sagili KD, Lima MA, Mitra G, Kumar D, Pandey K, Van Geertruyden JP, Boelaert M, Burza S (2015) Combination treatment for visceral leishmaniasis patients coinfected with human immunodeficiency virus in India. Clin Infect Dis 61(8):1255–1262. https://doi.org/10.1093/cid/civ530
- Masmoudi A, Maalej N, Boudaya S, Turki H, Zahaf A (2006) Adverse effects of intralesional glucantime in the treatment of cutaneous leishmaniosis. Med Mal Infect 36:226–228. https://doi.org/10. 1016/j.medmal.2005.11.018
- Mastroianni A (2004) Liposomal amphotericin B and rHuGM-CSF for treatment of visceral leishmaniasis in AIDS. Infez Med 12(3):197– 204
- Mayrink W, Botelho AC, Magalhães PA, Batista SM, Lima AO, Genaro O, Costa CA, Melo MN, Michalick MSM, Williams P, Dias M, Caiaffa WT, Nascimento E, Machado-Coelho GLL (2006) Immunotherapy, immunochemotherapy and chemotherapy for American cutaneous leishmaniasis treatment. Rev Soc Bras Med Trop (1):14–21. https://doi.org/10.1590/S0037-86822006000100003
- Melaku Y, Collin SM, Keus K, Gatluak F, Ritmeijer K, Davidson RN (2007) Treatment of kala-azar in Southern Sudan using a 17-day regimen of sodium stibogluconate combined with paromomycin: a retrospective comparison with 30-day sodium stibogluconate monotherapy. Am J Trop Med Hyg 77:89–94
- Monge-Maillo B, López-Vélez R (2013) Therapeutic options for visceral leishmaniasis. Drugs. (17):1863–1888. https://doi.org/10.1007/ s40265-013-0133-0
- Monjour L, Neogy AB, Vouldoukis I, Silva OA, Brito MEF, Lesot A, Vignot N, Martins JS, Jardim ML (1994) Exploitation of parasite derived antigen in therapeutic success of human cutaneous leishmaniasis in Brazil. Mem Inst Oswaldo Cruz 89(3):479–483. https://doi.org/10.1590/S0074-02761994000300034
- Mosleh IM, Geith E, Natsheh L, Schonian G, Abotteen MD, Krarabsheh S (2008) Efficacy of a weekly cryotherapy regimen to treat *Leishmania major* cutaneous leishmaniasis. J Am Acad Dermatol 58(4):617–624. https://doi.org/10.1016/j.jaad.2007.12.032
- Mueller M, Ritmeijer K, Balasegaram M, Koummuki Y, Santana MR, Davidson R (2007) Unresponsiveness to AmBisome in some Sudanese patients with kala-azar. Trans R Soc Trop Med Hyg 101(1):19–24. https://doi.org/10.1016/j.trstmh.2006.02.005
- Musa AM, Younis B, Fadlalla A, Royce C, Balasegaram M, Wasunna M, Hailu A, Edwards T, Omollo R, Mudawi M, Kokwaro G, El Hassan A, Khalil E (2010) Paromomycin for the treatment of visceral leishmaniasis in Sudan: a randomized, open-label, dose-finding study. Lockwood DNJ, editor. PLoS Negl Trop Dis 4(10). https://doi.org/ 10.1371/journal.pntd.0000855
- Musa AM, Khalil E, Hailu A, Olobo J, Balasegaram M, Omollo R, Edwards T, Rashid J, Mbui J, Musa B, Abuzaid AA, Ahmed O, Fadlalla A, El Hassan A, Muller M, Mucee G, Njoroge S, Manduku V, Mutuma G, Apadet L, Lodenyo H, Mutea D, Kirigu G, Yifru S, Mengistu G, Hurissa Z, Hailu W, Weldegebreal T, Tafes H, Mekonnen Y, Makonnem E, Ndegwa S, Sagaki P, Kimutai R,

Kesusu J, Owiti R, Ellis S, Wasunna M (2012) Sodium stibogluconate (SSG) & paromomycin combination compared to SSG for visceral leishmaniasis in East Africa: a randomised controlled trial. PLoS Negl Trop Dis 6(6):e1674. https://doi.org/10. 1371/journal.pntd.0001674

- Navin TR, Arana BA, Arana FE, de Merida AM, Castillo AL, Pozuelos JL (1990) Placebo-controlled clinical trial of meglumine antimoniate (glucantime) vs. localized controlled heat in the treatment of cutaneous leishmaniasis in Guatemala. Am J Trop Med Hyg 42:43–50. https://doi.org/10.4269/ajtmh.1990.42.43
- Negera E, Gadisa E, Hussein J, Engers H, Kuru T, Gedamu L, Aseffa A (2012) Treatment response of cutaneous leishmaniasis due to *Leishmania aethiopica* to cryotherapy and generic sodium stibogluconate from patients in Silti, Ethiopia. Trans R Soc Trop Med Hyg 106(8):496–503. https://doi.org/10.1016/j.trstmh.2012. 02.006
- Organización Panamericana de la Salud (OPS). Leishmaniasis em las Américas (2013) Recomendaciones para el tratamiento. OPS, Washington, pp 1–60
- Paula CDRD, Sampaio JHD, Cardoso DR, Sampaio RNR (2003) A comparative study between the efficacy of pentamidine isothionate given in three doses for one week and N-methil-glucamine in a dose of 20mgSbV/day for 20 days to treat cutaneous leishmaniasis https:// doi.org/10.1590/S0037-86822003000300009
- Pan American Health Organization (2018) Leishmaniasis in the Americas: treatment recommendations. PAHO, Washington, D.C http://iris.paho.org
- Pushpakom S, Iorio F, Eyers PA, Escott KJ, Hopper S, Wells A, Doig A, Guilliams T, Latimer J, McNamee C, Norris A, Sanseau P, Cavalla D, Pirmohamed M (2019) Drug repurposing: progress, challenges and recommendations. Nat Rev Drug Discov 18(1):41–58. https:// doi.org/10.1038/nrd.2018.168
- Prates FVDO, Dourado MEF, Silva SC, Shriefer A, Guimarães LH, Brito MGO, Almeida J, Carvalho EM, Machado PRL (2017) Fluconazole in the treatment of cutaneous leishmaniasis caused by *Leishmania braziliensis*: a randomized controlled trial. Clin Infect Dis 64(1):67– 71. https://doi.org/10.1093/cid/ciw662
- Ramalho DB, Silva RED, Senna MCR, Moreira HAS, Pedras MJ, Avelar DM, Saraiva L, Rabelle A, Cota G (2018) Meglumine antimoniate intralesional infiltration for localised cutaneous leishmaniasis: a single arm, open label, phase II clinical trial. Mem Inst Oswaldo Cruz 113(9):e180200. https://doi.org/10.1590/0074-02760180200
- Reithinger R, Dujardin JC, Louzir H, Pirmez C, Alexander B, Brooker S (2007) Cutaneous leishmaniasis. Lancet Infect Dis 7(9):581–596. https://doi.org/10.1016/S1473-3099(07)70209-8
- Ritmeijer K, Dejenie A, Assefa Y, Hundie TB, Mesure J, Boots G, den Boer M, Davidson RN (2006) A comparison of miltefosine and sodium stibogluconate for treatment of visceral leishmaniasis in an Ethiopian population with high prevalence of HIV infection. Clin Infect Dis 43(3):357–364. https://doi.org/10.1086/505217
- Roatt BM, Aguiar-Soares RD, Coura-Vital W, Gama Ker H, Moreira ND, Vitoriano-Souza J, Giunchetti RC, Carneiro CM, Reis AB (2014) Immunotherapy and immunochemotherapy in visceral leishmaniasis: promising treatments for this neglected disease. Front Immunol 5(272). https://doi.org/10.3389/fimmu.2014.00272
- Roussel M, Nacher M, Fremont G, Rotureau B, Clyti E, Sainte-Marie D, Carme B, Pradinaud R, Couppié P (2006) Comparison between one and two injections of pentamidine isethionate, at 7 mg/kg in each injection, in the treatment of cutaneous leishmaniasis in French Guiana. Ann Trop Med Parasitol 100(4):307–314. https://doi.org/ 10.1179/136485906X105561
- Sadeghian G, Nilfroushzadeh MA, Iraji F (2007) Efficacy of local heat therapy by radiofrequency in the treatment of cutaneous leishmaniasis, compared with intralesional injection of meglumine antimoniate. Clin Exp Dermatol 32:371–374. https://doi.org/10. 1111/j.1365-2230.2007.02405.x

- Saleem K, Khursheed Z, Hano C, Anjum I, Anjum S (2019) Applications of nanomaterials in leishmaniasis: a focus on recent advances and challenges. Nanomaterials 9(12):e1749. https://doi.org/10.3390/ nano9121749
- Sazgarnia A, Taheri AR, Soudmand S, Parizi AJ, Rajabi O, Darbandi MS (2013) Antiparasitic effects of gold nanoparticles with microwave radiation on promastigots and amastigotes of *Leishmania major*. Int J Hyperthermia. 29(1):79-86. https://doi.org/10.3109/02656736. 2012.758875
- Shamsi SM, Zandi S, Aghaie H, Heshmatkhah A (2011) Efficacy of CO(2) laser for treatment of anthroponotic cutaneous leishmaniasis, compared with combination of cryotherapy and intralesional meglumine antimoniate. J Eur Acad Dermatol Venereol 25:587– 591. https://doi.org/10.1111/j.1468-3083.2010.03781.x
- Sinha PK, Jha TK, Thakur CP, Nath D, Mukherjee S, Aditya AK, Sundar S (2011) Phase 4 pharmacovigilance trial of paromomycin injection for the treatment of visceral leishmaniasis in India. J Trop Med 2011(645203). https://doi.org/10.1155/2011/645203
- Soto J, Buffet P, Grogl M, Berman J (1994) Successful treatment of Colombian cutaneous leishmaniasis with four injections of pentamidine. Am J Trop Med Hyg 50(1):107–111. https://doi.org/10.4269/ ajtmh.1994.50.107
- Soto J, Arana BA, Toledo J, Rizzo N, Vega JC, Diaz A, Luz M, Gutierrez P, Arboleda M, Berman JD, Junge K, Engel J, Sindermann H (2004) Miltefosine for new world cutaneous leishmaniasis. Clin Infect Dis 38(9):1266–1272. https://doi.org/10.1086/383321
- Soto J, Paz D, Rivero D, Soto P, Quispe J, Toledo J, Berman J (2016) Intralesional pentamidine: a novel therapy for single lesions of Bolivian cutaneous leishmaniasis. Am J Trop Med Hyg 94(4): 852–856. https://doi.org/10.4269/ajtmh.15-0640
- Sousa AQ, Frutuoso MS, Moraes EA, Pearson RD, Pompeu MML (2011) High-dose oral fluconazole therapy effective for cutaneous leishmaniasis due to *Leishmania (Vianna) braziliensis*. Clin Infect Dis 53(7):693–695. https://doi.org/10.1093/cid/cir496
- Squires KE, Rosenkaimer F, Sherwood JA, Forni AL, Were JB, Murray HW (1993) Immunochemotherapy for visceral leishmaniasis: a controlled pilot trial of antimony versus antimony plus interferon-gamma. Am J Trop Med Hyg 48(5):666–669. https://doi.org/10.4269/ ajtmh.1993.48.666
- Srivastava S, Mishra J, Kumar GA (2017) Laboratory confirmed miltefosine resistant cases of visceral leishmaniasis from India. Parasit Vectors 10:49. https://doi.org/10.1186/s13071-017-1969-z
- Sundar S, Rosenkaimer F, Murray HW (1994) Successful treatment of refractory visceral leishmaniasis in India using antimony plus interferon-gamma. J Infect Dis 170(3):659–662. https://doi.org/10.1093/ infdis/170.3.659
- Sundar S, Murray HW (1995) Effect of treatment with interferon-gamma alone in visceral leishmaniasis. J Infect Dis 172(6):1627–1629. https://doi.org/10.1093/infdis/172.6.1627
- Sundar S (2000) Drug resistance in Indian visceral leishmaniasis. Tropical Med Int Health 6:849–854. https://doi.org/10.1046/j. 1365-3156.2001.00778.x
- Sundar S, Jha TK, Thakur CP, Engel J, Sindermann H, Fischer C, Junge K, Bryceson A, Berman J (2002) Oral miltefosine for Indian visceral leishmaniasis. N Engl J Med 347:1739–1746. https://doi.org/10. 1056/NEJMoa021556
- Sundar S, Chakravarty J, Rai VK, Agrawal N, Singh SP, Chauhan V, Murray HW (2007a) Amphotericin B treatment for Indian visceral leishmaniasis: response to 15 daily versus alternate-day infusions. Clin Infect Dis 45(5):556–561. https://doi.org/10.1086/520665
- Sundar S, Jha TK, Thakur CP, Sinha PK, Bhattacharya SK (2007b) Injectable paromomycin for visceral leishmaniasis in India. N Engl J Med 356(25):2571–2581. https://doi.org/10.1056/ NEJMoa066536
- Sundar S, Mondal D, Rijal S, Bhattacharya S, Ghalib H, Kroeger A, Boelaert M, Desjeux P, Richer-Airijoki H, Harms G (2008a)

Implementation research to support the initiative on the elimination of kala azar from Bangladesh, India and Nepal—the challenges for diagnosis and treatment. Tropical Med Int Health 13(1):2–5. https://doi.org/10.1111/j.1365-3156.2007.01974.x

- Sundar S, Chakravarty J (2008) Paromomycin in the treatment of leishmaniasis. Expert Opin Investig Drugs 17(5):787–794. https://doi. org/10.1517/13543784.17.5.787
- Sundar S, Rai M, Chakravarty J, Agarwal D, Agrawal N, Vaillant M, Olliaro P, Murray HW (2008b) New treatment approach in Indian visceral leishmaniasis: single-dose liposomal amphotericin B followed by short-course oral miltefosine. Clin Infect Dis 47: 1000–1006. https://doi.org/10.1086/591972
- Sundar S, Singh A, Rai M, Chakravarty J, Agarwal D, Vaillant M, Olliaro P, Murray HW (2012) Efficacy of miltefosine in the treatment of visceral leishmaniasis in India after a decade of use. Clin Infect Dis 55(4):543–550. https://doi.org/10.1093/cid/cis474
- Sundar S, Chakravarty J (2013) Leishmaniasis: an update of current pharmacotherapy. Expert Opin Pharmacother 14(1):53–63. https://doi. org/10.1517/14656566.2013
- Sundar S, Chakravarty J (2015) An update on pharmacotherapy for leishmaniasis. Expert Opin Pharmacother 16(2):237–225. https://doi.org/ 10.1517/14656566.2015.97380
- Taslimi Y, Zahedifard F, Rafati S (2018) Leishmaniasis and various immunotherapeutic approaches. Parasitology 145(4):497–507. https:// doi.org/10.1017/S003118201600216X
- Tunes LG, Morato RE, Garcia A, Schmitz V, Steindel M, Côrrea-Junior JD, Santos HF, Frézard F, Almeida MV, Silva H, Moreti NS, Barros ALB, Monte-Neto RL (2020) Preclinical gold complexes as oral drug candidates to treat leishmaniasis are potent trypanothione reductase inhibitors. ACS Infect Dis 8;6(5):1121–1139. https://doi. org/10.1021/acsinfecdis.9b00505
- Uzun S, Durdu M, Culha G, Allahverdiyev AM, Memisoglu HR (2004) Clinical features, epidemiology, and efficacy and safety of intralesional antimony treatment of cutaneous leishmaniasis: recent experience in Turkey. J Parasitol 90:853–859. https://doi.org/10. 1645/GE-185R
- Valencia BM, Miller D, Witzig RS, Boggild AK, Llanos-Cuentas A (2013) Novel low-cost thermotherapy for cutaneous leishmaniasis in Peru. PLoS Neg Trop Dis 7:e2196. https://doi.org/10.1371/ journal.pntd.0002196
- van Griensven J, Diro E (2019) Visceral leishmaniasis: recent advances in diagnostics and treatment regimens. Infect Dis Clin N Am 33(1):79– 99. https://doi.org/10.1016/j.idc.2018.10.005
- Verma NK, Dey CS (2004) Possible mechanism of miltefosine-mediated death of *Leishmania donovani*. Antimicrob Agents Chemother 48(8):3010–3015. https://doi.org/10.1128/AAC.48.8.3010-3015. 2004
- Wagner V, Minguez-Menendez A, Pena J, Fernández-Prada C (2019) Innovative solutions for the control of leishmaniases: nanoscale drug delivery systems. Curr Pharm Des 25(14):1582–1592. https://doi. org/10.2174/1381612825666190621154552
- Wasunna M, Njenga S, Balasegaram M, Alexander N, Omollo R, Edwards T, Dorlo TPC, Musa B, Ali MHS, Elamin MY, Kirigi G, Juma R, Kip AE, Schoone GJ, Hailu A, Olobo J, Ellis S, Kimutai R, Wells S, Khalil EAG, Wourgaft NS, Alves F, Musa A (2016) Efficacy and safety of AmBisome in combination with sodium stibogluconate or miltefosine and miltefosine monotherapy for African visceral leishmaniasis: phase II randomized trial. PLoS Negl Trop Dis 10(9):e0004880. https://doi.org/10.1371/journal. pntd.0004880
- WHO, Public consultation of experts to join the Guideline Development Group (GDG) for treatment of visceral leishmaniasis in HIVvisceral leishmaniasis coinfected persons in East Africa and South-East Asia—second round (2020). www.who.int/leishmaniasis/ news/GDG-treatment-visceral-leishmaniasis-HIV-visceralleishmaniasis/en/. Accessed 05 Jan 2020

- Wortmann G, Zapor M, Ressner R, Fraser S, Hartzell J, Pierson J, Weintrob A, Magill A (2010) Liposomal amphotericin B for treatment of cutaneous leishmaniasis. Am J Trop Med Hyg 83:1028– 1033. https://doi.org/10.4269/ajtmh.2010.10-0171
- Wolcott LE, Wheeler PC, Hardwicke HM, Rowley BA (1969) Accelerated healing of skin ulcer by electrotherapy: preliminary

clinical results. 62(7):795-801. https://doi.org/10.1097/00007611-196907000-00008

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