#### **MINI-REVIEW**



### Diversity of potentially exploitable pharmacological activities of the highly prized edible medicinal fungus *Antrodia camphorata*

Caicheng Wang<sup>1,2,3</sup> • Weiwei Zhang<sup>1,2,3</sup> • Jack Ho Wong<sup>4</sup> • Tzibun Ng<sup>4</sup> • Xiujuan Ye<sup>1,2,3</sup>

Received: 29 March 2019 / Revised: 8 July 2019 / Accepted: 9 July 2019 / Published online: 12 August 2019 © Springer-Verlag GmbH Germany, part of Springer Nature 2019

#### Abstract

Antrodia camphorata, also known as A. cinnamomea, is a precious medicinal basidiomycete fungus endemic to Taiwan. This article summarizes the recent advances in research on the multifarious pharmacological effects of A. camphorata. The mushroom exhibits anticancer activity toward a large variety of cancers including breast, cervical, ovarian, prostate, bladder, colorectal, pancreatic, liver, and lung cancers; melanoma; leukemia; lymphoma; neuroblastoma; and glioblastoma. Other activities encompass antiinflammatory, antiatopic dermatitis, anticachexia, immunoregulatory, antiobesity, antidiabetic, antihyperlipidemic, antiatherosclerotic, antihypertensive, antiplatelet, antioxidative, antiphotodamaging, hepatoprotective, renoprotective, neuroprotective, testis protecting, antiasthmatic, osteogenic, osteoprotective, antiviral, antibacterial, and wound healing activities. This review aims to provide a reference for further development and utilization of this highly prized mushroom.

Keywords Antrodia camphorata · Anticancer · Antiinflammatory · Antileukemia · Hepatoprotective · Neuroprotective

### Introduction

Mushrooms produce a spectacular array of enzymes, nonenzymatic proteins, proteins, polysaccharides, and other compounds with a myriad of activities beneficial to health and thus have arrested the attention of many researchers (Hu et al. 2017; Jedinak et al. 2011; Jiang and Sliva 2010; Lavi et al. 2010a,b; Liu et al. 2017a, b; Pan et al. 2013; Schwartz and Hadar 2014; Wasser 2014, 2017; Wong et al. 2010; Zhou et al. 2017, 2018).

Xiujuan Ye 2578754211@qq.com

- <sup>1</sup> State Key Laboratory of Ecological Pest Control for Fujian and Taiwan Crops, Fujian Agriculture and Forestry University, Fuzhou 350002, Fujian, China
- <sup>2</sup> Key Laboratory of Biopesticide and Chemical Biology, Ministry of Education, Fujian Agriculture and Forestry University, Fuzhou 350002, Fujian, China
- <sup>3</sup> Fujian Key Laboratory of Plant Virology, Institute of Plant Virology, Fujian Agriculture and Forestry University, Fuzhou 350002, Fujian, China
- <sup>4</sup> School of Biomedical Sciences, Faculty of Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China

Antrodia camphorata, also known as Antrodia cinnamomea, or Taiwanofungus camphoratus and Niuchang-chih or Niu-chang-ku in China, which means bull camphor mushroom and stout camphor fungus, is a precious medicinal fungus that belongs to Basidiomycetes. A. camphorata grows in areas 450 to 2000 m above the sea level on the mountains of Taiwan, in the hollow interior of the decayed trunk or the moist surface of the lodging trunk of Cinnamomum kanehirae tree (bull camphor tree) (Ao et al. 2009; Wu et al. 1997), an endemic native tree species in Taiwan. The mushroom has been used as a traditional remedy and a cancer-preventive herbal supplement in Taiwanese folk medicine. It is well known for its antidotal and antitumor functions and used to treat toxicities caused by food, alcohol, and drugs as well as conditions such as diarrhea, abdominal pain, hypertension, itchy skin, and tumors (Geethangili and Tzeng 2011). It has captured much attention in recent years owing to its multifarious health benefits embracing antiinflammatory, antioxidative, antileukemia, hepatoprotective, detoxifying, neuroprotective, and other effects, particularly its anticancer activity. In this article, the pharmacological effects of A. camphorata are presented from the above facets. Its active ingredients and pharmacological effects are summarized in Table 1, with the intent to provide a reference which may be useful for future work exploring the further development and utilization of A. camphorata.

## Pharmacological effects of A. camphorata demonstrated using purified ingredients

# Effect of 4,7-dimethoxy-5-methylbenzo [d][1,3] dioxole and its more polar analogs on some cancer cell lines

The synthetic forms of a secondary metabolite of *A. camphorata*, 4,7-dimethoxy-5-methylbenzo [d][1,3]dioxole, and its more polar analogs weakly suppressed the proliferation of some cancer cell lines in the NCI-60 screen at 10  $\mu$ M concentration (Yeung and Piggott, 2018).

### Effect of antroquinonol on several carcinoma cells

Antroquinonol induced apoptosis in carcinoma cells via impeding activation of Ras and Rho signaling protein. This effect was achieved by downregulating the expression level of isoprenyltransferase (Ho et al. 2014).

### Effect of methylantcinate A on oral cancer cells

A triterpenoid compound, methylantcinate A, extracted from the fruiting bodies of *A. camphorata* was investigated for the antiproliferative effect and the mechanism of induction of apoptosis in human oral cancer cell lines OEC-M1 and OC-2. The inhibitory effect on the growth of oral cancer cells was mediated by the Bax-related mitochondrial apoptotic pathway, but the specific mechanism of action needs further study (Tsai et al. 2010).

### Effect of 4,7-dimethoxy-5-methyl-1,3-benzodioxole, 1(10 $\rightarrow$ 6)abeo-ergosta-5,7,9,22-tetraen-3 $\alpha$ -ol, citreoanthrasteroid B, dankasterones A, dankasterones B, 4-acetylantroquinonol B, and antroquinonol on colon or colorectal cancer cells

A purified ingredient of fruiting bodies from *A. camphorata*, 4,7-dimethoxy-5-methyl-1,3-benzodioxole, exhibited an inhibitory effect on COLO-205 colon cancer cells in vitro by activating the p27/Kip1 signaling pathway mediated by p53 (Tu et al. 2012).

The effects of seven components from *A. camphorata* were explored. The results indicated that four of these compounds including  $1(10 \rightarrow 6)$ abeo-ergosta-5,7,9,22-tetraen-3 $\alpha$ -ol, citreoanthrasteroid B, dankasterones A, and dankasterones B exhibited a significant inhibitory effect on CT26 colorectal cancer cells and human K562 leukemia cells (Lee et al. 2014b).

4-Acetylantroquinonol B, also extracted from *A. camphorata* mycelia, exerted a suppressive effect on colorectal carcinoma cells. The activity likely involved the STAT3 signaling channel (Chang et al. 2015a). Antroquinonol

isolated from *A. camphorata* mycelia impeded the growth, migration, and invasion of colon carcinoma cells, probably by acting on the PI3K/AKT/ $\beta$  signaling pathway (Lin et al. 2017).

4-Acetylantroquinonol B did not affect the viability and proliferation of normal colon cells. In contrast, it diminished proliferation, migration, invasion, clonogenicity, and viability in colorectal cancer HCT116 and DLD1 cells. Re-expression of hsa-miR-324-5p and expression of E-cadherin and BAX2 were stimulated while expression of BcL-xL2, c-Myc, Ncadherin, SOD2, and vimentin was reduced. The expression of hsa-miR-324-5p in colorectal cancer cells was inhibited and demonstrated an inverse relationship with superoxide dismutase SOD2 expression. Enhanced expression of hsa-miR-324-5p in colorectal cancer cells repressed their in vitro and in vivo carcinogenicities. 4-Acetylantroquinonol B enhanced the antineoplastic activity of FOLFOX (composed of folinate (leucovorin), fluorouracil, and oxaliplatin) by eliciting the re-expression of SOD2-suppressed hsa-miR-324 and inhibiting SOD2-mediated tumorigenicity (Bamodu et al. 2018).

# Effect of antroquinonol, triterpenoids, polysaccharides, and 1,3-β-D-glucan on pancreatic adenocarcinoma cells

Antroquinonol isolated from *A. camphorata* exhibited antiproliferative activity in PANC-1 and AsPC-1 pancreatic cancer cells. The mechanism involved inhibition of PI3-kinase/ Akt/mTOR pathways and induction of G1 arrest in the cell cycle and apoptosis (Yu et al. 2012).

The ethanol-soluble and ethyl acetate-soluble compounds (mainly triterpenoids, polysaccharides, and 1,3- $\beta$ -D-glucan) extracted from *A. camphorata* fruiting bodies manifested a cytotoxic effect on BxPC-3 human pancreatic adenocarcinoma cells but not on normal cells. The extracts of *A. camphorata* hindered proliferation, promoted apoptosis, and suppressed migration of BxPC-3 human pancreatic adenocarcinoma cells. These effects were dependent on the mitochondrial apoptotic pathway. In particular, it was brought about by regulating the bax/bcl-2 ratio of the levels of expression of bax and bcl-2, augmenting the release of cytochrome c, and triggering the activation of caspase-9 and caspase-3 (Lee et al. 2014a).

## Effects of 4-acetylantroquinonol B on hepatocellular carcinoma cells

5'AMP-activated protein kinase (AMPK) and the mammalian target of rapamycin (mTOR) are targets for treatment of liver cancer with chemotherapy. The ranking of potency of antroquinonol against various liver cancer lines was HepG2 > HepG2.2.15 > Mahlavu > PLC/PRF/5 > SK-Hep1 > Hep3B. Antroquinonol curbed the translation but not transcription of G1 regulator proteins, encompassing Cdk2, Cdk4, cyclin D1, and cyclin E. The assembly of tuberous

sclerosis complex (TSC)-1/TSC2 was thwarted, resulting in arrest of protein biosynthesis via suppression of protein

 Table 1
 Pharmacological effects of some active ingredients isolated from Antrodia camphorata

Chemical structure	Compound name	Main pharmacological effects	Reference
H-CO.	2,3-Dimethoxy-5-	Anti-cancer effect through inducing A549	Chung et al. 2014
ĬĬ	methyl-1,4-benzoq	lung cell apoptosis and mediating ROS	
H <sub>3</sub> CO	uinone	production	
,OCH1	2,3,5-Trimethoxy-	Suppressing lung carcinoma cells	Lin et al. 2015a
н,со осн,	4-cresol	migration/invasion	
осн <sub>3</sub>	2,4-Dimethoxy-6-	Ameliorating atopic dermatitis-like lesion	Yang et al. 2018
	methylbenzene-1,3		
H <sub>3</sub> CO <sup>2</sup> CH <sub>3</sub>	-diol		
OCH <sub>3</sub>	4,7-Dimethoxy-5-	Inhibiting COLO-205 colon cancer cells;	Tu et al. 2012
	methyl-1,3-benzodi	Inhibiting LPS-induced inflammation in	Shie et al. 2016
OCH <sub>3</sub>	oxole	RAW264.7 cells	
	Antrocamphin A	Inhibitory synthesis and expression of	Buccini et al.
Н,СО ОСН.		TNF	2014
CH <sub>3</sub> O	Antrolone	Inhibiting LPS-induced inflammation in	Yen et al. 2018
HO OCH,		RAW264.7 cells	
	Antrodin A	Inhibiting replication of herpes simplex	He et al. 2016
		virus in Vero cells	
он 	3-Isobutyl-4-[4-(3-	Inhibiting Lewis lung carcinoma cells	Nakamura et al.
	methyl-2-butenylo		2004
	xy)phenyl]-1H-pyr		
	rol-1-ol-2,5-dione		
, H	3-Isobutyl-4-[4-(3-	Inhibiting Lewis lung carcinoma cells	Nakamura et al.
	methyl-2-butenylo		2004
	xy)phenyl]-1H-pyr	Preventing liver fibrosis both in vitro and in	Yang et al. 2017b
. • 0 •	role-2,5-dione	vivo in an early phase	
он 	Antrocinnamomins	Anti-inflammatory activity by restraining	Wu et al. 2013
	E	NO generation	
	$HO + GOCH_3$ $HO + GOCH_3$ $H_3CO + GO$	$ \begin{split} & \underset{k,co}{ \qquad \qquad$	$ \begin{split} & \underset{n \in \mathcal{F}}{\operatorname{hor}} + \underset{n \in \mathcal{F}}{\operatorname{hor}} & \underset{n \in \operatorname{hor}}{\operatorname{hor}} + \underset{n \in \operatorname{hor}}{\operatorname{hor}} + \underset{n \in \operatorname{hor}}{\operatorname{hor}} + \underset{n \in \operatorname{hor}}{\operatorname{hor}} & \underset{n \in \operatorname{hor}}{\operatorname{hor}} + \underset{hor}}{\operatorname{hor}} + \underset{hor}}{\operatorname{hor}} + \underset{hor}}{\operatorname{hor}}$

### Table 1(continued)

Pyrrole-2,5-dione	он   N	Antrocinnamomins	Anti-inflammatory activity by restraining	Wu et al. 2013
		F	NO generation	
Pyrrole-2,5-dione	0> / <sup>N</sup> > ~0	Antrocinnamomins	Anti-inflammatory activity by restraining	Wu et al. 2013
		G	NO generation	
	H <sub>J</sub> CO OH			
Pyrrole-2,5-dione	0~ ( ~ ~ 0	Antrocinnamomins	Anti-inflammatory activity by restraining	Wu et al. 2013
		Н	NO generation	
	H <sub>J</sub> CO OH			
Sesquiterpene		Antrocin	Thwarting cell proliferation and evoking	Yeh et al. 2013
lactone			apoptosis in the lung cancer cells	
Ubiquinone	H	4-Acetylantroquino	Inhibiting proliferation of hepatoma	Lin et al. 2010b
derivative		nol B	carcinoma cells;	
	H <sub>3</sub> CO OCOCH <sub>3</sub>		Inhibiting colorectal cancer tumorigenesis	Chang et al.
	l осн <sub>3</sub>		and suppressing cancer stem-like phenotype	2015a
Ubiquinone		Antroquinonol	Inhibiting colon cancer stem cell-like cells;	Lin et al. 2017
derivative			Suppressing breast tumor	Lee et al. 2015a
			migration/invasion;	
			Promoting learning and memory of	Chang et al.
	н,со		defective mice;	2015b
	OCH <sub>3</sub>		Blocking Ras and Rho signaling via the	Ho et al. 2014
			inhibition of protein isoprenyltransferase;	
			Inhibiting cell proliferation in pancreatic	Yu et al. 2012
	= , , , ,		cancer PANC-1 and AsPC-1 cells	
Ubiquinone		Antroquinonol D	Potential DNA demethylatingand anticancer	Wang et al. 2014
derivative	н,со		effects	
Lanostanoid		11α-Hydroxy-3,7-d	Anti-inflammatory activity by inhibiting NO	Liaw et al. 2013
	НО Инин	ioxolanost-8,24(E)-	production	
		dien-26-oic acid		
	Č A Č			

phosphorylation. AMPK activity was upregulated. AMPK plays a paramount role in the action of antroquinonol since

the effects of antroquinonol were abolished by an AMPK inhibitor. Disruption of mitochondrial membrane potential,

### Table 1(continued)

Lanostanoid		15-O-acetylganolu	Anti-inflammatory activity by inhibiting NO	Liaw et al. 2013
		cidate A	production	
Lanostanoid		3,7,11-Trioxo-5α-l	Anti-inflammatory activity by inhibiting NO	Liaw et al. 2013
	O OH	anosta-8,24(E)-die	production	
		n-26-oic acid		
anostanoid		Ethyl lucidenate A	Anti-inflammatory activity by inhibiting NO	Liaw et al. 2013
	OCH <sub>2</sub> CH <sub>3</sub>		production	
Unsaturated		7,9(11),22-Ergostat	Antidiabetic activity by regulating the	Kuo et al. 2015b
terol		rien-3β-ol	expression levels of GLUT4,	
		(Antrosterol)	phosphenolpyruvate carboxykinase,	
			glucose-6-phosphatase and AMPK	
			phosphorylation;	
			Anti-inflammatory effects in a mouse skin ischemia model;	Tsai et al. 2015
			Anti-inflammatory and anti-photodamaging	Kuo et al. 2016b
	но		effects; Protecting the liver from	Chang et al.
			CCl4-induced damage via enhancing	2017b
			antioxidant and anti-inflammatory capacities	
			Downregulating expression of the	Kao et al. 2018
			proinflammatory cytokines IL-1β and IL-6	
Unsaturated		7,22-Ergostadien-3	Downregulating expression of the	Kao et al. 2018
sterol		β-ol	proinflammatory cytokines IL-1β and IL-6	
Sterol with		1(10→6)Abeo-erg	Inhibiting CT26 colorectal cancer cells and	Lee et al. 2014b
aromatic ring B		osta-5,7,9,22-tetrae n-3α-ol	human K562 leukemia cell line	

7847

Sterol with	14.000 V	Citreoanthrasteroid	Inhibiting CT26 colorectal cancer cells and	Lee et al. 2014b
aromatic ring B		В	human K562 leukemia cell line	
Steroid	ном, соон	Antcin A	Triggering glucocorticoid receptor transfer to the nucleus to restrain inflammation	Chen et al. 2011
Steroid		Antcin B (Zhankuic acid A)	Inhibiting HL 60 leukemia cells	Du et al. 2012
Steroid		Antcin C	Inhibiting HL 60 leukemia cells	Du et al. 2012
Steroid		Antcin H (Zhankuic acid C)	Inhibiting HL 60 leukemia cells Inducing lymphoma cell line apoptosis	Du et al. 2012 Chen et al. 2018f
13(14→8) Abeo-ergostane-t ype steroid		Dankasterone A	Inhibiting CT26 colorectal cancer cells and human K562 leukemia cell line	Lee et al. 2014b
13(14→8) Abeo-ergostane-t ype steroid		Dankasterone B	Inhibiting CT26 colorectal cancer cells and human K562 leukemia cell line	Lee et al. 2014b
Triterpenoid		Dehydroeburicoic acid	Inhibiting HL 60 leukemia cells Suppressing inflammatory mediator expression in mice;	Du et al. 2012 Deng et al. 2013
	нот Хін		Hepatoprotective effects in a mouse model	Huang et al. 2013

#### Table 1 (continued)

(continued)				
			of acute hepatic injury;	
			Antidiabetic and	Kuo et al. 2015a
			antihyperlipidemic activities in	
			streptozotocin-induced mice in vitro;	
			Preventing diabetic and dyslipidemic	Kuo et al. 2016a
			state in high-fat-fed mice;	
Triterpenoid	HOOC	Eburicoic acid	Suppressing inflammatory mediator	Deng et al. 2013
			expression in mice	
ſ			Hepatoprotective effects in a mouse model	Huang et al. 2013
но			of acute hepatic injury	
Triterpenoid	Muhner COOCH3	Methylantcinate A	Inducing apoptosis in OEC-M1 and OC-2	Tsai et al. 2010
			human oral cancer cells	
0				
Triterpenoid	1	Methylantcinate B	Inhibiting Helicobacter pylori	Lin et al. 2013
-	COOCH3		cytotoxin-associated gene A	
			-induced pathogenesis	
0				
Lanostane	- 0	Ethyl	Anti-inflammatory activity by inhibiting NO	Liaw et al. 2013
triterpene	O <sup>4/I</sup>	3,7,11,12,15,23-he	production	
		xaoxo-5α-lanost-8-		
Ĺ		en-26-oate		
0				
Lanostane		Methyl	Anti-inflammatory activity by inhibiting NO	Liaw et al. 2013
triterpene		3,7,11,12,15,23-he	production	
		xaoxo-5α-lanost-8-		
0		en-26-oate		

arrest of the cancer cells at G1 phase of the cell cycle, and ultimately apoptosis were observed (Chiang et al. 2010).

4-Acetylantroquinonol B was the most effective active ingredient in the fermentation broth of cultured *A. camphorata* mycelia which was capable of suppressing proliferation of hepatoma carcinoma cells (Lin et al. 2010b).

4-Acetylantroquinonol B from *A. cinnamomea* mycelia augmented the anticancer action of dendritic cells against liver cancer stem cells and holds promise for the prevention and immunotherapy of hepatoma. 4-Acetylantroquinonol B downregulated epithelial cell adhesion molecule (EpCAM), alpha-fetoprotein (AFP), and related pathways of hepatoma HepG2 cells and suppressed tumorigenicity. It repressed  $\beta$ catenin expression and liberation of cytokines associated with immune escape and exerted proliferative activity toward immunocytes and enhanced the endocytotic activity of immature dendritic cells. When immature dendritic cells were cultured with EpCAM+ HepG2 cells, MHC class I and II expressions on both types of cells were upregulated by 4acetylantroquinonol B. The expression of CD80 costimulatory molecules in dendritic cells and cytokines associated with immunostimulation was upregulated (Li and Chiang, 2019).

## Effect of 4-acetylantroquinonol B on neuroblastoma and glioblastoma cells

Antroquinonol decreased cell viability in C6 glioma N18 as well as neuroblastoma cell lines and triggered apoptosis of the cancer cells. p53 and other pro-apoptotic proteins were upregulated while antiapoptotic proteins were suppressed. The protein levels of cdc42 FAK, pFAK, Rac1, Src, pSrc, and epithelial-to-mesenchymal-transition proteins were reduced in response to antroquinonol. Antroquinonol exerted an anticancer action in mice bearing xenograft C6 glioma (Thiyagarajan et al. 2015).

4-Acetylantroquinonol B downregulated expression of the following: Sox2 and Oct4 important for keeping the pluripotent embryonic stem cell phenotype;  $\beta$ -catenin, vimentin, and slug involved in epithelial-mesenchymal transition; the oncogene c-Myc and Krüppel-like factor 4 were involved in proliferation, differentiation, and apoptosis. 4-Acetylantroquinonol B suppressed the cancer-promoting catenin/LEF1/Stat3 signaling pathway and decreased viability in U87MG and DBTRG-05MG glioblastoma cells. Invasiveness of the cells and formation of colonies and tumorspheres were abated (Liu et al. 2018a).

### Effects of antroquinonol D, antroquinonol, and antrocin on breast cancer cells and action of ergostane-type triterpenoids on breast cancer cells

Antroquinonol D isolated from *A. camphorata* mycelia elicited DNA demethylation, induced cancer cell apoptosis, and suppressed the survival and migration of breast carcinoma cells (Wang et al. 2014). The impact of antroquinonol, (4R,5R,6R)-4-hydroxy-2,3-dimethoxy-6-methyl-5-[(2E,6E)-3,7,11– trimethyldodeca-2,6,10-trien-1-yl] cyclohex-2-en-1-one, a ubiquitin derivative extracted from the solid-state culture of *A. camphorata*, on human breast carcinoma cells including MCF7 and MDA-MB-231 cells, has been investigated. The results demonstrated that antroquinonol manifested a repressive effect on the migration and invasion of MCF7 and MDA-MB-231 cells to different degrees. The mechanism primarily entailed suppressing the expression of EMT protein and matrix metalloproteinase (MMP)-9 protein via mediating the ERK-AP-1 and AKT-NF-κB signaling pathways (Lee et al. 2015a).

Ergostane-type triterpenoids extracted from *A. camphorata* fruiting bodies possessed cytotoxicity against human breast cancer MDA-MB-231 cells (Huang et al. 2014b).

Antrocin suppressed the viability and production of migration colonies and mammospheres. It downregulated the expression of Notch1, Akt, and  $\beta$ -catenin. It is known that  $\beta$ catenin phosphorylation of AKT enhances invasiveness of cancer cells, elevated expression of Notch 1 is detected in cancer tissue, and that heightened Atk and  $\beta$ -catenin expression is associated with drug resistance and poor prognosis in breast cancer patients. Antrocin synergized with paclitaxel to reduce viability of breast cancer cells in vitro and exhibited an anticancer action in mice bearing breast cancer xenografts. The concurrent application of antrocin and paclitaxel awaits clinical trials (Chen et al. 2019a).

### Effect of polysaccharide on cervical cancer cells

A polysaccharide purified from water extraction of *A. cinnamomea* mycelia displayed inhibitory effects on HeLa cervical cancer cells. It induced apoptosis and cell cycle arrest, and the mechanism involved blocking of topoisomerase I/tyrosyl-DNA phosphodiesterase I-mediated DNA repair pathway (Zhang et al. 2018b).

## Ameliorative effect of antrodan and antrocin on prostatic hyperplasia

Antrodan extracted from *A. camphorata* mycelia could improve benign prostatic hyperplasia in the Sprague-Dawley rat model. It exerted its action through reducing collagen accumulation and decreasing the expression of sex hormones including testosterone and estradiol (Peng et al. 2015).

Concurrent application of antrocin and ionizing radiation produced synergistic antiproliferative and apoptotic effects in prostate cancer cells resistant to radiation. Antrocin downregulated PI3K/AKT and MAPK signaling pathways as well as suppressed type 1 insulin-like growth factor 1 receptor (IGF-1R)-mediated induction of  $\beta$ -catenin. Antrocin potentiated the efficacy of radiotherapy in mice bearing prostate cancer xenografts (Chen et al. 2019c).

### Effect of maleic and succinic acid derivatives, antrocin, 2,3-dimethoxy-5-methyl-1,4-benzoquinone, 2,3,5-trimethoxy-4-cresol, antrodan, antroquinonol, and sulfated glucan on lung carcinoma cells

Five new maleic and succinic acid derivatives were extracted from *A. camphorata* mycelia. It was found that 3-isobutyl-4-[4-(3-methyl-2-butenyloxy)phenyl]-1H-pyrrole-2,5-dione and 3-isobutyl-4-[4-(3-methyl-2-butenyloxy)phenyl]-1H-pyrrol-1-ol-2,5-dione displayed cytotoxic activity on Lewis lung carcinoma cells (Nakamura et al. 2004).

Studies have been conducted on the antilung cancer effect and the action mechanism of antrocin, which is 7,7-dimethyl-4-methylidene-3,3a,5,6,6a,8,9,10-octahydro benzo [h][2] benzofuran-1-one, a sesquiterpene lactone extracted from *A. camphorata* on H441 and H1975 non-small-cell lung cancer cells. Antrocin thwarted cell proliferation and evoked apoptosis in the lung cancer cells. Antrocin induced cell apoptosis via enhancing the activity of caspase-3, mediating the expression of apoptosis-related proteins and bringing about downregulation of the JAK2/STAT3 signal pathway (Yeh et al. 2013). The effect of an ethyl acetate extract of submerged cultivated *A. camphorata* mycelia against A549 human lung cancer cells was investigated. The results indicated that 2,3-dimethoxy-5-methyl-1,4-benzoquinone was the most effective active ingredient in the induction of apoptosis in A549 cells. 2,3-Dimethoxy-5-methyl-1,4-benzoquinone exhibited anticancer effect through inducing apoptosis of A549 cells and mediating ROS production (Chung et al. 2014). Ergostane-type triterpenoids extracted from *A. camphorata* fruiting bodies possessed cytotoxicity against lung carcinoma A549 cells (Huang et al. 2014b).

2,3,5-Trimethoxy-4-cresol obtained from the solid-state cultivation of *A. camphorata* mycelia inhibited the invasion and migration of lung carcinoma cells by mediating the expression levels of some molecules including protein kinase B, MMP-2, MMP-9, E-cadherin, and tissue inhibitor of metalloproteinase (TIMP)-1 (Lin et al. 2015a).

Antrodan isolated from *A. camphorata* mycelia exerted antimetastatic activity in lung carcinoma cells through regulating the expression of MMP-2, MMP-9, TIMP-1, TIMP-2, and nm23-H1 (Fa et al. 2015). Furthermore, antrodan performed well in adjuvant therapy when combined with cisplatin and ameliorated cisplatin-induced kidney dysfunction (Chen et al. 2018c).

In A549 cells, antroquinonol elicited mitochondrial membrane depolarization, increased arrest in Sub-G1 phase of the cell cycle, exerted an antiproliferative effect, increased cell shrinkage, and induced appearance of apoptotic vacuoles, pores, and TUNEL-positive cells. Antroquinonol downregulated cdc2, Bcl2, PI3K, and mTOR but did not affect the expression of pcdc2, pcdc25C cyclin B1, and cdc25C. PARP cleavage and caspase 3 were activated. The levels of expression of miRNAs antroquinonol were altered (Kumar et al. 2011b). In view of the action of antroquinonol on different kinds of cancers such as non-small cell lung cancer, Huang et al. (2017) suggested the use of antroquinonol in conjunction with anticancer drugs in the treatment of advanced and/or refractory solid tumors. Antroquinonol as a therapeutic agent for non-small-cell lung cancer is in phase II clinical trials in Taiwan and USA (Chou et al. 2019). 4-Hydroxybenzoic acid is the ring precursor for antroquinonol but not for 4-acetylantroquinonol B. Endogenous 4-hydroxybenzoic acid is employed by A. cinnamomea through the shikimate pathway for the formation of antroquinonol, but exogenous phenylalanine and tyrosine are used instead in the event of glyphosate blockade of the shikimate pathway. The benzoquinone ring of antroquinonol is produced through the polyketide pathway as well as the shikimate pathway whereas 4-acetylantroquinonol B is formed through only the polyketide pathway (Chou et al. 2019).

Sulfated glucan isolated from *A. cinnamomea* exhibited inhibitory activity toward lung cancer cells; it reduced cancer cell viability via inhibition of EGFR and mTOR activity and showed a synergistic effect when administered in conjunction with cisplatin (Lu et al. 2018). The glucan downregulated the TGF $\beta$ /FAK/AKT axis and triggered the breakdown of Slug in lung cancer H1975 cells and thus prevented the growth of the lung cancer cells (Lin et al. 2019b).

### Antileukemia activity of antcin C, dehydroeburicoic acid, zhankuic acid A, and zhankuic acid C on leukemia cells

The cytotoxicity of active ingredients from the ethanolic extract of A. camphorata on leukemia HL 60 cells was studied. The active ingredients were predominantly enriched in the triterpenoid fraction. Four major triterpenoids were isolated from the active components, namely, antcin C ((6R)-6-[(4S,5S,7S,10S,13R,14R,17R)-7-hydroxy-4,10,13-trimethyl-3,11-dioxo-2,4,5,6,7,12,14,15,16,17-decahydro-1H-cyclopenta [a]phenanthren-17-yl]-2-methyl-3-methylideneheptanoic acid), dehydroeburicoic acid ((2R)-2-[(3S,5R,10S,13R,14R,17R)-3hydroxy-4,4,10,13,14-pentamethyl-2,3,5,6,12,15,16,17octahydro-1H-cyclopenta [a]phenanthren-17-yl]-6-methyl-5methylideneheptanoic acid), zhankuic acid A ((6R)-2-methyl-3methylidene-6-[(4S,5S,10S,13R,14R,17R)-4,10,13-trimethyl-3,7,11-trioxo-1,2,4,5,6,12,14,15,16,17-decahydrocyclopenta [a]phenanthren-17-yl] heptanoic acid), and zhankuic acid C ((6R)-6-[(3R,4S,5S,10S,12R,13R,14R,17R)-3,12-dihydroxy-4,10,13-trimethyl-7,11-dioxo-2,3,4,5,6,12,14,15,16,17decahydro-1H-cyclpoenta [a]phenanthren-17-yl]-2-methyl-3methylideneheptanoic acid). Dehydroeburicoic acid demonstrated the most potent cytotoxicity through inducing DNA damage and apoptosis. Dehydroeburicoic acid also induced cell cycle (mainly G2/M phase) arrest and suppressed the expression of topoisomerase II (Du et al. 2012).

### Effect of antcin H on lymphoma cells

Antcin H inhibited JAK/STAT-related signaling induced by latent membrane protein 1 and induced apoptosis of lymphoma cells. Besides, it enhanced the cytotoxicity of methotrexate against lymphoma cells at a low concentration (Chen et al. 2018f).

### Immunomodulatory effect of polysaccharides, antroquinonol, antrodin D, and 2,4-dimethoxy-6-methylbenzene-1,3-diol

Through a series of analyses, it was revealed that the immunomodulatory effect of a hot water extract of fermentation culture obtained from *A. camphorata* mycelia was mainly related to the 10–20 kDa polysaccharides and adenosine (Kuo et al. 2008). The immunomodulatory activity and antiinfective effect of polysaccharides extracted from *A. camphorata* mycelia on *Schistosoma mansoni* were disclosed by studying cytokine expression and immune function in T1/T2 doubly transgenic mice (Cheng et al. 2008). Polysaccharides derived from *A. camphorata* mycelia could regulate the levels of indices of immune organs (spleen, thymus, and bursa), blood biochemical indices, and T lymphocyte proliferation and manifested an immunomodulatory effect on specific pathogen-free chickens (Song et al. 2014).

Antroquinonol exerted an antiproliferative activity toward CD8<sup>+</sup> T cells and inhibited the generation of proinflammatory cytokines interferon- $\gamma$  and interleukin-2 markers of T cell activation CD137 and CD69 in vitro. Antroquinonol attenuated the decrease in length of hair follicles, cutaneous thickness, and expression of tyrosinase involved in pigment formation and pigmentation caused by damage brought about by hydrogen peroxide in C57BL/6 mice. Antroquinonol downregulated cutaneous CD8<sup>+</sup> T cell infiltration, proinflammatory cytokine secretion, and expression of the chemokines CXCL10 and CXCR3 (Guan et al. 2017).

The cold water-soluble galactomannan of *A. cinnamomea* augmented the ability of mouse macrophages to phagocytose and kill *Escherichia coli* and elevate the production of the cytokines IL-6 and TNF- $\alpha$ . The galactomannan exerted its immunoenhancing action through protein kinase C- $\alpha$  and mitogen-activated protein kinases (MAPK) phosphorylation via Toll-like receptor 4 and the endotoxin tolerance-like effect via suppression of NF- $\kappa$ B (Perera et al. 2017, 2018). The ameliorative effects of polysaccharide obtained from *A. cinnamomea* against cyclophosphamide-induced immunosuppression were determined. It was found that polysaccharide administration enhanced the function of T cells and the cytotoxicity of natural killer cells and counteracted the effects induced by cyclophosphamide, including decreased levels of immunoglobulin A, G, and M and interleukin 2, 6, and 12 (Liu et al. 2018b).

Antrodin D and 2,4-dimethoxy-6-methylbenzene-1,3-diol suppressed nitric oxide production in mouse macrophages which had been stimulated by lipopolysaccharide but did not affect their viability, with IC<sub>50</sub> values of 26.3 and 32.2  $\mu$ g/mL, respectively. 4-Acetyl-antroquinonol B and antroquinonol B were more potent with IC<sub>50</sub> values of 14.7 and 16.2  $\mu$ g/mL, respectively, but cytotoxicity was detectable albeit at much higher concentrations. Slightly less than half of the macrophages were viable after exposure to 2,3-(methylenedioxy)-6-methylbenzene-1,4-diol at a concentration of 16.8  $\mu$ g/mL (Yang et al. 2009).

### Antiinflammatory activity of polysaccharides, antcin A, antrocinnamomins E-H, dehydroeburicoic acid, eburicoic acid, lanostanoids, lactone derivatives, methylantcinate B, antrocamphin A, 4,7-dimethoxy-5-methyl-1,3-benzodioxole, ergostatrien-3β-ol, and antrolone

The antiinflammatory effect of polysaccharides isolated from mycelia and fruiting bodies of *A. camphorata* in septic mice

has been explored. The polysaccharides undermined the inflammatory reaction mainly through diminishing the expression levels of proinflammatory cytokines such as IL-6, IL-10, and TNF- $\alpha$ . Polysaccharides extracted from fruiting bodies appeared to be more efficacious than their mycelial counterparts in attenuating the inflammatory reaction (Meng et al. 2012).

The antiinflammatory activity of active constituents including five major antcins (A, B, C, H, and K) isolated from the methanolic extract of *A. camphorata* fruiting bodies was examined. Antcin A (6R)-2-methyl-3-methylidene-6-[(4S,5S,10S,13R,14R,17R)-4,10,13-trimethyl-3,11-dioxo-2,4,5,6,7,12,14,15,16,17-decahydro-1H-cyclopenta [a] phenanthrene-17-yl] heptanoic acid) played a role in triggering glucocorticoid receptor transfer to the nucleus to curb inflammation like glucocorticoids, while antcins B, C, H, and K did not produce similar effects (Chen et al. 2011). As antiinflammatory drugs, however, glucocorticoids exert some short-term side effects. It awaits further research to ascertain whether antcin A exerts an adverse effect on the body.

The ethanol extract of *A. camphorata* mycelia restrained NO generation. The isolated main active ingredients were antrocinnamomins E, antrocinnamomins F, antrocinnamomins G, and antrocinnamomins H, respectively (Wu et al. 2013).

Eburicoic acid ((2R)-2-[(3S,5R,10S,13R,14R,17R)-3-hydroxy-4,4,10,13,14-pentamethyl-2,3,5,6,7,11,12, 15,16,17decahydro-1H-cyclopenta [a]phenanthren-17-yl]-6-methyl-5methylideneheptanoic acid) and dehydroeburicoic acid ((2R)-2-[(3S,5R,10S,13R,14R,17R)-3-hydroxy-4,4,10,13,14pentamethyl-2,3,5,6,12,15,16,17-octahydro-1H-cyclopenta [a] phenanthren-17-yl]-6-methyl-5-methylideneheptanoic acid) extracted from the solid culture of *A. camphorata* exhibited analgesic and antiinflammatory activities associated with the reduction of some inflammatory cytokines such as NO, TNF- $\alpha$ , and IL-1 $\beta$  and the augmented activities of some antioxidant enzymes including catalase, superoxide dismutase, and glutathione peroxidase (Deng et al. 2013).

Six lanostanoids and lactone derivatives were isolated from the coculture of mycelia and fruiting bodies of *A. camphorate.* Four known compounds were evaluated for antiinflammatory effect through measuring the level of NO generated by RAW 264.7 cells. It was found that six compounds (ethyl lucidenate A, 15-O-acetylganolucidate A, 11 $\alpha$ -hydroxy-3,7-dioxolanost-8,24(E)-dien-26-oic acid, 3,7,11-trioxo-5 $\alpha$ -lanosta-8,24(E)dien-26-oic acid, methyl 3,7,11,12,15,23-hexaoxo-5 $\alpha$ -lanost-8-en-26-oate and ethyl 3,7,11,12,15,23-hexaoxo-5 $\alpha$ -lanost-8en-26-oate, respectively) exerted antiinflammatory activity by inhibiting NO production (Liaw et al. 2013).

Methylantcinate B, purified from the chloroform extract of *A. camphorata* fruiting bodies, bridled the inflammatory reaction induced by cytotoxin-associated gene A through mediating some molecules including NF- $\kappa$ B, p65 NF- $\kappa$ B, I $\kappa$ Ba, and IL-8 (Lin et al. 2013).

There are numerous reports on the antiinflammatory effects of some active ingredients of *A. camphorata* such as the benzenoid compounds antrocamphin A and 4,7-dimethoxy-5methyl-1,3-benzodioxole (Buccini et al. 2014; Shie et al. 2016). Antrocamphin A derived from *A. camphorata* had an inhibitory effect on the biosynthesis and expression of TNF. 4,7-Dimethoxy-5-methyl-1,3-benzodioxole exerted an antiinflammatory action on LPS-stimulated RAW264.7 cells by inhibiting the production of pro-inflammatory molecules such as NO, TNF- $\alpha$ , and inter IL-1 $\beta$  mainly through downregulating the NF- $\kappa$ B and TLR4 signaling pathway. In addition, the induction of hemoxygenase-1 in RAW264.7 cells was related to the antiinflammatory effect of 4,7-dimethoxy-5-methyl-1,3-benzodioxole.

A. camphorata in wood culture, A. camphorata in solidstate culture, as well as its ingredient ergostatrien- $3\beta$ -ol, decreased in mice, following skin flap surgery, necrosis, and infiltration of inflammatory cells, in both the epidermis and the sub-dermis of the skin flap. The expression of proinflammatory genes such as interleukin-6, inducible nitric oxide synthase, NF- $\kappa$ appaB, and tumor necrosis factor- $\alpha$ was inhibited. The action of A. camphorata against inflammation may have application in hydrocolloid dressings (Tsai et al. 2015). Ergostatrien-3 $\beta$ -ol extracted from the submerged culture of A. camphorata possessed antiinflammatory and antiphotodamaging attributes. Treatment of hairless mice, topically irradiated with ultraviolet B with ergostatrien-3β-ol, disclosed that ergostatrien- $3\beta$ -ol played an antiinflammatory role through restraining the expression level of iNOS, IL-6, MMP-1, and NF-κB. Moreover, ergostatrien-3β-ol also defended the hairless murine skin against ultraviolet-induced photodamage (Kuo et al. 2016b). The crude extract of solid-statecultured mycelia of A. camphorata attenuated, in BALB/c mice, mite allergen Dermatogoides pteronyssinus-induced airway hyperresponsiveness, suppressed total serum immunoglobulin E levels, and inhibited recruitment of inflammatory cells to the bronchoalveolar lavage fluid through cytokine downregulation and modulation of Th1/Th2/Th17 response. The steroids (7,22-ergostadien-3 $\beta$ -ol and 7,9(11),22-ergostatrien-3 $\beta$ -ol) purified from the crude extract downregulated the expression of the proinflammatory cytokines IL-1 $\beta$  and IL-6 in lung macrophages (Kao et al. 2018).

The antiinflammatory effects of antrolone were determined in murine macrophage RAW264.7 cells. The results showed that antrolone significantly decreased the production of NO, prostaglandin E-2, pro-inflammatory cytokine, keratinocyte chemoattractant, the levels of the proteins inducible NO synthase and cyclooxygenase-2, and increased the level of nuclear factor erythroid-2-related factor 2 (Nrf2) and heme oxygenase-1. Moreover, antrolone inhibited the activation of the NF kappa B, MAPK, and AKT pathways (Yen et al. 2018).

### Antiasthmatic activity of polysaccharides from solid-state culture

Polysaccharides extracted from a solid-state culture of *A. camphorata* exhibited the potential for preventing ovalbumin (OVA)-induced asthma. Polysaccharides possessed an immunomodulatory treatment potential against allergic asthma (Liu et al. 2010).

### Antidiabetic and antihyperlipidemic activities of dehydroeburicoic acid, ergostatrien-3β-ol, and antroquinonol

A triterpenoid compound, dehydroeburicoic acid, isolated from the mycelia of A. camphorata, alleviated diabetes and dyslipidemia in streptozotocin-induced diabetic mice. The antidiabetic effect of dehydroeburicoic acid was associated with regulation of the expression level of glucose transporter 4 (GLUT4) protein and AMP-activated protein kinase (AMPK) phosphorylation and thereby lowering the levels of blood glucose and insulin. Dehydroeburicoic acid also exhibited hypolipidemic activity through mediating the expression of fatty acid synthase, peroxisome proliferator-activated receptor (PPAR)- $\alpha$ , and carnitine palmitoyl transferase Ia (Kuo et al. 2015a). Dehydroeburicoic acid also prevented diabetes and dyslipidemia in high-fat diet-fed mice. The mechanism involved regulating the levels of phosphorylation of GLUT4, PPAR- $\alpha$ , and AMPK (Kuo et al. 2016a).

Treating high-fat-diet fed mice with ergostatrien- $3\beta$ -ol ((3S,9S,10R,13R,14R,17R)-17-[(E,2R,5R)-5,6-dimethylhept-3-en-2-yl]-10,13-dimethyl-2,3,4,9,11,12,14,15,16,17-decahydro-1H-cyclopenta [a]phenanthren-3-ol) extracted from the submerged culture broth of *A. camphorata* mycelia revealed that ergostatrien- $3\beta$ -ol effectively reduced the blood levels of glucose and triglyceride. By a series of analysis including body weight, food intake, the blood levels of glucose and glycated hemoglobin, western blotting, and hepatic lipid analysis, the effects of ergostatrien- $3\beta$ -ol were associated with regulating the expression levels of glucose transporter 4, phosphoenol-pyruvate carboxykinase, glucose-6-phosphatase, and AMP-activated protein kinase phosphorylation (Kuo et al. 2015b).

Antroquinonol inhibited dipeptidyl peptidase IV activity as potently as sitagliptin. Antroquinonol and sitagliptin exhibited antihyperglycemic activity in diet-induced obese mice (Hsu et al. 2015). Antroquinonol possessed antidiabetic activity in vitro as well as in vivo (Sulake et al. 2015). Antroquinonol lowered activities of $\alpha$ -amylase and  $\alpha$ -glucosidase (Riyaphan et al. 2018).

## Activation of peroxisome proliferator-activated receptor $\boldsymbol{\alpha}$

Since peroxisome proliferator-activated receptor  $\alpha$  (PPAR $\alpha$ ) is involved in the control of the metabolism of lipids, its agonists like fibrates are therapeutic candidates for hyperlipidemia and other derangements of metabolism. It would be desirable to find agonists with reduced undesirable side effects compared with the existing agonists. Antcins B, H, and K, but not ergostatrien-3 $\beta$ -ol (EK100), are activators of PPAR $\alpha$ . His440 and Tyr314 located in the ligand-binding domain of PPAR $\alpha$  are necessitated to stabilize helix 12. The PPAR $\alpha$ -stimulating activity of antcins depends on binding which requires stabilization of helix 12. Antcins appear to be safe and promising PPAR $\alpha$  agonists (Wang et al. 2019).

## Ameliorative effects of triterpenoids on reproductive function in diabetic rats

It was reported that administration of nanoencapsulated triterpenoids extracted from petri dish-cultured *A. cinnamomea* ameliorated morphology of testicular seminiferous tubules and sperm morphology and motility in diabetic rats, as well as increased the plasma levels of testosterone, luteinizing hormone, and follicle stimulating hormone (Sudirman et al. 2018).

## Hepatoprotective effect of antroquinonol, antcin C, antcin K, dehydroeburicoic acid, eburicoic acid, antrodan, antrosterol, and maleic acid derivative

Antroquinonol inhibited ethanol (100 mM)-induced escalations in activities of alanine and aspartate aminotransferases, reactive oxygen species, nitric oxide, malondialdehyde production, and glutathione reduction in human liver cancer HepG2 cells. Antroquinonol enhanced Nrf-2 activation of nuclear factor (erythroid-derived 2)-like 2 (Nrf-2) and its downstream antioxidant gene heme oxygenase 1 via mitogenactivated protein kinase pathway (Kumar et al. 2011a).

Antcin C exerted a protective action on HepG2 cells against the apoptosis inducing activity of 2,2'-azobis (2amidinopropane) dihydrochloride (AAPH) by attenuating the generation of reactive oxygen species and the proapoptotic factors Bax, caspase-3, caspase-4, caspase 9, caspase-12, cytochrome c, and poly ADP ribose polymerase (PARP). In mice, antcin C attenuated the glutathione reduction and diminished the elevations in activities of hepatic aminotransferases and lipid peroxidation brought about by AAPH. Antioxidant genes such  $as\gamma$ -glutamylcysteine synthetase, hemeoxygenase-1, superoxide dismutase, and NAD(P)H: quinone oxidoreductase 1 were induced through stimulation of Nrf2 transcription. Antcin C stimulated nuclear factor (erythroid-derived 2)-like 2 (Nrf2) through upregulation of and phosphatidylinositol 3-kinase (PI3K) and c-Jun N-terminal kinases (JNK1/2) (Gokila Vani et al. 2013). The chief ingredient antcin K displayed a hepatoprotective action on alcoholintoxicated mice as evidenced by the diminished activities of alanine and aspartate aminotransferases and level of triglyceride (Wu et al. 2019).

Dehydroeburicoic acid and eburicoic acid derived from a solid-state culture of *A. camphorata* fruiting bodies shielded the liver from  $CCl_4$  (carbon tetrachloride)-induced oxidant stress and injury (Huang et al. 2013).

Antrodan, a glycoprotein extracted from *A. camphorata* mycelia, displayed a hepatoprotective effect on the Sprague-Dawley rat model induced by LPS. Low doses of antrodan had a protective effect on liver injury, while high doses produced some side effects (Ker et al. 2014).

Antrosterol extracted from the submerged culture of *A. camphorata* exerted a protective effect on the liver of chronic-alcohol fed mice, which was mainly related to the regulation of alcohol clearance, attenuation of inflammation, and oxidation in addition to lipid balance in the liver (Chang et al. 2017b).

A maleic acid derivative, procured from mycelia of *A. camphorata*, suppressed early hepatic fibrosis brought about by TGF- $\beta$ , mainly involved the inhibition of ROS, inflammation as well as the activation of hepatic stellate cells (Yang et al. 2017b).

### Neuroprotective effect of antroquinonol

Amyloid- $\beta$  peptide (A $\beta$ ) is a fragment cleaved from amyloid precursor protein and a triggering factor for early incidence of Alzheimer's disease. Antroquinonol isolated from *A. camphorata*, a ubiquinone derivative, promoted learning and memory of defective mice and reduced levels of hippocampal A $\beta$  and the degree of astrogliosis, mainly through upregulating the Nrf2 pathway and downregulating the level of histone deacetylase 2 (Chang et al. 2015b).

### **Renoprotective activity of antroquinonol**

Antroquinonol suppressed inflammatory changes of the kidneys and alleviated oxidative stress in a murine renal fibrosis model. It delayed the development of severe kidney damage in mice with induced accelerated and progressive IgA nephropathy. Antroquinonol upregulated the Nrf2 antioxidant pathway, suppressed activation of T cells and NLRP3 inflammasomes, reduced proteinuria and kidney histopathology, and ameliorated kidney function (Yang et al. 2013c).

Antroquinonol suppressed interleukin-1 $\beta$  and tumor necrosis factor- $\alpha$  in RAW 267.4 macrophages stimulated by lipopolysaccharide (Chang et al. 2011). Antroquinonol mitigated urinary protein excretion, kidney dysfunction, and glomerulopathological alterations in glomeruli; alleviated oxidative stress, infiltration of leukocytes, and expression of renal proteins associated with fibrosis; enhanced glutathione peroxidase activity and renal nuclear factor E2-related factor 2; prevented activation of kidney nuclear factor- $\kappa$ B; and reduced renal and serum transforming growth factor- $\beta$ 1 in a murine model of focal segmental glomerulosclerosis (Tsai et al. 2011).

In (NZB × NZW) F1 mice with experimental accelerated severe lupus nephritis brought about by *Salmonella*-type lipopolysaccharide, antroquinonol reduced urinary excretion of blood and protein, cellular crescent formation, fibrinoid necrosis, T cell activation/proliferation and neutrophil infiltration in the glomerulus, production of interleukin-18, nitric oxide, and reactive oxygen species. Renal inflammation and interstitial inflammation in the vicinity of the glomerulus were inhibited, but Treg cell suppression and Nrf2 activation were increased (Tsai et al. 2012). In rats with N $\omega$ -nitro-l-arginine methyl ester-induced hypertension, antroquinonol suppressed the rise of diastolic and systolic blood pressure, serum creatinine, renal malondialdehyde, endothelin-1, angiotensin II, NADPH oxidase heme oxygenase-1, greater nuclear factor erythroid-2, and arteriole thickening (Chen et al. 2018a).

### Antiviral activity of antrodin A

Several isolated constituents including antrodin A displayed an inhibitory effect on herpes simplex virus in the presence as well as in the absence of the antiviral drug acyclovir (He et al. 2016).

### Ameliorative effect of 2,4-dimethoxy-6-methylbenzene-1,3-diol on atopic dermatitis-like lesion

The ameliorative effects of 2,4-dimethoxy-6-methylbenzene-1,3-diol on atopic dermatitis-like lesion and its associated *Staphylococcus aureus* infection were investigated. The results showed that 2,4-dimethoxy-6-methylbenzene-1,3-diol ameliorated atopic dermatitis-like lesion, reduced chemokines in activated keratinocytes, and inhibited the growth of methicillin-resistant *S. aureus* (Yang et al. 2018).

### Safety assessment of Antrodia camphorata

Antroquinonol, provided at 50–600 mg daily for 1 month to patients with metastatic non-small-cell lung cancer, was regarded as well tolerated and generally safe, devoid of dose-limiting toxicities or mortality associated with treatment. The most common treatment-related adverse events, nausea, vomiting, and diarrhea, were the side effects (Lee et al. 2015b).

A. cinnamomea  $\beta$ -glucan provided orally at a daily dose of 2 g/kg for 3 months did not produce harmful effects on the

eyes, feed intake, weights of different organs, increase in body weight, hematological, serum, and urine parameters. Histopathological examination and tests of mutagenicity and genotoxicity did not disclose toxicity of *A. cinnamomea* (Chen et al. 2018d).

However, as the saying goes, "As long as the body intakes drugs, the body will produce three-tenths of the toxicity," the drugs consumed will undergo hepatic metabolism and may increase the burden on the liver if the drug intake is excessive. After all, if a person is healthy, there is no urgent need to take a lot of medicine. At present, there is no specific statement on the daily safe dose of *A. camphorata*; hence, safe dosages still require further study and clinical validation. At the same time, we need to popularize knowledge about *A. camphorata* as well as human pharmacology to consumers through various channels including newspapers, media, radio, and so on, so that people can acquire a better understanding and relevant knowledge and will not undertake a blind pursuit of the socalled God medicine and fall into the snare and be swindled.

### Other activities demonstrated using aqueous extract and alcoholic extract of mycelia or fruiting bodies and fermentation culture broth

In their investigations, some authors preferred to employ purified ingredients whereas others chose to use extracts and fermentation culture medium, probably because the purified ingredients were readily accessible to only some of the researchers. Different assays with dissimilar assay parameters were utilized. Nevertheless, the various investigations yielded different information which individually is useful in its own right. The following information is included because of reasons including (i) the corresponding information on purified ingredients is not available since the purified ingredients have not been assayed and (ii) the information gathered using aqueous extract and alcoholic extract of mycelia or fruiting bodies, and fermentation culture broth, for instance, the apoptotic pathway or proteins affected, is distinct from that collected using purified ingredients. All in all, the following information sheds new light on the activities in A. camphorata. This information will, together with the information above on pure compounds, give a more complete picture and enable one to gain a fuller understanding of the multifarious actions of A. camphorata.

### Effect of extract on oral cancer cells

A. camphorata extracts induced apoptosis in OC-2 human oral cancer cells and also induced the phosphorylation of mitogen activated protein kinases and influenced the regulation of  $Ca^{2+}$  ions (Huang et al. 2009).

## Effects of methanolic and ethanolic extract of fruiting bodies and mycelia on liver cancer cells

The apoptotic mechanism of the methanolic mycelial extract on HepG2 human hepatoma cells entailed activation of caspase-3 and caspase-8 and upregulation of the expression of Fas protein to transmit the death signal (Song et al. 2005). The ethanolic extract of fruiting bodies hampered the migration of human hepatocellular carcinoma cells, which depends on the ERp57, PGK-1, MAPK, and PI3K/Akt cellular pathways (Chen et al. 2015). The extract also brought about cell cycle arrest in Hep3B and HepJ5 human hepatocellular carcinoma cells through heightening the expression of P21s and P27 and triggering apoptosis via activating the expression of caspase-3. The extract potentiated the chemotherapeutic drugs (cisplatin and doxorubicin) (Lin et al. 2015c) and ginger extract (Chen et al. 2018e) in inhibition of hepatocellular carcinoma cells. An ethanolic extract of mycelia triggered apoptosis, induced loss of cell viability, and inhibited invasiveness in cultured liver cancer HepG2 and SMMC-7721 hepatoma cells. The extract induced apoptosis and suppressed JAK2/ STAT3 activation/ phosphorylation in cells as well as tumors. The expression of STAT3-targeted molecules, such as antiapoptotic proteins Bcl-2 and Bcl-xL, and matrix metalloproteinase-2 and matrix metalloproteinase-9, was diminished. The extract suppressed tumor growth but was devoid of toxicity on rats, nude mice, and normal human liverderived cells (Zhu et al. 2018).

# Effects of fermentation culture broth, ethanolic extract of fruiting bodies, and mycelia on breast cancer cells

The fermented culture broth of mycelia exhibited an undermining effect on cell proliferation and an inductive action on cell apoptosis in HER-2/neu-overexpressing MDA-MB-453 and BT-474 breast cancer cells. It did so by elevating the production of reactive oxygen species (ROS) in the cells, downregulating the HER-2/neu signal channel and inhibiting the activation of PI3K/Akt-dependent signaling pathway (Lee et al. 2012). The culture broth suppressed the Twist expression in human breast cancer cells (MDA-MB-231) and inhibited the epithelial-to-mesenchymal transition by downregulating mesenchymal marker proteins. Moreover, it restrained breast cancer metastasis to the lungs (Hseu et al. 2019).

The ethanol extract of fruiting bodies suppressed the growth of MCF-7 and tamoxifen-resistant MCF-7 breast cancer cells and induced their apoptosis. It inhibited the mRNA expression of S-phase kinase-associated protein 2 by increasing the expression of miR-21-5p, miR-26-5p, and miR-30-5p in these breast cancer cells (Lin et al. 2018b).

The ethanolic extract of artificially cultured *A. cinnamomea* exerted an antiproliferative activity toward T47D breast cancer

cells. The cells were arrested at G1 phase of the cell cycle. Autophagy of the cells was also observed. Expression of transcription factor FOXO1 and autophagic markers LC3 II and p62 was upregulated, whereas expression of cell-cycle-related proteins was downregulated. The mechanism also involved endoplasmic reticulum stress. The expression of C/EBP homologous protein, glucose regulating protein 78, and inositol-requiring enzyme  $1\alpha$  was enhanced but histone deacetylase activity was inhibited. The extract suppressed tumor growth without undesirable side effects (Chen et al. 2019d).

## Effect of extract of fruiting bodies and fermentation culture broth on ovarian cancer cells

The fruiting body extract exerted cytotoxicity and induced apoptosis in SKOV-3 and TOV-21G human ovarian cancer cells by mediating the expression of Bcl-2 family proteins; activating the activity of caspase-3, caspase-8, and caspase-9; and upregulating the release of mitochondrial cytochrome c. The crude extract enhanced the cytotoxicity of paclitaxel to both SKOV-3 and TOV-21G ovarian cancers (Liu et al. 2011). The fermentation culture broth retarded the proliferation and expedited the apoptosis of SKOV-3 human ovarian cancer cells. These effects were associated with the generation of ROS, downregulation of the HER-2/neu signaling pathway, and suppression of the PI3K/Akt cascade, which involved the attenuated expression of some proteins including HER-2/neu protein,  $\beta$ -catenin, cyclin D1, and p27 KIP1 (Yang et al. 2013a).

## Effect of extract of fruiting bodies on cervical cancer cells

The crude extract of fruiting body exerted cytotoxicity on HeLa and C-33A human cervical cancer cells and induced apoptosis of these cells, by activating caspase-3, caspase-8, and caspase-9, elevating the release of cytosolic cytochrome c and promoting the expression of the pro-apoptotic proteins Bak, Bim, and Bad (Yang et al. 2013b).

## Effect of ethanolic extract of fruiting bodies on lung cancer cells

The ethanolic extract hindered the proliferation of lung cancer cells through initiating endoplasmic reticulum stress and reduced the excretion of galectin-1 which plays a significant role in the survival and apoptosis of the cells (Wu et al. 2006).

The ethanol extract of mycelia exerted a dose-dependent cytotoxic action and apoptotic action in Lewis lung carcinoma cells. It upregulated Bax and p53 expression, caspase-3 and PARP cleavage, inhibited Bcl-2 and survivin expression, ad suppressed JAK2, and phosphorylated STAT3 levels in LLC

cells. Oral treatment of tumor bearing mice with the ethanol extract inhibited tumor growth and metastasis without altering body weight and serum parameters. The treatment upregulated caspase-3 cleavage and downregulated STAT3 phosphorylation in tumors reduced the growth of human tumor xeno-grafts in nude mice (Huang et al., 2019).

### Effect of ethanol extract on bladder cancer cells

The ethanol extract induced cell cycle arrest (at G2 and M phase) and prevented migration of T24 human bladder cancer cells (Peng et al. 2006). Testing of the ethanolic extract on bladder cancer cells exhibiting different degrees of invasiveness, including RT4 cells, TSGH-8301 cells, and T24 cells, revealed that the growth of these three types of bladder cancer cells could be inhibited to varying degrees and that the different inhibitory effects of the ethanolic extract were largely related to the degree of cell differentiation. The repressive effect of the ethanol extract on RT4 cells might be attributed to the mechanism of replicative senescence, while the growth inhibiting activity of the ethanolic extract on TSGH-8301 cells and T24 cells was probably dependent on the expression of Cdc2 and cyclin B1 complex (Peng et al. 2007).

### Effect of extracts on colorectal cancer cells

*A. cinnamomea* extracts showed cytotoxicity in colorectal cancer Caco-2, Colo205, HCT116, HT29, and SW480 cells. The mechanism involved autophagy mediated by the CHOP/TRB3/Akt/mTOR pathway (Tsai et al. 2018).

### Effect of ethanol extract on leukemia cells

The crude extracts exerted antileukemia activity and expedited immune responses through lengthening the survival rate and precluding the body weight loss in WEHI-3 leukemia bearing BALB/c mice (Lin et al. 2010a).

The ethanol extract derived from wild fruiting bodies of induced HL 60 cells apoptosis, by decreasing histone acetylation via raising the activity of histone deacetyltransferase 1 and lowering the activity of histone acetyltransferase (Lu et al. 2009).

### Effect of fermentation culture broth on melanoma cells

The fermented culture broth from *A. camphorata* restrained the proliferation and migration of the B16F1 and B16F10 melanoma cells in vitro. It triggered apoptosis and prevented metastasis of melanoma cells, mainly by regulation of the Wnt/ $\beta$ -catenin signaling pathways (Hseu et al. 2012).

## Ameliorative effect of ethanol extract on cachexia symptoms

Administration of the ethanol extract greatly rectified the problems of body weight loss and skeletal muscle atrophy induced by gemcitabine and cisplatin in lung tumor-bearing mice (Chen et al. 2018b).

### Immunomodulatory effect of aqueous extract and mycelial powder

When p185neu-overexpressed MBT-2 bladder tumor bearing mice were treated the aqueous extract together with a HER-2/ neu DNA vaccine, the inhibitory effect on tumor cell growth was enhanced and the survival of experimental mice was prolonged. These effects were associated with the immuno-modulatory activity of the *A. camphorata* extract (Huang et al. 2010).

The mycelial powder exhibited immunomodulatory effects. Oral administration of a health food product containing the mycelial powder could upregulate the immune function of the mouse, as evidenced by increased proliferation of splenic lymphocytes, half value of serum hemolysin, phagocytic activity of macrophages, and cytolytic activity of natural killer cells (Chen et al. 2018g; Lin et al. 2018a). In addition, the mycelial powder increased the levels of IL-2, TNF- $\alpha$ , INF- $\gamma$ , GM-CSF, serum OVA-IgG, and OVA-IgM (Chen et al. 2018g).

### Wound healing activity of ethanol extract

The ethanol extract of *A. camphorata* expedited wound healing in Sprague-Dawley rats through accelerating fibroblast proliferation (Amin et al. 2015).

### Antiinflammatory effect of ethanol extract

The antiinflammatory effects of the ethanol extract of A. camphorata grown on germinated brown rice against colitis symptoms induced by sodium dextran sulfate were investigated, which were evaluated through diminishing the disease activity index scores and preventing the body weight loss. The results indicated that the ethanol extract impeded the production of some molecules including nitric oxide and prostaglandin E2 in RAW264.7 cells and curtailed the protein level of some enzymes encompassing inducible nitric oxide synthase and cyclooxygenase-2. The expression of tumor necrosis factor (TNF)- $\alpha$  and interleukin (IL)-6 mRNA was also mitigated following exposure of lipopolysaccharide (LPS)-stimulated RAW264.7 cells to the ethanol extract. These antiinflammatory effects of ethanol extract may be attributed to the regulation of the nuclear factor kappa B (NF-KB) and mitogen-activated protein kinases (MAPK) signaling pathways (Park and Park 2013).

The ethanol extract depressed the expression of some inflammation-related cytokines including TNF- $\alpha$ , IL-1 $\beta$ , and IL-18. These effects were associated with MAPK and NF- $\kappa$ B signal pathways (Huang et al. 2014a).

### Antidiabetic activity of ethanolic extract

The ethanolic extract mitigated lipid peroxidation, oxidative stress, insulin resistance, and hyperglycemia in male rats rendered diabetic by streptozotocin and nicotinamide (Johnson et al. 2019).

### Antiobesity activity of water extract

Mice fed a high-fat diet and receiving treatment with a water extract for 2 months demonstrated a lower body weight, insulin resistance, serum levels of leptin and triglyceride, as well as inflammation markers (interleukin-1 $\beta$  and interleukin-6 and tumor necrosis factor- $\alpha$ ), but higher levels of adipokine than mice fed a high fat diet but without the extract. Treatment with the extract resulted in upregulation, in the small intestine, of the antimicrobial proteins lysozyme C and Reg3g and the intestinal tight junction proteins occludin and zonula occludens-1, giving rise to diminished endotoxin levels in the blood. The Firmicutes/Bacteroidetes ratio was decreased, but the populations of *Akkermansia muciniphila* and other antiinflammatory bacteria were escalated (Chang et al. 2018).

### Antiatherosclerotic activity of fermentation culture medium

Fermentation culture medium attenuated the production of ROS induced by cigarette smoke and lowered DNA damage as well as mitigated atherosclerosis in apolipoprotein E- deficient mice (Yang et al. 2017a).

### Antihypertensive activity of methanol extract of solid-state culture

The methanol extract of solid-state culture effectively lowered the blood pressure of spontaneously hypertensive rats including diastolic blood pressure and systolic blood pressure by restraining the activity of angiotensin-converting enzyme (Liu et al. 2007).

### Ameliorative effects of ethanol extract on reproductive function in diabetic rats

In male rats rendered diabetic by streptozotocin and nicotinamide, the ethanol extract restored seminiferous tubule structure, reduced damage to DNA, and ameliorated mitochondrial membrane potential in spermatozoa (Johnson et al. 2019).

# Hepatoprotective effect of mycelia, fruiting body, fermented product, aqueous extract, ethanolic extract, and *n*-hexane extract

The hepatoprotective activity of *A. camphorata* was evaluated after orally treating acute ethanol-induced rats with fermentation mycelia. *A. camphorata* fermentation culture remarkably suppressed the serum levels of alanine aminotransferase, aspartate transaminase, alkaline phosphatase, and bilirubin, which are all related to liver injury (Lu et al. 2007).

Feeding of alcohol-treated rats with A. camphorata powder accelerated alcohol clearance, which was associated with raised aldehyde dehydrogenase and catalase enzyme systems. In mice treated with ethanol (5 g/kg), heightened activities of alanine and aspartate aminotransferases in the blood, glutathione reduction, lipid peroxidation, enlargement, and degenerative changes in the liver were observed. Administration of ethanolic extracts of A. camphorata mycelia enhanced heme oxygenase-1 and Nrf-2 activation via mitogen-activated protein kinase and attenuated the aforementioned hepatic changes. Thus, the mushroom is useful for the treatment of alcoholic liver diseases (Kumar et al. 2011a). Metalloproteinases upregulated by TNF- $\alpha$  play a part in the development of liver fibrosis, while A. camphorata powder reduced the activity of metalloproteinase-9 and serum and lowered the expression of tumor necrosis factor- $\alpha$ , Krüppel-like factor 6, and transforming growth factor-\beta1; thereby, A. camphorata had a certain protective effect on the liver (Wu et al. 2011).

The fermentation culture exerted a hepatoprotective effect on liver damage and fibrosis in fibrotic rats via suppressing the levels of expression of lipid peroxidation, ROS, iNOS, and TNF- $\alpha$  (Wang et al. 2013).

Activation of hepatic stellate cells played a crucial part in liver fibrosis, while transforming growth factor (TGF)- $\beta$ 1 had a central role in the activation of hepatic stellate cells. The *n*hexane extract of *A. camphorata* inhibited the expression of  $\alpha$ -smooth muscle actin, collagen I, collagen III, and fibronectin stimulated by TGF- $\beta$ 1 in CFSC-8B cells (Geng et al. 2014). An aqueous extract of *A. cinnamomea*, CCM111, exhibited a hepatoprotective action against damage produced by carbon tetrachloride. CCM111 lowered the hepatic activities of alanine and aspartate aminotransferase; suppressed STAT3-, TGF- $\beta$ -, and Wnt-dependent profibrotic and proinflammatory mediators; and minimized collagen deposition and fibrotic changes in the liver (Lin et al. 2019a).

### Nephroprotective action of mycelial extract

The mycelial extract reduced urinary protein excretion and blood urea nitrogen levels and thickness of renal glomerular basement membrane in NZB/W F1 mice with a tendency to develop systemic lupus erythematosus (Chang et al. 2011).

### Neuroprotective effect of extract and fermentation culture

The neuroprotective effect of *A. camphorata* extract in a rat model of ischemic stroke, induced by middle cerebral artery occlusion, may occur through strengthening of the inhibitory effect on the expression of haem oxygenase-1, followed by conspicuous suppression of the expression of other molecules including Bax, iNOS, and caspase-3. Moreover, the formation of hydroxyl radicals was partly inhibited (Yang et al. 2015). The ability of the culture filtrate of *A. camphorata* to counteract amyloid  $\beta$ -peptide-induced neurotoxicity was augmented via increasing the mycelial secretion of some ingredients (Shi et al. 2016).

### **Radioprotective action of ethanolic extract**

The ethanolic extract removed irradiation-triggered reactive oxygen species by upregulating Nrf2 (a basic leucine zipper protein regulating expression of antioxidant proteins) and the downstream redox system enzymes in normal liver CL48 cells. The extract exerted a protective action on irradiation-induced acute hepatitis in tumor-bearing mice and carbon tetrachloride-induced hepatitis in normal mice (Kuo et al. 2019).

## Osteogenic and antiosteoporotic activities of alcohol extract of fruiting bodies

The ability of alcohol extract of fruiting bodies expedited osteogenesis and decreased bone loss in ovariectomized SAMP8 mice (Liu et al. 2016).

### Antifatigue activity of combination of A. camphorata and Panax ginseng

A fixed combination formula of *A. camphorata* and *Panax ginseng*, given orally to male ICR mice for 1 month at the daily dosages of 0.98, 2.95, and 5.90 g/kg, prolonged after a swimming exercise, exhaustive swimming time, enhanced forelimb grip strength, increased serum glucose level, suppressed serum levels of lactate and ammonia, reduced blood urea nitrogen level, and creatine kinase activity (Hsiao et al. 2018).

### Antiplatelet activity of crude extracts

The crude extracts largely mediated calcium and protein kinase C cascade reaction and protein kinase B signaling pathway, thereby inhibiting platelet activation and platelet aggregation to prevent thrombosis (Lu et al. 2014).

### Antibacterial activity of different extracts and antiviral activity of ethanol extract

The ethyl acetate and chloroform extracts exerted a remarkable inhibitory activity on the growth and adhesion of oral bacteria (*Streptococcus mutans*), while the inhibitory effects of 50% ethanol extract and 95% ethanol extract were only moderate (Lien et al. 2014).

## Antioxidant activity of aqueous extract, ethanolic extract, and fermented culture broth

The aqueous mycelial extract inhibited oxidative hemolysis of erythrocytes and lipid or protein peroxidation stimulated by peroxyl radicals as well as attenuated cytoplasmic depletion of the antioxidant tripeptide glutathione (Hseu et al. 2002). Moreover, the aqueous mycelial extract eliminated 2,2-diphenyl-1-picrylhydrozyl free radicals and also exhibited protective activity against DNA damage induced by hydroxyl radicals and antimutagenic effect against direct as well as indirect mutagens (Hsieh et al. 2015). The protective effect of the fermentation culture broth and the aqueous mycelial extract on endothelial cells damage elicited by free radicals was attested to by a decrease of DNA fragmentation, release of cytochrome c, upregulation of caspase-3, and regulation of Bcl-2 and Bax (Hseu et al. 2008).

Supplementing *A. cinnamomea* powder in the diet significantly enhanced the activity of superoxide dismutase and catalase in broiler chicken serum and upregulated the expression of antioxidant genes, including genes of heme oxygenase 1 and glutamate-cysteine ligase catalytic subunit (Lee et al. 2018).

The ethanolic extract removed irradiation-triggered reactive oxygen species by upregulating Nrf2 (a basic leucine zipper protein which regulates expression of antioxidant proteins) and the downstream redox system enzymes in normal liver CL48 cells. The extract exerted a protective action on irradiation-induced acute hepatitis in tumor-bearing mice and carbon tetrachloride-induced hepatitis in normal mice (Kuo et al. 2019).

### Antiphotodamaging activity of fermented extract

The fermented extract relieved cytotoxicity induced by radiation. By comparing the cellular viability of immune spleen cells, human colorectal cancer HT-29 cells, and human breast cancer BT-474 cells, it was revealed that the sensitivities of these three kinds of cells to radiation-inducible damage were different (Cheng et al. 2014).

### Safety assessment of fermented broth of mycelium and mycelial extract

When experimental rats treated with the fermented broth of mycelium were subjected to the acute toxicity test, the  $LD_{50}$ determined exceeded 15 g/kg (Lin et al. 2001). Oral administration of A. cinnamomea to mice at 16 mg to 1.6 g/kg body weight for 3 months had no effects on body weight, hematological and serum biochemical parameters, internal organs, and survival (Chang et al. 2013). The A. cinnamomea health food product Leader Deluxe Antrodia cinnamomea at 0.7-2.8 g/kg/day did not yield anomalous results in the following tests: Ames test, Chinese hamster ovary cells, ratio of immature to total red blood cells and number of micronuclei in immature murine red blood cells, and reproductive and developmental toxicology in pregnant rats and fetuses (Lin et al. 2015b). Oral supplementation of patients with chronic hepatitis C on ribavirin (pegRiba) therapy with A. cinnamomea mycelia did not produce a different outcome due to A. cinnamomea as evidenced by biochemical data, heavy metals, cytokines, and antioxidant status, immune function, and viral response (Chiu et al. 2017).

## Comparison of A. camphorata with other medicinal mushrooms

There are a number of medicinal mushrooms other than A. camphorata. These mushrooms in the raw form and also in the form of nutraceutical products are available. The literature on some of these medicinal mushrooms is fairly voluminous. da Silva de Souza et al. (2017) reviewed the antimutagenic, anticancer, immunomodulating, antiinflammatory, antioxidant, antidiabetic, hepatoprotective, antimicrobial, and antiparasitic activities of Agaricus blazei. Olatunji et al. (2018) and Wong et al. (2018) reviewed the analgesic, antimalarial, antifungal, antibacterial, anti-HIV, cytotoxic, antitumor, immunomodulatory, antiallergic, antiinflammatory, antioxidant, and antidiabetic activities in Cordyceps. The antitumor and immunomodulatory activities of Coriolus versicolor (Piotrowski et al. 2015; Chang et al. 2017a; Wong et al. 2018), as well as antidiabetic, antioxidative, immunomodulatory, and antitumor activities of Grifola frondosa (He et al. 2017, 2018; Rossi et al. 2018) have been reviewed. The diverse activities of Ganoderma lucidum comprising anticancer, antiangiogenic, immunomodulatory, antiinflammatory, antiulcer, analgesic, antidiabetic, antihyperlipidemic, antioxidant, antiatherosclerotic, antiarthritic, antifibrotic, hepatoprotective, antiosteoporotic, antiaging, antimicrobial, and antiherpetic activities have been reviewed (Ahmad, 2018; Rossi et al. 2018; Wong et al. 2018). By comparison, A. camphorata displays a repertoire of activities which is as impressive if not more impressive than those of the other aforementioned medicinal mushrooms. Hence, *A. camphorata* has enormous potential.

## Prospects for the development and utilization of *A. camphorata*

A. camphorata, as a highly prized medicinal fungus, which has tremendous medicinal potential and exploitation value for human health, has been shown in a multitude of studies that it can improve some conditions or treat some diseases, especially cancer. The mushroom demonstrates inhibitory effects on a wide range of cancers. It exerts effects on a variety of organs and produces an array of bioactive molecules with different activities, many of which manifest structural resemblance to steroids. This is reminiscent of the fact that slight variations in the chemical structures of the groups attached to the basic perhydrocyclopentano phenanthrene skeleton typical of cholesterol give rise to different steroid hormones with different activities including the glucocorticoids, mineralocorticoids, progestagens, and sex steroids. Structure function studies may enable the chemists to design potent products. Many mechanisms of its biological actions have been unraveled. Many other exploitable activities of A. camphorata await discovery. However, the growth conditions of the wild A. camphorata are incredibly harsh and its growth rate is extremely slow. Thus, the wild plant resources are scarce, and hence, the current trend is artificial culture (Chen et al. 2019b; Xia et al. 2019; Zhang et al. 2019).  $\alpha$ -Terpineol added to the culture medium of A. cinnamomea mycelia increased the yield and modified the composition of triterpenoids. The triterpenoids produced in the presence of  $\alpha$ -terpineol exhibited a more potent apoptosis-inducing activity than triterpenoids produced without  $\alpha$ -terpineol toward several tumor cell lines. p53, Bax, and caspase-3 in the mitochondrial apoptotic pathway were not affected. However, the production of tyrosyl-DNA phosphodiesterase I and topoisomerase I with a role in DNA repair, at transcriptional as well as translational levels, was downregulated (Zhang et al. 2018a). The optimal conditions for mycelial culture ensuing in an optimal yield await further investigations.

Although the technology of artificial culture has become increasingly mature; nevertheless, there is still some evidence that the content of active ingredients of the cultured *A*. *camphorata* differs from the wild mushroom and its pharma-cological effect is not the same. This is also a problem that calls for solution. The improvement of artificial culture techniques of *A. camphorata* will play a prominent part in future research and development.

Although *A. camphorata* has a diversity of biological activities, the current applications of *A. camphorata* are still mainly focused on the health food products, which are available in the market in the form of capsules of the fruiting body powder, solution of its extract and fermentation broth, or other dosage forms. Therefore, some of the safety assessments of consumption of A. camphorata products and many studies on the biological activity of A. camphorata were conducted only at the level of a mixture such as the fermentation broth or an extract. Because research on activity was based on the mixture, the effective components and content have not been elucidated, and consistency in composition of each batch of A. camphorata fermentation product cannot be guaranteed. Hence, the quality control and effectiveness of the products may not be assured. Reports on the evaluation of the safety of A. camphorata are not comprehensive. If safety evaluation is carried out on a mixture, it may not be easy to draw a firm conclusion because the content of toxic substances may be too low to cause a great impact on the experimental results. In addition, other than the active components of A. camphorata, there may be substances non-essential to human, and longterm intake of these will certainly increase the burden on hepatic metabolism and renal excretion. From the above two points of view, the contemporary research on A. camphorata still has some limitations. Moreover, although a host of studies have demonstrated that extracts derived from A. camphorata fruiting bodies or mycelia exhibited a constellation of effects including anticancer, antileukemia, antiinflammatory, and hepatoprotective activities, most of these experimental findings were derived from in vitro investigations which were not supported by data from clinical trials. Hence, further clinical validation is necessitated if one intends to utilize the active constituents from A. camphorata to prevent or treat certain diseases.

In order to create a greater value of A. camphorata, the development of its industry is bound to implicate upgrading from the traditional dietary health products to high-end new medications in the future, which will bring along more stringent requirements for the related research on A. camphorata. From the traditional health food to the field of medicine, the preliminary research idea is to separate and purify the components from the extract and fermentation broth of A. camphorata, determine the specific components and their structures with biological activity, and then pharmacological and toxicological studies of the active constituents will be carried out. Pharmacological research includes pharmacodynamics and pharmacokinetic studies. The former studies should clarify the mechanism of action of the active ingredients of A. camphorata and indicate the relationship between efficacy and structure, dose, and time. The latter studies focus on the absorption, metabolism, distribution and excretion of the active ingredient, the effects of different routes of administration on efficacy, and the differences in the efficacy of different species. Toxicological studies emphasize on the toxicity and toxicokinetics of the active ingredients and their metabolites, including acute toxicity, long-term toxicity, reproductive toxicity, genotoxicity, topical toxicity, carcinogenicity, dependence, and immunogenicity.

Toxicological studies serve to provide the relationship between the concentration of each active ingredient and its toxicity and determine the target organ of the toxic action and the mechanism of each of its components or its metabolites. From the above, the objective of the preclinical study is to analyze whether the active ingredients of *A. camphorata* have the possibility of new drug development and then it will be followed up by pharmaceutical research and clinical research.

**Funding** This work was supported by the University-Industry Cooperation Project of Fujian Science and Technology Department (2018N5005). We also gratefully acknowledge the award of HMRF research grant (no. 12131221) from Food and Health Bureau, the Government of Hong Kong Special Administrative Region, and a grant from National Natural Science Foundation of China (no. 81471927).

### **Compliance with ethical standards**

**Conflict of interest** The authors declare that there is no conflict of interest.

**Ethical statement** This work does not contain any studies with human participants or animals performed by any of the authors.

### References

- Ahmad MF (2018) Ganoderma lucidum: persuasive biologically active constituents and their health endorsement. Biomed Pharmacother 107:507–519. https://doi.org/10.1016/j.biopha.2018.08.036
- Amin ZA, Ali HM, Alshawsh MA, Darvish PH, Abdulla MA (2015) Application of Antrodia camphorata promotes rat's wound healing in vivo and facilitates fibroblast cell proliferation in vitro. Evid-Based Compl Alt 2015:317693 https://doi.org/10.1155/2015/ 317693
- Ao ZH, Xu ZH, Lu ZM, Xu HY, Zhang XM, Dou WF (2009) Niuchangchih (*Antrodia camphorata*) and its potential in treating liver diseases. J Ethnopharmacol 121(2):194–212. https://doi.org/ 10.1016/j.jep.2008.10.039
- Bamodu OA, Yang CK, Cheng WH, Tzeng DTW, Kuo KT, Huang CC, Deng L, Hsiao M, Lee WH, Yeh CT (2018) 4-Acetyl-antroquinonol B suppresses SOD2-enhanced cancer stem cell-like phenotypes and chemoresistance of colorectal cancer cells by inducing hsa-miR-324 re-expression. Cancers 10(8). https://doi.org/10.3390/ cancers10080269
- Buccini M, Punch KA, Kaskow B, Flematti GR, Skelton BW, Abraham LJ, Piggott MJ (2014) Ethynylbenzenoid metabolites of *Antrodia camphorata*: synthesis and inhibition of TNF expression. Org Biomol Chem 12(7):1100–1113. https://doi.org/10.1039/ c3ob42333f
- Chang JM, Lee YR, Hung LM, Liu SY, Kuo MT, Wen WC, Chen P (2011) An extract of *Antrodia camphorata* mycelia attenuates the progression of nephritis in systemic lupus erythematosus-prone NZB/W F1mice. Evid-Based Compl Alt:1–7. https://doi.org/10. 1093/ecam/nen057
- Chang JB, Wu MF, Lu HF, Chou J, Au MK, Liao NC, Chang CH, Huang YP, Wu CT, Chung JG (2013) Toxicological evaluation of *Antrodia cinnamomea* in BALB/c mice. In Vivo 27(6):739–745
- Chang TC, Yeh CT, Adebayo BOW, Lin YC, Deng L, Rao YK, Huang CC, Lee WH, Wu ATH, Hsiao M, Wu CH, Wang LS, Tzeng YM (2015a) 4-Acetylantroquinonol B inhibits colorectal cancer

tumorigenesis and suppresses cancer stem-like phenotype. Toxicol Appl Pharm 288(2):258–268. https://doi.org/10.1016/j.taap.2015. 07.025

- Chang WH, Chen MC, Cheng IH (2015b) Antroquinonol lowers brain amyloid-beta levels and improves spatial learning and memory in a transgenic mouse model of Alzheimer's disease. Sci Rep 5:15067. https://doi.org/10.1038/srep15067
- Chang Y, Zhang M, Jiang Y, Liu Y, Luo H, Hao C, Zeng P, Zhang L (2017a) Preclinical and clinical studies of *Coriolus versicolor* polysaccharopeptide as an immunotherapeutic in China. Discov Med 23(127):207–219
- Chang YY, Liu YC, Kuo YH, Lin YL, Wu YHS, Chen JW, Chen YC (2017b) Effects of antrosterol from *Antrodia camphorata* submerged whole broth on lipid homeostasis, antioxidation, alcohol clearance, and anti-inflammation in livers of chronic-alcohol fed mice. J Ethnopharmacol 202:200–207. https://doi.org/10.1016/j. jep.2017.03.003
- Chang CJ, Lu CC, Lin CS, Martel J, Ko YF, Ojcius DM, Wu TR, Tsai YH, Yeh TS, Lu JJ, Lai HC, Young JD (2018) Antrodia cinnamomea reduces obesity and modulates the gut microbiota in high-fat diet-fed mice. Int J Obesity 42(2):231–243. https://doi.org/ 10.1038/ijo.2017.149
- Chen YC, Liu YL, Li FY, Chang CL, Wang SY, Lee KY, Li SL, Chen YP, Jinn TR, Tzen JT (2011) Antcin A, a steroid-like compound from *Antrodia camphorata*, exerts anti-inflammatory effect via mimicking glucocorticoids. Acta Pharmacol Sin 32(7):904–911. https://doi. org/10.1038/aps.2011.36
- Chen YY, Liu FC, Wu TS, Sheu MJ (2015) Antrodia cinnamomea inhibits migration in human hepatocellular carcinoma cells: the role of ERp57 and PGK-1. Am J Chin Med 43(8):1671–1696. https://doi. org/10.1142/S0192415x15500950
- Chen JR, Ko J, Yeh WJ, Huang WC, Yang HY (2018a) Renoprotective effects of antroquinonol in rats with N (omega)-nitro-l-arginine methyl ester-induced hypertension. Nutrients 10(10). https://doi.org/10.3390/nu10101521
- Chen MC, Hsu WL, Chou TC (2018b) Anti-cachectic effect of Antrodia cinnamomea extract in lung tumor-bearing mice under chemotherapy. Oncotarget 9(28):19584–19596. https://doi.org/10.18632/ oncotarget.24680
- Chen PC, Chen CC, Ker YB, Chang CH, Chyau CC, Hu ML (2018c) Anti-metastatic effects of antrodan with and without cisplatin on Lewis lung carcinomas in a mouse xenograft model. Int J Mol Sci 19(6). https://doi.org/10.3390/ijms19061565
- Chen SN, Chang CS, Chen S, Soni M (2018d) Subchronic toxicity and genotoxicity studies of *Antrodia* mushroom beta-glucan preparation. Regul Toxicol Pharmacol 92:429–438. https://doi.org/10.1016/j. yrtph.2017.12.022
- Chen SY, Lee YR, Hsieh MC, Omar HA, Teng YN, Lin CY, Hung JH (2018e) Enhancing the anticancer activity of *Antrodia cinnamomea* in hepatocellular carcinoma cells via cocultivation with ginger: the impact on cancer cell survival pathways. Front Pharmacol 9:780 https://doi.org/10.3389/fphar.2018.00780
- Chen YF, Chang CH, Huang ZN, Su YC, Chang SJ, Jan JS (2018f) The JAK inhibitor antcin H exhibits direct anticancer activity while enhancing chemotherapy against LMP1-expressed lymphoma. Leukemia & lymphoma:1–11. https://doi.org/10.1080/10428194. 2018.1512709
- Chen YY, Lo CP, Lin CC, Hsieh YH (2018g) Effects of *Taiwanofungus camphoratus* on non-specific and specific immune activities in mice. Mycology 9(2):129–135. https://doi.org/10.1080/21501203. 2018.1437837
- Chen JH, Wu ATH, Tzeng DTW, Huang CC, Tzeng YM, Chao TY (2019a) Antrocin, a bioactive component from *Antrodia cinnamomea*, suppresses breast carcinogenesis and stemness via downregulation of beta-catenin/Notch1/Akt signaling.

Phytomedicine 52:70–78. https://doi.org/10.1016/j.phymed.2018. 09.213

- Chen L, Wang Z, Zhang B, Ge M, Ng H, Niu Y, Liu L (2019b) Production, structure and morphology of exopolysaccharides yielded by submerged fermentation of *Antrodia cinnamomea*. Carbohydr Polym 205:271–278. https://doi.org/10.1016/j.carbpol. 2018.10.070
- Chen YA, Tzeng DTW, Huang YP, Lin CJ, Lo UG, Wu CL, Lin H, Hsieh JT, Tang CH, Lai CH (2019c) Antrocin sensitizes prostate cancer cells to radiotherapy through inhibiting PI3K/AKT and MAPK signaling pathways. Cancers 11(1). https://doi.org/10.3390/ cancers11010034
- Chen YC, Liu YC, El-Shazly M, Wu TY, Chang JG, Wu YC (2019d) Antrodia cinnamomea, a treasured medicinal mushroom, induces growth arrest in breast cancer cells, T47D cells: new mechanisms emerge. Int J Mol Sci 20(4). https://doi.org/10.3390/ijms20040833
- Cheng PC, Hsu CY, Chen CC, Lee KM (2008) *In vivo* immunomodulatory effects of *Antrodia camphorata* polysaccharides in a T1/T2 doubly transgenic mouse model for inhibiting infection of Schistosoma mansoni. Toxicol Appl Pharm 227(2):291–298. https://doi.org/10.1016/j.taap.2007.10.023
- Cheng PC, Huang CC, Chiang PF, Lin CN, Li LL, Lee TW, Lin B, Chen IC, Chang KW, Fan CK, Luo TY (2014) Radioprotective effects of *Antrodia cinnamomea* are enhanced on immune cells and inhibited on cancer cells. Int J Radiat Biol 90(10):841–852. https://doi.org/10. 3109/09553002.2014.911989
- Chiang PC, Lin SC, Pan SL, Kuo CH, Tsai IL, Kuo MT, Wen WC, Chen P, Guh JH (2010) Antroquinonol displays anticancer potential against human hepatocellular carcinoma cells: a crucial role of AMPK and mTOR pathways. Biochem Pharmacol 79(2):162–171. https://doi.org/10.1016/j.bcp.2009.08.022
- Chiu CC, Chen CC, Huang HY, Chen CC, Lin TW, Chyau CC, Chiou YL, WS K (2017) Complementary efficacy of *Antrodia cinnamomea* mycelia on patients with chronic hepatitis C virus infection: a randomized controlled pilot clinical study. J Food Nutr Res 5(7):481–489
- Chou KC, Wu HL, Lin PY, Yang SH, Chang TL, Sheu F, Chen KH, Chiang BH (2019) 4-Hydroxybenzoic acid serves as an endogenous ring precursor for antroquinonol biosynthesis in *Antrodia cinnamomea*. Phytochemistry 161:97–106. https://doi.org/10.1016/ j.phytochem.2019.02.011
- Chung CH, Yeh SC, Chen CJ, Lee KT (2014) Coenzyme Q0 from Antrodia cinnamomea in submerged cultures induces reactive oxygen species-mediated apoptosis in A549 human lung cancer cells. Evid-Based Compl Alt 2014:246748 https://doi.org/10.1155/2014/ 246748
- da Silva de Souza AC, Correa VG, Goncalves GA, Soares AA, Bracht A, Peralta RM (2017) Agaricus blazei bioactive compounds and their effects on human health: benefits and controversies. Curr Pharm Design 23(19):2807–2834. https://doi.org/10.2174/ 1381612823666170119093719
- Deng JS, Huang SS, Lin TH, Lee MM, Kuo CC, Sung PJ, Hou WC, Huang GJ, Kuo YH (2013) Analgesic and anti-inflammatory bioactivities of eburicoic acid and dehydroeburicoic acid isolated from *Antrodia camphorata* on the inflammatory mediator expression in mice. J Agric Food Chem 61(21):5064–5071. https://doi.org/10. 1021/jf303820k
- Du YC, Chang FR, Wu TY, Hsu YM, El-Shazly M, Chen CF, Sung PJ, Lin YY, Lin YH, Wu YC, Lu MC (2012) Antileukemia component, dehydroeburicoic acid from *Antrodia camphorata* induces DNA damage and apoptosis *in vitro* and *in vivo* models. Phytomedicine 19(8–9):788–796. https://doi.org/10.1016/j.phymed.2012.03.014
- Fa KN, Yang CM, Chen PC, Lee YY, Chyau CC, Hu ML (2015) Antimetastatic effects of antrodan, the *Antrodia cinnamomea* mycelia glycoprotein, in lung carcinoma cells. Int J Biol Macromol 74: 476–482. https://doi.org/10.1016/j.ijbiomac.2015.01.004

- Geethangili M, Tzeng YM (2011) Review of pharmacological effects of *Antrodia camphorata* and its bioactive compounds. Evid-Based Compl Alt 1(17). https://doi.org/10.1093/ecam/nep108
- Geng Y, Wang J, Xie MF, Lu ZM, Xu HY, Shi JS, Xu ZH (2014) Screening and isolation for anti-hepatofibrotic components from medicinal mushrooms using TGF-beta 1-induced live fibrosis in hepatic stellate cells. Int J Med Mushrooms 16(6):529–539
- Gokila Vani M, Kumar KJ, Liao JW, Chien SC, Mau JL, Chiang SS, Lin CC, Kuo YH, Wang SY (2013) Antcin C from Antrodia cinnamomea protects liver cells against free radical-induced oxidative stress and apoptosis in vitro and in vivo through Nrf2-dependent mechanism. Evid Based Complement Alternat Med 2013:296082. https://doi.org/10.1155/2013/296082
- Guan C, Li Q, Song X, Xu W, Li L, Xu A (2017) Antroquinonol exerts immunosuppressive effect on CD8(+) T cell proliferation and activation to resist depigmentation induced by H<sub>2</sub>O<sub>2</sub>. Oxidative Med Cell Longev 2017:9303054 https://doi.org/10.1155/2017/9303054
- He YC, Lu ZH, Shi P, Hao JC, Zhao ZJ, Xie HT, Mao P, Chen SJ (2016) Anti-herpes simplex virus activities of bioactive extracts from *Antrodia camphorata* mycelia. Antivir Ther 21(5):377–383. https://doi.org/10.3851/Imp2988
- He X, Wang X, Fang J, Chang Y, Ning N, Guo H, Huang L, Huang X, Zhao Z (2017) Polysaccharides in *Grifola frondosa* mushroom and their health promoting properties: a review. Int J Biol Macromol 101:910–921. https://doi.org/10.1016/j.ijbiomac.2017.03.177
- He Y, Li X, Hao C, Zeng P, Zhang M, Liu Y, Chang Y, Zhang L (2018) Grifola frondosa polysaccharide: a review of antitumor and other biological activity studies in China. Discov Med 25(138):159–176
- Ho CL, Wang JL, Lee CC, Cheng HY, Wen WC, Cheng HHY, Chen MCM (2014) Antroquinonol blocks Ras and Rho signaling via the inhibition of protein isoprenyltransferase activity in cancer cells. Biomed Pharmacother 68(8):1007–1014. https://doi.org/10.1016/j. biopha.2014.09.008
- Hseu YC, Chang WC, Hseu YT, Lee CY, Yech YJ, Chen PC, Chen JY, Yang HL (2002) Protection of oxidative damage by aqueous extract from *Antrodia camphorata* mycelia in normal human erythrocytes. Life Sci 71(4):469–482. https://doi.org/10.1016/S0024-3205(02) 01686-7
- Hseu YC, Chen SC, Yech YJ, Wang L, Yang HL (2008) Antioxidant activity of *Antrodia camphorata* on free radical-induced endothelial cell damage. J Ethnopharmacol 118(2):237–245. https://doi.org/10. 1016/j.jep.2008.04.004
- Hseu YC, Tsou HT, Kumar KJS, Lin KY, Chang HW, Yang HL (2012) The antitumor activity of *Antrodia camphorata* in melanoma cells: modulation of Wnt/beta-catenin signaling pathways. Evid-Based Compl Alt 2012:197309 https://doi.org/10.1155/2012/197309
- Hseu YC, Chang GR, Pan JY, Rajendran P, Mathew DC, Li ML, Liao JW, Chen WTL, Yang HL (2019) Antrodia camphorata inhibits epithelial-to-mesenchymal transition by targeting multiple pathways in triple-negative breast cancers. J Cell Physiol 234(4):4125–4139. https://doi.org/10.1002/jcp.27222
- Hsiao CY, Hsu YJ, Tung YT, Lee MC, Huang CC, Hsieh CC (2018) Effects of Antrodia camphorata and Panax ginseng supplementation on anti-fatigue properties in mice. J Vet Med Sci 80(2):284– 291. https://doi.org/10.1292/jvms.17-0572
- Hsieh YL, Wu SP, Fang LW, Hwang TS (2015) Effects of Antrodia camphorata extracts on anti-oxidation, anti-mutagenesis and protection of DNA against hydroxyl radical damage. Bmc Complem Altern M 15. https://doi.org/10.1186/s12906-015-0768-3
- Hsu CY, Sulake RS, Huang PK, Shih HY, Sie HW, Lai YK, Chen C, Weng CF (2015) Synthetic (+)-antroquinonol exhibits dual actions against insulin resistance by triggering AMP kinase and inhibiting dipeptidyl peptidase IV activities. Br J Pharmacol 172(1):38–49. https://doi.org/10.1111/bph.12828
- Hu Y, Zhu M, Tian G, Zhao L, Wang H, Ng TB (2017) Isolation of a protease-resistant and pH-stable alpha-galactosidase displaying

hydrolytic efficacy toward raffinose family oligosaccharides from the button mushroom Agaricus bisporus. Int J Biol Macromol

- 104:576–583. https://doi.org/10.1016/j.ijbiomac.2017.06.077 Huang CC, Cheng HH, Wang JL, Cheng JS, Chai KL, Fang YC, Kuo CC, Chu ST, Ho CM, Lin KL, Tsai JY, Jan CR (2009) Effects of *Antrodia camphorata* extracts on the viability, apoptosis, [Ca<sup>2+</sup>](i), and MAPKs phosphorylation of OC2 human oral cancer cells. Chin J Physiol 52(3):128–135. https://doi.org/10.4077/Cjp.2009. Amh013
- Huang CH, Chang CC, Lin CM, Wang ST, Wu MT, Li EIC, Chang HC, Lin CC (2010) Promoting effect of *Antrodia camphorata* as an immunomodulating adjuvant on the antitumor efficacy of HER-2/ neu DNA vaccine. Cancer Immunol Immun 59(8):1259–1272. https://doi.org/10.1007/s00262-010-0852-y
- Huang GJ, Deng JS, Huang SS, Lee CY, Hou WC, Wang SY, Sung PJ, Kuo YH (2013) Hepatoprotective effects of eburicoic acid and dehydroeburicoic acid from *Antrodia camphorata* in a mouse model of acute hepatic injury. Food Chem 141(3):3020–3027. https://doi. org/10.1016/j.foodchem.2013.03.061
- Huang TT, Wu SP, Chong KY, Ojcius DM, Ko YF, Wu YH, Wu CY, Lu CC, Martel J, Young JD, Lai HC (2014a) The medicinal fungus *Antrodia cinnamomea* suppresses inflammation by inhibiting the NLRP3 inflammasome. J Ethnopharmacol 155(1):154–164. https://doi.org/10.1016/j.jep.2014.04.053
- Huang Y, Lin XH, Qiao X, Ji S, Liu KD, Yeh CT, Tzeng YM, Guo D, Ye M (2014b) Antcamphins A-L, ergostanoids from *Antrodia* camphorata. J Nat Prod 77(1):118–124. https://doi.org/10.1021/ np400741s
- Huang CY, Ju DT, Chang CF, Muralidhar Reddy P, Velmurugan BK (2017) A review on the effects of current chemotherapy drugs and natural agents in treating non-small cell lung cancer. Biomedicine 7(4):23–23. https://doi.org/10.1051/bmdcn/2017070423
- Huang TT, Lan YW, Chen CM, Ko YF, Ojcius DM, Martel J, Young JD, Chong KY (2019) *Antrodia cinnamomea* induces anti-tumor activity by inhibiting the STAT3 signaling pathway in lung cancer cells. Sci Rep 9(1). https://doi.org/10.1038/s41598-019-41653-9
- Jedinak A, Dudhgaonkar S, Wu QL, Simon J, Sliva D (2011) Antiinflammatory activity of edible oyster mushroom is mediated through the inhibition of NF-kappa B and AP-1 signaling. Nutr J 10. https://doi.org/10.1186/1475-2891-10-52
- Jiang J, Sliva D (2010) Novel medicinal mushroom blend suppresses growth and invasiveness of human breast cancer cells. Int J Oncol 37(6):1529–1536. https://doi.org/10.3892/ijo 00000806
- Johnson A, Cheng SC, Tsou D, Kong ZL (2019) Attenuation of reproductive dysfunction in diabetic male rats with timber cultured *Antrodia cinnamomea* ethanol extract. Biomed Pharmacother 112: 108684–108684. https://doi.org/10.1016/j.biopha.2019.108684
- Kao ST, Kuo YH, Wang SD, Hong HJ, Lin LJ (2018) Analogous corticosteroids, 9A and EK100, derived from solid-state-cultured mycelium of *Antrodia camphorata* inhibit proinflammatory cytokine expression in macrophages. Cytokine 108:136–144. https://doi.org/10. 1016/j.cyto.2018.03.035
- Ker YB, Peng CC, Chang WL, Chyau CC, Peng RY (2014) Hepatoprotective bioactivity of the glycoprotein, antrodan, isolated from *Antrodia cinnamomea* mycelia. PLoS One 9(4). https://doi. org/10.1371/journal.pone.0093191
- Kumar KJ, Chu FH, Hsieh HW, Liao JW, Li WH, Lin JC, Shaw JF, Wang SY (2011a) Antroquinonol from ethanolic extract of mycelium of *Antrodia cinnamomea* protects hepatic cells from ethanol-induced oxidative stress through Nrf-2 activation. J Ethnopharmacol 136(1): 168–177. https://doi.org/10.1016/j.jep.2011.04.030
- Kumar VB, Yuan TC, Liou JW, Yang CJ, Sung PJ, Weng CF (2011b) Antroquinonol inhibits NSCLC proliferation by altering PI3K/ mTOR proteins and miRNA expression profiles. Mutat Res-Fund Mol M 707(1–2):42–52. https://doi.org/10.1016/j.mrfmmm.2010. 12.009

- Kuo MC, Chang CY, Cheng TL, Wu MJ (2008) Immunomodulatory effect of Antrodia camphorata mycelia and culture filtrate. J Ethnopharmacol 120(2):196–203. https://doi.org/10.1016/j.jep. 2008.08.011
- Kuo YH, Lin CH, Shih CC (2015a) Antidiabetic and antihyperlipidemic properties of a triterpenoid compound, dehydroeburicoic acid, from *Antrodia camphorata in vitro* and in streptozotocin-induced mice. J Agric Food Chem 63(46):10140–10151. https://doi.org/10.1021/ acs.jafc.5b04400
- Kuo YH, Lin CH, Shih CC (2015b) Ergostatrien-3 beta-ol from Antrodia camphorata inhibits diabetes and hyperlipidemia in high-fat-diet treated mice via regulation of hepatic related genes, glucose transporter 4, and AMP-activated protein kinase phosphorylation. J Agric Food Chem 63(9):2479–2489. https://doi.org/10.1021/acs.jafc. 5b00073
- Kuo YH, Lin CH, Shih CC (2016a) Dehydroeburicoic acid from Antrodia camphorata prevents the diabetic and dyslipidemic state via modulation of glucose transporter 4, peroxisome proliferatoractivated receptor alpha expression and AMP-activated protein kinase phosphorylation in high-fat-fed mice. Int J Mol Sci 17(6). https://doi.org/10.3390/ijms17060872
- Kuo YH, Lin TY, You YJ, Wen KC, Sung PJ, Chiang HM (2016b) Antiinflammatory and antiphotodamaging effects of ergostatrien-3 beta-ol, isolated from *Antrodia camphorata*, on hairless mouse skin. Molecules 21(9). https://doi.org/10.3390/molecules21091213
- Kuo TH, Kuo YH, Cho CY, Yao CJ, Lai GM, Chuang SE (2019) Protective effect of Antrodia cinnamomea extract against irradiation-induced acute hepatitis. Int J Mol Sci 20(4). https://doi. org/10.3390/ijms20040846
- Lavi I, Levinson D, Peri I, Nimri L, Hadar Y, Schwartz B (2010a) Orally administered glucans from the edible mushroom *Pleurotus pulmonarius* reduce acute inflammation in dextran sulfate sodiuminduced experimental colitis. Brit J Nutr 103(3):393–402. https:// doi.org/10.1017/s0007114509991760
- Lavi I, Levinson D, Peri I, Tekoah Y, Hadar Y, Schwartz B (2010b) Chemical characterization, antiproliferative and antiadhesive properties of polysaccharides extracted from *Pleurotus pulmonarius* mycelium and fruiting bodies. Appl Microbiol Biotechnol 85(6):1977– 1990. https://doi.org/10.1007/s00253-009-2296-x
- Lee CC, Yang HL, Way TD, Kumar KJS, Juan YC, Cho HJ, Lin KY, Hsu LS, Chen SC, Hseu YC (2012) Inhibition of cell growth and induction of apoptosis by *Antrodia camphorata* in HER-2/neu-overexpressing breast cancer cells through the induction of ROS, depletion of HER–2/neu, and disruption of the PI3K/Akt signaling pathway. Evid-Based Compl Alt 2012:702857https://doi.org/10.1155/2012/ 702857
- Lee CI, Wu CC, Hsieh SL, Lee CL, Chang YP, Chang CC, Wang YZ, Wang JJ (2014a) Anticancer effects on human pancreatic cancer cells of triterpenoids, polysaccharides and 1,3-beta-D-glucan derived from the fruiting body of *Antrodia camphorata*. Food Funct 5(12):3224–3232. https://doi.org/10.1039/c4fo00720d
- Lee TH, Chen CC, Chen JJ, Liao HF, Chang HS, Sung PJ, Tseng MH, Wang SY, Ko HH, Kuo YH (2014b) New and cytotoxic components from *Antrodia camphorata*. Molecules 19(12):21378–21385. https://doi.org/10.3390/molecules191221378
- Lee WT, Lee TH, Cheng CH, Chen KC, Chen YC, Lin CW (2015a) Antroquinonol from *Antrodia camphorata* suppresses breast tumor migration/invasion through inhibiting ERK-AP-1-and AKT-NFkappa B-dependent MMP-9 and epithelial-mesenchymal transition expressions. Food Chem Toxicol 78:33–41. https://doi.org/10.1016/ j.fct.2015.01.012
- Lee YC, Ho CL, Kao WY, Chen YM (2015b) A phase I multicenter study of antroquinonol in patients with metastatic non-small-cell lung cancer who have received at least two prior systemic treatment regimens, including one platinum-based chemotherapy regimen. Mol Clin Oncol 3(6):1375–1380. https://doi.org/10.3892/mco.2015.642

- Lee MT, Lin WC, Wang SY, Lin LJ, Yu B, Lee TT (2018) Evaluation of potential antioxidant and anti-inflammatory effects of *Antrodia cinnamomea* powder and the underlying molecular mechanisms via Nrf2-and NF-kappa B-dominated pathways in broiler chickens. Poultry Sci 97(7):2419–2434. https://doi.org/10.3382/ps/pey076
- Li TY, Chiang BH (2019) 4-Acetylantroquinonol B from *Antrodia cinnamomea* enhances immune function of dendritic cells against liver cancer stem cells. Biomed Pharmacother 109:2262–2269. https://doi.org/10.1016/j.biopha.2018.11.101
- Liaw CC, Chen YC, Huang GJ, Tsai YC, Chien SC, Wu JH, Wang SY, Chao LK, Sung PJ, Huang HC, Kuo YH (2013) Anti-inflammatory lanostanoids and lactone derivatives from *Antrodia camphorata*. J Nat Prod 76(4):489–494. https://doi.org/10.1021/np300443p
- Lien HM, Tseng CJ, Huang CL, Lin YT, Chen CC, Lai YY (2014) Antimicrobial activity of *Antrodia camphorata* extracts against oral bacteria. PLoS One 9(8). https://doi.org/10.1371/journal.pone. 0105286
- Lin WC, Kuo SC, YW W (2001) Effects of 28-days repeated oral administration of the fermented extract of mycelia of *Antrodia camphorata* (CCRC 93032) on rats. J Chin Med 12:293–303
- Lin SY, Sheen LY, Chiang BH, Yang JS, Pan JH, Chang YH, Hsu YM, Chiang JH, Lu CC, Wu CL, Chung JG (2010a) Dietary effect of *Antrodia camphorata* extracts on immune responses in WEHI-3 leukemia BALB/c mice. Nutr Cancer 62(5):593–600. https://doi. org/10.1080/01635580903532341
- Lin YW, Pan JH, Liu RH, Kuo YH, Sheen LY, Chiang BH (2010b) The 4acetylantroquinonol B isolated from mycelium of *Antrodia cinnamomea* inhibits proliferation of hepatoma cells. J Sci Food Agr 90(10):1739–1744. https://doi.org/10.1002/jsfa.4010
- Lin CJ, Rao YK, Hung CL, Feng CL, Lane HY, Tzeng DTW, Hsu PN, Lai CH, Tzeng YM (2013) Inhibition of helicobacter pylori CagAinduced pathogenesis by methylantcinate B from *Antrodia camphorata*. Evid–Based Compl Alt 2013:682418 https://doi.org/ 10.1155/2013/682418
- Lin CC, Chen CC, Kuo YH, Kuo JT, Kumar KJS, Wang SY (2015a) 2,3, 5-Trimethoxy-4-cresol, an anti-metastatic constituent from the solidstate cultured mycelium of *Antrodia cinnamomea* and its mechanism. J Nat Med-Tokyo 69(4):513–521. https://doi.org/10.1007/ s11418-015-0916-6
- Lin CC, Kumar KJS, Liao JW, Kuo YH, Wang SY (2015b) Genotoxic, teratotoxic and oral toxic assessments of *Antrodia cinnamomea* health food product (leader deluxe *Antrodia cinnamomea*). Toxicol Rep 2:1409–1417. https://doi.org/10.1016/j.toxrep.2015.10.007
- Lin LT, Tai CJ, Su CH, Chang FM, Choong CY, Wang CK, Tai CJ (2015c) The ethanolic extract of Taiwanofungus camphoratus (*Antrodia camphorata*) induces cell cycle arrest and enhances cytotoxicity of cisplatin and doxorubicin on human hepatocellular carcinoma cells. Biomed Res Int 2015:415269https://doi.org/10.1155/ 2015/415269
- Lin HC, Lin MH, Liao JH, Wu TH, Lee TH, Mi FL, Wu CH, Chen KC, Cheng CH, Lin CW (2017) Antroquinonol, a ubiquinone derivative from the mushroom *Antrodia camphorata*, inhibits colon cancer stem cell-like properties: insights into the molecular mechanism and inhibitory targets. J Agric Food Chem 65(1):51–59. https:// doi.org/10.1021/acs.jafc.6b04101
- Lin YH, Kuo JT, Chen YY, Kumar KJS, Lo CP, Lin CC, Wang SY (2018a) Immunomodulatory effects of the stout camphor medicinal mushroom, *Taiwanofungus camphoratus* (agaricomycetes)-based health food product in mice. Int J Med Mushrooms 20(9):849– 858. https://doi.org/10.1615/IntJMedMushrooms.2018027389
- Lin YS, Lin YY, Yang YH, Lin CL, Kuan FC, Lu CN, Chang GH, Tsai MS, Hsu CM, Yeh RA, Yang PR, Lee IY, Shu LH, Cheng YC, Liu HT, Lee KD, Chang DC, Wu CY (2018b) Antrodia cinnamomea extract inhibits the proliferation of tamoxifen-resistant breast cancer cells through apoptosis and skp2/microRNAs pathway. Bmc Complem Altern M 18. https://doi.org/10.1186/s12906-018-2204-y

- Lin IY, Chiou YS, Wu LC, Tsai CY, Chen CT, Chuang WC, Lee MC, Lin CC, Lin TT, Chen SC, Pan MH, Ma N (2019a) CCM111 prevents hepatic fibrosis via cooperative inhibition of TGF-beta, Wnt and STAT3 signaling pathways. J Food Drug Anal 27(1):184–194. https://doi.org/10.1016/j.jfda.2018.09.008
- Lin TY, Tseng AJ, Qiu WL, Chao CH, Lu MK (2019b) A sulfated glucan from Antrodia cinnamomea reduces Slug expression through regulation of TGFbeta/AKT/GSK3beta axis in lung cancer. Carbohydr Polym 210:175–184. https://doi.org/10.1016/j.carbpol.2019.01.078
- Liu DZ, Liang YC, Lin SY, Lin YS, Wu WC, Hou WC, Su CH (2007) Antihypertensive activities of a solid-state culture of *Taiwanofungus* camphoratus (Chang-Chih) in spontaneously hypertensive rats. Biosci Biotechnol Biochem 71(1):23–30. https://doi.org/10.1271/ bbb.60268
- Liu KJ, Leu SJ, Su CH, Chiang BL, Chen YL, Lee YL (2010) Administration of polysaccharides from *Antrodia camphorata* modulates dendritic cell function and alleviates allergen-induced T helper type 2 responses in a mouse model of asthma. Immunology 129(3):351–362. https://doi.org/10.1111/j.1365-2567.2009.03175.x
- Liu FS, Yang PY, Hu DN, Huang YW, Chen MJ (2011) Antrodia camphorata induces apoptosis and enhances the cytotoxic effect of paclitaxel in human ovarian cancer cells. Int J Gynecol Cancer 21(7):1172–1179. https://doi.org/10.1097/IGC.0b013e31821f742c
- Liu HY, Huang CF, Li CH, Tsai CY, Chen WH, Wei HJ, Wang MF, Kuo YH, Cheong ML, Deng WP (2016) Osteoporosis recovery by *Antrodia camphorata* alcohol extracts through bone regeneration in SAMP8 mice. Evid-Based Compl Alt doi. https://doi.org/10. 1155/2016/2617868
- Liu F, Wang Y, Zhang K, Wang Y, Zhou R, Zeng Y, Han Y, Ng TB (2017a) A novel polysaccharide with antioxidant, HIV protease inhibiting and HIV integrase inhibiting activities from *Fomitiporia punctata* (P. karst.) murrill (Basidiomycota, hymenochaetales). Int J Biol Macromol 97:339–347. https://doi.org/10.1016/j.ijbiomac. 2017.01.030
- Liu Q, Zhu M, Geng X, Wang H, Ng TB (2017b) Characterization of polysaccharides with antioxidant and hepatoprotective activities from the edible mushroom *Oudemansiella radicata*. Molecules 22(2). https://doi.org/10.3390/molecules22020234
- Liu HW, Su YK, Bamodu OA, Hueng DY, Lee WH, Huang CC, Deng L, Hsiao M, Chien MH, Yeh CT, Lin CM (2018a) The disruption of the beta-catenin/TCF-1/STAT3 signaling axis by 4-acetylantroquinonol B inhibits the tumorigenesis and cancer stem-cell-like properties of glioblastoma cells, *in vitro* and *in vivo*. Cancers (Basel) 10(12). https://doi.org/10.3390/cancers10120491
- Liu Y, Yang A, Qu Y, Wang Z, Zhang Y, Liu Y, Wang N, Teng L, Wang D (2018b) Ameliorative effects of *Antrodia cinnamomea* polysaccharides against cyclophosphamide-induced immunosuppression related to Nrf2/HO-1 signaling in BALB/c mice. Int J Biol Macromol 116:8–15. https://doi.org/10.1016/j.ijbiomac.2018.04.178
- Lu ZM, Tao WY, Zou XL, Fu HZ, Ao ZH (2007) Protective effects of mycelia of Antrodia camphorata and Armillariella tabescens in submerged culture against ethanol-induced hepatic toxicity in rats. J Ethnopharmacol 110(1):160–164. https://doi.org/10.1016/j.jep. 2006.09.029
- Lu MC, Du YC, Chuu JJ, Hwang SL, Hsieh PC, Hung CS, Chang FR, Wu YC (2009) Active extracts of wild fruiting bodies of *Antrodia camphorata* (EEAC) induce leukemia HL 60 cells apoptosis partially through histone hypoacetylation and synergistically promote anticancer effect of trichostatin A. Arch Toxicol 83(2):121–129. https://doi.org/10.1007/s00204-008-0337-3
- Lu WJ, Lin SC, Lan CC, Lee TY, Hsia CH, Huang YK, Lee HC, Sheu JR (2014) Effect of *Antrodia camphorata* on inflammatory arterial thrombosis-mediated platelet activation: the pivotal role of protein kinase C. Sci World J 2014:745802. https://doi.org/10.1155/2014/ 745802

- Lu MK, Lin TY, Chang CC (2018) Chemical identification of a sulfated glucan from *Antrodia cinnamomea* and its anti-cancer functions via inhibition of EGFR and mTOR activity. Carbohyd Polym 202:536– 544. https://doi.org/10.1016/j.carbpol.2018.09.009
- Meng LM, Pai MH, Liu JJ, Yeh SL (2012) Polysaccharides from extracts of *Antrodia camphorata* mycelia and fruiting bodies modulate inflammatory mediator expression in mice with polymicrobial sepsis. Nutrition 28(9):942–949. https://doi.org/10.1016/j.nut.2012.01.006
- Nakamura N, Hirakawa A, Gao JJ, Kakuda H, Shiro M, Komatsu Y, Sheu CC, Hattori M (2004) Five new maleic and succinic acid derivatives from the mycelium of *Antrodia camphorata* and their cytotoxic effects on LLC tumor cell line. J Nat Prod 67(1):46–48. https://doi.org/10.1021/np030293k
- Olatunji OJ, Tang J, Tola A, Auberon F, Oluwaniyi O, Ouyang Z (2018) The genus Cordyceps: an extensive review of its traditional uses, phytochemistry and pharmacology. Fitoterapia 129:293–316. https://doi.org/10.1016/j.fitote.2018.05.010
- Pan WL, Wong JH, Fang EF, Chan YS, Ye XJ, Ng TB (2013) Differential inhibitory potencies and mechanisms of the type I ribosome inactivating protein marmorin on estrogen receptor (ER)-positive and ER-negative breast cancer cells. Biochim Biophys Acta 1833(5):987–996. https://doi.org/10.1016/j.bbamcr.2012.12.013
- Park DK, Park HJ (2013) Ethanol extract of Antrodia camphorata grown on germinated brown rice suppresses inflammatory responses in mice with acute DSS-induced colitis. Evid-Based Compl Alt 2013: 914524 https://doi.org/10.1155/2013/914524
- Peng CC, Chen KC, Peng RY, Su CH, Hsieh-Li HM (2006) Human urinary bladder cancer T24 cells are susceptible to the Antrodia camphorata extracts. Cancer Lett 243(1):109–119. https://doi.org/ 10.1016/j.canlet.2005.11.021
- Peng CC, Chen KC, Peng RY, Chyau CC, Su CH, Hsieh-Li HM (2007) Antrodia camphorata extract induces replicative senescence in superficial TCC, and inhibits the absolute migration capability in invasive bladder carcinoma cells. J Ethnopharmacol 109(1):93–103. https://doi.org/10.1016/j.jep.2006.07.009
- Peng CC, Lin YT, Chen KC, Chyau CC, Peng RY (2015) Antrodan, a beta-glucan obtained from *Antrodia cinnamomea* mycelia, is beneficial to benign prostate hyperplasia. Food Funct 6(2):635–645. https://doi.org/10.1039/c4fo00472h
- Perera N, Yang FL, Chang CM, Lu YT, Zhan SH, Tsai YT, Hsieh JF, Li LH, Hua KF, Wu SH (2017) Galactomannan from *Antrodia cinnamomea* enhances the phagocytic activity of macrophages. Org Lett 19(13):3486–3489. https://doi.org/10.1021/acs.orglett. 7b01468
- Perera N, Yang FL, Lu YT, Li LH, Hua KF, Wu SH (2018) Antrodia cinnamomea galactomannan elicits immuno-stimulatory activity through Toll-like receptor 4. Int J Biol Sci 14(10):1378–1388. https://doi.org/10.7150/ijbs.24564
- Piotrowski J, Jedrzejewski T, Kozak W (2015) Immunomodulatory and antitumor properties of polysaccharide peptide (PSP). Postepy Hig Med Dosw 69:91–97
- Riyaphan J, Jhong CH, Lin SR, Chang CH, Tsai MJ, Lee DN, Sung PJ, Leong MK, Weng CF (2018) Hypoglycemic efficacy of docking selected natural compounds against alpha-glucosidase and alphaamylase. Molecules 23(9). https://doi.org/10.3390/ molecules23092260
- Rossi P, Difrancia R, Quagliariello V, Savino E, Tralongo P, Randazzo CL, Berretta M (2018) B-glucans from *Grifola frondosa* and *Ganoderma lucidum* in breast cancer: an example of complementary and integrative medicine. Oncotarget 9(37):24837–24856. https://doi.org/10.18632/oncotarget.24984
- Schwartz B, Hadar Y (2014) Possible mechanisms of action of mushroom-derived glucans on inflammatory bowel disease and associated cancer. Ann Transl Med 2(2):19–19. https://doi.org/10. 3978/j.issn.2305-5839.2014.01.03

- Shi YC, Yang SY, Lee DY, Lee C (2016) Increasing anti-A beta-induced neurotoxicity ability of *Antrodia camphorata*-fermented product with deep ocean water supplementary. J Sci Food Agr 96(14): 4690–4701. https://doi.org/10.1002/jsfa.7687
- Shie PH, Wang SY, Lay HL, Huang GJ (2016) 4,7-Dimethoxy-5-methyl-1,3-benzodioxole from Antrodia camphorata inhibits LPS-induced inflammation via suppression of NF-kappa B and induction HO-1 in RAW264.7 cells. Int Immunopharmacol 31:186–194. https://doi. org/10.1016/j.intimp.2015.12.030
- Song TY, Hsu SL, Yeh CT, Yen GC (2005) Mycelia from Antrodia camphorata in submerged culture induce apoptosis of human hepatoma HepG2 cells possibly through regulation of Fas pathway. J Agric Food Chem 53(14):5559–5564. https://doi.org/10.1021/ jf050329+
- Song AR, Qin D, Zhao C, Sun XL, Huang F, Kong C, Yang S (2014) Immunomodulatory effect of polysaccharides extracted from the medicinal mushroom *Antrodia camphorata* (higher Basidiomycetes) in specific pathogen-free chickens. Int J Med Mushrooms 16(1):95–103
- Sudirman S, Hsu YH, Johnson A, Tsou D, Kong ZL (2018) Amelioration effects of nanoencapsulated triterpenoids from petri dish-cultured *Antrodia cinnamomea* on reproductive function of diabetic male rats. Int J Nanomedicine 13:5059–5073. https://doi.org/10.2147/ ijn.S172906
- Sulake RS, Lin HH, Hsu CY, Weng CF, Chen C (2015) Synthesis of (+)antroquinonol: an antihyperglycemic agent. J Org Chem 80(12): 6044–6051. https://doi.org/10.1021/acs.joc.5b00345
- Thiyagarajan V, Tsai MJ, Weng CF (2015) Antroquinonol targets FAKsignaling pathway suppressed cell migration, invasion, and tumor growth of C6 glioma. PLoS One 10(10):e0141285. https://doi.org/ 10.1371/journal.pone.0141285
- Tsai WC, Rao YK, Lin SS, Chou MY, Shen YT, Wu CH, Geethangili M, Yang CC, Tzeng YM (2010) Methylantcinate A induces tumor specific growth inhibition in oral cancer cells via Bax-mediated mitochondrial apoptotic pathway. Bioorg Med Chem Lett 20(20):6145– 6148. https://doi.org/10.1016/j.bmcl.2010.08.006
- Tsai PY, Ka SM, Chao TK, Chang JM, Lin SH, Li CY, Kuo MT, Chen P, Chen A (2011) Antroquinonol reduces oxidative stress by enhancing the Nrf2 signaling pathway and inhibits inflammation and sclerosis in focal segmental glomerulosclerosis mice. Free Radic Biol Med 50(11):1503–1516. https://doi.org/10.1016/j.freeradbiomed. 2011.02.029
- Tsai PY, Ka SM, Chang JM, Lai JH, Dai MS, Jheng HL, Kuo MT, Chen P, Chen A (2012) Antroquinonol differentially modulates T cell activity and reduces interleukin-18 production, but enhances Nrf2 activation, in murine accelerated severe lupus nephritis. Arthritis Rheum 64(1):232–242. https://doi.org/10.1002/art.33328
- Tsai TC, Tung YT, Kuo YH, Liao JW, Tsai HC, Chong KY, Chen HL, Chen CM (2015) Anti-inflammatory effects of *Antrodia camphorata*, a herbal medicine, in a mouse skin ischemia model. J Ethnopharmacol 159:113–121. https://doi.org/10.1016/j.jep.2014. 11.015
- Tsai DH, Chung CH, Lee KT (2018) *Antrodia cinnamomea* induces autophagic cell death via the CHOP/TRB3/Akt/mTOR pathway in colorectal cancer cells. Sci Rep 8(1):17424. https://doi.org/10.1038/ s41598-018-35780-y
- Tu SH, Wu CH, Chen LC, Huang CS, Chang HW, Chang CH, Lien HM, Ho YS (2012) *In vivo* antitumor effects of 4,7-dimethoxy-5-methyl-1,3-benzodioxole isolated from the fruiting body of *Antrodia camphorata* through activation of the p53-mediated p27/Kip1 signaling pathway. J Agric Food Chem 60(14):3612–3618. https://doi. org/10.1021/jf300221g
- Wang LC, Kuo IU, Tsai TY, Lee CL (2013) Antrodia camphoratafermented product cultured in deep ocean water has more liver protection against thioacetamide-induced fibrosis. Appl Microbiol

Biotechnol 97(23):9955–9967. https://doi.org/10.1007/s00253-013-5214-1

- Wang SC, Lee TH, Hsu CH, Chang YJ, Chang MS, Wang YC, Ho YS, Wen WC, Lin RK (2014) Antroquinonol D, isolated from *Antrodia camphorata*, with DNA demethylation and anticancer potential. J Agric Food Chem 62(24):5625–5635. https://doi.org/10.1021/ jf4056924
- Wang YJ, Lee SC, Hsu CH, Kuo YH, Yang CC, Lin FJ (2019) Antcins, triterpenoids from *Antrodia cinnamomea*, as new agonists for peroxisome proliferator-activated receptor alpha. J Food Drug Anal 27(1):295–304. https://doi.org/10.1016/j.jfda.2018.11.004
- Wasser SP (2014) Medicinal mushroom science: current perspectives, advances, evidences, and challenges. Biom J 37(6):345–356. https://doi.org/10.4103/2319-4170.138318
- Wasser SP (2017) Medicinal mushrooms in human clinical studies. Part I. Anticancer, oncoimmunological, and immunomodulatory activities: a review. Int J Med Mushrooms 19(4):279–317. https://doi.org/10. 1615/IntJMedMushrooms.v19.i4.10
- Wong JH, Ng TB, Cheung RCF, Ye XJ, Wang HX, Lam SK, Lin P, Chan YS, Fang EF, Ngai PHK, Xia LX, Ye XY, Jiang Y, Liu F (2010) Proteins with antifungal properties and other medicinal applications from plants and mushrooms. Appl Microbiol Biotechnol 87(4): 1221–1235. https://doi.org/10.1007/s00253-010-2690-4
- Wong JH, Sze SCW, Ng TB, Cheung RCF, Tam C, Zhang KY, Dan X, Chan YS, Shing Cho WC, Ng CCW, Waye MMY, Liang W, Zhang J, Yang J, Ye X, Lin J, Ye X, Wang H, Liu F, Chan DW, Ngan HYS, Sha O, Li G, Tse R, Tse TF, Chan H (2018) Apoptosis and anticancer drug discovery: the power of medicinal fungi and plants. Curr Med Chem 25(40):5613–5630. https://doi.org/10.2174/ 0929867324666170720165005
- Wu SH, Ryvarden L, Chang TT (1997) Antrodia camphorata (niu-changchih), new combination of a medicinal fungus in Taiwan. Bot Bull Acad Sinica 38(4):273–275
- Wu H, Pan CL, Yao YC, Chang SS, Li SL, Wu TF (2006) Proteomic analysis of the effect of *Antrodia camphorata* extract on human lung cancer A549 cell. Proteomics 6(3):826–835. https://doi.org/10. 1002/pmic.200401341
- Wu MT, Tzang BS, Chang YY, Chiu CH, Kang WY, Huang CH, Chen YC (2011) Effects of *Antrodia camphorata* on alcohol clearance and antifibrosis in livers of rats continuously fed alcohol. J Agric Food Chem 59(8):4248–4254. https://doi.org/10.1021/jf104561h
- Wu MD, Cheng MJ, Yech YJ, Yuan GF, Chen JJ (2013) Inhibitory effects of maleimide derivatives from the mycelia of the fungus *Antrodia cinnamomea* BCRC 36799 on nitric oxide production in lipopolysaccharide (LPS)-activated RAW264.7 macrophages. Chem Biodivers 10(3):434–441. https://doi.org/10.1002/cbdv.201200258
- Wu Y, Tian WD, Gao S, Liao ZJ, Wang GH, Lo JM, Lin PH, Zeng DQ, Qiu DR, Liu XZ, Zhou M, Lin T, Chen HF (2019) Secondary metabolites of petri-dish cultured *Antrodia camphorata* and their hepatoprotective activities against alcohol-induced liver injury in mice. Chin J Nat Med 17(1):33–42. https://doi.org/10.1016/s1875-5364(19)30007-x
- Xia Y, Chen Y, Liu X, Zhou X, Wang Z, Wang G, Xiong Z, Ai L (2019) Enhancement of antroquinonol production during batch fermentation using pH control coupled with an oxygen vector. J Sci Food Agr 99(1):449–456. https://doi.org/10.1002/jsfa.9206
- Yang SS, Wang GJ, Wang SY, Lin YY, Kuo YH, Lee TH (2009) New constituents with iNOS inhibitory activity from mycelium of *Antrodia camphorata*. Planta Med 75(5):512–516. https://doi.org/ 10.1055/s-0029-1185305
- Yang HL, Lin KY, Juan YC, Kumar KJS, Way TD, Shen PC, Chen SC, Hseu YC (2013a) The anti-cancer activity of *Antrodia camphorata* against human ovarian carcinoma (SKOV-3) cells via modulation of HER-2/neu signaling pathway. J Ethnopharmacol 148(1):254–265. https://doi.org/10.1016/j.jep.2013.04.023

- Yang PY, Hu DN, Liu FS (2013b) Cytotoxic effect and induction of apoptosis in human cervical cancer cells by *Antrodia camphorata*. Am J Chin Med 41(5):1169–1180. https://doi.org/10.1142/ S0192415x13500791
- Yang SM, Ka SM, Hua KF, Wu TH, Chuang YP, Lin YW, Yang FL, Wu SH, Yang SS, Lin SH, Chang JM, Chen A (2013c) Antroquinonol mitigates an accelerated and progressive IgA nephropathy model in mice by activating the Nrf2 pathway and inhibiting T cells and NLRP3 inflammasome. Free Radical Bio Med 61:285–297. https://doi.org/10.1016/j.freeradbiomed.2013.03.024
- Yang PS, Lin PY, Chang CC, Yu MC, Yen TL, Lan CC, Jayakumar T, Yang CH (2015) Antrodia camphorata potentiates neuroprotection against cerebral ischemia in rats via downregulation of iNOS/HO-1/ Bax and activated caspase-3 and inhibition of hydroxyl radical formation. Evid-Based Compl Alt 2015:232789 https://doi.org/10. 1155/2015/232789
- Yang HL, Korivi M, Chen CH, Peng WJ, Chen CS, Li ML, Hsu LS, Liao JW, Hseu YC (2017a) Antrodia camphorata attenuates cigarette smoke-induced ROS production, DNA damage, apoptosis, and inflammation in vascular smooth muscle cells, and atherosclerosis in ApoE–deficient mice. Environ Toxicol 32(8):2070–2084. https:// doi.org/10.1002/tox.22422
- Yang KL, Chang WT, Hong MY, Hung KC, Chuang CC (2017b) Prevention of TGF-beta-induced early liver fibrosis by a maleic acid derivative antioxidant through suppression of ROS, inflammation and hepatic stellate cells activation. PLoS One 12(4). https://doi. org/10.1371/journal.pone.0174008
- Yang SC, Huang TH, Chiu CH, Chou WL, Alalaiwe A, Yeh YC, Su KW, Fang JY (2018) The atopic dermatitis-like lesion and the associated MRSA infection and barrier dysfunction can be alleviated by 2,4dimethoxy-6-methylbenzene-1,3-diol from *Antrodia camphorata*. J Dermatol Sci 92(2):188–196. https://doi.org/10.1016/j.jdermsci. 2018.09.002
- Yeh CT, Huang WC, Rao YK, Ye M, Lee WH, Wang LS, Tzeng DTW, Wu CH, Shieh YS, Huang CYF, Chen YJ, Hsiao M, Wu ATH, Yang Z, Tzeng YM (2013) A sesquiterpene lactone antrocin from *Antrodia camphorata* negatively modulates JAK2/STAT3 signaling via microRNA let-7c and induces apoptosis in lung cancer cells. Carcinogenesis 34(12):2918–2928. https://doi.org/10.1093/carcin/ bgt255
- Yen IC, Shi LS, Chung MC, Ahmetaj-Shala B, Chang TC, Lee SY (2018) Antrolone, a novel benzoid derived from Antrodia cinnamomea, inhibits the LPS-induced inflammatory response in RAW264.7

macrophage cells by balancing the NF-kappa B and Nrf2 pathways. Am J Chin Med 46(6):1297–1313. https://doi.org/10.1142/S0192415x18500684

- Yeung SY, Piggott MJ (2018) Reprint of: Antiproliferative activity of the Antrodia camphorata secondary metabolite 4,7-dimethoxy-5methylbenzo [d][1,3]dioxole and analogues. Fitoterapia 126:40– 44. https://doi.org/10.1016/j.fitote.2018.04.001
- Yu CC, Chiang PC, Lu PH, Kuo MT, Wen WC, Chen P, Guh JH (2012) Antroquinonol, a natural ubiquinone derivative, induces a cross talk between apoptosis, autophagy and senescence in human pancreatic carcinoma cells. J Nutr Biochem 23(8):900–907. https://doi.org/10. 1016/j.jnutbio.2011.04.015
- Zhang Y, Li D, Wang Z, Zang W, Rao P, Liang Y, Mei Y (2018a) Alphaterpineol affects synthesis and antitumor activity of triterpenoids from *Antrodia cinnamomea* mycelia in solid-state culture. Food Funct 9(12):6517–6525. https://doi.org/10.1039/c8fo02079e
- Zhang Y, Wang Z, Li D, Zang W, Zhu H, Wu P, Mei Y, Liang Y (2018b) A polysaccharide from *Antrodia cinnamomea* mycelia exerts antitumor activity through blocking of TOP1/TDP1-mediated DNA repair pathway. Int J Biol Macromol 120:1551–1560. https://doi.org/10. 1016/j.ijbiomac.2018.09.162
- Zhang BB, Guan YY, Hu PF, Chen L, Xu GR, Liu L, Cheung PCK (2019) Production of bioactive metabolites by submerged fermentation of the medicinal mushroom *Antrodia cinnamomea*: recent advances and future development. Crit Rev Biotechnol:1–14. https:// doi.org/10.1080/07388551.2019.1577798
- Zhou R, Han Y-J, Zhang M-H, Zhang K-R, Ng TB, Liu F (2017) Purification and characterization of a novel ubiquitin-like antitumour protein with hemagglutinating and deoxyribonuclease activities from the edible mushroom *Ramaria botrytis*. AMB Express 7. https://doi.org/10.1186/s13568-017-0346-9
- Zhou R, Liu Z, Zhang YN, Ng TB, Liu F (2018) Research progress on bioactive proteins from edible mushrooms. Curr Protein Pept Sci. https://doi.org/10.2174/1389203719666180613090710
- Zhu PL, Fu XQ, Li JK, Tse AK, Guo H, Yin CL, Chou JY, Wang YP, Liu YX, Chen YJ, Hossen MJ, Zhang Y, Pan SY, Zhao ZJ, Yu ZL (2018) *Antrodia camphorata* mycelia exert anti-liver cancer effects and inhibit stat3 signaling *in vitro* and *in vivo*. Front Pharmacol 9: 1449. https://doi.org/10.3389/fphar.2018.01449

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.