Contamination Profiles of Antifouling Biocides in Selected Coastal Regions of Malaysia

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Abstract The concentrations of butyltins (BTs) in sediment from Peninsular Malaysia along the Strait of Malacca and their spatial distribution are discussed. The concentrations of BTs were high in the southern part of Peninsular Malaysia where there is a lot of ship traffic, because trade is prosperous. The concentrations of monobutyltin (MBT), dibutyltin (DBT), and tributyltin (TBT) in sediment from the coastal waters of Peninsular Malaysia were in the range $4.1-242 \ \mu g/kg dw$, respectively. A higher percentage of TBT was observed in the area where TBT concentrations were high. The

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Center for International Cooperation, Ocean Research Institute, The University of Tokyo, 1-15-1, Minamidai, Nakano, Tokyo 164-8639, Japan concentrations of monophenyltin (MPT), diphenyltin (DPT), and triphenyltin (TPT) were in the range <0.1-121 µg/kg dw, 0.4-27 µg/kg dw, and 0.1-34 µg/kg dw in sediment from Peninsular Malaysia, respectively. MPT was the dominant phenyltin species. MBT, DBT, and TBT in green mussel (Perna viridis) samples were detected in the range 41-102 µg/kg, 3-5 µg/kg, and 8- $32 \mu g/kg$, respectively. A tolerable average residue level (TARL) was estimated at 20.4 µg/kg from a tolerable daily intake (TDI) of 0.25 µg TBTO/kg body weight/day. The maximum value of TBT detected in green mussel samples was the value near the TARL. TPTs were not detected in green mussel samples. The concentrations of Diuron and Irgarol 1051 in sediment from Peninsular Malaysia were in the range $<0.1-5 \mu g/kg$ dw and <0.1-14 µg/kg dw, respectively. High concentrations of these compounds were observed in locations where the concentrations of TBT were high. Sea Nine 211, Dichlofluanid, and Pyrithiones were not detected in sediment. The concentrations of antifouling biocides in Melaka and the Strait of Johor were investigated in detail. BTs were found in similar concentrations among all sampling sites from Melaka, indicating that BT contamination spread off the coast. However, Sea Nine 211, Diuron, and Irgarol 1051 in the sediment from Melaka were high at the mouth of the river. BT concentrations at the Strait of Johor were higher than those in Peninsular Malaysia and Melaka and were high at the narrowest locations with poor flushing of water. The concentrations of antifouling biocides were compared among Malaysia, Thailand, and Vietnam. A higher concentration and wide variations of TBT and TPT in sediment from Malaysia were observed among these countries. The Irgarol 1051 concentrations in sediment from Malaysia were higher than those in Thailand and Vietnam.

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Organotin (OT) compounds have been utilized as an active biocide causing deleterious effects such as endocrine disruption in nontarget marine organisms (Bryan and Gibbs 1991; Laughlin and Linden 1985), and further environmental studies have indicated OT contamination of the marine environment on a worldwide scale (Clark et al. 1988). In the 1980s, the use of tributyltin (TBT) was regulated in some developed countries such as England, France and the United States, but TBT has still been detected in the aquatic environment of these countries at a contamination level that causes adverse effects on the aquatic environment. In developing countries, there have been no controls on the usage of TBT. In October 2001, the International Maritime Organization (IMO) adopted the Internal Convention on the Control of Harmful Antifouling Systems (AFS Convention), which prohibited the use of OTs as active ingredients in antifouling systems for ships. Following the international restrictions on the use of OT-based antifoulants, paint manufacturers have developed many alternative products.

In the last 20 years, the economies of some Southeast Asian countries have been growing rapidly. Environmental pollution is closely tied to economic growth, because human and industrial activities increase and trading flourishes when there is an increase of economic activity. In fact, the coastal areas of Thailand, the Philippines, and Vietnam are contaminated by chemical compounds such as persistant organic pollutants (POPs) and antifouling biocides (Kan-atireklap et al. 1997a, b, 1998; Lee et al. 1997; Monirith et al. 2003; Prudente et al. 1999). Malaysia is the fastest growing country in Southeast Asia., because Malaysia has an abundance of mineral resources and beautiful natural areas. The tourism industry and trade businesses are thriving, and the Strait of Malacca and the Strait of Johor in Malaysia have become the world's busiest waterways. Recent surveys show that the coastal waters of Malaysia are polluted by BTs (Hashimoto et al. 1998; Sudaryant et al. 2002, 2004; Tong et al. 1996).

Furthermore, contaminations by alternative biocides are also of concern, because many ocean liners enter the coastal areas of Malaysia. However, to date there have been no reports of pollution by alternative biocides.

In this study, the concentrations of OTs and alternative biocides are reported in sediment and green mussels from Malaysia and the distribution of antifouling biocides in Malaysia is also discussed.

Materials and Methods

Sampling Description

Malaysia is located in the center of Southeast Asia and consists of two geographical regions: Peninsular Malaysia

and Malavsian Borneo divided by the South China Sea. Mining has declined, but the electronic industry has flourished, and as a result, Malaysia has become famous in Southeast Asia as an exporter of electronic parts. In order to confirm the widespread contaminations, sediment and biological samples were taken in Peninsular Malaysia along the Strait of Malacca from September 7 to 15, 2006 (Fig. 1). Descriptions of the sampling sites are given in Table 1. The surface sediment samples (0-5 cm) were taken using a core sampler in 13 sites. Sediment samples were not sieved. The dry weight of the sediment samples was calculated, with the difference between the weight of the sediment after having been dried at 110°C for 2 h and the weight of the sediment at the beginning. Water content and ignition loss (organic substances content) are shown in Table 1.

The green mussel (*Perna viridis*) samples were taken at 10 sites. The shell lengths of the green mussels were in the range 61–110 mm. Three mussel samples in each site were homogenated before analysis. Sediment samples were also taken in the coastal area of Melaka and the Strait of Johor from September 28 to 30, 2005 (Fig.1). All samples were brought back to Japan in a cold box and stored in a freezer at -20° C until chemical analysis.

Chemical Analysis

Organotin Compounds

The method used for the determination of OTs in sediment and biological samples was based on that of Midorikawa et al. (2004), with some modification. Five grams of sediment and the homogenate soft tissues of the mussels were put in a centrifuge tube and 100 µl of mixed acetone solution containing 1 µg/ml each of monobutyltin trichloride (MBTCl)-d₉ dibutyltin dichloride (DBTCl)-d₁₈, tributyltin monochloride (TBTCl)-d₂₇, monophenyltin trichloride (MPTCl)-d₅, diphenyltin dichloride (DPTCl)-d₁₀, and triphenyltin monochloride (TPTCl)-d₁₅ were added to the centrifuge tube as a surrogate standard. The mixture was shaken with 15 ml of 1 M HCl-methanol/ethyl acetate (1/1) for 10 min. After centrifugation for 10 min, the residue was again extracted with 10 ml of 1 M HCl-methanol/ ethyl acetate (1/1). After centrifugation, the combined supernatants and 30 ml of saturated NaCl solution were transferred to a separatory funnel. The analytes were extracted twice with 15 ml of ethyl acetate/hexane (3/2)solution and the organic layer was combined. Fifty milliliters of hexane was mixed into the organic layer and left to stand for 20 min. After removal of the aqueous layer, the organic layer was dried with anhydrous Na₂SO₄ and concentrated up to trace level. The analytes were diluted with 5 ml of ethanol, 5 ml of acetic acid-sodium acetate buffer

Fig. 1 Sampling sites, Peninsular Malaysia, Melaka, and the Strait of Johor

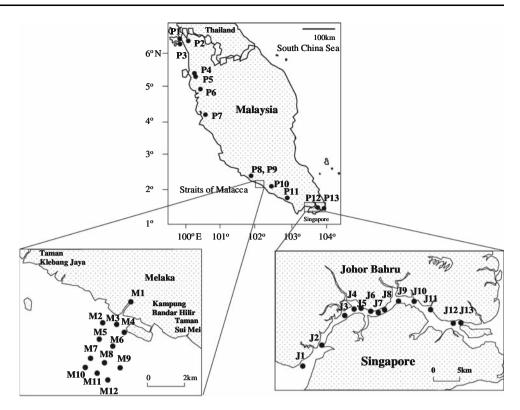


Table 1Characterization of
sampling sites and sediment
samples in Malaysia

Location	Coastal activity	Water content (%)	Ignition loss (%)
Perlis			
P1	Mix mangrove, ecotourism area	33	3.0
P2	Intertidal beach, fish landing, transit ports	63	10
P3	Intertidal beach, transit port, marina	32	3.8
Penang			
P4	Industrial and urban areas	52	15
P5	Industrial and urban areas	28	3.5
P6	Major mangrove area, aquaculture area (fishing boats mooring)	63	11
P7	Naval base, dockyard (active passenger, fishing boats and trading ships)	20	2.0
Negeri Se	embilan		
P8	Aquaculture area (small fishery boats mooring)	41	4.6
P9	Aquaculture area (sooring)	68	10
Melaka			
P10	Aquaculture area (small fishery boats mooring)	73	11
P11	Small and medium-size fishery and trading boats	42	4.0
Johor			
P12	Aquaculture area (small and medium-size fishery and passenger boats)	73	21
P13	Industrial, urban, and aquaculture areas, area dockyard (trading ships, and fishery and boats)	62	15

(pH 5.0), and 10 ml of distilled water and subsequently ethylated by shaking with 1 ml of 5% $NaBEt_4$ for 30 min. The solution containing ethylated OTs was saponificated

with 10 ml of 1 M KOH–ethanol solution by shaking for 1 h. Forty milliliters of distilled water and 20 ml of hexane were added to the solution and the mixture was shaken for

10 min. The ethylated OT residue in the aqueous layer was extracted again by shaking for 10 min with 20 ml of hexane. The combined organic layers were dried with anhydrous Na₂SO₄. After being concentrated up to 1 ml, the solution was cleaned by a florisil Sep-Pak column (Waters Association Co. Ltd.) The analytes were eluted with 10 ml of 5% diethyl ether/hexane. All eluting solvent was collected in a bottom flask. The solution was concentrated up to 0.5 ml after the addition of tetrabutyltin (TeBT)-d₃₆ and tetraphenyltin (TePT)-d₂₀ as an internal standard. The final solution was concentrated up to 0.5 ml.

A Hewlett-Packard 6890 series gas chromatography equipped with a mass spectrometry (5973 N) was used for analysis of the OTs. The separation was carried out in a capillary column coated with 5% phenyl methyl silicone (J&W Scientific Co.; 30 m length \times 0.25 mm inner diameter, 0.25 µm film thickness). The column temperature was held at 60°C for the first 2 min, then increased to 130°C at 20°C/min, to 210°C at 10°C/min, to 260°C at 5°C/min and to 300°C at 10°C/min. Finally, the column temperature was held at 300°C for 2 min. The interface temperature, ion source temperature, and ion energy were 280°C, 230°C and 70 eV, respectively. Selected ion monitoring was operated under this program. The monitoring ions of 235 (233) for MBT, 261 (263) for DBT, 263 (261) for TBT, 253 (255) for MPT, 303 (301) for DPT, and 351 (349) for TPT were used to quantify the concentrations of OTs, respectively. The parenthesis shows the qualifier ions. One microliter of the sample was injected by splitless injection. The concentrations of OTs in this study are expressed as Sn⁴⁺.

Alternative Biocides

The method used for the determination of alternative compounds in sediment was based on that of Harino et al. (2005). In the centrifuge tube, 5 g of wet sediment was placed together with 10 ml of acetonitrile. The mixture was shaken for 10 min in a mechanical shaker. After removal of the supernatant, the analytes were reextracted with 10 ml acetonitrile for 10 min and the mixture was then centrifuged. The combined supernatants were concentrated by a rotary evaporator up to 5 ml and 45 ml of distilled water was added to them. The analytes were extracted three times with 10 ml of dichloromethane. The organic layer was dried by anhydrous Na₂SO₄. After 10 ml of methanol and 100 μ l of atrazine-₁₃C₃ (1 mg/l) were added to the organic layer, the organic layer was concentrated up to 2 ml by a rotary evaporator. The analytes were determined by liquid chromatography/mass spectrometry-mass spectrometry (LC/MS-MS).

Liquid chromatography was performed on a high-performance LC (HPLC) apparatus equipped with an Agilent model 1100 series (Agilent; Yokogawa Analytical Systems, Tokyo, Japan). The separation was carried out on a narrow-bore C_{18} silica column (2.1 × 50 mm, 5 µm). The mobile phase was methanol–water run over a gradient (50% of methanol linear to 100% of methanol for 20 min and held for 10 min). The injection volume was 10 µl.

Electrospray mass spectrometry (ESI-MS-MS) analyses were carried out using a PE-Sciex API2000 (Sciex; Applied Biosystems Japan). Ionization of the analytes was achieved by electrospray in the positive ion mode. All the interface parameters were optimized by infusion of a standard solution (1 mg/l) of the analytes at the flow rate of 20 ml/min. The final electrospray conditions were as follows: nitrogen curtain gas, 40 l/min; ion spray voltage, 4800 V; ion source gas 1, 40 µl/min; gas 2, 70 µl/min; collision gas, 4 µl/min. LC/MS-MS acquisition was performed in the multiple reaction monitoring (MRM) mode and the precursor ion/product ion of Sea-Nine 211, Diuron, Dichlofluanid, Irgarol 1051, M1, and Pyrithiones were 282/ 170 (43), 233/46 (160), 333/224 (123), 254/198 (83), 214/ 158 (43), and 316/190 (189) respectively. The parentheses show the qualifier ion.

Results and Discussion

Evaluation of the Analytical Procedure

In order to examine the quality of the data obtained by the analytical procedure of OTs and alternative biocides, 0.1 μ g of OTs were spiked to 5 g of sediments and green mussels. The recoveries of the OTs in sediment and mussels were in the range 98–115% and 95–110%, respectively, and the relative standard deviations (RSDs) of the OTs were in the range 6–10% for sediment and 5–12% for mussels, respectively (Table 2). The recoveries of the alternative biocides in the sediment were in the range 75–95%, and their RSDs were in the range of 8–15%.

The detection limits were calculated from a signal-tonoise ratio of 3.The OT's detection limits were 0.1 μ g/kg dry weight (dw) for the sediment samples and 1 μ g/kg wet weight (ww) for the mussel samples. The detection limits of Sea-Nine 211, Diuron, Dichlofluanid, Irgarol 1051, M1, and Pyrithiones in the sediment samples were 0.04, 0.02, 0.1, 0.02 and 0.09 and 20 μ g/kg dw, respectively.

Distribution and Concentrations of Antifouling Biocides in Peninsular Malaysia

The coastal area of Peninsular Malaysia, which is called the commercial capital, is an area in which many ships sail and are moored. It can therefore be predicted that contamination by antifouling paint in the area has occurred.

	Amounts (g)	Amount spiked (µg)	Recov	Recovery rates (%)										
			MBT	DBT	TBT	MPT	DPT	ТРТ	Sea-Nine 211	Diuron	Dichlofluanid	Irgarol 1051	M1	Pyrithiones
Sediment	5	0.1	110	102	105	115	98	100	85	95	78	75	95	80
			(10)	(8)	(8)	(9)	(10)	(6)	(10)	(9)	(10)	(8)	(8)	(15)
Mussels	5	0.1	98	101	95	110	95	98	_	_	_	-	_	-
			(12)	(10)	(5)	(10)	(9)	(11)	-	-	-	_	-	-

Table 2 Recovery rates of antifouling biocides

Parenthesis shows relative standard deviation

Fig. 2 The spatial distribution of BTs and PTs in sediment from Peninsular Malaysia

Figure 2 shows the spatial distribution of BTs in sediment from Peninsular Malaysia along the Strait of Malacca. The concentrations of BTs were high in the southern part of Peninsular Malaysia. In the northern part of Peninsular Malaysia, tourism in a big industry because there are many mangrove forests and beautiful beaches, whereas in the southern part, there are fishing grounds, fish farms, and industrial ports. Severe BTs contamination was therefore found in the southern part. Sudaryanto et al. (2004) reported that elevated concentrations of BTs were found along the coast of the Strait of Malacca, particularly in locations having intensive maritime activities, such as Penang, Johor, and Johor Bahru. The concentrations of MBT, DBT, and TBT in sediment from the coastal waters of Peninsular Malaysia along the Strait of Malacca were in the range $4.1-242 \ \mu g/kg \ dw$, $1.1-186 \ \mu g/kg \ dw$, and $0.7-228 \ \mu g/kg \ dw$, respectively (Table 3). The maximum concentration of TBT was observed at St. P13, where there is heavy ship traffic. The TBT concentrations in sediment from Sts. P10-11, which contains aquaculture and fishery areas, were also high. We predicted that much TBT was used in this area in the past. Furthermore, the high percentages of TBT in sediment from Sts. P10-11 and P13 show that TBT is still being used in these areas (Table 3).

Sudaryanto et al. (2004) reported BT concentrations in sediment from the coastal areas of Malaysia in 1997. The levels of BTs in sediment between 1997 and 2006 were compared using these values, because the sampling sites by Sudaryanto et al. (2004) were similar to our study areas (Fig. 3). Remarkable changes of BTs during 10 years were not observed in sediment, suggesting that the use of BTs has persisted.

The spatial distribution of PTs in Peninsular Malaysia is shown in Fig. 2. Despite the detection of PTs in most sites, a characteristic trend was not observed in the spatial distribution. The concentrations of MPT, DPT, and TPT were in the range $<0.1-121 \mu g/kg dw$, $0.4-27 \mu g/kg dw$, and $0.1-34 \mu g/kg dw$ in sediment from the coastal waters of Peninsular Malaysia, respectively. Of total PTs, MPT was the dominant species. Kannan and Lee (1996) reported that TPT was used as a pesticide. The detections of PTs in Peninsular Malaysia might be due to its use as a pesticide and not as an antifouling biocide, because the spatial distribution of PTs is different from that of BTs.

Figure 4 shows the spatial distribution of BTs in green mussels from Peninsular Malaysia. Remarkable differences of BTs among sites were not observed. It was reported that TBTs in mussels reflected on those in water within 2 or 3 months (Short and Sharp 1989). It is presumed that BTs in water are at similar levels among the sampling sites. MBT, DBT and TBT in green mussel samples were detected in the range 41–102 μ g/kg, 3–5 μ g kg⁻¹ and 8–32 μ g kg⁻¹, respectively. Of all BTs, the percentage of

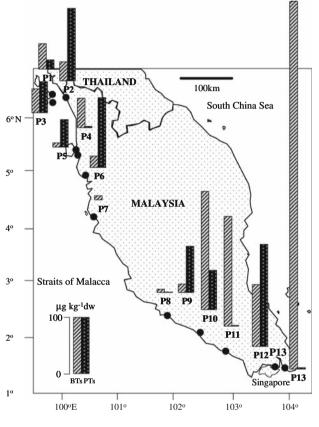


Table 3 Concentrations of antifouling biocides in sediment from Peninsular Malaysia (µg/kg dw)

Location	MBT	DBT	TBT	MPT	DPT	TPT	Seanine-211	Diuron	Dichlofluanid	Irgarol 1051	M1	Pyrithions
P1	19	7.1	19	16	0.4	0.2	0.05	< 0.02	<0.1	< 0.02	< 0.09	<20
P2	17	7.9	8.9	77	24	28	< 0.04	0.39	< 0.1	1.2	< 0.09	<20
P3	19	8.8	15	32	10	13	< 0.04	0.12	< 0.1	0.25	< 0.09	<20
P4	24	9.7	20	< 0.1	1.2	0.8	< 0.04	0.29	< 0.1	< 0.02	< 0.09	<20
Р5	5.5	1.4	0.9	30	8.2	11	< 0.04	0.08	< 0.1	0.2	< 0.09	<20
P6	14	4.7	1.9	81	19	24	< 0.04	0.14	< 0.1	0.40	< 0.09	<20
P7	5.3	1.3	1.5	< 0.1	0.4	0.1	< 0.04	< 0.02	< 0.1	0.17	< 0.09	<20
P8	4.1	1.1	0.7	< 0.1	0.5	0.2	< 0.04	< 0.02	< 0.1	< 0.02	< 0.09	<20
P9	12	2.5	1.1	44	17	21	< 0.04	< 0.02	< 0.1	0.80	< 0.09	<20
P10	42	13	160	65	4.2	0.8	1.7	4.8	< 0.1	14	< 0.09	<20
P11	25	20	150	< 0.1	0.7	0.4	< 0.04	< 0.02	< 0.1	0.25	< 0.09	<20
P12	46	23	41	121	27	34	< 0.04	0.86	< 0.1	1.9	< 0.09	<20
P13	240	190	230	< 0.1	1.7	0.9	< 0.04	4.3	<0.1	2.6	< 0.09	<20

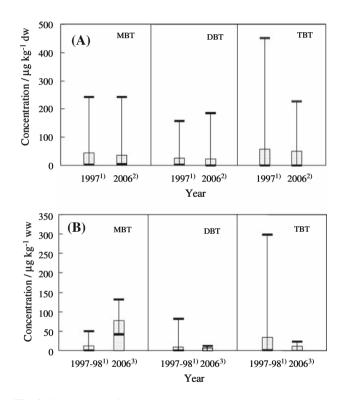


Fig. 3 Comparison of BT concentrations in sediment and mussel samples between 1997 and 2006: **a** sediment, **b** mussels. ¹⁾Sudaryanto et al. (2004), ²⁾this study, ³⁾Sudaryanto et al. (2002)

TBT was low, suggesting the decrease of TBT's input. Sudaryanto et al. (2002) reported the BT concentrations in green mussels from the coastal areas in Malaysia during 1997–1998. The levels of BTs in sediment between 1997– 1998 and 2006 were compared using these values (Fig. 3). During those 10 years, the TBT concentrations decreased and MBT concentrations increased, supporting the decreasing input of BTs.

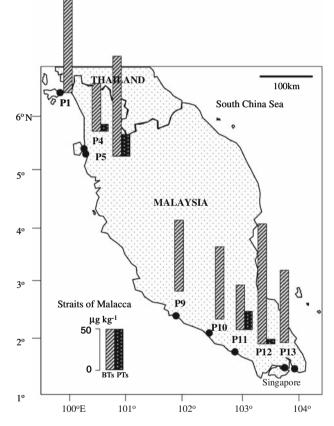


Fig. 4 The spatial distribution of BTs and PTs in green mussels from Peninsular Malaysia

Although MPT and DPT in mussel samples were detected in the range $<0.1-22 \mu g/kg$ and $<0.1-5 \mu g/kg$, respectively, TPT was not detected.

The mussels used in the present study are an important source of food. Therefore, the health risk to persons from the BT in mussels is a concern. Penninks (1993) derived a tolerable daily intake (TDI) of TBTO of $0.25 \ \mu g/kg$ body weight/day. This value was based on the observed effects of TBT on the immune function in rats with a safety factor of 100 to extrapolate the toxicity test from rats to humans. The tolerable average residue levels (TARLs) were calculated from TDI based on the following formula:

 $TARL = (TDI \times 60 \text{kg body weight})/$ average daily seafood consumption) (1)

The TARL for seafood in Malaysia were estimated to be 40.9 μ g/kg ww for an average person weighing 60 kg and consuming 146.6 g of seafood a day (Belfroid et al. 2000). The maximum value of TBT (32 μ g/kg ww) detected in mussel samples was the value near the TARL, suggesting the importance of further monitoring of the TBT concentrations in mussel samples.

As alternative compounds, Sea Nine 211, Diuron, Irgarol 1051, M1, and Pyrithione were measured in sediment from Peninsular Malaysia. The concentrations of Diuron in sediment were in the range $<0.1-5 \ \mu g/kg dw$. The concentrations of Diuron in sediment from the coastal waters of England were in the range $<12-395 \ \mu g/kg dw$, as reported by Boxall et al. (2000). Thomas et al. (2000, 2002) reported that the Diuron concentrations were in the range $<0.1-1.4 \ \mu g/kg dw$ in Southampton. Harino et al. (2005) reported that Diuron was detected in the range 0.64– 1350 $\mu g/kg dw$ in the Port of Osaka, Japan. The concentrations of Diuron in sediment from Peninsular Malaysia were lower than these values.

The concentrations of Irgarol 1051 in sediment from Malaysia were in the range $<0.1-14 \ \mu g/kg$ dw. The concentrations of Irgarol 1051 have been reported in various countries; for example, the concentrations in the Hamble estuary, Owell estuary, and Southampton, England ranged from 6 to 880 $\mu g/kg$ dw (Boxall et al. 2000; Gough et al. 1994; Thomas et al. 2000, 2002). The concentrations of Irgarol 1051 in sediment from Switzerland, Germany, and Japan were in the range $<0.2-8 \ \mu g/kg$ dw, $3-220 \ \mu g/kg$ dw, and $7-816 \ \mu g/kg$ dw, respectively (Biselli et al. 2000; Harino et al. 2005; Toth et al. 1996). The concentrations of Irgarol 1051 in Malaysia were lower than those in European countries and Japan.

Higher concentrations of Diuron and Irgarol 1051 were observed in Sts. P10 and P12–13, where the concentrations of TBT were high. This means that Diuron and Irgarol 1051 are used as an antifouling paint in Malaysia. Sea Nine 211 and Dichlofluanid in sediment from most sampling sites were not detected. However, the concentrations of alternative compounds were lower than those of TBT (Table 3). Judging from these findings, the contamination by OTs in Malaysia is a more serious problem than that by alternative compounds.

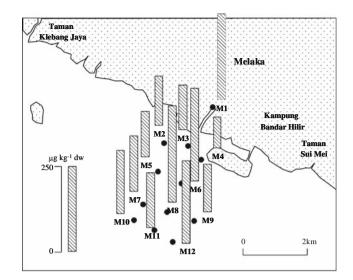


Fig. 5 The spatial distribution of BTs in sediment from Melaka

Distribution of Antifouling Biocides in Melaka and the Strait of Johor

The concentrations of antifouling biocides in Melaka and the Strait of Johor, where there is great traffic density, were investigated in detail. The spatial distribution of BTs in Melaka is shown in Fig. 5. There were similar concentrations of BTs among all sampling sites from Melaka, indicating that BTs contamination spread off the coast. The concentrations of PTs were lower than those of BTs, and the spatial distribution of PTs in Melaka was different from those of BTs (Table 4). The input of PTs to sediment might be due to its use as a pesticide, as described in the previous subsection. The concentrations of MBT, DBT, and TBT in sediment from Melaka were in the range 85-270 µg/kg dw, 3.6-17 µg/kg dw, and 2.4-31 µg/kg dw, respectively, which were similar to those in Peninsular Malaysia. MBT was the dominant species in sediment from Melaka. It is well known that TBT persists for a long time in sediment (Dowson et al. 1993; Harino et al. 1998; Maguire and Tkacz 1985). However, the higher percentage of MBT among BTs in this study shows the degradation of TBT. It was reported that in most of the locations tested in Malaysia, MBT generally occurred in the highest proportion (Sudaryanto et al. 2004). The degradation of TBT in sediment from Malaysia might be faster than those in other areas because of higher temperatures. Further study is needed to clarify why the proportion of MBT is high in sediment from Malaysia.

The spatial distribution of BTs in sediment from the Strait of Johor is shown in Fig. 6. BTs concentrations in sediment were high at Sts. J7–J11 with poor flushing of water. Sudaryanto et al. (2004) reported that higher concentrations of BTs were found in the sediment samples

Table 4 Concentrations of antifouling biocides in sediment from Melaka (µg/kg dw)

Location	MBT	DBT	TBT	MPT	DPT	TPT	Seanine-211	Diuron	Dichlofluanid	Irgarol 1051	M1	Pyrithions
M1	200	17	31	9.2	0.5	< 0.1	4.2	4.1	< 0.1	0.21	< 0.09	<20
M2	140	5.2	4.1	5.9	0.4	1.3	0.11	0.18	< 0.1	0.15	< 0.09	<20
M3	120	5.9	8.1	10	0.3	1.1	0.11	< 0.02	< 0.1	0.05	< 0.09	<20
M4	85	3.6	2.4	4.8	0.2	0.4	0.11	< 0.02	<0.1	0.06	< 0.09	<20
M5	120	5.7	3.8	3.6	0.3	0.3	< 0.04	< 0.02	<0.1	0.08	< 0.09	<20
M6	260	7.5	4.7	5.6	0.5	1.3	0.14	< 0.02	< 0.1	0.06	< 0.09	<20
M7	150	5.3	4.1	3.7	0.3	1.0	< 0.04	0.48	< 0.1	0.06	< 0.09	<20
M8	270	7.0	4.3	11	0.5	1.6	0.09	0.36	< 0.1	0.06	< 0.09	<20
M9	130	5.8	2.9	8.0	0.1	0.3	0.20	< 0.02	< 0.1	< 0.02	< 0.09	<20
M10	170	7.5	4.4	4.6	0.1	0.2	0.04	0.99	< 0.1	0.11	< 0.09	<20
M11	150	5.6	3.0	6.1	0.1	0.4	< 0.04	< 0.02	< 0.1	0.07	< 0.09	<20
M12	230	7.3	8.5	6.4	0.1	0.9	0.09	< 0.02	<0.1	0.14	0.49	<20

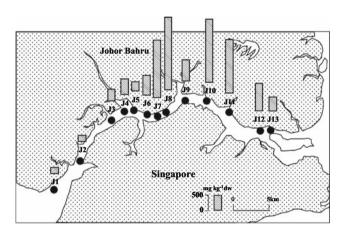


Fig. 6 The spatial distribution of BTs in sediment from the Strait of Johor

collected in the narrowest area of the Strait of Johor. The concentrations of MBT, DBT, and TBT in the Strait of Johor were in the range $83-542 \text{ }\mu\text{g/kg}$ dw, $30-232 \text{ }\mu\text{g/kg}$

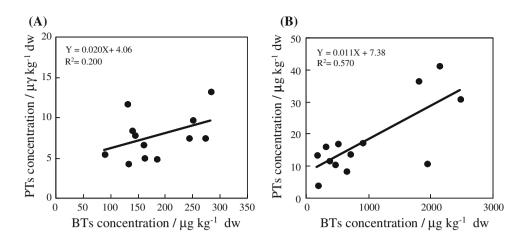
dw, and 41–492 µg/kg dw, respectively, which were higher than those in Peninsular Malaysia and Melaka (Table 5). MBT was the dominant compound at most sampling sites of the Strait of Johor, suggesting that the degradation rate of TBT is faster than the input rate in sediment from this area. The concentrations of MPT, DPT, and TPT were in the range 41–66 µg/kg dw, 5–29 µg/kg dw, and 0.3–34 µg/ kg dw from the Strait of Johor, respectively (Table 5). The concentrations of PTs in the Strait of Johor were similar to those in the coastal waters of Peninsular Malaysia and Melaka and MPT among PTs was the dominant compound in Strait of Johor.

The relationship between the BT and PT concentrations is shown in Fig. 7. The correlation coefficient between the BTs and PTs concentrations in the Strait of Johor $(R^2 = 0.570)$ was higher than that in Melaka $(R^2 = 0.200)$. It was found that PTs are used not only as pesticides but also as antifouling paint on ships that are sailed and moored in the Strait of Johor.

Table 5 Concentrations of antifouling biocides in sediment from the Strait of Johor (μ g/kg dw)

Location	MBT	DBT	TBT	MPT	DPT	TPT	Seanine-211	Diuron	Dichlofluanid	Irgarol 1051	M1	Pyrithions
J1	180	7.1	7.2	3.3	0.2	0.2	0.04	0.22	<0.1	0.04	< 0.09	<20
J2	150	19	20	12	0.4	0.8	< 0.04	1.0	<0.1	0.11	< 0.09	<20
J3	320	26	38	11	0.3	0.7	< 0.04	0.89	<0.1	0.11	< 0.09	<20
J4	460	32	28	15	0.5	1.1	0.04	0.99	<0.1	0.11	< 0.09	<20
J5	290	14	12	15	0.2	0.6	< 0.04	1.2	<0.1	0.08	< 0.09	<20
J6	480	90	87	8.2	< 0.1	< 0.1	< 0.04	1.8	<0.1	0.13	< 0.09	<20
J7	1,300	240	400	10	0.7	< 0.1	< 0.04	< 0.02	<0.1	< 0.02	< 0.09	<20
J8	1,000	590	880	25	0.7	4.9	0.31	< 0.02	<0.1	0.21	< 0.09	<20
J9	360	170	200	13	0.6	< 0.1	0.15	2.3	<0.1	0.34	< 0.09	<20
J10	1100	400	640	38	0.8	2.1	0.92	4.5	<0.1	0.90	< 0.09	<20
J11	640	230	940	34	1.7	1.2	0.47	9.9	<0.1	0.75	< 0.09	<20
J12	550	110	240	15	0.5	1.3	0.44	3.0	<0.1	0.32	< 0.09	<20
J13	350	51	61	4.3	0.2	6.0	0.10	2.1	<0.1	0.14	< 0.09	<20

Fig. 7 Relationships of concentration between BTs and PTs in sediment from Peninsular Malaysia: **a** Melaka, **b** the Strait of Johor



Sea Nine 211, Diuron, and Irgarol 1051 in the sediment from Melaka were detected in the range $<0.04-4.2 \ \mu g/kg$ dw, $<0.02-4.1 \ \mu g/kg$ dw, and $<0.02-0.21 \ \mu g/kg$ dw, respectively, whereas Dichlofluanid, M1, and Pyrithiones were not detected. The concentrations of Sea Nine 211, Diuron, and Irgarol 1051 were high at St. M1, which is located at the mouth of a river. In fact, the spatial distributions of alternative biocides were different from those of BTs. BT pollution in sediment was spread across a wide area because BTs have been used for a long time. However, the contamination by alternative compounds showed localized distribution, because alternative biocides have recently begun to be used.

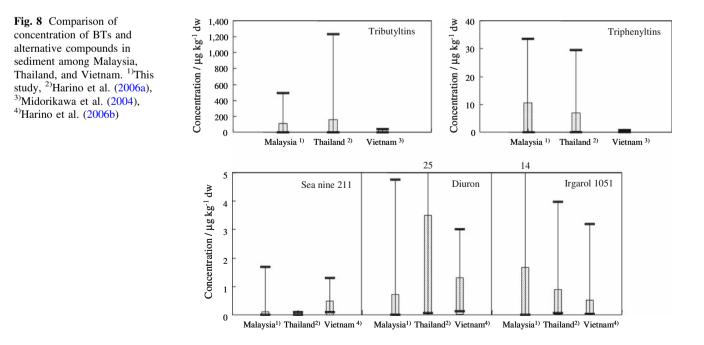
Sea Nine 211, Diuron, and Irgarol 1051 in the sediment from the Strait of Johor were detected in the range $<0.04-0.92 \ \mu g/kg \ dw$, $<0.02-9.9 \ \mu g/kg \ dw$, and $<0.02-0.90 \ \mu g/kg \ dw$, respectively, and Dichlofluanid, M1, and Pyrithiones

were not detected. The Diuron concentration was the highest among alternative biocides.

Comparison of the Concentrations of Antifouling Biocide Among Southeast Asian Countries

The concentrations of OTs were compared among Malaysia, Thailand, and Vietnam (Fig. 8). The higher concentrations and the wide variations of TBT and TPT in sediment from Malaysia and Thailand show that larger amounts of these compounds have been used in coastal waters from these countries in the past.

The level of each alternative compound in sediment was different in each country (Fig. 8). The higher concentrations were observed in Irgarol 1051 for Malaysia, Diuron for Thailand, and Sea Nine 211 for Vietnam. This reflects the



fact that the patterns of application of antifouling biocides vary among regions.

Conclusions

Judging from the concentration of antifouling biocides in sediment, contamination by OTs is a more serious issue than that by alternative compounds in Southeast Asia. Even now, higher concentrations of OTs were observed at the busiest locations for ship traffic. Information about the hazards of contamination by alternative biocides is also needed because an increase in the demand for alternative compounds is estimated in the near future. Therefore, further studies are needed to continue monitoring antifouling biocides in Southeast Asian countries and to evaluate the risk of alternative compounds.

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