Reduced Fitness of *Daphnia magna* Fed a Bt-Transgenic Maize Variety

Thomas Bøhn · Raul Primicerio · Dag O. Hessen · Terje Traavik

Received: 16 November 2007/Accepted: 11 February 2008/Published online: 18 March 2008 © Springer Science+Business Media, LLC 2008

Abstract Genetically modified (GM) maize expressing the Bt-toxin Cry1Ab (Bt-maize) was tested for effects on survival, growth, and reproduction of the water flea Daphnia magna, a crustacean arthropod commonly used as a model organism in ecotoxicological studies. In three repeated experiments, D. magna were fed 100% ground maize in suspension, using either GM or isogenic unmodified (UM) maize. D. magna fed GM-maize showed a significantly reduced fitness performance: The mortality was higher, a lower proportion of females reached sexual maturation, and the overall egg production was lower compared to D. magna fed UM isogenic maize. We conclude that the tested variety of Bt-maize and its UM counterpart do not have the same quality as food sources for this widely used model organism. The combination of a reduced fitness performance combined with earlier onset of reproduction of D. magna fed Bt-maize indicates a toxic effect rather than a lower nutritional value of the GMmaize.

T. Bøhn (⊠) · T. Traavik Genøk—Centre for Biosafety, The Science Park, P.O. Box 6418, Tromso 9294, Norway e-mail: thomas@genok.org

R. Primicerio Norwegian College of Fishery Science, University of Tromsø, Tromso 9037, Norway

D. O. Hessen Department of Biology, University of Oslo, Oslo, Norway

T. Traavik Department of Microbiology and Virology, School of Medicine, University of Tromsø, Tromsø 9037, Norway

$\underline{\underline{\hat{2}}}$ Springer

Introduction

A few herbivore insect species are considered important pests of major crop plants. These pests are targeted with genetic engineering by insertion of various *Bacillus thuringiensis* (Bt) *cry* genes. In the plant, these genes express Cry proteins with toxic effects on different target pest insects. The Cry1Ab protein is the most widely used toxin against the order *Lepidoptera*. The most common Cry1Ab transgenic plants, known as Bt-plants, are maize, rice, cotton, and potato. In maize, the most frequent modifications include insect resistance (Bt), herbicide tolerance, or the stacking of genes that provide both of these traits. Insect-resistant Bt-maize contribute to more than 50% of the transgenic maize grown globally, but the use of varieties with stacked genes are increasing (James 2006).

The purpose of Bt-plants is to selectively kill insect pest species, thereby increasing plant yield without affecting other (nontarget) species. For the environment, the purported selectivity and localized effect of Bt-plants might be an improvement over the use of sprayed pesticides. According to the US Environmental Protection Agency, Bt-crops pose no significant risk to the environment or to human health (Mendelson et al. 2003). However, the specificity of Bt-plant defense has yet to be established (Dutton et al. 2003, 2005), especially because the mode of action of Cry proteins, both in target and nontarget organisms, is not fully understood (Brandt et al. 2004; Bravo et al. 2007; Crickmore 2005; Hilbeck and Schmidt 2006).

Many nontarget organisms, including herbivores, pollinators, parasitoids, and predators will directly and indirectly be exposed to transgene products and altered interactions (Andow and Hilbeck 2004; Dutton et al. 2003;

Groot and Dicke 2002; Hilbeck 2001; O'Callaghan et al. 2005) and it is well justified to test these for potential harmful effects. Several reviews cover many groups of invertebrates, but nearly all of the tested species are terrestrial. However, a large number of studies show significant negative impacts on nontarget species after feeding transgenic plant parts. Without a clear mode of action of Cry1Ab on nontarget groups and negative effects found on predators (about 30% of the studies) and parasitoids (57% of the studies) (Lövei and Arpaia 2005) and on about 50% of all studies of Bt-plants (Hilbeck and Schmidt 2006), there is a need for further studies of nontarget organisms: Which species are exposed, which are sensitive, what is/are the mode(s) of action, and, ultimately, what are the consequences?

The list of nontarget groups extends if we include aquatic ecosystems adjacent to agricultural fields. These receive runoff material from agricultural fields and harbor nontarget organisms that are potentially exposed and affected by Bt-plants. The aquatic component of nontarget organisms has received little attention. However, transgenic cry1Ab genes have recently been shown to drain, persist, and possibly be transported long distances into aquatic freshwater ecosystems from agricultural fields with GM Bt-maize (Douville et al. 2007). Douville and colleagues concluded that the cry1Ab transgene is likely to be expressed in aquatic environments and recommended further monitoring strategies to characterize environmental exposure and effects. Rosi-Marshall et al. (2007) demonstrated that toxin-containing crop by-products are dispersed, decomposed, and consumed in aquatic environments adjacent to agricultural fields. They also demonstrated negative effect of Bt-plants on two nontarget stream insects (caddisflies). Negative effects of released Bt-crop by-products to aquatic environments have broad implications because such effects have been overlooked by previous research and management (Rosi-Marshall et al. 2007).

The uncertainties and lack of relevant research related to biological effects of Cry1Ab plant versions were reflected by a recent review concerning putative health effects of GM-plants. The author concluded with the question: "Where is the scientific evidence showing that GM plants/ food are toxicologically safe?" (Domingo 2007).

Daphnia magna is a crustacean (phylum: Arthropoda) invertebrate that inhabits ponds and lakes in most regions of the world. It is a common inhabitant of ponds in agricultural landscapes and will, like many other zooplankton and benthic arthropods, receive pollen and detritus from drainage water from agricultural fields (cf. Rosi-Marshall et al. 2007). *D. magna* feed nonselectively on a broad range of particles in the size range 1–50 μm, and where transgenic Bt-plants are grown, they might receive this in

their diet in the form of detrital particles and pollen. The clonal D. magna is commonly used in toxicological and ecotoxicological research (Atienzar et al. 2001; Barry 1996; Kramer et al. 2004) and has shown no treatmentrelated adverse effects to acute toxicity tests on transgenic Cry1Ab-maize pollen (Mendelson et al. 2003). However, a 48-h acute toxicity is only a first step in testing a GM crop plant because sublethal effects are precluded. We analyze in detail the performance of D. magna feeding on transgenic Cry1Ab-maize over the whole life cycle. More accurate assessment including potential fitness costs can be derived from analysis of life-history traits responses (i.e., responses on survival, growth, and reproduction). The rapid life cycle of *D. magna*, combined with a predominant asexual mode of reproduction and minimal genetic variation as well as easily measurable and plastic life-history traits, makes it an ideal model organism. Different species of the genus Daphnia have been used not only to evaluate pesticides, other toxicants, and pharmaceuticals (Nogueira et al. 2004) but recently also for investigating the role of toxins in human disease (Campbell et al. 2004). To increase the precision and level of detail in studies of responses, ecotoxicologists have started to include sublethal effects by using life histories and population fitness measures (Stark and Banks 2003). We transfer parts of this methodology to test comparatively the food quality of a GM versus the unmodified (UM) plant.

In the present study, we investigate whether Bt-maize might have negative impacts on a nontarget model organism, either by toxic effects or by reduced energy availability. We compare the fitness performance of *D. magna* that were fed either a variety of transgenic Cry1Ab-maize or its UM isogenic counterpart grown in the same environment. The measured response variables were (1) survival, (2) growth, (3) individual fecundity, (4) population fecundity, (5) frequency of maturation, and (6) age at maturation. We followed these response variables in three consecutive experiments, employing the same study design through the whole life cycle of the animals.

Methods

Feed

The transgenic Cry1Ab-maize was of the variety Dekalb 818 YG (a hybrid of MON 810) and a Philippine, local variety of maize called Dekalb 818. Both varieties (Dekalb 818 YG and Dekalb 818) were grown side by side in adjacent fields, divided by a small river, in Elizabeth Cruzara, near Iloilo City in 2003. Maize had been grown on these fields for many years. This was the very first year of GM-maize cultivation. The neighboring farmers delivering



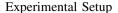
the GM- and UM-maize have stated that there was no external pesticides used in the fields. We inspected the fields and confirmed their GM and UM status by polymerase chain reaction (PCR) analyses of field-collected samples before buying adequate maize amounts from local farmers. The transgenic status of the MON810 event was further verified by DNA nucleotide sequencing, employing an Applied Biosystems 3130xl genetic analyzer (data not shown).

We subsampled a 50-kg bag of GM-maize and analyzed the expression of the cry1Ab gene with enzyme-linked immunosorbent assay (ELISA) on 15 samples from the bag (diluted 1:20–1:33), using a commercially available kit (Abraxis; http://www.abraxiskits.com). On average, the GM-maize expressed 67 (\pm 27 SD) ng Cry1Ab toxin per gram of dried grain tissue. All negative controls and the UM-maize showed negative results.

Maize feed were made twice (referred to as feed 1 and feed 2) from the same bag of kernels. Subsamples of GM-and UM-maize were drawn from 50-kg bags, and 35 g of dried kernels were grinded with separate coffee grinders (Petra Espresso), first on the coarsest grinding and then five times repeatedly on the finest setting. The resulting flour was sieved through a filter with a 250-µm mesh size. Eight hundred milligrams of the filtered maize flour were added to 250 mL of Aarchnia Daphnien Medium (ADAM) (Kluttgen et al. 1994), homogenized, and frozen in 10-mL test tubes. This is later referred to as the feed. All steps of the feed production were the same for GM- and UM-maize.

In culture, *D. magna* is generally raised on phytoplankton, but it might, like most zooplankton species, feed on a mixed diet of small particles. In the feeding trials, we chose food concentrations within a range that is normally encountered in nature ($<10~\text{mg C/L}^1$), and both the control and the Bt-maize tests were conducted with the same concentrations to avoid food quantity effects. The size spectrum of both food types was analyzed with a Fac-sCalibur flow-cytometer using 0.5- μ m beads as standard. These analyses revealed an almost identical particle size range for the Bt-maize and the control spanning 2–4 μ m, which is close to the optimal size range both for juvenile and adult *D. magna*.

In preliminary experiments, we had established that on a diet of 100% UM-maize, *D. magna* had a survival close to 80% in 20 days, grew up to about 3 mm in body length (about 20–30% smaller than on standard algal diet; *e.g.*, *Scenedesmus gracilis*), and reproduced up to a maximum of 6 clutches within 42 days. The clutch sizes were small and within the range of one to four eggs. We did not follow the survival of the neonates but used numbers of eggs as the reproductive end point in the study. With maize as the only food source, we eliminate any potential confounding effects associated with a composite diet.



All individuals of D. magna used in the experiments were born within 30 h from the third clutch of a single clonal population. Twenty juvenile individuals were randomly chosen and assigned to separate glasses with 60 mL autoclaved ADAM medium. Ten animals received the GM feed (treatment) and 10 received the UM feed (control) under identical environmental conditions in a climate chamber at 20°C and 24 h daylight (resembling the summer conditions at our latitude). Every third day, all individuals were transferred to new glasses with new medium using a broad-tipped pipette. Thereafter, the position of each individual was rerandomized. Each D. magna was fed daily and inspected for survival and number of eggs produced. The experiment lasted for 42 days. Care was taken to provide the same amount of food for all experimental units within and among GM and UM food recipients. All individual D. magna were fed daily with 100 µL of maize feed, corresponding to 0.4 mg dry weight of maize or about 0.2 mg C per individual per day (i.e., within the range recommended for laboratory experiments with Daphnia; Sims et al. 1993). Our experiments combined the study of survival (ecotoxicological approach) with growth and fecundity (feeding performance approach) to be able to estimate the overall fitness implications of the GM treatment diet.

This experimental setup was performed consecutively three times, hereafter referred to as experiment 1, 2, and 3. Feed 1 was used in experiment 1, and feed 2 was used in experiments 2 and 3. The same clonal line of *D. magna* was used in all experiments.

Measurements

Body length was measured 17, 29, and 42 days after the experiment was initiated. In experiment 1, individual D. magna from both groups (GM treatment and UM control) were measured for body length (distance from the top of the head to the base of the caudal spine) using a $40 \times$ binocular microscope. In experiments 2 and 3, body length was measured by image analysis (using ImageJ software) based on digital photographs of D. magna individuals. All length measurements were done "double-blind"; that is, the observer (and the experimental unit, the D. magna) did not know to which group the measured individual belonged.

Statistical Analysis

Survival

Statistical calculations were performed in R (2.5.0) and Systat (10.2) software. Survival analyses were



performed in R using the package "survival" and Cox proportional hazards (coxph test) for testing differences between GM and UM groups. For estimating the predicted mean survival, we used a survival model based on the *survreg*-function in R, with exponential error terms specified.

Growth and Reproduction

In *t*-tests (within each experiment), we used pooled variances. Because the variability was relatively high among experiments, we analyzed overall differences (all experiments combined) with *Experiment* as a covariate (ANCOVA) (*i.e.*, correcting for differences between the experiments). The analyses on fecundity were based on ln-transformed numbers of eggs. The percentage of females that reached maturation was tested with chisquare tests. The age at 50% maturation was estimated with logistic regression and the confidence limits were calculated with bootstrapping in R using the package "boot" with 1000 resamplings including bias correction and acceleration.

Results

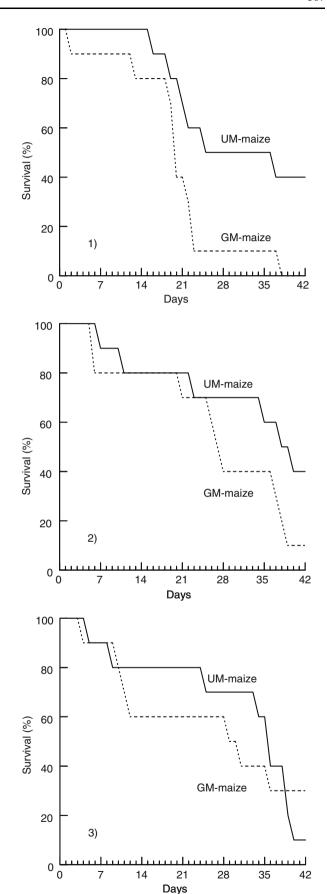
Survival

Daphnia magna had reduced survival when fed GM-maize compared with UM-maize (Figs. 1 and 2). The difference was statistically significant in experiment 1 (p = 0.031, coxph test) and in the combined data from all experiments (p = 0.029, coxph test). The predicted mean survival was lower in D. magna fed GM-maize (mean survival for pooled data: UM, 45 days; GM, 28.2 days).

Growth

Differences in body length between GM- and UM-fed D. magna were not consistent among experiments (Fig. 3). In experiment 1, significantly larger D. magna were found in the UM group at day 17 (p = 0.05, t-test). When testing the pooled data from all experiments at day 17 and using Experiment as a covariate (i.e., correcting for differences between experiments), a tendency for larger individuals of D. magna feeding on UM-maize relative to the GM-maize was found; yet the difference was not significant (p = 0.134, ANCOVA).

Fig. 1 Survival curves (%) of *D. magna* fed on GM-maize (dotted line) and UM-maize (solid line) in experiments 1, 2, and 3 (from top to bottom)





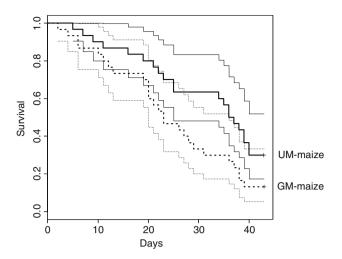


Fig. 2 Proportion of surviving *D. magna* fed on GM-maize (dotted lines) and UM-maize (solid lines) for pooled data (n=60). Thin lines represent 95% confidence intervals

Reproduction

Individual Fecundity

Daphnia magna fed GM-maize had a significantly higher mean number of eggs per female in experiment 3 (p = 0.043, t-test). Also in the pooled samples from all experiments, GM-fed animals had a higher fecundity (7.3) compared to the UM-fed females (5.1) (Fig. 4); yet the difference was not statistically significant (p = 0.255, ANCOVA).

Total Production of Eggs

The total number of eggs produced by GM-fed *D. magna* was lower in sum for all the experiments, compared to those fed UM-maize (80 vs, 96 eggs) (Fig. 5); this was largely due to very few eggs produced in the GM experiment 1.

Percentage of Females Reaching Maturation

The percentage of females that reached maturation (*i.e.*, produced eggs) was generally lower in the GM-fed D. magna, ranging from 20% to 60% in the different experiments (36.7% on average) compared to 50–80% (63.3% on average) in the UM-fed D. magna (Fig. 6). Statistically significant differences were found in experiment 3 (p = 0.025, chi-test) and for the combined data from all experiments (p = 0.039, chi-test).

Age at Maturation

The age at 50% maturation was generally lower for the GM-fed *D. magna* (15.3 days) than for the UM-fed group

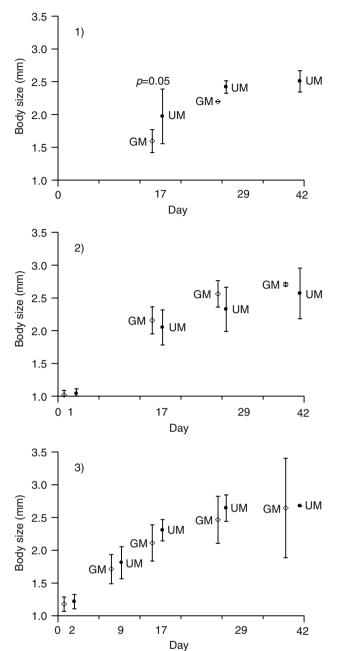


Fig. 3 Body size ($\pm 95\%$ confidence intervals) at days 1, 2, 9, 17, 29, and 42 of *D. magna* fed on GM-maize and UM-maize in the three different experiments. The *p*-value of a significant difference within experiment 1 is shown (*t*-test)

(17.5 days) and the 95% confidence intervals estimated from bootstrapping did not overlap between the two groups (Fig. 7).

Discussion

We detected negative effects of a transgenic Bt-maize line on fitness-related parameters of *D. magna* in repeated and



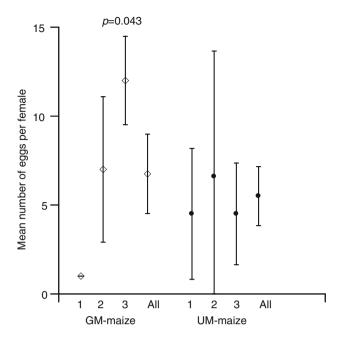


Fig. 4 Mean number of eggs ($\pm 95\%$ confidence intervals) per female *D. magna* fed on GM-maize and UM-maize in experiments 1, 2, and 3 and for all of them combined. The *p*-value of a significant difference within experiment 3 is shown (*t*-test)

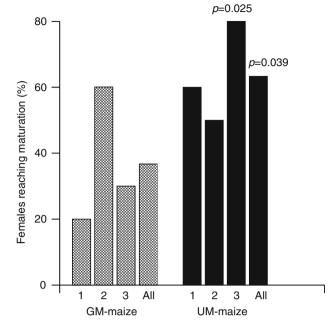


Fig. 6 Proportion of females that reached maturation in *D. magna* fed on GM-maize and UM-maize in experiments 1, 2, and 3 and for all of them combined. The *p*-values of significant differences (within experiment 3 and for the pooled samples) are shown (chi-square test)

19

18

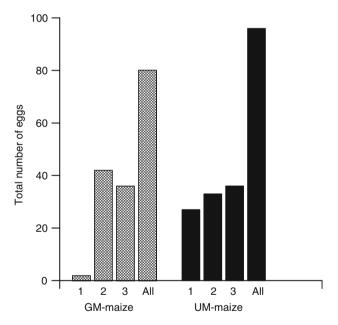


Fig. 5 Total number of eggs observed in *D. magna* fed on GM-maize and UM-maize in experiments 1, 2, and 3 and for all of them combined (total sum)

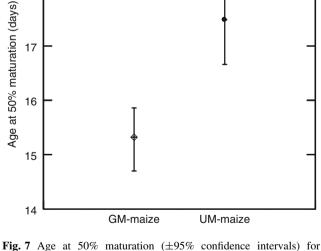


Fig. 7 Age at 50% maturation (±95% confidence intervals) for *D. magna* fed GM-maize and UM-maize. Confidence intervals were calculated from bootstrapping

fully randomized laboratory experiments. Three repeated studies, with the same experimental design, lasted over the entire life cycle of the test animals, allowing us to investigate the impact on a number of life-history traits.

Daphnia magna performed substantially better when fed the isogenic UM-maize compared to the GM-maize. The data pointed toward significantly reduced survival and a reduced proportion of females that reached maturity. These two variables are not independent, as the ability to become sexually mature clearly depends on the survival. Small differences were found in growth rates, but there was a tendency for the UM-fed animals to have higher growth



rates, and in one of the experiments (one out of six comparisons on adults), significantly larger body size was found in the UM-fed group compared to the group fed GM-maize. The individual fecundity showed a trend toward more eggs per female in the GM-fed group, and we observed a significantly higher fecundity in GM-fed animals in one of the experiments (one out of three). However, the overall reproductive output (*i.e.*, the total number of egg produced) was higher in the UM-fed groups. The group fed GM-maize reproduced, on average, significantly earlier than *D. magna* fed UM-maize.

The above results allow identifying the likely mechanism behind the negative fitness response of *D. magna* fed GM-maize on the basis of expectations from life-history theory corroborated by empirical data. The two mechanisms considered—limited nutrition versus low levels of toxicity—are expected to determine different configurations of life-history traits.

Reduced quantity as well as quality of food might lead to reduced clutch size and delayed reproduction in *D. magna* and *D. longispina* (Brett 1993; Enserink 1995), whereas only delayed reproduction was recorded for *D. galeata* (Vanni and Lampert 1992). Therefore, if the GM-maize in our experiments had a lower nutritional value than the UM-maize, we would expect *D. magna* to reproduce later and with smaller clutch sizes (*i.e.*, the opposite of our observations).

On the other hand, low-level toxicity is expected to reduce survival, a condition that, according to life-history theory, might trigger greater investment in reproduction early in the life cycle (Roff 2002). For example, higher juvenile mortality is generally linked to an early onset of reproduction (Stearns and Koella 1986). In organisms with plastic life-history traits, like *D. magna* (Enserink 1995), a response to low-level toxicity might thus lead to an allocation of resources to increase the fecundity early in the life cycle (Hansen et al. 1999; Mauri et al. 2003; Twombly et al. 1998). At high levels of toxicity, the general development of the organisms is severely impaired and all reproductive characters are negatively affected (*i.e.*, maturity is delayed and fecundity is reduced) (Enserink et al. 1995; Hansen et al. 1999; Mauri et al. 2003).

In our experiments, *D. magna* fed GM-maize reproduced earlier and had higher individual fecundity compared to *D. magna* fed UM-maize, but this was at the cost of survival and reproductive output later in life. From life-history theory, our results are likely to be explained by an allocation tradeoff after a toxic reaction (as opposed to reduced nutritional value) of *D. magna* to the GM-maize.

Our data represent a food/feed quality test of one Cry1Ab-transgenic versus the isogenic UM-maize variety grown in a specific environment. The strengths of the study include the following:

- 1. Its specificity. The diet consisted of 100% maize. Other feeding studies using GM feed have typically used diets of 60–76% from the transgenic material (Aeschbacher et al. 2005; Brake and Vlachos 1998; Ewen and Pusztai 1999; Reuter et al. 2002), sometimes as low as 25–33% (Hammond et al. 2006; Teshima et al. 2002). On the other hand, dozens of studies on terrestrial arthropods (*D. magna* is an arthropod) have used 100% plant diets to test impacts of transgenic plants, mostly related to Bt-plants (reviewed in Lövei and Arpaia, 2005). A summary of these studies show that 57% of the studied parameters showed significant negative impacts from the transgenic Bt-plants (Lövei and Arpaia 2005).
- 2. Its duration, through the whole life cycle of the animal: within 30 h after birth and up to a maximum of six reproductive events. Young and growing animals are likely to be more sensitive to food quality compared to adults because a healthy development depends on appropriate nutrition. Possible negative health effects of GM foods have been shown in young rats (Ewen and Pusztai 1999; Pusztai 2002) in studies that have been criticized and discussed in great detail but, unfortunately, not repeated. However, recent European regulations recognize the need for long-term feeding studies with and without spiking with the pure novel gene product (Knudsen and Poulsen 2007).
- 3. Its study design, with complete randomization (that corrects for potentially confounding factors in the experimental setup), lack of interactions between the experimental units (all experimental animals were independent of each other), double-blind measurements of body length (*i.e.*, the measurements were done without knowledge of which treatment group each animal belonged to), in a well-known model animal with minimal genetic variation (clonal) and with several fitness traits tested.

These strengths can also be seen as limitations; specificity comes at the cost of generality: (1) Only one species/variety/hybrid of GM-plant and its UM counterpart was used; (2) these were grown in only one type of environment; (3) only one type of model organism was used.

However, more that anything else, these limitations point to the need of further studies: Different varieties of different transgenic plants grown under a wide variety of environmental conditions should be tested with a comprehensive methodology. A reasonable test regime for genetically modified organisms must be based on a case-by-case approach (Kowalchuk et al. 2003), as also recognized by more than 140 countries committed to implementation of the Cartagena Protocol on Biosafety (Hill and Sendashonga 2006).



A natural follow-up of the present study would be to investigate dose responses of Bt-toxin on D. magna. At first sight, the Cry1Ab toxin seems to be the most likely cause of the reduced performance of the GM-maize-fed D. magna. However, the relatively low expression level of the transgenic product in our test maize (67 \pm 27 ng Cry1Ab toxin per gram of dried kernels), which is lower than what found even in pollen (<90 ng/g): http://www.agbios.com), might indicate that some other difference exists between the tested GM-maize and its UM isoline. An interesting observation in our study is that the GM-maize contained Bt-toxin (although at a low level) after more than 3 years in storage, indicating that Cry1Abtoxin is not effectively broken down in dried maize kernels.

In these feeding trials, we used maize as the sole food source. D. magna can survive on a multitude of diets, but, admittedly, this is an artificial situation. However, our study was not aimed at estimating the responses of D. magna under natural field conditions, where they would have a diverse diet. The intention was to perform an initial screening of potential Bt-maize effects on a nontarget organism. The results indicate that D. magna, and potentially also other related aquatic zooplankton species, might be vulnerable to transgenic Cry1Ab-maize. Although the causality of the observed effects still remains open, our results go along with others (Hilbeck and Schmidt 2006; Lövei and Arpaia 2005; Rosi-Marshall et al. 2007) and call for further testing of nontarget arthropods under varying conditions. Summary data from both the United States and Europe indicate that the expression of Bt-toxin is 20-30 times higher in maize leaves compared to maize grains (http://www.agbios.com). If Cry1Ab-toxin caused the effects on D. magna observed in this study, the considerably higher amount of expressed transgenic protein from the rest of the plant will have a larger potential to reach and affect nontarget organisms in the environment.

The large biomass of dead plants/roots that is left in the field after harvest represents a huge amount of exotic and potentially bioactive components for recipient ecosystems. Bt-toxin produced in GM-plants has been shown to retain 25-30% of its toxicity after 140 days when bound to soil (Palm et al. 1996). Adsorption and binding of Cry proteins to clay sometimes even enhance the insecticidal activity (Tapp and Stotzky 1995), thus increasing the likelihood that bioactive toxin is accumulated in the environment (Groot and Dicke 2002). At the same time, Bt-maize and isogenic controls have been corroborated to be substantially equivalent at the level of major nutrients and minerals and trace elements, both in kernels (Brake and Vlachos 1998; Sidhu et al. 2000) and in the whole plant (Clark and Ipharraguerre 2001). Clark and co-workers found a negative trend in growth for the terrestrial isopod Trachelipus rathkii when fed different Bt-maize varieties compared to their isolines, but they could not find any negative effect of even high concentrations of purified bacterial Bt-toxin on the same test organism (Clark et al. 2006). There are several possible explanations for such findings: (1) The Bt-toxin in the plant is different from the purified, bacterial version, due to, for instance, plant specific posttranslational modifications; (2) the nutritional quality of the transgenic plant and the UM isoline tissues are different, due to, for instance, overexpressed antinutrional factors; (3) some other difference(s) exist(s) between the GM and the isoline, due to, for instance, insertional effects on the expression levels of endogenous genes.

In conclusion, our study demonstrated significant and negative long-term effects after feeding a transgenic Bt-maize variety. The combination of life-history traits indicates a toxic response of *D. magna* to the GM-maize. Within our test system, we reject the null hypothesis that the tested GM-maize and UM-maize had the same quality as a food source. The observed effects of transgenic Bt-maize on *D. magna* call for greater attention, not only on the runoff material from transgenic agricultural fields but also on the sensitivity of aquatic nontarget organisms to transgenic products.

Acknowledgments We would like to thank Dr. Chito Medina and farmers from the Iloilo district in the Philippines for providing the maize samples used in the experiments. We are grateful to Professor Kaare M. Nielsen for valuable discussions related to the experiments. We also thank Dr. Idun Grønsberg, Marte Albrigtsen, Julia Eggert, and Elisabeth Olsen at the GenØk Lab in Tromsø and Dr. Morten Johansen and Kriss Rokkan Iversen at the Norwegian College of Fishery Science for practical assistance during the experiments. The studies were supported by a grant (Project No. 154504) from the Research Council of Norway.

References

Aeschbacher K, Messikommer R, Meile L, Wenk C (2005) Bt176 corn in poultry nutrition: physiological characteristics and fate of recombinant plant DNA in chickens. Poult Sci 84(3):385–394

Andow D, Hilbeck A (2004) Science-based risk assessment for nontarget effects of transgenic crops. Bioscience 54(7):637–649
Atienzar FA, Cheung VV, Vadesh NJ, Epledge MH (2001) Fitness parameters and DNA effects are sensitive indicators of copperinduced toxicity in *Daphnia magna*. Toxicol Sci 59:241–250

Barry MJ (1996) Effects of an organochlorine pesticide on different levels of organization in *Daphnia*. Ecotoxicology and environmental safety. Environ Safety 34:239–251

Brake J, Vlachos D (1998) Evaluation of transgenic event 176 "Bt" corn in broiler chickens, Poult Sci 77(5):648-653

Brandt SL, Coudron TA, Habibi J et al (2004) Interaction of two *Bacillus thuringiensis* delta-endotoxins with the digestive system of lygus hesperus. Curr Microbiol 48(1):1–9

Bravo A, Gill SS, Soberon M (2007) Mode of action of *Bacillus thuringiensis* Cry and Cyt toxins and their potential for insect control. Toxicon 49:423–435

Brett MT (1993) Resource quality effects on Daphnia-Longispina offspring fitness. J Plankton Res 15(4):403-412



- Campbell AK, Wann KT, Matthews SB (2004) Lactose causes heart arrhythmia in the water flea *Daphnia pulex*. Comp Biochem Physiol B: Biochem Mol Biol 139(2):225–234
- Clark BW, Prihoda KR, Coats JR (2006) Subacute effects of transgenic Cry1Ab Bacillus thuringiensis corn litter on the isopods Trachelipus rathkii and Armadillidium nasatum. Environ Toxicol Chem 25(10):2653–2661
- Clark JH, Ipharraguerre IR (2001) Biotech crops as feeds for livestock. Abstr Papers Am Chem Soc 222:U67
- Crickmore N (2005) Using worms to better understand how *Bacillus* thuringiensis kills insects. Trends Microbioly 13(8):347–350
- Domingo JL (2007) Toxicity studies of genetically modified plants: a review of the published literature. Crit Rev Food Sci Nutr 47:721–733
- Douville M, Gagne F, Blaise C, Andre C (2007) Occurence and persistence of *Bacillus thuringiensis* (Bt) and transgenic Bt corn *cry1Ab* gene from an aquatic environment. Ecotoxicol Environ Safety 66:195–203
- Dutton A, Romeis J, Bigler F (2003) Assessing the risks of insect resistant transgenic plants on entomophagous arthropods: Bt-maize expressing Cry1Ab as a case study. Biocontrol 48(6):611–636
- Dutton A, Romeis J, Bigler F (2005) Effects of Bt maize expressing Cry1Ab and Bt spray on Spodoptera littoralis. Entomol Exp Applic 114(3):161–169
- Enserink EL, Kerkhofs MJJ, Baltus CAM, Koeman JH (1995) Influence of food quantity and lead-exposure on maturation in Daphnia-Magna: evidence for a trade-off mechanism. Funct Ecol 9(2):175–185
- Ewen SWB, Pusztai A (1999) Effect of diets containing genetically modified potatoes expressing *Galanthus nivalis* lectin on rat small intestine. Lancet 354(9187):1353–1354
- Groot AT, Dicke M (2002) Insect-resistant transgenic plants in a multi-trophic context. Plant J 31(4):387–406
- Hammond BG, Dudek R, Lemen JK, Nemeth MA (2006) Results of a 90-day safety assurance study with rats fed grain from corn borer-protected corn. Food Chem Toxicol 44(7):1092–1099
- Hansen FT, Forbes VE, Forbes TL (1999) Effects of 4-*n*-nonylphenol on life-history traits and population dynamics of a polychaete. Ecol Appl 9(2):482–495
- Hilbeck A (2001) Implications of transgenic, insecticidal plants for insect and plant biodiversity. Perspect Plant Ecol Evol Syst 4(1):43-61
- Hilbeck A, Schmidt JEU (2006) Another view on Bt proteins: how specific are they and what else might they do? Biopestic Int 2(1):1-50
- Hill R, Sendashonga C (2006) Conservation biology, genetically modified organisms and the biosafety protocol. Conserv Biol 20(6):1620–1625
- James C (2006) Global status of commercialized biotech/GM crops. ISAAA Brief No. 35. 2006. ISAAA, Ithaca, NY
- Kluttgen B, Dulmer U, Engels M, Ratte HT (1994) Adam, an artificial fresh-water for the culture of zooplankton. Water Res 28(3):743–746
- Knudsen I, Poulsen M (2007) Comparative safety testing of genetically modified foods in a 90-day rat feeding study design allowing the distinction between primary and secondary effects of the new genetic event. Regul Toxicol Pharmacol 49(1):53-62
- Kowalchuk GA, Bruinsma M, van Veen JA (2003) Assessing responses of soil microorganisms to GM plants. Trends Ecol Evol 18(8):403–410

- Kramer KJM, Jak RG, van Hattum B, Hooftman RN, Zwolsman JJG (2004) Copper toxicity in relation to surface water-dissolved organic matter: biological effects to *Daphnia magna*. Environ Toxicol Chemistry 23(12):2971–2980
- Lövei GL, Arpaia S (2005) The impact of transgenic plants on natural enemies: a critical review of laboratory studies. Entomol Exp Applic 114:1–14
- Mauri M, Baraldi E, Simonini R (2003) Effects of zinc exposure on the polychaete *Dinophilus gyrociliatus*: a life-table response experiment. Aquat Toxicol 65(1):93–100
- Mendelson M, Kough J, Vaituzis Z, Matthews K (2003) Are *Bt* crops safe? Nature Biotechnol 21(9):1003–1009
- Nogueira ICG, Saker ML, Pflugmacher S, Wiegand C, Vasconcelos VM (2004) Toxicity of the cyanobacterium *Cylindrospermopsis* raciborskii to *Daphnia magna*. Environ Toxicol 19:453–459
- O'Callaghan M, Glare TR, Burgess EPJ, Malone LA (2005) Effects of plants genetically modified for insect resistance on nontarget organisms. Annu Rev Entomol 50:271–292
- Palm CJ, Schaller DL, Donegan KK, Seidler RJ (1996) Persistence in soil of transgenic plant produced *Bacillus thuringiensis* var kurstaki delta-endotoxin. Can J Microbiol 42(12):1258–1262
- Pusztai A (2002) Can science give us the tools for recognizing possible health risks of GM food? Nutr Health 16:73–84
- Reuter T, Aulrich K, Berk A, Flachowsky G (2002) Investigations on genetically modified maize (Bt-maize) in pig nutrition: chemical composition and nutritional evaluation. Arch Anim Nutr 56(1):23–31
- Roff DA (2002) Life history evolution. Sinauer Associates, Sunderland, MA
- Rosi-Marshall EJ, Tank JL, Royer TV et al (2007) Toxins in transgenic crop byproducts may affect headwater stream ecosystems. Proc Natl Acad Sci USA 104(41):16,204–16,208
- Sidhu RS, Hammond BG, Fuchs RL et al (2000) Glyphosate-tolerant corn: the composition and feeding value of grain from glyphosate-tolerant corn is equivalent to that of conventional corn (*Zea mays* L.). J Agric Food Chem 48(6):2305–2312
- Sims IR, Watson S, Holmes D (1993) Toward a standard Daphnia juvenile production test. Environ Toxicol Chem 12(11):2053–2058
- Stark JD, Banks JE (2003) Population-level effects of pesticides and other toxicants on arthropods. Annu Rev Entomol 48:505– 519
- Stearns SC, Koella JC (1986) The evolution of phenotypic plasticity in life-history traits: predictions of reaction norms for age and size at maturity. Evolution 40(5):893–913
- Tapp H, Stotzky G (1995) Insecticidal activity of the toxins from Bacillus-Thuringiensis subspecies kurstaki and tenebrionis adsorbed and bound on pure and soil clays. Appl Environl Microbiol 61(5):1786–1790
- Teshima R, Watanabe T, Okunuki H et al (2002) Effect of subchronic feeding of genetically modified corn (CBH351) on immune system in BN rats and B10A mice. J Food Hyg Soc Japan 43(5):273–279
- Twombly S, Clancy N, Burns CW (1998) Life history consequences of food quality in the freshwater copepod *Boeckella triarticulata*. Ecology 79(5):1711–1724
- Vanni MJ, Lampert W (1992) Food quality effects on life-history traits and fitness in the generalist herbivore Daphnia. Oecologia 92(1):48–57

