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The low levels of circulating hepatocyte growth factor in nephrolithiasis cases: independent from gene polymorphism

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Abstract Environmental and genetic factors are important in development of nephrolithiasis. In a recent study, it has been demonstrated that hepatocyte growth factor (HGF) has an anti-apoptotic effect and thus can reduce the adhesion of calcium oxalate monohydrate crystals to renal epithelial cells. The aim of this study was to evaluate the HGF serum levels and its two gene polymorphisms and possible association of the two in patients with nephrolithiasis. One hundred and five patients with nephrolithiasis and 70 healthy volunteers with similar demographic features were included in this study. Serum HGF levels were measured, and HGF intron 13 C>A (in 102 stone patients and 68 healthy subjects) and intron 14 T>C (in 99 stone patients and 56 healthy subjects) polymorphisms were determined using real-time polymerase chain reaction with TaqMan allelic discrimination method. There were no statistically significant differences in HGF intron 13 C>A and intron 14 T>C polymorphisms between the control and patient groups ($X^2 = 1.72$ *df* = 2; *p* = 0.42, and $X^2 = 0.68$ $df = 2$; $p = 0.71$, respectively). Mean serum HGF concentration was significantly lower in the stone disease patients than in the control subjects (1.05 \pm 0.63 pg/mL and 1.35 ± 0.58 ng/mL respectively, $p = 0.0001$). When allele distribution frequency between stone patients and healthy subjects was compared, there were no significant

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differences in intron 13 and intron 14 allele distributions between two groups ($p = 0.43$ and $p = 0.44$, respectively). It may be concluded from the findings that decrease in HGF levels may play a role in renal stone formation, independent from gene polymorphisms.

Keywords Nephrolithiasis · Hepatocyte growth factor · Gene polymorphism · Serum hepatocyte growth factor level

Abbreviations

COM Calcium oxalate monohydrate HGF Hepatocyte growth factor

Introduction

Urinary tract stone disease is a complex, multifactorial disorder with an incidence of >10 % in general population and the incidence may have some geographical variations [[1\]](#page-4-0). The anatomic, genetic and environmental factors, which include dietary intake of salt, protein, calcium and other nutrients, fluid intake, urinary tract infections, socioeconomic status of the individual, lifestyle and climate, are important in development of this disease [\[2](#page-4-1)]. The prevalence of symptomatic kidney stone formation is 10 % for men and 5 % for women. After a first stone, the risk of recurrence is 40 % by 5 years, and 75 % by 20 years [\[3](#page-4-2)]. Calcium is a major component of 85 % of kidney stones, largely as either calcium oxalate or calcium phosphate [[3,](#page-4-2) [4](#page-4-3)].

The stone formation mechanism is not yet clearly understood. It is thought that adhesion of calcium oxalate monohydrate (COM) crystals to the surface of renal tubular cells and the subsequent cellular response may constitute critical

pathogenic steps in the development of nephrolithiasis [\[5](#page-4-4)]. High levels of oxalate and COM crystals lead to injury and apoptosis in renal tubular cells, which thus increases crystal adhesion to the renal epithelial cells [\[5](#page-4-4)[–7](#page-4-5)].

Although initially thought to be a liver-specific mitogen, hepatocyte growth factor (HGF) was later reported to have mitogenic, motogenic (enhancement of cell motility), morphogenic, proliferative and anti-apoptotic activities in various cell types [[5,](#page-4-4) [8–](#page-4-6)[11\]](#page-4-7). HGF has protection and regeneration functions in various organs and systems such as kidney, liver, lung, gastrointestinal tissues, and cardiovascular, cutaneous and nervous systems [[12,](#page-4-8) [13\]](#page-4-9). For this reason, blood and tissue levels may increase after acute injury and illness [\[13](#page-4-9)].

Tei et al. [\[5](#page-4-4)] have shown that HGF reduces cell damage and apoptosis formed by oxalate and COM crystals and inhibits adhesion of COM crystals to the cells of renal tubules in stone-forming rats. Furthermore, they determined that HGF gene transfer significantly reduced crystal deposits and the number of apoptotic cells on the renal tubules in stone-forming rat kidneys [\[5](#page-4-4)].

Studies have shown that stone disease is a polygenic disease and calcification inhibitor protein genes such as osteopontin [[14\]](#page-4-10), fetuin-A [[15\]](#page-4-11), and matrix Gla protein [\[16](#page-4-12)] and polymorphisms of urokinase [\[17](#page-4-13)] and claudin-14 gene [[18\]](#page-4-14) may be in important development of urinary tract stones. Similarly, some receptor single nucleotide polymorphisms associated with calcium metabolism, such as calcium-sensing receptor $[19]$ $[19]$ and vitamin D receptor $[20]$ $[20]$, were investigated in nephrolithiasis cases. There is no clear evidence about the role of HGF gene polymorphism in stone disease.

HGF gene has been associated with kidney stone formation in experimental rat studies [[5\]](#page-4-4). The intron 13 C>A and intron 14 T>C substitutions are relatively common polymorphisms in HGF gene, and these polymorphisms have been related to several diseases including essential hypertension [[21\]](#page-4-17).

The aim of this study was to evaluate the possible association of hepatocyte growth factor serum levels in nephrolithiasis cases. We also examined its two polymorphisms in these patients.

Patients and methods

One hundred and five patients (68 males, 37 females) with nephrolithiasis and stone composition of COM or calcium oxalate dihydrate, who were treated at Urology Department of Ataturk University, and 70 healthy volunteers with similar demographic features, were included in the study. The control group consisted of healthy volunteers without a history of kidney stone or family history of stone disease. Stone samples were obtained either after extracorporeal

shock wave lithotripsy or surgery for treatment, and all patients had stone analysis by X-ray diffraction.

The study protocol was approved by the local institutional ethics board, and written informed consent was obtained from all subjects before participation. Before any intervention, blood samples were taken after an overnight fasting. Serum samples obtained from blood were stored at −80 °C until HGF assay. Genomic DNA was extracted from blood samples in EDTA tubes using a commercial spin column kit (PureLink Genomic DNA Mini Kit, Invitrogen, Inc. Carlsbad, CA).

Analyses of the HGF gene C>A polymorphism (rs2074725) in intron 13 and T>C polymorphism (rs2074724) in intron 14 were performed by TaqMan allelic discrimination method using real-time polymerase chain reaction (Applied Biosystems 7300). Allele specific TaqMan probes that utilized in analysis process were as follows: for intron 13 C/A of HGF gene 5′-FAM-GCACAAATTATAGTCCAGAGCTTACcGT CTGGCAAGCAGATGTGATCAGCT-Tamra-3′ and 5′-VIC GCACAAATTATAGTCCAGAGCTTACaGTCTGGCAA GCAGATGTGATC AGCT-Tamra-3′; forward primer 5′-CT ACCTCTGGAGGCACAAATTA-3′ and reverse primer 5′-GGGTACAACCTTCAGGACCA-3′ and for intron 14 T/C 5′-FAM-CTACAGGAGAAAGAAGTAGTGAGGAtTG AAAAAAGCCTATTGACAATTTAG-Tamra-3′ and 5′-VIC-CTACAGGAGAAAGAAGTAGTGAGGAcTGAAAAAA GCCTATTGACA ATTTAG-Tamra-3′.

HGF in serum samples were measured using an ELISA (BioVendor, GmbH, Heidelberg, Germany) according to the manufacturer's standard protocol. Intra-assay and interassay variations were 4.8 % and 5.4 %, respectively. Serum HGF levels were calculated as pg/mL.

Statistical analysis

Statistical analysis was done using SPSS version 11.5 (SPSS Inc, Chicago, USA). Distribution of continuous variables was determined with Kolmogorov–Smirnov test. Comparison of HGF levels was analyzed between the groups using Mann–Whitney U test and Kruskal–Wallis test. Chi-squared test was used for comparison of the distribution of HGF gene polymorphism. *p* value <0.05 was regarded the statistically significant.

Results

A total of 105 patient samples and 70 control samples were analyzed for serum HGF levels. Mean serum HGF concentration was significantly lower in the stone disease patients than in control subjects (1.05 \pm 0.63 ng/mL for patient group and 1.35 ± 0.58 ng/mL for control group, $p = 0.0001$.

Genotype distributions at HGF intron 13 C>A and intron 14 T>C gene polymorphisms in control and patient groups are shown in Table [1.](#page-2-0) There were no statistically significant differences in these gene polymorphisms between the control and patient groups $(p = 0.42, \text{ and})$ $p = 0.71$, respectively). When the allele distribution frequency between stone patients and healthy subjects were compared, there were no significant differences in intron 13 and intron 14 allele distributions between two groups $(p = 0.43$ and $p = 0.44$, respectively) (Table [2](#page-2-1)). For HGF intron 13 C>A, genotype could not be detected in three patients and two controls, thus the results of 102 patients and 68 control subjects were statistically analyzed. Also, intron 14 T>C genotype could be determined in 99 patients and 56 control subjects and statistical evaluation was done in these subjects.

In addition, the effect of HGF intron 13 C>A and HGF intron 14 T>C genotypes on serum levels of HGF was investigated. For HGF intron 13 C>A, serum HGF levels were not significantly different between CC, CA and AA genotype carriers among healthy subjects and patients with urinary tract stones ($p = 0.78$ for the control group, $p = 0.44$ for the patient group). In both groups, the serum HGF levels were not significantly different among TT, CT and CC genotype carriers for HGF intron 14 T>C genotypes ($p = 0.22$ for the control group, $p = 0.12$ for the patient group) (Fig. [1\)](#page-3-0).

Table 1 Genotype distributions at HGF intron 13 C>A and intron 14 T>C gene polymorphisms in control and patient groups

Polymorphism	Patients $(\%)$	Controls $(\%)$	\boldsymbol{p}
Intron 13			
CC	68 (60)	46(40)	$X^2 = 1.72, df = 2$
AA	8(80)	2(20)	$p = 0.42$
CA	28 (58)	20(42)	
Intron 14			
TT	70 (65)	37(35)	$X^2 = 0.68, df = 2$
CT	26(62)	16(38)	$p = 0.71$
CC	3(50)	3(50)	

Table 2 Comparison of allele distribution frequency at HGF rs2074725 and rs2074724 SNPs in patients and controls

Further, comparison of CC genotype patients with A allele carriers at rs2074725 polymorphism in HGF intron 13 showed no significant difference in terms of serum HGF levels ($p = 0.42$, $Z = -0.79$). Also comparison of TT genotype patients with the C allele carriers at rs2074724 polymorphism in HGF intron 14 showed no significant difference in terms of serum HGF levels ($p = 0.60$, $Z = -0.51$) (Fig. [2;](#page-3-1) Table [3](#page-4-18)).

Discussion

In the present study, we investigated the importance of C>A polymorphism in HGF gene intron 13 and T>C polymorphism in intron 14 in patients with urinary tract stone disease. We detected that C>A polymorphism in HGF gene intron 13 and T>C polymorphism in intron 14 did not correlate with urinary tract stone disease. Additionally, A and C allele frequency for intron 13 and C and T allele for intron 14 in HGF gene of patients was not different from healthy controls. Furthermore, in patients with nephrolithiasis, serum HGF concentrations were significantly lower when compared to the control group. In addition, we found that HGF intron 13 and 14 genotypes had no effect on serum HGF levels.

It has been suggested that HGF's anti-apoptotic effect might result in the reduction of adhesion of COM crystals [\[5](#page-4-4)]. Furthermore, patients with nephrolithiasis had lower levels of serum HGF than the healthy control group in our study. It may be speculated that decrease in HGF production may be important factor in the formation of stone in kidneys.

Recent studies have stated that HGF is a multifunctional molecule [\[22](#page-4-19)[–24](#page-5-0)]. HGF gene polymorphism studies are limited in the literature. Polymorphism studies were performed in diseases such as hypertension [\[21](#page-4-17)], myopia [[25,](#page-5-1) [26](#page-5-2)], autism [[27\]](#page-5-3) and retinopathy of prematurity [\[28](#page-5-4)], and some HGF gene region polymorphisms were found to be associated with these diseases. For example, Motone et al. [\[24](#page-5-0)] studied HGF gene C/A substitution in intron 13, T/C substitution in intron 14 and T/A substitution in intron 8 in patients with essential hypertension. They found that genotype distributions are 83 % for CC, 16 % for CA and 1 % for AA in HGF gene intron 13 C/A. They have suggested that the prevalence of hypertension was significantly higher in individuals with the CC genotype, and the presence of A allele reduced the risk of essential hypertension in lean or female subjects. In the same study, authors could not find any significant association between the distributions of allele with hypertension. Additionally in some studies, it has been shown that serum HGF concentration was significantly higher in the hypertensive subjects than in the normotensive ones [[29,](#page-5-5) [30\]](#page-5-6). HGF gene polymorphisms did not

Fig. 1 Serum HGF concentrations of both control and study groups in CC, CA and AA genotype carriers for HGF rs2074725 polymorphism (**a**) and TT, CT and CC genotype carriers for HGF rs2074724 polymorphism (**b**)

Fig. 2 *Boxplot* shows serum HGF levels of the CC genotype patients and the A allele carriers at rs2074725 polymorphism in HGF intron 13 *Boxblot* shows serum HGF levels of the TT genotype patients and the C allele carriers at rs2074724 polymorphism in HGF intron 14

affect the risk of development of autism in many regions of the intron from 1 to 15 is shown by the Toyoda et al. [\[27](#page-5-3)].

HGF seems to function in protection of the myocardium with preventing apoptosis of myocardial cells and by increasing the formation of a blood vessel in ischemic field [[31\]](#page-5-7). Studies have shown that serum HGF levels were markedly elevated in patients with acute myocardial infarction [\[31](#page-5-7), [32\]](#page-5-8). Chen et al. [\[33](#page-5-9)] showed that HGF gene transfer before ischemia reduced the myocardial infarct size and protected left ventricular function via multiple **Table 3** Serum HGF concentrations of both control and study groups in CC, CA and AA genotype carriers for HGF rs2074725 polymorphism and TT, CT and CC genotype carriers for HGF rs2074724 polymorphism

beneficial actions, such as antioxidant, anti-apoptotic, and anti-fibrotic actions, and increased expression of bcl-2 and angiogenesis.

It may be concluded from our findings that decrease in HGF levels has a role in renal stone formation independently from gene polymorphisms. We think that decrease of HGF levels may be a cause of the crystal precipitation via increase of apoptosis in kidney cells. Further studies investigating polymorphism of HGF gene regions different from this study are need.

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Conflict of interest The authors declare that they have no conflict of interest.

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