

## C6-Like and C3-Like Molecules from the Cephalochordate, Amphioxus, Suggest a Cytolytic Complement System in Invertebrates

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**Abstract.** The mammalian immune system has cytotoxic mechanisms, both cellular and humoral, that destroy the membrane integrity of target cells. The main effector molecules of these cytolytic mechanisms—perforin, used by killer lymphocytes, and the membrane attack complex (MAC) components of the complement system—share a unique module called the MAC/perforin module. Until now, both immunological cytotoxicity and the MAC/perforin module have been reported only in jawed vertebrates. Here, we report the identification of a protein containing the MAC/perforin module from the invertebrate cephalochordate, amphioxus (*Branchiostoma belcheri*), using expressed sequence tag (EST) analysis of the notochord. The deduced amino acid sequence of this molecule is most similar to the primary structure of human complement component C6 and is designated AmphiC6. AmphiC6 shares a unique modular structure, including the MAC/perforin module, with human C6 and other MAC components. Another EST clone predicts the presence of a thioester-containing protein with the closest structural similarity to vertebrate C3 (therefore designated AmphiC3). AmphiC3 retains most of the functionally important residues of vertebrate C3 and is shown by phylogenetic analysis to be derived directly from the common ancestor of vertebrate C3, C4, and C5. Only opsonic activity has been assigned to the invertebrate complement system until now. Therefore,

this is the first molecular evidence for complement-mediated immunological cytotoxicity in invertebrates.

**Key words:** Immunological cytotoxicity — Modular structure — Complement — Amphioxus — Membrane attack complex

### Introduction

The mammalian immune system is equipped with both cell-mediated and humoral cytotoxic mechanisms. Cell-mediated cytotoxicity is mediated by cytotoxic T cells and natural killer (NK) cells, and perforin acts as an effector molecule (Lichtenheld et al. 1988; Shinkai et al. 1988). Upon activation of these cells, perforin stored in their lytic granules is released and polymerizes to generate transmembrane pores in target cell membranes. Humoral cytotoxicity is instigated by the assembly of the terminal components of complement, leading to the formation of a similar pore called the “membrane attack complex” (MAC) (Plumb and Sodetz 1998). The complement components involved in MAC formation are C5, C6, C7, C8 $\alpha$ – $\gamma$ , and C9, and several of these (C6, C7, C8 $\alpha$ , C8 $\beta$ , and C9) share a characteristic modular structure that includes the MAC/perforin module, which is also found in perforin. Therefore, all pivotal molecules involved in immunological cytotoxicity seem to have originated from a common ancestral molecule containing the MAC/perforin module. These molecules have been identified only in mammalian species so far, except for the identification of MAC (Nonaka et al. 1981) and

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cDNA clones for C8 and C9 (Katagiri et al. 1999; Stanley et al. 1985; Yeo et al. 1997) in bony fish. Therefore, the evolutionary origins of the immunological cytotoxic molecules are largely unknown. Recent progress in genome science has cast some light on the evolution of the immunological cytotoxic molecules containing the MAC/perforin module. There are only six copies of this module in the human genome (Venter et al. 2001), indicating that only perforin and the MAC components contain this module. Moreover, there is no MAC/perforin module in the *Drosophila* genome (Adams et al. 2000). Therefore, this module seems to have been established in the deuterostome lineage, although its presence in invertebrate deuterostomes is yet to be demonstrated.

Two major parts of the vertebrate immune system, adaptive immunity and innate immunity, have quite different evolutionary origins. The former originated in the jawed vertebrate lineage, whereas the latter has been traced back to the protostomes (Hoffmann et al. 1999). Although the mammalian complement system is intrinsically involved in both innate and adaptive immunity, accumulating evidence indicates that this system was established as one of the major components of innate immunity in the deuterostome lineage, well before the emergence of adaptive immunity (Nonaka 2001). The central component C3 and/or its activating enzyme factor B (Bf) have been identified in sea urchins (echinoderms) (Al-Sharif et al. 1998; Smith et al. 1998), ascidians (urochordates) (Nonaka et al. 1999), and lamprey (Nonaka and Takahashi 1992; Nonaka et al. 1994) and hagfish (Ishiguro et al. 1992) (both cyclostomes). In contrast, the recently published *Drosophila* (Adams et al. 2000) and *Caenorhabditis elegans* (*C. elegans* Sequencing Consortium 1998) genomes do not contain identifiable C3 and Bf homologues, suggesting that the complement system is unique to the deuterostomes. The mammalian complement system has three activation pathways, referred to as the classical, alternative, and lectin pathways, and one cytolytic effector pathway that leads to membrane damage (Volanakis 1998). All jawed vertebrates so far analyzed possess all four pathways, whereas only components of the alternative and lectin pathways have been identified in the apparently primitive complement systems of the sea urchin, ascidian, and cyclostomes (Nonaka 2001). Therefore, the evolution of the complement system seems to involve an increase in the number of components and activation pathways, probably via gene duplications, with the emergence of jawed vertebrates. Moreover, from a functional viewpoint, only opsonic activity that enhances phagocytosis has been identified in the ascidian and lamprey complement systems (Nonaka et al. 1984, 1999). It is still unclear whether these primitive complement systems also have cytolytic activity.

Cephalochordata is classified as a subphylum of Chordata, together with Urochordata and Vertebrata.

Chordata, Hemichordata, and Echinodermata are considered the only deuterostome phyla. Although the phylogeny of the deuterostomes is still in dispute, recent molecular data indicate that Vertebrata is monophyletic and a sister clade of Cephalochordata. Urochordata is also monophyletic and constitutes a sister group to Vertebrata and Cephalochordata (Cameron et al. 2000; Wada and Satoh 1994). The presence of myotomes in vertebrates and cephalochordates and the presence of the tunic in the urochordates further support this phylogenetic relationship. Therefore, Cephalochordata occupies an important position in the deuterostome phylogeny as the closest relative to Vertebrata and provides a unique opportunity to analyze the evolution of the vertebrate immune system. However, there is still no report on the cephalochordate immune system at the protein or cDNA level.

## Materials and Methods

**Animals and cDNA Cloning.** Adult amphioxus, *Branchiostoma belcheri*, were collected near the National Research Institute of Aquaculture, Mie, Japan. Partial cDNA clones for possible complement genes of amphioxus were originally derived from a *B. belcheri* notochord cDNA library by EST analysis (Suzuki and Satoh 2000). Full-length clones were isolated from the same Uni-ZAP XR library (Stratagene, La Jolla, CA) using the partial cDNAs as probes and were converted into pBluescript SK(-) phagemids using a rapid excision kit (Stratagene). The complete sequences of the inserts in these clones were determined using a Big-Dye Terminator Cycle Sequencing Ready Reaction kit and an ABI PRISM 377 DNA sequencer (Perkin Elmer, Norwalk, CT). Each sequence was determined at least twice from both strands.

**Phylogenetic Analysis.** The deduced amino acid sequences of the amphioxus complement genes were aligned with the corresponding sequences of various animals using the CLUSTAL W software (Thompson et al. 1994). Based on this alignment, a phylogenetic tree was constructed using the neighbor joining method (Saitou and Nei 1987).

## Results

**Identification of Two Possible Complement Component Genes from Amphioxus.** Expressed sequence tag (EST) analysis of the amphioxus notochord (Suzuki and Satoh 2000) resulted in the identification of two possible complement genes that, according to a BlastX homology search, are most similar to vertebrate complement components C6 and C3. The complete primary structures of these proteins, designated AmphiC6 and AmphiC3, respectively, were deduced from the nucleotide sequences of full-length cDNA clones.

**Structure of AmphiC6.** One of two AmphiC6 EST clones contained a full-length protein-coding region within a 3.5-kb-long insert. The deduced complete primary structure of AmphiC6 predicts a characteristic

|   |      |
|---|------|
| ACCAGGTTGGACAGACTTCAACATCGCCACAAGAAGAGATTTTCCCTACTTCACTGTCTCCACTGTACTTCCACAGACATGAGGTGCATGTTTCCGCTGCTGGTGTGGCGCGGTT       | 120  |
| <b>M F R L L V L L A A V</b>  | 10   |
| GTGCGGTTCTGACCGTACCGCTTCAGCGCTGGCGGCGGCGAGGAAGAAGAGCTGCCCGCGCCACCTCTCCACCCTGCCAGTCCGCGGGCTGGTCTCCCTGGTCCACATGT            | 240  |
| <b>Leader Sequence</b>  |      |
| <u>V A V L T V P S</u> D G W R R R R R R R R R R A P P P P P P F V H C Q V G G W S P W S T C                              | 250  |
| TCCAGTCTTTGTCAGGCGGGACCCAGGCGCGCAATCCCGCAATCCCGCATGCGCCGACAGGACCGCGGTCCACCTCTCCGTCGAATCAGCCAGGTCCAACTTACAGACATGT          | 360  |
| <b>TS1</b>  | 90   |
| CCCATCGACTGCAATTTGGGAACGTTTGGGAACAGGCAGCAGGCGCACCCCTGTGAGAGGAAGGAGGAGCGTGACCGGCTGGTGTACAGGCCAGTCAAGGGCCAGCGGCTGCC         | 480  |
| <b>TS2</b>  | 130  |
| GCCGAAGCTGCAGAGACGGCGGCTGCCTCACCAACGAGCGCTGCCCGCGCAAGGACAGCTGCAGGGAGAACGAGTTCACCTGCCAAAACAAGCGGTGCATCCCGGAGTCCAGACATGT    | 600  |
| <b>LDLRA</b>  | 170  |
| AACGGGATAACGACTGTGGAGACTTCTCCGACGAGCGGCGGTGTGAGCAGTCTTTGACGTGTGAGCGGACAGGACTTCGACGCTATCCCAACATTTAGTTCCGTGGCAGCGGGTAC      | 720  |
| <b>MAC/P</b>  | 210  |
| AACATCTGTCGGGAGAGVTGTCGGGAAGTTCGGAGAACAGGCTGACGGACAGCTGTCTACAGCGGAGACCACGGACAGGACTCCCGTCCGAGAGAACCTACAA                   | 840  |
| <b>EGF</b>  | 250  |
| AACTACCGTTCCAAGTTCTCGTGACAACTCTTCGCCACCAGCGGTACACAGCGCGGGAATACTACCAGGACGAGGTCCAACTCCAGCGGAACCTCAAGGTCCGCGGGTCCG           | 960  |
| <b>TS3</b>  | 290  |
| GGCAGTTCTTCGTCAATCAGGCGAGCGCGGCTGCGCACAGCGTCCCGCATGACTCAGGAAGTCAATCGAGTCCGCGAGAAAATAGACTCCAAGTACTCCAAGTATGAACACA          | 1080 |
| <b>TS4</b>  | 330  |
| GTTGAGTGGCTCAGTTCAAGTATGCGCAGAAGCGCTCAACCCTCCGACATCTTCTTCGCGGCATGAAAGGACTTACCTGTCTACTACAACACTTGGACTACAGTTTCTTATTGA        | 1200 |
| <b>TS5</b>  | 370  |
| GACTTGGCAGCAGCTACTTTTCTCGGGAAGTCTAGGGCAGTATGAGTACGTTACCCGATCAGCAGAGCCAGTCTGCTCAGCTCTGCTGACAGGAGGAAAGAAACAGTCGTGT          | 1320 |
| <b>TS6</b>  | 410  |
| CTGAGTCCCGAAGTCAAGSCCTCGTTCTTCAGTTCCTCCGCGCAGCAGCAGCGCAGCAGGTCGCAAGAAGAAGCGGTTTCACAGAGGAACAGCGAAGTTCACCTTCGTCCGAG         | 1440 |
| <b>TS7</b>  | 450  |
| TCCTTCTCCAGCTGAAAGGGCGGCTGTCGGAAATCCGCGCGCAGTGGCTTCGCCAAGGACGACCCGAGGAGTACGAGGCGTGGATCCAGGAGCTCAACCGGAAACCCCGCTATC        | 1560 |
| <b>TS8</b>  | 490  |
| ATCAGCTACGAGTACCGCCATTTCTGAGCTTGTGCGGCTCCCTACCGCGCATCAAGCGGGCTAACATGGAGAAGGCTCTGGTAATACCTGCACTTCCAGCTCCCTGCAAG            | 1680 |
| <b>EGF</b>  | 530  |
| GTGAATCTCCTGTCAGAACAGCGCGAGCCATCATGATCGGCACGGAATGCGCTGCGCTGCAAGCGGGAAGTCAACGCGTCTCCGCGAGCTGGGAGCCAGGTGACAAAGTCGGT         | 1800 |
| <b>TS9</b>  | 570  |
| ACTGCACTCAGCGAAGTGGCGGTTCCTGGGCTTCGGGCGCGTGCAGTAAGTCTCTGGGTCAGGACAGCGGGTCCGCGCGCAGCTGCGCAACCCCAACCCCGGAACACGGC            | 1920 |
| <b>TS10</b>   | 610  |
| CAGACCTGCCCGGAGCAGGATCCGAGACCGGAGCTTAACCTCACACCCATGCCCTCACGCAATCGCCGAGGACCGGAGGCTACTACTCCGTCAGGCTAGCAGCGGTCAGCGGACCTCGCGC | 2040 |
| <b>Unknown</b>  | 650  |
| GCGCCTTCAGACCTCCCGGCGCAGAAGACTGCTGCGGCTTCGCCGCGGAAAACACCGGCGGCTCCGCAACGATCTGTTCAAGACTCTGTTGAAGTGAAGAGCTGTTGA              | 2160 |
| <b>TS11</b>   | 690  |
| TCAGACCGCGGCTTCCTGCGTCAGGCTGGTCTTCAGGATCAGCAGGCTCAAGAACAGGTCGCGGCGGATAGGCCAGAGATTCTTCGGCTCCAGAGGCACTAATAGCTAGCTATAGT      | 2280 |
| <b>TS12</b>   | 730  |
| GACGGGCAACTCCGGTATCCAGTGGCTCGTCACTGGTGTAGAGTGGGTGAAGACAGACAGTATAGGACACGATATGATACAGCTCAAGCGACAGCTTGAAGAGGACGATGATCAA       | 2400 |
| <b>TS13</b>   | 770  |
| ACAGACCGCCATCCGCAATCCCGTTCGAGACACCAACGGAACATCTTCCGCACTCCCGGAGCGGTGCGGAGGAGTCCCGGAACTACCGAGCGGCGAACCGATATA                 | 2520 |
| <b>TS14</b>   | 810  |
| TCTGATGAGAAAGCAGGATCTCGAGCGGAGTGGTGTCTTCAGGATGTCTTCAGGTCCTGATCTGACACGTCACCAGGAGTCTGACAGTTCACCGTCCAGGAGATGCT               | 2640 |
| <b>TS15</b>   | 850  |
| GAAAGCTCACAGGAGATCTGACAGTCCAGGAGATGCTGAAACGCTCACAGGAGATGCTGAAACGCTCACAGGAGATGCTGAAACGCTCACAGGAGATGCTGACAGTCCAGGAG         | 2760 |
| <b>TS16</b>   | 890  |
| GATGCTGAAACGTCACCGAGATCTGATGACTCACTCAAGATCCCGGACACCGAGAGAACGAGATGACTACTACTATGATTATGATGATGAATCGACATCGTATGTCAA              | 2880 |
| <b>TS17</b>   | 921  |
| TATCTACTCGAAGGAGAATTCGAAGAGTCTGTCCGACATACAAAGAGTGAATAGGAAGCTGAATAGTCGAAATATTAGATTTTTACTGTTTCAACATTTATGCTTTT               | 3000 |
| <b>TS18</b>   | 3120 |
| CCGAAATTTTACAGGATAAACAAGACTTCGATTCTTTTAAGAAAAGTGACATTTTATGATTAAAGGAAGATTTGATACATGTATGACATGTCACAAAGCTTTTATACGTACAAG        | 3240 |
| <b>TS19</b>   | 3360 |
| AGTGAATCTCCCTTATGAGGAAATGCTTTTACAAATCAATGTTTCCTCAAATACATACATTTATCTGACATTTTCAGATACATGCTGCTTTTTCATTGCGCCTGGTAGGAAATG        | 3480 |
| <b>TS20</b>   | 3526 |
| CTTCACCTTCCCTTATTAGACGCTTAGCATCTTACATGCTGCTGCATAGGATGATGCTTTTATTAACGACCGTCCAGTATGAAACAGTGTACATGTCACAGACT                  | 3600 |
| <b>TS21</b>   | 3526 |
| AACGTCAATTAAGTATGGTCAAATGAAAAAATAAAAAA  |      |

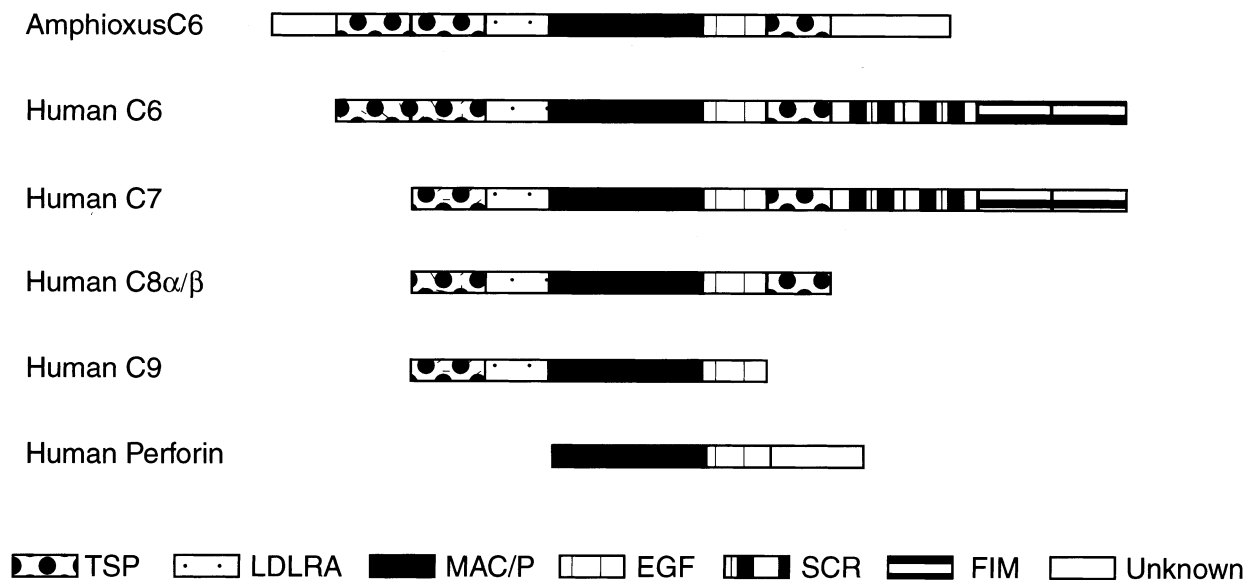
**Fig. 1.** Nucleotide and deduced amino acid sequences of AmphiC6. A putative leader sequence is shown in *boldface* and *underlined*. The N-terminal ends of each module are indicated by arrows below the amino acid sequence. The characteristic consecutive R residues (22–

28), consecutive P residues (30–36), and DAD/ETSPG repeats (835–897) are shown in *boldface*. Also shown in bold are the polyadenylation signal and the poly(A) tail at the 3' end.

module structure in its central portion (Figs. 1 and 2). This module structure, reading from the N-terminal end, comprises two TSP (thrombospondin type 1) modules, one LDLRA (low-density lipoprotein-receptor class A) module, one MAC/perforin (membrane attack complex/perforin) module, one EGF (epidermal growth factor) module, and one TSP module and is identical to the six-N-terminal module structure of human C6 (DiScipio and Hugli 1989; Haefliger et al. 1989) (Fig. 2). The four C-terminal modules of human C6, two SCR (short consensus repeats) and two FIM (factor I modules), are missing from AmphiC6. Compared with the module structures of human C8 $\alpha$  (Rao et al. 1987) and C8 $\beta$  (Haefliger et al. 1987; Howard et al. 1987), the central module structure of AmphiC6 has one extra TSP module at its N terminus. The module structure of other complement MAC components, C7 (DiScipio et al. 1988) and C9 (Stanley et al. 1985), and perforin (Lichtenheld et al. 1988; Shinkai et al. 1988) are less similar to that of

AmphiC6 (Fig. 2). From the shared module structure, it is apparent that AmphiC6 is evolutionarily related to perforin and the MAC components of the mammalian complement system.

For phylogenetic analysis, the amino acid sequences of the TSP, LDLRA, MAC/perforin, and EGF modules shared by AmphiC6 and the human MAC components and the corresponding region of perforin were aligned using the CLUSTAL W software. Part of this alignment, corresponding to the entire MAC/perforin module, is shown in Fig. 3. In the MAC/perforin module, AmphiC6 shares 36, 33, 30, 27, 25, and 27% amino acid identity with human C6, C7, C8 $\alpha$ , C8 $\beta$ , C9, and perforin, respectively. Four cysteine residues in the MAC/perforin module of AmphiC6 are perfectly conserved in all human MAC components and perforin. The membrane-interaction site identified in human C9 (Peitsch et al. 1990) is marked by a line over the alignment (Fig. 3). The degree of sequence similarity between AmphiC6



**Fig. 2.** Module structures of AmphiC6, human MAC components, and perforin. Each module is shown by a different shading pattern, with its name given at the bottom.

and the human MAC components and perforin is highest in this region, implicating it in a membrane-interacting function.

The phylogenetic tree based on this alignment of the four modules was drawn using the neighbor-joining method. As shown in Fig. 4A, AmphiC6 forms a cluster with human C6, and this clustering is supported by a high bootstrap value (97%). This result suggests that gene duplications among the MAC components predated the divergence of the cephalochordates from the vertebrates. However, this is an unrooted tree and further studies, which may include a search for C7, C8, or C9 genes in amphioxus, are required to clarify this point. A phylogenetic tree constructed using only the MAC/perforin module had the same topology (data not shown). In addition to the central module structure, AmphiC6 has unique sequences in its N- and C-terminal regions. These unique sequences show no significant similarity to any known sequence when analyzed by a BLAST search of GenBank. The N-terminal region is short and contains Arg and Pro stretches. The C-terminal region is about 300 amino acid residues long and contains nine consecutive repeats of the heptapeptide DAD/ETSPG (Fig. 1; bold-face letters) at its C-terminal end.

**Structure of AmphiC3.** The predicted prepro-AmphiC3 is 1792 amino acids long (Fig. 5) and shows sequence homology with C3, C4, and C5 of other species throughout its entire length. The amino acid sequence identity is 29% with human C3 (de Bruijn and Fey 1985), 29% with human C4 (Belt et al. 1984), 26% with human C5 (Haviland et al. 1991), and 25% with human  $\alpha$ 2-macroglobulin ( $\alpha$ 2M) (Kan et al. 1985). There is a possible  $\beta$ - $\alpha$  processing site, and the typical thioester site containing a catalytic His residue (Dodds et al. 1996).

The possible C3a region contains six Cys residues, perfectly conserved in vertebrate C3a, C4a, and C5a but not conserved in the ascidian (Nonaka et al. 1999) or sea urchin (Al-Sharif et al. 1998) C3 sequences. Furthermore, the C-terminal region of AmphiC3 shows the characteristic distribution of eight cysteine residues that are shared by all the C3, C4, and C5 sequences of vertebrates and invertebrates so far determined, but are not found in  $\alpha$ 2M, including the *C. elegans* and *Drosophila* thioester proteins (*C. elegans* Sequencing Consortium 1998; Lagueux et al. 2000). Therefore, AmphiC3 is identified conclusively as a member of the C3/C4/C5 family, and not as  $\alpha$ 2M. Phylogenetic analysis using the neighbor-joining method with human  $\alpha$ 2M as an outgroup shows AmphiC3 located outside the vertebrate C3, C4, and C5 cluster (Fig. 4B). This result strongly suggests that the gene for AmphiC3 was derived directly from the common ancestor of the vertebrate C3, C4, and C5 genes and that the gene duplications giving rise to the latter occurred in the vertebrate lineage after the divergence of Cephalochordata.

## Discussion

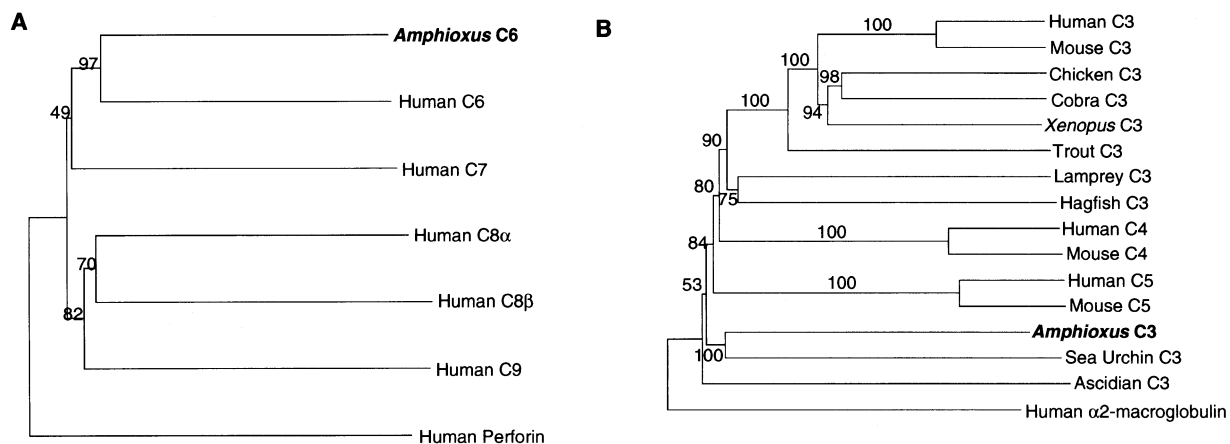
This is the first report of an invertebrate gene encoding the MAC/perforin module, suggesting an ancient origin for the cytotoxic pore-forming mechanism that uses this module. All other modules found in AmphiC6 (TSP, LDLRA, and EGF) are present in the *Drosophila* or *C. elegans* genomes, whereas the MAC/perforin module does not occur in these protostome genomes (Venter et al. 2001). Therefore, it is probable that the MAC/perforin module arose in the deuterostome lineage and that the TSP, LDLRA, and EGF modules were recruited to es-

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282
AmC6 -- QIFDVCGDQDFAIPIPIEFAGSGNIIISGEVAG--KVLNRLYG--GTCTNITVYS--GDHGKSFRLPENLQNYRFOVL-ADNSFAITAYNSAEAYRETRNSQ
HuC6 --RTKAVCT-RKYNPIPSVQIMGMGFHFLAGEPRG--EVLDNSFTG--GICKTVKS-----SRTSNPYRVEANLENVYFVQTAEDDLKTFYKDLTSLGHNEQOQS
HuC7 DSERRPSCD--IDKPPNPIELTGTNGYNELTGQFVN--RVINTKSFQ--CQCRKVFSS-----GDGKDFYRLSGNVLSYTFQVK-INNDNFYEFYNSFTWSYVKHTSTFHT
HuC8a VRAIDEDCS--QYEPPIPGSQAAALGYNILITQEDAQ--SVVDASYG--CQCEYVINGEWRELRYDSTCERLYYGDDEKFKYFRKPNFLKYHFEAL-ADTGISSEFYDNANDLLSKVKDKDS
HuC8b --RIYKCKQ--HEMDOYMGIGSLASGINLFTNSFEG--PVLDRHYA--GGCSPHYI-----LNTFRKPKYVNESYTPQIQ--GKVEFILLKEYESYSDRBRNVTEKMA
HuC9 --EPRPPCRORVVESELARTAGYGINILGMDPLS--TFPDNEFYN--GLCNRRDRD-----GNTLYYRFRPWNVAASLIYETK-GEKNFRTEHYEBEQIEAFKSIIQEKT
HuPF ---RSECK--RSHKFPVPGAWLAGEGVDT*SLRRSGSFPVD*TRQFLRPDGT*CTILCEN-----ALQEGTLQRLPLALITNWRAGSGCQRHVTRAKVSSTEAVARDAARSIR
378
AmC6 RNVKVGSGS-----FFVKAASGVSHSQSRMTHIEBSAQKIDSYPKVMNTVELAQKMR--SDLNPSDIFLRRMKDLPVYNYL---DYSFLIEDFGFTHYFS
HuC6 FSSOGGSFVSP-----IFYSSKRSEININHS--AFKQAIQASHKKDSSFRIHKVMKVLNFTTKA-KDLHLSDVFLKALNHLPLEYNSA---LYSRIFFDFGTHYFT
HuC7 SSSR---KRS-----FFRSSSSSSRSYTS-----HTNEIHKGKSYQLLVVENTVEAQQFINNPEFLQAEFPWKELSHLPSLYDYS--AVRRLDQYGHYLLQ
HuC8a AGSPLLVGVGVSHSQSDT*SFNLELNKYNKFKIFTRIFTKVQTAHFMRK--DDIMLDEGLQSMELPQYNYG--MWAKFINDYGHYIT
HuC8b SKSGFSFGFKIP-----GIFELGISSQSDRGG--HYTRRTKRFSSHOKSVFLHARSDELVAHFKLP--RSLMLHYEFLQRMFLPLEYSYG---EYRDLFRDFGTHYIT
HuC9 SNFNAAI*SLKFTPTETNKAEQCCCE*ETASSI*SLHGKGSFRF*SYSKNETYQL*FL*YS*SSKKEKMF*HL*VK*GEL*HL*GF*VMRN--RDVLT*TT*V*DD*IK*AL*PT*YEKG---EY*FA*LE*TY*GH*Y*SS
HuPF NDWKVGLDVT*PK-----PTS*NVHVS*VAG*SH*SQ*AA*FAAQ*TKHQDQY*SF*ST*DT*VE*CR*FY*SF*H*V*HT*P--P*L*H*P*D--*F*K*RA*L*G*L*D*L*P*H*H*F*NA*ST*Q*P*AY*LR*LI*SN*Y*G*TH*F*IR
490
AmC6 SGLGGQYEVYRYSRADLSHSGL*TEEQK*CLS-----AEAKA*FFF*P*F*SG*SS*GR*KEN*AL*SQ*RN*SG*FT*LI*SA*SE*F*SH*V*KG*SS*SA*G*Q*LA*F*AN*G*PN--*PEK--*YE*AM*IQ*DK*RN*P*AI
HuC6 SGLGGVYDILLYQFSSEELKN*SGL*TEEQ*AKH*CVR-----I*ET*KR*VL*FA*KK*TK*V*E*HR*CT*TN*KL*SE*KK*H*E*G*SF*IO*GA*E*K*SI*LI*RG*SR*SE*Y*GA*AL*AW*E*KS*G*SG*LE*E*KT--*P*SE*W*LE*SV*KE*NP*AV
HuC7 SGLGGEYRVL*FY*VD*SE*KL*KQ*ND*EN*SV*E*E*KK*CK-----S*SS--*G*W*H*F*V*V*K*F*SS--*H*G*CK*E*LE*N*AL*K*AA*SG*TO*NN*V*LR*G*E*P*FI*RG*G*AG*FI*SG*LT*Y*LE*LD*NP*AG*N*KR*RY*SA*W*AB*SV*TN*L*P*QV
HuC8a SGMGGI*Y*E*Y*IL*V*ID*K*AK*ME*SL*G*IT*SR*DI*TT*CF*G--G*SL*G*I*Q*Y*E*DK*IN*GG*GL*G*DH*CK*FK*G*G*K*TER*AK*MA*VED*II*SR*V*RG*SS*G*W*G*H*LA*Q*NR*ST*IT-----Y*RS*W*G*RS*LK*Y*NP*V*V
HuC8b EAVLGGI*Y*E*Y*TL*V*MN*KE*AM*ER*GD*Y*TL*NN*V*HA*CA*KN*DF*IK*G*AI*BE*Y*Y*SL*G*V*G*Y*G*CK*IG*IL*NE*IK*DR*NR*DT*W*E*DI*V*LV*LR*G*G*AS*H*IT*TL*AY*Q*EL*P*TA*DL-----M*O*E*W*G*DA*V*O*Y*NP*AI
HuC9 SGLGLLYELI*Y*VL*DK*AS*MK*RK*G*V*EL*K*DI*KR*CL*G*Y*H*LD*Y*SL*AF*SEI*SV*GA*E*F*NK*DD*CK*V*KR*G*E*G*RA*V*NI*P*SE*NI*LD*DV*Y*LI*RG*G*TR*Y*AF*EL*KE*KL*RG*TV*ID*V*DF*V*W*AS*SI*ND*AP*VL
HuPF AVELGGRI*SA*LT*AL*RT*CE*LA*LE*GL*TD*NE*VE*DE*CL*LT-----V*EA*Q*V*NI*GI*H*G*IS*SA*E*AK*AK*E*E*E*K*K*H*H*MT*AS*F*H*Q*Y*R*ER*H*SE*V*V*G*H*H*TS*IN*DI*LL*F*IQ*AG*PE*Q-----Y*SA*W*NS*V*P*G*SP*GL
524
AmC6 ISYEITPISELV-VGIPYADIKR--RNMEKALV*EY*LYH
HuC6 IDFELAPIVDLIV-RNIPCAVTKR--NNLRKALQ*EY*AA
HuC7 IKOKLTPLYELIV-KEVPCASVKK--LYLKWALREYLD
HuC8a IDFEMQPIHEVL-RHTSLGPLEAKRQNLRRALDQYLM
HuC8b IKVVEPLVELV-TATDFAYSS*TVRQNMKQALEEFQK
HuC9 ISQKLSPIVNLIVPVKMKNAHLKK--QNLERAIEDYIN
HuPF VDYTTLEPLHLV-----

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**Fig. 3.** Amino acid sequence alignment of the MAC/perforin modules of AmphiC6, the human MAC components, and perforin. Amino acid residue numbers of the rightmost residues of each lane of AmphiC6 are shown at the *right*. Cysteine residues are shown in *boldface* and *asterisks* indicate the completely conserved residues. The *line* above the alignment indicates the human C9 region identified as a membrane-interacting region.



**Fig. 4.** Phylogenetic tree calculated from the sequences of MAC/perforin module-containing proteins and C3 family proteins. **A** MAC/perforin module-containing proteins. Only the four domains shared by all mammalian MAC components and AmphiC6 (TSP, LDLRA, MAC/P, and EGF) were used in the calculation. For perforin, which lacks the TSP and LDLRA modules, the same number of amino acid residues

was used for comparison. **B** C3 family. The complete amino acid sequences of C3, C4, and C5 of various animals were aligned using the CLUSTAL W software, and a phylogenetic tree was constructed by the neighbor joining method. Human  $\alpha$ 2-macroglobulin was used as the outgroup. Numbers indicate the proportion of bootstrap replicates supporting the given branching pattern.

establish the ancestral immunological cytolytic gene. AmphiC6 shows a closer similarity in modular structure to the vertebrate MAC components, especially C6, than to vertebrate perforin. Although further searches are required in amphioxus for proteins containing the MAC/perforin module, our results suggest that the complement-based cytotoxic mechanisms have a more ancient evolutionary origin than the lymphocyte-based cytotoxic mechanisms. Accumulating evidence indicates that adaptive immunity was established in the jawed vertebrate lineage (Kasahara et al. 1997), whereas the complement system is more ancient (Nonaka 2001). The present results give further support to this evolutionary scenario of the immune system.

Recently, C3-like thioester-containing proteins (TEP) have been reported in insects (Lagueux et al. 2000; Levashina et al. 2001), suggesting a protostome origin for the complement system. However, insect TEPs and vertebrate C3, C4, and C5 share no derived structural characteristics, indicating that they arose independently from their common ancestor,  $\alpha$ 2M (Sottrup-Jensen et al. 1985). In contrast, authentic complement genes have been identified in deuterostome invertebrates, such as sea urchins and ascidians, indicating that the complement system was established at an early stage of deuterostome evolution (Nonaka 2001). Therefore, it is highly probable that AmphiC3 and AmphiC6, and possibly additional components such as Bf and MASP, found in other

invertebrate deuterostomes, constitute a multicomponent complement system in amphioxus. Another intriguing possibility, however, is that AmphiC3 and AmphiC6 act independently. In this context, it is worth noting that there are unique sequences at the N and C termini of AmphiC6. At the N terminus, the unique RRRRRRRA-PPPPPPP sequence immediately follows a possible leader sequence. The unique C-terminal sequence contains nine consecutive repeats of the DAD/ETSPG motif. A similar heptapeptide repeat is present in the C-terminal region of the largest subunit of RNA polymerase II and is a known phosphorylation site that is important for transcriptional regulation (Uptain et al. 1997). Although the functional significance of these unique sequences in AmphiC6 is unknown, they may be involved in an activation process of AmphiC6. If this is the case, the activation mechanism for AmphiC6 is quite different from that of the terminal pathway of the vertebrate complement system, even if AmphiC6 is a complement component of amphioxus. The third possibility, that AmphiC6 has a novel function different from that of complement terminal components or perforin, has not yet been ruled out. Further studies of invertebrates are required, especially at the protein level, to clarify the original function of the MAC/perforin-module-containing proteins.

The cDNA clones for AmphiC3 and AmphiC6 were originally isolated from the notochord. However, a preliminary in situ hybridization analysis indicates that both

**Fig. 5.** Nucleotide and deduced amino acid sequences of AmphiC3. The nucleotide sequence of AmphiC3 is presented together with the deduced amino acid sequence. The initiation methionine codon was identified by comparison with vertebrate C3, and a putative leader sequence is shown in *boldface* and *underlined*. The nucleotide and amino acid numbers of the rightmost residues are shown for each lane.

The  $\beta$ - $\alpha$  processing site (amino acid residues 660–663), the thioester site (1012–1016), and the His residue (1129) associated with the thioester are shown in *boldface* and *underlined*. Also shown in *boldface* and *underlined* are six cysteine residues perfectly conserved in vertebrate C3a, C4a, and C5a (683–719) and the conserved cysteine residues in the C-terminal C3/C4/C5-specific region (1554–1790).



AmphiC3 and AmphiC6 are expressed mainly in the epipharyngeal groove, which is located just below the notochord (data not shown). Since no significant signal for AmphiC3 and AmphiC6 was detected in the notochord, the original EST clones probably originated from contamination with mRNA from the epipharyngeal groove. Although the function of the epipharyngeal groove is still not clear, the present results imply a secretory function for this organ. Again, this point is still to be tested at the protein level.

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