

Evidence for a pharmacogenetic adapted dose of oral anticoagulant in routine medical practice

Laurent Becquemont

Received: 20 March 2008 / Accepted: 8 July 2008 / Published online: 30 August 2008
© Springer-Verlag 2008

Abstract Oral anticoagulants (OA) are a leading cause of fatal haemorrhagic adverse events in relation with an important interindividual variability of response to these drugs. Besides several clinical factors, this interindividual variability of response to OA has a pharmacogenetic basis. Carriers of cytochrome P450 2C9 (*CYP2C9*)-deficient alleles have a reduced clearance of warfarin and are exposed to dramatic overdoses in the first weeks of treatment. Genetic polymorphisms of vitamin K epoxide reductase (*VKORC1*), the target of OA, identify patients with a high sensitivity to OA who are at risk of early overdose. Most pharmacogenetic evidence is presently restricted to warfarin. Several warfarin dosing algorithms have been constructed, adapted on *CYP2C9* and *VKORC1* genotypes and clinical factors, to predict the best dose for each patient. Carriers of one of allelic variant need a 20–30% reduction of warfarin dose. However, definite evidence concerning the usefulness of these algorithms in terms of reducing the frequency of major bleeding episodes is still lacking. Ongoing prospective randomised trials will ascertain definitive answer over the coming years.

Keywords Cytochrome P450 2C9 · *VKORC1* · Oral anticoagulants · Vitamin K antagonists · Pharmacogenetics · Dosing algorithm · Warfarin · Acenocoumarol · Phenprocoumon · Fluindione

Introduction

Oral anticoagulants (OA) have proven to be highly effective antithrombotic drugs for the treatment or prevention of deep venous thrombosis, pulmonary embolism, stroke and myocardial infarction for more than 50 years. Besides these well-demonstrated clinical benefits, OA are associated with a high risk of major bleeding, which unfortunately limits their use, particularly in the elderly [1]. In France, OA are associated with 17,000 hospitalisations for major bleeding annually. In the United States, more than 20 million patients are treated with OAs and 29,000 visits for bleeding complications are observed each year. Warfarin (WA) represents the first cause of lethal adverse drug reaction in the United States [2]. Several risk factors of bleeding have been identified, such as age, female gender, drug interactions, and previous bleeding [2]. Besides these classical risk factors, pharmacogenetic risk factors have been identified in the past decade and were introduced in the summarised product characteristics of WA by the US Food and Drug Administration (FDA) in 2007, indicating that lower initiation doses should be considered for patients carrying allelic variants of *CYP2C9* and vitamin K epoxide reductase (*VKORC1*) [3]. Additional genetic variants of other genes have been identified (*CYP4F2*, *GGCX*, *APOE*, *PROC* and clotting factors [4–7]), but they play a minor role in OA pharmacodynamics and are out of the scope of this review considering practical applications of pharmacogenetics tests.

L. Becquemont
Pharmacology Department, Faculté de médecine Paris Sud,
Université Paris-Sud,
Le Kremlin Bicêtre, France

L. Becquemont (✉)
Unité de recherche clinique Paris Sud,
Assistance Publique Hôpitaux de Paris, Hôpital Bicêtre,
78, rue du Général Leclerc,
94275 Le Kremlin Bicêtre Cedex, France
e-mail: laurent.becquemont@bct.ap-hop-paris.fr

Table 1 Pharmacokinetic profile of vitamin K antagonists: partly based on Ufer et al. [10]

	Mean daily dose	Excretion as unchanged drug	CYP2C9 metabolism	Influence of <i>CYP2C9</i> allelic variants	Phase 2 enzymes	Half life
Warfarin (enantiomeric)	6 mg	<5%	Intensive	<i>CYP2C9</i> *2 <i>CYP2C9</i> *3	No	S-W=30 h R-W=40 h
Phenprocoumon (enantiomeric)	2–3 mg	40%	Medium	<i>CYP2C9</i> *3	Yes	S-P=120 h R-P=120 h
Acenocoumarol (enantiomeric)	4 mg	1%	Intensive	<i>CYP2C9</i> *3	No	S-AC=2 h R-AC=7 h
Fluindione	15 mg	Below 10%	Medium	(<i>CYP2C9</i> *3 ?)	No	31 h

S-W S-warfarin, *R-W* R-warfarin, *S-P* S-phenprocoumon, *R-P* R-phenprocoumon, *S-AC* S-acenocoumarol, *R-AC* R-acenocoumarol

Different oral anticoagulant drugs

WA is available all over the world and represents the most prescribed OA in North America, Asia, and England, whereas acenocoumarol (AC), phenprocoumon, and fluindione are more commonly used in several European countries [8, 9]. WA, AC and phenprocoumon are administered as racemate, and their S-enantiomers retain the highest anticoagulant activity. There are some significant differences in the pharmacokinetic profile of the different OA [10], which can be classified as short- or long-acting drugs based on their terminal half-life (Table 1). They all act by decreasing vitamin-K-dependant clotting factors (II, VII, IX and X) via the inhibition of vitamin K epoxide reductase (VKOR), which recycles reduced vitamin K (inactive) into active oxidised vitamin K (Fig. 1). All OA have a similar tolerance profile, except for fluindione, an indanedione derivative, which is associated to more frequent immunoallergic adverse drug reaction such as hepatitis and nephritis [11].

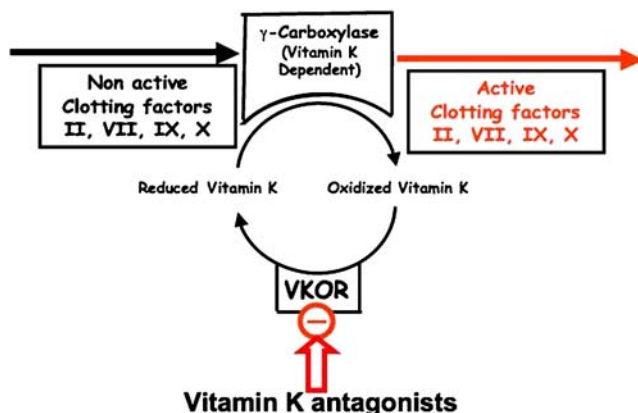


Fig. 1 Mechanism of action of vitamin-K antagonists. *VKOR* vitamin K epoxide reductase

CYP2C9 and oral anticoagulants metabolism

Cytochrome P4502C9 (*CYP2C9*) represents quantitatively the second CYP isoform expressed in the liver [12] and is involved in the metabolic clearance of different drugs such as oral hypoglycemic agents, nonsteroidal anti-inflammatory drugs, angiotensin II blockers, anticonvulsivants and antidepressant drugs [13]. Its role in the metabolism of WA was highlighted at the end of last century [14–16].

In vitro data concerning *CYP2C9*-dependant metabolism of oral anticoagulants

CYP2C9 metabolises S-warfarin, the the most active enantiomer, whereas *CYP3A4*, *CYP2C19* and *CYP1A2* are involved in R-warfarin metabolism [17–19]. Concerning acenocoumarol, *CYP2C9* seems to be the most potent isoform involved in the metabolism of both enantiomers [20, 21]. Acenocoumarol *CYP2C9*-dependant intrinsic clearance is far higher compared with WA, which explains its shorter half-life in humans. The role of *CYP2C9* in phenprocoumon metabolic clearance seems to be less pronounced compared with the two previous OA, with a similar involvement of *CYP3A4* and *CYP2C9* [10, 22]. Furthermore, phenprocoumon is also extensively conjugated, implicating phase-2 enzymes in its metabolic clearance. No data are available for the metabolism of fluindione, except that it is mainly cleared as hydroxylated and conjugated metabolites that have never been identified.

Two main functional *CYP2C9*-deficient allelic variants were identified in the early 1990s: *CYP2C9**2 (Arg₁₄₄ into Cys) and *CYP2C9**3 (Ile₃₅₉ to Leu) [23]. Both polymorphisms were rapidly shown to be associated with decreased S-warfarin hepatic clearance and, as a consequence, to need lower steady-state daily doses to achieve a usual therapeutic target [International Normalised Ratio (INR) 2–3] [14, 16, 24]. The influence of *CYP2C9* genetic

polymorphism has also been observed for AC [25] and phenprocoumon, whereas no data are available for flutidione. Compared with WA, the influence of the *CYP2C9*-deficient allele *CYP2C9*2* seems absent [26, 27] or less important for acenocoumarol [10]. Furthermore, the contribution of *CYP2C9*-dependant clearance to the overall oral clearance of phenprocoumon seems to be lower than that observed with WA [10, 28]. The importance of *CYP2C9* genetic polymorphism is probably higher among patients of Caucasian origin, in whom the frequencies of the *CYP2C9*-deficient alleles is almost the double compared with other ethnic groups. Indeed, the allelic frequencies of *CYP2C9*2* and *CYP2C9*3* are 0.1 and 0.08, 0.04 and 0.02 and 0.01 and 0.04 among subjects of Caucasian, African and Asiatic origins [29–31]. The *CYP2C9*-dependant pharmacogenetic effect observed for OA has three major clinical consequences, which are described below.

CYP2C9 and daily dose requirements

In 1999, Aithal et al. described for the first time that patients carrying the *CYP2C9*2* and/or *CYP2C9*3* allele needed much lower doses of WA at steady state [24] compared with noncarriers. These results were further confirmed by many different independent groups [32, 33]. A similar relationship between *CYP2C9* genetic polymorphism and AC daily dose requirement was observed [26, 34, 35]. Concerning phenprocoumon, slightly lower dose requirements were described for carriers of *CYP2C9*-deficient alleles [36, 37], but this was not observed by all authors [38], probably in relation with the lower impact of *CYP2C9*-dependant metabolism of phenprocoumon compared with WA and acenocoumarol.

CYP2C9 and time to reach stable anticoagulation

Aithal et al. were the first to detect that patients treated with WA and carrying one *CYP2C9*-deficient allele had more difficulties reaching stable anticoagulation [24]. This observation was also confirmed by other groups [33] and with the other related OA, AC [34, 39] and phenprocoumon [36].

CYP2C9 and risk of bleeding

As a consequence of this difficulty finding the right dose and to achieve stable anticoagulation, it has been shown that carriers of *CYP2C9* allelic variants have a 3.68 increased risk of major bleeding with WA [24]. The *CYP2C9*-dependant pharmacogenetic risk of bleeding is maximal during the initiation phase of the anticoagulant treatment. Increased risk of major haemorrhage with WA conferred by *CYP2C9* allelic variants has been estimated to

be 5.3 before stabilisation of therapy and 2.2 after stabilisation [32, 33]. A similar increased number of overanticoagulation episodes (INR>6) and bleeding events has been described for carriers of *CYP2C9* variants treated with AC [39, 40]. More conflicting data exist concerning the *CYP2C9*-dependant risk of bleeding with phenprocoumon [36, 40, 41], which is probably smaller than that observed with WA and AC due to a smaller involvement of *CYP2C9* in phenprocoumon oral clearance [28].

VKORC1, the target of oral anticoagulants

The gene that encodes VKOR (*VKORC1*), the target enzyme for OA (Fig. 1), was identified in 2004 [42, 43]. Rost et al. identified the *VKORC1* gene from four independent families whose members were considered resistant to WA. One patient needed more than 17 mg/day of WA to achieve adequate anticoagulation, two others about 40 mg/d and the fourth did not respond to any dose of WA. They carried rare mutations of *VKORC1* of amino acids 29 (Val/Leu), 45 (Val/Ala), 58 (Arg/Gly) and 128 (Leu/Arg) [42]. Several months later, other groups described that common *VKORC1* single nucleotide polymorphisms (SNPs) were strongly associated with OA sensitivity [44–48]. *VKORC1* genetic polymorphism has the same influence for all OA [37, 45, 49].

Several SNPs in complete linkage disequilibrium identify a haplotype combination called A [32, 48], which is associated with OA hypersensitivity. Two of these SNPs (–1639G>A, rs9934438 or 1173C>T, rs17878363) are usually used to identify (tag) this *VKORC1* A haplotype group, which is associated with lower levels of hepatic VKORC expression [48]. Therefore, patients carrying the *VKORC1* A haplotype have lower levels of hepatic VKOR and as a consequence are more sensible to OA. The allelic frequency of this *VKORC1* A haplotype is rather low among subjects from African origin (0.15), high in Caucasians (0.42) and extremely frequent among Asians [32, 45, 47, 50]. As observed for *CYP2C9*, *VKORC1* genetic polymorphism has three major clinical consequences on the pharmacodynamics of OA, which are described below.

VKORC1 and daily dose requirements

Since *VKORC1* genetic polymorphism modulates WA sensitivity, it also influences the dose of WA at steady state to reach stable anticoagulation [48, 51]. Swartz et al. observed a clear gene–dose effect with *VKORC1* A associated with a higher INR response, requiring lower doses of WA [51]. From a large cohort of patients treated with WA, it has been estimated that the dose should be reduced by 28% for each *VKORC1* A allele [52]. Similar

Table 2 Consequences of *CYP2C9* and *VKORC1* polymorphisms on OAs pharmacokinetics and pharmacodynamics

	<i>CYP2C9</i>				<i>VKORC1</i>			
	PK	Daily dose	Time to stable INR	Overdose and bleeding risk	PK	Daily dose	Time to stable INR	Overdose and bleeding risk
Warfarin	Yes (for S-warfarin) [64, 65]	Yes [14, 24, 33, 51, 57, 64, 66]	Yes [33, 59, 60, 67]	Yes [24, 33]	No	Yes [44, 51, 68]	Yes [32, 69, 70]	Yes [32]
Phenprocoumon	Yes (Minor) [10, 28]	Yes (minor) [36, 37, 40, 41]	Yes (minor) [36, 41, 37]	Yes (minor) [36, 37, 40]	No	Yes [37, 53]	No data	Yes [37, 53]
Acenocoumarol	Yes [25, 27, 71]	Yes [26, 34, 35, 39, 54]	Yes [34, 39]	Yes [38–40]	No	Yes [53, 54, 72]	Yes [39]	Yes [54, 55]
Fluindione	No data	No data	No data	Inconclusive [73]	No	No data	No data	Yes [55]

INR International Normalised Ratio

findings have been obtained for other OA, such as AC [53, 54] and phenprocoumon [37, 53]. *VKORC1* genetic polymorphism also mainly explains why steady-state doses of OA differ between patients of different ethnic origins: *VKORC1* A (linked to OA sensitivity) is rare among blacks, who need higher doses of OA; the same A haplotype is present in 90% of patients of Asian origin, who need the smallest doses to achieve stable anticoagulation.

VKORC1 and time to reach stable anticoagulation

WA-treated patients homozygous for the *VKORC1* A/A haplotype (sensitive to OA) achieve their first INR in the target range 2.4 sooner compared with others and reach an INR above 4, 2.5 sooner [51]. They also spend twice as much time above the INR upper limit of 4 compared with other patients [51] and are thus more difficult to equilibrate.

VKORC1 and risk of bleeding

As a consequence of this difficulty to obtain of stable INR and a stable dose, *VKORC1* genetic polymorphism has been rapidly recognised as an important risk for bleeding in

patients treated with OA. This higher frequency of WA overdose among *VKORC1* A/A patients was mainly observed during the first month following the introduction of the OA [51]. A similar increased risk of bleeding linked to *VKORC1* genetic polymorphism has been observed with phenprocoumon [37, 53] and AC [54, 55].

Pharmacogenetic-based dosing algorithms

Different algorithms have been developed to determine the right dose for the right patient [56]. The first algorithms, which were employed for a long time, were based on clinical variables such as INR target, patient age, history of previous bleeding, reason for prescription (atrial fibrillation or venous thrombosis), associated diseases and concomitant drugs prescribed. As it has been estimated that *CYP2C9* and *VKORC1* genetic polymorphisms explain 35–50% of the interindividual variability of OA response during the initiation phase of the treatment, dosing algorithms rapidly integrated genotype information. One of the first of these pharmacogenetic-based dosing normograms including *VKORC1* and *CYP2C9* genotypes was proposed by Sconce

Table 3 Predicted steady-state warfarin daily dose requirements based on pharmacogenetic-based algorithms

	<i>VKORC1</i> non-A/non-A	<i>VKORC1</i> non-A/ A	<i>VKORC1</i> A/ A	<i>VKORC1</i> non-A/non-A	<i>VKORC1</i> non-A/non A
	<i>CYP2C9</i> *1/*1	<i>CYP2C9</i> *1/*1	<i>CYP2C9</i> *1/*1	<i>CYP2C9</i> *1/*3	<i>CYP2C9</i> *3/*3
Sconce et al. [57]	5 mg/d	4 mg/d	3 mg/d	3 mg/d	2 mg/d
Gage et al. [52]	6 mg/d	4 mg/d	3 mg/d	4 mg/d	3 mg/d

Additional information introduced in the algorithm of Sconce et al.: age 70 years, height 1.7 m. Additional information introduced in the algorithm of Gage et al.: age 70 years, height 1.7 meters, weight 75 kg, target INR 2.5, white origin, nonstatin and no concomitant amiodarone, nonsmoker, male gender

VKORC1 vitamin K epoxide reductase

et al. [57], which allowed in a replication cohort to explain about 80% of the interindividual variability of the WA dose needed to achieve an INR between 2 and 3. Different other pharmacogenetic-based normograms have been developed [58] for Caucasian and non-Caucasian ethnic groups [59]. At the time, one of the best validated dosing algorithm was proposed by Gage et al. [52] and includes not only *CYP2C9* and *VKORC1* genetic data but also numerous clinically relevant items such as associated drugs, smoking status and ethnic origin. It explains 53–54% of the variability in the WA dose and was prospectively validated in a replication cohort. It was obtained from one of the largest collaborative cohorts (1,015 patients) and was made available online at www.warfarindosing.org. Tables 2 and 3 compare the dose of WA that should be introduced in a 70-year-old white nonsmoking man without concomitant drugs (statins or amiodarone) of 75 kg and 1.70 m tall based on two different dosing normograms. Both algorithms predict relatively similar doses. However, Gage et al.'s algorithm allows the dose to be adapted to much more relevant clinical factors. Pharmacogenetic-based algorithms for OA other than WA are missing at this time.

Pharmacogenetic prospective randomised clinical trials

Unfortunately, only three small-scale prospective randomised trials were performed to determine whether an initial pharmacogenetic-adapted dose could improve the time to reach stable anticoagulation and the frequency of bleeding episodes during initiation of OA therapy [60–62]. The first study, which included only 38 patients, was just a pilot trial testing the feasibility of *CYP2C9* genotyping before introducing WA; no differences could be observed between the *CYP2C9*-adapted and nonadapted patients [62]. The second study was aimed to decrease by 50% the time spent out of the INR range during the initiation of WA treatment [60]. The authors used a WA loading dose during the first 2 days of treatment and randomised the patients in two arms: one according to a standard clinical titration (99) and the other based on a *CYP2C9*- and *VKORC1*-adapted algorithm (101). The authors could not confirm their primary hypothesis in relation to a lower frequency of the clinical event compared with what was expected in the control group and a too ambitious planned difference between both groups (underpowered study for the main outcome). However, they could detect that patients in the pharmacogenetic-guided arm needed less INR controls and had less dose modifications. No differences could be observed concerning the bleeding events. The last trial was planned to reduce the time to reach the first therapeutic INR range and the time to reach stable anticoagulation [61]. Patients were randomised in

two arms: one with a standardised dose of WA (142), and the other (141) with a pharmacogenetic-adapted dose of WA, which included only *CYP2C9* genotype in its normogram. Both study endpoints were reached sooner (2.23 days and 18.1 days, respectively) in the pharmacogenetic-adapted arm. Furthermore, in the pharmacogenetic-adapted arm, patients spent more time within the INR therapeutic range (80.4% vs 63.4%) and experienced less minor bleeding (3.2% vs 12.5%).

One might be surprised by the low number of randomised trials in this field despite strong scientific evidence indicating that *CYP2C9* and *VKORC1* explain about 50% of the interindividual variability of WA response. Unfortunately, this is a common problem in the field of pharmacogenetics, which lacks randomised trials. One reason is the recent discovery in 2005 of the influence of *VKORC1* polymorphisms on OA response. Several randomised trials, some including a larger number of patients, are ongoing and are available at www.clinicaltrials.gov. The results of these trials will be available in the coming years and will hopefully definitively demonstrate the usefulness of a pharmacogenetically adapted dose of WA. The next explanation for so few randomised trials in the field is that the ultimate objective of such a study is to reduce the frequency of major bleeding episodes (the most clinically relevant endpoint), which occur in clinical trials in 1% of the study population per year [63]. Such an event frequency implicates inclusion of at least 1,000 patients. Another difficulty in performing such randomised trials is the need for rapid pharmacogenetic tests, the results of which must be sent to the physician within a few hours. Such rapid genotyping is for the moment restricted to several university hospitals, but this situation should change rapidly with the development of validated diagnostic kits, some of which have been approved by the FDA.

Future prospective randomised trials will also bring extremely important information concerning the frequency of recurrent thrombotic events. All pharmacogenetic efforts have focused on reducing OA overdose and the risk of bleeding. But difficulties attaining stable anticoagulation also includes INR episodes below 2 and possibly increased frequency of thrombotic recurrence. Furthermore, a pharmacogenetically adapted dose might limit the bleeding episodes but at the same time increase the early recurrence of thrombotic events, which might totally blunt the benefit of the pharmacogenetically adapted dosing. Therefore, attention must be paid to precisely evaluate the real risk/benefit ratio of a pharmacogenetically adapted dosing.

No randomised trials have been performed with OA other than WA. A prospective randomised trial is planned in Europe to compare the utility of a pharmacogenetic-based prescribing between WA, AC and phenprocoumon.

Conclusion

A large number of studies have demonstrated that the genetic polymorphisms of *CYP2C9* and *VKORC1* significantly influence OA pharmacokinetics and pharmacodynamics. However, most of the evidence is restricted to WA, and even for this drug, a definite demonstration of the utility of systematic genotyping to adapt its dose is still lacking. The benefit of such pharmacogenetic testing remains to be validated in terms of costs with a pharmacoeconomic approach.

The definitive answer concerning the usefulness of the pharmacogenetically based WA dosing will be available in the coming years. This process will probably take some time for the OA used in Europe. Meanwhile, physicians can benefit from the www.warfarindosing.org normogram, which has the advantage of including the first INR values and a delayed genotype.

References

- Shireman T, Mahnken J, Howard P, Kresowik T, Hou Q, Ellerbeck E (2006) Development of a contemporary bleeding risk model for elderly warfarin recipients. *Chest* 130:1390–1396
- Wysowski D, Nourjah P, Swartz L (2007) Bleeding complications with warfarin use. *Arch Intern Med* 167:1414–1419
- FDA. New Labeling Information for Warfarin (marketed as Coumadin). <http://www.fda.gov/cder/drug/infopage/warfarin/default.htm> 2007
- Caldwell M, Awad T, Johnson J, Gage B, Falkowski M, Gardina P, Hubbard J, Turpaz Y, Langaee T, Eby C, King C, Brower A, Schmelzer J, Glurich I, Vidaillet H, Yale S, Qi Zhang K, Berg R, Burmester J (2008) CYP4F2 genetic variant alters required warfarin dose. *Blood* 111:4106–4112
- Cooper G, Johnson J, Langaee T, Feng H, Stanaway I, Schwarz U, Ritchie M, Stein C, Roden D, Smith J, Veenstra D, Rettie A, Rieder M (2008) A genome-wide scan for common genetic variants with a large influence on warfarin maintenance dose. *Blood Jun 5* [Epub ahead of print]
- Wadelius M, Chen L, Eriksson N, Bumpstead S, Ghori J, Wadelius C, Bentley D, McGinnis R, Deloukas P (2007) Association of warfarin dose with genes involved in its action and metabolism. *Hum Genet* 121:23–24
- Wadelius M, Chen L, Lindh J, Eriksson N, Ghori M, Bumpstead S, Holm L, McGinnis R, Rane A, Deloukas P (2008) The largest prospective warfarin-treated cohort supports genetic forecasting. *Blood Jun 23* [Epub ahead of print]
- Ansell J, Hollowell J, Pengo V, Martinez-Brotons F, Caro J, Drouet L (2007) Descriptive analysis of the process and quality of oral anticoagulation management in real-life practice in patients with chronic non-valvular atrial fibrillation: the international study of anticoagulation management (ISAM). *J Thromb Thrombolysis* 23:83–91
- Pengo V, Pegoraro C, Cucchini U, Iliceto S (2006) Worldwide management of oral anticoagulant therapy: the ISAM study. *J Thromb Thrombolysis* 21:73–77
- Ufer M (2005) Comparative pharmacokinetics of vitamin K antagonists: warfarin, phenprocoumon and acenocoumarol. *Clin Pharmacokinet* 44:1227–1246
- Sparsa A, Bédane C, Benazahary H, De Vencay P, Gauthier M, Le Brun V, Boulinguez S, Loustaud-Ratti V, Soria P, Vidal E, Bonnetblanc J (2001) Drug hypersensitive syndrome caused by fluidione. *Ann Dermatol Venereol* 128:1014–1018
- Shimada T, Yamazaki H, Mimura M, Inui Y, Guengerich FP (1994) Interindividual variations in human liver cytochrome P-450 enzymes involved in the oxydation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. *J Pharmacol Exp Ther* 270:414–423
- Flockhart D. Cytochrome P450 drug interaction table, <http://medicine.iupui.edu/flockhart/table.htm>
- Furuya H, Fernandez-Salguero P, Gregory W, Taber H, Steward A, Gonzalez FJ, Idle JR (1995) Genetic polymorphism of CYP2C9 and its effect on warfarin maintenance dose requirement in patients undergoing anticoagulation therapy. *Pharmacogenetics* 5:389–392
- Rettie AE, Wienkers LC, Gonzalez FJ, Trager WF, Korzekwa KR (1994) Impaired (S)-warfarin metabolism catalysed by the R144C allelic variant of CYP2C9. *Pharmacogenetics* 4:39–42
- Steward DJ, Haining RL, Henne KR, Davis G, Rushmore TH, Trager WF, Rettie AE (1997) Genetic association between sensitivity to warfarin and expression of CYP2C9*3. *Pharmacogenetics* 7:361–367
- Rettie A, Korzekwa K, Kunze K, Lawrence R, Eddy A, Aoyama T, Gelboin H, Gonzalez F, Trager W (1992) Hydroxylation of warfarin by human cDNA-expressed cytochrome P-450: a role for P-4502C9 in the etiology of (S)-warfarin-drug interactions. *Chem Res Toxicol* 5:54–59
- Zhang Z, Fasco M, Huang Z, Guengerich F, Kaminsky L (1995) Human cytochromes P4501A1 and P4501A2: R-warfarin metabolism as a probe. *Drug Metab Dispos* 23:1339–1346
- Zhou Q, Zhou S, Chan E (2005) Effect of omeprazole on the hydroxylation of warfarin enantiomers in human: in-vitro studies with liver microsomes and cDNA-expressed cytochrome P450 isozymes. *Curr Drug Metab* 6:399–411
- Hermans JJ, Thijssen HH (1993) Human liver microsomal metabolism of the enantiomers of warfarin and acenocoumarol: P450 isozyme diversity determines the differences in their pharmacokinetics. *Br J Pharmacol* 110:482–490
- Thijssen HH, Flinois JP, Beaune PH (2000) Cytochrome P4502C9 is the principal catalyst of racemic acenocoumarol hydroxylation reactions in human liver microsomes. *Drug Metab Dispos* 28:1284–1290
- Ufer M, Svensson J, Krausz K, Gelboin H, Rane A, Tybring G (2004) Identification of cytochromes P450 2C9 and 3A4 as the major catalysts of phenprocoumon hydroxylation in vitro. *Eur J Clin Pharmacol* 60:173–182
- Ingelman-Sundberg M, Daly A, Nebert D. Home Page of the Human Cytochrome P450 (CYP) Allele Nomenclature Committee <http://www.imm.ki.se/CYPalleles/>
- Aithal GP, Day CP, Kesteven PJ, Daly AK (1999) Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. *Lancet* 353:717–719
- Thijssen HH, Ritzen B (2003) Acenocoumarol pharmacokinetics in relation to cytochrome P450 2C9 genotype. *Clin Pharmacol Ther* 74:61–68
- Hermida J, Zarza J, Alberca I, Montes R, Lopez ML, Molina E, Rocha E (2002) Differential effects of 2C9*3 and 2C9*2 variants of cytochrome P-450 CYP2C9 on sensitivity to acenocoumarol. *Blood* 99:4237–4239
- Morin S, Bodin L, Lorient MA, Thijssen HH, Robert A, Strabach S, Verstuyft C, Tregouet DA, Dubert L, Laurent-Puig P, Funck-Brentano C, Jaillon P, Beaune PH, Becquemont L (2004) Pharmacogenetics of acenocoumarol pharmacodynamics. *Clin Pharmacol Ther* 75:403–414

28. Kirchheiner J, Ufer M, Walter EC, Kammerer B, Kahlich R, Meisel C, Schwab M, Gleiter CH, Rane A, Roots I, Brockmoller J (2004) Effects of CYP2C9 polymorphisms on the pharmacokinetics of R- and S-phenprocoumon in healthy volunteers. *Pharmacogenetics* 14:19–26
29. London SJ, Daly AK, Leathart JB, Navidi WC, Idle JR (1996) Lung cancer risk in relation to the CYP2C9*1/CYP2C9*2 genetic polymorphism among African-Americans and Caucasians in Los Angeles County, California. *Pharmacogenetics* 6:527–533
30. Scordo M, Aklillu E, Yasar U, Dahl M, Spina E, Ingelman-Sundberg M (2001) genetic polymorphism of cytochrome P450 2C9 in a Caucasian and black African population. *Br J Clin Pharmacol* 52:447–450
31. Yang JQ, Morin S, Verstuyft C, Fan LA, Zhang Y, Xu CD, Barbu V, Jaillon P, Becquemont L (2003) Frequency of cytochrome P450 2C9 allelic variants in the Chinese and French populations. *Fundam Clin Pharmacol* 17:373–376
32. Limdi N, McGwin G, Goldstein J, Beasley T, Arnett D, Adler B, Baird M, Acton R (2008) Influence of CYP2C9 and VKORC1 1173C/T Genotype on the Risk of Hemorrhagic Complications in African-American and European-American Patients on Warfarin. *Clin Pharmacol Ther* 83:312–321
33. Higashi MK, Veenstra DL, Kondo LM, Wittkowsky AK, Srinouanprachanh SL, Farin FM, Rettie AE (2002) Association between CYP2C9 genetic variants and anticoagulation-related outcomes during warfarin therapy. *JAMA* 287:1690–1698
34. Tassies D, Freire C, Pijoan J, Maragall S, Monteagudo J, Ordinas A, Reverter JC (2002) Pharmacogenetics of acenocoumarol: cytochrome P450 CYP2C9 polymorphisms influence dose requirements and stability of anticoagulation. *Haematologica* 87:1185–1191
35. Thijssen H, Verkooyen I, Frank H (2000) The possession of the CYP2C9*3 allele is associated with low dose requirement of acenocoumarol. *Pharmacogenetics* 10:757–760
36. Schalekamp T, Oosterhof M, van Meegen E, van Der Meer FJ, Conemans J, Hermans M, Meijerman I, de Boer A (2004) Effects of cytochrome P450 2C9 polymorphisms on phenprocoumon anticoagulation status. *Clin Pharmacol Ther* 76:409–417
37. Schalekamp T, Brasse BP, Roijers JF, van Meegen E, van der Meer FJ, van Wijk EM, Egberts AC, de Boer A (2007) VKORC1 and CYP2C9 genotypes and phenprocoumon anticoagulation status: interaction between both genotypes affects dose requirement. *Clin Pharmacol Ther* 81:185–193
38. Visser L, van Vliet M, van Schaik R, Kasbergen A, De Smet P, Vulto A, Hofman A, van Duijn C, Stricker B (2004) The risk of overanticoagulation in patients with cytochrome P450 CYP2C9*2 or CYP2C9*3 alleles on acenocoumarol or phenprocoumon. *Pharmacogenetics* 14:27–33
39. Schalekamp T, van Geest-Daalderop JH, de Vries-Goldschmeding H, Conemans J, Bernsen Mj M, de Boer A (2004) Acenocoumarol stabilization is delayed in CYP2C9*3 carriers. *Clin Pharmacol Ther* 75:394–402
40. Visser L, van Schaik R, van Vliet M, Trienekens P, De Smet P, Vulto A, Hofman A, van Duijn C, Stricker B (2004) The risk of bleeding complications in patients with cytochrome P450 CYP2C9*2 or CYP2C9*3 alleles on acenocoumarol or phenprocoumon. *Thromb Haemost* 92:61–66
41. Hummers-Pradier E, Hess S, Adham I, Papke T, Pieske B, Kochen M (2003) Determination of bleeding risk using genetic markers in patients taking phenprocoumon. *Eur J Clin Pharmacol* 59:213–219
42. Rost S, Fregin A, Ivaskevicius V, Conzelmann E, Hortnagel K, Pelz HJ, Lappégard K, Seifried E, Scharrer I, Tuddenham EG, Muller CR, Strom TM, Oldenburg J (2004) Mutations in VKORC1 cause warfarin resistance and multiple coagulation factor deficiency type 2. *Nature* 427:537–541
43. Li T, Chang C-Y, Jin D-Y, Lin P-J, Khvorova A, Stafford DW (2004) Identification of the gene for vitamin K epoxide reductase. *Nature* 427:541–543
44. D’Andrea G, D’Ambrosio RL, Di Perna P, Chetta M, Santacroce R, Brancaccio V, Grandone E, Margaglione M (2005) A polymorphism in the VKORC1 gene is associated with an interindividual variability in the dose-anticoagulant effect of warfarin. *Blood* 105:645–649
45. Bodin L, Verstuyft C, Tregouet DA, Robert A, Dubert L, Funck-Brentano C, Jaillon P, Beaune P, Laurent-Puig P, Becquemont L, Lorient MA (2005) Cytochrome P450 2C9 (CYP2C9) and vitamin K epoxide reductase (VKORC1) genotypes as determinants of acenocoumarol sensitivity. *Blood* 106:135–140
46. Wadelius M, Chen LY, Downes K, Ghori J, Hunt S, Eriksson N, Wallerman O, Melhus H, Wadelius C, Bentley D, Deloukas P (2005) Common VKORC1 and GGX polymorphisms associated with warfarin dose. *Pharmacogenomics J* 5:262–270
47. Yuan HY, Chen JJ, Lee MT, Wung JC, Chen YF, Chang MJ, Lu MJ, Hung CR, Wei CY, Chen CH, Wu JY, Chen YT (2005) A novel functional VKORC1 promoter polymorphism is associated with inter-individual and inter-ethnic differences in warfarin sensitivity. *Hum Mol Genet* 14:1745–1751
48. Rieder MJ, Reiner AP, Gage BF, Nickerson DA, Eby CS, McLeod HL, Blough DK, Thummel KE, Veenstra DL, Rettie AE (2005) Effect of VKORC1 haplotypes on transcriptional regulation and warfarin dose. *N Engl J Med* 352:2285–2293
49. Becquemont L (2007) Pharmacogenetics of Vitamin K antagonists. Joint Cold Spring Harbor Laboratory/Wellcome Trust conference on Pharmacogenomics, Hinxton :4
50. Larramendy-Gozalo C, Yang JQ, Verstuyft C, Bodin L, Dubert L, Zhang Y, Xu C, Fan L, Jaillon P, Becquemont L (2006) Genetic Polymorphism of Vitamin K Epoxide Reductase (VKORC1) 1173C>T in a Chinese and a Caucasian Population. *Basic Clin Pharmacol Toxicol* 98:611–613
51. Schwartz U, Ritchie M, Bradford Y, Li C, Dudek S, Frye-Anderson A, Kim R, Roden D, Stein C (2008) Genetic determinants of response to warfarin during initial anticoagulation. *N Engl J Med* 358:999–1008
52. Gage B, Eby C, Johnson J, Deych E, Rieder M, Ridker P, Milligan P, Grice G, Lenzini P, Rettie A, Aquilante C, Grosso L, Marsh S, Langae T, Farnett E, Voora D, Veenstra D, Glynn R, Barrett A, McLeod H (2008) Use of Pharmacogenetic and Clinical Factors to Predict the Therapeutic Dose of warfarin. *Clin Pharmacol Ther* Feb 27 [Epub ahead of print]
53. Reitsma P, van der Heijden J, Groot A, Rosendaal F, Büller H (2005) A C1173T Dimorphism in the VKORC1 Gene Determines Coumarin Sensitivity and Bleeding Risk. *PLOS Medicine* 2:996–998
54. Schalekamp T, Brasse BP, Roijers JF, Chahid Y, van Geest-Daalderop JH, de Vries-Goldschmeding H, van Wijk EM, Egberts AC, de Boer A (2006) VKORC1 and CYP2C9 genotypes and acenocoumarol anticoagulation status: interaction between both genotypes affects overanticoagulation. *Clin Pharmacol Ther* 80:13–22
55. Quteineh L, Verstuyft C, Descot C, Dubert L, Robert A, Jaillon P, Becquemont L (2005) Vitamin K epoxide reductase (VKORC1) genetic polymorphism is associated to oral anticoagulant overdose. *Thromb Haemost* 94:690–691
56. Crowther M (2003) The impact of CYP2C9 and VKORC1 genetic polymorphism and patient characteristics upon warfarin dose. *Semin Vasc Med* 3:255–260
57. Sconce E, Khan T, Wynne H, Avery P, Monkhouse L, King B, Wood P, Kesteven P, Daly A, Kamali F (2005) The impact of CYP2C9 and VKORC1 genetic polymorphism and patient characteristics upon warfarin dose requirements: proposal for a new dosing regimen. *Blood* 106:2329–2333

58. Wu A (2007) Use of genetic and nongenetic factors in warfarin dosing algorithms. *Pharmacogenomics* 8:851–861
59. Wen MS, Lee M, Chen JJ, Chuang HP, Lu LS, Chen CH, Lee TH, Kuo CT, Sun FM, Chang YJ, Kuan PL, Chen YF, Charng MJ, Ray CY, Wu JY, Chen YT (2008) Prospective Study of Warfarin Dosage Requirements Based on CYP2C9 and VKORC1 Genotypes. *Clin Pharmacol Ther*
60. Anderson JL, Home BD, Stevens SM, Grove AS, Barton S, Nicholas ZP, Kahn SF, May HT, Samuelson KM, Muhlestein JB, Carlquist JF (2007) Randomized trial of genotype-guided versus standard warfarin dosing in patients initiating oral anticoagulation. *Circulation* 116:2563–2570
61. Caraco Y, Blotnick S, Muszkat M (2008) CYP2C9 genotype-guided warfarin prescribing enhances the efficacy and safety of anticoagulation: a prospective randomized controlled study. *Clin Pharmacol Ther* 83:460–470
62. Hillman MA, Wilke RA, Yale SH, Vidaillet HJ, Caldwell MD, Glurich I, Berg RL, Schmelzer J, Burmester JK (2005) A prospective, randomized pilot trial of model-based warfarin dose initiation using CYP2C9 genotype and clinical data. *Clin Med Res* 3:137–145
63. Kearon C, Ginsberg J, Kovacs M, Anderson D, Wells P, Julian J, MacKinnon B, Weitz J, Crowther M, Dolan S, Turpie A, Geerts W, Solymoss S, van Nguyen P, Demers C, Kahn S, Kassis J, Rodger M, Hambleton J, Gent M (2003) Comparison of low-intensity warfarin therapy with conventional-intensity warfarin therapy for long-term prevention of recurrent venous thromboembolism. *N Eng J Med* 349:675–683
64. Scordo MG, Pengo V, Spina E, Dahl ML, Gusella M, Padriani R (2002) Influence of CYP2C9 and CYP2C19 genetic polymorphisms on warfarin maintenance dose and metabolic clearance. *Clin Pharmacol Ther* 72:702–710
65. Takahashi H, Wilkinson GR, Caraco Y, Muszkat M, Kim RB, Kashima T, Kimura S, Echizen H (2003) Population differences in S-warfarin metabolism between CYP2C9 genotype-matched Caucasian and Japanese patients. *Clin Pharmacol Ther* 73:253–263
66. Taube J, Halsall D, Baglin T (2000) Influence of cytochrome P-450 CYP2C9 polymorphisms on warfarin sensitivity and risk of over-anticoagulation in patients on long-term treatment [In Process Citation]. *Blood* 96:1816–1819
67. Kamali F, Khan T, King B, Frearson R, Kesteven P, Wood P, Daly A, Wynne H (2004) Contribution of age, body size, and CYP2C9 genotype to anticoagulant response to warfarin. *Clin Pharmacol Ther* 75:204–212
68. Obayashi K, Nakamura K, Kawana J, Ogata H, Hanada K, Kurabayashi M, Hasegawa A, Yamamoto K, Horiuchi R (2006) VKORC1 gene variations are the major contributors of variation in warfarin dose in Japanese patients. *Clin Pharmacol Ther* 80:169–178
69. Millican EA, Lenzini PA, Milligan PE, Grosso L, Eby C, Deych E, Grice G, Clohisy JC, Barrack RL, Burnett RS, Voora D, Gatchel S, Tiemeier A, Gage BF (2007) Genetic-based dosing in orthopedic patients beginning warfarin therapy. *Blood* 110:1511–1515
70. Carlquist JF, Horne BD, Muhlestein JB, Lappe DL, Whiting BM, Kolek MJ, Clarke JL, James BC, Anderson JL (2006) Genotypes of the cytochrome p450 isoform, CYP2C9, and the vitamin K epoxide reductase complex subunit 1 conjointly determine stable warfarin dose: a prospective study. *J Thromb Thrombolysis* 22:191–197
71. Thijssen HH, Drittij MJ, Vervoort LM, de Vries-Hanje JC (2001) Altered pharmacokinetics of R- and S-acenocoumarol in a subject heterozygous for CYP2C9*3. *Clin Pharmacol Ther* 70:292–298
72. Montes R, Ruiz de Gaona E, Martínez-González M, Alberca I, Hermida J (2006) The c.-1639G > A polymorphism of the VKORC1 gene is a major determinant of the response to acenocoumarol in anticoagulated patients. *Br J Haematol* 133:183–187
73. Verstuyft C, Robert A, Morin S, Lorient MA, Flahault A, Beaune P, Funck-Brentano C, Jaillon P, Becquemont L (2003) Genetic and environmental risk factors for oral anticoagulant overdose. *Eur J Clin Pharmacol* 58:739–745