INVASIVE SPECIES - ORIGINAL PAPER



# **Understanding the invasion success of** *Sargassum muticum***: herbivore preferences for native and invasive** *Sargassum* **spp**

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**Abstract** Several ecological models explain the success of introduced seaweeds by herbivore–prey interactions. The 'enemy release hypothesis' states that invaders benefit from a lack of natural enemies compared to the recipient community. The 'novel weapons hypothesis,' however, highlights the importance of chemical defense compounds of introduced species that are more effective in invaded regions than native counterparts. In order to explain the tremendous invasion success of the brown alga *Sargassum muticum*, we compared the palatability and nutritional value of *S. muti‑ cum* individuals from their native and invaded habitat (Japan and North Sea, respectively) with noninvasive congeneric species from Japan (*S. fusiforme, S. horneri*), and a native competitor from the North Sea (*Fucus vesiculosus*). Different feeding assays using artificial food with either freezedried algae or algal extracts and three dominant North Sea mesograzers were performed to detect feeding preferences. All herbivores preferred the local brown alga *F. vesiculosus*, followed by North Sea *S. muticum*, while *Sargassum* spp.



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from Japan were the least preferred. Since nutritional value did not correlate with feeding preference and algal extracts had the same effect as algal powder, we could demonstrate a deterrent activity of algal secondary metabolites. The preference of herbivores for the sympatric *S. muticum* population compared to the allopatric Japanese population could indicate a resource allocation from chemical defense to reproduction and growth. Due to the low palatability of *Sar‑ gassum* spp. from Japan, it might be reasonable to include additional *Sargassum* species in North Sea monitoring to prevent their establishment in European waters.

# **Introduction**

Introduction of non-indigenous marine species (NIMS) by anthropogenic influences is one of the key pressures that threaten marine biodiversity and resources of the world's oceans (Vitousek et al. [1997;](#page-12-0) Millennium Ecosystem Assessment [2005](#page-11-0); Halpern et al. [2008;](#page-10-0) Carlton [2009](#page-10-1); Davidson et al. [2015\)](#page-10-2). From all detected NIMS of all major animal, plant and algal phyla, macroalgae not only constitute a large component of the globally introduced biota, but also cause significant economic and environmental damage over which we have only limited post-invasion control and management options (Ribera and Boudouresque [1995](#page-11-1); Thresher [1999](#page-11-2); Schaffelke et al. [2006](#page-11-3); Anderson [2007](#page-9-0); Schaffelke and Hewitt [2007\)](#page-11-4).

To enable a targeted monitoring and adaptive invasion management, the search for specific invasive traits is of increasing interest. Although invasion success has usually been correlated with numerous species traits such as reproduction strategy, growth rate, and dispersal potential (Rejmánek and Richardson [1996](#page-11-5); Nyberg and Wallentinus [2005\)](#page-11-6), several studies contradict this assumption (Smith and Knapp [2001;](#page-11-7) Daehler [2003](#page-10-3); Seabloom et al. [2003](#page-11-8)) and rather highlight the link between the success of marine bioinvasion and the presence of chemical defenses by introduced organisms (Wikström et al. [2006;](#page-12-1) Cassano et al. [2008\)](#page-10-4). In order to verify the importance of chemical defenses for the invasion success, more investigations are needed.

Since predator–prey interactions deeply shape marine ecosystems, the effectivity of antipredator metabolites might help to explain the success of widespread NIMS. Several theories have been proposed to explain the linkage between invasion success and enemy defense. The 'enemy release hypothesis' (ERH) states that introduced species in the new range are less affected by local enemies, as these fail to recognize them as adequate food or hosts (Keane and Crawley [2002\)](#page-10-5). The 'evolution of increased competitive ability hypothesis' (EICA) extends this assumption by assuming that plants experiencing reduced herbivory in a new range should be under strong selection to allocate less resources to defenses and more to growth and reproduction, resulting in an evolution of increased competitive ability in these plant populations (Blossey and Notzold [1995](#page-10-6)). The ERH has been contradicted by the notion that native generalist herbivores prefer alien to native species because the newcomers have not been selected to resist their new enemies (Parker and Hay [2005](#page-11-9); Morrison and Hay [2011](#page-11-10)). However, recent studies corroborated that aliens may have an advantage over native competitors since they contain potent defense compounds toward which the native consumers are evolutionarily naïve. This so-called novel weapons hypothesis (NWH) is indirectly supported by correlation studies, showing that highly invasive plants are more likely to possess novel chemical defenses compared to noninvasive species (Cappuccino and Carpenter [2005;](#page-10-7) Cappuccino and Arnason [2006](#page-10-8)). Enge et al. ([2012\)](#page-10-9) corroborated the NWH showing that the low consumption of the successful invader *Bonnemaisonia hamifera* might be attributed to a chemical defense toward which the indigenous consumers are evolutionarily naïve.

Despite increasing research on traits which promote invasion success and on links between invaders' secondary metabolites and feeding preference of local herbivores, there is still little direct evidence in support of the NWH or the evolutionary adaptation of chemical defenses to the invaded ecosystem. Most studies investigating correlations between plant invasion and palatability originate from terrestrial organisms and have produced inconsistent results (Dietz and Edwards [2006\)](#page-10-10). In this study, we compared the palatability of an invasive alga with closely related noninvasive species in order to find further support for the ERH or the NWH.

We used the brown algal species *Sargassum muticum* (Yendo) Fensholt as a model for a highly invasive seaweed. Originating from the Pacific (Chinese and Japanese Sea), it has been non-intentionally introduced to Europe with oysters in the early 1970 s (Critchley [1983\)](#page-10-11). Thereupon, it was reported from various locations along the Atlantic coast of Western Europe from Portugal to Scandinavia. *S. muticum* was first recorded in the German part of the North Sea in 1988 (Kornmann and Sahling [1994](#page-11-11); Karlsson and Loo [1999\)](#page-10-12) and is nowadays present in suitable habitats (i.e., rocky shores and artificial hard substrata) on most European Atlantic coasts (Engelen et al. [2015](#page-10-13)). It is still extending its range and has recently arrived on the coast of Morocco (Sabour et al. [2013\)](#page-11-12). *S. muticum* produces several defense compounds effective against bacteria, fungi, algal competitors and herbivores (Plouguerné et al. [2008](#page-11-13); Bazes et al. [2009](#page-10-14); Plouguerné et al. [2010\)](#page-11-14) and is a rich source of phlorotannins (Tanniou et al. [2014\)](#page-11-15), suggesting that the invasive success of *S. muticum* might be partly explained by its effective chemical defense. A low herbivore preference for *S. muticum* has been assumed to give the invader a competitive advantage over the native algal community along the Portuguese coast, thereby contributing to the invasiveness of *S. muticum* (Monteiro et al. [2009\)](#page-11-16). Several studies evaluated herbivory on invasive algal species in their introduced range, and in most cases, grazers from the introduced range preferred native to non-native algae (Weinberger et al. [2008](#page-12-2); Monteiro et al. [2009;](#page-11-16) Cacabelos et al. [2010;](#page-10-15) Engelen et al. [2011](#page-10-16); Nejrup et al. [2012](#page-11-17)). However, only few studies compared native and introduced populations of the same—or of closely related algal species. The study of Wikström et al. ([2006\)](#page-12-1) revealed that the brown alga *Fucus evanescens* is less exposed to herbivory in its introduced range in Sweden than in its native range in Iceland, probably due to increased chemical defenses in introduced populations. Similarly, Hammann et al. ([2013\)](#page-10-17) found a higher palatability of native compared to invasive *Gracilaria vermiculophylla* but with high regional variability between the native populations and without considering different life history stages. Thus, differences in palatability might also be explained by different herbivore preferences for haploid, diploid or even polyploid stages (Pereira and da Gama [2008](#page-11-18)).

Palatability relies not only on chemical defensive metabolites but also on the nutritional value or structural characteristics of algal food. Since herbivores are usually nitrogen-limited, they are assumed to prefer algae with a high protein content and a high proportion of nitrogen, i.e., with a low carbon-to-nitrogen (C:N) ratio (Mattson [1980](#page-11-19); Cruz-Rivera and Hay [2003](#page-10-18)). In general, herbivores look for energy-rich compounds and nutrients, such as proteins, nitrogen, and carbohydrates, and the availability of these primary compounds is therefore expected to affect food preference strongly (Mattson [1980\)](#page-11-19).

In this study, we compared the palatability and nutritional value of invasive *S. muticum* and two *Sargassum* <span id="page-2-0"></span>**Table 1** Extracted dry weight (dw), volume, yield of crude extract and the specific tissue concentration of freeze-dried algal tissue of *Sargassum* spp. from Japan and Germany and of *F. vesiculosus* from Germany



species from Japan: *Sargassum fusiforme* (Harvey) Setchell, which is currently not recorded as invasive, and *Sargas‑ sum horneri* (Turner) C. Agardh, which has recently been recorded along the southern region of the Pacific coast but has not yet reached the North Sea (Marks et al. [2015](#page-11-20)). Feeding preference was also compared to the local dominant North Sea brown alga *Fucus vesiculosus* Linnaeus 1753. Additionally, we compared intraspecific differences in palatability and chemistry of an invasive *S. muticum* population from the German North Sea and a native population from Japan and looked for possible adaptations and resulting chemical modification during the invasion process. We performed feeding assays and chemical analyses to test the following hypotheses: (1) the invasive alga *S. muticum* is less preferred by North Sea herbivores compared to a cooccurring competing brown alga as stated by the NWH; (2) the invasive alga *S. muticum* is less preferred by North Sea herbivores compared to close relatives which might explain its invasiveness; (3) *S. muticum* originating from invasive populations is in general less palatable than *S. muticum* from native populations because it has been selected for resistance against North Sea herbivores during the invasion process and (4) the evolved increased herbivore resistance is due to a different chemistry of invasive and native *S. muticum* due to changing environmental conditions and grazing pressure.

# **Materials and methods**

## **Collection of organism and extract preparation**

Whole algal thalli of adult *Sargassum* species were sampled before maturation in Japan and Germany. Three *Sar‑ gassum* species (Phaeophyceae), *S. fusiforme*, *S. horneri* and *S. muticum* were obtained from Japan by Scuba diving at a depth of 1–3 m in the Seto Inland Sea, Oshima, Japan (33°55′04.4″N132°27′42.7″E) during March 2013. The invasive *Sargassum muticum* from Germany was collected in June 2014 in the sheltered rocky intertidal zone of Heligoland, Germany (54° 11′05.3″N, 7°52′31.4″E). *Fucus vesiculosus* was collected by hand from Nassau harbor in Wilhelmshaven, Germany (53°30′54.7″N8°08′57.8″E) in April 2012. After collection, all *Sargassum* specimens were immediately transferred to the laboratory in insulated boxes filled with seawater, where they were gently washed with sterile seawater and cleaned from associated epibiota. Individuals of each algal species were pooled, wet weight  $(\pm 0.01 \text{ g})$  was measured with a balance, and the volume of each species was determined by water displacement in a graduated cylinder (Table [1\)](#page-2-0). Since we wanted to assess herbivore preferences based on chemical characteristics, the algae were subsequently frozen and freeze-dried to avoid changes in algal chemistry due to transport. Freezedried *Sargassum* spp. from Japan were sent to Germany for further processing and ground into fine powder to increase extraction efficiency. One part of the freeze-dried algal powder from Japan and from Germany was kept for the feeding assays, while the other was used for extraction of secondary metabolites.

Freeze-dried algal powder was extracted three times with ethylacetate (EtOAc)/methanol (MeOH) (AppliChem GmbH) (1:1, v/v) and finally with 100 % methanol. For each gram of algal weight, 20 mL of solvent was used. Each extract was filtered through Watman No. 1 filters, and remaining solvents were removed by rotary evaporation. The obtained extracts were combined, dried in a centrifugal vacuum concentrator, weighed and stored in a freezer at −20 °C until used in feeding assays (Table [1\)](#page-2-0).

# **Feeding assay**

# *Study organisms*

The palatability of the invasive *S. muticum* from the North Sea, hereafter *S. muticum* (inv.), was compared with the two noninvasive *Sargassum* species (*S. fusiforme, S. horneri*) and native *S. muticum* from Japan, hereafter *S. muticum* (nat.), using two common North Sea herbivores: *Littorina littorea* (Gastropoda), and *Psammechinus mil‑ iaris* (Echinodermata), and the omnivore: *Idotea baltica* (Isopoda). These species occur abundantly on seaweeds and are considered to be generalist consumers (Toth et al. [2007](#page-11-21); Enge et al. [2012;](#page-10-9) Kelly and Scheibling [2012;](#page-10-19) Nylund et al. [2012\)](#page-11-22). Isopods were taken from an *I. baltica* culture of the Alfred Wegener Institute, Bremerhaven, Germany.

Isopods were fed with customary fish food and the brown alga *Ascophyllum nodosum*, and maintained in an aerated 200 L flow-through tank with a 12/12 h light/dark cycle within a constant temperature room of 15 °C. New *I. bal‑ tica* individuals from drift algae collected in the Heligoland Bight were constantly introduced into the culture. During the experimental phase, isopods were maintained on their natural diet *A. nodosum* to avoid a preconditioning to any of our test algae.

*Littorina littorea* were collected between June and September 2014 from tide pools in the higher intertidal zone of the North Sea (53°30′46.9″N8°08′39.8″E). Snails were directly transferred to 27 L aerated seawater aquaria with a 12/12 h light/dark cycle and fed with *Ulva* sp. and *A. nodosum* up to 2 days before being used in experiments. Although it is known that some animals change their feeding preferences when they are starved, previous studies revealed similar feeding preferences for fed and 2 weeks starved *L. littorea* (Imrie et al. [1990;](#page-10-20) unpublished data). We used this fasting period to ensure feeding within 24 h. Water was exchanged manually two times a week, and temperature varied between 18–20 °C.

*Psammechinus miliaris* were collected around the German North Sea archipelago Heligoland (54°10′57″N 7°53′07″E) in July 2014. Sea urchins were maintained in 27 L aerated flow-through outdoor aquaria with flow-through seawater from the adjacent North Sea. Sea urchins were fed with fresh *Ulva* sp. and *A. nodo‑ sum* during a 4 weeks acclimation phase and between experiments.

Prior to the experiments, all herbivores were offered artificial food containing agar and pulverized *Ulva* sp. to get used to the agar-based diet. Snails were left without food 2 days before the experiment to ensure active feeding during the assays. Individual snails and isopods were only used once for the feeding experiments. Due to the smaller stock of sea urchins, they were applied in different feeding assays but with an interval of at least 4 weeks.

#### *Artificial food preparation*

Two kinds of artificial algal food were tested to analyze the feeding preferences of *L. littorea*, *I. baltica* and *P. mil‑ iaris*: powdered algae and algal extracts. Artificial food pellets containing powdered algae were produced to exclude the morphological characteristics. Consequently, feeding preferences were based on chemical and nutritional algal characteristics only. Food pellets with algal extracts were prepared to exclude both the morphologic structure and the nutritional value of *Sargassum* species to ensure that feeding preferences were predominantly due to chemical characteristics (Hay et al. [1994\)](#page-10-21).

Artificial food preparation was adapted from Hay et al. [\(1994](#page-10-21)) and Schupp and Paul ([1994\)](#page-11-23). To prepare the artificial food, 1.08 g agar was dissolved in 30 mL distilled  $H_2O$ and heated in a microwave. Dried and ground algae were added at natural concentrations corresponding to 30 mL volume to obtain the algal powder food pellets. Natural concentration was calculated as algal dry weight per 1 mL algal volume times 30 (Table [1](#page-2-0)). For the food with algal extracts, lyophilized *Ulva* sp., a local palatable green alga, served as basis for the artificial food. 3.3 g *Ulva* sp. powder was added to the microwaved agar and stirred rapidly for even distribution of the algal powder. Crude extracts of *Sar‑ gassum* spp. and *F. vesiculosus* were redissolved in 5 mL methanol and incorporated into the *Ulva*-agar mixture at natural volumetric concentration. Most of the methanol evaporates when added to the hot algal mixture, minimizing potential toxic effects. Additionally, the Ulva-agar mixture was treated with the same amount of methanol and consumed readily (data not shown). To achieve pieces of algal food in equal size and thickness, the hot agar-algae mixture was pressed onto a stainless steel window screen for sea urchin and on plastic window screen for snail and isopod food. After cooling, the agar food was firmly attached to the window screen and cut into pieces of  $2 \times 2$  cm for sea urchins and  $1 \times 1$  cm for snails and isopods and immediately used in feeding assays (Hay et al. [1994](#page-10-21); Rohde et al. [2004](#page-11-24))

#### *Two‑choice feeding assays*

In a crossed design, *Sargassum* spp. and *F. vesiculosus* were tested in two-choice feeding preference tests  $(n = 20)$ with each herbivore. Each of the three herbivores was allowed to choose between the same two varieties of artificial agar food, either as algal powder or extract. The assays were conducted in  $17 \times 13$  cm boxes filled with filtered seawater and one herbivore per box. In the case of *L. lit‑ torea,* three snails were inserted in each replicate due to low consumption rates. Herbivores were allowed to feed until half of one agar piece was consumed or until 48 h had passed. Preference was quantified as the number of entirely eaten window screen squares. Replicates where no food was consumed were excluded from statistical analysis.

## **Algal chemical defense and nutritional characteristics**

Algal nutritional value was characterized by measuring the wet/dry weight ratio (ww/dw), the amount of phenolic compounds, proteins and mannitol concentration. All the chemical analyses were run from algal material which was freeze-dried immediately after collection, pulverized and stored at −20 °C until final determination.

Such a procedure is advised to preserve the chemical constituents in the best possible way (Waterman and Mole 1994). Phenolic, protein and mannitol content were measured photometrically using a microplate reader (see below, Thermo Fisher Scientific Inc., Waltham, MA., USA).

## *Quantification of polyphenols*

The total phenolic content of all species used in the feeding assays was determined with a microplate-adapted Folin–Cioalteu assay following the procedure described in Zhang et al. [\(2006](#page-12-3)). Phloroglucinol (1,3,5-trihydroxybenzene, Sigma-Aldrich, Germany) was used as a standard, and a calibration curve was generated with concentrations of 0, 6.25, 12.5, 25, 50 and 100  $\mu$ g mL<sup>-1</sup>. Total phenolic contents (TPCs) were expressed as percentages of phenolic compounds per algal dw. The Folin–Ciocalteu method quantifies non-phenolic hydroxylated aromatic compounds as well, but since these interfering substances make up <5 % of the total reactive compounds, they were neglected (van Alstyne [1995\)](#page-12-4). In the following, the term 'phlorotannins' is used for total phenolics, since brown algae are not known to contain others polyphenols (Targett and Arnold [1998](#page-11-25)).

## *Protein determination*

The protein content was quantified using the microplateadapted Bradford method (Bradford [1976\)](#page-10-22). For the calibration curve, Bovine serum albumin (BSA) was used as a standard in concentrations of 20, 30, 40, 50, 60, 80 and 100 μg mL<sup>-1</sup>. Algal samples (0.1 g) were extracted with 1 M sodium hydroxide, and the absorbance was measured at 595 nm.

## *Mannitol determination*

To determine mannitol concentrations, 0.1 g of each algal sample was extracted and analyzed after Vas'kovskii and Isai ([1972\)](#page-12-5), with the difference that periodate oxidation was stopped after 10 s. The short reaction time is important since this method is not specific for mannitol and it also stains some other polyols. However, the formation of formaldehyde from mannitol is particularly fast, which allows the quantification of algal mannitol content.

# *C/N ratio*

Nitrogen and carbon contents were determined by using an elemental analyzer (Flash EA 1112, Thermo Fisher, Germany).

#### **Statistics**

Statistical calculations were performed using SPSS IBM Statistics version 23, Illinois, USA. Normality and homogeneity of variances were determined using the Kolmogorov–Smirnov test and Levene's tests, respectively. Since no homoscedasticity was achieved in the feeding preference assays, means of the Wilcoxon signed-ranks paired test have been used to identify significant differences in the feeding preference assay.

A one-way ANOVA was used to assess differences of algal characteristics in terms of phlorotannin, mannitol and protein contents of *Sargassum* spp. and *F. vesiculosus* extracts. Pairwise differences were analyzed using Tukey post hoc tests.

Spearman's rank correlation was performed to examine relationships between feeding preferences of the three herbivore species and determined algal characteristics.

In all cases, the threshold for significance was  $\alpha = 0.05$ .

# **Results**

#### **Feeding preferences**

All herbivore species demonstrated very similar feeding preferences in the two-choice feeding assays. All herbivores preferred the local brown algae *F. vesiculosus* and the invasive population of *S. muticum* (inv.), while the *S. muticum* population from Japan, as well as *Sargassum fusi‑ forme* and *Sargassum horneri*, were less consumed in all assays. Not all combinations were tested since the assumption that  $A > C$  when  $A > B$  and  $B > C$  seems to be justified when the preferences among five species could be linearly arranged without contradiction between preference results. However, at both extremes of the proposed preference gradient, preferences were not always clear (Fig. [1](#page-5-0)a–c). Thus, the resulting ranking of the Japanese species should be considered with some caution (Table [2\)](#page-6-0). Especially, the gastropod *L. littorea* and the echinoderm *P. miliaris* were overall less selective than the isopod *I. baltica* and most choices were not significant. Nevertheless, *L. littorea* significantly preferred *F. vesiculosus* over every *Sargassum* species and *S. muticum* (inv.) over *Sargassum* spp. from Japan (Fig. [1b](#page-5-0), *p* < 0.05). Surprisingly, both *L. littorea* and *P. miliaris* clearly differentiated between the invasive and native *S. muticum* with 97 and 81 % more consumption of the artificial food containing North Sea *S. muticum* extracts, respectively (Fig. [1b](#page-5-0), c).

Artificial food type (algal powder or extract) and herbivore species had overall no significant influence on the ratio of consumed algae (Table [2\)](#page-6-0). In the case of *L. littorea*



<span id="page-5-0"></span>**Fig. 1** Mean consumption  $(+SD, n = 20)$  by the North Sea herbivores **a** *Idotea baltica*, **b** *Littorina littorea*, and **c** *Psammechinus mil‑ iaris.* Herbivory was tested in two-choice experiments with artificial food containing either algal powder or algal extracts of the species *S. fusiforme* (SF; striped), *S. horneri* (SH; *dotted*), *S. muticum* (SM

(nat)) from Japan (blank), as well as *S. muticum* (SM (inv.; *black*) and *F. vesiculosus* (FV; *gray*) from the North Sea. Significant consumption differences are indicated by asterisks above the *bars* (n.s. = nonsignificant, \**p* < 0.05, \*\**p* < 0.01, \*\*\**p* < 0.001, Wilcoxon signedranks test)

<span id="page-6-0"></span>**Table 2** Ranking of feeding preferences of the North Sea herbivores *Idotea baltica*, *Littorina littorea* and *Psammechinus miliaris* when fed with two types of artificial food containing either algal powder or algal crude extract of *F. vesiculosus* (FV), the *Sargassum* species *S. fusiforme* (SF), *S. horneri* (SH) and *S. muticum* (SM). SM included

specimens both from the North Sea population (inv.) and a population from Japan (nat.). Preference ranking derived from food consumption in two-choice feeding trials is defined as the difference of consumed food whether significant or not. 5 is the most and 1 the least preferred species



#### <span id="page-6-1"></span>**Fig. 2** Tissue contents

(means  $+$  SD;  $n = 3$ ) of **a** phlorotannin, **b** protein, **c** mannitol and **d** C/N ratio for *Sargassum* spp. from Japan (*white bars*; *S. muticum* (SM (nat), *S. fusiforme* (SF), *S. horneri* (SH)) and *F. vesiculosus* (FV) and *S. muti‑ cum* (SM (inv.) from the North Sea (*black bars*). Different letters above the *bars* indicate significant differences between tissue concentrations (ANOVA, Tukey post hoc test,  $p < 0.05$ )



and *P. miliaris,* the ranking of algal species remained equal with changing food type. Rank four and five as well as two and three changed during assays with *I. baltica* when offered food pellets containing algal extracts compared to algal powder.

# **Algal characteristics**

Mean phlorotannin concentration of *F. vesiculosus* and native *S. muticum* ranged from 3.32 to 5.87 % dw, respectively, and exceeded other *Sargassum* spp. by up to 6.7 times in the case of *S. horneri* (Fig. [2](#page-6-1)a). The protein content of *F. vesiculosus* (3.58 % dw) was significantly lower compared to *Sargassum* spp. and was around 33 % below the native *S. muticum,* which exhibited the highest protein concentration with 10.64 % dw (Fig. [2b](#page-6-1)). The native *S.* 

*muticum* and *S. fusiforme* are characterized by elevated protein content, while native and invasive *S. muticum* exhibited higher mannitol contents (Fig. [2](#page-6-1)b, c). Mannitol content of the native *S. muticum* was about 21 times higher compared to *S. horneri* but was even exceeded by the mannitol concentration of *F. vesiculosus* which averaged 12.24 % dw (Fig. [2c](#page-6-1)). The C/N ratio ranged between the lowest ratio for *S. fusiforme* with 9.33 and the high ratios for the native *S. muticum* and *F. vesiculosus* with 19.84 and 22.08, respectively (Fig. [2d](#page-6-1)).

A Spearman's rank-order correlation was run to determine the relationship between algal preference ranking and the nutritional value of powdered algal food (Table [3](#page-7-0)). There was a significant positive correlation between isopod feeding preference, phlorotannin ( $r_s = 0.655$ ,  $p = 0.008$ ), mannitol content ( $r_s = 0.895$ ,  $p < 0.001$ ) and C/N ratio

<span id="page-7-0"></span>

 $(r<sub>s</sub> = 0.753, p = 0.001)$ . While protein content did not significantly correlate with isopod feeding preference  $(p = 0.187)$ , it was negatively correlated with the feeding preference of *L. littorea* ( $r_s = -0.655$ ,  $p = 0.01$ ) and *P. miliaris* ( $r_s = -0.862$ ,  $p < 0.001$ ).

### **Discussion**

Herbivory is especially intense in marine environments, with approximately 70 % of benthic primary production being consumed by herbivores globally (Poore et al. [2012](#page-11-26)). Therefore, feeding preferences of native generalist herbivores may determine community composition and could influence algal invasion success in two opposing ways: Native generalists could either preferentially attack exotics which are not defended against herbivores in their new range and consequently suppress their abundance (e.g., Lind and Parker [2010;](#page-11-27) Morrison and Hay [2011](#page-11-10)), or release less preferred invaders from predation (Wikström et al. [2006](#page-12-1); Monteiro et al. [2009;](#page-11-16) Nejrup et al. [2012](#page-11-17)).

In our study, the three tested local herbivores clearly preferred the local brown alga *F. vesiculosus* over *S. muticum* (inv.), while the three *Sargassum* species from Japan were less consumed in all assays. The detected preference for a native alga is consistent with the ERH and several studies which quantified grazing on invasive seaweed species in their new range. In most cases, grazers from the invaded region preferred native over non-native algae (Cappuccino and Carpenter [2005;](#page-10-7) Weinberger et al. [2008;](#page-12-2) Monteiro et al. [2009](#page-11-16); Cacabelos et al. [2010](#page-10-15); Engelen et al. [2011;](#page-10-16) Nejrup et al. [2012](#page-11-17)). The lower grazing pressure on *S. muticum* compared to its abundant competitor *F. vesiculosus* might partly explain the invasive success of *S. muticum* in the North Sea. Engelen et al. ([2011\)](#page-10-16) investigated whether the low food preference for *S. muticum* reported by Monteiro et al. ([2009\)](#page-11-16) also holds when tested with different foods and herbivores. Their results confirmed the avoidance of *S. muticum* compared to native algae by all tested herbivores. Additionally, the high growth rate of *S. muticum* was less affected by the grazers compared to native seaweeds which showed a decreased growth rate in the presence of grazers. Similarly as in our study, Johnsen et al. [\(2013](#page-10-23)) found an overall tendency for the avoidance of *S. muticum* compared to native algae by *P. miliaris*. Nevertheless, the ERH does not only relate to the complete absence of grazing on a non-native species, but also to relative grazing pressure on non-native species when compared with native, co-occurring species. Preference for and higher consumption of the dominant brown alga *F. vesiculosus* may lower the grazing pressure on the non-native *S. muticum* and even release it from potential competition with indigenous species, thereby increasing the overall fitness and the space extension of invasive *S. muticum* within its new range. Since *S. muticum* (inv.) was not completely refused by the tested herbivores in our study, it can be argued that it was the reduction in herbivore intensity rather than the escape from (native) herbivores that provided the invasive macroalga with an advantage over the local *F. vesiculosus* (Vermeij et al. [2009\)](#page-12-6). Since there are no publications about herbivory of *S. muticum* in its native range, the threat of predation by those diverse herbivores inhabiting *S. muticum's* native range can merely be assumed (Trowbridge et al. [2009\)](#page-11-28).

Artificial algal food type (powder or extract) had overall no significant influence on the ratio of consumed algae (Table [2\)](#page-6-0), thus indicating that algal chemistry is responsible for herbivore preference. This is concurrent with the NWH, since *S. muticum* might possess exotic metabolites to which North Sea herbivores are evolutionarily naïve. Additionally, chemical metabolites seem to be efficient against a broad range of generalist herbivores, given that different herbivores did not change the general pattern of preference for the local North Sea algae compared to *Sargassum* species from Japan.

Low preference of herbivores for *S. muticum* has been linked to the presence of secondary metabolites, such as the relatively high levels of phenolic compounds (Monteiro et al. [2009](#page-11-16)). This correlates with our result that the least preferred *S. muticum* (nat.) exhibited the highest phlorotannin content compared to all other tested species. However, the native *S. horneri* and *S. fusiforme* were similarly avoided by herbivores and revealed the lowest phlorotannin contents. Previous studies have shown opposing effects of phlorotannin on herbivores (Targett and Arnold [1998;](#page-11-25) Deal et al. [2003\)](#page-10-24), suggesting that adaptations to utilize phlorotannin-rich algae for food have evolved among marine herbivores. Especially, *I. baltica* has been shown to be adapted to feed on phlorotannin-rich species. This herbivore even

preferred those algae with the highest phlorotannin content in feeding choice experiments (Jormalainen et al. [2001](#page-10-25)). Furthermore, we found a positive correlation between preference ranking and phlorotannin content, meaning that preference increases with increasing phlorotannin concentrations, and also suggesting that other metabolites are responsible for feeding deterrence of Japanese *Sargassum* species. Protein content decreased with increasing feeding preferences of *I. baltica, L. littorea* and *P. miliaris*. Thus, the most preferred *F. vesiculosus* and *S. muticum* (inv.) exhibited the lowest protein per dry weight. *I. baltica* feeding preference and C/N ratio is however positively correlated, meaning that most preferred *F. vesiculosus* exhibited the highest C/N ratio. Herbivores are generally nitrogenlimited and depend on the intake of food with a high protein content and a high proportion of nitrogen, thus with a low (C/N) ratio (Mattson [1980;](#page-11-19) Cruz-Rivera and Hay [2003](#page-10-18)). Therefore, the preference for algae with low nitrogen content might be another indication that algal palatability is determined by other algal characteristics, such as the quality and quantity of secondary metabolites. Only algal mannitol content might contribute to feeding preference with a significant positive correlation between *I. baltica* feeding preference and an increasing amount of mannitol per dry weight. Mannitol-containing food has previously been shown to attract *I. baltica* (Weinberger et al. [2011\)](#page-12-7) since mannitol food concentration positively co-varied with egg size and therefore plays an important role in the reproductive performance of isopods (Hemmi and Jormalainen [2002](#page-10-26), [2004](#page-10-27)).

Interestingly, both *L. littorea* and *P. miliaris* clearly differentiate between the invasive and native *S. muticum* with a 97 and 81 % higher consumption of food containing North Sea *S. muticum* extracts, respectively. This result is contrary to our initial hypothesis that invasive *S. muticum* populations are in general less palatable than *S. muticum* from native populations because they have been selected for resistance against herbivores during the invasion process. It seems rather likely that the defensive level of *S. muticum* decreased after the establishment in the North Sea. Since there is not much known about *S. muticum*–herbivore interactions in Japan, we only can hypothesize what happened during its invasion process. Japanese shores have a higher density of macroalgae and herbivores, probably resulting in more species interactions compared to other temperate shores (Trowbridge et al. [2009\)](#page-11-28). Thus, Japanese *S. muticum* might invest more energy in costly defenses against herbivores and competitors to persist in this highly competitive environment. Anti-feeding compounds might have aided the establishment of *S. muticum,* which benefited from the higher grazing pressure on its strongest competitor *F. vesiculosus*. These suggestions are in line with the 'novel weapons hypothesis' (NWH). Many studies suggest that well-defended introduced species can get a significant advantage over native competitors and rapidly become abundant in communities dominated by generalist consumers (Callaway and Ridenour [2004;](#page-10-28) Cappuccino and Carpenter [2005;](#page-10-7) Cappuccino and Arnason [2006](#page-10-8); Forslund et al. [2010](#page-10-29)). After the positive establishment of *S. muticum* in its new range, it might have come to a resource allocation shift from the production of defense metabolites to reproduction and/or growth (i.e., evolution of increased competitive ability or EICA hypothesis; Blossey and Notzold [1995\)](#page-10-6). A terrestrial metaanalysis by (Hawkes [2007\)](#page-10-30) which compared conspecifics in native and introduced ranges provides further clues for possible evolutionary adaptations of introduced organisms due to altered environmental conditions. Furthermore, the author's analyses suggested that conspecifics were generally larger and allocated more to reproduction in the introduced range compared with the native range. In fact, *S. muticum* is one of the smaller *Sargassum* species in Japan (75–120 cm; Josefsson and Jansson [2011](#page-10-31); Yendo [1907](#page-12-8)), but grows considerably larger when introduced into new areas and reaches a length of >4 m in the German North Sea (Polte and Buschbaum [2008\)](#page-11-29). However, the size of *S. muticum* in its native range is controversial. The sampled *S. muticum* used in the present study was about 1–1.5 m long and reached maximum length of 2 m after maturation (pers. communication), but there are recent findings of large specimens of up to 3.2 m (Engelen et al. [2015](#page-10-13)). It seems that there have been two phases of invasion (Dietz and Edwards [2006\)](#page-10-10) into the North Sea. In phase one, the pre-adapted highly defended *S. muticum* arrived in the new range where it benefited from grazer avoidance compared to the local dominant brown alga *F. vesiculosus*. Grazer attacks decreased growth rates of *F. vesiculosus* and therefore facilitated the rapid establishment of *S. muticum* in the German North Sea. In phase two, the continuous low grazing pressure resulted in a resource allocation shift from the production of defense metabolites to reproduction or growth, which is reflected by the higher length of *S. muti‑ cum* in the North Sea compared to Japan. This caused a selection of the less defended but more vigorous or competitive genotypes (Blossey and Notzold [1995;](#page-10-6) Callaway and Ridenour [2004](#page-10-28); Cano et al. [2009](#page-10-32)). However, this mechanism is highly speculative and other studies found opposite results. Non-native populations of the East Asian seaweed *Gracilaria vermiculophylla* have been shown to be generally less palatable to marine herbivores than native populations, and the increased capacity for activated chemical defense is suggested to have contributed to their invasion success (Hammann et al. [2016\)](#page-10-33). Furthermore, Le Cam et al. [\(2015](#page-11-30)) found limited genetic polymorphism of *S. muticum* populations in the invaded ranges and a perfect match of the invaded genotype in one of the native population, implying a restricted probability of genetic adaptation due to the release from predation. A resource allocation shift might therefore rather result from gene expression regulation than from real adaptation through gene modification.

Additionally, differences in feeding preference might arise from other confounding factors. The more apical and younger areas of the thallus are usually defended more strongly (Pavia et al. [2002\)](#page-11-31), and the sampling of *Sargassum* populations of different developmental stages might result in more or less feeding deterrence. Phenolic content of *S. muticum* in Brittany peaked during the reproductive period, which was hypothesized as providing a maximum protection of the fertile receptacles from both grazing and solar radiation (Plouguerné et al. [2006\)](#page-11-32). Thus, different tissue types or developmental stages of native and invasive *S. muticum* might result in different feeding preferences. We tried to minimize these factors in this study by sampling whole algal thalli of *Sargassum* spp. North Sea and Japanese populations and before maturation only. Furthermore, algal metabolites can differ among populations over scales of hundreds of kilometers. Phlorotannin concentration of *Ascophyllum nodu‑ sum* populations differed significantly at scales of meters up to one kilometer, whereas only little variations were found among populations located 1000 km apart (Pavia and Åberg [1996\)](#page-11-33). Therefore, general assumptions about invasive and native *Sargassum* spp. should be handled with caution.

While *S. muticum* was significantly less preferred to *F. vesiculosus* by all tested herbivores, there is no clear evidence that it is better defended than its native, non- or weakly-invasive relatives. *S. muticum* (nat.) has in fact been the least preferred by two out of three tested herbivores. However, this is just a weak indication for explaining the invasion success of *S. muticum* compared to *S. horneri* or *S. fusiforme*. There have been a number of other traits besides, or in conjunction with, herbivore defense (Carpenter and Cappuccino [2005;](#page-10-34) Engelen et al. [2011\)](#page-10-16) that have been found to be important for the establishment and dispersal of introduced species: Faster growth or higher fecundity (van Kleunen et al. [2010](#page-12-9); Engelen et al. [2011\)](#page-10-16), positive response to disturbance (Deysher and Norton [1982\)](#page-10-35), and invasion melt-downs where non-native herbivores selectively suppress native plants and facilitate invasion by non-native plants that have evolved with these invasive herbivores (Parker et al. [2006\)](#page-11-34). *S. muticum* shows several of these characteristics with its high reproductive output, fast growth and great potential to colonize uninhabited areas (Deysher and Norton [1982](#page-10-35); Wernberg et al. [2000](#page-12-10); Engelen et al. [2011](#page-10-16)). Additionally, *S. muticum* exerts other adverse effects on the local algal community. The introduction of *S. muticum* in Limfjorden has led to a decline in slow growing perennial species, such as *Fucus vesiculosus*, *Fucus serratus, Saccharina latissima* and *Codium fragile*. It has been suggested that the reduction of these algae is due to competition for space with *S. muticum* (Stæhr et al. [2000\)](#page-11-35). Furthermore, *S. muticum* is less affected

by herbivory and can compensate grazing losses through high growth rates, while growth of local co-occurring species is reduced due to grazing. On the other hand, it can be argued that the well-defended Japanese *S. fusiforme* and *S. horneri* might invade the North Sea in future as well. The high growth rates of *S. muticum* of 4 cm day<sup>-1</sup> under optimal conditions (Jephson and Gray [1977](#page-10-36)) has not yet been achieved by *S. horneri* (0.1–0.9 cm day−<sup>1</sup> in situ; Gao and Hua  $1997$ ) or *S. fusiforme* (0.2 cm day<sup>-1</sup> in the laboratory; Zou [2005](#page-12-11)). However, *S. horneri* has recently been detected in California, USA, from where it spread rapidly along the southern region of the Pacific Coast and might achieve higher growth rates with further dispersal (Miller et al. [2007](#page-11-36); Marks et al. [2015](#page-11-20)). It might therefore be reasonable not to focus on *S. muticum* only, but to include other *Sargassum* species in North Sea monitoring programs to prevent their establishment in European waters.

In conclusion, the invasion success of *S. muticum* in the North Sea might have been facilitated by feeding preferences of local herbivores. The invasive *S. muticum* was less preferred compared to its local competitor *F. vesiculo‑ sus* which might have facilitated the expansion of *S. muti‑ cum* in its new range. The native *S. muticum* population from Japan was less preferred compared to its allopatric population from the North Sea. This might be the result of a resource allocation shift due to reduced grazing pressure in the North Sea.

More comparisons between invasive and noninvasive related species should be conducted in various environments differing in grazing pressure and competition in order to get a closer insight into invasive traits and evolutionary adaptations.

Since other *Sargassum* species from Japan are equally defended against grazers like *S. muticum* from Japan, they should be included in European monitoring programs.

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#### **Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no potential conflict of interest.

**Ethical approval** All applicable international, national, and/or institutional guidelines for the collection, care and use of organisms were followed.

#### **References**

<span id="page-9-0"></span>Anderson LWJ (2007) Control of invasive seaweeds. Bot Mar 50:418–437. doi[:10.1515/BOT.2007.045](http://dx.doi.org/10.1515/BOT.2007.045)

- <span id="page-10-14"></span>Bazes A, Silkina A, Douzenel P et al (2009) Investigation of the antifouling constituents from the brown alga *Sargassum muticum* (Yendo) Fensholt. J Appl Phycol 21:395–403. doi[:10.1007/](http://dx.doi.org/10.1007/s10811-008-9382-9) [s10811-008-9382-9](http://dx.doi.org/10.1007/s10811-008-9382-9)
- <span id="page-10-6"></span>Blossey B, Notzold R (1995) Evolution of increased competitive ability in invasive nonindigenous plants—a hypothesis. J Ecol 83:887–889
- <span id="page-10-22"></span>Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72:248–254. doi[:10.1016/0003-2697\(76\)90527-3](http://dx.doi.org/10.1016/0003-2697(76)90527-3)
- <span id="page-10-15"></span>Cacabelos E, Olabarria C, Incera M, Troncoso JS (2010) Do grazers prefer invasive seaweeds? J Exp Mar Bio Ecol 393:182–187. doi[:10.1016/j.jembe.2010.07.024](http://dx.doi.org/10.1016/j.jembe.2010.07.024)
- <span id="page-10-28"></span>Callaway RM, Ridenour WM (2004) Novel weapons: invasive success and the evolution of increased competitive ability. Front Ecol Environ 2:436–443. doi:10.1890/1540-9295(2004)002[0436:NWI SAT]2.0.CO;2
- <span id="page-10-32"></span>Cano L, Escarré J, Vrieling K, Sans FX (2009) Palatability to a generalist herbivore, defence and growth of invasive and native *Senecio* species: testing the evolution of increased competitive ability hypothesis. Oecologia 159:95–106. doi[:10.1007/](http://dx.doi.org/10.1007/s00442-008-1182-z) [s00442-008-1182-z](http://dx.doi.org/10.1007/s00442-008-1182-z)
- <span id="page-10-8"></span>Cappuccino N, Arnason JT (2006) Novel chemistry of invasive exotic plants. Biol Lett 2:189–193. doi[:10.1098/rsbl.2005.0433](http://dx.doi.org/10.1098/rsbl.2005.0433)
- <span id="page-10-7"></span>Cappuccino N, Carpenter D (2005) Invasive exotic plants suffer less herbivory than non-invasive exotic plants. Biol Lett 1:435–438. doi[:10.1098/rsbl.2005.0341](http://dx.doi.org/10.1098/rsbl.2005.0341)
- <span id="page-10-1"></span>Carlton JT (2009) Deep invasion ecology and the assembly of communities in historical time. Biol Invasions Mar Ecosyst 13–56. doi[:10.1007/978-3-540-79236-9\\_2](http://dx.doi.org/10.1007/978-3-540-79236-9_2)
- <span id="page-10-34"></span>Carpenter D, Cappuccino N (2005) Herbivory, time since introduction and the invasiveness of exotic plants. J Ecol 93:315–321. doi[:10.1111/j.1365-2745.2005.00973.x](http://dx.doi.org/10.1111/j.1365-2745.2005.00973.x)
- <span id="page-10-4"></span>Cassano V, De-Paula JC, Fujii MT et al (2008) Sesquiterpenes from the introduced red seaweed *Laurencia caduciramulosa* (Rhodomelaceae, Ceramiales). Biochem Syst Ecol 36:223–226. doi[:10.1016/j.bse.2007.07.005](http://dx.doi.org/10.1016/j.bse.2007.07.005)
- <span id="page-10-11"></span>Critchley AT (1983) *Sargassum muticum*—a taxonomic history including world-wide and western Pacific distributions. J Mar Biol Assoc United Kingdom 63:617–625
- <span id="page-10-18"></span>Cruz-Rivera E, Hay ME (2003) Prey nutritional quality interacts with chemical defenses to affect consumer feeding and fitness. Ecol Monogr 73:483–506. doi:[10.1890/0012-9615\(2003\)073](http://dx.doi.org/10.1890/0012-9615(2003)073)
- <span id="page-10-3"></span>Daehler CC (2003) Performance comparisons of co-occurring native and alien invasive plants: implications for conservation and restoration. Annu Rev Ecol Evol Syst 34:183–211. doi[:10.1146/132403](http://dx.doi.org/10.1146/132403)
- <span id="page-10-2"></span>Davidson AD, Campbell ML, Hewitt CL, Schaffelke B (2015) Assessing the impacts of nonindigenous marine macroalgae: an update of current knowledge. Bot Mar 58:55–79. doi[:10.1515/bot-2014-0079](http://dx.doi.org/10.1515/bot-2014-0079)
- <span id="page-10-24"></span>Deal MS, Hay ME, Wilson D, Fenical W (2003) Galactolipids rather than phlorotannins as herbivore deterrents in the brown seaweed *Fucus vesiculosus*. Oecologia 136:107–114. doi[:10.1007/](http://dx.doi.org/10.1007/s00442-003-1242-3) [s00442-003-1242-3](http://dx.doi.org/10.1007/s00442-003-1242-3)
- <span id="page-10-35"></span>Deysher L, Norton TA (1982) Dispersal and colonization in *Sargas‑ sum muticum* (Yendo) Fensholt. J Exp Mar Bio Ecol 56:179– 195. doi[:10.1016/0022-0981\(81\)90188-X](http://dx.doi.org/10.1016/0022-0981(81)90188-X)
- <span id="page-10-10"></span>Dietz H, Edwards PJPJ (2006) Recognition that causal processes change during plant invasion helps explain conflicts in evidence. Ecology 87:1359–1367. doi:10.1890/0012-9658 (2006)87[1359:RTCPCD]2.0.CO;2
- <span id="page-10-9"></span>Enge S, Nylund GM, Harder T, Pavia H (2012) An exotic chemical weapon explains low herbivore damage in an invasive alga. Ecology 93:2736–2745. doi[:10.1890/12-0143.1](http://dx.doi.org/10.1890/12-0143.1)
- <span id="page-10-16"></span>Engelen AH, Henriques N, Monteiro C, Santos R (2011) Mesograzers prefer mostly native seaweeds over the invasive brown seaweed

*Sargassum muticum*. Hydrobiologia 669:157–165. doi[:10.1007/](http://dx.doi.org/10.1007/s10750-011-0680-x) [s10750-011-0680-x](http://dx.doi.org/10.1007/s10750-011-0680-x)

- <span id="page-10-13"></span>Engelen AH, Serebryakova A, Ang P, Britton-Simmons K, Mineur F, Pedersen MF, Arenas F et al (2015) Circumglobal invasion by the brown seaweed *Sargassum muticum*. Ocean Mar Biol Ann Rev 53:81–126
- <span id="page-10-29"></span>Forslund H, Wikström SA, Pavia H (2010) Higher resistance to herbivory in introduced compared to native populations of a seaweed. Oecologia 164:833–840. doi:[10.1007/s00442-010-1767-1](http://dx.doi.org/10.1007/s00442-010-1767-1)
- <span id="page-10-37"></span>Gao K, Hua W (1997) In Situ growth rates of *Sargassum horneri* (Fucales, Phaeophyta). Phycol. Res. 45:55–57
- <span id="page-10-0"></span>Halpern BS, Walbridge S, Selkoe KA et al (2008) A global map of human impact on marine ecosystems. Science 319(80):948–953. doi[:10.1126/science.1149345](http://dx.doi.org/10.1126/science.1149345)
- <span id="page-10-17"></span>Hammann M, Wang G, Rickert E et al (2013) Invasion success of the seaweed *Gracilaria vermiculophylla* correlates with low palatibility. Mar Ecol Prog Ser 486:93–103. doi:[10.3354/meps10361](http://dx.doi.org/10.3354/meps10361)
- <span id="page-10-33"></span>Hammann M, Rempt M, Pohnert G, Wang G, Boo SM, Weinberger F (2016) Increased potential for wound activated production of Prostaglandin E2 and related toxic compounds in non-native populations of *Gracilaria vermiculophylla*. Harmful Algae 51:81–88. doi:[10.1016/j.hal.2015.11.009](http://dx.doi.org/10.1016/j.hal.2015.11.009)
- <span id="page-10-30"></span>Hawkes CV (2007) Are invaders moving targets? The generality and persistence of advantages in size, reproduction, and enemy release in invasive plant species with time since introduction. Am Nat 170:832–843. doi:[10.1086/522842](http://dx.doi.org/10.1086/522842)
- <span id="page-10-21"></span>Hay ME, Kappel QE, Fenical W (1994) Synergisms in plant defenses against herbivores: Interactions of chemistry, calcification, and plant quality. Ecology 75:1714–1726
- <span id="page-10-26"></span>Hemmi A, Jormalainen V (2002) Nutrient enhancement increases performance of a marine herbivore via quality of its food alga. Ecology 83:1052–1064. doi:10.1890/0012-9658(2002)083[1052:NEI POA]2.0.CO;2
- <span id="page-10-27"></span>Hemmi A, Jormalainen V (2004) Geographic covariation of chemical quality of the host alga *Fucus vesiculosus* with fitness of the herbivorous isopod *Idotea baltica*. Mar Biol 145:759–768. doi[:10.1007/s00227-004-1360-4](http://dx.doi.org/10.1007/s00227-004-1360-4)
- <span id="page-10-20"></span>Imrie DW, McCrohan CR, Hawkins SJ (1990) Feeding behaviour in *Littorina littorea*: a study of the effects of ingestive conditioning and previous dietary history on food preference and rates of consumption. Hydrobiologa 193:191–198
- <span id="page-10-36"></span>Jephson NA, Gray PWG (1977) Aspects of the ecology of *Sargassum muticum* (Yendo) Fensholt, in the Solent Region of the British Isles. 1. The growth cycle and epiphytes. In: Keegan BF, Boaden PJS, Ceidigh PO (eds) Biology of benthic organisms. 11th European symposium on marine biology. Pergamon Press, Oxford, pp 367–375
- <span id="page-10-23"></span>Johnsen KL, Halle LL, Karling ND (2013) Can grazer avoidance explain the invasiveness of the brown alga *Sargassum muti‑ cum* in Limfjorden, Denmark? Thesis, Roskilde Universitet, pp 12–17
- <span id="page-10-25"></span>Jormalainen V, Honkanen T, Heikkilä N (2001) Feeding preferences and performance of a marine isopod on seaweed hosts: cost of habitat specialization. Mar Ecol Prog Ser 220:219–230. doi[:10.3354/meps220219](http://dx.doi.org/10.3354/meps220219)
- <span id="page-10-31"></span>Josefsson M, Jansson K (2011) NOBANIS—invasive Alien Species fact sheet—*Sargassum muticum*. In: Online Database Eur. Netw. Invasive Alien Species—NOBANIS [www.nobanis.org](http://www.nobanis.org)
- <span id="page-10-12"></span>Karlsson J, Loo LO (1999) On the distribution and continuous expansion of the Japanese seaweed—*Sargassum muticum*—in Sweden. Bot Mar 42:285–294. doi[:10.1515/BOT.1999.032](http://dx.doi.org/10.1515/BOT.1999.032)
- <span id="page-10-5"></span>Keane RM, Crawley MJ (2002) Exotic plant invasions and the enemy release hypothesis. Trends Ecol Evol 17:164–170. doi[:10.1016/](http://dx.doi.org/10.1016/S0169-5347(02)02499-0) [S0169-5347\(02\)02499-0](http://dx.doi.org/10.1016/S0169-5347(02)02499-0)
- <span id="page-10-19"></span>Kelly JR, Scheibling RE (2012) Fatty acids as dietary tracers in benthic food webs. Mar Ecol Prog Ser 446:1–22. doi[:10.3354/](http://dx.doi.org/10.3354/meps09559) [meps09559](http://dx.doi.org/10.3354/meps09559)
- <span id="page-11-11"></span>Kornmann P, Sahling PH (1994) Meeresalgen von Helgoland: Zweite Ergänzung. Helgoländer Meeresuntersuchungen 48:365–406. doi[:10.1007/BF02366253](http://dx.doi.org/10.1007/BF02366253)
- <span id="page-11-30"></span>Le Cam S, Thiebaut CD, Bouchemousse S, Viard F (2015) Elucidating unresolved invasion history with genome-wide sequencing approach: the case of the global invader *Sargassum muticum*. Eur J Phycol 50:24–25. doi:[10.1080/09670262.2015.1069489](http://dx.doi.org/10.1080/09670262.2015.1069489)
- <span id="page-11-27"></span>Lind EM, Parker JD (2010) Novel weapons testing: are invasive plants more chemically defended than native plants? PLoS ONE. doi:[10.1371/journal.pone.0010429](http://dx.doi.org/10.1371/journal.pone.0010429)
- <span id="page-11-20"></span>Marks LM, Salinas-Ruiz P, Reed DC et al (2015) Range expansion of a non-native, invasive macroalga *Sargassum horneri* (Turner) C. Agardh, 1820 in the eastern Pacific. BioInvasions Rec 4:243–248
- <span id="page-11-19"></span>Mattson WJJ (1980) Herbivory in relation to plant nitrogen content. Annu Rev Ecol Syst 11:119–161
- <span id="page-11-0"></span>Millennium Ecosystem Assessment (2005) Ecosystems and human well-being: synthesis. Island Press, Washington
- <span id="page-11-36"></span>Miller KA, Engle JM, Uwai S, Kawai H (2007) First report of the Asian seaweed *Sargassum filicinum* Harvey (Fucales) in California, USA. Biol Invasions 9:609–613. doi[:10.1007/](http://dx.doi.org/10.1007/s10530-006-9060-2) [s10530-006-9060-2](http://dx.doi.org/10.1007/s10530-006-9060-2)
- <span id="page-11-16"></span>Monteiro CA, Engelen AH, Santos ROP (2009) Macro- and mesoherbivores prefer native seaweeds over the invasive brown seaweed *Sargassum muticum*: a potential regulating role on invasions. Mar Biol 156:2505–2515. doi:[10.1007/s00227-009-1275-1](http://dx.doi.org/10.1007/s00227-009-1275-1)
- <span id="page-11-10"></span>Morrison WE, Hay ME (2011) Herbivore preference for native vs. exotic plants: generalist herbivores from multiple continents prefer exotic plants that are evolutionarily naïve. PLoS ONE. doi[:10.1371/journal.pone.0017227](http://dx.doi.org/10.1371/journal.pone.0017227)
- <span id="page-11-17"></span>Nejrup LB, Pedersen MF, Vinzent J (2012) Grazer avoidance may explain the invasiveness of the red alga *Gracilaria vermicu‑ lophylla* in Scandinavian waters. Mar Biol 159:1703–1712. doi[:10.1007/s00227-012-1959-9](http://dx.doi.org/10.1007/s00227-012-1959-9)
- <span id="page-11-6"></span>Nyberg CD, Wallentinus I (2005) Can species traits be used to predict marine macroalgal introductions? Biol Invasions 7:265–279. doi[:10.1007/s10530-004-0738-z](http://dx.doi.org/10.1007/s10530-004-0738-z)
- <span id="page-11-22"></span>Nylund GMN, Pereyra RT, Wood HL, Johannesson K (2012) Increased resistance towards generalist herbivory in the new range of a habitat-forming seaweed. Ecosphere 3:1–13. doi[:10.1890/ES12-00203.1](http://dx.doi.org/10.1890/ES12-00203.1)
- <span id="page-11-9"></span>Parker JD, Hay ME (2005) Biotic resistance to plant invasions? Native herbivores prefer non-native plants. Ecol Lett 8:959–967. doi[:10.1111/j.1461-0248.2005.00799.x](http://dx.doi.org/10.1111/j.1461-0248.2005.00799.x)
- <span id="page-11-34"></span>Parker JD, Burkepile DE, Hay ME (2006) Opposing effects of native and exotic herbivores on plant invasions. Science 311(80):1459–1461
- <span id="page-11-33"></span>Pavia H, Åberg P (1996) Spatial variation in polyphenolic content of *Ascophyllum nodosum* (Fucales, Phaeophyta). Hydrobiologia 326:199–203
- <span id="page-11-31"></span>Pavia H, Toth GB, Åberg P (2002) Optimal defense theory: elasticity analysis as a tool to predict intraplant variation in defenses. Ecology 83:891–897. doi:[10.2307/3071898](http://dx.doi.org/10.2307/3071898)
- <span id="page-11-18"></span>Pereira RC, da Gama BAP (2008) Macroalgal chemical defenses and their roles in structuring tropical marine communities. In: Amsler CD (ed) Algal chemical ecology. Springer, Berlin, Heidelberg, pp 25–49
- <span id="page-11-32"></span>Plouguerné E, Le Lann K, Connan S, Jechoux G, Deslandes E, Stiger-Pouvreau V (2006) Spatial and seasonal variation in density, reproductive status, length and phenolic content of the invasive brown macroalga *Sargassum muticum* (Yendo) Fensholt along the coast of Western Brittany (France). Aquat Bot 85:337–344
- <span id="page-11-13"></span>Plouguerné E, Hellio C, Deslandes E et al (2008) Anti-microfouling activities in extracts of two invasive algae: *Grateloupia turuturu* and *Sargassum muticum*. Bot Mar 51:202–208. doi[:10.1515/](http://dx.doi.org/10.1515/BOT.2008.026) [BOT.2008.026](http://dx.doi.org/10.1515/BOT.2008.026)
- <span id="page-11-14"></span>Plouguerné E, Ioannou E, Georgantea P et al (2010) Anti-microfouling activity of lipidic metabolites from the invasive brown alga *Sargassum muticum* (Yendo) Fensholt. Mar Biotechnol (NY) 12:52–61. doi:[10.1007/s10126-009-9199-9](http://dx.doi.org/10.1007/s10126-009-9199-9)
- <span id="page-11-29"></span>Polte P, Buschbaum C (2008) Native pipefish *Entelurus aequoreus* are promoted by the introduced seaweed *Sargassum muticum* in the northern Wadden Sea, North Sea. Aquat Biol 3:11–18. doi[:10.3354/ab00071](http://dx.doi.org/10.3354/ab00071)
- <span id="page-11-26"></span>Poore AGB, Campbell AH, Coleman RA et al (2012) Global patterns in the impact of marine herbivores on benthic primary producers. Ecol Lett 15:912–922. doi:[10.1111/j.1461-0248.2012.01804.x](http://dx.doi.org/10.1111/j.1461-0248.2012.01804.x)
- <span id="page-11-5"></span>Rejmánek M, Richardson DM (1996) What attributes make some plant species more invasive? Ecology 77:1655–1661. doi[:10.2307/2265768](http://dx.doi.org/10.2307/2265768)
- <span id="page-11-1"></span>Ribera M, Boudouresque C (1995) Introduced marine plants, with special reference to macroalgae: mechanisms and impact. Prog Phycol Res 11:187–268
- <span id="page-11-24"></span>Rohde S, Molis M, Wahl M (2004) Regulation of anti-herbivore defence by *Fucus vesiculosus* in response to various cues. J Ecol 92:1011–1018. doi:[10.1111/j.0022-0477.2004.00936.x](http://dx.doi.org/10.1111/j.0022-0477.2004.00936.x)
- <span id="page-11-12"></span>Sabour B, Reani A, Magouri HEL, Haroun R (2013) *Sargassum muticum* (Yendo) Fensholt (Fucales, Phaeophyta) in Morocco, an invasive marine species new to the Atlantic coast of Africa. Aquat Invasions 8:97–102. doi:[10.3391/ai.2013.8.1.11](http://dx.doi.org/10.3391/ai.2013.8.1.11)
- <span id="page-11-4"></span>Schaffelke B, Hewitt CL (2007) Impacts of introduced seaweeds. Bot Mar 50:397–417. doi:[10.1515/9783110211344](http://dx.doi.org/10.1515/9783110211344)
- <span id="page-11-3"></span>Schaffelke B, Smith JE, Hewitt CL (2006) Introduced macroalgae a growing concern. J Appl Phycol 18:529–541. doi[:10.1007/](http://dx.doi.org/10.1007/s10811-006-9074-2) [s10811-006-9074-2](http://dx.doi.org/10.1007/s10811-006-9074-2)
- <span id="page-11-23"></span>Schupp PJ, Paul VJ (1994) Calcium carbonate and secondary metabolites in tropical seaweeds: variable effects on herbivorous fishes. Ecology 75:1172–1185
- <span id="page-11-8"></span>Seabloom EW, Harpole WS, Reichman OJ, Tilman D (2003) Invasion, competitive dominance, and resource use by exotic and native California grassland species. Proc Natl Acad Sci USA 100:13384–13389. doi[:10.1073/pnas.1835728100](http://dx.doi.org/10.1073/pnas.1835728100)
- <span id="page-11-7"></span>Smith MD, Knapp AK (2001) Physiological and morphological traits of exotic, invasive exotic, and native plant species in tallgrass prairie. Int J Plant Sci 162:785–792. doi:[10.1086/320774](http://dx.doi.org/10.1086/320774)
- <span id="page-11-35"></span>Stæhr PA, Pedersen MF, Thomsen MS et al (2000) Invasion of *Sar‑ gassum muticum* in Limfjorden (Denmark) and its possible impact on the indigenous macroalgal community. Mar Ecol Prog Ser 207:79–88. doi[:10.3354/meps207079](http://dx.doi.org/10.3354/meps207079)
- <span id="page-11-15"></span>Tanniou A, Vandanjon L, Incera M et al (2014) Assessment of the spatial variability of phenolic contents and associated bioactivities in the invasive alga *Sargassum muticum* sampled along its European range from Norway to Portugal. J Appl Phycol 26:1–16. doi[:10.1007/s10811-013-0198-x](http://dx.doi.org/10.1007/s10811-013-0198-x)
- <span id="page-11-25"></span>Targett NM, Arnold TM (1998) Predicting the effects of brown algal phlorotannins on marine herbivores in tropical and temperate oceans. J Phycol 34:195–205. doi[:10.1046/j.1529-8817.1998.340195.x](http://dx.doi.org/10.1046/j.1529-8817.1998.340195.x)
- <span id="page-11-2"></span>Thresher RE (1999) Key threats from marine bioinvasions: a review of current and future issues. In: Pederson J (ed) Marine bioinvasions: proceedings of the first national conference. Massachussetts Institute of Technology, Sea Grant College Program, Boston, pp 24–36
- <span id="page-11-21"></span>Toth GB, Karlsson M, Pavia H (2007) Mesoherbivores reduce net growth and induce chemical resistance in natural seaweed populations. Oecologia 152:245–255. doi[:10.1007/](http://dx.doi.org/10.1007/s00442-006-0643-5) [s00442-006-0643-5](http://dx.doi.org/10.1007/s00442-006-0643-5)
- <span id="page-11-28"></span>Trowbridge CD, Hirano YM, Hirano YJ (2009) Interaction webs of marine specialist herbivores on Japanese shores. J Mar Biol Assoc United Kingdom 89:277. doi[:10.1017/](http://dx.doi.org/10.1017/S002531540900318X) [S002531540900318X](http://dx.doi.org/10.1017/S002531540900318X)
- <span id="page-12-4"></span>van Alstyne KL (1995) Comparison of three methods for quantifying brown algal polyphenolic compounds. J Chem Ecol 21:45–58. doi[:10.1007/BF02033661](http://dx.doi.org/10.1007/BF02033661)
- <span id="page-12-9"></span>van Kleunen M, Weber E, Fischer M (2010) A meta-analysis of trait differences between invasive and non-invasive plant species. Ecol Lett 13:235–245. doi:[10.1111/j.1461-0248.2009.01418.x](http://dx.doi.org/10.1111/j.1461-0248.2009.01418.x)
- <span id="page-12-5"></span>Vas'kovskii VE, Isai SV (1972) Determination of the amount of mannitol in brown seaweeds. Chem Nat Compd 8:596–600. doi[:10.1007/BF00564297](http://dx.doi.org/10.1007/BF00564297)
- <span id="page-12-6"></span>Vermeij MJA, Smith TB, Dailer ML, Smith CM (2009) Release from native herbivores facilitates the persistence of invasive marine algae: a biogeographical comparison of the relative contribution of nutrients and herbivory to invasion success. Biol Invasions 11:1463–1474. doi:[10.1007/s10530-008-9354-7](http://dx.doi.org/10.1007/s10530-008-9354-7)
- <span id="page-12-0"></span>Vitousek PM, Dantonio CM, Loope LL et al (1997) Introduced species: a significant component of human-caused global change. N Z J Ecol 21:1–16
- <span id="page-12-2"></span>Weinberger F, Buchholz B, Karez R, Wahl M (2008) The invasive red alga *Gracilaria vermiculophyll*a in the Baltic Sea: adaptation to brackish water may compensate for light limitation. Aquat Biol 3:251–264. doi:[10.3354/ab00083](http://dx.doi.org/10.3354/ab00083)
- <span id="page-12-7"></span>Weinberger F, Rohde S, Oschmann Y et al (2011) Effects of limitation stress and of disruptive stress on induced antigrazing defense in the bladder wrack *Fucus vesiculosus*. Mar Ecol Prog Ser 427:83–94. doi:[10.3354/meps09044](http://dx.doi.org/10.3354/meps09044)
- <span id="page-12-10"></span>Wernberg T, Thomsen MS, Stæhr PA, Pedersen MF (2000) Comparative phenology of *Sargassum muticum* and *Halidrys siliquosa* (Phaeophyceae: Fucales) in Limfjorden, Denmark. Bot Mar 44:31–39. doi:[10.1515/BOT.2001.005](http://dx.doi.org/10.1515/BOT.2001.005)
- <span id="page-12-1"></span>Wikström SA, Steinarsdóttir MB, Kautsky L, Pavia H (2006) Increased chemical resistance explains low herbivore colonization of introduced seaweed. Oecologia 148:593–601
- <span id="page-12-8"></span>Yendo K (1907) The Fucaceae of Japan. J Coll Sci Tokyo Imp Univ 21(12):1–174
- <span id="page-12-3"></span>Zhang Q, Zhang J, Shen J et al (2006) A Simple 96-well microplate method for estimation of total polyphenol content in seaweeds. J Appl Phycol 18:445–450. doi[:10.1007/s10811-006-9048-4](http://dx.doi.org/10.1007/s10811-006-9048-4)
- <span id="page-12-11"></span>Zou D (2005) Effects of elevated atmospheric CO2 on growth, photosynthesis and nitrogen metabolism in the economic brown seaweed, *Hizikia fusiforme* (Sargassaceae, Phaeophyta). Aquaculture 250:726–735. doi:[10.1016/j.aquaculture.2005.05.014](http://dx.doi.org/10.1016/j.aquaculture.2005.05.014)