SHORT NOTE

Large‑scale spatial variation in epilithic algal matrix cryptofaunal assemblages on the Great Barrier Reef

M. J. Kramer · D. R. Bellwood · O. Bellwood

Received: 7 February 2014 / Accepted: 3 July 2014 / Published online: 19 July 2014 © Springer-Verlag Berlin Heidelberg 2014

Abstract Spatial variation in the epilithic algal matrix (EAM) cryptofauna was investigated at three locations on the Great Barrier Reef: two inner shelf—Orpheus Island and the Turtle Island group—and a mid-shelf location, Lizard Island. Although the EAM appears to be a relatively simple and consistent habitat, significant differences in cryptofaunal assemblages were found between locations. EAM assemblages from Orpheus Island were markedly different. This appears to be a function of the sediment profile characteristics (grain size >60 µm) at Orpheus Island, as many cryptofaunal taxa displayed positive relationships with sediment volume. However, sediment volumes did not differ significantly between the three locations, highlighting the possibility of cyclonic activity affecting the sediment profile at Orpheus Island in the months preceding the study, in addition to the nutrient input from major terrigenous sources. The results of this study show that EAM cryptofaunal assemblages are not uniform across the Great Barrier Reef and suggest that dissolved nutrients, sediment loads and distance from river systems may be significant drivers of cryptobenthic faunal compositions.

Communicated by M. Huettel.

M. J. Kramer $(\boxtimes) \cdot$ D. R. Bellwood \cdot O. Bellwood School of Marine and Tropical Biology, James Cook University, Townsville, QLD 4811, Australia e-mail: michael.kramer@my.jcu.edu.au

M. J. Kramer · D. R. Bellwood Australian Research Council Centre of Excellence for Coral Reef Studies, James Cook University, Townsville, QLD 4811, Australia

Introduction

Investigations of marine assemblages across large spatial scales often reveal distinct patterns of species distributions and community structure, particularly in latitudinal or crossshelf studies (Stevens [1989;](#page-7-0) Bellwood and Wainwright [2001](#page-6-0); Wismer et al. [2009](#page-7-1)). Spreading nearly 2,000 km along the coastline and up to 200 km offshore, Australia's Great Barrier Reef shows clear divisions between inshore, mid- and offshore reefs. This pattern holds true for a wide variety of organisms, ranging from fishes (Bellwood and Wainwright [2001;](#page-6-0) Hoey and Bellwood [2008\)](#page-6-1) and benthic macroalgae (Wismer et al. [2009](#page-7-1)), to crustose coralline algae (Fabricius and De'ath [2001\)](#page-6-2) and crustaceans (Preston and Doherty [1994\)](#page-7-2).

Factors that may drive the divisions in the cross-shelf distributions include wave energy (Bellwood and Wainwright [2001\)](#page-6-0), benthic composition (Wismer et al. [2009](#page-7-1)), the availability of nutrients (Uthicke and Nobes [2008](#page-7-3); Uthicke and Altenrath [2010\)](#page-7-4), turbidity (Cooper et al. [2007](#page-6-3)), and sedimentation (Preston and Doherty [1994;](#page-7-2) McCook [1996](#page-7-5); Fabricius and De'ath [2001](#page-6-2)). Relatively high rates of sedimentation on inner shelf reefs appear to be particularly influential on benthic organisms (McCook [1996](#page-7-5); Fabricius and De'ath [2001;](#page-6-2) McCulloch et al. [2003](#page-7-6)). For example, in high sedimentation locations, crustose coralline algae cover is low (Fabricius and De'ath [2001](#page-6-2)) and crustacean communities in dead corals have distinctly different assemblages and lower abundances (Preston and Doherty [1994](#page-7-2)). Sediment, per se, does little to explain the large-scale community structure of Foraminifera, however, turbidity and nutrient profiles appear to have a much greater influence on these benthic organisms (Cooper et al. [2007](#page-6-3); Uthicke and Nobes [2008](#page-7-3); Uthicke and Altenrath [2010](#page-7-4)).

The epilithic algal matrix (EAM) is a significant, and often dominant, component of benthic assemblages on coral reefs, covering approximately 30–70 % of the available surface area (Klumpp and McKinnon [1992;](#page-6-4) Goatley and Bellwood [2011](#page-6-5)). Within the EAM, short filamentous and crustose algal species are common (Scott and Russ [1987](#page-7-7); Connell et al. [2014\)](#page-6-6) in addition to nutrient-rich detritus (Crossman et al. [2001](#page-6-7); Wilson et al. [2003\)](#page-7-8), inorganic sediments and a community of small organisms (Kramer et al. [2012](#page-6-8)). Once thought to be exploited solely by herbivorous fishes, the EAM has progressively gained recognition as being a valuable resource for detritivorous (Wilson et al. [2003](#page-7-8), [2004](#page-7-9)) and small carnivorous fishes (Kramer et al. [2013](#page-6-9)). Although the EAM is structurally simple, based on a short algal turf ranging from 1 to 6 mm in height (Bonaldo and Bellwood [2011\)](#page-6-10) it supports a surprisingly diverse and abundant community of motile taxa (Kramer et al. [2012\)](#page-6-8) that are likely to be important contributors to ecosystem function (Kennedy and Jacoby [1999;](#page-6-11) Schratzberger et al. [2000](#page-7-10)). In the present study, the term 'cryptofauna' will be used to include both meiofauna (0.06–0.5 mm; Giere [2009\)](#page-6-12) and larger taxa (up to 5 mm).

Environmental factors are capable of having both positive and negative effects on the abundance of organisms and the overall structure of a community. One such factor is the effect of sediments, which consist of both organic and inorganic settled particulate matter. The amount of sediment is particularly important to coral reefs, as high levels are known to place corals under metabolic stress (Rogers [1990\)](#page-7-11), inhibit coral settlement (Birrell et al. [2005](#page-6-13)) and suppress grazing by herbivorous fishes (Bellwood and Fulton [2008](#page-6-14)). The influence of sediment and dissolved nutrients on the abundance of small organisms is relatively well known (Preston and Doherty [1994](#page-7-2); Cooper et al. [2007](#page-6-3); Uthicke and Altenrath [2010;](#page-7-4) Kramer et al. [2012\)](#page-6-8). However, whilst the abundance of cryptofauna is reported to remain relatively temporally consistent (Klumpp et al. [1988](#page-6-15); Logan et al. [2008\)](#page-7-12) not much is known about spatial variability. The present study aims to investigate the spatial differences in cross- and alongshelf variation of EAM cryptofauna using two inner shelf locations and a typical mid-shelf location on the GBR. In addition, sediment volumes were measured and proximity to major river systems was determined to examine the relationship between sediment, nutrients and EAM cryptofauna at these locations.

Methods

Sampling locations

Samples of EAM fauna were collected from three locations across the Great Barrier Reef: Orpheus Island, the Turtle Island group and Lizard Island (Fig. [1\)](#page-2-0). Orpheus Island is located on the inner Great Barrier Reef (18° 36′40″S, 146° 29′20″E), 16 km from the mainland. Samples were collected from four sites across the reef of Pioneer Bay, Orpheus Island. A detailed description of Pioneer Bay habitats is given in Fox and Bellwood ([2007\)](#page-6-16). The Turtle Island group is similarly located on the inner-shelf of the GBR, 12 km offshore and approximately 450 km north of Orpheus Island (14° 43′55″S, 145° 11′00″). Composed of nine small islands and reefs, two islands (2 sites each) were sampled for EAM cryptofauna. Lizard Island is located on the mid-shelf GBR (14° 40′40″S, 145° 26′ 55″E) 30 km offshore and on similar latitude to the Turtle Island group. Samples were collected from the reef in Mermaid Cove and the Lagoon Entrance (2 sites each). Sampling at all locations was conducted during the austral summer months (November–March) to reduce variation due to seasonal effects, although seasons usually have a minimal influence on cryptofaunal populations (Klumpp et al. [1988](#page-6-15); Logan et al. [2008\)](#page-7-12). For consistency, sampling at all locations was conducted from similar depths (2–3 m depth at high tide) on the fringing reef crest of the leeward shore where live coral and EAM were dominant and macroalgae sparse. All locations are marine protected areas where fishing is prohibited and removal of organisms is only granted for approved research. As such, the marine communities are considered to be relatively intact and include all major functional groups of herbivorous fishes (Bellwood et al. [2004](#page-6-17); Green and Bellwood [2009\)](#page-6-18), thereby subjecting the EAM to similar grazing pressure. The EAM at all sites was of a similar height (4–6 mm) and morphology (short, filamentous Chlorophyta and Rhodophyta). Samples were taken from open planar areas away from territorial damselfish territories that may modify EAM composition (Klumpp et al. [1988](#page-6-15)). Sampling on Orpheus Island occurred 1 month after a tropical cyclone (Cyclone Yasi, Category 5) passed over the island (Great Barrier Reef Marine Park Authority [2011](#page-6-19)) and 3 months after extensive coastal flooding due to Cyclone Tasha (Hayes and Goonetilleke [2012](#page-6-20)).

Sample collection

Samples from all locations were collected using SCUBA from the reef crest zone. A total of 60 samples was obtained, consisting of five individual samples from each of the four sites at each of the three locations. An underwater vacuum apparatus based on the design described in Kramer et al. ([2012\)](#page-6-8) was utilised to remove all particulate material within a defined area. The sampling area was delimited by a section of PVC pipe 51 mm in diameter, representing 20.4 cm² of EAM. The sampling area was vacuumed thoroughly for 30 s, during which time the resident organisms and particulate matter were drawn into the apparatus and retained by a 60 µm plankton mesh filter bag. The filter bag

Fig. 1 Sampling locations. **a** Queensland, Australia, indicating regions where sampling was conducted. **b** Orpheus Island. **c** Turtle Island group. **d** Northern sampling locations: Turtle Island group and

was then sealed, transported to the surface and the contents fixed in 4 % formaldehyde solution in seawater.

Sample and data analysis

Samples were stained with eosin erythrosin to aid in distinguishing organisms amongst the particulate matter. Collected material was washed onto a petri dish, which had a grid affixed to the bottom to prevent observing the same organism twice. Samples were examined using an Olympus SZ40 binocular microscope under $40\times$ magnification to identify taxa to the lowest functional taxonomic level (usually order). This level of taxonomic resolution was deemed appropriate for the purpose of detecting spatial differences

Lizard Island. **e** Lizard Island. *Dashed lines* indicate fringing reefs and stars are sampling sites. Two sites were sampled at *each star*

in the EAM fauna and comparing overall community composition (cf. Beattle and Oliver [1994;](#page-6-21) Krell [2004](#page-6-22)). The abundance of the respective organisms was recorded and standardised to 100 cm^2 . The sediment volume of each sample was calculated following Kramer et al. [\(2012](#page-6-8)). Using digital vernier calipers (accuracy: ± 0.02 mm), the depth of the settled particulates within the sample vial was measured and the volume estimated based on a calibrated vial.

The dataset used for the analysis incorporated abundant organisms and excluded those taxa with fewer than 10 individuals 100 cm−² . Thus, taxa considered in the analysis were Amphipoda, Cumacea, Gastropoda, Harpacticoida, Isopoda, Polychaeta, Ostracoda and Tanaidacea. Organisms

that were observed, but not included in the analysis due to low abundances, were Chaetognatha, Chironomida, Cumacea and Decapoda. The cryptofaunal community data were explored using a non-metric multidimensional scaling (nMDS) ordination of a Manhattan distance matrix based on standardised data, thus representing proportions rather than raw abundance and preventing the over-representation of abundant taxa. Taxa vectors (based on maximum correlation with taxon variables) were added to the ordination to aid interpretation of the data (Oksanen et al. [2013\)](#page-7-13). A permutational multivariate analysis of variance (PER-MANOVA) was conducted to elucidate differences in sites and locations. Spearman rank correlations were conducted to investigate the relationship between sediment and the abundance of organisms. Additionally, a one-way ANOVA was used to test the difference in sediment volume among locations. Multivariate analyses, Spearman rank correlations and ANOVA were conducted using the R packages vegan (Oksanen et al. [2013\)](#page-7-13) and Hmisc (Harrell [2014](#page-6-23)). Pairwise comparisons were calculated using Primer 6 with PERMANOVA.

Results

PERMANOVA analysis found no significant difference in cryptofaunal assemblages among sites within locations, or in the interaction between sites and locations. Site data were therefore pooled within locations for the remainder of the analyses. Investigation of the nMDS revealed that each location could be separated into distinct ordination groupings. This is supported by the PERMANOVA, which indicates that locations were statistically significant (PERMANOVA, *P* < 0.001). The pairwise PERMANOVA

further indicated that all locations were significantly different from each other (pairwise PERMANOVA, *P* < 0.05). From the vectors fitted to the nMDS ordination (Fig. [2](#page-3-0)), Orpheus Island showed strong gradients for Polychaeta and Gastropoda, whereas the Turtle Islands and Lizard Island are represented by a gradient towards the Harpacticoida. Vectors also indicate that Amphipoda, Cumacea and Tanaidacea exhibit strong gradients toward the Turtle Islands. Although Orpheus Island did not exhibit any shared multivariate space with the other locations, the Turtle Islands and Lizard Island did have a small degree of overlap, indicating that some sampling locations between these two regions were somewhat similar in EAM community composition.

Data were further explored by investigating the relationship between sediment and the abundance of organisms in the EAM. The only taxon not to have a significant correlation with sediment was Isopoda (Spearman rank correlation, $r_s = 0.125$, $N = 60$, $P = 0.340$). All other taxa had a significant, positive, relationship with sediment, ranging from $(r_s = 0.571, N = 60, P < 0.001)$ to $(r_s = 0.280,$ $N = 60$, $P = 0.03$) for Polychaeta and Amphipoda, respectively (Fig. [3](#page-4-0)). However, sediment volumes among locations revealed no significant difference in the volume of sediment at each location (ANOVA, $F_{(2, 57)} = 1.819$, $P = 0.171$.

Discussion

Cross-shelf studies on the GBR have consistently revealed a distinct separation between inner, mid and outer shelf reefs. This pattern applies to community assemblages (Done [1982](#page-6-24); Preston and Doherty [1994;](#page-7-2) Bellwood and Wainwright [2001](#page-6-0)), benthic composition (McCook [1996](#page-7-5);

Dimension 1

Fig. 2 Non-metric multidimensional scaling (nMDS) ordination of epilithic algal matrix fauna from Orpheus Island (*dashed line*), Lizard Island (*dotted line*) and Turtle Island group (*solid line*). *Vectors* indicate strength and direction of taxa gradients. All locations are significantly different (pairwise PERMANOVA, *P* < 0.05)

Fig. 3 Spearman rank correlations of the relationships between sediment volume and abundance of cryptofaunal organisms. Correlation coefficients and statistical significance values for each relationship are inserted onto the respective correlations

Fabricius and De'ath [2001;](#page-6-2) Wismer et al. [2009\)](#page-7-1) and ecosystem processes (Russ and McCook [1999;](#page-7-14) Hoey and Bellwood [2008](#page-6-1)). The preliminary observations in the present study suggest that there is not only a cross-shelf difference in EAM cryptofauna composition, but also a distinct along-shelf separation. Somewhat surprisingly, the nMDS suggests that an inner shelf reef (Orpheus Island) differs more from another inner shelf location (Turtle Island group) than the Turtle Islands differ from a nearby mid-shelf system (Lizard Island). This result contrasts with other cross-shelf studies, which typically report that features such as hydrodynamics (Bellwood and Wainwright [2001](#page-6-0)), sediments (Preston and Doherty [1994;](#page-7-2) Fabricius and De'ath [2001](#page-6-2)), abundance of macroalgae (Wismer et al. [2009](#page-7-1)), or a combination of the above (Hoey and Bellwood [2008](#page-6-1)), drive the differences across large spatial scales. The samples of the EAM collected in the present study were taken from largely similar environments, thereby controlling for the effect of depth, reef zone, season, proximity of macroalgae and hydrodynamics. Yet, the EAM cryptofauna differed significantly among all locations.

Whilst these results were unexpected, there are indications that this variation in EAM cryptofauna may be a result of the water quality and nature of sediments at each location. The coastline adjacent to the sampling locations is subject to tropical seasonal fluctuations, namely a dry season and a wet season. During the wet season, extensive flooding is common, where large amounts of terrigenous sediments and anthropogenic contaminants are flushed into major rivers and out into coastal waters (Devlin and Brodie [2005](#page-6-25); Brodie et al. [2010\)](#page-6-26). The resulting sediment plumes typically move in a northerly direction along the Queensland coastline (McCulloch et al. [2003](#page-7-6); Brodie et al. [2010](#page-6-26)). The catchments that are most likely to affect the locations in the present study are the Mossman–Daintree and a small section of the Northeastern Cape York catchment (Turtle Island group and Lizard Island) and the Burdekin-Haughton and Ross-Black catchments (Orpheus Island) (Neil et al. [2002;](#page-7-15) Bainbridge et al. [2012](#page-6-27)). It is important to note the difference in catchment areas that deliver floodwaters into the GBR lagoon. River catchments that affect the Turtle Island group and Lizard Island are much smaller than those that influence Orpheus Island (Neil et al. [2002](#page-7-15)). In addition, of all the rivers along the Queensland coast, the Burdekin River produces the highest volume of suspended sediments to the GBR lagoon, emitting an estimated 3×10^8 kg year⁻¹ (Neil et al. [2002](#page-7-15); McCulloch et al. [2003](#page-7-6); Kroon et al. [2012\)](#page-6-28). Although Orpheus Island is not in the direct path of the flood plume stemming from the Burdekin River, excessive nutrients and (to a lesser extent) suspended sediments from the Burdekin River have been documented at considerable distances that easily extend past, and encompass, Orpheus Island (Devlin and Brodie [2005](#page-6-25); Devlin et al. [2008](#page-6-29); Bainbridge et al. [2012\)](#page-6-27).

In the present study, although almost all taxa displayed significant positive relationships with sediment volume, four taxa in particular appear to be influenced most strongly: Gastropoda, Harpacticoida, Ostracoda and Polychaeta. Of these, Gastropoda and Polychaeta were characteristic of Orpheus Island, whereas Harpacticoida and Ostracoda were dominant taxa at the Turtle Islands and at Lizard Island. The major trophic role of these four taxa is primary consumption (Ruppert et al. [2004](#page-7-16)), thus it would be expected that an increase in primary productivity due to nutrient input would increase populations (Montagna et al. [1995\)](#page-7-17). It is likely that the sediment composition and nutrient profile created by local terrigenous inputs (i.e. flood plumes from the Burdekin-Haughton and Ross-Black catchments) produces a distinctive EAM environment at Orpheus Island. Furthermore, it appears that Harpacticoida and Ostracoda are abundant EAM taxa in all locations, therefore it is the paucity of Gastropoda and Polychaeta at the Turtle Islands and Lizard Island locations that most clearly drove the distinctly different ordination groups. Future analyses that incorporate a finer taxonomic resolution, particularly of these four Orders (Gastropoda, Harpacticoida, Ostracoda and Polychaeta), may provide greater detail in regards to how families, genera or species respond to specific environmental conditions, or how the abundance of specific trophic categories may vary.

Although the sediment volumes were not significantly different among locations, it is likely that it was the composition (i.e. particle size or nutrient profiles) rather than volume, per se, that most heavily influenced the abundance of certain taxa and thus, community composition. In this regard, the effect of cyclonic activity on the GBR may also be important. Cyclones subject the reef to substantial hydrodynamic action, causing resuspension and flushing of sediment from the reef, especially in shallow waters (Wolanski et al. [2005\)](#page-7-18). As the majority of the fringing reef of Orpheus Island is less than 10 m depth, extensive resuspension and removal of sediment from the reef crest during cyclones is likely. Additionally, extensive flooding is often associated with cyclones, which increases the amount of terrigenous material being flushed out into the GBR lagoon. There is a strong possibility that the sediment profile at Orpheus Island observed in the present study was influenced by the combined effect of Cyclone Tasha's coastal flooding (December 2010) and Cyclone Yasi's wind-generated waves (February 2011). Nutrients and sediments previously flushed into the GBR lagoon may have been resuspended and re-released by the cyclonic events (Gagan et al. [1987](#page-6-30), [1990;](#page-6-31) Russ and McCook [1999](#page-7-14)), causing an increase in the availability of primary producers and subsequently the abundance of primary consumers in the EAM cryptofauna at Orpheus Island. Furthermore, sediments that were previously established within the EAM may have been relocated due to major wave action (Wolanski et al. [2005](#page-7-18)). Physical disturbance of the cryptofauna itself is not expected to have a great impact on the populations, as these small organisms are known to recover to pre-disturbance abundances in <24 h (Sherman and Coull [1980](#page-7-19); Johnson et al. [2007\)](#page-6-32).

While the effect of sediment and dissolved nutrients on corals, algae and cryptofauna are well documented on faunal communities, knowledge of the impact of sediment and nutrients on the EAM is in its infancy. The apparent sensitivity of most cryptofauna to sediment emphasises the

potential of natural and human-induced modification of sediment and water quality to impact coastal ecosystems. Although the EAM is increasingly recognised as an important contributor to reef processes in terms of productivity of detritus and algae (Klumpp and McKinnon [1992](#page-6-4); Wilson et al. [2003](#page-7-8); Bonaldo and Bellwood [2011\)](#page-6-10), the present study provides the first, preliminary, account of the cross- and along-shelf variation in the cryptofauna of the EAM. Data presented herein provides a baseline by which to compare the condition of EAM communities in the respective locations. The present study indicates that EAM cryptofaunas are not uniform across the GBR and that particulate and dissolved nutrient input, sediment loads and position relative to catchment areas may be more important than shelf position in shaping cryptobenthic communities.

Acknowledgments We wish to thank the staff of Orpheus Island and Lizard Island research stations, S Brandl, C Mirbach, J Rizzari and J Welsh for field support and two anonymous reviewers for helpful comments on earlier drafts. This work was supported by the Australian Research Council (D.R.B.).

References

- Bainbridge ZT, Wolanski E, Álvarez-Romero JG, Lewis SE, Brodie JE (2012) Fine sediment and nutrient dynamics related to particle size and floc formation in a Burdekin River flood plume, Australia. Mar Pollut Bull 65:236–248
- Beattle AJ, Oliver I (1994) Taxonomic minimalism. Trends Ecol Evol 9:488–490
- Bellwood DR, Fulton CJ (2008) Sediment-mediated suppression of herbivory on coral reefs: decreasing resilience to rising sea levels and climate change? Limnol Oceanogr 53:2695–2701
- Bellwood DR, Wainwright P (2001) Locomotion in labrid fishes: implications for habitat use and cross-shelf biogeography on the Great Barrier Reef. Coral Reefs 20:139–150
- Bellwood DR, Hughes TP, Folke C, Nystrom M (2004) Confronting the coral reef crisis. Nature 429:827–833
- Birrell CL, McCook LJ, Willis BL (2005) Effects of algal turfs and sediment on coral settlement. Mar Pollut Bull 51:408–414
- Bonaldo R, Bellwood D (2011) Spatial variation in the effects of grazing on epilithic algal turfs on the Great Barrier Reef, Australia. Coral Reefs 30:381–390
- Brodie J, Schroeder T, Rohde K, Faithful J, Masters B, Dekker A, Brando V, Maughan M (2010) Dispersal of suspended sediments and nutrients in the Great Barrier Reef lagoon during river-discharge events: conclusions from satellite remote sensing and concurrent flood-plume sampling. Mar Fresh Res 61:651–664
- Connell SD, Foster MS, Airoldi L (2014) What are algal turfs? Towards a better description of turfs. Mar Ecol Prog Ser 495:299–307
- Cooper TF, Uthicke S, Humphrey C, Fabricius KE (2007) Gradients in water column nutrients, sediment parameters, irradiance and coral reef development in the Whitsunday Region, central Great Barrier Reef. Estuar Coast Shelf Sci 74:458–470
- Crossman DJ, Choat JH, Clements KD, Hardy T, Jason M (2001) Detritus as food for grazing fishes on coral reefs. Limnol Oceanogr 46:1596–1605
- Devlin MJ, Brodie J (2005) Terrestrial discharge into the Great Barrier Reef Lagoon: nutrient behavior in coastal waters. Mar Pollut Bull 51:9–22
- Devlin M, Brodie J, Bainbridge Z, Lewis S (2008) Flood plumes in the GBR: the Burdekin and Fitzroy flood plumes, 2007/08—case studies for marine monitoring program. Australian Centre for Tropical Freshwater Research, James Cook University, Townsville
- Done TJ (1982) Patterns in the distribution of coral communities across the central Great Barrier Reef. Coral Reefs 1:95–107
- Fabricius K, De'ath G (2001) Environmental factors associated with the spatial distribution of crustose coralline algae on the Great Barrier Reef. Coral Reefs 19:303–309
- Fox RJ, Bellwood DR (2007) Quantifying herbivory across a coral reef depth gradient. Mar Ecol Prog Ser 339:49–59
- Gagan M, Sandstrom M, Chivas A (1987) Restricted terrestrial carbon input to the continental shelf during Cyclone Winifred: implications for terrestrial runoff to the Great Barrier Reef Province. Coral Reefs 6:113–119
- Gagan MK, Chivas AR, Herczeg AL (1990) Shelf-wide erosion, deposition, and suspended sediment tranport during Cyclone Winifred, central Great Barrier Reef, Australia. J Sediment Petrol 60:456–470
- Giere O (2009) Meiobenthology: the microscopic motile fauna of aquatic sediments. Springer, Berlin
- Goatley CHR, Bellwood DR (2011) The roles of dimensionality, canopies and complexity in ecosystem monitoring. PLoS One 6:e27307
- Great Barrier Reef Marine Park Authority (2011) Impacts of tropical cyclone Yasi on the Great Barrier Reef: a report on the findings of a rapid ecological impact assessment, July 2011, GBRMPA, Townsville
- Green AL, Bellwood DR (2009) Monitoring functional groups of herbivorous reef fishes as indicators of coral reef resilience—a practical guide for coral reef managers in the Asia Pacific region. In: IUCN working group on climate change and Coral Reefs. IUCN, Gland, Switzerland
- Harrell FE (2014) The Hmisc Package. R package version 3.14-4. <http://cran.r-project.org/web/packages/Hmisc/>
- Hayes J, Goonetilleke A (2012) Building community resilience learning from the 2011 floods in Southeast Queensland, Australia. In: Kakimoto R, Yamada F (eds) 8th annual conference of international institutes for infrastructure, renewal and reconstruction: international conference on disaster management, Kumamoto University, Kumamoto, Japan pp 51–60
- Hoey A, Bellwood D (2008) Cross-shelf variation in the role of parrotfishes on the Great Barrier Reef. Coral Reefs 27:37–47
- Johnson GEL, Attrill MJ, Sheehan EV, Somerfield PJ (2007) Recovery of meiofauna communities following mudflat disturbance by trampling associated with crab-tiling. Mar Environ Res 64:409–416
- Kennedy A, Jacoby C (1999) Biological indicators of marine environmental health: meiofauna—a neglected benthic component? Environ Monit Assess 54:47–68
- Klumpp DW, McKinnon AD (1992) Community structure, biomass and productivity of epilithic algal communities on the Great Barrier Reef: dynamics at different spatial scales. Mar Ecol Prog Ser 86:77–89
- Klumpp DW, McKinnon AD, Mundy CN (1988) Motile cryptofauna of a coral reef—abundance, distribution and trophic potential. Mar Ecol Prog Ser 45:95–108
- Kramer MJ, Bellwood DR, Bellwood O (2012) Cryptofauna of the epilithic algal matrix on an inshore coral reef, Great Barrier Reef. Coral Reefs 31:1007–1015
- Kramer MJ, Bellwood O, Bellwood DR (2013) The trophic importance of algal turfs for coral reef fishes: the crustacean link. Coral Reefs 32:575–583
- Krell F-T (2004) Parataxonomy versus taxonomy in biodiversity studies—pitfalls and applicability of 'morphospecies' sorting. Biodivers Conserv 13:795–812
- Kroon FJ, Kuhnert PM, Henderson BL, Wilkinson SN, Kinsey-Henderson A, Abbott B, Brodie JE, Turner RDR (2012) River loads of

suspended solids, nitrogen, phosphorus and herbicides delivered to the Great Barrier Reef lagoon. Mar Pollut Bull 65:167–181

- Logan D, Townsend KA, Townsend K, Tibbetts IR (2008) Meiofauna sediment relations in leeward slope turf algae of Heron Island reef. Hydrobiologia 610:269–276
- McCook LJ (1996) Effects of herbivores and water quality on *Sargassum* distribution on the central Great Barrier Reef: cross-shelf transplants. Mar Ecol Prog Ser 139:179–192
- McCulloch M, Fallon S, Wyndham T, Hendy E, Lough J, Barnes D (2003) Coral record of increased sediment flux to the inner Great Barrier Reef since European settlement. Nature 421:727–730
- Montagna PA, Blanchard GF, Dinet A (1995) Effect of production and biomass of intertidal microphytobenthos on meiofaunal grazing rates. J Exp Mar Biol Ecol 185:149–165
- Neil DT, Orpin AR, Ridd PV, Yu B (2002) Sediment yield and impacts from river catchments to the Great Barrier Reef lagoon: a review. Mar Fresh Res 53:733–752
- Oksanen J, Blanchet FG, Kindt R, Legendre P, Minchin PR, O'Hara RB, Simpson GL, Peter Solymos M, Stevens HH, Wagner H (2013) Vegan: community ecology package. R package version 2.0-7. <http://cran.r-project.org/web/packages/vegan/index.html>
- Preston NP, Doherty PJ (1994) Cross-shelf patterns in the community structure of coral-dwelling crustacea in the central region of the Great Barrier Reef II. cryptofauna. Mar Ecol Prog Ser 104:27–38
- Rogers CS (1990) Responses of coral reefs and reef organisms to sedimentation. Mar Ecol Prog Ser 62:185–202
- Ruppert EE, Fox RS, Barnes RD (2004) Invertebrate zoology: a functional evolutionary approach. Brooks/Cole, Belmont
- Russ GR, McCook LJ (1999) Potential effects of a cyclone on benthic algal production and yield to grazers on coral reefs across the central Great Barrier Reef. J Exp Mar Biol Ecol 235:237–254
- Schratzberger M, Gee JM, Rees HL, Boyd SE, Wall CM (2000) The structure and taxonomic composition of sublittoral meiofauna assemblages as an indicator of the status of marine environments. J Mar Biol Assoc UK 80:969–980
- Scott FJ, Russ GR (1987) Effects of grazing on species composition of the epilithic algal community on coral reefs of the central Great Barrier Reef. Mar Ecol Prog Ser 39:293–304
- Sherman KM, Coull BC (1980) The response of meiofauna to sediment disturbance. J Exp Mar Biol Ecol 46:59–71
- Stevens GC (1989) The latitudinal gradient in geographical range: how so many species coexist in the tropics. Am Nat 133:240–256
- Uthicke S, Altenrath C (2010) Water column nutrients control growth and C: N ratios of symbiont-bearing benthic foraminifera on the Great Barrier Reef, Australia. Limnol Oceanogr 55:1681–1696
- Uthicke S, Nobes K (2008) Benthic Foraminifera as ecological indicators for water quality on the Great Barrier Reef. Estuar Coast Shelf Sci 78:763–773
- Wilson SK (2004) Growth, mortality and turnover rates of a small detritivorous fish. Mar Ecol Prog Ser 284:253–259
- Wilson SK, Bellwood DR, Choat JH, Furnas MJ (2003) Detritus in the epilithic algal matrix and its use by coral reef fishes. Oceanogr Mar Biol Annu Rev 41:279–309
- Wismer S, Hoey AS, Bellwood DR (2009) Cross-shelf benthic community structure on the Great Barrier Reef: relationships between macroalgal cover and herbivore biomass. Mar Ecol Prog Ser 376:45–54
- Wolanski E, Fabricius K, Spagnol S, Brinkman R (2005) Fine sediment budget on an inner-shelf coral-fringed island, Great Barrier Reef of Australia. Estuar Coast Shelf Sci 65:153–158