#### **REVIEW**



# **Role of Vitamin K in Bone and Muscle Metabolism**

**N. Alonso1 · A. Meinitzer1 · E. Fritz‑Petrin1 · D. Enko<sup>1</sup> · M. Herrmann1**

Received: 10 December 2021 / Accepted: 26 January 2022 / Published online: 12 February 2022 © The Author(s) 2022

#### **Abstract**

Vitamin K, a cofactor for the  $\gamma$ -glutamyl carboxylase enzyme, is required for the post-translational activation of osteocalcin and matrix Gla protein, which play a key role in bone and muscle homeostasis. In vivo and in vitro models for osteoporosis and sarcopenia suggest the vitamin K could exert a positive efect in both conditions. In bone, it increases osteoblastogenesis, whilst decreases osteoclast formation and function. In muscle, it is associated with increased satellite cell proliferation and migration and might play a role in energy metabolism. Observational trials suggest that high levels of vitamin K are associated with increased bone mineral density and reduced fracture risk. However, interventional studies for vitamin K supplementation yielded conficting results. Clinical trials in sarcopenia suggest that vitamin K supplementation could improve muscle mass and function. One of the main limitations on the vitamin K studies are the technical challenges to measure its levels in serum. Thus, they are obtained from indirect sources like food questionnaires, or levels of undercarboxylated proteins, which can be afected by other environmental or biological processes. Although current research appoints to a benefcial efect of vitamin K in bone and muscle, further studies overcoming the current limitations are required in order to incorporate this supplementation in the clinical management of patients with osteosarcopenia.

**Keywords** Vitamin K · Vitamin K metabolites · Osteoporosis · Sarcopenia · Mass spectrometry

## **Introduction**

Osteoporosis is a complex disease characterized by low bone mineral density and alterations in the bone quality, which leads to the appearance of fractures. Bone mineral density is assessed by Dual-energy X-ray absorptiometry (DXA) scan and patients with osteoporosis present T-score values below − 2.5 standard deviations from the average value of young healthy women. Bone quality is defned as the combination of bone composition and structure that contributes to bone strength [[1\]](#page-14-0). Osteoporosis is associated with an increased morbidity and mortality and causes a major public health burden. It commonly appears after the 5th decade of life and can be also associated with loss of muscle function and mass, known as sarcopenia [\[2](#page-14-1)]. To date, there are diferent treatments for osteoporosis which range from dietary supplements like vitamin D and calcium to pharmacological approaches, including antiresorptive (i.e. bisphosphonates)

 $\boxtimes$  M. Herrmann markus.herrmann@medunigraz.at and anabolic drugs (i.e. teriparatide). Some countries, like Japan, have also included vitamin K in their recommendations for patients with osteoporosis [[3\]](#page-14-2). Vitamin K is a liposoluble compound with a well-established function in blood clotting  $[4]$  $[4]$ , which has also shown some beneficial effects in bone and muscle metabolism, especially in in vitro and in vivo models. Observational and interventional studies in patients with osteoporosis and sarcopenia have yielded conficting results. This review aims to summarize the latest fndings on the role of vitamin K in bone and muscle metabolism, including both preclinical and clinical studies published from 2015.

# **Chemical and Physiological Characteristics of Vitamin K**

Vitamin K (VK) is an essential cofactor for the  $\gamma$ -glutamyl carboxylase (GGCX) enzyme that converts glutamic acid (Glu) residues into γ-carboxyglutamic acid (Gla) in the Gla protein family [[5\]](#page-15-1), which includes seventeen members, namely osteocalcin (OCN), matrix Gla protein (MGP), growth arrest specifc protein 6 (Gas6), periostin,

<sup>&</sup>lt;sup>1</sup> Clinical Institute of Medical and Chemical Laboratory Diagnostics, Medical University of Graz, Graz, Austria

periostin-like factor, Gla-rich proteins, seven proteins involved in blood clotting, two proline-rich Gla proteins (PRGP1, PRGP2) and two transmembrane Gla proteins (TMG3, TMG4) [[6\]](#page-15-2). GGCX is located in the endoplasmatic reticulum [[5\]](#page-15-1) and expressed in diferent tissues, including liver, brain, heart, kidney, lung, pancreas and skeletal muscle [[7\]](#page-15-3).

The term vitamin K comprises a heterogeneous group of chemical compounds of diferent origin. All of them share an unsubstituted naphthoquinone core structure with a free methyl group in position 2. Diferent lipophilic side chains in position 3 characterize the individual members of the VK family. These side chains vary in length and the degree of saturation, and thus determine the fat solubility of each compound  $[8]$ . Vitamin K-1 (VK-1) and vitamin K-2 (VK-2) are the two main forms of VK that difer only in their side chain in position 3. VK-1 has a phytyl side chain, whereas VK-2 has a side chain with a varying number of isoprenyl units (Fig. [1](#page-1-0)), an unsaturated hydrocarbon motif with the formula CH2=C(CH3)−CH=CH2. VK-1, known as phylloquinone, is mostly found in green plants, like spinach, broccoli, green leaf lettuces or parsley, and accounts for more than 50% of the dietary intake of VK [[9\]](#page-15-5). VK-2 (menaquinone) is found in fermented soybeans (natto), egg yolk and chicken breast, and it can also be produced by the gut bacteria. Several forms of VK-2 are known, usually abbreviated as MK-n, where 'n' is the number of isoprenyl groups in the side chain. These include MK-1, MK-4, MK-7, MK-9 and MK-11, whereby the properties and activities depend on the lipophilic side chain and the aromatic methyl-group ring. Vitamin K-3 (VK-3), also known as menadione (2-methyl-1,4-naphtoquinone)

is a water-soluble synthetic form of VK which has no side chain and is classifed as a pro-vitamin [[10](#page-15-6)]. Small amounts of VK-1 can be converted into MK-4 either directly within certain tissues or via the VK-3 intermediate which is prenylated to be converted into MK-4 by the ubiquitously expressed UBIAD1 enzyme [[11](#page-15-7)].

Dietary VK-1 is absorbed in the intestine by active transport in the jejunum, whilst VK-2 is absorbed in the small intestine by difusion. Bile salts and products of pancreatic lipase are required for optimal absorption. Therefore, extrahepatic cholestasis and pancreatic insufficiency may result in VK deficiency  $[12]$  $[12]$ . The absorption rates of VK metabolites decrease significantly with the length of the side chain [\[13](#page-15-9)]. In the blood circulation, VK is transported to the diferent tissues by lipoproteins (Fig. [2](#page-2-0)). Bioavailability and pharmacokinetics vary widely amongst the diferent forms of VK. Some of them have very short half-life, like VK-1 [\[14](#page-15-10)], and can be found in the circulation from 4 to 10 h after ingestion, whilst others, like MK-7, are present for more than 96 h  $[15]$  $[15]$ . Tissue distribution also differs between the short-chain and long-chain VKs. VK-1 can be found in all tissues, but mostly in liver and heart. MK-4, instead, is most abundant in pancreas, salivary gland and sternum [[16\]](#page-15-12).

Once the VK is incorporated into the rough endoplasmic reticulum of the cells, it hydroxylates into the biologically active form hydroquinone. This form acts as a cofactor of GGCX and is converted into VK-2,3 epoxide, then reduced into the respective quinone by VK epoxide reductase complex subunit 1 (VKORC1), and, fnally, it is transformed back into hydroquinone by the VK reductase. Then, the cycle starts again. Various drugs, such as coumarin (i.e. warfarin), inhibit the VK epoxide reductase (Fig. [3](#page-2-1)) [\[17](#page-15-13)].

<span id="page-1-0"></span>**Fig. 1** Chemical structure of vitamin K species. The isoprenyl unit is shown in brackets





<span id="page-2-0"></span>**Fig. 2** Vitamin K uptake and elimination in the body: After oral intake of vitamin K, it is absorbed in the intestine into the thoracic duct and then transported by the bloodstream to the liver (where it undergoes further metabolism and participates in various carboxyla-

tion reactions of the blood coagulation factors), adipose tissue (probably for storage), muscles and bones. Approximately 50% of the excretion is reabsorbed, 30% is eliminated via the faeces and approximately 20% via the urine



<span id="page-2-1"></span>**Fig. 3** Schematic representation of the vitamin K cycle

The daily recommended intake of VK it is not well established yet and varies from country to country. It ranges from 90 µg (women) and 120 µg (men)/day in the USA to 70 ug/ day in all adults including pregnant and lactating women in Europe (European Food Safety Authority, EFSA 2017) [\[9](#page-15-5), [18\]](#page-15-14). VK-1 accounts for 50%, MK-4 for 10% and MK-7, MK-8 and MK-9 for 40% of the absorbed VK [\[19](#page-15-15)]. However, it is unclear whether these recommendations can be sufficient for the total of all carboxylation reactions. Recent studies have shown that only MK-7 can achieve the γ-carboxylation of the extrahepatic vitamin K-dependent proteins (VKDPs) with the recommended daily requirement [\[20\]](#page-15-16).

Although diet is the major source of vitamin K, some VK metabolites are produced by the gut microbiota. *Bacteroides fragilis* produces MK-10, MK-11 and MK-12, *Eubacterium lentum* generates MK-6 and *Lactococcus lactis ssp. lactis* and *ssp.cremoris* synthesize MK-8 and MK-9 [[21,](#page-15-17) [22](#page-15-18)]. Moreover, most aerobic Gram-positive and anaerobic gut bacteria use VK-2 in their electron transport pathways and are also responsible for the length and saturation of the VK side chain [[23\]](#page-15-19). However, the exact mechanism is not fully understood yet.

#### **Analytical Methods of Vitamin K Metabolites**

VK metabolites are present in serum at very low concentrations, which limits their quantifcation with the methodology available to date. Besides, they show high affinity for lipoproteins, making their separation technically challenging. Therefore, most of the studies on VK assess its levels by food questionnaires or other proxy parameters, like prothrombin rate or levels of undercarboxylated proteins.

The frst determination of VK-1 in serum was reported in 1979 by Lefevere et al. [[24](#page-15-20)] using a classical chromatographic method with direct ultraviolet detection that required large sample volumes and a time-consuming pre-analytical procedure. This method sufered from a limited sensitivity and selectivity. A few years later, Ueno et al. developed a frst HPLC method with electrochemical detection that offered improved sensitivity and specificity  $[25]$  $[25]$ . Through the employment of a glassy carbon electrode and an Ag/AgCl reference electrode, they were able to detect 2.2 nmol/L of VK-1 in serum. The next signifcant improvement in the analysis of VK was the introduction of a post-column reduction reactor flled with metallic zinc coupled to a fuorescence detector [\[26](#page-15-22)]. This widely used approach results in an enhanced sensitivity and selectivity. The improvement in separation columns over the last 20 years allowed the use of less complex sample preparation procedures for the removal of interfering substances. Today, the HPLC approach with post-column reduction reactor and fuorescence detector is the most commonly used method for the measurement of VK in medical laboratories. In the absence of a standard reference material and an accepted reference measurement procedure, the comparability of results within a single laboratory as well as between laboratories is limited and depends on the preparation of the post-column reactor, the mobile phases and the choice of separation column.

Liquid chromatography-tandem mass spectrometry (LC–MS/MS) has brought signifcant progress in analytical chemistry through its high selectivity and sensitivity. However, its use for the measurement of VK is hampered by the non-polar structure of VK and the high number of accompanying substances that suppress ionization and thus limit sensitivity. In order to optimize sensitivity, LC–MS/ MS methods usually employ the same sample preparation as HPLC, which consists of liquid–liquid extraction followed by a solid-phase purifcation step [[27\]](#page-15-23). In addition to this rather complex sample preparation, Suhara et al. used a 75-min chromatographic separation protocol with extensive wash and re-equilibration steps. Compared to previous methods, this approach resulted in improved sensitivity and selectivity. With 500 µL of sample volume, they achieved a limit of detection (LOD) of 0.09 nmol/L for VK-1, 0.11 nmol/L for MK-4 and 0.12 nmol/mL for MK-7. Furthermore, the serum concentrations for VK-1, MK-4 and MK-7 correlated well with the HPLC method. When employing this LCMS method in a cohort study of 396 Japanese women, Tsugawa et al. found mean plasma concentrations of VK-1, MK-4 and MK-7 that ranged from 0.88 for MK-4 to 14.3 nmol/L for MK-7 [[28](#page-15-24)]. Despite its improved sensitivity and selectivity, this rather complex method is not suitable for clinical practice.

In 2014, Karl et al. [\[29\]](#page-15-25) developed a method for the simultaneous quantitation of 11 VK species (VK-1, MK-4 to MK-13) by LC–MS/MS with atmospheric pressure chemical ionization (APCI). However, the advantage of detecting multiple VK species in one run is fawed by a limited sensitivity (LODs between 1 and 30 nmol/L). Consequently, most VK compounds could only be quantitated in stool, but not in serum.

Riphagen et al. have tried to develop a less complex, but sufficiently sensitive, method for the measurement of VK by focussing on the most abundant species VK-1, MK-4 and MK-7 [[30](#page-15-26)]. The limits of quantifcation (LOQ) using 350 µL plasma ranged from 0.14 nmol/L for VK-1 and MK-4 to 4.4 nmol for MK-7. In 2019, Dunovska et al. [[31\]](#page-15-27) published a method with an improved sensitivity for these three compounds with LOQs of 0.07 nmol/L for VK-1 and MK-4, and 0.05 nmol for MK-7.

In summary, the measurement of VK is highly complex and requires a compromise between sensitivity and throughput (Table [1](#page-4-0)). Amongst all methods published to date, the HPLC method from Haaron et al. and the LC–MS/

Study, year [ref]	Volume	Instrumentation/detec- tion	Sample preparation	$LOQ$ (nmol/L)	Fasting human serum levels $(nmol/L)$
Lefevere et al., 1979 [24]	2 mL	HPLC / UV 248 nm	Methanol/n-hexan $&$ precleaning on a silica column	$1.1$ nmol/L VK-1	11 to $66$ nmol/L
Ueno et al., 1983 [25]	$1.5$ mL	HPLC / ECD	Ethanol/n-hexan $&$ precleaning on a silica column	$0.7$ nmol/L VK-1	2.2 nmol/L $(n=26)$
Haroon et al., 1986 [26]	$0.5 -$ $1 \text{ mL}$	HPLC/FLD Zn post-column reactor	Ethanol/n-hexan $&$ solid- phase extraction (SPE)	$0.1$ nmol/L VK-1	1.23 nmol/L $(n=22)$
Suhara et al., 2005 [27]	$1$ mL	LC-APCI-MS/MS	Ethanol/n-hexan & solid- phase extraction (SPE)	0.09 VK-1; 0.11 MK-4, $0.12$ MK-7	$2.68 \pm 1.25$ VK-1; $0.88 \pm 1.04$ MK-4; $4.3 \pm 11.5$ MK-7 $(n=20)$
Riphagen et al., 2016 [30]	$0.35$ mL	LC-APCI-MS/MS	Precipitation with ethanol & solid-phase extraction (SPE)	$0.14$ VK-1; 0.14 MK-4, 4.4 MK-7	$1.35(0.89-2.32)$ VK-1,; $0.20(0.17-0.25)$ $MK-4$ : $<$ 4.40 MK-7 (n = 60)
Klapkova et al., 2018 $\lceil 101 \rceil$	$0.5 -$ $1 \text{ mL}$	HPLC/FLD Zn post-column reactor	Ethanol/n-hexan & solid- phase extraction (SPE)	0.07 VK-1; 0.07 MK-4; $0.05$ MK-7	$1.08 \pm 0.88$ VK-1; $1.85 \pm 0.60$ MK-4; $1.83 \pm 1.66$ MK-7 $(n=158)$
Dunovska et al., 2019 $\left[31\right]$	$0.5$ mL	LC-ESI-MS/MS	Ethanol/n-hexan $&$ solid- phase extraction (SPE)	$0.07$ VK-1; 0.07 MK-4; $0.05$ MK-7	$0.44(0.07-4.8)$ VK-1; $0.42(0.11-6.3)$ MK-4; $0.14(0.08-2.1)$ MK-7 $(n=191)$

<span id="page-4-0"></span>**Table 1** Overview of the development of the vitamin K determination in human serum and the technologies used

MS method from Dunovska et al. are best suited for clinical practice. The lack of standardization and appropriate external quality assurance programmes limit the comparability of results obtained by diferent laboratories. So far, the European Vitamin K External Quality Assurance (KEQAS) programme is the only one that allows an independent control of VK-1 results, but does not provide a certifed target concentration. Other VK species, such as MK-4, MK-7 and vitamin K1 2,3-epoxide are only available as a pilot scheme with concentrations outside the physiological range. Therefore, some laboratories consider assessing the levels of vitamin K using indirect methods, like measuring prothrombin time (PT) or undercarboxylated vitamin K-dependent proteins. Vitamin K defciency in adults is clinically characterized by a bleeding tendency due to the low activity of blood coagulation factors, resulting in an increase of prothrombin time or partial thromboplastin time. In healthy adults, this can occur more than 2–3 weeks after the very low phylloquinone intake (i.e.  $<$  10 lb/day) [\[32](#page-15-28)]. However, this measure is not a reliable indicator of vitamin K status, because an abnormal prothrombin time can also arise from hepatic dysfunction or haematological conditions, independently of vitamin K (reviewed by Booth and Al Rajabi [[33\]](#page-15-29)). Alternatively, the determination of undercarboxylated vitamin K-dependent proteins is also used as a proxy to identify vitamin K defciency. Serum concentration of undercarboxylated osteocalcin (ucOC) [[34,](#page-15-30) [35\]](#page-15-31) or desphospho-uncarboxylated MGP (dp-ucMGP) are two examples. These undercarboxylated proteins can be determine alone, or together with the total amount of each protein expressing the results as percentage. Although the direct quantifcation of dp-ucMGP is capable of showing a reduction in response to vitamin K supplementation [\[36–](#page-15-32)[38\]](#page-15-33), expressing dp-ucMGP as percentage of the total MGP concentration is more precise because it takes into consideration the biological variability of the total MGP concentration. Moreover, the PIVKA-II (Protein Induced by Vitamin K Absence II) concentration has also been proposed as a biomarker for vitamin K status in some renal, bone and cardiovascular diseases [\[39](#page-15-34)[–41](#page-16-0)]. Although the Expert Group of the EFSA [[18\]](#page-15-14) acknowledges that phylloquinone intake at certain doses changes the concentration of the abovementioned proteins, it has to be mentioned that these doses are considerably higher than those recommended in Europe. Another issue is that commercially available PIVKA-II assays are not sufficiently sensitive to measure normal concentrations, which are  $\leq 2 \mu g/L$ . Also, the optimal degree of carboxylation of PIVKA-II, OC or MGP is unknown and reliable information on vitamin K intake required for c-carboxylation of these proteins is lacking. In summary, the inherent limitations of indirect vitamin K surrogate markers make them rather unreliable and thus they should be used with caution. The direct measurement of vitamin K and its metabolites can overcome virtually all of these issues and is thus the preferred way to assess vitamin K status.

# **Pre‑clinical Studies on the Role of Vitamin K in the Musculoskeletal System**

#### **Vitamin K and Bone**

Preclinical research has provided essential insights into the metabolic activities of VK in the musculoskeletal systems. Cell culture studies indicate an anabolic role of VK in bone by increasing osteoblast activity and decreasing osteoclas togenesis. Already more than 20 years ago, in vitro studies found that VK-2 promotes mesenchymal stem cell prolifera t[ion](#page-16-1) and diferentiation into osteoblasts, but not adipocytes [[42\]](#page-16-1), by enhancing the expression of cartilage-associated Growth diferentiation factor 15 (Gdf15) and Stanniocalcin 2 (Stc2) [[43\]](#page-16-2). It also prevents osteoblast apoptosis by inhib iting the apoptotic genes *Fas* and *Bax* [\[42\]](#page-16-1) and stimulates autophagy via Lc3 and Beclin1 [[44](#page-16-3)]. A very recent study has shown that MK-7 is required for osteogenic diferentia tion at early stages, when it upregulates RUNX2 through an OCN-independent pathway [[45](#page-16-4)] (Fig. [4](#page-5-0)). Apart from enhancing osteoblast diferentiation and activity, VK also regulates osteoclasts, both directly and indirectly. It inhibits osteoclastogenesis by IκBα-dependent NF-κB downregula tion  $[46]$  and induces osteoclast apoptosis  $[47]$  $[47]$  $[47]$ , as well as reduces the *RANKL/OPG* mRNA ratio in osteocytes, leading to a decrease in osteoclast activity [\[48](#page-16-7)] (Fig. [4\)](#page-5-0).

In diferentiated osteoblasts, VK acts primarily as a cofac tor for GCCX, which activates bone-related proteins, like osteocalcin (OCN) and matrix Gla protein (MGP). OCN is the most abundant non-collagenous protein in the bone matrix. It is synthesized by osteoblasts and released to bind hydroxyapatite crystals in the extracellular matrix, promot ing mineralization. Its affinity for calcium depends on the carboxylation of three Glu residues by the two enzymes GGCX and VKORC1 [\[49](#page-16-8)]. Most of the carboxylated OCN is bound in the extracellular bone matrix, but during bone resorption, the low pH in the resorption lacunae can decar boxlyte Gla residues of OCN and resulting in the release of OCN, and, in particular, undercarboxylated OCN, into the blood. Therefore, the OCN concentration in serum is used as a biomarker for bone turnover, and undercarboxylated OCN has been proposed as an indicator of the VK levels [[50](#page-16-9)]. Recent results from an OCN knockout mouse model suggest that OCN is required for aligning the hydroxyapatite crystals between collagen fbres and thus contributes to bone quality (Fig. [4](#page-5-0)) [\[51](#page-16-10)]. However, the importance of the VK-dependent OCN carboxylation for bone mineralization is questioned by the results of another conditional knockout (KO) model for *GGCX* (*Col1-Cre Ggcx*−/−) [[52\]](#page-16-11). In this model, the lack of GGCX in the osteoblastic lineage resulted in increased trabecular and cortical bone formation and extra skeletal calcifcation, but no reduction in bone mineral density.

fiber area<br>Alterations in insulin and glucose<br>expenditure Regulation of muscle mass and y-carboxylation of **OCN** Improvement of mitochondrial<br>electron transfer with increased Regulation of energy metabolism Muscle of ATP production Reduction of lactate dehydrogenase Increase of myogenin and myoD expression<br>Reduction of differentiation Regulation of early Increase of proliferation myogenesis Increase of migration Vitamin K Inhibition of osteoclastogenesis<br>Induction of osteoclast apoptosis<br>Decrease of RANKL expression Increase of OPG expression Inhibitory effects<br>on bone resorption and collagen fibers<br>Regulation of bone mineralisation Production of osteocalcin (OCN) Alignment of apatite crystals matrix Gla-protein (MGP) y-carboxylation Cofactor of Bone Carboxylation of Induction of osteoblast differentiation<br>Increase of alkaline phosphatase  $\arcsin y$  (ALP)  $$\tilde{}$$  Prevention of osteoblast apoptosis Stimulation of osteogenesis Improvement of<br>osteoblast function  $\ddot{\phantom{a}}$ 

<span id="page-5-0"></span>**Fig. 4** Role of vitamin K in bone and muscle metabolism from in vitro and in vivo studies. Dashed line indicates a questionable role

Fig. 4 Role of vitamin K in bone and muscle metabolism from in vitro and in vivo studies. Dashed line indicates a questionable role

In addition to OCN, VK is also required to carboxylate MGP, expressed in cartilage, bone and soft tissue. This protein binds the organic matrix and hydroxyapatite crystals and inhibits vascular calcifcation [[53](#page-16-12)]. Apart of its role in inhibiting bone formation, it also regulates osteoclastogenesis. MGP inhibition leads to an increased osteoclast differentiation and bone resorption as demonstrated in vivo in a *Mgp−/−* mouse model [[54\]](#page-16-13). Moreover, VK is needed for the carboxylation of the growth arrest specifc gene product (Gas6), which regulates the activity of osteoclasts and protein S, a cofactor for the anticoagulant protein C. It shows a high degree of homology with Gas6, but its role in bone is still unknown [[55\]](#page-16-14).

Through binding to the steroid and xenobiotic receptor (SXR; also known as pregnane X receptor, Pxr, in mice), VK also exerts effects that are unrelated to  $γ$ -carboxylation [\[56](#page-16-15)]. SXR is a xenobiotic sensor associated with detoxification and drug excretion that activates Tsukushi, Matrilin-2 and CD14 proteins, involved in collagen accumulation, osteoblast- and osteoclastogenesis (Fig. [4](#page-5-0)) [[5\]](#page-15-1). Several Pxr KO mouse models have been developed, showing diferent phenotypes. One of them presented hypophosphatemia, as Pxr directly activates the cotransporter Na/Pi Slc34a2 [\[57](#page-16-16)], whilst the other did not show any change in serum calcium or phosphate, but an osteoporotic phenotype: reduced bone mineral density (BMD) and decreased trabecular and cortical thickness with increased bone fragility [[58\]](#page-16-17).

Last, but not least, there is also some evidence that VK-1 and VK-2 are involved in the antioxidant defence against reactive oxygen species that are produced by metabolically active osteoblasts [\[45](#page-16-4), [59](#page-16-18)]. MK-7 also increases the expression of PMCA, a calcium transporting ATPase, promoting normal bone mineralization even under oxidative stress conditions [\[59\]](#page-16-18).

In addition to the isolated effects of VK in bone, several in vitro and in vivo studies have investigated if VK modifes the efects of common anti-osteoporotic treatments. For example, combining vitamin D3 and vitamin K has resulted in additive anabolic efects in murine models and osteopenic patients. In a murine model for diabetes, the two vitamins together increased calcium deposits, bone anabolic markers and bone formation transcription factors, suggesting that this combination could be useful for the treatment of diabetes-associated osteoporosis [[60](#page-16-19)]. In cultured precursor cells from osteopenic patients, the combined treatment with VK and vitamin D increased osteoblastogenesis and MSC diferentiation into osteoblasts and decreased osteoclastogenesis. After one year of treatment, these patients presented an increased lumbar spine and femoral neck BMD and a decreased risk of fracture [\[61](#page-16-20)].

Genistein, an estrogenic agent, also showed an increased bone formation capacity when combined with vitamin K. Both treatments increased OCN and alkaline phosphatase

activity in MC3T3 cells, suggesting that this treatment could promote osteoblast activity [\[62](#page-16-21)].

Teriparatide is a well-established anabolic treatment for patients with severe osteoporosis. The combined treatment of ovariectomized rats, an established model for osteoporosis, with teriparatide and VK-2 increased serum γ-carboxylated OCN, showed higher BMD [\[63](#page-16-22)] and femoral bone strength [\[64](#page-16-23)] than each of the two treatments by itself. In vitro analysis of primary bone marrow-derived stromal cells from these animals showed that the combined treatment prevented bone loss and enhanced bone formation and mineralization better that teriparatide alone [[65\]](#page-16-24).

Moreover, vitamin K also seems to have beneficial efects when combined with zoledronic acid, an antiresorptive drug for osteoporosis. VK-2 pre-treatment of primary murine osteoblasts reduces Sost expression and partially prevents the inhibition of bone formation indices (BMD, calcium content and bone strength) caused by zoledronic acid [\[66](#page-16-25)]. In vivo studies using ovariectomized Wistar rats confrmed these results, as pre-treatment with VK increased the calcium content, improved trabecular structure and antipressure strength of bone after bisphosphonate treatment. However, no effect was detected when the treatments were administered at the same time [\[66](#page-16-25)].

Taken together, existing evidence from preclinical studies is limited and in many ways inconclusive. However, there seem to be some anabolic effects that are mediated by stimulating osteoblasts and inhibiting osteoclasts. In addition, VK seems to improve the efficacy of common bone-preserving compounds, such as vitamin D, teriparatide or bisphosphonates. The underlying mechanisms are poorly understood and subject of ongoing research.

#### **Vitamin K and Skeletal Muscle**

Little is known about the role of VK supplementation in the skeletal muscle. Only few studies have been performed to date, suggesting that VK exerts a protective efect. Most of the research, however, has focussed on how the carboxylation of bone-derived OCN could afect muscle health, a role that lately has been questioned [[51,](#page-16-10) [67](#page-16-26)]. In vitro studies by Rønning et al. have shown that MK-4 increases proliferation and migration of bovine muscle cells [[68\]](#page-16-27). Furthermore, MK increases the expression of myogenin and myoD, and reduces the diferentiation of bovine satellite cells. In this cell culture model, MK-4 also reduces lactate dehydrogenase, a marker for cell lysis and toxicity, indicating a stabilizing efect on muscle cells. Expression of Lrp1 and Ldlr receptor was increased shortly after the treatment with MK-4, and the authors suggest that they could be involved in the VK-driven changes in muscle cells, although the exact mechanism is still unknown (Fig. [4\)](#page-5-0).

Some studies also suggested that VK could regulate energy metabolism in the skeletal muscle. VK-2 (MK-4) improves the transfer of electrons in the mitochondria, increasing ATP production [[69](#page-16-28)]. It could as well maintain the balance of the skeletal muscle mitochondria, which is a major factor to prevent sarcopenia [\[70](#page-16-29)].

Moreover, for long time OCN has been considered a bone hormone that could regulate diferent tissues, including muscle [\[71\]](#page-17-1). This work has been based on a knockout mouse model for OCN [\[72](#page-17-2)], which showed reduced muscle mass and decreased muscle fbre area compared to wild type animals. These animals also presented alterations in insulin secretion and glucose expenditure by peripheral tissues, like liver, muscle and adipose tissue (reviewed in [[73](#page-17-3)]). However, last year, two independent OCN KO mouse models failed to validate these changes [\[51,](#page-16-10) [67\]](#page-16-26). The animals exhibited normal muscle mass and fbre area, and no alterations in glucose or lipid metabolism, which questions the hormonal role of OCN.

## **Role of Vitamin K in Human Studies**

#### **Vitamin K and Fragility Fractures**

Preclinical analyses suggest that VK is involved in bone metabolism and has anabolic efects. However, it is unclear if VK has an infuence on bone metabolism and bone quality in humans. Over the years, a number of studies has been published with conficting results. Several recent observational studies suggest a relationship between a high VK intake and increased BMD (Table [2\)](#page-8-0), in both East Asian and European populations. However, most of these studies are limited by a rather small cohort size and the lack of VK measurements in serum or plasma. In addition, the study populations were rather heterogeneous including healthy elderly individuals, osteoporotic women, cystic fbrosis patients, renal patients and children with low trauma fractures. Apart from three studies [[74–](#page-17-4)[76\]](#page-17-5), VK status was estimated indirectly by food questionnaires or serum levels of undercarboxylated proteins (OCN or MGP). When measuring serum VK by HPLC or LC–MS/MS, no relationship with BMD or biochemical bone turnover markers was found [\[74](#page-17-4), [75](#page-17-6)].

Additional information can be derived from supplementation studies. All 15 VK treatment studies that have been published since 2015 are summarized in Table [3.](#page-9-0) The results of these studies are inconsistent. Whilst 6 studies reported an increase in BMD upon VK treatment, the others did not. In a meta-analysis of 19 randomized controlled trials, including 6,759 Asian and Caucasian participants, Huang et al. found an increase of vertebral BMD in postmenopausal women with osteoporosis [\[77\]](#page-17-7). In non-osteoporotic individuals this efect was not present. Although this meta-analysis suggests that the evidence for VK efect on BMD is robust, it should be mentioned that all osteoporotic patients were of Asian origin, mostly Japanese, whilst the controls were mostly Caucasians. This leaves ample room for ethnicity-related efects, such as diferences in pharmacogenomics and lifestyle [\[77\]](#page-17-7). In fact, studies in Caucasians are limited to a few rather small studies. The best evidence in Caucasians comes from a randomized placebo-controlled, study involving 142 healthy postmenopausal Danish women [\[78](#page-17-8)]. In this study, 12 months of VK-2 supplementation together with calcium and vitamin D did not change BMD. In order to assess whether a longer treatment was needed to induce signifcant efects, the authors extended their study for another two years [[79](#page-17-9)]. However, even after 3 years of treatment BMD at total hip, femoral neck or lumbar spine was not afected by VK supplementation. Only a small increase in trabecular thickness was observed. The combination of VK-2 and risedronate also failed to improve BMD when compared with risedronate alone in a prospective, multicentre, open-labelled, randomized trial including 1983 patients (Table [3\)](#page-9-0) [[80\]](#page-17-10).

Bone turnover biomarkers were only analysed in a subset of the available studies (Table [3\)](#page-9-0), with variable results. In some cases, VK increased P1NP and decreased CTx levels [[61,](#page-16-20) [81,](#page-17-11) [82](#page-17-12)], but these results were not consistent across the trials [\[78](#page-17-8), [83](#page-17-13)]. Only undercarboxylated OCN was consistently reduced by VK, which confirms the efficacy of the supplementation.

Quantitative ultrasound sonometry is a widely used tool to assess bone quality. One of the parameters obtained is the speed of sound (SOS) which informs about the bone density. Suzuki et al. [[84\]](#page-17-14) found that levels of undercarboxylated OCN negatively correlate with SOS in 49 healthy Japanese females,  $84\%$  of which presented VK-1 insufficiency. The mechanism involved is still unknown, but the authors propose that VK could regulate bone quality either as a coenzyme of Gla proteins involved in mineralization [\[49](#page-16-8)] or as a ligand for the SXR receptor, to stimulate the expression of TSK, which is involved in collagen assembly and could afect the bone scafold [[56\]](#page-16-15).

Fractures are the typical result of reduced bone quality. Therefore, several studies have investigated if VK is related to fracture risk, but the results do not permit a clear answer yet (as reviewed in [[85](#page-17-15), [86](#page-17-16)]). Observational studies, including predominantly healthy and osteoporotic Caucasians, suggest that the serum VK concentration is inversely related to fracture risk in line with previous multicentre studies [[87](#page-17-17), [88\]](#page-17-18). In a cross-sectional study of 374 women with postmenopausal osteoporosis, Moore et al. showed a 45% reduction in fracture risk per ug/L higher serum VK-1 concentration [[74](#page-17-4)]. This association remained signifcant after correcting for age, BMI, vitamin D and lifestyle factors. In addition, VK-1 was also

<span id="page-8-0"></span>

*ucOCN*undercarboxylated OCN, *fx* fracture, *TBS*trabecular bone score, *CKD*chronic kidney disease, *LCMSMS*liquid chromatography tandem mass spectrometry

ucOCN undercarboxylated OCN, fx fracture, TBS trabecular bone score, CKD chronic kidney disease, LCMSMS liquid chromatography tandem mass spectrometry



<span id="page-9-0"></span> $\underline{\textcircled{\tiny 2}}$  Springer





 $\underline{\textcircled{\tiny 2}}$  Springer

alkaline phosphatase, *P1NP*total procollagen type I N-terminal propeptide, *cOCN*carboxylated OCN

associated with hip geometry and indices of mechanical strength. These results are in line with the NOREPOS study (Norwegian Epidemiologic Osteoporosis Study) [[76\]](#page-17-5). This large epidemiologic study includes 21,774 men and women from four population-based studies aged 65–79 years. After adjustment for age and gender, low serum concentrations of VK-1 and vitamin D were associated with a 50% higher risk for hip fractures.

Intervention studies also have been conducted to investigate the efect of VK supplementation in preventing fractures. The results of these studies are mixed. Whilst some found a signifcant reduction in fracture risk [\[61](#page-16-20), [77](#page-17-7), [89](#page-17-20)], others did not [[80,](#page-17-10) [90\]](#page-17-19). A frst meta-analysis of osteopenic/ osteoporotic patients and controls showed reduction of fracture risk in patients, but not in controls [[77\]](#page-17-7). However, this meta-analysis is limited by the heterogeneity of the studies included. For example, all osteopenic/osteoporotic patients were of East-Asian origin, whereas the healthy controls were mostly Caucasian controls. Furthermore, some studies analysed rather small cohorts, with as few as 20 patients [\[52](#page-16-11)]. A later meta-analysis performed by Mott et al. in 2019, included 36 studies with a total of 11,112 participants. In this analysis VK reduced the risk of all clinical fractures in postmenopausal women and osteoporotic patients (OR 0.72, 95% CI 0.55 to 0.95) [[89\]](#page-17-20), although after excluding studies with a high risk of bias, the efect was no longer signifcant (OR 0.76, 95% CI 0.58 to 1.01). However, it should be mentioned that this meta-analysis included a very broad spectrum of healthy individuals and patients with bone, liver and kidney disease from diferent ethnical groups. The absence of a signifcant VK efect on fracture risk is further supported by another meta-analysis of Asian cohorts that included 70% of postmenopausal women [[90](#page-17-19)]. A general issue comparing VK supplementation studies is the wide range of doses that were administered, the frequent combination with other micronutrients, anti-osteoporotic drugs and a highly variable study duration.

Additional clues regarding the efect of VK on fracture risk can be drawn from patients that are treated with Warfarin, an inhibitor of GGCX and VKORC1, which causes an inhibition of VK function. Existing studies reported mixed results. However, a meta-analysis including 22 observational studies and one randomized controlled trial, with more than one million participants in total, failed to identify any association between VK antagonists or DOACs and fracture risk, even during long-term treatment. Nevertheless, in selected sub-groups, such as female VK antagonist users (OR 1.11 [95%CI 1.02–1.21]) and patients over 65 years of age (OR 1.07 [95%CI 1.01–1.14]), this meta-analysis found small, but signifcant associations. Although existing data suggests that the risk of fracture does not seem a major consideration in anticoagulated patients, it should be point out that most existing studies are of observational nature.

Despite the lack of convincing evidence that VK modifies BMD and fracture risk, some recent studies raised the hypothesis that the gut microbiome might be related to fracture risk through its efects on VK metabolism. A small study in 38 postmenopausal women with no conditions or medication afecting their bone metabolism or gut microbiota showed that Bacteroides, which produce MK-10, MK-11 and MK-12, were associated with VK-2 levels and fracture risk [[91](#page-17-21)]. Since no diferences in VK intake were found between patients with low and high VK-2 levels, these results suggest a role of gut microbiota in reducing fractures risk by regulating VK-2 levels. The relevance of this observation is further supported by a case report of a 48-year-old Asian man with bisphosphonate-associated fractures. An incomplete atypical femoral fracture on his left leg was treated with 15 mg of VK-2 three times a day. After 4 months, X-rays showed healing of the fracture line, sup-porting an anabolic effect of VK-2 [[92\]](#page-17-22).

#### **Role of Vitamin K in Sarcopenia**

Imbalances of the serum VK status have also been discussed to adversely afect muscle mass and function [[93](#page-17-23)]. Since 2015, only fve clinical trials have assessed the role of VK in muscle health (Table [4\)](#page-13-0). Observational studies found that better physical performance is associated with higher plasma concentrations of VK [[94](#page-17-24)]. However, intervention studies have not yet confrmed a causal relationship [[95,](#page-17-25) [96\]](#page-17-26).

The frst study on the role of VK in lower extremity function was carried out in 2016 [[94\]](#page-17-24). In 1,089 elderly participants of the Health, Ageing and Body Composition Study (Health ABC), lower extremity muscle function was assessed by a short physical performance battery (SPPB), gait speed over 20 m, endurance over 400 m and knee extensor strength. Individuals with a plasma VK-1 concentration over or equal to 1 nmol/L (corresponding to 90–120 µg/ day dietary intake) showed better lower extremity physical performance, both cross-sectionally and over 5 years of follow-up. In addition, the ucMGP concentration was associated with SPPB cross-sectionally, but not longitudinally. The potential role of ucMGP as a surrogate marker of VK status and physical performance has also been investigated in an ongoing, longitudinal population-based study in Dutch adults (the Longitudinal Aging Study Amsterdam, LASA). In this study, higher ucMGP concentrations (indicating a reduced VK status) were associated with lower handgrip strength and calf circumference. Moreover, in women, high ucMGP levels were also related to a lower physical performance [\[97](#page-17-27)].

Whilst some studies support a link between VK and muscle function, others do not. In a 6-months intervention study, treatment of elderly vascular patients with 100 μg/day of VK-2 failed to induce any change in vascular or physical



 $\overline{\phantom{a}}$ 

<span id="page-13-0"></span> $\overline{ }$ 

 $\underline{\textcircled{\tiny 2}}$  Springer

*ucOCN*undercarboxylated OCN, *ucMGP*undercarboxylated MGP

outcome parameters [\[95](#page-17-25)]. Similarly, in another randomized controlled supplementation study, 3 years of VK-1 administration did not show any improvement in lean mass or fat mass [\[96](#page-17-26)]. However, this treatment reduced undercarboxylated OCN levels between 58 and 61% in both women and men [\[98](#page-17-28)].

In order to investigate the role of undercarboxylated OCN in muscle and whether it could be a biomarker for bonemuscle interaction, a randomized controlled multicentre crossover trial has been set up very recently [\[99](#page-17-29)]. This study aims to recruit men and women over 60-year-old, measure the undercarboxylated OCN levels in serum before and after acute exercise, and correlate these values with parameters of muscle quality. In a previous study, a low undercarboxylated OCN/OCN ratio was associated with lower muscle strength in young women [[100](#page-17-30)], but so far, this relationship has not been tested in older adults yet.

### **Limitations**

Many clinical studies have investigated the role of VK in bone and muscle homeostasis with conficting results, due, in part, to multiple methodological diferences between the studies. These include the genetic background and lifestyle of the studied populations, the age of the individuals, the variability of data from self-reported fractures, the presence of bone and muscle diseases, and the changes in other vitamins and triglycerides that could afect VK levels. Furthermore, the power of many studies is compromised by a small sample size and the lack of VK measurement in plasma. Food questionnaires, prothrombin rates, or the carboxylation levels of selected target proteins were often used as surrogates for the participants VK status. However, these markers provide at best a very rough estimate of the supply with VK and do not give any information on individual VK species. Finally yet importantly, many VK treatment studies are hampered by the simultaneous administration of other micronutrients, such as vitamin D and therapeutic drugs that interfere with bone and muscle metabolism.

### **Conclusion**

Despite a substantial body of literature, our knowledge about VK and its role in bone and muscle is still very limited. This is mainly due to the lack of broadly available analytical methods that allow a reliable quantitation of diferent VK species in serum and other biological matrices. Furthermore, high quality clinical studies are rare, which limits the value of existing meta-analysis. Preclinical studies suggest some anabolic effects that are mediated by osteoblasts and osteoclasts. However, the underlying mechanisms and the relevance of these efects in humans is still poorly understood. Sufficiently powered high quality studies are needed to further explore the role of VK in the musculoskeletal system. In particular, the VK status should always be assessed by direct measurement with a fully validated and quality controlled method. Supplementation studies should avoid co-treatment with other micronutrients and include a proper control group. A potential additive efect with established pharmaceutical anti-osteoporotic drugs should also be explored. Systematic preclinical studies can help to unravel the molecular efects of VK on bone and muscle cells in vitro and in vivo.

**Acknowledgements** We want to thank Dr. Giovanny Rodriguez Blanco for his comments and advice on the manuscript preparation.

**Authors Contribution** MH conceived the idea; NA performed the literature search and wrote the bone and muscle sections; EF-P, AM and DE performed the literature search and wrote the chemical, physiological and methodological sections; all authors critically reviewed the manuscript.

**Funding** Open access funding provided by Medical University of Graz. The authors did not receive support from any organization for the submitted work.

**Data Availability** Availability of data and material is not applicable.

#### **Declarations**

**Conflict of interest** There is no confict of interest to declare.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by/4.0/>.

## **References**

- <span id="page-14-0"></span>1. Compston J (2006) Bone quality: what is it and how is it measured? Arq Bras Endocrinol Metabol 50:579–585. [https://doi.org/](https://doi.org/10.1590/s0004-27302006000400003) [10.1590/s0004-27302006000400003](https://doi.org/10.1590/s0004-27302006000400003)
- <span id="page-14-1"></span>2. Cruz-Jentoft AJ, Baeyens JP, Bauer JM et al (2010) Sarcopenia: European consensus on defnition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. Age Ageing 39:412–423.<https://doi.org/10.1093/ageing/afq034>
- <span id="page-14-2"></span>3. Orimo H, Nakamura T, Hosoi T et al (2012) Japanese 2011 guidelines for prevention and treatment of osteoporosis–executive summary. Arch Osteoporos 7:3–20. [https://doi.org/10.1007/](https://doi.org/10.1007/s11657-012-0109-9) [s11657-012-0109-9](https://doi.org/10.1007/s11657-012-0109-9)
- <span id="page-15-0"></span>4. Swanson JC, Suttie JW (1982) Vitamin K dependent in vitro production of prothrombin. Biochemistry 21:6011–6018. [https://doi.](https://doi.org/10.1021/bi00266a044) [org/10.1021/bi00266a044](https://doi.org/10.1021/bi00266a044)
- <span id="page-15-1"></span>5. Azuma K, Ouchi Y, Inoue S (2014) Vitamin K: novel molecular mechanisms of action and its roles in osteoporosis. Geriatr Gerontol Int 14:1–7.<https://doi.org/10.1111/ggi.12060>
- <span id="page-15-2"></span>6. Danziger J (2008) Vitamin K-dependent proteins, warfarin, and vascular calcifcation. Clin J Am Soc Nephrol 3:1504–1510. <https://doi.org/10.2215/CJN.00770208>
- <span id="page-15-3"></span>7. Wu SM, Staford DW, Frazier LD et al (1997) Genomic sequence and transcription start site for the human gamma-glutamyl carboxylase. Blood 89:4058–4062
- <span id="page-15-4"></span>8. Shearer MJ, Newman P (2008) Metabolism and cell biology of vitamin K. Thromb Haemost 100:530–547
- <span id="page-15-5"></span>9. Booth SL (2012) Vitamin K: food composition and dietary intakes. Food Nutr Res.<https://doi.org/10.3402/fnr.v56i0.5505>
- <span id="page-15-6"></span>10. Hirota Y, Tsugawa N, Nakagawa K et al (2013) Menadione (vitamin K3) is a catabolic product of oral phylloquinone (vitamin K1) in the intestine and a circulating precursor of tissue menaquinone-4 (vitamin K2) in rats. J Biol Chem 288:33071–33080. <https://doi.org/10.1074/jbc.M113.477356>
- <span id="page-15-7"></span>11. Nakagawa K, Hirota Y, Sawada N et al (2010) Identifcation of UBIAD1 as a novel human menaquinone-4 biosynthetic enzyme. Nature 468:117–121.<https://doi.org/10.1038/nature09464>
- <span id="page-15-8"></span>12. Olson RE (1984) The function and metabolism of vitamin K. Annu Rev Nutr 4:281–337. [https://doi.org/10.1146/annurev.nu.](https://doi.org/10.1146/annurev.nu.04.070184.001433) [04.070184.001433](https://doi.org/10.1146/annurev.nu.04.070184.001433)
- <span id="page-15-9"></span>13. Conly JM, Stein K (1992) Quantitative and qualitative measurements of K vitamins in human intestinal contents. Am J Gastroenterol 87:311–316
- <span id="page-15-10"></span>14. Novotny JA, Kurilich AC, Britz SJ et al (2010) Vitamin K absorption and kinetics in human subjects after consumption of 13C-labelled phylloquinone from kale. Br J Nutr 104:858–862. <https://doi.org/10.1017/S0007114510001182>
- <span id="page-15-11"></span>15. Sato T, Schurgers LJ, Uenishi K (2012) Comparison of menaquinone-4 and menaquinone-7 bioavailability in healthy women. Nutr J 11:93.<https://doi.org/10.1186/1475-2891-11-93>
- <span id="page-15-12"></span>16. Thijssen HH, Drittij-Reijnders MJ (1994) Vitamin K distribution in rat tissues: dietary phylloquinone is a source of tissue menaquinone-4. Br J Nutr 72:415–425. [https://doi.org/10.1079/](https://doi.org/10.1079/bjn19940043) [bjn19940043](https://doi.org/10.1079/bjn19940043)
- <span id="page-15-13"></span>17. Staford DW (2005) The vitamin K cycle. J Thromb Haemost 3:1873–1878.<https://doi.org/10.1111/j.1538-7836.2005.01419.x>
- <span id="page-15-14"></span>18. Efsa Panel on Dietetic Products N, Allergies TD et al (2017) Dietary reference values for vitamin K. EFSA J 15:e04780. [https://](https://doi.org/10.2903/j.efsa.2017.4780) [doi.org/10.2903/j.efsa.2017.4780](https://doi.org/10.2903/j.efsa.2017.4780)
- <span id="page-15-15"></span>19. Harshman SG, Shea MK (2016) The role of vitamin K in chronic aging diseases: infammation, cardiovascular disease, and osteoarthritis. Curr Nutr Rep 5:90–98. [https://doi.org/10.1007/](https://doi.org/10.1007/s13668-016-0162-x) [s13668-016-0162-x](https://doi.org/10.1007/s13668-016-0162-x)
- <span id="page-15-16"></span>20. DiNicolantonio JJ, Bhutani J, O'Keefe JH (2015) The health benefts of vitamin K. Open Heart 2:e000300. [https://doi.org/](https://doi.org/10.1136/openhrt-2015-000300) [10.1136/openhrt-2015-000300](https://doi.org/10.1136/openhrt-2015-000300)
- <span id="page-15-17"></span>21. Suttie JW (1995) The importance of menaquinones in human nutrition. Annu Rev Nutr 15:399–417. [https://doi.org/10.1146/](https://doi.org/10.1146/annurev.nu.15.070195.002151) [annurev.nu.15.070195.002151](https://doi.org/10.1146/annurev.nu.15.070195.002151)
- <span id="page-15-18"></span>22. Usui Y, Tanimura H, Nishimura N et al (1990) Vitamin K concentrations in the plasma and liver of surgical patients. Am J Clin Nutr 51:846–852.<https://doi.org/10.1093/ajcn/51.5.846>
- <span id="page-15-19"></span>23. Collins MD, Jones D (1981) Distribution of isoprenoid quinone structural types in bacteria and their taxonomic implication. Microbiol Rev 45:316–354. [https://doi.org/10.1128/mr.45.2.](https://doi.org/10.1128/mr.45.2.316-354.1981) [316-354.1981](https://doi.org/10.1128/mr.45.2.316-354.1981)
- <span id="page-15-20"></span>24. Lefevere MF, De Leenheer AP, Claeys AE (1979) High-performance liquid chromatographic assay of vitamin K in human

serum. J Chromatogr 186:749–762. [https://doi.org/10.1016/](https://doi.org/10.1016/s0021-9673(00)95293-2) [s0021-9673\(00\)95293-2](https://doi.org/10.1016/s0021-9673(00)95293-2)

- <span id="page-15-21"></span>25. Ueno T, Suttie JW (1983) High-pressure liquid chromatographicreductive electrochemical detection analysis of serum trans-phylloquinone. Anal Biochem 133:62–67. [https://doi.org/10.1016/](https://doi.org/10.1016/0003-2697(83)90222-1) [0003-2697\(83\)90222-1](https://doi.org/10.1016/0003-2697(83)90222-1)
- <span id="page-15-22"></span>26. Haroon Y, Bacon DS, Sadowski JA (1986) Liquid-chromatographic determination of vitamin K1 in plasma, with fuorometric detection. Clin Chem 32:1925–1929
- <span id="page-15-23"></span>27. Suhara Y, Kamao M, Tsugawa N et al (2005) Method for the determination of vitamin K homologues in human plasma using high-performance liquid chromatography-tandem mass spectrometry. Anal Chem 77:757–763. [https://doi.org/10.1021/ac048](https://doi.org/10.1021/ac0489667) [9667](https://doi.org/10.1021/ac0489667)
- <span id="page-15-24"></span>28. Tsugawa N, Shiraki M, Suhara Y et al (2006) Vitamin K status of healthy Japanese women: age-related vitamin K requirement for gamma-carboxylation of osteocalcin. Am J Clin Nutr 83:380– 386. <https://doi.org/10.1093/ajcn/83.2.380>
- <span id="page-15-25"></span>29. Karl JP, Fu X, Dolnikowski GG et al (2014) Quantifcation of phylloquinone and menaquinones in feces, serum, and food by high-performance liquid chromatography-mass spectrometry. J Chromatogr B Analyt Technol Biomed Life Sci 963:128–133. <https://doi.org/10.1016/j.jchromb.2014.05.056>
- <span id="page-15-26"></span>30. Riphagen IJ, van der Molen JC, van Faassen M et al (2016) Measurement of plasma vitamin K1 (phylloquinone) and K2 (menaquinones-4 and -7) using HPLC-tandem mass spectrometry. Clin Chem Lab Med 54:1201–1210. [https://doi.org/10.1515/](https://doi.org/10.1515/cclm-2015-0864) [cclm-2015-0864](https://doi.org/10.1515/cclm-2015-0864)
- <span id="page-15-27"></span>31. Dunovska K, Klapkova E, Sopko B et al (2019) LC-MS/MS quantitative analysis of phylloquinone, menaquinone-4 and menaquinone-7 in the human serum of a healthy population. PeerJ 7:e7695.<https://doi.org/10.7717/peerj.7695>
- <span id="page-15-28"></span>32. Udall JA (1965) Human sources and absorption of vitamin K in relation to anticoagulation stability. JAMA 194:127–129
- <span id="page-15-29"></span>33. Booth SL, Al Rajabi A (2008) Determinants of vitamin K status in humans. Vitam Horm 78:1–22. [https://doi.org/10.1016/S0083-](https://doi.org/10.1016/S0083-6729(07)00001-5) [6729\(07\)00001-5](https://doi.org/10.1016/S0083-6729(07)00001-5)
- <span id="page-15-30"></span>34. Ferland G, Sadowski JA, O'Brien ME (1993) Dietary induced subclinical vitamin K deficiency in normal human subjects. J Clin Invest 91:1761–1768.<https://doi.org/10.1172/JCI116386>
- <span id="page-15-31"></span>35. Booth SL, Martini L, Peterson JW et al (2003) Dietary phylloquinone depletion and repletion in older women. J Nutr 133:2565– 2569.<https://doi.org/10.1093/jn/133.8.2565>
- <span id="page-15-32"></span>36. Cranenburg EC, Koos R, Schurgers LJ et al (2010) Characterisation and potential diagnostic value of circulating matrix Gla protein (MGP) species. Thromb Haemost 104:811–822. [https://](https://doi.org/10.1160/TH09-11-0786) [doi.org/10.1160/TH09-11-0786](https://doi.org/10.1160/TH09-11-0786)
- 37. Shea MK, O'Donnell CJ, Vermeer C et al (2011) Circulating uncarboxylated matrix gla protein is associated with vitamin K nutritional status, but not coronary artery calcium, in older adults. J Nutr 141:1529–1534. [https://doi.org/10.3945/jn.111.](https://doi.org/10.3945/jn.111.139634) [139634](https://doi.org/10.3945/jn.111.139634)
- <span id="page-15-33"></span>38. Dalmeijer GW, van der Schouw YT, Magdeleyns E et al (2012) The effect of menaquinone-7 supplementation on circulating species of matrix Gla protein. Atherosclerosis 225:397–402. [https://](https://doi.org/10.1016/j.atherosclerosis.2012.09.019) [doi.org/10.1016/j.atherosclerosis.2012.09.019](https://doi.org/10.1016/j.atherosclerosis.2012.09.019)
- <span id="page-15-34"></span>39. Elliott MJ, Booth SL, Hopman WM et al (2014) Assessment of potential biomarkers of subclinical vitamin K defciency in patients with end-stage kidney disease. Can J Kidney Health Dis 1:13.<https://doi.org/10.1186/2054-3581-1-13>
- 40. Kuwabara A, Fujii M, Kawai N et al (2011) Bone is more susceptible to vitamin K defciency than liver in the institutionalized elderly. Asia Pac J Clin Nutr 20:50–55
- <span id="page-16-0"></span>41. Rapp N, Brandenburg VM, Kaesler N et al (2021) Hepatic and vascular vitamin K status in patients with high cardiovascular risk. Nutrients.<https://doi.org/10.3390/nu13103490>
- <span id="page-16-1"></span>42. Urayama S, Kawakami A, Nakashima T et al (2000) Efect of vitamin K2 on osteoblast apoptosis: vitamin K2 inhibits apoptotic cell death of human osteoblasts induced by Fas, proteasome inhibitor, etoposide, and staurosporine. J Lab Clin Med 136:181–193.<https://doi.org/10.1067/mlc.2000.108754>
- <span id="page-16-2"></span>43. Ichikawa T, Horie-Inoue K, Ikeda K et al (2007) Vitamin K2 induces phosphorylation of protein kinase A and expression of novel target genes in osteoblastic cells. J Mol Endocrinol 39:239–247. <https://doi.org/10.1677/JME-07-0048>
- <span id="page-16-3"></span>44. Li W, Zhang S, Liu J et al (2019) Vitamin K2 stimulates MC3T3E1 osteoblast diferentiation and mineralization through autophagy induction. Mol Med Rep 19:3676–3684. [https://doi.](https://doi.org/10.3892/mmr.2019.10040) [org/10.3892/mmr.2019.10040](https://doi.org/10.3892/mmr.2019.10040)
- <span id="page-16-4"></span>45. Akbulut AC, Wasilewski GB, Rapp N et al (2020) Menaquinone-7 supplementation improves osteogenesis in pluripotent stem cell derived mesenchymal stem cells. Front Cell Dev Biol 8:618760.<https://doi.org/10.3389/fcell.2020.618760>
- <span id="page-16-5"></span>46. Yamaguchi M, Weitzmann MN (2011) Vitamin K2 stimulates osteoblastogenesis and suppresses osteoclastogenesis by suppressing NF-kappaB activation. Int J Mol Med 27:3–14. [https://](https://doi.org/10.3892/ijmm.2010.562) [doi.org/10.3892/ijmm.2010.562](https://doi.org/10.3892/ijmm.2010.562)
- <span id="page-16-6"></span>47. Kameda T, Miyazawa K, Mori Y et al (1996) Vitamin K2 inhibits osteoclastic bone resorption by inducing osteoclast apoptosis. Biochem Biophys Res Commun 220:515–519. [https://doi.org/](https://doi.org/10.1006/bbrc.1996.0436) [10.1006/bbrc.1996.0436](https://doi.org/10.1006/bbrc.1996.0436)
- <span id="page-16-7"></span>48. Atkins GJ, Welldon KJ, Wijenayaka AR et al (2009) Vitamin K promotes mineralization, osteoblast-to-osteocyte transition, and an anticatabolic phenotype by {gamma}-carboxylation-dependent and -independent mechanisms. Am J Physiol Cell Physiol 297:C1358–C1367.<https://doi.org/10.1152/ajpcell.00216.2009>
- <span id="page-16-8"></span>49. Hauschka PV, Lian JB, Cole DE et al (1989) Osteocalcin and matrix Gla protein: vitamin K-dependent proteins in bone. Physiol Rev 69:990–1047. [https://doi.org/10.1152/physrev.1989.69.3.](https://doi.org/10.1152/physrev.1989.69.3.990) [990](https://doi.org/10.1152/physrev.1989.69.3.990)
- <span id="page-16-9"></span>50. Beulens JW, Booth SL, van den Heuvel EG et al (2013) The role of menaquinones (vitamin  $K(2)$ ) in human health. Br J Nutr 110:1357–1368. <https://doi.org/10.1017/S0007114513001013>
- <span id="page-16-10"></span>51. Moriishi T, Ozasa R, Ishimoto T et al (2020) Osteocalcin is necessary for the alignment of apatite crystallites, but not glucose metabolism, testosterone synthesis, or muscle mass. PLoS Genet 16:e1008586. <https://doi.org/10.1371/journal.pgen.1008586>
- <span id="page-16-11"></span>52. Azuma K, Shiba S, Hasegawa T et al (2015) Osteoblast-specifc gamma-glutamyl carboxylase-defcient mice display enhanced bone formation with aberrant mineralization. J Bone Miner Res 30:1245–1254.<https://doi.org/10.1002/jbmr.2463>
- <span id="page-16-12"></span>53. Murshed M, Schinke T, McKee MD et al (2004) Extracellular matrix mineralization is regulated locally; diferent roles of two gla-containing proteins. J Cell Biol 165:625–630. [https://doi.org/](https://doi.org/10.1083/jcb.200402046) [10.1083/jcb.200402046](https://doi.org/10.1083/jcb.200402046)
- <span id="page-16-13"></span>54. Zhang Y, Zhao L, Wang N et al (2019) Unexpected role of matrix gla protein in osteoclasts: inhibiting osteoclast diferentiation and bone resorption. Mol Cell Biol. [https://doi.org/10.1128/MCB.](https://doi.org/10.1128/MCB.00012-19) [00012-19](https://doi.org/10.1128/MCB.00012-19)
- <span id="page-16-14"></span>55. Nakamura YS, Hakeda Y, Takakura N et al (1998) Tyro 3 receptor tyrosine kinase and its ligand, Gas6, stimulate the function of osteoclasts. Stem Cells 16:229–238. [https://doi.org/10.1002/](https://doi.org/10.1002/stem.160229) [stem.160229](https://doi.org/10.1002/stem.160229)
- <span id="page-16-15"></span>56. Ichikawa T, Horie-Inoue K, Ikeda K et al (2006) Steroid and xenobiotic receptor SXR mediates vitamin K2-activated transcription of extracellular matrix-related genes and collagen accumulation in osteoblastic cells. J Biol Chem 281:16927–16934. <https://doi.org/10.1074/jbc.M600896200>
- <span id="page-16-16"></span>57. Konno Y, Moore R, Kamiya N et al (2010) Nuclear xenobiotic receptor PXR-null mouse exhibits hypophosphatemia and represses the Na/Pi-cotransporter SLC34A2. Pharmacogenet Genomics 20:9–17. [https://doi.org/10.1097/FPC.0b013e3283](https://doi.org/10.1097/FPC.0b013e328333bb28) [33bb28](https://doi.org/10.1097/FPC.0b013e328333bb28)
- <span id="page-16-17"></span>58. Azuma K, Casey SC, Ito M et al (2010) Pregnane X receptor knockout mice display osteopenia with reduced bone formation and enhanced bone resorption. J Endocrinol 207:257–263. <https://doi.org/10.1677/JOE-10-0208>
- <span id="page-16-18"></span>59. Muszynska M, Ambrozewicz E, Gegotek A et al (2020) Protective efects of vitamin K compounds on the proteomic profle of osteoblasts under oxidative stress conditions. Molecules. [https://](https://doi.org/10.3390/molecules25081990) [doi.org/10.3390/molecules25081990](https://doi.org/10.3390/molecules25081990)
- <span id="page-16-19"></span>60. Poon CC, Li RW, Seto SW et al (2015) In vitro vitamin K(2) and 1alpha,25-dihydroxyvitamin D(3) combination enhances osteoblasts anabolism of diabetic mice. Eur J Pharmacol 767:30–40. <https://doi.org/10.1016/j.ejphar.2015.09.048>
- <span id="page-16-20"></span>61. Maria S, Swanson M H, Enderby L T, et al. (2017) Melatoninmicronutrients Osteopenia Treatment Study (MOTS): a translational study assessing melatonin, strontium (citrate), vitamin D3 and vitamin K2 (MK7) on bone density, bone marker turnover and health related quality of life in postmenopausal osteopenic women following a one-year double-blind RCT and on osteoblast-osteoclast co-cultures. Aging (Albany NY) 9:256–285.<https://doi.org/10.18632/aging.101158>
- <span id="page-16-21"></span>62. Katsuyama M, Demura M, Katsuyama H et al (2017) Genistein and menaquinone-4 treatment-induced alterations in the expression of mRNAs and their products are benefcial to osteoblastic MC3T3-E1 cell functions. Mol Med Rep 16:873–880. <https://doi.org/10.3892/mmr.2017.6632>
- <span id="page-16-22"></span>63. Nagura N, Komatsu J, Iwase H et al (2015) Efects of the combination of vitamin K and teriparatide on the bone metabolism in ovariectomized rats. Biomed Rep 3:295–300. [https://doi.org/](https://doi.org/10.3892/br.2015.431) [10.3892/br.2015.431](https://doi.org/10.3892/br.2015.431)
- <span id="page-16-23"></span>64. Li H, Zhou Q, Bai BL et al (2018) Efects of combined human parathyroid hormone (1–34) and menaquinone-4 treatment on the interface of hydroxyapatite-coated titanium implants in the femur of osteoporotic rats. J Bone Miner Metab 36:691–699. <https://doi.org/10.1007/s00774-017-0893-9>
- <span id="page-16-24"></span>65. Weng SJ, Yan DY, Gu LJ et al (2019) Combined treatment with vitamin K2 and PTH enhanced bone formation in ovariectomized rats and increased diferentiation of osteoblast in vitro. Chem Biol Interact 300:101–110. [https://doi.org/10.1016/j.cbi.](https://doi.org/10.1016/j.cbi.2019.01.012) [2019.01.012](https://doi.org/10.1016/j.cbi.2019.01.012)
- <span id="page-16-25"></span>66. Zhao B, Zhao W, Wang Y et al (2018) Prior administration of vitamin K2 improves the therapeutic efects of zoledronic acid in ovariectomized rats by antagonizing zoledronic acidinduced inhibition of osteoblasts proliferation and mineralization. PLoS ONE 13:e0202269. [https://doi.org/10.1371/journ](https://doi.org/10.1371/journal.pone.0202269) [al.pone.0202269](https://doi.org/10.1371/journal.pone.0202269)
- <span id="page-16-26"></span>67. Diegel CR, Hann S, Ayturk UM et al (2020) An osteocalcindeficient mouse strain without endocrine abnormalities. PLoS Genet 16:e1008361. [https://doi.org/10.1371/journal.pgen.](https://doi.org/10.1371/journal.pgen.1008361) [1008361](https://doi.org/10.1371/journal.pgen.1008361)
- <span id="page-16-27"></span>68. Ronning SB, Pedersen ME, Berg RS et al (2018) Vitamin K2 improves proliferation and migration of bovine skeletal muscle cells in vitro. PLoS ONE 13:e0195432. [https://doi.org/10.](https://doi.org/10.1371/journal.pone.0195432) [1371/journal.pone.0195432](https://doi.org/10.1371/journal.pone.0195432)
- <span id="page-16-28"></span>69. Vos M, Esposito G, Edirisinghe JN et al (2012) Vitamin K2 is a mitochondrial electron carrier that rescues pink1 defciency. Science 336:1306–1310. [https://doi.org/10.1126/science.12186](https://doi.org/10.1126/science.1218632) [32](https://doi.org/10.1126/science.1218632)
- <span id="page-16-29"></span>70. Boengler K, Kosiol M, Mayr M et al (2017) Mitochondria and ageing: role in heart, skeletal muscle and adipose tissue. J Cachexia Sarcopenia Muscle 8:349–369. [https://doi.org/10.](https://doi.org/10.1002/jcsm.12178) [1002/jcsm.12178](https://doi.org/10.1002/jcsm.12178)
- <span id="page-17-1"></span>71. Lee NK, Sowa H, Hinoi E et al (2007) Endocrine regulation of energy metabolism by the skeleton. Cell 130:456–469. [https://](https://doi.org/10.1016/j.cell.2007.05.047) [doi.org/10.1016/j.cell.2007.05.047](https://doi.org/10.1016/j.cell.2007.05.047)
- <span id="page-17-2"></span>72. Ducy P, Desbois C, Boyce B et al (1996) Increased bone formation in osteocalcin-defcient mice. Nature 382:448–452. [https://](https://doi.org/10.1038/382448a0) [doi.org/10.1038/382448a0](https://doi.org/10.1038/382448a0)
- <span id="page-17-3"></span>73. Lin X, Brennan-Speranza TC, Levinger I et al (2018) Undercarboxylated osteocalcin: experimental and human evidence for a role in glucose homeostasis and muscle regulation of insulin sensitivity. Nutrients. [https://doi.org/10.3390/nu100](https://doi.org/10.3390/nu10070847) [70847](https://doi.org/10.3390/nu10070847)
- <span id="page-17-4"></span>74. Moore AE, Kim E, Dulnoan D et al (2020) Serum vitamin K1 (phylloquinone) is associated with fracture risk and hip strength in post-menopausal osteoporosis: A cross-sectional study. Bone 141:115630. <https://doi.org/10.1016/j.bone.2020.115630>
- <span id="page-17-6"></span>75. Tejero S, Cejudo P, Quintana-Gallego E et al (2016) The role of daily physical activity and nutritional status on bone turnover in cystic fbrosis: a cross-sectional study. Braz J Phys Ther 20:206–212. <https://doi.org/10.1590/bjpt-rbf.2014.0154>
- <span id="page-17-5"></span>76. Finnes TE, Lofthus CM, Meyer HE et al (2016) A combination of low serum concentrations of vitamins K1 and D is associated with increased risk of hip fractures in elderly Norwegians: a NOREPOS study. Osteoporos Int 27:1645–1652. [https://doi.](https://doi.org/10.1007/s00198-015-3435-0) [org/10.1007/s00198-015-3435-0](https://doi.org/10.1007/s00198-015-3435-0)
- <span id="page-17-7"></span>77. Huang ZB, Wan SL, Lu YJ et al (2015) Does vitamin K2 play a role in the prevention and treatment of osteoporosis for postmenopausal women: a meta-analysis of randomized controlled trials. Osteoporos Int 26:1175–1186. [https://doi.org/10.1007/](https://doi.org/10.1007/s00198-014-2989-6) [s00198-014-2989-6](https://doi.org/10.1007/s00198-014-2989-6)
- <span id="page-17-8"></span>78. Ronn SH, Harslof T, Pedersen SB et al (2016) Vitamin K2 (menaquinone-7) prevents age-related deterioration of trabecular bone microarchitecture at the tibia in postmenopausal women. Eur J Endocrinol 175:541–549. [https://doi.org/10.1530/](https://doi.org/10.1530/EJE-16-0498) [EJE-16-0498](https://doi.org/10.1530/EJE-16-0498)
- <span id="page-17-9"></span>79. Ronn SH, Harslof T, Oei L et al (2021) The efect of vitamin MK-7 on bone mineral density and microarchitecture in postmenopausal women with osteopenia, a 3-year randomized, placebocontrolled clinical trial. Osteoporos Int 32:185–191. [https://doi.](https://doi.org/10.1007/s00198-020-05638-z) [org/10.1007/s00198-020-05638-z](https://doi.org/10.1007/s00198-020-05638-z)
- <span id="page-17-10"></span>80. Tanaka S, Miyazaki T, Uemura Y et al (2017) Comparison of concurrent treatment with vitamin K2 and risedronate compared with treatment with risedronate alone in patients with osteoporosis: Japanese Osteoporosis Intervention Trial-03. J Bone Miner Metab 35:385–395.<https://doi.org/10.1007/s00774-016-0768-5>
- <span id="page-17-11"></span>81. Torbergsen AC, Watne LO, Frihagen F et al (2019) Efects of nutritional intervention upon bone turnover in elderly hip fracture patients. Randomized controlled trial Clin Nutr ESPEN 29:52–58.<https://doi.org/10.1016/j.clnesp.2017.11.012>
- <span id="page-17-12"></span>82. Morato-Martinez M, Lopez-Plaza B, Santurino C et al (2020) A dairy product to reconstitute enriched with bioactive nutrients stops bone loss in high-risk menopausal women without pharmacological treatment. Nutrients. [https://doi.org/10.3390/nu120](https://doi.org/10.3390/nu12082203) [82203](https://doi.org/10.3390/nu12082203)
- <span id="page-17-13"></span>83. Barnuevo MD, Marhuenda J, Aldeguer M et al (2018) In vivo randomized trial of three marketed milk preparations enriched with calcium and vitamins (D and K) on bone mass and bone turnover markers from biological fuids in premenopausal Caucasian women. Nutr Hosp 35:1174–1185
- <span id="page-17-14"></span>84. Suzuki Y, Maruyama-Nagao A, Sakuraba K et al (2017) Level of serum undercarboxylated osteocalcin correlates with bone quality assessed by calcaneal quantitative ultrasound sonometry in young Japanese females. Exp Ther Med 13:1937–1943. [https://](https://doi.org/10.3892/etm.2017.4206) [doi.org/10.3892/etm.2017.4206](https://doi.org/10.3892/etm.2017.4206)
- <span id="page-17-15"></span>85. Fusaro M, Cianciolo G, Brandi ML et al (2020) Vitamin K and osteoporosis. Nutrients.<https://doi.org/10.3390/nu12123625>
- <span id="page-17-16"></span>86. Fusaro M, Cianciolo G, Evenepoel P et al (2021) Vitamin K in CKD bone disorders. Calcif Tissue Int 108:476–485. [https://doi.](https://doi.org/10.1007/s00223-020-00792-2) [org/10.1007/s00223-020-00792-2](https://doi.org/10.1007/s00223-020-00792-2)
- <span id="page-17-17"></span>87. Cheung AM, Tile L, Lee Y et al (2008) Vitamin K supplementation in postmenopausal women with osteopenia (ECKO trial): a randomized controlled trial. PLoS Med 5:e196. [https://doi.org/](https://doi.org/10.1371/journal.pmed.0050196) [10.1371/journal.pmed.0050196](https://doi.org/10.1371/journal.pmed.0050196)
- <span id="page-17-18"></span>88. Fusaro M, Noale M, Viola V et al (2012) Vitamin K, vertebral fractures, vascular calcifcations, and mortality: VItamin K Italian (VIKI) dialysis study. J Bone Miner Res 27:2271–2278. <https://doi.org/10.1002/jbmr.1677>
- <span id="page-17-20"></span>89. Mott A, Bradley T, Wright K et al (2019) Efect of vitamin K on bone mineral density and fractures in adults: an updated systematic review and meta-analysis of randomised controlled trials. Osteoporos Int 30:1543–1559. [https://doi.org/10.1007/](https://doi.org/10.1007/s00198-019-04949-0) [s00198-019-04949-0](https://doi.org/10.1007/s00198-019-04949-0)
- <span id="page-17-19"></span>90. Su S, He N, Men P et al (2019) The efficacy and safety of menatetrenone in the management of osteoporosis: a systematic review and meta-analysis of randomized controlled trials. Osteoporos Int 30:1175–1186. [https://doi.org/10.1007/](https://doi.org/10.1007/s00198-019-04853-7) [s00198-019-04853-7](https://doi.org/10.1007/s00198-019-04853-7)
- <span id="page-17-21"></span>91. Ozaki D, Kubota R, Maeno T et al (2021) Association between gut microbiota, bone metabolism, and fracture risk in postmenopausal Japanese women. Osteoporos Int 32:145–156. [https://doi.](https://doi.org/10.1007/s00198-020-05728-y) [org/10.1007/s00198-020-05728-y](https://doi.org/10.1007/s00198-020-05728-y)
- <span id="page-17-22"></span>92. Wang B, Tang J, Weng S et al (2021) Is vitamin K2 a treatment choice for atypical femoral fractures in patients with secondary osteoporosis? J Int Med Res 49:3000605211001643. [https://doi.](https://doi.org/10.1177/03000605211001643) [org/10.1177/03000605211001643](https://doi.org/10.1177/03000605211001643)
- <span id="page-17-23"></span>93. Beaudart C, Reginster JY, Petermans J et al (2015) Quality of life and physical components linked to sarcopenia: The SarcoPhAge study. Exp Gerontol 69:103–110. [https://doi.org/10.1016/j.exger.](https://doi.org/10.1016/j.exger.2015.05.003) [2015.05.003](https://doi.org/10.1016/j.exger.2015.05.003)
- <span id="page-17-24"></span>94. Shea MK, Loeser RF, Hsu FC et al (2016) Vitamin K status and lower extremity function in older adults: the health aging and body composition study. J Gerontol A Biol Sci Med Sci 71:1348–1355.<https://doi.org/10.1093/gerona/glv209>
- <span id="page-17-25"></span>95. Fulton RL, McMurdo ME, Hill A et al (2016) Efect of vitamin K on vascular health and physical function in older people with vascular disease–a randomised controlled trial. J Nutr Health Aging 20:325–333.<https://doi.org/10.1007/s12603-015-0619-4>
- <span id="page-17-26"></span>96. Shea MK, Dawson-Hughes B, Gundberg CM et al (2017) Reducing undercarboxylated osteocalcin with vitamin K supplementation does not promote lean tissue loss or fat gain over 3 years in older women and men: a randomized controlled trial. J Bone Miner Res 32:243–249. <https://doi.org/10.1002/jbmr.2989>
- <span id="page-17-27"></span>97. van Ballegooijen AJ, van Putten SR, Visser M et al (2018) Vitamin K status and physical decline in older adults-The Longitudinal Aging Study Amsterdam. Maturitas 113:73–79. [https://doi.](https://doi.org/10.1016/j.maturitas.2018.04.013) [org/10.1016/j.maturitas.2018.04.013](https://doi.org/10.1016/j.maturitas.2018.04.013)
- <span id="page-17-28"></span>98. Booth SL, Dallal G, Shea MK et al (2008) Efect of vitamin K supplementation on bone loss in elderly men and women. J Clin Endocrinol Metab 93:1217–1223. [https://doi.org/10.1210/](https://doi.org/10.1210/jc.2007-2490) [jc.2007-2490](https://doi.org/10.1210/jc.2007-2490)
- <span id="page-17-29"></span>99. Smith C, Lin X, Scott D et al (2021) Uncovering the bone-muscle interaction and its implications for the health and function of older adults (the wellderly project): protocol for a randomized controlled crossover trial. JMIR Res Protoc 10:e18777. [https://](https://doi.org/10.2196/18777) [doi.org/10.2196/18777](https://doi.org/10.2196/18777)
- <span id="page-17-30"></span>100. Levinger I, Scott D, Nicholson GC et al (2014) Undercarboxylated osteocalcin, muscle strength and indices of bone health in older women. Bone 64:8–12. [https://doi.org/10.1016/j.bone.](https://doi.org/10.1016/j.bone.2014.03.008) [2014.03.008](https://doi.org/10.1016/j.bone.2014.03.008)
- <span id="page-17-0"></span>101. Klapkova E, Cepova J, Dunovska K et al (2018) Determination of vitamins K1, MK-4, and MK-7 in human serum of

postmenopausal women by HPLC with fuorescence detection. J Clin Lab Anal 32:e22381.<https://doi.org/10.1002/jcla.22381>

- <span id="page-18-0"></span>102. Kim MS, Kim ES, Sohn CM (2015) Dietary intake of vitamin K in relation to bone mineral density in Korea adults: The Korea National Health and Nutrition Examination Survey (2010–2011). J Clin Biochem Nutr 57:223–227. [https://doi.org/10.3164/jcbn.](https://doi.org/10.3164/jcbn.14-98) [14-98](https://doi.org/10.3164/jcbn.14-98)
- <span id="page-18-1"></span>103. Popko J, Karpinski M, Chojnowska S et al (2018) Decreased levels of circulating carboxylated osteocalcin in children with low energy fractures: a pilot study. Nutrients. [https://doi.org/10.](https://doi.org/10.3390/nu10060734) [3390/nu10060734](https://doi.org/10.3390/nu10060734)
- <span id="page-18-2"></span>104. Evenepoel P, Claes K, Meijers B et al (2019) Poor vitamin K status is associated with low bone mineral density and increased fracture risk in end-stage renal disease. J Bone Miner Res 34:262–269. <https://doi.org/10.1002/jbmr.3608>
- <span id="page-18-3"></span>105. Nalevaiko JZ, Marques JVO, Oliveira MF et al (2021) Bone density and quality in patients treated with direct-acting oral anticoagulants versus warfarin. Bone 150:116000. [https://doi.org/10.](https://doi.org/10.1016/j.bone.2021.116000) [1016/j.bone.2021.116000](https://doi.org/10.1016/j.bone.2021.116000)
- <span id="page-18-4"></span>106. Jaghsi S, Hammoud T, Haddad S (2018) Relation between circulating vitamin K1 and osteoporosis in the lumbar spine in syrian post-menopausal women. Open Rheumatol J 12:1–9. [https://doi.](https://doi.org/10.2174/1874312901812010001) [org/10.2174/1874312901812010001](https://doi.org/10.2174/1874312901812010001)
- <span id="page-18-5"></span>107. Ebina K, Noguchi T, Hirao M et al (2016) Comparison of the efects of 12 months of monthly minodronate monotherapy and monthly minodronate combination therapy with vitamin K2 or eldecalcitol in patients with primary osteoporosis. J Bone Miner Metab 34:243–250.<https://doi.org/10.1007/s00774-015-0710-2>
- <span id="page-18-6"></span>108. Shikano K, Kaneko K, Kawazoe M et al (2016) Efficacy of vitamin K2 for glucocorticoid-induced osteoporosis in patients with systemic autoimmune diseases. Intern Med 55:1997–2003. <https://doi.org/10.2169/internalmedicine.55.6230>
- <span id="page-18-7"></span>109. Kodama Y, Okamoto Y, Kubota T et al (2017) Efectiveness of vitamin K2 on osteoporosis in adults with cerebral palsy. Brain Dev 39:846–850.<https://doi.org/10.1016/j.braindev.2017.05.012>
- <span id="page-18-8"></span>110. Bartstra JW, Draaisma F, Zwakenberg SR et al (2021) Six months vitamin K treatment does not affect systemic arterial calcification or bone mineral density in diabetes mellitus 2. Eur J Nutr 60:1691–1699.<https://doi.org/10.1007/s00394-020-02412-z>
- <span id="page-18-9"></span>111. Zhang Y, Liu Z, Duan L et al (2020) Efect of low-dose vitamin K2 supplementation on bone mineral density in middle-aged and elderly Chinese: a randomized controlled study. Calcif Tissue Int 106:476–485.<https://doi.org/10.1007/s00223-020-00669-4>
- <span id="page-18-10"></span>112. Beaudart C, Locquet M, Touvier M et al (2019) Association between dietary nutrient intake and sarcopenia in the SarcoPhAge study. Aging Clin Exp Res 31:815–824. [https://doi.](https://doi.org/10.1007/s40520-019-01186-7) [org/10.1007/s40520-019-01186-7](https://doi.org/10.1007/s40520-019-01186-7)

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.