REVIEW

Bone-Targeted Therapies in Cancer-Induced Bone Disease

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Abstract Cancer-induced bone disease is a major source of morbidity and mortality in cancer patients. Thus, effective bone-targeted therapies are essential to improve disease-free, overall survival and quality of life of cancer patients with bone metastases. Depending of the cancertype, bone metastases mainly involve the modulation of osteoclast and/or osteoblast activity by tumour cells. To inhibit metastatic bone disease effectively, it is imperative to understand its underlying mechanisms and identify the target cells for therapy. If the aim is to prevent bone metastasis, it is essential to target not only bone metastatic features in the tumour cells, but also tumour-nurturing bone microenvironment properties. The currently available bone-targeted agents mainly affect osteoclasts, inhibiting bone resorption (e.g. bisphosphonates, denosumab). Some agents targeting osteoblasts begin to emerge which target osteoblasts (e.g. romosozumab), activating bone formation. Moreover, certain drugs initially thought to target only osteoclasts are now known to have a dual action (activating osteoblasts and inhibiting osteoclasts, e.g. proteasome inhibitors). This review will focus on the evolution of

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bone-targeted therapies for the treatment of cancer-induced bone disease, summarizing preclinical and clinical findings obtained with anti-resorptive and bone anabolic therapies.

Keywords Bone metastasis · RANKL · Sclerostin · DKK1 · Bisphosphonates · mTOR inhibitors · Denosumab · Romosozumab - Radium 223 - Cathepsin k inhibitors c-Src inhibitors

Introduction

Bone metastases are frequent complications of solid tumours [\[1](#page-15-0), [2](#page-15-0)]. For example, the incidence of bone metastases is 65–75% in breast cancer [[1\]](#page-15-0), 65–90% in prostate cancer [[2\]](#page-15-0), 20–25% in renal cell carcinoma [\[3](#page-15-0)], 14–45% in melanoma [\[4](#page-15-0)], 65% in thyroid cancer [\[4](#page-15-0)], 17–64% in lung cancer [\[4](#page-15-0)], 40% in bladder cancer [[4\]](#page-15-0), 10% in colorectal cancer and the incidence of cancer-induced bone disease in cases of multiple myeloma is 70–95% [\[4](#page-15-0)]. Moreover, osteosarcoma, a primary bone cancer accounts for 2% of childhood cancers, and the average 5-year survival rate for patients with localized disease is 60–80%, and for metastatic patients 15–30% [\[5](#page-15-0)]. Therefore, providing appropriate relief and/or preventing the appearance of bone metastases or cancer-induced bone disease in highrisk cancer patients is a major challenge in the field. Adding to the deleterious effects of bone metastases (e.g. bone pain, fractures, spinal cord compression, hypercalcemia), certain anti-cancer therapeutic regimens also worsen bone health (e.g. treatment induced bone loss [\[6](#page-15-0)]). Thus, bone-targeted agents are also needed in such cases to preserve bone quality.

The multifactorial, multi-step nature of bone metastasis has unveiled different cellular and molecular targets in the

metastatic cascade such as osteoclasts, osteoblasts, osteocytes, endothelial cells and other bone microenvironment elements (e.g. immune cells, extracellular matrix). This bone tropism of solid tumours, such as breast and prostate cancers, has been explained by several concepts: 1- the "seed and soil" theory; 2- the osteomimicry of tumour cells; and 3- the establishment of a vicious cycle between tumour and bone cells. Bone marrow acts as a fertile soil for the seeding of circulating tumour cells (CTCs), expressing anchoring receptors and providing growth factors for the establishment and subsequent growth of metastatic foci. Recent developments in the cancer research field highlighted the concept of pre-metastatic niches in which primary tumour cells secrete factors or activate immune cells, that prime distant sites, rendering them more nurturing to CTCs [\[7–9](#page-15-0)]. Several lines of evidence have shown that breast and prostate cancer cells, under the selective pressure of the bone microenvironment, acquire an osteoblast-like phenotype by overexpressing bone-related genes, which ultimately facilitate tumour cells to thrive and subsequently cause overt bone metastases [\[10](#page-15-0), [11\]](#page-16-0). In bone, tumour cells activate osteoclast differentiation and bone-resorbing activity leading to the release of growth factors previously embedded in the bone matrix, stimulating tumour growth, inhibiting osteoblast activity and thereby perpetuating a cycle of osteoclast activation and tumour growth, ultimately leading to bone destruction (Fig. 1) [\[12](#page-16-0)].

Because bone destruction is the main skeletal-related event associated with bone metastasis bone-targeted therapies have been essentially aimed at inhibiting the bone resorptive activity of osteoclasts. Anti-resorptive drugs used in the treatment of bone metastasis in preclinical and clinical settings include bisphosphonates, the anti-RANKL antibody denosumab, cathepsin K inhibitors, mTOR inhibitors and Src inhibitors (Fig. 1). However, with the growing evidence of the involvement of other cell types, particularly in the early steps of the bone metastatic cascade, new targets arose in osteoblasts, osteocytes, endothelial cells, immune cells, etc. Moreover, some osteoclast-targeted agents were also found to affect osteoblasts, macrophages and other cells directly or indirectly [\[13](#page-16-0), [14](#page-16-0)].

This review will cover currently approved bone-targeted therapies in cancer-induced bone disease, bone-targeted therapies currently in the clinical trial pipeline, as well as emerging therapies in the discovery/preclinical phases.

Approved Bone-Targeting Therapies

Molecular-Targeted Therapies

Bisphosphonates

Bisphosphonates (BPs) have a high affinity for hydroxyapatite crystals and mainly target bone-resorbing

Fig. 1 Cancer-induced disease bone marrow microenvironmental targets

osteoclasts. Depending on their structure the bone mineral affinity, potency and mechanisms of action of BPs are different [\[15](#page-16-0)]. Non-nitrogen containing BPs (non-N-BPs) are metabolized to AppCp-type ATP analogues, which are cytotoxic via inhibition of the mitochondrial adenine nucleotide translocase (ANT), thereby inducing apoptosis. Nitrogen containing BPs (N-BPs) inhibit the mevalonate pathway enzyme farnesyl pyrophosphate synthase (FPPS) leading to inhibition of protein prenylation and accumulation of isopentenyl pyrophosphate (IPP) and triphosphoric acid 1-adenosin-5'-yl ester 3-(3-methylbut-3-enyl) ester (ApppI) [\[16](#page-16-0)]. ApppI evokes apoptosis similarly to the ATP analogues formed from non-nitrogen containing BPs. Table 1 provides a list of the different bisphosphonates and their anti-tumour and/or bone-preserving activity.

In the past 2–3 decades, novel cellular targets for BPs emerged both in vitro and in vivo. Among such targets are tumour-associated macrophages [[26\]](#page-16-0), neutrophils [[27\]](#page-16-0), $\gamma\delta$ -T cells [\[28–31](#page-16-0)], endothelial cells [[24\]](#page-16-0) and osteoblasts [\[32](#page-16-0)]. The direct anti-tumour effects of N-BPs relied on frequent low doses of the drugs, which lead to high concentrations within tumours [\[33](#page-16-0)].

Preclinical evidence demonstrated the benefit of bisphosphonates in the management of established bone metastases of solid tumour and multiple myeloma. This has prompted several clinical trials in the late 90s early 2000s, leading to the EMEA (2001) and FDA (2002) approval of bisphosphonates, particularly zoledronic acid (ZOL) for the prevention of skeletal-related events (SRE) in patients with advanced malignancies involving bone and treatment of tumour-induced hypercalcaemia.

Further preclinical [\[34](#page-16-0)] and clinical studies (Table [2\)](#page-3-0) showed that bisphosphonates prevent bone metastasis as well as have extra-skeletal benefits in defined microenvironmental contexts, such as oestrogen deprivation.

A recent meta-analysis of individual patient data from randomized trials of adjuvant BP use in early breast cancer has further proved a reduction in the risk of bone and other metastases and breast cancer mortality only in older or oestrogen-deprived patients [[35\]](#page-16-0).

Two prospective clinical studies [[36,](#page-16-0) [37](#page-16-0)] explored the anti-tumour potential of ZOL in the neoadjuvant setting. The first study by Winter and colleagues [\[36](#page-16-0)] showed a synergistic effect of ZOL treatment followed by chemotherapy possibly due to an increased apoptosis and reduced proliferation and a reduction of the VEGF levels. The second study [\[37](#page-16-0)], enrolling fifty-three breast cancer patients (thirty-three with locally advanced and twenty with a first bone-only relapse) demonstrated that a single 4 mg dose of ZOL 14 days prior to any further treatment increased the number of apoptotic CTCs and primary tumour cells, reduced tumour and endothelial cell proliferation. ZOL antiangiogenic effects in a neoadjuvant setting were also suggested in prostate cancer patients, where ZOL treatment decreased the number of circulating endothelial and endothelial precursor cells, both markers of ongoing pathological neoangiogenesis [[38\]](#page-16-0).

Additionally, a meta-analysis of randomized trials of ZOL plus neoadjuvant chemotherapy in breast cancer patients revealed benefits in terms of pathological complete response in the breast only for postmenopausal women [\[39](#page-17-0)].

Overall, the use of BPs (mainly oral CLO and intravenous ZOL) in the adjuvant and neoadjuvant settings of large phase-III, prospective clinical trials in early breast cancer shows that these agents exhibit anti-cancer activity in patients with hormone-responsive breast cancer who had low levels of reproductive hormones at study entry, achieved either through natural menopause or ovarian suppression therapy (Table 1). The mechanisms behind the improved overall survival of these patients in a low oestrogen environment who received a BP are unknown. In stark contrast, the use of ZOL in the adjuvant treatment of patients with high-risk, castration-sensitive prostate cancer,

Table 1 Bisphosphonate classes and their anti-tumour and/or bone-preserving activity

BP class	BP	Anti-tumour or bone-preserving activity
Non-N-BPs	Clodronate	In experimental breast cancer bone metastasis, daily regimens decrease skeletal tumour growth $[17]$
$N-BPs$	Alendronate	Decrease breast cancer cell adhesion to bone matrices [18]
	Pamidronate	
	Olpadronate	
	Ibandronate	Decreases breast cancer cell adhesion to bone matrices and bone metastasis via induction of osteoclast and breast cancer cell apoptosis [18, 19]
	Zoledronate	Induces breast, prostate, lung, colon, osteosarcoma and myeloma cancer cell apoptosis $[20-22]$ and decreases osteolytic lesions and bone tumour burden, preserving bone structure in breast cancer bone metastasis models $[23-25]$

BP bisphosphonates, Non-N-BPs non-nitrogen containing BPs, N-BPs nitrogen containing BPs

Clinical trial	Patient population	BP (dose, treatment duration)	Results
Breast cancer			
Oral CLO in adjuvant treatment $[40]$	Operable breast cancer patients	CLO (1600 mg/day po, 3 years)	↑Recurrence-free, bone metastasis-free and non- bone metastasis-free intervals in women > 50 years
Z/ZO-FAST [41, 42]	Postmenopausal early breast cancer patients	Immediate or delayed ZOL $(4 \text{ mg } iv \text{ q})$ 6 months, 5 years)	Preserved BMD and ¹ DFS
AZURE [43]	Early breast cancer patients	ZOL (4 mg <i>iv</i> q 3-4 weeks \times 6, 4 mg <i>iv</i> q 3 months \times 8, 4 mg iv q 6 months, 5 years)	TIDFS and lextra-skeletal metastases in postmenopausal women Bone metastases in the whole population
ABCSG-12 [44]	Premenopausal early breast cancer patients	ZOL $(4 \text{ mg } iv \ q \ 6 \text{ months}, 3 \text{ years})$	TDFS in oestrogen-deprived women
			ZOL Tefficacy of tamoxifen
Prostate cancer			
Adjuvant Effect of IV CLO [45]	Castration-sensitive prostate cancer patients	CLO (1500 mg iv q 3 months for $50-124$ months)	Delayed time to first bone metastasis
MRC PR04, PR05 studies [46]	Castration-sensitive prostate cancer patients	CLO (2080 mg/day po , 5 years)	No prevention of bone metastasis
RADAR [47]	Castration-sensitive prostate cancer patients	ZOL $(4 \text{ mg } iv \ q \ 3 \text{ months}, 18 \text{ months})$	ZOL treatment prevented the sustained BMD loss caused by 18 months of ADT
ZEUS study $[48]$	High-risk, castration- sensitive prostate cancer patients	ZOL (4 mg iv q 3 months, 4 years)	Ineffective in the prevention bone metastasis
STAMPEDE study $[49]$	High-risk, castration- sensitive prostate cancer patients	ZOL (4 mg iv q 3 weeks \times 6, 4 mg iv q 4 weeks, 2 years)	ZOL shows no evidence of survival improvement

Table 2 Effects of bisphosphonates in adjuvant and neoadjuvant settings of breast and prostate cancer clinical trials

BP bisphosphonate, CLO clodronate, q every, po per os, iv intravenously, ZOL zoledronic acid, BMD bone mineral density, DFS disease-free survival, IDFS invasive disease-free survival, OS overall survival, SRE skeletal-related events, ADT androgen-deprivation treatment

regardless of whether these men received or not androgendeprivation therapy, does not provide a benefit on diseasefree survival (Table 2). The reasons for these marked differences in the clinical outcome of breast and prostate cancer patients receiving an adjuvant BP treatment are unclear.

Despite being the gold standard in bone-targeted therapy, bisphosphonates have side-effects, which may limit their use in certain patient groups. The most common adverse effects are fatigue, fever, nausea/vomiting, anaemia, bone/joint pain, osteonecrosis of the jaw and atypical femur fractures. Osteonecrosis of the jaw and atypical femur are rare but preoccupying side-effects that are associated with bisphosphonate long-term use [\[50](#page-17-0)].

Due to their high affinity to hydroxyapatite, bisphosphonates are also exploited as bone-targeting moieties, for decades as radiotracers, and more recently to deliver chemotherapy and other toxic cargo to bone resident tumour cells. So far, the latter approach remains in the

preclinical realm [[51\]](#page-17-0). However, future studies may change this scenario.

Anti-RANKL Antibody (Denosumab)

The triad receptor activator of nuclear factor- $K\beta$ (RANK), RANK ligand (RANKL) and osteoprotegerin (OPG, RANKL decoy receptor) has been shown to regulate osteoclast maturation, differentiation and survival. RANK is expressed by osteoclasts and osteoclast precursors, whereas RANKL is produced by osteoblasts and osteocytes [[52\]](#page-17-0). The generation of the RANK- and RANKL-knockout mice revealed that, other than osteopetrosis, the other tissue compartments that exhibited functional defects were the peripheral lymphatic tissue and the development of the mammary gland during pregnancy [[53\]](#page-17-0). These findings are in line with the observation that activated T cells and mammary epithelial cells express RANK/RANKL [\[54\]](#page-17-0). Additionally, RANK and RANKL mRNAs are expressed in other tissues, including skeletal muscle, thymus, liver, heart, brain and adrenal glands [[54](#page-17-0)].

RANKL binding to RANK activates osteoclastogenesis and promotes osteoclast survival and activity, leading to increased bone resorption, a feature of osteoporosis and breast cancer-induced bone disease. 1,25-dihydroxyvitamin D3, parathyroid hormone-related protein (PTHrP), interleukins 1 and 6 (IL-1, IL-6), tumour necrosis factor (TNF), prolactin, corticosteroids and prostaglandin E2 increase RANKL expression. Oestrogens, calcitonin, transforming growth factor β (TGF β), platelet-derived growth factor (PDGF) and calcium-induced OPG decrease RANKL/ RANK binding thus preventing excessive bone resorption [\[54](#page-17-0)]. Upon RANK/RANKL interaction osteoblasts secrete further RANKL, which initiates a cancer cell osteoblast osteoclast vicious cycle and promotes osteoclastogenesis and osteoclast activity. Increased osteoclast-mediated bone resorption further fuels the vicious cycle by the release of growth factors from the resorbed bone, which stimulate tumour growth (Fig. 2).

Originally, due to the osteoclast-activating role of RANK/RANKL, and the known part played by osteoclasts in mediating osteolytic bone metastasis, OPG a decoy receptor for RANKL was used in preclinical breast cancer bone metastatic models [\[55](#page-17-0)]. Due to the ubiquitous expression of RANK in a variety of tissues, targeting the circulating and local levels of RANKL seems a safer therapeutic approach than directly inhibiting the receptor. OPG-Fc was the first of such attempts, but the induction of an immune response against OPG was a potential safety risk. Its development was therefore discontinued in favour of anti-RANKL human monoclonal antibodies with better pharmacokinetic profiles and higher anti-RANKL affinity/ specificity e.g. denosumab. The fully human anti-RANKL IgG2 antibody, denosumab, binds RANKL as the endogenous decoy receptor OPG, blocking RANKL-RANK interaction and thus inhibiting osteoclastogenesis and osteoclast activity, which in turn reduces bone resorption.

As the murine RANKL is not recognized by human monoclonal antibodies against RANKL, the preclinical development of such therapeutic agents relied on the use of primates (cynomolgus monkey) [[56,](#page-17-0) [57](#page-17-0)] or humanized mice [[58,](#page-17-0) [59\]](#page-17-0). Most of the preclinical evidence of blocking RANKL in oncology derived from OPG-Fc and RANK-Fc treatments [\[60](#page-17-0), [61](#page-18-0)] and only recently made use of the humanized mouse model suitable for denosumab treatments [\[62](#page-18-0)]. A strong inhibition of osteoclast differentiation and activity with these experimental treatments has prompted the use of denosumab in several oncology phase-III clinical trials. These trials focused on the prevention of SRE by denosumab in comparison with ZOL in patients with advanced cancer and bone metastasis, including multiple myeloma [[63–66\]](#page-18-0), breast [[63,](#page-18-0) [66–69\]](#page-18-0), prostate [\[70–73](#page-18-0)] and other metastatic cancers [\[64](#page-18-0), [74\]](#page-18-0). Denosumab was found to be superior to ZOL in delaying the onset of the first SRE in breast and prostate cancer. In other metastatic solid tumours (excluding breast and prostate cancer) and multiple myeloma, denosumab was non-inferior to ZOL [[69\]](#page-18-0). Denosumab has also shown positive results in the treatment of patients in which bone loss is prevalent, including women with breast cancer receiving aromatase inhibitor therapy and men with prostate cancer undergoing androgen-deprivation therapy (Table [3](#page-5-0)). In these placebocontrolled randomized phase-III trials denosumab increases bone mineral density (BMD) and decreases bone resorption (Table [3\)](#page-5-0). Denosumab also increases bone metastasis-free survival and delays time to first bone metastasis compared with placebo in men with non-metastatic castration-resistant prostate cancer at high risk of bone metastasis (prostate-specific antigen $[PSA] \geq 8.0 \mu g/L$ or PSA doubling time ≤ 10.0 months, or both) (Table [3\)](#page-5-0).

An ongoing clinical trial (D-CARE, NCT01077154) is testing denosumab in high-risk early breast cancer patients receiving neoadjuvant or adjuvant therapy (Table [3\)](#page-5-0). Its

Fig. 2 RANK-RANKL signalling in cancer-induced bone disease

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Clinical trial	Patient population	Dose	Results
Breast cancer			
HALT, NCT00089661 [75, 76]	Hormone receptor-positive non- metastatic breast cancer patients	Denosumab (60 mg sc q 6 months) versus placebo plus AI	TBMD and J bone remodelling markers
ABCSG-18 [77]	Postmenopausal breast cancer patients	Denosumab (60 mg sc q 6 months) versus \downarrow AI-induced fractures placebo plus AI	
Prostate cancer			
NCT00089674 [78]	Non-metastatic CRPC patients	Denosumab (60 mg sc q 6 months) plus androgen-deprivation therapy	Rapid and sustained decrease of bone turnover markers by denosumab
Smith et al. $[72]$	Non-metastatic CRPC patients at high risk of bone metastasis	Denosumab (120 mg sc q 4 weeks) versus placebo, plus ADT	Increase bone metastasis-free survival
D-CARE NCT01077154	High-risk early breast cancer patients	Denosumab (120 mg sc q 4 weeks, 6 months, then 120 mg sc q 3 months, 18 months) versus placebo, plus neoadjuvant or adjuvant therapy	August 2017

Table 3 Effects of denosumab in adjuvant settings in breast and prostate cancer clinical trials

sc subcutaneously, q every, AI aromatase inhibitors, ADT androgen-deprivation therapy, BMD bone mineral density, CRPC castration-resistant prostate cancer

estimated primary completion date is August 2017 and its estimated study completion date August 2022.

Cumulative in vitro, in vivo and retrospective clinical data link RANK overexpression by breast cells with stem cell properties, epithelial to mesenchymal transition [\[79](#page-18-0)], malignant transformation and ultimately with disease progression, particularly in progesterone driven breast cancers [\[53](#page-17-0)] and BRCA mutants [[80\]](#page-18-0). Therefore, certain experts suggest the use of denosumab for breast cancer chemoprevention in high-risk women (BRCA1, BRCA2 mutation carriers) as an alternative to radical prophylactic mastectomy. An ongoing prospective clinical trial has started to address the efficacy of such a strategy (BRCA-D trial, ACTRN12614000694617).

In oncology denosumab is currently FDA approved for (1) treatment of adults and skeletally mature adolescents with giant cell tumour of bone that is unresectable or where surgical resection is likely to result in severe morbidity,(2) prevention of SREs (pathological fracture, radiation to bone, spinal cord compression or surgery to bone) in adults with bone metastases from solid tumours and (3) increasing bone mass in patients at high risk for fracture due to ADT for non-metastatic prostate cancer or AI therapy for breast cancer. Similar indications have been approved by EMEA, except for its use in unresectable giant cell tumour of bone and breast cancer patients receiving AI.

The most common adverse effects of denosumab are similar to those of BPs: nausea, diarrhoea, fatigue and osteonecrosis of the jaw, the latter being equally rare [\[81](#page-18-0)].

Mammalian Target of Rapamycin (mTOR) Inhibitors

The dysregulation of the phosphoinositide 3-kinase (PI3K)/ protein kinase B (AKT)/mTOR pathway is a common feature of cancer [[82\]](#page-18-0). Additionally, it has been implicated in normal and pathological osteoclastogenesis [\[83](#page-18-0)]. Indeed, mTOR is an antiapoptotic downstream target of M-CSF, RANKL and TNF- α , which is essential for osteoclast differentiation, survival and activity [\[84](#page-19-0), [85](#page-19-0)]. mTOR inhibition leads to increased OPG expression, osteoclast apoptosis and might also promote osteoblastogenesis [\[86](#page-19-0), [87](#page-19-0)].

Rapamycin and its analogues (e.g. sirolimus, temsirolimus, everolimus, deforolimus) are mTOR inhibitors, which block the translation of survival factors and apoptosis inhibitors [[84\]](#page-19-0).

Preclinical evidence demonstrated an effect of mTOR inhibition in cancer-induced bone diseases. For instance, in the 4T1 orthotopic breast cancer model, rapamycin treatment of tumour-bearing animals reduces the number of osteolytic lesions and increased survival [\[88](#page-19-0)]. In osteoblastic and osteolytic osteosarcoma models, everolimus plus ZOL combination treatment slowed tumour progression and increased bone mass [[89\]](#page-19-0). In prostate cancer, everolimus alone or in combination with docetaxel and/or ZOL decreased tumour burden and cachexia [[90\]](#page-19-0). In oral squamous cell carcinoma, temsirolimus slowed tumour growth and inhibited osteolysis [\[91](#page-19-0)]. In neuroblastoma, despite the known direct effects of mTOR inhibition on osteoclasts and osteoblasts and their respective precursors the overall effects on cancer-induced bone disease may well be a combination of direct anti-tumour effects and microenvironmental effects as the PI3K/AKT/mTOR pathway also regulates cell growth and apoptosis.

Due to its in vivo synergistic effect with other bone resorption inhibitors, chemotherapy and hormonal therapy, several clinical trials evaluated the effect of mTOR inhibitors in advanced cancers. In breast cancer, the efficacy of everolimus vs placebo in combination with trastuzumab plus paclitaxel as first-line treatment for patients with HER2-positive advanced breast cancer (BOLERO-1 trial) was investigated [\[92\]](#page-19-0). Although everolimus did not improve progression-free survival (PFS) of the whole population, compared to placebo there was a 7-month PFS prolongation in hormone receptor negative HER-2 positive breast cancer patients [\[92](#page-19-0)]. The BOLERO-2 trial led to the approval of everolimus in combination with exemestane, for patients with advanced hormone receptor-positive/ HER2-negative (HER2-) breast cancer who progressed on prior endocrine therapy with either letrozole or anastrozole [\[93–95](#page-19-0)]. Additionally, exploratory analyses in BOLERO-2 evaluated the effect of everolimus on bone marker levels and progressive disease in bone [[6\]](#page-15-0). Data obtained show that, compared to exemestane alone, everolimus plus exemestane has beneficial effects on bone turnover and progressive disease in bone in patients, irrespective of the use of bisphosphonates [[6\]](#page-15-0). The effect of everolimus, as a single agent, was also studied in a double-blind, placebocontrolled, phase-II trial in HER2-negative breast cancer patients with bone metastases only (RADAR study) [\[96](#page-19-0)]. The results suggest that patients with bone metastases only may retrieve long-term benefit from everolimus.

The efficacy and safety of everolimus has also been examined in other advanced cancers with bone metastasis, including renal cell carcinoma, non-small cell lung carcinoma and prostate carcinoma [\[97–99](#page-19-0)]. Compared to everolimus alone, the combination of everolimus and ZOL significantly reduces bone resorption markers and prolongs tumour control in renal cell carcinoma patients with bone metastases [[97](#page-19-0)].

Similar synergistic effects of everolimus and ZOL in delaying bone metastasis and prolonging OS were seen in patients with advanced non-small cell lung carcinoma and bone metastases [[99\]](#page-19-0). In contrast, the addition of everolimus to carboplatin demonstrated minimal clinical efficacy in metastatic castrate-resistant prostate cancer patients. However, in this trial, patients were not pre-selected for: PTEN deletion, TSC1 mutations, and lack of pAKT staining, all potential biomarkers for mTOR inhibition response. Thus, the studied population could be biased to everolimus non-responders [\[100](#page-19-0)].

Phase-I clinical trials also explored the use of everolimus in relapsed and/or refractory multiple myeloma patients [[101,](#page-19-0) [102](#page-19-0)]. Although these trials were designed to evaluate feasibility, anti-myeloma activity, defined as clinical benefit, was documented when everolimus was used as a single agent or in combination with lenalidomide [\[102](#page-19-0)].

The adverse events profile of mTOR inhibitors includes stomatitis, infection, rash, non-infectious pneumonitis, hyperglycaemia and hyperlipidaemia, most reversible by dose adjustment and supportive care measures. The metabolic abnormalities are of utmost concern in postmenopausal women, who have an already increased risk for such conditions. Thus, special attention must be paid in the management of this patient population [\[103](#page-19-0)].

Radium-223 and Other Radiopharmaceuticals

External beam radiation therapy has been in clinical practice to treat bone pain in metastatic patients. However, its use is limited to localized disease due to the toxicity to healthy surrounding tissues. Low linear energy transfer β emitter radionucleotides like strontium-89 (calcium mimetic that binds to bone mineral) and bisphosphonateconjugated samarium-153 enabled targeted delivery of radiotherapy to bone and showed benefits in the palliative treatment of bone metastatic patients. The wide radiation range of the latter two agents causes dose-limiting toxicities, thus their use is restricted. On the contrary, high linear energy transfer radionucleotides (particularly α -emitters) such as radium-223 have advantages in terms of relative biological effectiveness, enhanced bystander effect, reduced oxygen enhancement ratio and shorter range of radiation emission. This decreases toxicity and delivers higher energy to target tumour cells even in hypoxic sites, with lower probability to target healthy bone marrow cells [\[104](#page-19-0), [105](#page-19-0)].

Radionucleotides act by inducing DNA double strand breaks which when accumulated in target cells lead to cell death. Due to unknown mechanisms, bystander cells not directly irradiated may also undergo apoptosis, via the formation of foci of γ -phosphorylated histone protein $(\gamma H2A)$ [[104\]](#page-19-0).

Preclinical studies demonstrated radium-223 distribution to osseous sites [[106,](#page-19-0) [107\]](#page-19-0) and its effectiveness in decreasing osteolytic lesions in experimental models of bone metastasis [\[108](#page-19-0)]. In the clinic, a phase-II trial in castration-resistant prostate cancer patients with symptomatic bone metastases (ALSYMPCA) showed that radium-223 improves OS, compared to the placebo group receiving the best standard of care including bisphosphonates or denosumab [\[109](#page-19-0), [110](#page-20-0)]. Additionally, radium-223 improves quality of life, provides pain relief and is well tolerated in this patient population. Results from the ALSYMPCA study led to the FDA approval of radium-223 as a single agent in patients with hormone refractory prostate cancer and bone metastases. Similar encouraging results were observed in metastatic breast cancer [\[111–113](#page-20-0)]. Experimentally, radium-223 alone or in combination with ZOL or doxorubicin, increases OS, decreases osteolysis and skeletal tumour burden and prevents tumourinduced cachexia in a breast cancer model of bone metastasis [[111\]](#page-20-0). In an open-label phase-IIa study on breast cancer patients with bone-dominant disease, who have progressed on endocrine therapy, radium-223 treatment consistently reduced bone resorption and bone formation markers as well as metabolic changes associated with osteoblastic bone metastases [\[113](#page-20-0)]. Large randomized clinical trials in bone metastatic breast cancer patients are ongoing in order to examine the effect of radium-223 in combination with endocrine therapy and exemestane plus everolimus (NCT02258464 and NCT02258451 trials, respectively) [[105](#page-19-0)]. Little is known on the effect of radium-223 in other cancer types. However, trials are ongoing in osteosarcoma (NCT01833520) and thyroid cancer with refractory bone metastases (NCT02390934) [[105\]](#page-19-0).

At the recommended doses (six injections of radium-223 at 50 kBq/kg every 4 weeks) data from the ALSYMPCA trial indicate low-grade diarrhoea and low-grade myelosuppression as the most common adverse effects of radium-223 [[110\]](#page-20-0).

Proteasome Inhibitors (Bortezomib)

The proteasome is an ATP-dependent enzymatic complex responsible for the degradation of ubiquitinated proteins. Malignant cells are generally more sensitive to proteasome inhibition due to their higher proliferation and protein synthesis rates. This is particularly true in multiple myeloma [[114\]](#page-20-0).

Bortezomib was the first proteasome inhibitor approved in the treatment of multiple myeloma. Bortezomib reversibly binds to the chymotrypsin-like subunit of the proteasome inhibiting its catalytic activity. Bortezomib dosedependently inhibits RANKL-induced osteoclastogenesis as it prevents the degradation of the NF- κ B inhibitor I- κ B, which blocks the binding of NF - κ B to the promoters of target genes. Additionally, bortezomib promotes bone formation via an increase in both Runx2 activity and expression of osteoblast markers such as type I collagen, as well as by increasing BMP-2 secretion by osteoblasts and inhibiting multiple myeloma-induced osteocyte death [\[114](#page-20-0)]. Due to its direct anti-tumour and anti-osteoclastic effects and its stimulatory effect on osteoblasts, bortezomib (as a proteasome inhibitor) seems ideal in the management of multiple myeloma. However, cancer patients have intrinsic or acquired resistance to bortezomib therapy. The mechanisms for this resistance are not fully understood, but include: upregulation of constitutive proteasome; point mutations in the bortezomib binding pocket; downregulation of the immunoproteasome; cellular extrusion of bortezomib by the drug efflux transporter P-glycoprotein; activation of pro-survival pathways (particularly in the bone marrow microenvironment: MAPK, insulin like growth factor and Akt/PI3 K/signalling upregulation, increased IL-6 secretion, and expression of miRNA 15a); loss of Xbp1; increased expression of (phosphorylated) MARCKS; and autophagy [\[115](#page-20-0)].

Hundreds of clinical trials with bortezomib in combination therapy regimens are ongoing in multiple myeloma. Addressing them all is out of the scope of this review. A recent meta-analysis demonstrated that bortezomib improves OS, PFS and response rate in multiple myeloma patients, compared to those who did not receive bortezomib. The most common side-effects of bortezomib are increased risk of thrombocytopenia, neutropenia, gastrointestinal toxicities, peripheral neuropathy, infection and fatigue [[116\]](#page-20-0).

Hormone-Related Therapies

Anti-androgens

Sex steroids are essential for bone homeostasis. In males, testosterone is the most abundant sex steroid and it confers protection against osteoporotic fractures mainly via aromatization to oestradiol, which will act on oestrogen receptors. Testosterone also acts on the androgen receptor present in osteoblasts and osteocytes as well as in other cell types [[117\]](#page-20-0). Abiraterone acetate is an irreversible inhibitor of cytochrome P17, which blocks androgen biosynthesis (Fig. [3\)](#page-8-0) leading to undetectable levels of androgens [\[118](#page-20-0)].

Enzalutamide is an inhibitor of the androgen receptor (Fig. [3\)](#page-8-0), which has shown to have bone specific effects, blocking the aromatization independent effects of testosterone in bone [[119\]](#page-20-0).

Prostate cancer is an androgen-dependent pathology, which initially responds to surgical or chemical castration. Androgen altered signalling is a validated therapeutic target in metastatic castration-resistant prostate cancer.

Initially thought to target only prostate cancer cells, abiraterone acetate directly inhibits osteoclastogenesis and promotes osteoblastogenesis and bone matrix deposition [\[120](#page-20-0)]. The use of anti-androgens like abiraterone acetate increases OS and delays time to development of SRE in metastatic castration-resistant prostate cancer patients [\[121](#page-20-0)]. However, despite their approved use in metastatic castration-resistant prostate cancer, the existence of androgen receptor splice variants (e.g. AR.V7) and mutants leads to the quick development of resistance to anti-androgens [\[122](#page-20-0)].

Fig. 3 Simplified mechanisms of action of c-SRC inhibitors, MET inhibitors and anti-androgens in prostate cancer-induced bone disease

Bone-Targeting Therapies in Clinical Trials

Molecular-Targeted Therapies

Cathepsin K Inhibitors

Cathepsin K is a lysosomal cysteine proteinase essential for osteoclast activity. Osteoclast secrete protons providing the optimal acidic microenvironment for cathepsin K degradation of the demineralized collagenous matrix by cleavage at multiple sites and release of N-telopeptide collagen fragments. Moreover, cathepsin K is also expressed by bone metastatic breast and prostate cancer cells, thus cathepsin K activity is elevated in osteolytic bone metastases and its targeting may provide dual cell targeting (osteoclasts and cancer cells) [\[123–125](#page-20-0)]. Additionally, in osteosarcoma patients, cathepsin K seems to be predictive of poor prognosis [[126\]](#page-20-0).

The design of cathepsin K inhibitors was initially challenging due to high similarity of substrate and mechanisms between members of the cysteine cathepsin family. Additionally, species differences (87–88% human/rodent homology) make human cathepsin K inhibitors less potent than rodent cathepsin K inhibitors. The initial cathepsin K inhibitors bound to the catalytic site irreversibly, which made them non-attractive to chronic use. Later generations of inhibitors (e.g. dutacatib, odanacatib, balicatib) reversibly bind to the catalytic site of cathepsin K, blocking substrate binding and subsequent cleavage. Another issue with these drugs is their high lysosomotropism. Prolonged lysosomal trapping of cathepsin K inhibitors leads to offtarget effects and to skin adverse effects, common to

lysosomal targeting drugs [\[127](#page-20-0)]. For instance, balicatib a basic lysosomotropic cathepsin K inhibitor successfully completed a phase-II clinical trial for osteoarthritis (NCT00371670), increasing BMD and decreasing bone resorption, but was discontinued due to rash and morphealike skin changes [\[127](#page-20-0)].

In preclinical models of breast cancer bone metastasis, treatment and preventive protocols with the cathepsin K inhibitor dutacatib (AFG495) 50 mg/kg, twice daily intraperitoneally alone or in combination with ZOL (single dose of 100 μ g/kg) decreased tumour-induced osteolysis and skeletal tumour burden and did not affect primary breast tumour growth [\[124](#page-20-0)]. Cathepsin K inhibition did not affect breast cancer cell proliferation, thus pointing to a microenvironmental effect of decreased bone resorption leading to a less nurturing soil for tumour cell implantation $[124]$ $[124]$. Odanacatib is another cathepsin K inhibitor that is as effective as ZOL to reduce bone resorption in breast cancer patients with bone metastases, as assessed by measuring urinary N-telopeptide of type I collagen [\[128](#page-20-0)]. Interestingly, cathepsin K inhibitor L-235, which is structurally related to odanacatib but with higher potency against the rodent enzyme, was tested in a breast cancer model of bone metastasis and results were very similar to those previously reported for AFG495 [[124,](#page-20-0) [129\]](#page-20-0). Additionally, L-235 inhibited breast cancer cell invasion in vitro, thus pointing to a dual targeting of breast cancer cells and osteoclasts [\[129](#page-20-0)].

The promising preclinical results obtained with nonbasic/non-lysosomotropic cathepsin K inhibitors such as odanacatib led to several clinical trials in osteoporosis and cancer-associated bone metastases. As aforementioned, the safety and efficacy of odanacatib on inhibition of biochemical markers of bone turnover in patients with breast cancer and established bone metastases has been reported [\[128](#page-20-0)]. Similarly, a phase-II study in postmenopausal osteoporosis reported that a 5-year continuous therapy with odanacatib inhibited bone resorption and increased BMD [\[130](#page-20-0)]. A phase-III Long-Term Odanacatib Fracture Trial (LOFT) enrolling 16,713 participants from 387 centres was therefore conducted [\[131](#page-20-0)]. Unfortunately, phase-III results showed that while the drug could reduce osteoporotic fractures, it also increased the risk of atrial fibrillation and stroke. The big pharma Merck, the odanacatib manufacturer, therefore decided to discontinue the development of this drug. For undisclosed reasons the metastatic bone disease trials (NCT01552122, NCT00691899 and NCT00692458) were also withdrawn.

c-Src Inhibitors

c-Src was the first proto-oncogene to be identified and it is a non-receptor protein tyrosine kinase crucial in several cellular processes such as proliferation, migration, invasion and survival. c-Src knockout mice have defective osteoclasts, impaired bone remodelling, are osteopetrotic and toothless [[132\]](#page-20-0). Osteoclastic c-Src activation is important for osteoclast differentiation and activity; it recruits several signalling proteins for motility and cytoskeletal rearrangement (e.g. podosome formation). RANK/RANKL signalling leads to c-Src activation, which in turn triggers PI3 K/Akt/mTOR signalling, promoting osteoclast survival [\[133](#page-20-0)]. Therefore, c-Src inhibition has the potential to impair osteoclast-mediated bone resorption. Additionally, c-Src inhibition has been shown to affect osteoblasts, suppressing proliferation and enhancing differentiation (Fig. [3\)](#page-8-0) [[134\]](#page-20-0).

Several orally active c-Src/multikinase inhibitors are FDA-approved (bosutinib, dasatinib, ponatinib and vandetanib) or are in clinical trials for various malignancies. Due to the high similarity of Src family tyrosine kinases, none of the current inhibitors is uniquely selective for Src, Table [4](#page-10-0) shows their known targets and potential therapeutic indications [[135\]](#page-20-0).

Pharmacological inhibition of c-Src or intra-cardiac injection of dominant-negative kinase dead c-Src breast cancer cells, reduces bone and visceral metastatic incidence as well as morbidity and lethality. Additionally, subcutaneous injection of c-Src overexpressing breast cancer cells originates larger and more proliferating breast tumours than wild type cells. Breast cancer cell c-Src overexpression affects osteoclastogenesis (via osteoblast inhibition) and endothelial proliferation. Thus, c-Src pharmacological inhibition may decrease bone metastasis by acting directly in tumour cells, osteoclasts, osteoblasts and endothelial cells [[136\]](#page-20-0).

In addition to osteoclasts, platelets and tumour cells, c-Src is also expressed by neurons. Therefore, a recent preclinical study evaluated the use of saracatinib in cancerinduced bone pain. Intra-tibial injection of rat mammary cancer cells induced thermal hyperalgesia and mechanical allodynia via phosphorylation of the GluN1 subunit of the N-methyl-D-aspartate receptor. Saracatinib treatment reduced this phosphorylation levels and reversed the thermal hyperalgesia while having no anti-tumour or bone preservation effect at any of the doses used [\[137](#page-20-0)]. This prompted the initiation of a clinical trial for the use of saracatinib in cancer-induced bone pain NCT02085603.

Preclinical studies with the PC-3 human bone metastatic prostate cancer model and two different c-Src inhibitors bosutinib [[142\]](#page-21-0) and saracatinib [[143\]](#page-21-0) demonstrated that c-Src inhibition decreases prostate cancer cell proliferation, migration and invasion in vitro and reduces tumour burden (orthotopically and in bone) and tumour-induced osteolysis in bone metastatic animals. Moreover, c-Src inhibition decreased the phosphorylation levels of several signalling

Inhibitor	Known targets	Potential therapeutic indications	Clinical trials in bone disease
Bosutinib	BCR-Abl, Src, Lyn, Hck, Kit, PDGFR	Ph ⁺ , CML, ALL, breast cancer, glioblastoma	Yes [138]
Dasatinib	BCR-Abl, Src, Fyn, Yes, Lck, Arg, Kit, EphA2, EGFR, PDGFR β	Ph ⁺ , CML, ALL, breast, colorectal, endometrial, head and Yes [139–141] neck, ovarian, and small cell lung cancers, glioblastoma, melanoma and NSCLC	
Ponatinib	BCR-Abl, Src family kinases, VEGFR, PDGFR, FGFR, Eph, Kit, RET, Tie2, Flt3	Ph ⁺ , CML, ALL, endometrial, GIST, hepatic biliary, small No cell lung and thyroid cancers	
Vandetanib	RET, Src family kinases, EGFR, VEGFRs, Brk, Tie2, EphRs	Medullary thyroid cancer, breast, head and neck, kidney cancers, NSCLC and several other solid tumours	Yes (NCT00659438)
Saracatinib (AZD0530)	Src. BCR-Abl	Colorectal, gastric, ovarian, small cell lung cancers, NSCLC, and metastatic osteosarcoma in lung	Yes (NCT00559507, NCT00397878. NCT02085603. NCT00558272)

Table 4 c-Src/multikinase inhibitors (adapted from [\[135](#page-20-0)])

 Ph^+ Philadelphia chromosome positive leukaemia, CML chronic myelogenous leukaemia, ALL acute lymphoblastic leukaemia, NSCLC nonsmall cell lung cancers, GIST gastrointestinal stromal tumour

molecules (AKT, mitogen-activated protein kinase MAPK, focal adhesion kinase FAK) as well as the transcription of genes essential for tumour progression (urokinase receptor uPAR, matrix metalloproteinase 2 MMP-2, MMP-6, bone morphogenetic protein 2 BMP-2, BMP-6, interleukin 8 IL-8 and TGF-b) in prostate cancer cells. An orally bioavailable c-Src inhibitor (KX2-391), which targets the substrate binding site instead of the ATP-binding site, as all the other c-Src inhibitors, has been also investigated in castration-resistant prostate cancer patients with bone metastases. KX2-391 40 mg po twice daily did not however have an anti-tumour effect and had only modest effects on inhibition of bone turnover markers [\[144](#page-21-0)]. The study of its pharmacokinetic demonstrated that the median maximum concentration (C_{max}) achieved was inferior to the C_{max} necessary for inhibition of tubulin polymerization, which may explain these modest inhibitory effects [[144](#page-21-0)].

Overall, phase-I/II trials in bone metastatic patients, using dasatinib and saracatinib showed these drugs are safe, well tolerated and have encouraging results in terms of delaying disease progression. The use of these drugs in large phase-III trials is awaited.

Anti-sclerostin Antibodies

Sclerostin (SOST) is an extracellular wingless pathway (Wnt) inhibitor, which acts by binding to low-density lipoprotein receptor-related proteins 5 and 6 (LRP5 and LRP6), preventing Wnt ligands binding and activation of canonical Wnt signalling (Fig. [4\)](#page-11-0) in bone thus decreasing bone formation through osteoblast inhibition. SOST is a protein mainly secreted by osteocytes [[145\]](#page-21-0). Nevertheless,

multiple myeloma cells [\[146](#page-21-0)] and breast cancer cells also secrete SOST [[147](#page-21-0)], SOST becomes therefore an interesting target in cancer-induced bone disease, as SOST inhibition could potentially lead to increased bone. For example, anastrozole treatment of postmenopausal women with hormone receptor-positive non-metastatic early breast cancer leads to increased SOST serum levels [[148\]](#page-21-0). This suggests a role for osteocytes in bone turnover of breast cancer patients, which could be therefore modulated by anti-SOST therapy. Similarly, in symptomatic multiple myeloma patients, high SOST circulating levels positively correlated with altered bone remodelling and advanced disease [[149,](#page-21-0) [150](#page-21-0)].

The role of SOST in prostate cancer is more controversial. Studies show an increase of serum SOST levels in prostate cancer patients, particularly those under androgendeprivation therapy [\[151](#page-21-0)] or with high bone turnover [\[152](#page-21-0)]. However, a transcriptional and proteomic study shows no significant difference in the local levels of SOST in prostate cancer bone osteolytic and osteoblastic metastases [\[153](#page-21-0)]. A confounding factor could be the use of androgen depriving therapy, which could mask differences between baseline circulating and local SOST levels. Additionally, according to Hudson and colleagues SOST and Dickkopf 1(DKK1, another Wnt inhibitor) have opposing effects in prostate cancer bone metastasis. DKK1 promotes prostate cancer invasion and metastasis and SOST evokes the opposite effect [\[154](#page-21-0)]. This surprising result warrants further investigations into the role of SOST in prostate cancer, especially as studies demonstrate that Wnt activation evokes prostate cancer soft tissue and bone metastasis [\[155](#page-21-0)].

There is some preclinical evidence that anti-SOST antibodies exhibit bone anabolic effects in animal models of bone loss [[156–158\]](#page-21-0). A study in a multiple myeloma model demonstrated that an anti-SOST antibody can increase bone volume in tumour-bearing mice to levels similar to naïve mice and it decreased tumour burden, thus slowing disease progression [\[159](#page-21-0)]. Human SOST neutralizing monoclonal antibodies, such as romosozumab, blosozumab and BPS804 are in clinical development for osteoporosis and osteogenesis imperfecta [\[160–162](#page-21-0)].

Pending of the results of these ongoing clinical trials, anti-SOST therapy in cancer patients with bone metastasis could be then considered.

Emerging Bone-Targeting Therapies

Molecular-Targeted Therapies

Anti-Dikkopf-1 Antibody

Dikkopf-1 (DKK1), another extracellular Wnt inhibitor, produced by breast [[163\]](#page-21-0), prostate and multiple myeloma cancer cells leads to pronounced osteoblast inhibition, which is essential for the establishment of osteolytic bone disease. Wnt inhibition evokes OPG downregulation and RANKL upregulation, thus indirectly controls osteoclastogenesis [[145\]](#page-21-0).

Preclinical evidence in multiple myeloma shows that DKK1 inhibition reduces cancer-induced bone destruction and promotes bone formation [[164\]](#page-21-0). In breast cancer bone metastasis DKK1 inhibition is controversial, as in vitro studies show that DKK1 inhibits tumour growth [\[165](#page-21-0)], which poses issues to the clinical utility of an anti-DKK1 treatment. In a PDX model of human osteosarcoma, the use

of a monoclonal antibody against human DKK1 (BHQ880) slowed the orthotopical tumour growth and inhibited metastasis, while increasing bone differentiation markers [\[166](#page-22-0)].

Phase-I (NCT00741377) and -II (NCT01302886, NCT01337752) clinical trials were conducted in multiple myeloma with the BHQ880, alone or in combination with ZOL and anti-myeloma therapy. The phase-I trial showed that BHQ880 is well tolerated and has potential clinical activity in multiple myeloma [[167\]](#page-22-0). Phase-III clinical trials are missing to further ascertain the clinical utility of BHQ880.

Due to compensatory mechanisms in the expression of Wnt inhibitors upon their inhibition in monotherapy, a bispecific monoclonal antibody targeting SOST and DKK1 is in preclinical development. So far the dual inhibition of the Wnt inhibitors by bispecific antibodies has shown a further increase in bone mass (Fig. [4](#page-11-0)) [\[168](#page-22-0)].

MET/VEGFR Inhibitors

MET also known as hepatocyte growth factor receptor (HGFR) and vascular endothelial growth factor receptors (VEGFRs) are tyrosine protein kinase receptors. Expression of VEGFRs especially by bone marrow-derived endothelial progenitor cells and hematopoietic progenitor cells has been linked with the establishment of pre-metastatic niches. Particularly, $VEGFR1⁺$ cells are required for extravasation, and $VEGFR2⁺$ cells for neovascularization [\[169](#page-22-0)]. Moreover, VEGF/VEGFR signalling contributes to prostate cancer-induced osteoblast differentiation and activity [\[170](#page-22-0)]. MET is overexpressed in prostate cancer primary tumours and bone metastases and correlates with higher tumour grade [[171\]](#page-22-0). MET/VEGFR targeting has thus the potential to impair tumour-induced neoangiogenesis, pre-metastatic niche formation, and osteoblast differentiation and activity (Fig. [3](#page-8-0)).

Initial studies with VEGFR inhibitors (e.g. sunitinib, sorafenib) and anti-VEGF therapy (e.g. bevacizumab) evoked an increased invasion and metastasis probably due to a rebound vascularization via activation of the MET pathway.

Cabozantinib is a tyrosine protein kinase receptor inhibitor, which preferentially targets VEGFR2 and MET. AXL, FLT-3, KIT and RET tyrosine kinase receptors are also inhibited but with less potency [\[172](#page-22-0)]. Cabozantinib treatment suppresses tumour growth and angiogenesis in multiple tumour types. Contrary to VEGFR/VEGF inhibition, cabozantinib also decreases metastasis [[172\]](#page-22-0). In bone metastatic prostate cancer models, cabozantinib inhibited bone- and subcutaneous tumour growth. Additionally, cabozantinib altered bone remodelling in tumour free and tumour-bearing bones. In tumour free bones, cabozantinib increased bone volume/tissue volume ratio (BV/TV). In tumour-bearing bones, cabozantinib effects on bone remodelling were dependent of the tumour type: in LuCaP 23.1 tumours (osteoblastic, androgen sensitive) cabozantinib decreased BV/TV; in C4-2B tumours (mixed osteoblastic/osteolytic, castration resistant) it tended to increase BV/TV. The authors speculated that the different effects seen in the LuCap 23.1 and C4-2B models were due to the higher bone remodelling induced by the former, thus an effect on the tumour growth (smaller bone tumours, lower osteoblastic reaction, lower bone formation) led to a decrease on BV/TV. However, since the treatment also affected normal bone remodelling, one should consider that, the overall effect is a combination of cabozantinib on tumour and bone cells [[173\]](#page-22-0). Dai and colleagues have obtained similar results with the additional demonstration of an effect of cabozantinib in osteoblasts [[174\]](#page-22-0). Further studies in normal bones have shown that this agent rever-

sibly: increases bone volume; increases osteoblast numbers (in male mice); decreases the number of osteoclasts (in female mice); and alters the bone marrow composition of treated animals, originating vascular ectasia, spillage of mature red blood cells in the extra vascular bone marrow and increasing the number of megakaryocytes [[32\]](#page-16-0). Additional proof for the direct effect of cabozantinib in prostate cancer tumour cells, endothelial cells and osteoblasts was provided by patient derived xenografts and clinical trials [\[175](#page-22-0)].

A study with TAS-115, another MET/VEGFR inhibitor that also inhibits FMS, demonstrated an osteolysis prevention effect (PC-3 model) due to prostate cancer cell and osteoclast targeting (the latter via inhibition of FMS-dependent RANKL-induced pre-osteoclast to osteoclast differentiation) [[176\]](#page-22-0). Similar results emerged from a model of bone metastatic lung carcinoma [\[177](#page-22-0)].

At least eight clinical trials of cabozantinib in bone metastatic cancers (prostate, breast, lung, multiple myeloma) are currently active. In the phase-III clinical trial in heavily treated (with docetaxel and Abiraterone acetate and/or enzalutamide) bone metastatic castration-resistant prostate cancer patients the results of cabozantinib versus prednisone were somehow disappointing, as the primary outcome of improved OS was not met. Cabozantinib improved bone scan response, radiographic PFS, symptomatic skeletal events, CTCs and bone biomarkers. The investigators argue that patient selection may be an issue and that treatment discontinuation in the cabozantinib group may be a confounding factor [\[178](#page-22-0)]. Nevertheless, the results of this trial led to the termination of several other trials in bone metastatic castration-resistant prostate cancer. Further clinical development of cabozantinib is pending on the use of a specific biomarker to select patients, on the clarification of the best diagnostic tool to determine its effectiveness in bone metastatic castration-resistant prostate cancer (as bone scans seem inappropriate), and of its use in combination therapy [[179,](#page-22-0) [180\]](#page-22-0).

The most common adverse events of cabozantinib reported in the phase-III clinical trial were decreased appetite, nausea, diarrhoea, fatigue, vomiting, asthenia, decreased weight, constipation and anaemia [\[178](#page-22-0)].

Anti-integrins

Integrins are heterodimeric cell-surface receptors that mediate adhesion to the extracellular matrix and immunoglobulin superfamily molecules. Structurally, integrins are composed of non-covalently bound α and β subunits forming 24 different heterodimers. Each has an extracellular domain, a single trans-membrane region and a short cytoplasmic tail. The extracellular domain shows high affinity for a defined RGD-motif expressed by vitronectin, fibronectin, osteopontin and other extracellular matrix components. Ligand binding propagates outside-in and inside-out intracellular signalling. The unique integrin repertoire of a given cell determines the extent of adhesion and migration of that cell in different matrices. Abnormal integrin overexpression by tumour and/or host cells has been associated with tumour proliferation, survival, angiogenesis, migration and metastasis [[181\]](#page-22-0). Particularly, in osteotropic tumours, overexpression of $\alpha \nu \beta$ 3 integrin has been linked with increased bone colonization by breast, prostate and lung cancer cells and osteomimetism by multiple myeloma cells. The interaction of cancer cells with stromal cells and immune cells via $\alpha \nu \beta$ 3 integrin induces the production of cytokines and growth factors that recruit and differentiate osteoclast precursors, thus induc-ing lytic lesions [[182\]](#page-22-0). Other integrins, such as $\alpha \nu \beta 5$, $\alpha \nu \beta 6$, α 5 β 1, α 6 β 4, α 4 β 1, have also been implicated in the tumour progression of different tumour types [[181\]](#page-22-0). Therefore, integrin-targeting strategies either by the use of RGD-motif peptidomimetics or of monoclonal antibodies has reached preclinical and clinical development. $\alpha \nu \beta$ 3 integrin is the predominant integrin in osteoclasts and it is involved in osteoclast attachment to the bone matrix [[183\]](#page-22-0).

Preclinical evidence has established $\alpha \nu \beta$ 3 as a target in breast cancer bone metastasis. Tumour $\alpha v \beta 3$ [[184,](#page-22-0) [185\]](#page-22-0) expression was essential for bone homing but not for bone colonization of breast and ovarian cancer cells. Short-term preventive treatment with a peptidomimetic antagonist of $\alpha v\beta3$ (PSK1404) inhibited tumour cell invasion in vivo while it did not inhibited bone resorption in ovariectomized mice. However, tumour-induced bone resorption was inhibited via the decrease in the secretion of osteoclastactivating factors by tumour cells. Continuous treatment, with a dose that inhibits bone resorption in ovariectomized mice, led to an even more effective reduction in skeletal

tumour burden probably due to a dual inhibitory effect in tumour cells and osteoclasts [[184\]](#page-22-0).

Treatment with the selective peptidomimetic inhibitor for av integrins, cilengitide, leads to decreased lung metastasis in an osteosarcoma model. In osteosarcoma patient specimens, $\alpha \nu \beta 5$ and $\alpha \nu \beta 3$ integrins are expressed by tumour cells and stromal cells, respectively. Dual inhibition of these integrins did not however affect primary tumour growth. In contrast, it decreased lung metastasis, thus advocating for a role of cilengitide in the treatment metastatic osteosarcoma patients [[186\]](#page-22-0).

Inhibition of α 5 β 1 integrin with a novel peptidomimetic antagonist also showed preclinical benefits in breast cancer cell invasion and angiogenesis, decreasing lung colonization and bone metastasis progression [\[187](#page-22-0)].

A peptidomimetic of the α 2 β 1 integrin-binding domain is efficacious at inhibiting ovariectomy induced bone loss, and impairs breast primary tumour growth and bone metastasis in mice. Combination with a suboptimal dose of doxorubicin increased OS to levels similar to the optimal doxorubicin dose [[188\]](#page-22-0).

A Phase-II clinical trial of MEDI-522 (also known as etaracizumab), a human monoclonal antibody directed against the human $\alpha v\beta3$ integrin, in combination with docetaxel, prednisone, and ZOL in the treatment of patients with metastatic castration-resistant prostate cancer has been completed (NCT00072930) although no results are publicly available. Phase-I/II Clinical studies in colorectal cancer and melanoma have also been completed. The outcome of phase-II clinical trials showed increased median survival of metastatic melanoma patients with minimal side-effects. An in vitro study demonstrated that anti- α v β 3 integrin antibody treatment reduced bone resorption without affecting osteoclastogenesis, simply by the inhibition of osteoclast attachment to bone surfaces [[189\]](#page-22-0).

Cilengitide in asymptomatic bone metastatic castrationresistant prostate cancer patients was well tolerated in a phase-II clinical trial, and the higher dose had a modest clinical effect in stabilizing disease, with no apparent effects on bone markers [\[190](#page-22-0)].

Overall, no clinical trials point to an efficacy of an integrin-targeting strategy being efficient in the management of cancer-induced bone disease, at least not in monotherapy. However, large phase-III clinical trials are lacking. Additionally, integrin targeting could be of use in theranostics. Integrin peptidomimetic inhibitors conjugated with radioligands have been used to target tumour and tumour endothelial cells aiding imaging and targeted radiotherapy in breast [[191\]](#page-22-0) and prostate cancer [\[192](#page-22-0)].

Hormone-Related Therapies

Activin A Inhibitors

Activin A is a member of the $TGF\beta$ superfamily. In vitro, activin A promotes bone remodelling, increasing both osteoclastogenesis and osteoblastogenesis [[193\]](#page-23-0). In vivo, blocking activin A increases bone formation [[194\]](#page-23-0). Activin A binds to activin receptor type IIA, recruits activin-like kinase 4 which phosphorylates Smad2/3 leading to the nuclear translocation of Smad 4. In cancer, activin A has paradoxical effects. Despite decreasing cell proliferation, activin A promotes migration and invasion of cancer cells in bone matrices. Increased levels of circulating and intratumour activin A are associated with prostate [[195\]](#page-23-0) and breast cancer bone metastasis, as well with multiple myeloma-induced bone disease [\[196](#page-23-0), [197](#page-23-0)]. Moreover, the decreased ratio of inhibins/activins is linked with the increased bone resorption, observed around perimenopause. Activin A is generally secreted by bone marrow stromal cells, inhibiting osteoblastogenesis and increasing osteoclastogenesis. Multiple myeloma cells stimulate this production [\[194](#page-23-0), [198](#page-23-0)]. Inhibition of activin A signalling rescues multiple myeloma- and breast cancerinduced bone lytic disease [[194,](#page-23-0) [196](#page-23-0)].

Cumulative preclinical evidence steered clinical trials in multiple myeloma and metastatic breast cancer (terminated due to slow recruitment) of sotatercept, a recombinant activin receptor type IIA (ActRIIA) ligand trap comprising the extracellular domain of the high-affinity human ActRIIA and the human immunoglobulin G Fc domain which binds activin A/B and other TGF β superfamily members with high affinity. In the phase-II clinical trial sotatercept showed anabolic improvements in bone mineral density, while barely affecting bone resorption. The most serious adverse events seen in sotatercept-treated patients (grade 4) were neutropenia, granulocytopenia, and atrial fibrillation. Excessive increases in haemoglobin levels led to dose interruption in certain patients and suggested the use of sotatercept as an erythropoietic agent [\[199](#page-23-0)].

Lenalidomide, an approved multiple myeloma therapy, was shown to increase activin A production by bone marrow cells and therefore decrease osteoblastogenesis. Preclinical evidence suggested a combination of lenalidomide with sotatercept to promote bone formation. This led to the ongoing clinical trial of the combination treatment of lenalidomide or pomalidomide with dexamethasone plus sotatercept in refractory multiple myeloma (NCT01562405) [\[200](#page-23-0)].

Conclusion

Thanks to the existing clinically approved bone-targeted therapies, bone metastatic patients currently have fewer skeletal-related events and prolonged disease-free survival than in the era where such agents were unavailable. Table [5](#page-14-0) provides a selection of ongoing clinical trials of bone-targeted therapies in cancer-induced bone disease, whose results may contribute to the approval of new drugs or new indications for currently used agents. With the growing improvement of bone anabolic agents and constant discovery of novel cellular and molecular targets in the bone metastatic cascade, one may expect that in the forthcoming years we will be able to further improve patient overall survival, quality of life and ideally prevent cancer-induced bone disease.

Conflict of interest The authors declare that they have no conflict of interest.

References

- 1. Gerratana L, Fanotto V, Bonotto M, Bolzonello S, Minisini AM, Fasola G, Puglisi F (2015) Pattern of metastasis and outcome in patients with breast cancer. Clin Exp Metastasis 32:125–133
- 2. Sternberg C, Baskin-Bey E, Watson M, Worsfold A, Rider A, Tombal B (2013) Treatment patterns and characteristics of European patients with castration-resistant prostate cancer. BMC Urol 13:58
- 3. Hoffmann NE, Gillett MD, Cheville JC, Lohse CM, Leibovich BC, Blute ML (2008) Differences in organ system of distant metastasis by renal cell carcinoma subtype. J Urol 179:474–477
- 4. Coleman RE (2001) Metastatic bone disease: clinical features, pathophysiology and treatment strategies. Cancer Treat Rev 27:165–176
- 5. Mirabello L, Troisi RJ, Savage SA (2009) International osteosarcoma incidence patterns in children and adolescents, middle ages, and elderly persons. Int J Cancer 125:229–234
- 6. Gnant M, Baselga J, Rugo HS, Noguchi S, Burris HA, Piccart M, Hortobagyi GN, Eakle J, Mukai H, Iwata H, Geberth M, Hart LL, Hadji P, El-Hashimy M, Rao S, Taran T, Sahmoud T, Lebwohl D, Campone M, Pritchard KI (2013) Effect of everolimus on bone marker levels and progressive disease in bone in BOLERO-2. J Natl Cancer Inst 105:654–663
- 7. Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, MacDonald DD, Jin DK, Shido K, Kerns SA, Zhu Z, Hicklin D, Wu Y, Port JL, Altorki N, Port ER, Ruggero D, Shmelkov SV, Jensen KK, Rafii S, Lyden D (2005) VEGFR1 positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. Nature 438:820–827
- 8. Peinado H, Lavotshkin S, Lyden D (2011) The secreted factors responsible for pre-metastatic niche formation: old sayings and new thoughts. Semin Cancer Biol 21:139–146
- 9. Cox TR, Rumney RMH, Schoof EM, Perryman L, Høye AM, Agrawal A, Bird D, Latif NA, Forrest H, Evans HR, Huggins ID, Lang G, Linding R, Gartland A, Erler JT (2015) The hypoxic cancer secretome induces pre-metastatic bone lesions through lysyl oxidase. Nature 522:106–110
- 10. González Á, García de Durango C, Alonso V, Bravo B, Rodríguez de Gortázar A, Wells A, Forteza J, Vidal-Vanaclocha

F (2017) Distinct osteomimetic response of androgen-dependent and independent human prostate cancer cells to mechanical action of fluid flow: prometastatic implications. Prostate 77:321–333

- 11. Tan C, Li G, Tan L, Du X, Li X, He R, Wang Q, Feng Y (2016) Breast cancer cells obtain an osteomimetic feature via epithelialmesenchymal transition that have undergone BMP2/RUNX2 signaling pathway induction. Oncotarget 7:79688–79705
- 12. Guise TA (2002) The vicious cycle of bone metastases. J Musculoskelet Neuronal Interact 2:570–572
- 13. Plotkin LI, Bivi N, Bellido T (2011) A bisphosphonate that does not affect osteoclasts prevents osteoblast and osteocyte apoptosis and the loss of bone strength induced by glucocorticoids in mice. Bone 49:122–127
- 14. Plotkin LI, Manolagas SC, Bellido T (2006) Dissociation of the pro-apoptotic effects of bisphosphonates on osteoclasts from their anti-apoptotic effects on osteoblasts/osteocytes with novel analogs. Bone 39:443–452
- 15. Rogers MJ, Crockett JC, Coxon FP, Mönkkönen J (2011) Biochemical and molecular mechanisms of action of bisphosphonates. Bone 49:34–41
- 16. Mönkkönen H, Auriola S, Lehenkari P, Kellinsalmi M, Hassinen IE, Vepsäläinen J, Mönkkönen J (2006) A new endogenous ATP analog (ApppI) inhibits the mitochondrial adenine nucleotide translocase (ANT) and is responsible for the apoptosis induced by nitrogen-containing bisphosphonates. Br J Pharmacol 147:437–445
- 17. Daubiné F, Le Gall C, Gasser J, Green J, Clézardin P (2007) Antitumor effects of clinical dosing regimens of bisphosphonates in experimental breast cancer bone metastasis. J Natl Cancer Inst 99:322–330
- 18. Van Der Pluijm G, Vloedgraven H, van Beek E, van der Wee-Pals L, Löwik C, Papapoulos S (1996) Bisphosphonates inhibit the adhesion of breast cancer cells to bone matrices in vitro. J Clin Invest 98:698–705
- 19. Hiraga T, Williams PJ, Mundy GR, Yoneda T (2001) The bisphosphonate ibandronate promotes apoptosis in MDA-MB-231 human breast cancer cells in bone metastases. Cancer Res 61:4418–4424
- 20. Yuen T, Stachnik A, Iqbal J, Sgobba M, Gupta Y, Lu P, Colaianni G, Ji Y, Zhu L-, Kim S-, Li J, Liu P, Izadmehr S, Sangodkar J, Bailey J, Latif Y, Mujtaba S, Epstein S, Davies TF, Bian Z, Zallone A, Aggarwal AK, Haider S, New MI, Sun L, Narla G, Zaidi M (2014) Bisphosphonates inactivate human EGFRs to exert antitumor actions. Proc Natl Acad Sci USA 111:17989–17994
- 21. Räikkönen J, Mönkkönen H, Auriola S, Mönkkönen J (2010) Mevalonate pathway intermediates downregulate zoledronic acid-induced isopentenyl pyrophosphate and ATP analog formation in human breast cancer cells. Biochem Pharmacol 79:777–783
- 22. Ory B, Blanchard F, Battaglia S, Gouin F, Redini F, Heymann D (2007) Zoledronic acid activates the DNA S-phase checkpoint and induces osteosarcoma cell death characterized by apoptosisinducing factor and endonuclease-G translocation independently of p53 and retinoblastoma status. Mol Pharmacol 71:333–343
- 23. Brown HK, Ottewell PD, Evans CA, Coleman RE, Holen I (2012) A single administration of combination therapy inhibits breast tumour progression in bone and modifies both osteoblasts and osteoclasts. J Bone Oncol 1:47–56
- 24. Coscia M, Quaglino E, Iezzi M, Curcio C, Pantaleoni F, Riganti C, Holen I, Mönkkönen H, Boccadoro M, Forni G, Musiani P, Bosia A, Cavallo F, Massaia M (2010) Zoledronic acid repolarizes tumour-associated macrophages and inhibits mammary carcinogenesis by targeting the mevalonate pathway. J Cell Mol Med 14:2803–2815
- 25. Melani C, Sangaletti S, Barazzetta FM, Werb Z, Colombo MP (2007) Amino-biphosphonate-mediated MMP-9 inhibition breaks the tumor-bone marrow axis responsible for myeloidderived suppressor cell expansion and macrophage infiltration in tumor stroma. Cancer Res 67:11438–11446
- 26. Junankar S, Shay G, Jurczyluk J, Ali N, Down J, Pocock N, Parker A, Nguyen A, Sun S, Kashemirov B, McKenna CE, Croucher PI, Swarbrick A, Weilbaecher K, Phan TG, Rogers MJ (2015) Real-time intravital imaging establishes tumor-associated Macrophages as the extraskeletal target of bisphosphonate action in cancer. Cancer Discov 5:35–42
- 27. Kalyan S, Chandrasekaran V, Quabius ES, Lindhorst TK, Kabelitz D (2014) Neutrophil uptake of nitrogen-bisphosphonates leads to the suppression of human peripheral blood $\gamma \delta$ T cells. Cell Mol Life Sci 71:2335–2346
- 28. Hewitt RE, Lissina A, Green AE, Slay ES, Price DA, Sewell AK (2005) The bisphosphonate acute phase response: rapid and copious production of proinflammatory cytokines by peripheral blood $\gamma\delta$ T cells in response to aminobisphosphonates is inhibited by statins. Clin Exp Immunol 139:101–111
- 29. Cabillic F, Toutirais O, Lavoué V, De La Pintière CT, Daniel P, Rioux-Leclerc N, Turlin B, Mönkkönen H, Mönkkönen J, Boudjema K, Catros V, Bouet-Toussaint F (2010) Aminobisphosphonate-pretreated dendritic cells trigger successful $V\gamma9V\delta2$ T cell amplification for immunotherapy in advanced cancer patients. Cancer Immunol Immunother 59:1611–1619
- 30. Gutman D, Epstein-Barash H, Tsuriel M, Golomb G (2011) Alendronate liposomes for antitumor therapy: activation of $\gamma\delta$ T cells and inhibition of tumor growth. Adv Exp Med Biol 733:165–179
- 31. Benzaïd I, Mönkkönen H, Stresing V, Bonnelye E, Green J, Mönkkönen J, Touraine JL, Clézardin P (2011) High phosphoantigen levels in bisphosphonate-treated human breast tumors promote $V\gamma9V\delta2$ T-cell chemotaxis and cytotoxicity in vivo. Cancer Res 71:4562–4572
- 32. Haider MT, Hunter KD, Robinson SP, Graham TJ, Corey E, Dear TN, Hughes R, Brown NJ, Holen I (2015) Rapid modification of the bone microenvironment following short-term treatment with Cabozantinib in vivo. Bone 81:581–592
- 33. Clézardin P (2007) Frequent low-dose bisphosphonate therapy. Bone 41:901–902
- 34. Ottewell PD, Wang N, Brown HK, Reeves KJ, Fowles CA, Croucher PI, Eaton CL, Holen I (2014) Zoledronic acid has differential antitumor activity in the pre- and postmenopausal bone microenvironment in vivo. Clin Cancer Res 20:2922–2932
- 35. Early Breast Cancer Trialists' Collaborative Group (EBCTCG) (2015) Adjuvant bisphosphonate treatment in early breast cancer: meta-analyses of individual patient data from randomised trials. Lancet 386:1353–1361
- 36. Winter MC, Wilson C, Syddall SP, Cross SS, Evans A, Ingram CE, Jolley IJ, Hatton MQ, Freeman JV, Mori S, Holen I, Coleman RE (2013) Neoadjuvant chemotherapy with or without zoledronic acid in early breast cancer-a randomized biomarker pilot study. Clin Cancer Res 19:2755–2765
- 37. Foroni C, Milan M, Strina C, Cappelletti M, Fumarola C, Bonelli M, Bertoni R, Ferrero G, Maldotti M, Takano E, Andreis D, Venturini S, Brugnoli G, Petronini PG, Zanoni V, Pritzker L, Pritzker K, Parissenti A, Santini D, Fox SB, Bottini A, Generali D (2014) Pure anti-tumor effect of zoledronic acid in naïve bone-only metastatic and locally advanced breast cancer: proof from the ''biological window therapy''. Breast Cancer Res Treat 144:113–121
- 38. Santini D, Zoccoli A, Gregorj C, Di Cerbo M, Iuliani M, Pantano F, Zamarchi R, Sergi F, Flammia G, Buscarini M, Rizzo S, Cicero G, Russo A, Vincenzi B, Avvisati G, Tonini G (2013) Zoledronic acid induces a significant decrease of circulating

endothelial cells and circulating endothelial precursor cells in the early prostate cancer neoadjuvant setting. Oncology 85:342–347

- 39. Kroep JR, Charehbili A, Coleman RE, Aft RL, Hasegawa Y, Winter MC, Weilbaecher K, Akazawa K, Hinsley S, Putter H, Liefers GJ, Nortier JWR, Kohno N (2016) Effects of neoadjuvant chemotherapy with or without zoledronic acid on pathological response: a meta-analysis of randomised trials. Eur J Cancer 54:57–63
- 40. Paterson AHG, Anderson SJ, Lembersky BC, Fehrenbacher L, Falkson CI, King KM, Weir LM, Brufsky AM, Dakhil S, Lad T, Baez-Diaz L, Gralow JR, Robidoux A, Perez EA, Zheng P, Geyer CE, Swain SM, Costantino JP, Mamounas EP, Wolmark N (2012) Oral clodronate for adjuvant treatment of operable breast cancer (National Surgical Adjuvant Breast and Bowel Project protocol B-34): a multicentre, placebo-controlled, randomised trial. Lancet Oncol 13:734–742
- 41. Brufsky AM, Harker WG, Beck JT, Bosserman L, Vogel C, Seidler C, Jin L, Warsi G, Argonza-Aviles E, Hohneker J, Ericson SG, Perez EA (2012) Final 5-year results of Z-FAST trial: adjuvant zoledronic acid maintains bone mass in postmenopausal breast cancer patients receiving letrozole. Cancer 118:1192–1201
- 42. Coleman R, De Boer R, Eidtmann H, Llombart A, Davidson N, Neven P, Von Minckwitz G, Sleeboom HP, Forbes J, Barrios C, Frassoldati A, Campbell I, Paija O, Martin N, Modi A, Bundred N (2013) Zoledronic acid (zoledronate) for postmenopausal women with early breast cancer receiving adjuvant letrozole (ZO-FAST study): final 60-month results. Ann Oncol 24:398–405
- 43. Coleman R, Cameron D, Dodwell D, Bell R, Wilson C, Rathbone E, Keane M, Gil M, Burkinshaw R, Grieve R, Barrett-Lee P, Ritchie D, Liversedge V, Hinsley S, Marshall H (2014) Adjuvant zoledronic acid in patients with early breast cancer: final efficacy analysis of the AZURE (BIG 01/04) randomised open-label phase 3 trial. Lancet Oncol 15:997–1006
- 44. Gnant M, Mlineritsch B, Stoeger H, Luschin-Ebengreuth G, Knauer M, Moik M, Jakesz R, Seifert M, Taucher S, Bjelic-Radisic V, Balic M, Eidtmann H, Eiermann W, Steger G, Kwasny W, Dubsky P, Selim U, Fitzal F, Hochreiner G, Wette V, Sevelda P, Ploner F, Bartsch R, Fesl C, Greil R (2015) Zoledronic acid combined with adjuvant endocrine therapy of tamoxifen versus anastrozol plus ovarian function suppression in premenopausal early breast cancer: final analysis of the Austrian Breast and Colorectal Cancer Study Group Trial 12. Ann Oncol 26:313–320
- 45. Rodrigues P, Hering FO, Meller A (2011) Adjuvant effect of IV clodronate on the delay of bone metastasis in high-risk prostate cancer patients: a prospective study. Cancer Res Treat 43:231–235
- 46. Dearnaley DP, Mason MD, Parmar MK, Sanders K, Sydes MR (2009) Adjuvant therapy with oral sodium clodronate in locally advanced and metastatic prostate cancer: long-term overall survival results from the MRC PR04 and PR05 randomised controlled trials. Lancet Oncol 10:872–876
- 47. Denham JW, Nowitz M, Joseph D, Duchesne G, Spry NA, Lamb DS, Matthews J, Turner S, Atkinson C, Tai KH, Gogna NK, Kenny L, Diamond T, Smart R, Rowan D, Moscato P, Vimieiro R, Woodfield R, Lynch K, Delahunt B, Murray J, D'Este C, McElduff P, Steigler A, Kautto A, Ball J (2014) Impact of androgen suppression and zoledronic acid on bone mineral density and fractures in the Trans-Tasman Radiation Oncology Group (TROG) 03.04 Randomised Androgen Deprivation and Radiotherapy (RADAR) randomized controlled trial for locally advanced prostate cancer. BJU Int 114:344–353
- 48. Wirth M, Tammela T, Cicalese V, Gomez Veiga F, Delaere K, Miller K, Tubaro A, Schulze M, Debruyne F, Huland H, Patel A, Lecouvet F, Caris C, Witjes W (2015) Prevention of bone metastases in patients with high-risk nonmetastatic prostate cancer treated with zoledronic acid: efficacy and Safety Results of the Zometa European Study (ZEUS). Eur Urol 67:482–491
- 49. James ND, Sydes MR, Clarke NW, Mason MD, Dearnaley DP, Spears MR, Ritchie AWS, Parker CC, Russell JM, Attard G, de Bono J, Cross W, Jones RJ, Thalmann G, Amos C, Matheson D, Millman R, Alzouebi M, Beesley S, Birtle AJ, Brock S, Cathomas R, Chakraborti P, Chowdhury S, Cook A, Elliott T, Gale J, Gibbs S, Graham JD, Hetherington J, Hughes R, Laing R, McKinna F, McLaren DB, O'Sullivan JM, Parikh O, Peedell C, Protheroe A, Robinson AJ, Srihari N, Srinivasan R, Staffurth J, Sundar S, Tolan S, Tsang D, Wagstaff J, Parmar MKB (2016) Addition of docetaxel, zoledronic acid, or both to first-line longterm hormone therapy in prostate cancer (STAMPEDE): survival results from an adaptive, multiarm, multistage, platform randomised controlled trial. Lancet 387:1163–1177
- 50. Reyes C, Hitz M, Prieto-Alhambra D, Abrahamsen B (2015) Risks and benefits of bisphosphonate therapies. J Cell Biochem 117:20–28
- 51. Cole LE, Vargo-Gogola T, Roeder RK (2016) Targeted delivery to bone and mineral deposits using bisphosphonate ligands. Adv Drug Deliv Rev 99:12–27
- 52. Jones DH, Nakashima T, Sanchez OH, Kozieradzki I, Komarova SV, Sarosi I, Morony S, Rubin E, Sarao R, Hojilla CV, Komnenovic V, Kong YY, Schreiber M, Dixon SJ, Sims SM, Khokha R, Wada T, Penninger JM (2006) Regulation of cancer cell migration and bone metastasis by RANKL. Nature 440:692–696
- 53. Gonzalez-Suarez E, Jacob AP, Jones J, Miller R, Roudier-Meyer M, Erwert R, Pinkas J, Branstetter D, Dougall WC (2010) RANK ligand mediates progestin-induced mammary epithelial proliferation and carcinogenesis. Nature 468:103–107
- 54. Leibbrandt A, Penninger JM (2008) RANK/RANKL: regulators of immune responses and bone physiology. Ann N Y Acad Sci 1143:123–150
- 55. Pfitzner BM, Branstetter D, Loibl S, Denkert C, Lederer B, Schmitt WD, Dombrowski F, Werner M, Rüdiger T, Dougall WC, Von Minckwitz G (2014) RANK expression as a prognostic and predictive marker in breast cancer. Breast Cancer Res Treat 145:307–315
- 56. Kostenuik PJ, Smith SY, Jolette J, Schroeder J, Pyrah I, Ominsky MS (2011) Decreased bone remodeling and porosity are associated with improved bone strength in ovariectomized cynomolgus monkeys treated with denosumab, a fully human RANKL antibody. Bone 49:151–161
- 57. Ominsky MS, Stouch B, Schroeder J, Pyrah I, Stolina M, Smith SY, Kostenuik PJ (2011) Denosumab, a fully human RANKL antibody, reduced bone turnover markers and increased trabecular and cortical bone mass, density, and strength in ovariectomized cynomolgus monkeys. Bone 49:162–173
- 58. Kostenuik PJ, Nguyen HQ, McCabe J, Warmington KS, Kurahara C, Sun N, Chen C, Li L, Cattley RC, Van G, Scully S, Elliott R, Grisanti M, Morony S, Tan HL, Asuncion F, Li X, Ominsky MS, Stolina M, Dwyer D, Dougall WC, Hawkins N, Boyle WJ, Simonet WS, Sullivan JK (2009) Denosumab, a fully human monoclonal antibody to RANKL, inhibits bone resorption and increases BMD in knock-in mice that express chimeric (murine/human) RANKL. J Bone Miner Res 24:182–195
- 59. Rinotas V, Niti A, Dacquin R, Bonnet N, Stolina M, Han CY, Kostenuik P, Jurdic P, Ferrari S, Douni E (2014) Novel genetic models of osteoporosis by overexpression of human RANKL in transgenic mice. J Bone Miner Res 29:1158–1169
- 60. Canon J, Bryant R, Roudier M, Branstetter DG, Dougall WC (2012) RANKL inhibition combined with tamoxifen treatment

increases anti-tumor efficacy and prevents tumor-induced bone destruction in an estrogen receptor-positive breast cancer bone metastasis model. Breast Cancer Res Treat 135:771–780

- 61. Ottewell PD, Wang N, Brown HK, Fowles CA, Croucher PI, Eaton CL, Holen I (2015) OPG-Fc inhibits ovariectomy-induced growth of disseminated breast cancer cells in bone. Int J Cancer 137:968–977
- 62. Canon JR, Bryant R, Roudier M, Dougall WC (2013) Abstract 3947: AMG 161, a fully human monoclonal antibody to human RANKL, inhibits tumor-induced osteoclastogenesis and reduces skeletal tumor burden in mice that express chimeric (murine/ human) RANKL. Cancer Res 73:3947
- 63. Body JJ, Facon T, Coleman RE, Lipton A, Geurs F, Fan M, Holloway D, Peterson MC, Bekker PJ (2006) A study of the biological receptor activator of nuclear factor-kappaB ligand inhibitor, denosumab, in patients with multiple myeloma or bone metastases from breast cancer. Clin Cancer Res 12:1221–1228
- 64. Henry DH, Costa L, Goldwasser F, Hirsh V, Hungria V, Prausova J, Scagliotti GV, Sleeboom H, Spencer A, Vadhan-Raj S, Von Moos R, Willenbacher W, Woll PJ, Wang J, Jiang Q, Jun S, Dansey R, Yeh H (2011) Randomized, double-blind study of denosumab versus zoledronic acid in the treatment of bone metastases in patients with advanced cancer (excluding breast and prostate cancer) or multiple myeloma. J Clin Oncol 29:1125–1132
- 65. Raje N, Vadhan-Raj S, Willenbacher W, Terpos E, Hungria V, Spencer A, Alexeeva Y, Facon T, Stewart AK, Feng A, Braun A, Balakumaran A, Roodman GD (2016) Evaluating results from the multiple myeloma patient subset treated with denosumab or zoledronic acid in a randomized phase 3 trial. Blood Cancer J 6:e378
- 66. Lipton A, Steger GG, Figueroa J, Alvarado C, Solal-Celigny P, Body JJ, de Boer R, Berardi R, Gascon P, Tonkin KS, Coleman RE, Paterson AHG, Gao GM, Kinsey AC, Peterson MC, Jun S (2008) Extended efficacy and safety of denosumab in breast cancer patients with bone metastases not receiving prior bisphosphonate therapy. Clin Cancer Res 14:6690–6696
- 67. Stopeck AT, Lipton A, Body J-, Steger GG, Tonkin K, De Boer RH, Lichinitser M, Fujiwara Y, Yardley DA, Viniegra M, Fan M, Jiang Q, Dansey R, Jun S, Braun A (2010) Denosumab compared with zoledronic acid for the treatment of bone metastases in patients with advanced breast cancer: a randomized, double-blind study. J Clin Oncol 28:5132–5139
- 68. Stopeck AT, Fizazi K, Body J-, Brown JE, Carducci M, Diel I, Fujiwara Y, Martín M, Paterson A, Tonkin K, Shore N, Sieber P, Kueppers F, Karsh L, Yardley D, Wang H, Maniar T, Arellano J, Braun A (2016) Safety of long-term denosumab therapy: results from the open label extension phase of two phase 3 studies in patients with metastatic breast and prostate cancer. Support Care Cancer 24:447–455
- 69. Lipton A, Fizazi K, Stopeck AT, Henry DH, Smith MR, Shore N, Martin M, Vadhan-Raj S, Brown JE, Richardson GE, Saad F, Yardley DA, Zhou K, Balakumaran A, Braun A (2016) Effect of denosumab versus zoledronic acid in preventing skeletal-related events in patients with bone metastases by baseline characteristics. Eur J Cancer 53:75–83
- 70. Fizazi K, Lipton A, Mariette X, Body J, Rahim Y, Gralow JR, Gao G, Wu L, Sohn W, Jun S (2009) Randomized phase II trial of denosumab in patients with bone metastases from prostate cancer, breast cancer, or other neoplasms after intravenous bisphosphonates. J Clin Oncol 27:1564–1571
- 71. Fizazi K, Carducci M, Smith M, Damião R, Brown J, Karsh L, Milecki P, Shore N, Rader M, Wang H, Jiang Q, Tadros S, Dansey R, Goessl C (2011) Denosumab versus zoledronic acid for treatment of bone metastases in men with castration-resistant

prostate cancer: a randomised, double-blind study. Lancet 377:813–822

- 72. Smith MR, Saad F, Coleman R, Shore N, Fizazi K, Tombal B, Miller K, Sieber P, Karsh L, Damião R, Tammela TL, Egerdie B, Van Poppel H, Chin J, Morote J, Gómez-Veiga F, Borkowski T, Ye Z, Kupic A, Dansey R, Goessl C (2012) Denosumab and bone-metastasis-free survival in men with castration-resistant prostate cancer: results of a phase 3, randomised, placebo-controlled trial. Lancet 379:39–46
- 73. Smith MR, Halabi S, Ryan CJ, Hussain A, Vogelzang N, Stadler W, Hauke RJ, Monk JP, Saylor P, Bhoopalam N, Saad F, Sanford B, Kelly WK, Morris M, Small EJ (2014) Randomized controlled trial of early zoledronic acid in men with castrationsensitive prostate cancer and bone metastases: results of CALGB 90202 (Alliance). J Clin Oncol 32:1143–1150
- 74. Diel IJ, Body J-, Stopeck AT, Vadhan-Raj S, Spencer A, Steger G, von Moos R, Goldwasser F, Feng A, Braun A (2015) The role of denosumab in the prevention of hypercalcaemia of malignancy in cancer patients with metastatic bone disease. Eur J Cancer 51:1467–1475
- 75. Ellis GK, Bone HG, Chlebowski R, Paul D, Spadafora S, Smith J, Fan M, Jun S (2008) Randomized trial of denosumab in patients receiving adjuvant aromatase inhibitors for nonmetastatic breast cancer. J Clin Oncol 26:4875–4882
- 76. Ellis GK, Bone HG, Chlebowski R, Paul D, Spadafora S, Fan M, Kim D (2009) Effect of denosumab on bone mineral density in women receiving adjuvant aromatase inhibitors for non-metastatic breast cancer: subgroup analyses of a phase 3 study. Breast Cancer Res Treat 118:81–87
- 77. Gnant M, Pfeiler G, Dubsky PC, Hubalek M, Greil R, Jakesz R, Wette V, Balic M, Haslbauer F, Melbinger E, Bjelic-Radisic V, Artner-Matuschek S, Fitzal F, Marth C, Sevelda P, Mlineritsch B, Steger GG, Manfreda D, Exner R, Egle D, Bergh J, Kainberger F, Talbot S, Warner D, Fesl C, Singer CF (2015) Adjuvant denosumab in breast cancer (ABCSG-18): a multicentre, randomised, double-blind, placebo-controlled trial. Lancet 386:433–443
- 78. Smith MR, Egerdie B, Toriz NH, Feldman R, Tammela TLJ, Saad F, Heracek J, Szwedowski M, Ke C, Kupic A, Leder BZ, Goessl C (2009) Denosumab in men receiving androgen-deprivation therapy for prostate cancer. N Engl J Med 361:745–755
- 79. Palafox M, Ferrer I, Pellegrini P, Vila S, Hernandez-Ortega S, Urruticoechea A, Climent F, Soler MT, Munoz P, Vinals F, Tometsko M, Branstetter D, Dougall WC, Gonzalez-Suarez E (2012) RANK induces epithelial-mesenchymal transition and stemness in human mammary epithelial cells and promotes tumorigenesis and metastasis. Cancer Res 72:2879–2888
- 80. Sigl V, Owusu-Boaitey K, Joshi PA, Kavirayani A, Wirnsberger G, Novatchkova M, Kozieradzki I, Schramek D, Edokobi N, Hersl J, Sampson A, Odai-Afotey A, Lazaro C, Gonzalez-Suarez E, Pujana MA, Cimba F, Heyn H, Vidal E, Cruickshank J, Berman H, Sarao R, Ticevic M, Uribesalgo I, Tortola L, Rao S, Tan Y, Pfeiler G, Lee EYHP, Bago-Horvath Z, Kenner L, Popper H, Singer C, Khokha R, Jones LP, Penninger JM (2016) RANKL/RANK control Brca1 mutation-driven mammary tumors. Cell Res 26:761–774
- 81. Ford JA, Jones R, Elders A, Mulatero C, Royle P, Sharma P, Stewart F, Todd R, Mowatt G (2013) Denosumab for treatment of bone metastases secondary to solid tumours: systematic review and network meta-analysis. Eur J Cancer 49:416–430
- 82. Easton JB, Houghton PJ (2006) mTOR and cancer therapy. Oncogene 25:6436–6446
- 83. Bertoldo F, Silvestris F, Ibrahim T, Cognetti F, Generali D, Ripamonti CI, Amadori D, Colleoni MA, Conte P, Del Mastro L, De Placido S, Ortega C, Santini D (2014) Targeting bone

metastatic cancer: role of the mTOR pathway. Biochim Biophys Acta 1845:248–254

- 84. Glantschnig H, Fisher JE, Wesolowski G, Rodan GA, Reszka AA (2003) M-CSF, TNF alpha and RANK ligand promote osteoclast survival by signaling through mTOR//S6 kinase. Cell Death Differ 10:1165–1177
- 85. Kneissel M, Luong-Nguyen N, Baptist M, Cortesi R, Zumstein-Mecker S, Kossida S, O'Reilly T, Lane H, Susa M (2004) Everolimus suppresses cancellous bone loss, bone resorption, and cathepsin K expression by osteoclasts. Bone 35:1144–1156
- 86. Mogi M, Kondo A (2009) Down-regulation of mTOR leads to up-regulation of osteoprotegerin in bone marrow cells. Biochem Biophys Res Commun 384:82–86
- 87. Lee KW, Yook JY, Son MY, Kim MJ, Koo DB, Han YM, Cho YS (2010) Rapamycin promotes the osteoblastic differentiation of human embryonic stem cells by blocking the mTOR pathway and stimulating the BMP/Smad pathway. Stem Cells Dev 19:557–568
- 88. Hussein O, Tiedemann K, Murshed M, Komarova SV (2012) Rapamycin inhibits osteolysis and improves survival in a model of experimental bone metastases. Cancer Lett 314:176–184
- 89. Moriceau G, Ory B, Mitrofan L, Riganti C, Blanchard F, Brion R, Charrier C, Battaglia S, Pilet P, Denis MG, Shultz LD, Mönkkönen J, Rédini F, Heymann D (2010) Zoledronic acid potentiates mTOR inhibition and abolishes the resistance of osteosarcoma cells to RAD001 (everolimus): pivotal role of the prenylation process. Cancer Res 70:10329–10339
- 90. Morgan TM, Pitts TE, Gross TS, Poliachik SL, Vessella RL, Corey E (2008) RAD001 (Everolimus) inhibits growth of prostate cancer in the bone and the inhibitory effects are increased by combination with docetaxel and zoledronic acid. Prostate 68:861–871
- 91. Okui T, Shimo T, Fukazawa T, Kurio N, Hassan NM, Honami T, Takaoka M, Naomoto Y, Sasaki A (2010) Antitumor effect of temsirolimus against oral squamous cell carcinoma associated with bone destruction. Mol Cancer Ther 9:2960–2969
- 92. Hurvitz SA, Andre F, Jiang Z, Shao Z, Mano MS, Neciosup SP, Tseng L, Zhang Q, Shen K, Liu D, Dreosti LM, Burris HA, Toi M, Buyse ME, Cabaribere D, Lindsay M, Rao S, Pacaud LB, Taran T, Slamon D (2015) Combination of everolimus with trastuzumab plus paclitaxel as first-line treatment for patients with HER2-positive advanced breast cancer (BOLERO-1): a phase 3, randomised, double-blind, multicentre trial. Lancet Oncol 16:816–829
- 93. Baselga J, Campone M, Piccart M, Burris HA, Rugo HS, Sahmoud T, Noguchi S, Gnant M, Pritchard KI, Lebrun F, Beck JT, Ito Y, Yardley D, Deleu I, Perez A, Bachelot T, Vittori L, Xu Z, Mukhopadhyay P, Lebwohl D, Hortobagyi GN (2012) Everolimus in postmenopausal hormone-receptor-positive advanced breast cancer. N Engl J Med 366:520–529
- 94. Yardley DA, Noguchi S, Pritchard KI, Burris HA 3rd, Baselga J, Gnant M, Hortobagyi GN, Campone M, Pistilli B, Piccart M, Melichar B, Petrakova K, Arena FP, Erdkamp F, Harb WA, Feng W, Cahana A, Taran T, Lebwohl D, Rugo HS (2013) Everolimus plus exemestane in postmenopausal patients with $HR(+)$ breast cancer: BOLERO-2 final progression-free survival analysis. Adv Ther 30:870–884
- 95. Hortobagyi GN (2015) Everolimus plus exemestane for the treatment of advanced breast cancer: a review of subanalyses from BOLERO-2. Neoplasia 17:279–288
- 96. Maass N, Harbeck N, Mundhenke C, Lerchenmüller C, Barinoff J, Lück H, Ettl J, Aktas B, Kümmel S, Rösel S, Wagner S, Müller L, Bischoff J, Lübbe K, Schwedler K, Schmidt M, Bauerschlag D, Nekljudova V, von Minckwitz G, Loibl S (2013) Everolimus as treatment for breast cancer patients with bone

metastases only: results of the phase II RADAR study. J Cancer Res Clin Oncol 139:2047–2056

- 97. Amato RJ, Flaherty A, Zhang Y, Ouyang F, Mohlere V (2014) Clinical prognostic factors associated with outcome in patients with renal cell cancer with prior tyrosine kinase inhibitors or immunotherapy treated with everolimus. Urol Oncol 32:345–354
- 98. Broom RJ, Hinder V, Sharples K, Proctor J, Duffey S, Pollard S, Fong PCC, Forgeson G, Harris DL, Jameson MB, O'Donnell A, North RT, Deva S, Hanning FJ, Grey A, Findlay MPN (2015) Everolimus and zoledronic acid in patients with renal cell carcinoma With bone metastases: a randomized first-line phase II trial. Clin Genitourin Cancer 13:50–58
- 99. Yu Y, Song Z, Yang S, Yang X, Zhang J, Lu S (2014) Everolimus and zoledronic acid—a potential synergistic treatment for lung adenocarcinoma bone metastasis. Acta Biochim Biophys Sin (Shanghai) 46:792–801
- 100. Vaishampayan U, Shevrin D, Stein M, Heilbrun L, Land S, Stark K, Li J, Dickow B, Heath E, Smith D, Fontana J (2015) Phase II trial of carboplatin, everolimus, and prednisone in metastatic castration-resistant prostate cancer pretreated with docetaxel chemotherapy: a prostate cancer clinical trial consortium study. Urology 86:1206–1211
- 101. Günther A, Baumann P, Burger R, Kellner C, Klapper W, Schmidmaier R, Gramatzki M (2015) Activity of everolimus (RAD001) in relapsed and/or refractory multiple myeloma: a phase I study. Haematologica 100:541–547
- 102. Yee AJ, Hari P, Marcheselli R, Mahindra AK, Mahindra AK, Cirstea DD, Scullen TA, Burke JN, Rodig SJ, Hideshima T, Laubach JP, Ghobrial IM, Schlossman RL, Munshi NC, Anderson KC, Weller EA, Richardson PG, Raje NS (2014) Outcomes in patients with relapsed or refractory multiple myeloma in a phase I study of everolimus in combination with lenalidomide. Br J Haematol 166:401–409
- 103. Rugo HS, Pritchard KI, Gnant M, Noguchi S, Piccart M, Hortobagyi G, Baselga J, Perez A, Geberth M, Csoszi T, Chouinard E, Srimuninnimit V, Puttawibul P, Eakle J, Feng W, Bauly H, El-Hashimy M, Taran T, Burris HA III (2014) Incidence and time course of everolimus-related adverse events in postmenopausal women with hormone receptor-positive advanced breast cancer: insights from BOLERO-2. Ann Oncol 25:808–815
- 104. Turner PG, O'Sullivan JM (2015) (223)Ra and other bone-targeting radiopharmaceuticals-the translation of radiation biology into clinical practice. Br J Radiol 88:20140752
- 105. Coleman R (2016) Treatment of metastatic bone disease and the emerging role of radium-223. Semin Nucl Med 46:99–104
- 106. Henriksen G, Fisher DR, Roeske JC, Bruland OS, Larsen RH (2003) Targeting of osseous sites with alpha-emitting 223Ra: comparison with the beta-emitter 89Sr in mice. J Nucl Med 44:252–259
- 107. Abou DS, Ulmert D, Doucet M, Hobbs RF, Riddle RC, Thorek DLJ (2015) Whole-body and microenvironmental localization of radium-223 in naive and mouse models of prostate cancer metastasis. J Natl Cancer Inst. doi:[10.1093/jnci/djv380](https://doi.org/10.1093/jnci/djv380)
- 108. Henriksen G, Breistol K, Bruland OS, Fodstad O, Larsen RH (2002) Significant antitumor effect from bone-seeking, alphaparticle-emitting (223)Ra demonstrated in an experimental skeletal metastases model. Cancer Res 62:3120–3125
- 109. Nilsson S, Franzén L, Parker C, Tyrrell C, Blom R, Tennvall J, Lennernäs B, Petersson U, Johannessen DC, Sokal M, Pigott K, Yachnin J, Garkavij M, Strang P, Harmenberg J, Bolstad B, Bruland ØS (2007) Bone-targeted radium-223 in symptomatic, hormone-refractory prostate cancer: a randomised, multicentre, placebo-controlled phase II study. Lancet Oncol 8:587–594
- 110. Sartor O, Coleman R, Nilsson S, Heinrich D, Helle SI, O'Sullivan JM, Fossa˚ SD, Chodacki A, Wiechno P, Logue J, Widmark A, Johannessen DC, Hoskin P, James ND, Solberg A, Syndikus I, Vogelzang NJ, O'Bryan-Tear CG, Shan M, Bruland ØS, Parker C (2014) Effect of radium-223 dichloride on symptomatic skeletal events in patients with castration-resistant prostate cancer and bone metastases: results from a phase 3, double-blind, randomised trial. Lancet Oncol 15:738–746
- 111. Suominen MI, Rissanen JP, Kakonen R, Fagerlund KM, Alhoniemi E, Mumberg D, Ziegelbauer K, Halleen JM, Kakonen SM, Scholz A (2013) Survival benefit with radium-223 dichloride in a mouse model of breast cancer bone metastasis. J Natl Cancer Inst 105:908–916
- 112. Takalkar A, Adams S, Subbiah V (2014) Radium-223 dichloride bone-targeted alpha particle therapy for hormone-refractory breast cancer metastatic to bone. Exp Hematol Oncol. doi:[10.](https://doi.org/10.1186/2162-3619-3-23) [1186/2162-3619-3-23](https://doi.org/10.1186/2162-3619-3-23)
- 113. Coleman R, Aksnes A, Naume B, Garcia C, Jerusalem G, Piccart M, Vobecky N, Thuresson M, Flamen P (2014) A phase IIa, nonrandomized study of radium-223 dichloride in advanced breast cancer patients with bone-dominant disease. Breast Cancer Res Treat 145:411–418
- 114. Accardi F, Toscani D, Bolzoni M, Dalla Palma B, Aversa F, Giuliani N (2015) Mechanism of action of bortezomib and the new proteasome inhibitors on myeloma cells and the bone microenvironment: impact on myeloma-induced alterations of bone remodeling. Biomed Res Int 2015:172458
- 115. Niewerth D, Jansen G, Assaraf YG, Zweegman S, Kaspers GJ, Cloos J (2015) Molecular basis of resistance to proteasome inhibitors in hematological malignancies. Drug Resist Updat 18:18–35
- 116. Scott K, Hayden PJ, Will A, Wheatley K, Coyne I (2016) Bortezomib for the treatment of multiple myeloma. Cochrane Database Syst Rev 4:CD010816
- 117. Vanderschueren D, Laurent MR, Claessens F, Gielen E, Lagerquist MK, Vandenput L, Borjesson AE, Ohlsson C (2014) Sex steroid actions in male bone. Endocr Rev 35:906–960
- 118. Haidar S, Ehmer PB, Barassin S, Batzl-Hartmann C, Hartmann RW (2003) Effects of novel 17alpha-hydroxylase/C17, 20-lyase (P450 17, CYP 17) inhibitors on androgen biosynthesis in vitro and in vivo. J Steroid Biochem Mol Biol 84:555–562
- 119. Wu J, Moverare-Skrtic S, Borjesson AE, Lagerquist MK, Sjogren K, Windahl SH, Koskela A, Grahnemo L, Islander U, Wilhelmson AS, Tivesten A, Tuukkanen J, Ohlsson C (2016) Enzalutamide reduces the bone mass in the axial but not the appendicular skeleton in male mice. Endocrinology 157:969–977
- 120. Iuliani M, Pantano F, Buttigliero C, Fioramonti M, Bertaglia V, Vincenzi B, Zoccoli A, Ribelli G, Tucci M, Vignani F, Berruti A, Scagliotti GV, Tonini G, Santini D (2015) Biological and clinical effects of abiraterone on anti-resorptive and anabolic activity in bone microenvironment. Oncotarget 6:12520–12528
- 121. Fizazi K, Scher HI, Molina A, Logothetis CJ, Chi KN, Jones RJ, Staffurth JN, North S, Vogelzang NJ, Saad F, Mainwaring P, Harland S, Goodman OB Jr, Sternberg CN, Li JH, Kheoh T, Haqq CM, de Bono JS, COU-AA-301 Investigators (2012) Abiraterone acetate for treatment of metastatic castration-resistant prostate cancer: final overall survival analysis of the COU-AA-301 randomised, double-blind, placebo-controlled phase 3 study. Lancet Oncol 13:983–992
- 122. McCrea E, Sissung TM, Price DK, Chau CH, Figg WD (2016) Androgen receptor variation affects prostate cancer progression and drug resistance. Pharmacol Res 114:152–162
- 123. Littlewood-Evans AJ, Bilbe G, Bowler WB, Farley D, Wlodarski B, Kokubo T, Inaoka T, Sloane J, Evans DB, Gallagher JA (1997) The osteoclast-associated protease cathepsin K is

expressed in human breast carcinoma. Cancer Res 57:5386–5390

- 124. Le Gall C, Bellahcène A, Bonnelye E, Gasser JA, Castronovo V, Green J, Zimmermann J, Clézardin P (2007) A cathepsin K inhibitor reduces breast cancer-induced osteolysis and skeletal tumor burden. Cancer Res 67:9894–9902
- 125. Brubaker KD, Vessella RL, True LD, Thomas R, Corey E (2003) Cathepsin K mRNA and protein expression in prostate cancer progression. J Bone Miner Res 18:222–230
- 126. Husmann K, Muff R, Bolander ME, Sarkar G, Born W, Fuchs B (2008) Cathepsins and osteosarcoma: expression analysis identifies cathepsin K as an indicator of metastasis. Mol Carcinog 47:66–73
- 127. Podgorski I (2009) Future of anticathepsin K drugs: dual therapy for skeletal disease and atherosclerosis? Future Med Chem 1:21
- 128. Jensen AB, Wynne C, Ramirez G, He W, Song Y, Berd Y, Wang H, Mehta A, Lombardi A (2010) The cathepsin K inhibitor odanacatib suppresses bone resorption in women with breast cancer and established bone metastases: results of a 4-week, double-blind, randomized, controlled trial. Clin Breast Cancer 10:452–458
- 129. Duong LT, Wesolowski GA, Leung P, Oballa R, Pickarski M (2014) Efficacy of a cathepsin K inhibitor in a preclinical model for prevention and treatment of breast cancer bone metastasis. Mol Cancer Ther 13:2898–2909
- 130. Langdahl B, Binkley N, Bone H, Gilchrist N, Resch H, Rodriguez Portales J, Denker A, Lombardi A, Le Bailly DeTilleghem C, DaSilva C, Rosenberg E, Leung A (2012) Odanacatib in the treatment of postmenopausal women with low bone mineral density: five years of continued therapy in a phase 2 study. J Bone Miner Res 27:2251–2258
- 131. Bone HG, Dempster DW, Eisman JA, Greenspan SL, McClung MR, Nakamura T, Papapoulos S, Shih WJ, Rybak-Feiglin A, Santora AC, Verbruggen N, Leung AT, Lombardi A (2015) Odanacatib for the treatment of postmenopausal osteoporosis: development history and design and participant characteristics of LOFT, the Long-Term Odanacatib Fracture Trial. Osteoporos Int 26:699–712
- 132. Soriano P, Montgomery C, Geske R, Bradley A (1991) Targeted disruption of the c-src proto-oncogene leads to osteopetrosis in mice. Cell 64:693–702
- 133. Horne WC, Sanjay A, Bruzzaniti A, Baron R (2005) The role(s) of Src kinase and Cbl proteins in the regulation of osteoclast differentiation and function. Immunol Rev 208:106–125
- 134. Lee YC, Huang CF, Murshed M, Chu K, Araujo JC, Ye X, deCrombrugghe B, Yu-Lee LY, Gallick GE, Lin SH (2010) Src family kinase/abl inhibitor dasatinib suppresses proliferation and enhances differentiation of osteoblasts. Oncogene 29:3196–3207
- 135. Roskoski R Jr (2015) Src protein-tyrosine kinase structure, mechanism, and small molecule inhibitors. Pharmacol Res 94:9–25
- 136. Rucci N, Recchia I, Angelucci A, Alamanou M, Del Fattore A, Fortunati D, Susa M, Fabbro D, Bologna M, Teti A (2006) Inhibition of protein kinase c-Src reduces the incidence of breast cancer metastases and increases survival in mice: implications for therapy. J Pharmacol Exp Ther 318:161–172
- 137. De Felice M, Lambert D, Holen I, Escott KJ, Andrew D (2016) Effects of Src-kinase inhibition in cancer-induced bone pain. Mol Pain. doi:[10.1177/1744806916643725](https://doi.org/10.1177/1744806916643725)
- 138. Campone M, Bondarenko I, Brincat S, Hotko Y, Munster PN, Chmielowska E, Fumoleau P, Ward R, Bardy-Bouxin N, Leip E, Turnbull K, Zacharchuk C, Epstein RJ (2012) Phase II study of single-agent bosutinib, a Src/Abl tyrosine kinase inhibitor, in patients with locally advanced or metastatic breast cancer pretreated with chemotherapy. Ann Oncol 23:610–617
- 139. Yu EY, Massard C, Gross ME, Carducci MA, Culine S, Hudes G, Posadas EM, Sternberg CN, Wilding G, Trudel GC, Paliwal P, Fizazi K (2011) Once-daily dasatinib: expansion of phase II study evaluating safety and efficacy of dasatinib in patients with metastatic castration-resistant prostate cancer. Urology 77:1166–1171
- 140. Yu EY, Duan F, Muzi M, Deng X, Chin BB, Alumkal JJ, Taplin ME, Taub JM, Herman B, Higano CS, Doot RK, Hartfeil D, Febbo PG, Mankoff DA (2015) Castration-resistant prostate cancer bone metastasis response measured by 18F-fluoride PET after treatment with dasatinib and correlation with progressionfree survival: results from American College of Radiology Imaging Network 6687. J Nucl Med 56:354–360
- 141. Mitri Z, Nanda R, Blackwell K, Costelloe CM, Hood I, Wei C, Brewster AM, Ibrahim NK, Koenig KB, Hortobagyi GN, Van Poznak C, Rimawi MF, Moulder-Thompson S, Translational Breast Cancer Research Consortium (2016) TBCRC-010: phase I/II study of dasatinib in combination with zoledronic acid for the treatment of breast cancer bone metastasis. Clin Cancer Res 22:5706–5712
- 142. Rabbani SA, Valentino ML, Arakelian A, Ali S, Boschelli F (2010) SKI-606 (Bosutinib) blocks prostate cancer invasion, growth, and metastasis in vitro and in vivo through regulation of genes involved in cancer growth and skeletal metastasis. Mol Cancer Ther 9:1147–1157
- 143. Yang JC, Bai L, Yap S, Gao AC, Kung HJ, Evans CP (2010) Effect of the specific Src family kinase inhibitor saracatinib on osteolytic lesions using the PC-3 bone model. Mol Cancer Ther 9:1629–1637
- 144. Antonarakis ES, Heath EI, Posadas EM, Yu EY, Harrison MR, Bruce JY, Cho SY, Wilding GE, Fetterly GJ, Hangauer DG, Kwan MF, Dyster LM, Carducci MA (2013) A phase 2 study of KX2-391, an oral inhibitor of Src kinase and tubulin polymerization, in men with bone-metastatic castration-resistant prostate cancer. Cancer Chemother Pharmacol 71:883–892
- 145. Baron R, Kneissel M (2013) WNT signaling in bone homeostasis and disease: from human mutations to treatments. Nat Med 19:179–192
- 146. Colucci S, Brunetti G, Oranger A, Mori G, Sardone F, Specchia G, Rinaldi E, Curci P, Liso V, Passeri G, Zallone A, Rizzi R, Grano M (2011) Myeloma cells suppress osteoblasts through sclerostin secretion. Blood Cancer J 1:e27
- 147. Mendoza-Villanueva D, Zeef L, Shore P (2011) Metastatic breast cancer cells inhibit osteoblast differentiation through the Runx2/CBFbeta-dependent expression of the Wnt antagonist, sclerostin. Breast Cancer Res 13:R106
- 148. Kyvernitakis I, Rachner TD, Urbschat A, Hars O, Hofbauer LC, Hadji P (2014) Effect of aromatase inhibition on serum levels of sclerostin and dickkopf-1, bone turnover markers and bone mineral density in women with breast cancer. J Cancer Res Clin Oncol 140:1671–1680
- 149. Terpos E, Christoulas D, Katodritou E, Bratengeier C, Gkotzamanidou M, Michalis E, Delimpasi S, Pouli A, Meletis J, Kastritis E, Zervas K, Dimopoulos MA (2012) Elevated circulating sclerostin correlates with advanced disease features and abnormal bone remodeling in symptomatic myeloma: reduction postbortezomib monotherapy. Int J Cancer 131:1466–1471
- 150. Eda H, Santo L, Wein MN, Hu DZ, Cirstea DD, Nemani N, Tai YT, Raines SE, Kuhstoss SA, Munshi NC, Kronenberg HM, Raje NS (2016) Regulation of sclerostin expression in multiple myeloma by Dkk-1; a potential therapeutic strategy for myeloma bone disease. J Bone Miner Res 31:1225–1234
- 151. Garcia-Fontana B, Morales-Santana S, Varsavsky M, Garcia-Martin A, Garcia-Salcedo JA, Reyes-Garcia R, Munoz-Torres M (2014) Sclerostin serum levels in prostate cancer patients and their relationship with sex steroids. Osteoporos Int 25:645–651
- 153. Larson SR, Zhang X, Dumpit R, Coleman I, Lakely B, Roudier M, Higano CS, True LD, Lange PH, Montgomery B, Corey E, Nelson PS, Vessella RL, Morrissey C (2013) Characterization of osteoblastic and osteolytic proteins in prostate cancer bone metastases. Prostate 73:932–940
- 154. Hudson BD, Hum NR, Thomas CB, Kohlgruber A, Sebastian A, Collette NM, Coleman MA, Christiansen BA, Loots GG (2015) SOST inhibits prostate cancer invasion. PLoS ONE 10:e0142058
- 155. Jiang Y, Dai J, Zhang H, Sottnik JL, Keller JM, Escott KJ, Sanganee HJ, Yao Z, McCauley LK, Keller ET (2013) Activation of the wnt pathway through AR79, a GSK3ß inhibitor, promotes prostate cancer growth in soft tissue and bone. Mol Cancer Res 11:1597–1610
- 156. Veverka V, Henry AJ, Slocombe PM, Ventom A, Mulloy B, Muskett FW, Muzylak M, Greenslade K, Moore A, Zhang L, Gong J, Qian X, Paszty C, Taylor RJ, Robinson MK, Carr MD (2009) Characterization of the structural features and interactions of sclerostin: molecular insight into a key regulator of Wnt-mediated bone formation. J Biol Chem 284:10890–10900
- 157. Nioi P, Taylor S, Hu R, Pacheco E, He YD, Hamadeh H, Paszty C, Pyrah I, Ominsky MS, Boyce RW (2015) Transcriptional profiling of laser capture microdissected subpopulations of the osteoblast lineage provides insight into the early response to sclerostin antibody in rats. J Bone Miner Res 30:1457–1467
- 158. Taylor S, Ominsky MS, Hu R, Pacheco E, He YD, Brown DL, Aguirre JI, Wronski TJ, Buntich S, Afshari CA, Pyrah I, Nioi P, Boyce RW (2016) Time-dependent cellular and transcriptional changes in the osteoblast lineage associated with sclerostin antibody treatment in ovariectomized rats. Bone 84:148–159
- 159. Reagan MR, McDonald M, Terry R, Pettitt J, Le L, Mohanty S, Kneissel M, Kramer I, Brooks D, Bouxsein M, Rosen CJ, Ghobrial IM, Croucher P (2015) Anti-sclerostin treatment prevents multiple myeloma induced bone loss and reduces tumor burden. Blood 126:119
- 160. McClung MR, Grauer A, Boonen S, Bolognese MA, Brown JP, Diez-Perez A, Langdahl BL, Reginster J-, Zanchetta JR, Wasserman SM, Katz L, Maddox J, Yang Y-, Libanati C, Bone HG (2014) Romosozumab in postmenopausal women with low bone mineral density. New Engl J Med 370:412–420
- 161. Recker RR, Benson CT, Matsumoto T, Bolognese MA, Robins DA, Alam J, Chiang AY, Hu L, Krege JH, Sowa H, Mitlak BH, Myers SL (2015) A randomized, double-blind phase 2 clinical trial of blosozumab, a sclerostin antibody, in postmenopausal women with low bone mineral density. J Bone Miner Res 30:216–224
- 162. Lewiecki EM (2014) Role of sclerostin in bone and cartilage and its potential as a therapeutic target in bone diseases. Ther Adv Musculoskelet Dis 6:48–57
- 163. Voorzanger-Rousselot N, Goehrig D, Journe F, Doriath V, Body JJ, Clezardin P, Garnero P (2007) Increased Dickkopf-1 expression in breast cancer bone metastases. Br J Cancer 97:964–970
- 164. Heath DJ, Chantry AD, Buckle CH, Coulton L, Shaughnessy JD, Evans HR, Snowden JA, Stover DR, Vanderkerken K, Croucher PI (2009) Inhibiting Dickkopf-1 (Dkk1) removes suppression of bone formation and prevents the development of osteolytic bone disease in multiple myeloma. J Bone Miner Res 24:425–436
- 165. Rachner TD, Gobel A, Benad-Mehner P, Hofbauer LC, Rauner M (2014) Dickkopf-1 as a mediator and novel target in malignant bone disease. Cancer Lett 346:172–177
- 166. Goldstein SD, Trucco M, Bautista Guzman W, Hayashi M, Loeb DM (2016) A monoclonal antibody against the Wnt signaling inhibitor dickkopf-1 inhibits osteosarcoma metastasis in a preclinical model. Oncotarget 7:21114–21123
- 167. Iyer SP, Beck JT, Stewart AK, Shah J, Kelly KR, Isaacs R, Bilic S, Sen S, Munshi NC (2014) A phase IB multicentre dosedetermination study of BHQ880 in combination with antimyeloma therapy and zoledronic acid in patients with relapsed or refractory multiple myeloma and prior skeletal-related events. Br J Haematol 167:366–375
- 168. Florio M, Gunasekaran K, Stolina M, Li X, Liu L, Tipton B, Salimi-Moosavi H, Asuncion FJ, Li C, Sun B, Tan HL, Zhang L, Han CY, Case R, Duguay AN, Grisanti M, Stevens J, Pretorius JK, Pacheco E, Jones H, Chen Q, Soriano BD, Wen J, Heron B, Jacobsen FW, Brisan E, Richards WG, Ke HZ, Ominsky MS (2016) A bispecific antibody targeting sclerostin and DKK-1 promotes bone mass accrual and fracture repair. Nat Commun 7:11505
- 169. Kaplan RN, Rafii S, Lyden D (2006) Preparing the ''soil'': the premetastatic niche. Cancer Res 66:11089–11093
- 170. Kitagawa Y, Dai J, Zhang J, Keller JM, Nor J, Yao Z, Keller ET (2005) Vascular endothelial growth factor contributes to prostate cancer-mediated osteoblastic activity. Cancer Res 65:10921–10929
- 171. Nakashiro K, Hayashi Y, Oyasu R (2003) Immunohistochemical expression of hepatocyte growth factor and c-Met/HGF receptor in benign and malignant human prostate tissue. Oncol Rep 10:1149–1153
- 172. Yakes FM, Chen J, Tan J, Yamaguchi K, Shi Y, Yu P, Qian F, Chu F, Bentzien F, Cancilla B, Orf J, You A, Laird AD, Engst S, Lee L, Lesch J, Chou YC, Joly AH (2011) Cabozantinib (XL184), a novel MET and VEGFR2 inhibitor, simultaneously suppresses metastasis, angiogenesis, and tumor growth. Mol Cancer Ther 10:2298–2308
- 173. Nguyen HM, Ruppender N, Zhang X, Brown LG, Gross TS, Morrissey C, Gulati R, Vessella RL, Schimmoller F, Aftab DT, Corey E (2013) Cabozantinib inhibits growth of androgen-sensitive and castration-resistant prostate cancer and affects bone remodeling. PLoS ONE 8:e78881
- 174. Dai J, Zhang H, Karatsinides A, Keller JM, Kozloff KM, Aftab DT, Schimmoller F, Keller ET (2014) Cabozantinib inhibits prostate cancer growth and prevents tumor-induced bone lesions. Clin Cancer Res 20:617–630
- 175. Varkaris A, Corn PG, Parikh NU, Efstathiou E, Song JH, Lee Y, Aparicio A, Hoang AG, Gaur S, Thorpe L, Maity SN, Bar Eli M, Czerniak BA, Shao Y, Alauddin M, Lin S, Logothetis CJ, Gallick GE (2016) Integrating murine and clinical trials with cabozantinib to understand roles of MET and VEGFR2 as targets for growth inhibition of prostate cancer. Clin Cancer Res 22:107
- 176. Watanabe K, Hirata M, Tominari T, Matsumoto C, Fujita H, Yonekura K, Murphy G, Nagase H, Miyaura C, Inada M (2016) The MET/vascular endothelial growth factor receptor (VEGFR) targeted tyrosine kinase inhibitor also attenuates FMS-dependent osteoclast differentiation and bone destruction induced by prostate cancer. J Biol Chem 291:20891–20899
- 177. Fujita H, Gomori A, Fujioka Y, Kataoka Y, Tanaka K, Hashimoto A, Suzuki T, Ito K, Haruma T, Yamamoto-Yokoi H, Harada N, Sakuragi M, Oda N, Matsuo K, Inada M, Yonekura K (2016) High potency VEGFRs/MET/FMS triple blockade by TAS-115 concomitantly suppresses tumor progression and bone destruction in tumor-induced bone disease model with lung carcinoma cells. PLoS ONE 11:e0164830
- 178. Smith M, De Bono J, Sternberg C, Le Moulec S, Oudard S, De Giorgi U, Krainer M, Bergman A, Hoelzer W, De Wit R, Bogemann M, Saad F, Cruciani G, Thiery-Vuillemin A,

Feyerabend S, Miller K, Houede N, Hussain S, Lam E, Polikoff J, Stenzl A, Mainwaring P, Ramies D, Hessel C, Weitzman A, Fizazi K (2016) Phase III study of cabozantinib in previously treated metastatic castration-resistant prostate cancer: COMET-1. J Clin Oncol 34:3005–3013

- 179. Doran MG, Spratt DE, Wongvipat J, Ulmert D, Carver BS, Sawyers CL, Evans MJ (2014) Cabozantinib resolves bone scans in tumor-naive mice harboring skeletal injuries. Mol Imaging. doi:[10.2310/7290.2014.00026](https://doi.org/10.2310/7290.2014.00026)
- 180. Graham TJ, Box G, Tunariu N, Crespo M, Spinks TJ, Miranda S, Attard G, de Bono J, Eccles SA, Davies FE, Robinson SP (2014) Preclinical evaluation of imaging biomarkers for prostate cancer bone metastasis and response to cabozantinib. J Natl Cancer Inst. doi[:10.1093/jnci/dju033](https://doi.org/10.1093/jnci/dju033)
- 181. Desgrosellier JS, Cheresh DA (2010) Integrins in cancer: biological implications and therapeutic opportunities. Nat Rev Cancer 10:9–22
- 182. Stucci S, Tucci M, Passarelli A, Silvestris F (2015) Avβ3 integrin: pathogenetic role in osteotropic tumors. Crit Rev Oncol Hematol 96:183–193
- 183. Davies J, Warwick J, Totty N, Philp R, Helfrich M, Horton M (1989) The osteoclast functional antigen, implicated in the regulation of bone resorption, is biochemically related to the vitronectin receptor. J Cell Biol 109:1817–1826
- 184. Zhao Y, Bachelier R, Treilleux I, Pujuguet P, Peyruchaud O, Baron R, Clement-Lacroix P, Clezardin P (2007) Tumor Av β 3 integrin is a therapeutic target for breast cancer bone metastases. Cancer Res 67:5821–5830
- 185. Carter RZ, Micocci KC, Natoli A, Redvers RP, Paquet-Fifield S, Martin AC, Denoyer D, Ling X, Kim SH, Tomasin R, Selistrede-Araujo H, Anderson RL, Pouliot N (2015) Tumour but not stromal expression of β 3 integrin is essential, and is required early, for spontaneous dissemination of bone-metastatic breast cancer. J Pathol 235:760–772
- 186. Gvozdenovic A, Boro A, Meier D, Bode-Lesniewska B, Born W, Muff R, Fuchs B (2016) Targeting $\alpha v \beta$ 3 and alphavbeta5 integrins inhibits pulmonary metastasis in an intratibial xenograft osteosarcoma mouse model. Oncotarget 7:55141–55154
- 187. Yao H, Veine DM, Livant DL (2016) Therapeutic inhibition of breast cancer bone metastasis progression and lung colonization: breaking the vicious cycle by targeting α 5 β 1 integrin. Breast Cancer Res Treat 157:489–501
- 188. Rucci N, Capulli M, Olstad OK, Önnerfjord P, Tillgren V, Gautvik KM, Heinegård D, Teti A (2015) The α 2 β 1 binding domain of chondroadherin inhibits breast cancer-induced bone metastases and impairs primary tumour growth: a preclinical study. Cancer Lett 358:67–75
- 189. Gramoun A, Shorey S, Bashutski JD, Dixon SJ, Sims SM, Heersche JN, Manolson MF (2007) Effects of Vitaxin, a novel therapeutic in trial for metastatic bone tumors, on osteoclast functions in vitro. J Cell Biochem 102:341–352
- 190. Bradley DA, Daignault S, Ryan CJ, Dipaola RS, Cooney KA, Smith DC, Small E, Mathew P, Gross ME, Stein MN, Chen A, Pienta KJ, Escara-Wilke J, Doyle G, Al-Hawary M, Keller ET, Hussain M (2011) Cilengitide (EMD 121974, NSC 707544) in asymptomatic metastatic castration resistant prostate cancer patients: a randomized phase II trial by the prostate cancer clinical trials consortium. Invest New Drugs 29:1432–1440
- 191. Kenny LM, Coombes RC, Oulie I, Contractor KB, Miller M, Spinks TJ, McParland B, Cohen PS, Hui AM, Palmieri C, Osman S, Glaser M, Turton D, Al-Nahhas A, Aboagye EO (2008) Phase I trial of the positron-emitting Arg-Gly-Asp (RGD) peptide radioligand 18F-AH111585 in breast cancer patients. J Nucl Med 49:879–886
- 192. Zhang J, Niu G, Lang L, Li F, Fan X, Yan X, Yao S, Yan W, Huo L, Chen L, Li Z, Zhu Z, Chen X (2017) Clinical translation

of a dual integrin $\alpha \beta$ 3- and gastrin-releasing peptide receptortargeting pet radiotracer, 68 Ga-BBN-RGD. J Nucl Med 58:228–234

- 193. Gaddy-Kurten D, Coker JK, Abe E, Jilka RL, Manolagas SC (2002) Inhibin suppresses and activin stimulates osteoblastogenesis and osteoclastogenesis in murine bone marrow cultures. Endocrinology 143:74–83
- 194. Chantry AD, Heath D, Mulivor AW, Pearsall S, Baud'huin M, Coulton L, Evans H, Abdul N, Werner ED, Bouxsein ML, Key ML, Seehra J, Arnett TR, Vanderkerken K, Croucher P (2010) Inhibiting activin-A signaling stimulates bone formation and prevents cancer-induced bone destruction in vivo. J Bone Miner Res 25:2633–2646
- 195. Kang HY, Huang HY, Hsieh CY, Li CF, Shyr CR, Tsai MY, Chang C, Chuang YC, Huang KE (2009) Activin A enhances prostate cancer cell migration through activation of androgen receptor and is overexpressed in metastatic prostate cancer. J Bone Miner Res 24:1180–1193
- 196. Vallet S, Mukherjee S, Vaghela N, Hideshima T, Fulciniti M, Pozzi S, Santo L, Cirstea D, Patel K, Sohani AR, Guimaraes A, Xie W, Chauhan D, Schoonmaker JA, Attar E, Churchill M, Weller E, Munshi N, Seehra JS, Weissleder R, Anderson KC,

Scadden DT, Raje N (2010) Activin A promotes multiple myeloma-induced osteolysis and is a promising target for myeloma bone disease. Proc Natl Acad Sci USA 107:5124–5129

- 197. Olsen OE, Wader KF, Hella H, Mylin AK, Turesson I, Nesthus I, Waage A, Sundan A, Holien T (2015) Activin A inhibits BMP-signaling by binding ACVR2A and ACVR2B. Cell Commun Signal. doi:[10.1186/s12964-015-0104-z](https://doi.org/10.1186/s12964-015-0104-z)
- 198. Silbermann R, Bolzoni M, Storti P, Guasco D, Bonomini S, Zhou D, Wu J, Anderson JL, Windle JJ, Aversa F, Roodman GD, Giuliani N (2014) Bone marrow monocyte-/macrophagederived activin A mediates the osteoclastogenic effect of IL-3 in multiple myeloma. Leukemia 28:951–954
- 199. Abdulkadyrov KM, Salogub GN, Khuazheva NK, Sherman ML, Laadem A, Barger R, Knight R, Srinivasan S, Terpos E (2014) Sotatercept in patients with osteolytic lesions of multiple myeloma. Br J Haematol 165:814–823
- 200. Scullen T, Santo L, Vallet S, Fulciniti M, Eda H, Cirstea D, Patel K, Nemani N, Yee A, Mahindra A, Raje N (2013) Lenalidomide in combination with an activin A-neutralizing antibody: preclinical rationale for a novel anti-myeloma strategy. Leukemia 27:1715–1721