REVIEW ARTICLE

Edible insect‑processing techniques: a strategy to develop nutritional food products and novelty food analogs

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Abstract

Edible insects have been part of the eating habits of several cultures over the years. They are recognized as a promising nutritional and sustainable alternative food source. The transformation of insects into safer and more acceptable food ingredients depends on the processing techniques and the fnal food system. Conventional methodologies such as blanching, boiling, drying, and milling are primarily used for material pre-conditioning and powder preparation, mostly for preparing bakery products. Most advanced technologies are preferred for extraction of insect derivatives such as lipids, proteins, polyphenols, and chitosan due to their better-quality preservation, higher yield, and more environmentally friendly (solvent residues). Insect derivatives (mainly lipids and proteins) have been used to enhance the nutritional value of processed products and to produce food analogs, principally for meat and less investigated for milk and dairy products. This literature overview summarizes the efect of diferent processing techniques on edible insects' safety and quality and their use for the development of processed products and derivatives for food analogs production.

Keywords Edible insects · Insect processing · Insect-based food products · Insect derivatives · Insect-based food analogs

Introduction

The food demand is projected to increase drastically as the world population grows exponentially; the United Nations (UN) estimations for 2050 indicate nearly 10 billion people. This overpopulation requires more natural resources and food, compromising food security. Globally, the food systems currently use at least 70% of freshwater and nearly 50% of land resources, mainly for animal feeding. It is well known that livestock farming has the most signifcant GHG emissions in the food supply chain, besides their afection for soil erosion, deforestation, and water pollution. For instance, cattle produce 4.6 gigatons $CO₂$ -eq, representing about 65% of the emissions [\[1\]](#page-10-0). The FAO estimates that the per capita consumption of animal protein will increase globally; thus, more animal-based production will be needed to meet human requirements [[2](#page-11-0)]. In recent years, the food industry has introduced more sustainable protein sources for

feeding and human consumption, such as biomass residues [[3\]](#page-11-1), microalgae [\[4](#page-11-2)], fungi [[5\]](#page-11-3), and, more recently, insects [[6,](#page-11-4) [7\]](#page-11-5). The FAO has encouraged the use of edible insects as food due to their high nutritional value and environmentally friendly breeding conditions [[8\]](#page-11-6). Some other advantages are that insect rearing has a high feed conversion efficiency and fecundity, low GHG and ammonia emissions, and less land, water, and energy requirements than poultry and livestock [[9,](#page-11-7) [10\]](#page-11-8). In this framework, edible insects can be considered a sustainable alternative to contribute to food security, providing high-nutritional quality ingredients for food product development, especially in developing countries facing animal-based protein shortages, leading to malnutrition and growth defciencies [[11\]](#page-11-9).

Entomophagy, which is the practice of eating insects, is a common feeding habit in several regions in Africa, Asia, and Latin America [[12\]](#page-11-10) compared to Western Europe, in which the edible insects are minimally cooked for direct consumption or used as ingredients to produce traditional dishes [[13\]](#page-11-11). For instance, insect consumption in Mexico, which stands out as the country with more than 500 identifed edible insect species, is culturally accepted and these species are mostly included in the gastronomic culture. The primary strategy to increase consumer acceptance

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is their conversion to a safe and familiar food product by processing. Minimal processing techniques such as drying and grinding have been used to produce material (mainly insect powders) incorporated in snacks [\[14](#page-11-12)], beverages [\[15](#page-11-13)], bakery $[16, 17]$ $[16, 17]$ $[16, 17]$ $[16, 17]$ $[16, 17]$ and pasta $[18]$ $[18]$ $[18]$ and meat products $[19–21]$ $[19–21]$ $[19–21]$ $[19–21]$. Also, insect derivatives, such as proteins, could be used as potential functional agents in the several food formulations due to their technological properties (emulsifying, gelling, foaming, water, and oil absorption) [\[10](#page-11-8)]. Nowadays, innovative food products formulated with edible insects are more acceptable and market-positioned, such as food analogs all over the world [\[22\]](#page-11-19). The production of food analogs aims to mimic or resemble a specifc type of food, principally animal-based commercial products such as meat and meat derivatives and milk and dairy products.

Edible insects: a nutritious alternative for food security

Population growth has impacted food security primarily in low- and middle-income economies. Over 820 million people worldwide suffer from hunger, and almost 2.3 billion face food insecurity at moderate or severe levels [[2](#page-11-0)]. Food insecurity is one of the most serious and widespread problems, substantially afecting people across developing nations [\[23\]](#page-11-20). The permanent availability, accessibility, and afordability of nutritious food is key for human well-being and is an international priority for vulnerable populations $[8, 24]$ $[8, 24]$ $[8, 24]$ $[8, 24]$. Insects are pointed out as an economical and efficient alternative to prevent nutritional insecurity due to their composition, accessibility, and sustainability. In addition, insect rearing requires elementary techniques, and the growing rates are fast [[8\]](#page-11-6).

Humans currently consume several species of edible insects (over 2000) because of their taste and availability. The most consumed worldwide include Coleoptera (beetles, 31%), Lepidoptera (caterpillars, 18%), and Hymenoptera (bees, wasps, and ants, 14%), followed by other main species, including Orthoptera (grasshoppers, locusts, and crickets, 13%), Hemiptera (cicadas, leafhoppers, planthoppers, scale insects, and true bugs, 10%), Isoptera (termites, 3%), Odonata (dragonfies, 3%), Diptera (fies, 2%) [[25\]](#page-11-22).

Edible insects are recognized mainly as a good source of protein, lipids, and some micronutrients (vitamins and minerals); however, their nutritional profile may differ depending on their metamorphosis stages, gender, feed composition, habitat, environmental factors, processing and preservation techniques, and measurement methods performed [\[8](#page-11-6), [26\]](#page-11-23).

Insect proteins have been reported as highly digestible by humans [[27\]](#page-11-24) and balanced in most essential amino acids such as tryptophan, tyrosine, and lysine [[28\]](#page-11-25). On average, the protein content on a dry basis varies between 35% (some species of Isoptera) and 61% (some species of Orthoptera) [\[29\]](#page-11-26). The highest protein noted is 77% in some of the Orthoptera species (*Melanoplus femurrubrum*) [\[28](#page-11-25)], and lipid content ranges from 9.12% (*Zonocerus variegates*) to 67.25% (*Homorocoryphus nitidulus*) on dry matter [\[30](#page-11-27)]. Some species of Lepidoptera (*Phassus triangularis*) have been documented to exhibit a lipid content reaching as high as 77% [[28](#page-11-25)]. Lipids consist mainly of unsaturated fatty acids, including monounsaturated fatty acids (palmitoleic acid and oleic acid), polyunsaturated fatty acids (linoleic and α-linolenic acids); however, some species, such as black soldier fy (*Hermetia illucens*) is rich in lauric acid (saturated fatty acid). From diferent points of view, this derivative compound could be used to replace palm kernel oil ingredients in developing some food products and represent an alternative source of essential fatty acids, especially in countries with poor access to fsh and fsh-based food products [\[8](#page-11-6)]. Regarding the diversity of unsaturated fatty acid compositions in diferent insect species, Womeni et al. [\[30](#page-11-27)] investigated the composition of oils extracted from some species consumed in Cameroon, and the authors concluded that the primary fatty acids in termites (*Macrotermes* sp.) were palmitic acid (30.47%), oleic acid (47.52%), and linoleic acid (8.79%), in grasshoppers (*Zonocerus variegates*) were palmitoleic acid (23.83%), oleic acid (10.71%), linoleic acid (21.07%), α -linolenic acid (14.76%), and γ -linolenic acid (22.54%), and in caterpillars were palmitic acid (30.80%) and linolenic acid (41.79%).

Regarding micronutrients, edible insects have been considered a good source of complex B vitamins [[31,](#page-11-28) [32](#page-11-29)]. Adult house crickets provide a valuable source of vitamin B_{12} (5.4 µg per 100 g), contributing to the recommended daily allowance (RDA) of vitamin B_{12} for adults (2.4 μ g/ day) [[33,](#page-11-30) [34](#page-11-31)]. It has been reported that silkworm pupae are an excellent source of thiamin (0.07 mg per 100 g), ribofavin (2.23 mg per 100 g), and niacin (2.2 mg per 100 g). Silkworm pupae also contain about 10 mg of tocopherols per 100 g, which is a much higher level than that found in animal products such as milk (0.21 mg per 100 g), chicken (0.67 mg per 100 g) and pork (0.34 mg per 100 g) [\[35](#page-11-32)]. The escamoles, which are *Liometopum apiculatum* M ant eggs, have a high content of vitamins A, D, and E, containing approximately 505 μg/100 g of retinol, 3.31 μg/100 g of cholecalciferol, and 2.22 mg/100 g of alpha-tocopherol [[36](#page-11-33)].

In the case of minerals, it has been found that iron content in Mopane caterpillars (*Imbrasia belina*) is higher $(31–77 \text{ mm}/100 \text{ g})$ than in beef $(6 \text{ mg}/100 \text{ mg})$ [\[37](#page-11-34)]. Similarly, locusts' species contain between 8 and 20 mg per 100 g [\[38\]](#page-11-35). Under this perspective, edible insects could contribute to iron daily intake and reduce its deficiency, mainly in low-income countries with limited afordability to meat. In addition, an investigation on the mineral content of four insects (*Bombay locust*, *Patanga succincta*, scarab beetle *Holotrichia* sp., house cricket *Acheta domesticus*, and mulberry silkworm *Bombyx mori*) has also shown not only high amounts of iron but also magnesium, zinc, and calcium [[39](#page-11-36)]. Based on their analyses, Ghosh et al. [\[40\]](#page-11-37) concluded that insects *Allomyrina dichotoma*, *Protaetia brevitarsis*, *Tenebrio molitor*, *Teleogryllus emma*, and *Gryllus bimaculatus* could be a good source of minerals, especially magnesium (82–304 mg/100 g dry weight) and zinc (10.26–22.43 mg/100 g dry weight). Zielinska et al. [[41\]](#page-12-0) analyzed the composition of some selected species: adult cricket (*Gryllodes sigillatus*), larvae of mealworm (*T. molitor*), and adult locust (*Schistocerca gregaria*) are very rich in copper (1.86–6.32 mg/100 g dry weight) and zinc (11.2–18.6 mg/100 g dry weight). Also, the calcium content of these species varies between 40 and 130 mg per 100 g of dry weight, while beef has on average 4–27 mg/100 g dry weight, pork 5–28, and poultry 5–14 mg/100 g $[41]$ $[41]$.

Another compound is chitin, a complex polysaccharide commonly found in nature, particularly in crustaceans, fungal cell walls, and insect cuticles, which is the insoluble fber (present in their exoskeleton) [\[42\]](#page-12-1). Edible insects can be considered as a potential source of dietary fber in the human health due to their high content of chitin representing more than 10% (dry basis). For example, yellow-winged grasshoppers (*Arphia fallax*) and adult yellow mealworms (*Tenebrio molitor*) contain approximately 12% and 16% fber in their composition (dry matter), respectively [[28\]](#page-11-25). Also, the prebiotic properties of chitin derivative (chitosan) have been reported to beneft the immune system by slowing down the growth of some potentially pathogenic microorganisms [\[43](#page-12-2)]. Additionally, chitin and chitosan can be produced all year round and are available as a cheap byproduct with industrial insect rearing. However, commercial production of these components and the risk of allergic reactions have yet to be established and fully described.

Food legislation for edible insects' production and consumption

The production of insects as food for human consumption is generally at small-scale levels. In some tropical Southeast Asia countries such as Thailand, Vietnam, and Laos, crickets are mainly reared as food in backyard sheds with no need for expensive materials. However, to replace traditional animal sources, edible insects must be produced on an industrial scale [[44\]](#page-12-3). The mealworms, wax moths, silkworms, crickets, and black soldier fy larvae are currently reared as food and are usually recommended as potential insect species for industrial-scale production because of their high oil and protein content and quality [[45,](#page-12-4) [46\]](#page-12-5).

In Europe, several companies and startups have been generated to commercialize insects as food for human consumption in the last decade. This framework rapidly created a regulatory response because of interest in commercial edible insects in the European market. The production and marketing of insects (Whole insects, their parts, and their derived ingredients) as food is governed by the 'Novel Foods' legislation—Regulation (EU) No 2015/2283 in the European Union (EU) [\[47](#page-12-6)[–49\]](#page-12-7). According to current legislation, after the safety evaluation by the European Food Safety Agency (EFSA) [[50–](#page-12-8)[55](#page-12-9)] and a favorable vote given by the EU Member States (MS), the market authorization of these insect-based food products can be granted by European Commission (EC) [[47\]](#page-12-6). There are currently four authorized species of edible insects as novel food in the EU. EFSA. Insects frozen, dried, and powdered for *Tenebrio molitor* larva (the yellow mealworm), *Locusta migratoria* (the migratory locust), *Acheta domesticus* (the house cricket), and *Alphitobius diaperinus* larvae (the lesser mealworm) [[56](#page-12-10)[–60\]](#page-12-11). However, comprehensive and systematic global research is needed to support and verify the favorable social and economic impact on the public of edible insect farming, primarily related to improving the future food security of low-income countries [\[8\]](#page-11-6).

Edible insect consumer's concern: safety and acceptance

Eating insects is a widespread practice in many countries worldwide, especially in some specifc tropical regions or in some ethnic communities [\[61\]](#page-12-12). However, accepting and adapting edible insects poses a primary challenge in the Western world. Several factors affect consumer reluctance to edible insect-based food products, for instance, its cultural unfamiliarity as a diet habit [\[26,](#page-11-23) [62\]](#page-12-13), its association with health risk, and neophobia, which is the fear and rejection of trying new foods [[47\]](#page-12-6).

Even though consumers nowadays seem to be more open to novel food, disgust or yucky is still a core obstacle [[63,](#page-12-14) [64](#page-12-15)]. Studies have shown that displaying a complete image of insects as a marketing strategy for insect-based food products in retail negatively afects consumers' willingness to purchase those specifc food products [[47](#page-12-6), [65\]](#page-12-16). A strategy to minimize neophobia's psychological barriers and cultural aversions, the research has mainly focused on processing technologies, developing conventional food prototypes, and novel formulations for improving the sensorial properties of the fnal product and increasing consumer consumption [[66](#page-12-17)]. One way to develop this product type is by using insect ingredients such as proteins, lipids, and vitamins. These ingredients could be isolated and incorporated into a product without visual changes [[67\]](#page-12-18). Microbial contamination is one of the primary consumers' concerns about eating insects. Edible insects can be related to microorganisms in two ways: their internal microbiota (digestive system,

cuticle, other anatomical parts) and external microbiome (rearing environment, nature). They can act as vectors for the spread of hazardous microorganisms, especially bacteria, and afect the total microbial load, safety, and shelf-life of food products they are processed [[68](#page-12-19)]. Most studies have been focused on determining the bacterial biota in edible insects [[69\]](#page-12-20). The most important food pathogens related to insects are *Staphylococcus aureus*, *Clostridium* spp., and species of the *Bacillus cereus* group [\[70\]](#page-12-21). There is still a lack of knowledge on the fungi, viruses, protozoa, and prion species associated with edible insects, as several food products such as fsh, peanuts, eggs, crustaceans, and insects can also cause risks to consumers with allergies. Many diverse allergens are identifed in edible insects, such as tropomyosin, arginine kinase, phospholipase, A fructose-bisphosphate aldolase, hyaluronidase, myosin light chain, α-tubulin, and β-tubulin [[71,](#page-12-22) [72\]](#page-13-0). Tropomyosin, a protein complex found in the muscles of many invertebrate species such as crustaceans and insects, is considered a signifcant allergen [\[73](#page-13-1)]. Tropomyosin can cross-react with primarily other consumed foods like crustaceans or with common invertebrate inhalant allergens like house dust mites and their metabolites [[74](#page-13-2)]. These allergens can induce non-specifc symptoms, such as itching and fushed skin, eczema, urticaria, allergic asthma, hypotension, gastrointestinal symptoms, loss of consciousness, and tachycardia. Therefore, the allergic potential of edible insects should be observed to eliminate these risks, and edible insect-based food products must be labeled appropriately, not mislead consumers, and minimize allergic reactions [[8](#page-11-6), [25\]](#page-11-22). Even though the consumption of edible insects has been associated with some microbial, toxic, allergenic, and chemical risks, EFSA has established safety assessments for these novel food products to reduce these risks and published its positive opinions [[49–](#page-12-7)[55\]](#page-12-9).

Can processing technologies afect edible insect‑food products' safety and consumer acceptance?

As the production of edible insects expands on an industrial scale, the implementation of suitable post-harvest and processing technologies that can ensure the safety, preservation, quality enhancement, fractionation, and proper storage of insects and insect derivatives becomes essential [\[75](#page-13-3)].

After immediate harvesting, edible insects can be traditionally consumed as whole insects (raw or cooked). Also, they can be processed or transformed into non-recognizable and more palatable forms, like powder and paste. In addition, isolated proteins, lipids, or fber (chitin) can be utilized as food ingredients to enhance existing food products such as bread, pasta, and biscuits or produce novel products such as oil, beverages, and confectioneries. Therefore, processing could contribute to their introduction to the daily human diet, especially for consumers in the Western world, and ensure safety [[76\]](#page-13-4).

Basic food processing techniques have often been applied to improve safety, increase shelf-life, and sensorial and nutritional quality. Principally, thermal techniques such as roasting, frying, boiling, blanching, drying, toasting, and steaming, followed by mechanical techniques such as grinding, milling, centrifuging, and sieving [[77\]](#page-13-5). In the case of extraction and isolation of insect derivatives more novel processing novel technologies are applied (Fig. [1](#page-3-0)).

Safety and quality perspective

Microbial aspects

The prior condition associated with the thermal processing of edible insects is microbial and enzymatic inactivation [[68\]](#page-12-19). Vandeweyer et al. [[78](#page-13-6)] observed that blanching followed by microwave drying resulted in signifcant microbial reductions (total viable count, psychotropic aerobic count, Enterobacteriaceae, yeasts and molds, and lactic acid bacteria) regardless of treatment times (10, 20, 30, and 40 s). Nevertheless, aerobic bacterial endospores were more resistant to the treatments implemented than vegetative cells. Similarly, Wynants et al. [\[79](#page-13-7)] reported that the number of Enterobacteriaceae, lactic acid bacteria, and fungi dropped signifcantly, and there was a reduction in the total microbial count of 4.0 log cfu/g, but aerobic endospores persisted after blanching. Larouche et al. [[80\]](#page-13-8) compared diferent

Fig. 1 Diferent processing methods applied to edible insects

processing methods for black soldier fy larvae, including blanching, freezing, desiccation, grinding, drowning using various gases (100% $CO₂$, 100% N₂, vacuum conditioning), and high hydrostatic pressure and found that blanching, achieved by boiling the larvae in water for 40 s, was the most efective method to reduce microbial load. The total microbial load of the *T. molitor* of 7.72 log10 cfu/g was reduced to 4.73 log10 cfu/g following cold atmospheric pressure plasma treatment for 15 min. In contrast, equally long thermal treatments at 120 and 140 °C were found to completely inactivate the native microorganism flora [[81](#page-13-9)]. Boiling and toasting reduced *Staphylococcus aureus* by 4.8–6.4 log cycles and total viable counts by 4–6 log cycles and eliminated the yeasts and molds, $Lac +$ enteric bacteria, and Salmonella [\[82](#page-13-10)]. Megido et al. [[83\]](#page-13-11) investigated the impact of boiling, frying, and under vacuum cooking on the microbial load of mealworms, showing a reduction of the total microbial load from 8.5 to 1.6, 3.3, and 3.9 log cfu/g, respectively.

Quality and nutritional aspects

The processing techniques may have diferent infuences on the quality characteristics of insects. Freeze-drying and microwave-drying methods caused only minor changes in protein, fat, and ash content of the larvae, while microwave drying reduced the vitamin B_{12} content of the mealworms compared to freeze-dried samples, and the fat fraction of freeze-dried samples showed higher oxidation than the microwave-dried samples [\[84\]](#page-13-12). Furthermore, in all dried mealworm samples, the water activity was below 0.60 (for blanched plus freeze-dried and freeze-dried samples, the water activity was even below 0.30). Azzolini et al. [\[85\]](#page-13-13) reported that blanching had no signifcant diferences in proximate composition between fresh and blanched samples (dry matter). Kröncke et al. [\[86](#page-13-14)] showed that all drying methods tested (microwave-drying, freeze-drying, vacuum-drying, fluidized-bed drying, and conventional hot air drying) caused minor changes in protein, fat, and fber content of the mealworms, freeze-dried mealworms showed significantly highest oxidation status than the other dried samples, while protein solubility was highest at freeze-drying. Megido et al. [\[83\]](#page-13-11) compared several cooking techniques (boiling, vacuum cooking, frying, and oven cooking) for preparing mealworms and demonstrated that frying reduced protein content and digestibility, while boiling and cooking under vacuum increased polyunsaturated fatty acids. Singh et al. [[87\]](#page-13-15) observed no diference in lipid concentration in house cricket between blanching (40 s at 100 °C) and freezing (2 h 10 min at −20 °C). Although tested methods increased the browning index, they had no detrimental effects on product quality. The browning index of microwave-dried mealworms remained stable in contrast to that of freeze-dried during four months of storage [[84](#page-13-12)]. The lightness and yellowness color values decreased when the insects were subjected to blanching+freeze-drying and blanching +air-drying treatments compared to fresh (or only) freeze-dried samples. However, there were no signifcant changes observed in redness (a ∗). In terms of color preservation, oven-drying at 50 °C, followed by a blanching treatment, offered an economical approach compared to freeze-drying [[85\]](#page-13-13). Blanching and microwave-assisted drying noted a dark color appearance, and the products were considered over-processed [[78\]](#page-13-6). Fombong et al. [[88\]](#page-13-16) studied the impact of freeze-drying and oven-drying on the nutrient composition of blanched grasshoppers (*Ruspolia diferens*). They found that both approaches showed the same nutritional quality (proximate, amino acid, mineral, and fatty acid compositions) and could be used to formulate food products containing insects without noticeable sensorial quality.

Khampakool et al. [\[89](#page-13-17)] pre-treated larvae with infrared before freeze-drying, and they found that lower hardness, chewiness, and higher protein content levels, and betterpreserved glutamic acid (6.30–7.29 g/100 g) and proline $(3.84-5.54 \text{ g}/100 \text{ g})$ concentrations compared to non-pretreated freeze-dried and conventionally dried samples (oven). Additionally, the surface appearance of the dried larvae after the infrared-assisted freeze-drying technique showed more air pores and volume expansion compared to the hot air drying-derived product, which might result in a good consumer appeal for dried snack products.

Novel processing alternatives for edible insect derivatives

Besides using edible insect powders and their incorporation into a food product, diferent processing technologies (Table [1](#page-5-0)) have also been applied to produce insect-based ingredients such as protein, lipids, and fber (chitin/chitosan). These derivatives are utilized to produce food analogs, fortifed products, and as techno-functional ingredients in complex systems to enhance sensorial characteristics and stability of fnal products. In general, the derivatives can be used as foaming, gelling, thickening, structuring, and emulsifying agents [\[10](#page-11-8), [66,](#page-12-17) [77](#page-13-5), [90](#page-13-18)]. However, the processing might afect the structure of the compounds. Therefore, the selecting of appropriate methods for the preparation of derivatives with high functional and nutritional value is an essential step.

Proteins

The basic methodology for extracting globular proteins is alkaline solubilization, precipitation (acid or thermal), and ultrafltration or dialysis to increase the purity. However, additional steps, such as defatting and assisted alternative

non-thermal techniques (ultrasound and HPP), are used to pretreat the starting material to increase the protein yield [\[103,](#page-13-31) [110\]](#page-14-2). It has been stated that both processing parameters and insect species (composition) can determine the protein characteristics (nutritional and functional) [[92,](#page-13-20) [111](#page-14-3)].

The protein content in proteinaceous insect powders from grasshoppers (*Schistocerca gregaria*) and honeybees *(Apis mellifera*) increased after defatting (hexane) and sonicationassisted alkaline extraction [\[96](#page-13-24)]. The authors also indicated that protein-emulsifying properties were enhanced compared to whey proteins. Ndiritu et al. [\[91](#page-13-19)] concluded that hexane extraction recorded higher protein yield, color values (lightness and hue angle), crude protein content, and crude ash than aqueous extraction. Nevertheless, aqueous extraction showed better protein emulsifying and foaming capacities and stabilities than hexane extraction. Psarianos et al. [[98\]](#page-13-26) conducted a study on the impact of pulsed electric felds (PEF) on the extraction of the protein from cricket (*Acheta domesticus*) powder. They reported that this treatment (4.90 kJ/kg, 60 min) resulted in a signifcant increase in protein yield $(-46 g$ protein/100 g dry matter) compared to the untreated one $(\sim 39 \text{ g protein}/100 \text{ g dry matter})$, improved techno-functional properties by increasing oil binding and emulsifying capacity over 40% and 70%, respectively, did not afect water binding capacity and foaming capacity. Zhang et al. [[112\]](#page-14-4) applied ultrasound-assisted alkaline extraction using diferent times (10, 20, 30, 40, and 50 min) to obtain protein from *Tenebrio molitor* and observed that ultrasonication could efectively alter the structural properties of proteins (primarily secondary and tertiary structures, thermal stability) which might result in improved functional properties, and was the optimal ultrasound parameter 30 min.

Lipids

Although more conventional lipid extraction methodologies are carried out using organic solvents (hexane, ethanol, methanol, among others), mechanical pressing and novel technologies such as supercritical carbon dioxide have emerged as green and environmentally friendly alternatives [\[103,](#page-13-31) [110\]](#page-14-2).

Tzompa-Sosa et al. [[100\]](#page-13-28) studied the chemical composition of lipids extracted from four insect species (*Tenebrio molitor*, *Alphitobius diaperinus*, *Acheta domesticus*, and *Blaptica dubia*) using various extraction methods and found that *T. molitor* contained the highest lipid content among the four insect species, the highest lipid yield was obtained via Folch extraction while aqueous extraction followed by centrifugation gave the lowest lipid yield, but the highest ω-3 fatty acids content and lowest ω-6/ω-3 fatty acids ratio. However, although the Folch procedure could be an efective method for lipid fractions characterization, it is not considered a food-grade recovering method (because of using chloroform as a solvent). Sun et al. [[102\]](#page-13-30) used ultrasound-assisted aqueous extraction and reported that oil had lower peroxide values, higher polyunsaturated fatty acids contents, and thermal stability and showed stronger antioxidant activities than Soxhlet extraction-derived oil samples. Another study using ultrasound-assisted extraction showed that higher lipids extraction from crickets and mealworms was found when pressurized liquid extraction was used compared to ultrasound-assisted extraction; however, using ultrasound improved the fatty acid profle (in terms of linoleic, oleic, and palmitic acids) of both insect species [\[106](#page-13-34)]. Purschke et al. [[101](#page-13-29)] applied the supercritical carbon dioxide extraction method and efectively achieved the maximal oil extraction yield (95%) from mealworms at high pressure and moderate temperature (400/250 bar, 45 °C, and 105 min). Nevertheless, there was no signifcant diference in yield and oil composition when using hexane.

Chitin

Chitin and its deacetylated form, chitosan, have widely attracted attention thanks to their beneficial biological properties such as nontoxic, antimicrobial, antioxidant, biocompatible, biodegradable, and edible properties in the food industry [\[113\]](#page-14-5). Kaya et al. [[109](#page-14-1)] extracted chitin and chitosan from exoskeletons of *Pimelia* spp. They concluded that both chitin and chitosan showed antimicrobial activity against the common food pathogen bacteria (*Listeria monocytogenes*) and yeast (*Candida albicans*) and, therefore, have the potential for use in food protection. Also, they suggested that both chitin and chitosan can be used to design potential oil-binding food supplements as their oil-binding properties are well documented. Chitin was extracted from the adult beetle (*Holotrichia Parallela Motschulsky*) and compared with the commercial α -chitin from shrimp by Liu et al. [[108\]](#page-14-0). Results indicated that the extracted chitin from the H. parallela exhibited similar characteristics to the chitin obtained from shrimp and could be used as an alternative source of chitin. Psarianos et al. [\[98\]](#page-13-26) studied the effect of pulsed electric felds on chitin extraction from cricket powder as a pre-treatment. They concluded that although chitin yield was not afected by the pulsed electric felds treatment, the morphology of chitin was afected by this treatment.

Application of edible insect derivatives

Functionality in food systems

Due to their technological properties (emulsifying, stabilizing, gelling, water, and oil absorption), insect derivatives such as proteins are used as fat replacers, structuring agents, binders, and extensors. These compounds are also used to (i) fortify food products for enhancing their nutritional quality (mainly through protein supplementation), (ii) develop products for special nutritional needs (such as gluten-free products, or (iii) improve their physicochemical and sensorial properties (Fig. [2](#page-7-0)) [\[10](#page-11-8), [13,](#page-11-11) [66](#page-12-17), [77\]](#page-13-5). Edible insects are mainly utilized for ingredient replacement, to increase the nutritional value, and for food product reformulation (Table [2](#page-8-0)).

Food analogous

During the last few years, the analogs sector has experienced growth in the market. Currently, food analogs commercially available in the market are meat and meat products (burgers, strips, patties, sausages, chunks, chicken-like blocks, nuggets, ground beef-like products and steaks), and milk and dairy analogs (soya bean milk, almond milk, coconut milk, cheese, yogurt) $[126-129]$ $[126-129]$.

Meat analogs intend to imitate the taste, texture, appearance, and functionality of given meat products in diferent forms (grounds, emulsions, and crumbles) [[129](#page-14-7)]. For this purpose, water, fats, oils, textured plant-based proteins (soy, pea, and wheat gluten), coloring and binding agents (gums, hydrocolloids, and enzymes), favors, and spices are among the most utilized analog ingredients to mimic the structural, nutritional, and sensorial characteristics of original meat products. These products are available in Europe, mainly in Germany, The Netherlands, France, Italy, Sweden, and the United Kingdom. Therefore, the availability of these palatable and healthy products plays an important role in

satisfying both vegetarian and meat product consumers' demands [[126,](#page-14-6) [130](#page-14-8)].

Several reasons pointed to the need to develop milk and dairy analogs, which include limited accessibility of milk in low-income regions, eating habits, religion, health issues related to protein allergy, lactose intolerance, and sustainability. The producers of milk and dairy analogs may use several additional ingredients to standardize their nutritional profle (lipids, proteins, sugars, and micronutrients mainly) and stabilize the product (hydrocolloids, lecithin) to approach similar properties of the original product. Usually, milk substitutes made from plant-based sources such as soybean, oats, rice, hemp, almond, and coconut are the most popular milk analogs in the market [[127\]](#page-14-9).

Cheese analogs, usually made by mixing given ingredients such as non-dairy fats, oils, or proteins to manufacture cheese-like products, offer nutritional benefits due to high unsaturated fatty acids, no cholesterol, and fewer calories. In addition, these products are considered cost-efective as cheaper vegetable products can replace the selected milk ingredients. Also, they have easier production compared with traditional cheese production. Cheese analogs, margarine, ice creams, and whipping creams are lower-cost and nutritionally advantageous alternatives [\[131](#page-14-10), [132\]](#page-14-11).

Even though the studies on insect-based food systems are increasing, mainly for meat products, other alternatives, such as milk and dairy products, have been less studied (Table [3\)](#page-10-1) [[22,](#page-11-19) [133,](#page-14-12) [134\]](#page-14-13).

Cho and Ryu [\[138\]](#page-14-14) investigated the effects of different *T.molitor* larva contents (0, 15, and 30%) and extrusion process parameters, including die temperature (140 and 150 °C) and moisture content (40% and 50%) using twin-screw extruder on the physicochemical properties of the extruded meat analog. Texture profle analysis and meat analog oxidation activity decreased with increased larva content, while water-holding capacity, protein digestibility, and antioxidant activity signifcantly increased. Lower die temperature and higher moisture content enhanced the textural properties but decreased protein digestibility. Azzolini et al. [\[137](#page-14-15)] studied different processing parameters $(CaCl₂$ concentration and processing temperature) to design insect-based-meat analog using protein from larvae of *Alphitobius diaperinus* and reported that around 74 g of coagulum could be obtained from 100 g fresh insects. The resulting product (at 100 °C, at 20 mmol of $CaCl₂$) demonstrated the desired texture and mouthfeel as the gelling properties of the insect proteins contributed to the formation of large protein aggregates and stranded structures.

Minced meat-like from mealworm larvae (*T. molitor* and *A. diaperinus*), insects were produced using diferent processing techniques (steam, stir, and pan-fried) and storage using air and modifed atmosphere (MAP) packaging. Fig. 2 Uses of edible insect derivatives
The total aerobic count for fresh insect meat analogous (3.8
The total aerobic count for fresh insect meat analogous (3.8

Table 2 Insect-based food products

Table 2 (continued)

log cfu/g) was lower than beef minced meat (4.9 log cfu/g), and after 28 days of storage, air-packed samples showed no microorganism growth [[139\]](#page-14-24). Insect margarine was successfully formulated by replacing 75% oil plants with *H. illucens* and *T. molitor* lipids without adversely affecting spreading and color [\[133\]](#page-14-12). Some enterprising entrepreneurs may seek to start black soldier fy farms less for waste removal and more for production of the larvae, if not for livestock feed, then for high-protein and low-carb food to combat unhealthy diets, or for more creative cuisine such as EntoMilk™ (Cape Town, South Africa) [\[142\]](#page-14-25). Tello et al. [[22](#page-11-19)] developed an insect-based milk analog using frozen *T. molitor* larvae. Although the resulting product is standardized with insect-extracted fat, ascorbic acid (as a preservative), and sunfower lecithin (as an emulsifer), it had low protein content (1.19%) compared to bovine milk (around 3.5%), the authors indicated this analog showed the fexibility of insects

as a food source and underlined their potential for product development.

Conclusion

Edible insects are a promising sustainable source of ingredients for food product development for human consumption. Based on food security, sustainability, and nutritional quality in the food industry, insect-based food alternatives could contribute to the globally increased food demand. This review shows that bakery goods are the most studied insectcontaining food systems, followed by meat products. In the case of food analogs, only meat prototypes have been extensively explored. More research should be conducted on other products, such as milk and dairy, representing an important niche market. Insect-based milk and dairy analogs (yogurt, ice cream, whipped cream, and cheese, among others) are nutritionally advantageous because they contain unsaturated fatty acids, protein quality, and are lactose free.

Table 3 Food analogs containing edible insects

Author contributions AK: writing—original draft preparation, NN: literature search and review, IK: review and editing, DB: conceptualization, review and editing.

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Declarations

Conflict of interest There is no confict of interest/competing interest.

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Research studies involving with humans and/or animals Not applicable.

Consent to participate Not applicable.

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References

1. Gerber PJ et al (2013) Tackling climate change through livestock: a global assessment of emissions and mitigation opportunities. Food and Agriculture Organization of the United Nations (FAO), Rome

- 2. Food and Agriculture Organization of the United Nations (2017) The future of food and agriculture: trends and challenges. FAO, Rome
- 3. Ramírez-Rodrigues MM, Metri-Ojeda JC, González-Ávila M, Ruiz-Álvarez BE, Baigts-Allende DK (2022) Digestibility and bioaccessibility of leaf protein concentrates and their impact on children gut microbiota. Waste Biomass Valor 13(1):299–314
- 4. Bleakley S, Hayes M (2021) Functional and bioactive properties of protein extracts generated from *Spirulina platensis* and *Isochrysis galbana* T-Iso. Appl Sci 11(9):3964
- 5. Stofel F, de Oliveira Santana W, Fontana RC, Camassola M (2021) Use of *Pleurotus albidus* mycoprotein four to produce cookies: evaluation of nutritional enrichment and biological activity. Innov Food Sci Emerg Technol 68:102642
- 6. Luna GC, Martin-Gonzalez FS, Mauer LJ, Liceaga AM (2021) Cricket (*Acheta domesticus*) protein hydrolysates' impact on the physicochemical, structural and sensory properties of tortillas and tortilla chips. J Insects Food Feed 7(1):109–120
- 7. Van der Spiegel M, Noordam M, Van der Fels-Klerx HJ (2013) Safety of novel protein sources (insects, microalgae, seaweed, duckweed, and rapeseed) and legislative aspects for their application in food and feed production. Comp Rev Food Sci Food Safe 12(6):662–678
- Van Huis A (2013) Potential of insects as food and feed in assuring food security. Annu Rev Entomol 58:563–583
- 9. de Castro RJS, Ohara A, dos SantosAguilar JG, Domingues MAF (2018) Nutritional, functional and biological properties of insect proteins: processes for obtaining, consumption and future challenges. Trends Food Sci Technol 76:82–89
- 10. Gravel A, Doyen A (2020) The use of edible insect proteins in food: challenges and issues related to their functional properties. Innov Food Sci Emerg Technol 59:102272
- 11. FAO (2021) Looking at edible insects from a food safety perspective. Challenges and opportunities for the sector. FAO, Rome, Italy
- 12. Hartmann C, Shi J, Giusto A, Siegrist M (2015) The psychology of eating insects: a cross-cultural comparison between Germany and China. Food Qua Prefer 44:148–156
- 13. Borges MM, da Costa DV, Trombete FM, Câmara AKFI (2022) Edible insects as a sustainable alternative to food products: an insight into quality aspects of reformulated bakery and meat products. Curr Opin Food Sci 46:100864
- 14. Severini C, Azzollini D, Albenzio M, Derossi AJ (2018) On printability, quality and nutritional properties of 3D printed cereal based snacks enriched with edible insects. Food Res Int 106:666–676
- 15. Sotelo-Díaz L et al (2022) Cricket four in a traditional beverage (*chucula*): emotions and perceptions of Colombian consumers. J Insects Food Feed 8(6):659–671
- 16. Osimani A et al (2018) Bread enriched with cricket powder (*Acheta domesticus*): a technological, microbiological and nutritional evaluation. Innov Food Sci Emerg Technol 48:150–163
- 17. Cheseto X, Baleba SB, Tanga CM, Kelemu S, Torto B (2020) Chemistry and sensory characterization of a bakery product prepared with oils from African edible insects. Foods 9(6):800
- 18. Carcea MJF (2020) Quality and nutritional/textural properties of durum wheat pasta enriched with cricket powder. Foods 9:1298
- 19. Megido RC et al (2016) Consumer acceptance of insect-based alternative meat products in Western countries. Food Qual Prefer 52:237–243
- 20. Choi Y-S et al (2017) Optimization of replacing pork meat with yellow worm (*Tenebrio molitor* L.) for frankfurters. Korean J Food Sci Anim Resour 37(5):617
- 21. Scholliers J, Steen L, Fraeye I (2020) Partial replacement of meat by superworm (*Zophobas morio* larvae) in cooked sausages: efect of heating temperature and insect: Meat ratio on structure and physical stability. Innov Food Sci Emerg Technol 66:102535
- 22. Tello A, Aganovic K, Parniakov O, Carter A, Heinz V, Smetana S (2021) Product development and environmental impact of an insect-based milk alternative. Future Foods 4:100080
- 23. Miladinov G (2023) Impacts of population growth and economic development on food security in low-income and middle-income countries. Front Hum Dyn 5:1121662
- 24. Tao J, Li YO (2018) Edible insects as a means to address global malnutrition and food insecurity issues. Food Qual Saf 2(1):17–26
- 25. Lange KW, Nakamura Y (2021) Edible insects as future food: chances and challenges. J Future Foods 1(1):38–46
- 26. Bessa LW, Pieterse E, Sigge G, Hoffman LC (2020) Insects as human food; from farm to fork. J Sci Food Agric 100(14):5017–5022
- 27. Belluco S et al (2013) Edible insects in a food safety and nutritional perspective: a critical review. Comp Rev Food Sci Food Safe 12(3):296–313
- 28. Ordoñez-Araque R, Egas-Montenegro E (2021) Edible insects: A food alternative for the sustainable development of the planet. Int J Gastronomy Food Sci 23:100304
- 29. Liceaga AM, Aguilar-Toalá JE, Vallejo-Cordoba B, González-Córdova AF, Hernández-Mendoza A (2022) Insects as an alternative protein source. Annu Rev Food Sci Technol 13:19–34
- 30. Womeni HM et al (2009) Oils of insects and larvae consumed in Africa: potential sources of polyunsaturated fatty acids. Oléagineux Corps Gras Lipid 16(4-5–6):230–235
- 31. Rumpold BA, Schlüter OK (2013) Nutritional composition and safety aspects of edible insects. Mol Nutr Food Res 57(5):802–823
- 32. Zamudio-Flores PB, Hernández-Gonzaléz M, García-Cano VJ (2019) Food supplements from a grasshopper: a developmental stage-wise evaluation of amino acid profle, protein and vitamins in *Brachystola magna* (Girard). Emir J Food Agric 31:561–568
- 33. Kouřimská L, Adámková A (2016) Nutritional and sensory quality of edible insects. NFS J 4:22–26
- 34. Institute of Medicine (US) Standing Committee on the Scientifc Evaluation of Dietary Reference Intakes (1998) Uses of dietary reference intakes. In: Dietary reference intakes for thiamin, ribofavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline. National Academies Press (US), Washington, DC
- 35. Wu X, He K, Velickovic TC, Liu Z (2021) Nutritional, functional, and allergenic properties of silkworm pupae. Food Sci Nutr 9(8):4655–4665
- 36. Melo-Ruiz V, Quirino-Barreda T, Calvo-Carrillo C, Sánchez-Herrera K, Sandoval-Trujillo HJ (2013) Assessment of nutrients of escamoles ant eggs *Limotepum apiculatum* M. by spectroscopy methods. J Chem Chem Eng 7(12):1181
- 37. Bukkens SGF (2005) Insects in the human diet: nutritional aspects. Ecological implications of minilivestock: potential of insects, rodents, frogs and snails, pp 545–577
- 38. Oonincx DG, Van Itterbeeck J, Heetkamp MJ, Van Den Brand H, Van Loon JJ, Van Huis AJ (2010) An exploration on greenhouse gas and ammonia production by insect species suitable for animal or human consumption. PLoS ONE 5(12):e14445
- 39. Köhler R, Kariuki L, Lambert C, Biesalski H (2019) Protein, amino acid and mineral composition of some edible insects from Thailand. J Asia-Pac Entomol 22(1):372–378
- 40. Ghosh S, Lee S-M, Jung C, Meyer-Rochow V (2017) Nutritional composition of fve commercial edible insects in South Korea. J Asia-Pac Entomol 20(2):686–694
- 41. Zielińska E, Baraniak B, Karaś M, Rybczyńska K, Jakubczyk A (2015) Selected species of edible insects as a source of nutrient composition. Food Res Int 77:460–466
- 42. da Silva Lucas AJ, Oreste EQ, Costa HLG, López HM, Saad CDM, Prentice C (2021) Extraction, physicochemical characterization, and morphological properties of chitin and chitosan from cuticles of edible insects. Food Chem 343:128550
- 43. Lopez-Santamarina A, del Carmen Mondragon A, Lamas A, Miranda JM, Franco CM, Cepeda A (2020) Animal-origin prebiotics based on chitin: An alternative for the future? A critical review. Foods 9(6):782
- 44. Van Huis A et al (2013) Edible insects: future prospects for food and feed security (no. 171). Food and agriculture organization of the United Nations, Rome
- 45. Van Huis A (2020) Insects as food and feed, a new emerging agricultural sector: a review. J Insects Food Feed 6:27–44
- 46. Wade M, Hoelle J (2020) A review of edible insect industrialization: scales of production and implications for sustainability. Environ Res Lett 15(12):123013
- 47. Sogari G, Dagevos H, Amato M, Taufk D (2022) Consumer perceptions and acceptance of insects as feed and food: current fndings and future outlook. In: Scafardi L, Formici G (eds) Novel foods and edible insects in the European Union: an interdisciplinary analysis. Springer International Publishing, Cham, pp 147–169
- 48. European Commission (2023) Approval of fourth insect as a novel food [https://food.ec.europa.eu/safety/novel-food/autho](https://food.ec.europa.eu/safety/novel-food/authorisations/approval-insect-novel-food_en) [risations/approval-insect-novel-food_en.](https://food.ec.europa.eu/safety/novel-food/authorisations/approval-insect-novel-food_en) Accessed 25 December 2023
- 49. Precup G et al (2022) The safety assessment of insects and products thereof as novel foods in the European Union. In: Scafardi L, Formici G (eds) Novel foods and edible insects in the European Union: an interdisciplinary analysis. Springer International Publishing, New York, pp 123–146
- 50. EFSA NDA Panel et al (2021) Safety of dried yellow mealworm (*Tenebrio molitor* larva) as a novel food pursuant to regulation (EU) 2015/2283. EFSA J 19(1):e06343
- 51. EFSA NDA Panel et al (2021) Safety of frozen and dried formulations from whole yellow mealworm (*Tenebrio molitor* larva) as a novel food pursuant to regulation (EU) 2015/2283. EFSA J 19(8):e06778
- 52. EFSA NDA Panel et al (2021) Safety of frozen and dried formulations from migratory locust (*Locusta migratoria*) as a novel food pursuant to regulation (EU) 2015/2283. EFSA J 19(7):e06667
- 53. EFSA NDA Panel et al (2021) Safety of frozen and dried formulations from whole house crickets (*Acheta domesticu*s) as a novel food pursuant to regulation (EU) 2015/2283. EFSA J 19(8):e06779
- 54. EFSA NDA Panel et al (2022) Safety of partially defatted house cricket (*Acheta domesticus*) powder as a novel food pursuant to Regulation (EU) 2015/2283. EFSA J 20(5):e07258
- 55. EFSA Nda Panel et al (2022) Safety of frozen and freeze-dried formulations of the lesser mealworm (*Alphitobius diaperinus* larva) as a Novel food pursuant to Regulation (EU) 2015/2283. EFSA J 20(7):e07325
- 56. Commission Implementing Regulation (EU) 2022/169 of 8 February 2022 authorising the placing on the market of frozen, dried and powder forms of yellow mealworm (*Tenebrio molitor* larva) as a novel food under regulation (EU) 2015/2283 of the European Parliament and of the Council, and Amending Commission Implementing Regulation (EU) 2017/2470 (Text with EEA Relevance). 2022; vol 28. [http://data.europa.eu/eli/reg_impl/2022/](http://data.europa.eu/eli/reg_impl/2022/169/oj/eng) [169/oj/eng](http://data.europa.eu/eli/reg_impl/2022/169/oj/eng). Accessed 25 December 2023
- 57. Commission Implementing Regulation (EU) 2021/1975 of 12 November 2021 authorising the placing on the market of frozen,

dried and powder forms of *Locusta migratoria* as a novel food under regulation (EU) 2015/2283 of the European Parliament and of the Council and Amending Commission Implementing Regulation (EU) 2017/2470 (text with EEA relevance). 2021; vol 402. [http://data.europa.eu/eli/r](http://data.europa.eu/eli/)eg_impl/2021/1975/oj/eng. Accessed 25 December 2023

- 58. Commission Implementing Regulation (EU) 2022/188 of 10 February 2022 authorising the placing on the market of frozen, dried and powder forms of *Acheta domesticus* as a novel food under regulation (EU) 2015/2283 of the European Parliament and of the Council and amending commission implementing regulation (EU) 2017/2470 (text with EEA relevance). 2022; vol 030. <http://data.europa.eu/eli/>reg_impl/2022/188/oj/eng. Accessed 25 December 2023
- 59. Commission Implementing Regulation (EU) 2023/5 of 3 January 2023 authorising the placing on the market of *Acheta domesticus* (house cricket) partially defatted powder as a novel food and amending implementing regulation (EU) 2017/2470 (text with EEA Relevance). 2023; vol 002. [http://data.europa.eu/eli/reg_](http://data.europa.eu/eli/reg_impl/2023/5/oj/) [impl/2023/5/oj/](http://data.europa.eu/eli/reg_impl/2023/5/oj/)eng. Accessed 25 December 2023
- 60. Commission Implementing Regulation (EU) 2023/58 of 5 January 2023 authorising the placing on the market of the frozen, paste, dried and powder forms of *Alphitobius diaperinus* larvae (lesser mealworm) as a novel food and amending implementing regulation (EU) 2017/2470 (text with EEA relevance). 2023; vol 5. [http://data.europa.eu/eli/reg_impl/2023/58/oj.](http://data.europa.eu/eli/reg_impl/2023/58/oj) Accessed 25 December 2023
- 61. Baigts-Allende D et al (2021) Insect protein concentrates from Mexican edible insects: Structural and functional characterization. LWT 152:112267
- 62. Mancini S, Sogari G, Espinosa Diaz S, Menozzi D, Paci G, Moruzzo RJF (2022) Exploring the future of edible insects in Europe. Foods 11(3):455
- 63. Ardoin R, Prinyawiwatkul W (2021) Consumer perceptions of insect consumption: a review of western research since 2015. Int J Food Sci Technol 56(10):4942–4958
- 64. Puteri B, Jahnke B, Zander KJA (2023) Booming the bugs: How can marketing help increase consumer acceptance of insect-based food in western countries? Appetite 187:106594
- 65. Baker MA, Shin JT, Kim YW (2016) An exploration and investigation of edible insect consumption: the impacts of image and description on risk perceptions and purchase intent. Psychol Market 33(2):94–112
- 66. Baiano A (2020) Edible insects: an overview on nutritional characteristics, safety, farming, production technologies, regulatory framework, and socio-economic and ethical implications. Trends Food Sci Technol 100:35–50
- 67. Meshulam-Pascoviche D, David-Birman T, Refael G, Lesmes U (2022) Big opportunities for tiny bugs: processing efects on the techno-functionality and digestibility of edible insects. Trends Food Sci Technol 122:265–274
- 68. Gnana Moorthy Eswaran U, Karunanithi S, Gupta RK, Rout S, Srivastav PP (2023) Edible insects as emerging food products processing and product development perspective. J Food Sci Technol 60(8):2105–2120
- 69. Garofalo C, Milanović V, Cardinali F, Aquilanti L, Clementi F, Osimani A (2019) Current knowledge on the microbiota of edible insects intended for human consumption: a state-of-the-art review. Food Res Int 125:108527
- 70. Vandeweyer D, De Smet J, Van Looveren N, Van Campenhout L (2021) Biological contaminants in insects as food and feed (in English). J Insects Food Feed 7(5):807–822
- 71. Ribeiro JC, Cunha LM, Sousa-Pinto B, Fonseca J (2018) Allergic risks of consuming edible insects: a systematic review. Mol Nutr Food Res 62(1):1700030
- 72. Gałęcki R, Bakuła T, Gołaszewski J (2023) Foodborne diseases in the edible insect industry in Europe—new challenges and old problems. Foods 12(4):770
- 73. Baigts-Allende DK, Stathopoulos C (2023) Overcoming obstacles in insect utilization. Eur Food Res Technol 249(4):849–860
- 74. Pali-Schöll I et al (2019) Edible insects: cross-recognition of IgE from crustacean-and house dust mite allergic patients, and reduction of allergenicity by food processing. World Allergy Organ 12(1):100006
- 75. Ojha S, Bußler S, Psarianos M, Rossi G, Schlüter O (2021) Edible insect processing pathways and implementation of emerging technologies. J Insects Food Feed 7(5):877–900
- 76. Castro-López C et al (2020) An insight to fermented edible insects: a global perspective and prospective. Food Res Int 137:109750
- 77. Van Huis A, Rumpold B, Maya C, Roos N (2021) Nutritional qualities and enhancement of edible insects. Annu Rev Nutr 41:551–576
- 78. Vandeweyer D, Lenaerts S, Callens A, Van Campenhout L (2017) Efect of blanching followed by refrigerated storage or industrial microwave drying on the microbial load of yellow mealworm larvae (*Tenebrio molitor*). Food Control 71:311–314
- 79. Wynants E et al (2018) Microbial dynamics during production of lesser mealworms (*Alphitobius diaperinus*) for human consumption at industrial scale. Food Microbiol 70:181–191
- 80. Larouche J, Deschamps M-H, Saucier L, Lebeuf Y, Doyen A, Vandenberg GW (2019) Effects of killing methods on lipid oxidation, colour and microbial load of black soldier fy (*Hermetia illucens*) larvae. Animals 9(4):182
- 81. Bußler S et al (2016) Cold atmospheric pressure plasma processing of insect four from *Tenebrio molitor*: Impact on microbial load and quality attributes in comparison to dry heat treatment. Innov Food Sci Emerg Technol 36:277–286
- 82. Nyangena DN et al (2020) Efects of traditional processing techniques on the nutritional and microbiological quality of four edible insect species used for food and feed in East Africa. Foods 9(5):574
- 83. Megido RC et al (2018) Efect of household cooking techniques on the microbiological load and the nutritional quality of mealworms (*Tenebrio molitor* L. 1758). Food Res Int 106:503–508
- 84. Lenaerts S, Van Der Borght M, Callens A, Van Campenhout L (2018) Suitability of microwave drying for mealworms (*Tenebrio molitor*) as alternative to freeze drying: impact on nutritional quality and colour. Food Chem 254:129–136
- 85. Azzollini D, Derossi A, Severini C (2016) Understanding the drying kinetic and hygroscopic behaviour of larvae of yellow mealworm (*Tenebrio molitor*) and the effects on their quality. J Insects Food Feed 2(4):233–243
- 86. Kröncke N, Böschen V, Woyzichovski J, Demtröder S, Benning R (2018) Comparison of suitable drying processes for mealworms (*Tenebrio molitor*). Innov Food Sci Technol 50:20–25
- 87. Singh Y, Cullere M, Kovitvadhi A, Chundang P, DalleZotte A (2020) Efect of diferent killing methods on physicochemical traits, nutritional characteristics, in vitro human digestibility and oxidative stability during storage of the house cricket (*Acheta domesticus* L.). Innov Food Sci Emerg Technol 65:102444
- 88. Fombong FT, Van der Borght M, Van den Broeck J (2017) Infuence of freeze-drying and oven-drying post blanching on the nutrient composition of the edible insect *Ruspolia diferens*. Insects 8(3):102
- 89. Khampakool A, Soisungwan S, You S, Park SH (2020) Infrared assisted freeze-drying (IRAFD) to produce shelf-stable insect food from *Protaetia brevitarsis* (white-spotted fower chafer) larva. Food Sci Anim Resour 40(5):813–830
- 90. Liceaga AM (2021) Processing insects for use in the food and feed industry. Curr Opin Insect Sci 48:32–36
- 91. Ndiritu AK, Kinyuru JN, Kenji GM, Gichuhi PN (2017) Extraction technique infuences the physico-chemical characteristics and functional properties of edible crickets (*Acheta domesticus*) protein concentrate. J Food Meas Characterization 11:2013–2021
- 92. Zhao X, Vázquez-Gutiérrez JL, Johansson DP, Landberg R, Langton MJ (2016) Yellow mealworm protein for food purposesextraction and functional properties. PLoS ONE 11(2):e0147791
- 93. Purschke B, Sanchez YDM, Jäger H (2018) Centrifugal fractionation of mealworm larvae (*Tenebrio molitor*, L.) for protein recovery and concentration. LWT 89:224–228
- 94. Purschke B, Tanzmeister H, Meinlschmidt P, Baumgartner S, Lauter K, Jäger H (2018) Recovery of soluble proteins from migratory locust (*Locusta migratoria*) and characterisation of their compositional and techno-functional properties. Food Res Int 106:271–279
- 95. Purschke B, Brüggen H, Scheibelberger R, Jäger H (2018) Efect of pre-treatment and drying method on physico-chemical properties and dry fractionation behaviour of mealworm larvae (*Tenebrio molitor* L.). Eur Food Res Technol 244:269–280
- 96. Mishyna M, Martinez J-JI, Chen J, Benjamin OJ (2019) Extraction, characterization and functional properties of soluble proteins from edible grasshopper (*Schistocerca gregaria*) and honey bee (*Apis mellifera*). Food Res Int 116:697–706
- 97. Mariod AA, Fadul H (2015) Extraction and characterization of gelatin from two edible Sudanese insects and its applications in ice cream making. Food Sci Technol Int 21(5):380–391
- 98. Psarianos M et al (2022) Efect of pulsed electric felds on cricket (*Acheta domesticus*) four: extraction yield (protein, fat and chitin) and techno-functional properties. Innov Food Sci Emerg Techno 76:102908
- 99. Choi BD, Wong NA, Auh J-H (2017) Defatting and sonication enhances protein extraction from edible insects. Korean J Food Sci Anim Resour 37(6):955
- 100. Tzompa-Sosa DA, Yi L, van Valenberg HJ, van Boekel MA, Lakemond CM (2014) Insect lipid profile: aqueous versus organic solvent-based extraction methods. Food Res Int 62:1087–1094
- 101. Purschke B, Stegmann T, Schreiner M, Jäger HJE (2017) Pilotscale supercritical CO2 extraction of edible insect oil from *Tenebrio molitor* L. larvae–infuence of extraction conditions on kinetics, defatting performance and compositional properties. Eur J Lipid Sci Technol 119(2):1600134
- 102. Sun M, Xu X, Zhang Q, Rui X, Wu J, Dong MJ (2018) Ultrasonic-assisted aqueous extraction and physicochemical characterization of oil from *Clanis bilineata*. J Oleo Sci 67(2):151–165
- 103. Smets R, Goos P, Claes J, Van Der Borght MJS (2021) Optimisation of the lipid extraction of fresh black soldier fy larvae (*Hermetia illucens*) with 2-methyltetrahydrofuran by response surface methodology. Sep Purif Technol 258:118040
- 104. Ramos-Bueno RP, González-Fernández MJ, Sánchez-Muros-Lozano MJ, García-Barroso F, Guil-Guerrero JL (2016) Fatty acid profles and cholesterol content of seven insect species assessed by several extraction systems. Eur J Food Res Technol 242:1471–1477
- 105. Kim D, Oh I (2022) The characteristic of insect oil for a potential component of oleogel and its application as a solid fat replacer in cookies. Gels 8(6):355
- 106. Otero P, Gutierrez-Docio A, Del Hierro JN, Reglero G, Martin D (2020) Extracts from the edible insects Acheta domesticus and Tenebrio molitor with improved fatty acid profle due to ultrasound assisted or pressurized liquid extraction. Food Chem 314:126200
- 107. Ugur AE, Bolat B, Oztop MH, Alpas H (2021) Efects of high hydrostatic pressure (HHP) processing and temperature on physicochemical characterization of insect oils extracted from *Acheta*

domesticus (house cricket) and *Tenebrio molitor* (yellow mealworm). Waste Biomass Valor 12:4277–4286

- 108. Liu S et al (2012) Extraction and characterization of chitin from the beetle Holotrichia parallela Motschulsky. Molecules 17(4):4604–4611
- 109. Kaya M, Sargin I, Erdonmez D (2016) Microbial bioflm activity and physicochemical characterization of biodegradable and edible cups obtained from abdominal exoskeleton of an insect. Innov Food Sci Emerg Technol 36:68–74
- 110. Kim T-K, Yong HI, Kim Y-B, Jung S, Kim H-W, Choi Y-S (2021) Efects of organic solvent on functional properties of defatted proteins extracted from *Protaetia brevitarsis* larvae. Food Chem 336:127679
- 111. van Huis AJ (2022) Edible insects: challenges and prospects. Entomol Res 52(4):161–177
- 112. Zhang F et al (2023) Mechanisms underlying the efects of ultrasound-assisted alkaline extraction on the structural properties and in vitro digestibility of *Tenebrio molitor* larvae protein. Ultrason Sonochem 94:106335
- 113. Younes I, Rinaudo M (2015) Chitin and chitosan preparation from marine sources. Structure, properties and applications. Mar Drugs 13(3):1133–1174
- 114. González CM, Garzón R, Rosell CM (2019) Insects as ingredients for bakery goods. A comparison study of *H. illucens*, *A.* domestica and *T. molitor* flours. Innov Food Sci Emerg Technol 51:205–210
- 115. Biró B, Fodor R, Szedljak I, Pásztor-Huszár K, Gere A (2019) Buckwheat-pasta enriched with silkworm powder: technological analysis and sensory evaluation. LWT 116:108542
- 116. Biró B, Sipos MA, Kovács A, Badak-Kerti K, Pásztor-Huszár K, Gere A (2020) Cricket-enriched oat biscuit: technological analysis and sensory evaluation. Foods 9(11):1561
- 117. Indriani S, Ab Karim MSB, Nalinanon S, Karnjanapratum S (2020) Quality characteristics of protein-enriched brown rice four and cake afected by Bombay locust (*Patanga succincta* L.) powder fortifcation. LWT 119:108876
- 118. Delicato C, Schouteten JJ, Dewettinck K, Gellynck X, Tzompa-Sosa DA (2020) Consumers' perception of bakery products with insect fat as partial butter replacement. Food Qual Prefer 79:103755
- 119. Çabuk B (2021) Infuence of grasshopper (*Locusta migratoria*) and mealworm (*Tenebrio molitor*) powders on the quality characteristics of protein rich muffins: nutritional, physicochemical, textural and sensory aspects. J Food Meas Characterization 15(4):3862–3872
- 120. Kim HW, Setyabrata D, Lee Y, Jones OG, Kim YHB (2017) Efect of house cricket (*Acheta domesticus*) four addition on physicochemical and textural properties of meat emulsion under various formulations. J Food Sci 82(12):2787–2793
- 121. Park Y-S et al (2017) Physicochemical properties of meat batter added with edible silkworm pupae (*Bombyx mori*) and transglutaminase. Korean J Food Sci Anim Resour 37(3):351–359
- 122. Kim H-W, Setyabrata D, Lee Y-J, Jones OG, Kim YHB (2016) Pre-treated mealworm larvae and silkworm pupae as a novel protein ingredient in emulsion sausages. Innov Food Sci Emerg Technol 38:116–123
- 123. Tzompa-Sosa DA, Dewettinck K, Gellynck X, Schouteten JJ (2021) Replacing vegetable oil by insect oil in food products: Efect of deodorization on the sensory evaluation. Food Res Int 141:110140
- 124. Tzompa-Sosa DA, Dewettinck K, Gellynck X, Schouteten JJ (2022) Consumer acceptance towards potato chips fried in yellow mealworm oil. Food Qual Prefer 97:104487
- 125. Tzompa-Sosa DA, Provijn P, Gellynck X, Schouteten J (2023) Frying dough with yellow mealworm oil: aroma profle and

consumer perception at a central location test and at home. J Food Sci 88(S1):A130–A146

- 126. Kyriakopoulou K, Dekkers B, van der Goot AJ (2019) Plantbased meat analogues. Sustainable meat production and processing. Elsevier, Amsterdam, pp 103–126
- 127. Paul AA, Kumar S, Kumar V, Sharma RJ (2020) Milk analog: plant based alternatives to conventional milk, production, potential and health concerns. Crit Rev Food Sci Nutr 60(18):3005–3023
- 128. Lima M, Costa R, Rodrigues I, Lameiras J, Botelho G (2022) A narrative review of alternative protein sources: highlights on meat, fsh, egg and dairy analogues. Foods 11(14):2053
- 129. Pingali P, Boiteau J, Choudhry A, Hall AJ (2023) Making meat and milk from plants: a review of plant-based food for human and planetary health. World Dev 170:106316
- 130. Maningat CC, Jeradechachai T, Buttshaw MR (2022) Textured wheat and pea proteins for meat alternative applications. Cereal Chem 99(1):37–66
- 131. Bachmann H-P (2001) Cheese analogues: a review. Int Diary J 11(4–7):505–515
- 132. Kamath R, Basak S, Gokhale J (2022) Recent trends in the development of healthy and functional cheese analogues—a review. Lebensmittel-Wissensch Technol 155:112991
- 133. Smetana S, Leonhardt L, Kauppi S-M, Pajic A, Heinz V (2020) Insect margarine: processing, sustainability and design. J Clean Prod 264:121670
- 134. Moura M, Martins BA, Oliveira GP, Takahashi JA (2022) Alternative protein sources of plant, algal, fungal and insect origins for dietary diversifcation in search of nutrition and health. Crit Rev Food Sci Nutr 63(31):10691–10708
- 135. Kim TK, Yong HI, Cha JY, Park SY, Jung S, Choi YS (2022) Drying-induced restructured jerky analog developed using a combination of edible insect protein and textured vegetable protein. Food Chem 373(Pt B):131519
- 136. Smetana S et al (2018) Structure design of insect-based meat analogs with high-moisture extrusion. J Food Eng 229:83–85
- 137. Azzollini D, Wibisaphira T, Lakemond C, Fogliano V (2019) Toward the design of insect-based meat analogue: the role of calcium and temperature in coagulation behavior of *Alphitobius diaperinus* proteins. LWT 100:75–82
- 138. Cho SY, Ryu GH (2021) Efects of mealworm larva composition and selected process parameters on the physicochemical properties of extruded meat analog. Food Sci Nutr 9(8):4408–4419
- 139. Stoops J et al (2017) Minced meat-like products from mealworm larvae (*Tenebrio molitor* and *Alphitobius diaperinus*): microbial dynamics during production and storage. Innov Food Sci Emerg Technol 41:1–9
- 140. Gould J, Wolf B (2018) Interfacial and emulsifying properties of mealworm protein at the oil/water interface. Food Hydrocolloids 77:57–65
- 141. Chou TH, Nugroho DS, Cheng YS, Chang JY (2020) Development and characterization of nano-emulsions based on oil extracted from black soldier fy larvae. Appl Biochem Biotechnol 191(1):331–345
- 142. Shelomi M (2020) Potential of black soldier fy production for pacifc small island developing states. Animals 10(6):1038

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