ORIGINAL PAPER



Characterization of the key aroma compounds in three world-famous black teas

Zuobing Xiao^{1,2} · Xueying Cao¹ · Jiancai Zhu¹ · Feng Chen³ · Yunwei Niu¹

Received: 13 March 2022 / Revised: 18 April 2022 / Accepted: 23 April 2022 / Published online: 24 May 2022 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2022

Abstract

The volatile compounds of three world-famous black teas (Darjeeling, DJL, Keemun, KM, and Ceylon, CL) were extracted by stir bar sorptive extraction (SBSE), and analyzed by gas chromatography–olfactometry (GC–O), gas chromatography– mass spectrometry (GC–MS). The results indicated that 78, 76, and 69 volatile compounds were detected in the three tea infusions. And 9 sulfur compounds in black teas were identified by gas chromatography–flame photometric detection (GC–FPD). In addition, a total of 42 aroma compounds were perceived and 38 compounds were identified as important aroma compounds due to their high odor activity values (OAVs), such as 3-methylbutanal (OAV: 24–82), linalool (OAV: 24–64), geraniol (OAV: 2–97), β -ionone (OAV: 54–122), and *cis*-jasmone (OAV: 2–119). According to the results of aroma recombination and omission experiments, 2-methylbutanal, linalool, methyl salicylate and β -cyclocitral were confirmed to be the key aroma compounds in Darjeeling black tea, 3-methylbutanal, hexanal, β -myrcene, and methyl salicylate were the key aroma compounds in Keemun, while β -ionone, linalool, 2-methylbutanal, and salicylaldehyde were the key aroma compounds in Ceylon black tea.

Keywords Black tea \cdot OAV \cdot Omission experiment \cdot Aroma recombination

Introduction

Tea, originating in China, is one of the world's three major non-alcoholic beverages along with coffee and cocoa. Owing to its unique taste and special aroma, tea has been popular for centuries [1]. Internationally, according to the degree of fermentation, tea can be classified into three main types: unfermented tea (green tea), semi-fermented tea (oolong tea), and fermented tea (black tea) [2]. Black tea is fermented, and the process of black tea production consists of four steps: withering, rolling, fermentation, and firing [3]. Besides, because black tea has an abundant aroma and a long cultural history, it is loved by consumers all over the world

⊠ Yunwei Niu yunweiniu@163.com

- ¹ Department of Perfume and Aroma Technology, Shanghai Institute of Technology, Shanghai, People's Republic of China
- ² School of Agriculture and Biology, Shanghai Jiao Tong University, Shanghai, People's Republic of China
- ³ Department of Food, Nutrition and Packaging Sciences, Clemson University, Clemson, USA

and has become one of the most widely consumed teas [4]. Black tea can be divided into many types, according to the geographical origin [5], growth climate [6], degree of fermentation [7], and tea variety [3].

Although the aroma compounds in tea only account for 0.01% of the dry weight of tea leaves [8], aroma is one of the essential factors for sensory evaluation of black tea [9]. Different characteristics and styles of black tea are formed due to the different types, contents, ratios and interactions of aroma compounds. Among the numerous types of black teas, Keemun from China, Darjeeling from India, and Ceylon from Sri Lanka are honored as the top three of higharoma black teas in the world, due to their rich and unique aroma qualities [10]. According to the literature, Darjeeling black tea has a musky scent, Keemun black tea shows a smoky smell, while Ceylon black tea presents a typical floral and sweet fragrance [11]. The aroma compounds in black tea have been extensively studied. For example, Magagna et al. [12] analyzed the volatiles in Ceylon black tea using two-dimensional gas chromatography-mass spectrometry (GC×GC-MS), and a total of 123 compounds were identified. The volatile compounds in Chinese congou black tea were investigated through aroma extract dilution analysis

and odor active values, indicating that *cis*-3-hexen-1-ol, 1-octen-3-ol, *trans*-linalooloxide, linalool, benzyl alcohol, phenylethyl alcohol, valeraldehyde, hexanal, etc. make important contributions to the aroma of black tea [10]. However, the characteristic compounds and differences among the three world-famous black teas have not been studied systematically. The identification of characteristic aroma compounds can enrich the theoretical basis of tea flavor chemistry, enhance the comprehension of the chemical compounds which contribute to the tea's aroma quality, and provide a theoretical basis for quality identification among black tea specimen.

Since the twentieth century, gas chromatography–mass spectrometry (GC–MS) has developed rapidly to analyze the aroma compounds of tea. At present, more than 600 substances have been identified, including alcohols, aldehydes, acids, ketones, esters, sulfur, furans, lactones, etc. [13] Combining GC–MS with olfactometry can not only isolate and identify the aroma compounds in black tea, but also can sense the aroma characteristics of aroma compounds [14]. By GC–MS and GC–O (gas, chromatography–olfactometry), the volatile compounds in Hanzhong black tea were analyzed, indicating that linalooloxide, 1-octen-3-one, (E, E)-2,4-nonadienal, etc. were considered as the characteristic aroma compounds in tea infusion [15].

In particular, the identification of aroma compounds will vary greatly when different extraction methods are used, therefore, choosing appropriate extraction methods are important to the identification of aroma compounds in black tea. Commonly, the extraction methods applied in tea include simultaneous distillation extraction (SDE) [16], solvent-assisted flavor evaporation (SAFE) [17], and headspace solid phase micro-extraction (HS-SPME) [18, 19]. Since SDE specimens might introduce new substances under high temperature for a long time, it is unsuitable for extracting volatile compounds from tea. SAFE is carried out at a low temperature and low pressure, which will not destroy the original aroma, but it will consume significant volumes of solvents. SPME can quickly and easily absorb aroma compounds [20], but it is not suitable for aroma compounds with high boiling point. Noticeably, stir bar sorptive extraction (SBSE) is a new technique for enrichment of volatile and semi-volatile compounds from aqueous samples [21]. Besides, according to the literature, SBSE has the advantages of large extraction coating volume, high extraction capacity, high sensitivity, small sample amount, and environmental friendliness [22]. In recent years, SBSE has been widely used in the detection of aroma compounds in water, wine, fruits, etc. [23] Thus, SBSE might have great potential in the aroma analysis of black tea. Unfortunately, few people use SBSE to study the aroma of black tea, and there are few systematic comparisons of the characteristic aromas of the top three of high-aroma black teas in the world.

Therefore, the main purposes of this research were as follows: (1) to identify the volatile compounds of the three world-famous black teas by GC–MS and GC–FPD; (2) to determine the characteristic aroma compounds by gas chromatography–olfactometry (GC–O) and OAV; (3) to confirm the key aroma compounds by aroma recombination and omission experiments. The analysis of the characteristic aroma of these black teas will not only complement the flavor map of black tea, but also enable a more comprehensive understanding of the differences between them.

Materials and methods

Materials

Three kinds of black teas (Darjeeling black tea (DJL), Keemun black tea (KM) and Ceylon black teas (CL)) used in the experiment were purchased from tea trade (Redsunion Co., Ltd. (Qimen City, Anhui Province, China), Whittard Co., Ltd. (London, UK) and Basilur Co., Ltd. (Sri Lanka, India), respectively). Before the experiment, all black tea samples were stored in aluminum foil, and kept in a refrigerator at 4 °C. Each black tea sample was ground into powder using mortar before experiments.

Chemicals

The information about standard compounds for qualitative and quantitative analysis has been given in Supporting Information. All the standard compounds used were GC quality. N-alkanes (C_6-C_{30}) for linear retention index determination were purchased from Shanghai Titan technology Co., Ltd. (Shanghai, China). Pure water was obtained from Milli-Q purification system (Millipore, Bedford, MA, USA).

Stir bar sorptive extraction

The aroma compounds in black teas were extracted by Stir Bar Sorptive Extraction (SBSE) method, and an ACAR/ PDMS twister (10 mm length and 0.5 mm thickness, Gerstel, Germany) was used. The SBSE extraction method was slightly modified on the basis of previous literature [24]. For each sample, 0.3 g of tea powder was weighted and poured into a 20 mL sample bottle, then 15 mL hot water (90 °C), 0.5 g NaCl and 5 μ L internal standard solution 2-octanol (400 μ g/mL in ethanol) were added. After 5 min of equilibration, the twister was immersed in the black tea infusion, and then the sample bottle was placed in a water bath magnetic stirrer (SHJ-4D, Yuhua, Tianjin, China), and extracted at 60 °C for 60 min with an agitation rate of 1250 rpm. Afterward, the twister was washed and dried with a lintfree tissue and then placed in the thermal desorption unit (TDU, Gerstel, German) for subsequent GC–O and GC–MS analysis.

Thermal desorption

The parameters used for the thermal desorption unit (TDU, Gerstel, German) were: the desorption program was held at 40 °C for 1 min, and raised to 260 °C at a rate of 60 °C/min, then held for 5 min, in splitless mode. The cooled injection system (Gerstel CIS-4 PTV injector, German) was maintained at -70 °C using liquid nitrogen (99.999%). Then the temperature of CIS-4 was raised to 260 °C at a rate of 12 °C/s and held for 5 min.

GC-O analysis

In the GC-O experiment, a GC (7890A, Agilent, Santa Clara, CA) equipped with flame ionization detector (FID) and Olfactory Detector Port (ODP-3, Gerstel, Mulheim an der Ruhr, Germany) was used for aroma sniffing analysis. After chromatographic separation, the extracted volatile compounds were split between the FID and the GC-O sniffing port in a ratio of 1:1. The length between the splitter and the sniffing port was 107 cm. The chromatographic columns in GC-O experiment were HP-Innowax (60 m \times 0.25 mm \times 0.25 µm) and DB-5 $(60 \text{ m} \times 0.25 \text{ mm} \times 0.25 \text{ } \mu\text{m})$. The chromatographic columns were purchased from Agilent (Santa, Clara, CA). Helium (99.999%) was used as the carrier gas, and the flow rate was 2 mL/min. The temperature program of oven was maintained at 40 °C for 6 min, then raised to 150 °C at a rate of 3 °C/ min, and finally raised to 230 °C at a rate of 6 °C/min and kept it for 15 min. The splitless mode was adopted. The temperature of the injector was 250 °C, while the temperature of the FID detector was 280 °C.

The GC-O analysis was performed by an experienced sensory panel. The sensory panel consisted of 5 healthy and non-smoking members (two males and three females, with an average age of 24 years old) who performed GC-O sniffing. All of the panelists belonged to the School of Perfume and Aroma Technology of Shanghai Institute of Technology (Shanghai, China). They all had received aroma training at least 5 years. Before carrying out the formal experiment, the sensory panel was trained for odor characteristics and aroma intensity using standard compounds solution to be familiar with odor characteristics. During the training, group panelists were trained to unify the aroma intensity and description of the aroma. After training, five panelists performed GC-O sniffing on three tea samples. When an odor was perceived, sensory panelists recorded the retention time, aroma intensity (AI) and aroma characteristics. According to previous researches, each panelist scored the aroma intensity (AI) using an intensity scale from 0 to 10,

where "0" meant no aroma, "5" meant medium intensity, and "10" meant extremely strong [25]. Each panelist repeated the experiment three times, and the average scores of the intensity were taken as the aroma intensity of the aroma compounds.

GC–MS analysis

The aroma compounds in black teas were isolated and identified by GC–MS (7890-5977B, Agilent, Santa Clara, CA). The types of the chromatographic column and the temperature procedure of oven were consistent with GC–O. The temperature of injection port was set at 250 °C with split-less mode. The ion source, quadrupole mass filter, and default values of electron impact energy were 230 °C, 150 °C and 70 eV, respectively. Mass scanning range was 30–400 amu. The carrier gas was helium (99.999%) at a rate of 1 mL/min. The experiment was repeated in triplicate. The aroma substances were identified through matching the retention time of authentic standards, retention indices (RIs) and the MS in the NIST11 database. The RIs were calculated by retention time of the homogeneous serious of alkanes (C₆-C₃₀) [26].

HS-SPME-GC-FPD analysis

HS–SPME was applied to extract the sulfur compounds in black teas. The tea infusions were the same as SBSE method. 6.5 g of tea infusion of each sample and 10 μ L of internal standard solution (dipropyl disulfide, 4 mg/L) were added into screw-capped vials fitted with PTEE silicone septa. Then 75 μ m carboxyl-poly-dimethyl siloxane (CAR-PDMS) fiber purchased from Supelco, Inc. (Bellefonte, PA, USA) was exposed to the headspace of the tea infusions in a water bath at 60 °C for 50 min. Then SPME fiber was inserted into the GC injector for desorption and analyzed at 250 °C for 5 min.

The sulfur compounds in black teas were identified by Agilent 7890A GC equipped with a flame photometric detector (FPD). The columns and oven temperature procedure were consistent with GC–MS. The GC was operated in constant flow mode at a rate of 1 mL/min with helium as carrier gas. The temperature of the FPD detector was 250 °C, while the PMT voltage was set to 500 V. The desorption time was 5 min and the injection port was operated in a splitless mode. Sulfur compounds in black tea samples were determined by comparing with retention time of authentic standards and retention indices (RIs). The experiment was carried out in triplicate.

Calibration of standard curves

According to previous literature, the standard curves of aroma compounds were made with a slight modification

[27]. The reconstituted sample consisting of all aroma compounds detected by GC-MS in the three specimens was dissolved in deionized water according to the concentration calculated by the internal standard concentration. And then the mixture was diluted with water according to the ratio of 1:2, 1:5, 1:10, 1:20, 1: 50, 1:75 and 1:100. Afterward, 5 µL of the internal standard solution containing 2-octanol $(400 \ \mu g/mL)$ was introduced to the diluted solution. Then each ratio of the solution was adsorbed by SBSE to construct the standard curve of aroma compounds and the analysis conditions were consistent with the aroma analysis of black tea. The calibration curves were established by the ratio of aroma compounds to internal standard. The abscissa was the ratio of the concentration of compound to internal standard, and the ordinate was the ratio of the peak area of compound to internal standard. All detected aroma compounds were quantified by the established standard curves.

Similarly, the standard curves of sulfur compounds were constructed the same as the non-sulfur compounds, except that the internal standard solution was changed to dipropyl disulfide (4 mg/L), and the extraction method was changed to HS–SPME.

Odor activity values (OAVs)

The odor activity value (OAV) was the ratio of the concentration of aroma compounds to its threshold. OAV method was often applied to evaluate the contributions of aroma compounds. In this experiment, the threshold values in water were collected from the literature [28].

Aroma recombination and electronic nose

To further confirm the key aroma compounds in the black tea samples were correctly identified and quantified, the aroma combination method was applied. According to the results of GC–MS, GC–O and GC–FPD, the aroma compounds with OAV \geq 1 or AI > 0 were dissolved in water as their quantitative concentration.

A total of 15 volunteers (7 men and 8 women, aged 22–28) were recruited. These panelists were also selected from the School of Perfume and Aroma Technology of Shanghai Institute of Technology (Shanghai, China). Sensory evaluation was conducted in sensory laboratory (25 °C). A volume of 50 mL black tea infusion was placed in odorless plastic cup. Each panelists described the sensory properties of black tea infusions. Finally, panelists discussed their sensory results and adopted 8 descriptors: fruity, green, sweet, floral, mushroom-like, baked, caramel and woody. The intensity of these attributes was rated from 0 to 10, where "0" represented none, "5" represented moderate, and "10" is strongly perceivable [29]. Each panelist conducted each evaluation in triplicate, with 30 min rest after each test.

A HERACLES electronic nose (Alpha M.O.S., Toulouse, France) was applied to compare the difference between black tea infusion and aroma recombination model. 6.5 g of tea infusions or aroma recombination solutions were added into a 20 mL headspace vial. The parameters of the auto-sampler were: headspace heating for 20 min, headspace temperature was 50 °C, the stirring speed was 500 rpm, the stirring opening time was 5 s, the stirring closing time was 2 s, and the washing time was 90 s. Sample headspace injection volume was 5000 μ L, sampling speed was 125 μ L/s, and injection temperature was 200 °C. The electronic nose analysis was repeated three times.

Omission experiments

To further explore the contribution of the characteristic aroma compounds of black tea to the overall aroma, the omission experiments were adopted [26]. All aroma compounds with $OAV \ge 1$ or AI > 0 were selected for the omission experiments. The concentrations of all selected compounds were quantified by the standard curve. The differences between the omission model and aroma recombination model were evaluated through a triangle test according to the International Standard Method [30]. There were three bottles solution in each omission experiment. One bottle missed one aroma compound and the other two contained the whole aroma. The sensory panel of the omission experiment consisted of 15 members (7 males, 8 females, aged between 22 and 28), and each panelist was asked to select the different one from the three bottles. Each experiment was repeated three times.

Statistical analysis

The aroma intensity and concentration of volatile compounds in three black tea samples were evaluated by analysis of variance (ANOVA). All statistical analyses were conducted by Duncan's multiple comparison tests in SPSS 21.0 software (SPSS Inc., Chicage, USA, 2019). The statistically significant level was 5% (p < 0.05). The radar map was drawn using Origin 19 software (Originlab Corporation, Northampton, Massachusetts, USA).

Results and discussion

Identification and aroma intensity of aroma-active compounds in three world-famous black teas by GC–O

As a new sample preparation technique, SBSE had been successfully applied to the analysis of aroma profile in green tea [24]. GC–O was a method that combines GC

with the human nose, and had been widely used in the analysis of food flavor. It was generally believed that the aroma compounds perceived by GC-O were usually regarded as the key aroma compounds [31]. To explore the characteristic aroma compounds in these three high-aroma black teas, the volatile organic compounds were studied by SBSE-TDU/GC-O. As shown in Table 1, a total of 42 aroma compounds were perceived by GC-O, and their aroma intensity ranged from 0.3 to 7.9. These aroma substances were determined by RI, retention time of authentic standard, and aroma description. Among these aromaactive compounds, there were 11 aldehydes, 11 alcohols, 5 ketones, 4 terpenes, 2 esters, 2 lactones, 2 pyrroles, 1 oxide, 1 acid, 1 indole, and 2 unknown compounds. Aldehydes and alcohols were the predominant aroma compounds in these black teas. Aldehydes were usually considered to impart "green, grassy, fatty, and citrus" notes, while alcohols generally presented "green, sweet, and alcoholic" notes. According to the results of GC-O, it could be seen that the floral, green, and sweet aromas had stronger intensities than other notes. Among these compounds, 2-methylbutanal (AI: 4.5-6.8), 2-methyl-2-butenal (AI: 4.3-6.2), cis-3-hexen-1-ol (AI: 4.2-6.7), (E, E)-2, 4-heptadienal (AI: 3.9-5.7), methyl salicylate (AI: 4.0-6.4), geraniol (AI: 0.5-7.3), and phenylethyl alcohol (AI: 5.8–7.9) presented the highest aroma intensities among volatile compounds. Therefore, these compounds might make significant contributions to the aroma and flavor of the three world-famous black teas.

In addition, a total of 39, 38, and 33 aroma compounds were perceived by GC–O in three tea specimens (DJL, KM and CL, respectively). Among the three world's famous black teas, two compounds were unidentified. They all imparted a floral, green odor, but which specific substances they were remained unknown. Among the three types of black teas, phenylethyl alcohol had the highest aroma intensity. According to the analysis result of GC–O, phenyl alcohol was described as imparting a "sweet, rose-like" odor, which might relate to the floral note of black tea.

Among the three world-famous black teas, Keemun black tea had the highest aroma intensity and a unique "Keemun odor", resembling flowers, fruits, and honey. In Keemun black tea, phenylethyl alcohol exhibited the strongest intensity (7.9), followed by geraniol (7.3), 2-methylbutanal (6.8), methyl salicylate (6.4), and phenylacetaldehyde (5.8).

In Darjeeling black tea, *cis*-3-hexen-1-ol demonstrated the highest intensity (6.7), followed by phenylethyl alcohol (6.3), (E, E)-2,4-heptadienal (5.7), methyl salicylate (5.4), and 2-methylbutanal (5.2). While in Ceylon black tea, 2-methyl-2-butenal exhibited the highest intensity (6.2), followed by phenylethyl alcohol (5.8), 2-acetylpyrrole (5.6), phenylacetaldehyde (5.2), and cinnamaldehyde (5.1). Acetophenone was only perceived in Ceylon black tea.

Ionone generally had a warm woody and violet-like floral odor. There were three isomers of ionone: α -ionone, β -ionone, and γ -ionone. Specifically, β -ionone was distributed in different teas. In this experiment, the odor of β -ionone was described as iris-like. The ionone in tea was produced by the oxidation of catechins and the degradation of carotenoids [32]. Among these world-famous black teas, Ceylon black tea had the strongest aroma intensity of β -ionone, while Darjeeling black tea had the weakest. In addition, dihydroactinidiolide was one of the characteristic aroma compounds in black tea, which was derived from β -ionone. β -Ionone could be oxidized to 5,6-epoxy- β -ionone, leading to the formation of saturated triol, which was then oxidized to dihydroactinidiolide by intramolecular cyclization [33].

Noticeably, 3-methylindole was described as elegant floral scent of jasmine in this experiment, which was widely present in black tea [34]. According to previous studies, 3-methylindole was also considered to be the characteristic aroma compound in Longjing tea. 3-Methylindole is also known as skatole, and its odor is concentration-dependent. At a high concentration, 3-methylindole would have a fecal, animal-like, unpleasant odor, while it would present a pleasant natural jasmine odor when diluted to a certain extent [27].

Although GC–O was widely applied in the aroma analysis of food, it also had some disadvantages: it relies mainly on the discriminant resolving power of the human nose, making the experimental results subjective. In addition, GC–O used air as the medium, which was different from its original water matrix. Therefore, other methods were required to further determine the characteristic aroma of black tea. To ensure the accuracy of the experimental results, OAV method was applied to further determine the key aroma compounds in these famous black teas in the world.

Quantitative analysis and OAVs of volatile compounds in three world-famous black teas

Subsequently, the aroma compounds of the three worldfamous black teas were identified by SBSE-TDU/GC–MS. As shown in Table 2, a total of 89 aroma compounds were identified in the three tea samples, including 27 aldehydes, 16 alcohols, 13 ketones, 12 esters, 9 olefins, 4 acids, 3 lactones, and 5 others (indole, 3-methylindole, *cis*-linaloloxide, 2-acetylpyrrole, and 2,5-dimethylpyrazine). Besides, a total of 78, 76, and 69 volatile compounds were identified in three black tea samples (DJL, KM, and CL, respectively). Related quantitative information, such as the standard curve and evaluation index (\mathbb{R}^2) of fitting regression of each aroma compounds, was summarized in the Supplementary Information. It could be seen from the quantitative results that *cis*linaloloxide (32.68–364.49 µg/kg), (E, E)-2,4-heptadienal

Table 1	Aroma-active compounds	s identified by GC-C) in three world-	famous black teas
---------	------------------------	----------------------	-------------------	-------------------

No	Compound	RI ^a		Aroma description ^c	Identification ^d	Aroma	intens	ity			
		HP-Innowax	DB-5			DJL	SD ^g	KM	SD	CL	SD
1	2-Methylbutanal	945	638	Coffee, nutty	RI, Std, AD	5.2b ^e	0.17	6.8a	0.32	4.5c	0.13
2	3-Methylbutanal	948	630	Fatty, chocolate	RI, Std, AD	4.3ab	0.56	5.1a	0.63	3.9b	0.19
3	Hexanal	1076	778	Green, fatty	RI, Std, AD	2.9b	0.14	4.5a	0.26	1.5c	0.20
4	2-Methyl-2-butenal	1100	717	Green, fruity	RI, Std, AD	4.3c	0.25	5.2b	0.18	6.2a	0.58
5	β-Myrcene	1149	968	Terpene, spicy	RI, Std, AD	4.1b	0.31	5.3a	0.72	1.9c	0.26
6	(+)-Limonene	1173	1006	Citrus, orange	RI, Std, AD	2.0a	0.25	2.2a	0.64	2.0a	0.08
7	2-Heptanone	1177	869	Fruity, spicy, sweet	RI, Std, AD	1.7a	0.47	1.8a	0.32	2.2a	0.26
8	Heptanal	1198	880	Green, fatty,	RI, Std, AD	0.3b	0.15	2.2a	0.11	0.3b	0.08
9	(Z)-3,7-Dimethylocta-1,3,6,-triene	1214	1023	Herbal	RI, Std, AD	2.7	0.55	4.0	0.41	-	
10	Pentanol	1269	746	Oil, sweet	RI, Std, AD	2.8	0.22	2.3	0.56	_	
11	6-Methylhept-5-en-2-one	1318	963	Citrus, green	RI, Std, AD	2.3b	0.19	3.6a	0.61	3.8a	0.16
12	1-Hexanol	1353	850	Sweet, green	RI, Std, AD	2	0.84	-		1.5	0.22
13	Unknown 1	1354	_b	Green, floral	AD	_f		4.3	0.72	_	
14	cis-3-Hexen-1-ol	1363	837	Green, grass, fresh	RI, Std, AD	6.7a	0.72	4.8b	0.36	4.2b	0.86
15	trans-2-Hexen-1-ol	1398	847	Green, herbal	RI, Std, AD	4.5	0.55	-		3	0.89
16	cis-Linaloloxide	1433	1050	Sweet, woody	RI, Std, AD	4.7a	0.59	5.2a	0.63	2.6b	0.22
17	1-Octene-3-ol	1450	983	Mushroom, earthy	RI, Std, AD	1.4	0.34	0.5	0.02	_	
18	Furfural	1457	809	Woody, almond	RI, Std, AD	2.6b	0.41	5.3a	0.30	2.7b	0.35
19	Acetic acid	1464	621	Sour, vinegar	RI, Std, AD	3.4b	0.27	4.1a	0.44	3.6ab	0.18
20	(E, E)-2,4-Heptadienal	1478	988	Green, vegetable	RI, Std, AD	5.7a	0.73	5.4a	0.47	3.9b	0.46
21	2-Acetylfuran	1496	886	Sweet, coffee	RI, Std, AD	_		3.5	0.49	4.6	0.42
22	Benzaldehyde	1507	942	Almond	RI, Std, AD	4.5a	0.51	4.4a	0.30	3.6b	0.23
23	Linalool	1532	1077	Green, floral	RI, Std, AD	4.7a	0.32	4.1a	0.53	2.5b	0.17
24	5-Methyl furfural	1564	939	Sweet, caramel	RI, Std, AD	4.0b	0.31	5.5a	0.39	4.1b	0.36
25	Unknown 2	1592	_	Sweet, ocimene	AD	3.5	0.74	5.6	0.67	_	
26	1-Ethyl-2-pyrrolecarboxaldehyde	1595	1025	Roasted, smoky	RI, Std, AD	0.5	0.04	1.2	0.26	_	
27	Phenylacetaldehyde	1631	1021	Green, sweet, floral	RI, Std, AD	4.9a	0.78	5.8a	0.42	5.2a	0.29
28	Acetophenone	1636	1021	Sweet, almond	RI, Std, AD	_		_		3.1	0.56
29	α-Terpineol	1688	1172	Terpene, woody	RI, Std, AD	4.0a	0.62	2.4b	0.15	4.0a	0.53
30	Methyl salicylate	1765	1170	Wintergreen mint	RI, Std, AD	5.4a	0.51	6.4a	0.66	4.0b	0.65
31	Geraniol	1833	1225	Sweet, floral, rose	RI, Std, AD	4.6b	0.38	7.3a	0.67	0.5c	0.03
32	Nerol	1840	1200	Sweet, citrus	RI, Std, AD	2.3c	0.41	5.5a	0.27	4.4b	0.24
33	Benzyl alcohol	1871	1012	Floral	RI, Std, AD	2.2	0.19	3.1	0.58	_	
34	Phenylethyl alcohol	1905	1090	Rose, sweet	RI, Std, AD	6.3b	0.25	7.9a	0.32	5.8b	0.63
35	β-Ionone	1939	1450	Floral, orris	RI, Std, AD	2.1c	0.22	2.7ab	0.37	3.3a	0.48
36	2-Acetylpyrrole	1971	1043	Nut, yeast	RI, Std, AD	4.1b	0.26	4.5b	0.54	5.6a	0.43
37	γ-Nonalactone	2001	1332	Coconut, sweet	RI, Std, AD	2.7	0.28	3.0	0.34	-	20
38	2-Formylpyrrole	2026	992	Sweet, coffee	RI, Std, AD	3.4	0.30	5.2	0.41	_	
39	Cinnamaldehyde	2026	1246	Sweet, spice	RI, Std, AD	3.2	0.47	_	0.11	5.1	0.32
40	Dihydroactinidiolide	2375	1502	Fruit, woody	RI, Std, AD	1.3b	0.19		0.14	2.0a	0.32
40 41	Coumarin	2483	1409	Sweet, hay, bean	RI, Std, AD	1.6b	0.12	2.9a	0.14	1.8b	0.09
r 1	3-Methylindole	2483	1360	Floral, jasmine	RI, Std, AD	1.00 1.1b	0.22	2.9a 1.6a	0.27	1.0b	0.09

^aRetention index of compounds on HP-Innowax and DB-5 columns

^bThe aroma compounds not identified on DB-5 column

^cThe description of aroma compounds by sensory panel

^dMethod of identification: RI: retention index; Std, confirmed by authentic standards; AD: aroma description

^eValues with different Roman letters (a–c) in the same row are significantly different according to the Duncan test (p < 0.05)

^fNot detected in samples

^gThe standard deviation of aroma intensity of compounds

(64.29–102.87 µg/kg), benzaldehyde (60.66–267.13) µg/kg), linalool (145.18–385.8 µg/kg), methyl salicylate (181.65–340.64 µg/kg), geraniol (12.49–640.62 µg/kg), and phenylethyl alcohol (67.88–202.43 µg/kg) showed higher concentrations than other aroma compounds. As reported in the literature, linalool was an important aroma compound in black teas, such as Turkish black tea (70–241 µg/kg), Indian black tea (77–626 µg/kg) and Chinese black tea (12.7–2764 µg/kg) [35, 36]. Besides, geraniol and phenylethyl alcohol were present in high concentrations in most black tea infusions [10].

The qualitative and quantitative analysis of black tea (Table 2) indicated that the types and concentrations of aroma compounds in different black tea samples were different. The aroma compounds in black tea infusion depended on the glycoside aroma precursor in the black tea leaves, and were also related to the activity of glucosidase. These factors were all related to the origin of the black tea, the soil, climate, and the type of tea tree [37]. Tea leaves grown at high altitudes were prone to possess a richer aroma, and their tea infusions also had better quality [34]. Both Ceylon black tea and Darjeeling black tea plantations were located at high altitudes, especially Darjeeling black tea plantations, with an average altitude of 2100 m, endowing it with the characteristics of abundant fragrance, fresh taste, and a golden color.

The contribution of aroma compounds was not only affected by their concentrations, but also by their threshold. Generally, aroma compounds with $OAV \ge 1$ were considered to make a significant contribution to the overall aroma [26]. The quantitative data pertaining to of the top three of high-aroma black teas in the world allowed the calculation of OAVs (Table 3). A total of 37 aroma compounds with $OAV \ge 1$ were determined therein. In particular, 3-methylbutanal (OAV: 24-82), linalool (OAV: 24-64), 3-methylnonane-2,4-dione (OAV: 18-28), geraniol (OAV: 2-97), β-ionone (OAV: 54-122), cisjasmone (OAV: 2-119), and (\pm) -dihydroactinidiolide (OAV: 54–122) had higher OAVs than other compounds. This indicated that these compounds might be the key aroma compounds of black tea. From Tables 2 and 3, the OAVs of low-concentration compounds were found to be not necessarily low. For instance, though the concentration of 3-methylnonane-2,4-dione among the three black tea samples was very low, its threshold was only about 0.00003 mg/kg, which resulted in its high OAV. In Darjeeling black tea, there were 27 aroma compounds with OAVs \geq 1, while there were 29 and 26 aroma compounds with $OAVs \ge 1$ in Keemun black tea and Ceylon black tea, respectively. In addition, the OAVs of aroma compounds showed significant differences in different tea infusions. For example, in Darjeeling black tea, linalool, β-ionone and 3-methylbutanal had the highest OAVs, cis-jasmone, geraniol and *cis*-4-heptenal had the highest OAVs in Keemun black tea, while β -ionone, *cis*-jasmone, and 3-methylbutanal had the highest OAVs in Ceylon black tea.

Furthermore, the sulfur compounds in black tea were detected by sulfur-specific FPD. FPD is a detector which designed for the detection of phosphorus and sulfurs specially. It has high selectivity and sensitivity, only responds to phosphorus and sulfurs, and other elements have no or little interference with it [29]. As shown in Table 4, a total of 9 sulfur compounds were identified (the detailed quantitative information was listed in the Supporting Information). All sulfur concentrations were present at trace levels, and these sulfur compounds were not detected in GC-MS and GC-O. Among them, dimethyl sulfide (3.639-7.948 µg/kg), 2-methylthiophene (0.0292-1.658 µg/kg), and methionol (0.120-1.640 µg/ kg) were present in higher concentrations than other sulfur compounds. Besides, the OAVs of dimethyl sulfide methionol, and 3-mercaptohexanol were greater than 1. Therefore, although these sulfur compounds in black tea samples were at trace levels, they were likely to be the key aroma compounds of black tea. Remarkably, the sulfur compounds in black tea were mainly produced by Strecker degradation [38], among which methionine played an important role in the formation of sulfur compounds. Methional was generally regarded as the precursor of methanethiol. Besides, methanethiol was the direct precursor of many low-threshold sulfur compounds [39]. Dimethyl sulfide has been found in black tea, green tea, and oolong tea, and presented a garlic-like aroma [40]. According to the literature, methylmethionine sulfonium salt was considered to be the precursor of dimethyl sulfide [41].

3-Mercapto-1-hexanol possesses characteristic aroma of passion fruit and grapefruit, which might contribute to the fruity note of black tea. Among the black tea samples, the concentration of 3-mercapto-1-hexanol ranged from $0.038 \ \mu g/kg$ in KM to $0.042 \ \mu g/kg$ in DJL. The different concentrations of 3-mercapto-1-hexanol may arise from the different tree varieties, climate, planting conditions and soil environments. Although trace amounts of 3-mercapto-1-hexanol were detected, the amounts were greater than its threshold. The results demonstrated that 3-mercapto-1-hexanol was the characteristic compounds in black tea. The identification of sulfur compounds not only enriched the aroma profile of black tea, but provided a way of evaluating the quality of black tea.

Aroma recombination and E-nose

Aroma recombination was an essential part of flavor analysis, and it had been proved to be an important method for verification of key aroma compounds [42]. The volatile

	2-Methylbutanal 3-Methylbutanal Pentanal (+)-Camphene Hexanal 5-4-Carene β-Myrcene β-Myrcene 1-Penten-3-ol (+)-Limonene 2-Heptanone Methyl hexanoate Heptanal	29, 41, <i>57</i> 41, 43, 44 29, 44, 58	HP-Innowax	DB-5		DIL	SD^{e}	KM	SD	CL	SD
0 - 0 = 4	ylbutanal ylbutanal al mphene u lyl-2-butenal rene ene monee anone hexanoate	29, 41, <i>57</i> 41, 43, 44 29, 44, 58 70, 03, 171									
0 - 0 = 4	ylbutanal al mphene ul yl-2-butenal rene rene an-3-ol nonene anone hexanoate	41, 43, 44 29, 44, 58 70, 03, 121	945	639	MS, RI, Std	77.46b ^c	2.56	88.09a	4.63	48.36c	3.17
0 - 0 = 4	ıl mphene ul yl-2-butenal rene ::n-3-ol monene anone hexanoate al	29, 44, 58 70, 03, 121	948	630	MS, RI, Std	40.48b	3.23	90.56a	8.73	26.92c	2.76
0 - 0 - 0	mphene ul yl-2-butenal rene xn-3-ol nonene anone hexanoate al	70 03 171	779	670	MS, RI, Std	12.92	1.34	16.62	10.85	I	
0 - 0 = 4	l yl-2-butenal rene cene sn-3-ol nonene anone hexanoate al	12, 77, 141	1071	930	MS, RI, Std	0.69c	0.05	0.90b	0.05	1.70a	0.09
0 - 0 - 4	yl-2-butenal rene ene m-3-ol nonene anone hexanoate al	41, 44, 56	1077	778	MS, RI, Std	58.52b	3.27	92.57a	8.84	32.23c	3.12
0 - 0 = 4	rene ene m-3-ol nonene anone hexanoate al	29, 55, 84	1101	718	MS, RI, Std	2.70c	0.09	4.78b	0.61	9.95a	0.46
0 - 0 - 7 + 1	ene x1-3-ol nonene anone hexanoate al	93, 121, 136	1142	994	MS, RI	1.55	0.12	6.37	0.35	I	
	:n-3-ol nonene anone hexanoate al	41, 69, 93	1149	968	MS, RI, Std	82.43b	5.63	180.46a	12.51	60.59c	4.25
	nonene anone hexanoate al	29, 57	1165	656	MS, RI, Std	15.84a	1.12	10.87b	0.74	4.80c	0.20
	anone hexanoate al	67, 68, 93	1173	1006	MS, RI, Std	41.67ab	3.90	48.73a	3.92	37.41b	2.63
	hexanoate al	43, 58	1177	869	MS, RI, Std	8.18c	0.38	10.20b	1.03	15.30a	1.16
	al	43, 74, 87	1192	006	MS, RI, Std	2.13	0.45	2.83	0.22	I	
		41, 44, 70	1198	880	MS, RI, Std	5.34b	0.37	8.71a	0.65	5.22b	0.44
	(3E)-3,7-Dimethyl-1,3,6-octatriene	41, 79, 93	1199	1011	MS, RI, Std	37.57	2.66	57.97	4.37	I	
15 trans-2-1	trans-2-Hexenal	39, 41, 42	1212	833	MS, RI, Std	55.01b	3.27	79.20a	6.30	20.25c	1.93
16 (Z)-3,7-l	(Z)-3,7-Dimethylocta-1,3,6,-triene	41, 79, 93	1214	1023	MS, RI, Std	15.02b	1.06	48.14a	3.84	3.22c	0.19
	Ethyl hexanoate	29, 43, 88	1245	978	MS, RI, Std	p_		I		5.83	0.28
18 cis-4-Heptenal	leptenal	29, 41, 68	1264	878	MS, RI, Std	0.62	0.02	2.35	0.13	I	
19 2-Octanone	none	41, 43, 58	1264	973	MS, RI, Std	I		I		38.00	2.41
20 Pentanol	lc	41, 42, 55	1269	746	MS, RI, Std	16.06	1.14	7.11	0.76	I	
21 p-Cymene	ene	91, 119, 134	1288	1000	MS, RI, Std	10.56b	0.98	18.25a	1.92	18.38a	1.27
22 2,5-Dim	2,5-Dimethylpyrazine	39, 42, 108	1311	891	MS, RI, Std	1.60	0.02	4.39	0.20	Ι	
	6-Methylhept-5-en-2-one	41, 43, 69	1318	963	MS, RI, Std	28.14a	2.12	30.02a	2.61	31.19a	2.79
	(4E,6Z)-2,6-Dimethylocta-2,4,6-triene	105, 121, 136	1330	1100	MS, RI, Std	2.27c	0.16	20.28a	1.98	10.72b	1.06
25 1-Hexanol	nol	41, 43, 56	1353	850	MS, RI, Std	28.77	1.25	7.67	0.66	I	
	cis-3-Hexen-1-ol	39, 41, 67	1382	837	MS, RI, Std	75.07a	5.59	39.28b	3.41	23.07c	1.74
27 2,4-Hexadienal	xadienal	39, 41, 81	1393	888	MS, RI, Std	6.83b	0.41	8.46a	0.52	6.94b	0.45
28 trans-2-]	trans-2-Hexen-1-ol	41, 57	1410	847	MS, RI, Std	24.82a	2.23	5.96c	0.44	21.34b	1.98
29 (2E)-2-Octenal	Octenal	29, 41, 55	1418	1035	MS, RI, Std	5.00b	0.37	5.40b	0.28	34.46a	2.64
30 cis-Lina	cis-Linaloloxide	59, 43	1438	1050	MS, RI, Std	223.91b	20.64	364.49a	32.68	110.01c	9.15
31 1-Octene-3-ol	ne-3-ol	57	1450	983	MS, RI, Std	6.32a	0.34	4.34b	0.31	2.16c	0.11
32 Furfural	ų	39, 95, 96	1457	809	MS, RI, Std	7.00b	0.81	18.05a	0.75	4.71c	0.23
33 Acetic acid	acid	43, 45, 60	1464	621	MS, RI, Std	28.24b	2.12	47.75a	2.91	12.88c	1.09
34 (E, E)-2	(E, E)-2,4-Heptadienal	81, 110	1478	988	MS, RI, Std	102.87a	9.54	92.32a	8.06	64.29b	4.44

Table 2 The concentration of volatile compounds detected by GC-MS in three world-famous black teas

Compounds Quantitative ions 2-Acetylfuran $39, 5, 110$ 3.5 -Octadiene-2-one $3, 195, 110$ 3.5 -Octadiene-2-one $43, 81, 95$ Decanal $77, 105, 106$ Linalool $77, 105, 106$ Linaly acctate $43, 81, 95$ Linaly acctate $43, 81, 95$ S-Methyl furtural $77, 105, 106$ Linalyl acctate $43, 81, 95$ Cyclocitral $41, 123$ Hortienol $93, 105, 119$ $-$ Ethyl furtural $93, 105, 119$ $-$ Ethyl forcate $43, 81, 01$ 6 -Cyclocitral $10, 9, 110$ $(-)$ -Cedtene $93, 105, 119$ 1 -Ethyl furcate $93, 105, 119$ 1 -Ethyl furcate $93, 105, 119$ 1 -Ethyl furcate $91, 41, 123$ 1 -Ovelocitral 109 1 -Provide $94, 123$ 9 -Cyclocitral $10, 140$ 1 -Ethyl furcate $91, 41, 123$ 1 -Ethyl furcate $51, 71, 105$ 1 -Ethyl furcate $51, 71, 105$ <												
2-Acetylfuran $3, 5$ -Octadiene-2-one $3, 5$ -Octadiene-2-one $3, 5$ -Octadiene-2-one $3, 5$ -Octadiene-2-one $4, 3, 57$ Benzaldehyde $77, 105, 106$ Linalol $77, 105, 106$ Linalol $77, 105, 106$ Linalol $55, 11, 93$ Linalol $55, 11, 93$ Linalyl acetate $43, 80, 93$ (E, E)- $3, 5$ -Octadien-2-one $43, 81, 95$ 5 -Methyl furfural $55, 119$ $1-Ethyl-2-pyrrolecarboxaldehyde 93, 94, 123 9, 2Cyclocitral 99, 41, 123 9, 10, 110 (-)-Cedrene 93, 94, 123 9, 111 Phenylacctaldehyde 93, 94, 123 9, 111 Phenylacctaldehyde 93, 94, 123 9, 111 Phenylacctaldehyde 93, 94, 123 1-Costrene 31, 41, 123 94, 123 1-Cyclocitral 10, 100 77, 105, 122 1-Cyclocitral 10, 100 77, 105, 122 1-Cyclocitral 10, 100 77, 105, 122 1-Cyclocitral 20, 12, 120 111 Prenylacctaldehyde 88, 101 $	No	Compounds	Quantitative ions	RI ^a		Identification ^b	Concentral	Concentration (µg/kg)				
2-Acetyffuran $3, 5$. Octadiene-2-one $3, 5$. Soctadiene-2-one $4, 43, 57$ Benzaldehyde $77, 105, 106$ Linabol $77, 105, 106$ Linabol $55, 71, 93$ Linabol $55, 71, 93$ Linabol $55, 71, 93$ Linabol $55, 71, 93$ Linabyl acetate $55, 109, 110$ Cyclocitral $93, 105, 119$ Hyl furtural $55, 119, 123$ Bernyl-2-pyrrolecarboxaldehyde $93, 94, 123$ β -Cyclocitral $10, 9$ Hyl furoate $91, 21, 120$ Ethyl furoate $71, 105, 122$ Ethyl bernzoate $88, 101$ Prenylacetaldehyde $91, 22, 120$ Ethyl bernzoate $51, 77, 105$ Ethyl bernzoate $51, 71, 122$ Salicylaldehyde $91, 92, 67, 93$ Citral $72, 102, 122$ Salicylaldehyde $91, 43, 69$ Citral $72, 120, 152$ Salicylaldehyde				HP-Innowax	DB-5		DJL	SD^{e}	KM	SD	CL	SD
3.5 -Octadiene-2-one $43, 81, 95$ 3.5 -Octadiene-2-one $41, 43, 57$ $Benzaldehyde77, 105, 1061.inalol55, 71, 931.inalol55, 71, 931.inalol55, 71, 931.inalol53, 109, 110(-)-Cettene93, 105, 1191-Ethyl-2-pyrrolecarboxaldehyde93, 105, 119(-)-Cettene93, 105, 1191-Ethyl-2-pyrrolecarboxaldehyde39, 41, 123\beta-Cyclocitral10, 100(-)-Cettene93, 101, 100(-)-Cettene93, 101, 100(-)-Cettene93, 101, 100(-)-Cettene93, 101, 100(-)-Cettral11, 123\beta-Cyclocitral10, 100(-)-Cettene93, 101, 123\beta-Cyclocitral10, 1001-Ethyl-2-pyrrolecarboxaldehyde39, 41, 123\beta-Cyclocitral1001-Ethyl-2-pyrrolecarboxaldehyde39, 41, 123\beta-Cyclocitral1001-Ethyl-2-pyrrolecarboxaldehyde39, 41, 123\beta-Cyclocitral1001-Cyclocitral111, 1231-Cyclocitral111, 1231-Cyclocitral11, 1231-Cyclocitral11, 1231-Cyclocitral11, 1231-Cyclocitral11, 1231-Cyclocitral11, 1231-Cyclocitral11, 1231-Cyclocitral11, 1231-Cyclocitral11, 1062-Cyclocitral11, 1062-Cyclocitr$	35	2-Acetylfuran	39, 95, 110	1496	886	MS, RI, Std	0.88c	0.04	2.80b	0.03	3.45a	0.22
Decanal $41, 43, 57$ Benzaldehyde $55, 71, 93$ Linalool $55, 71, 93$ Linalyl acetate $55, 71, 93$ Linalyl acetate $43, 80, 93$ $(E, E)-3,5-Octadien-2-one53, 109, 110(-)-Cedrene93, 105, 119(-)-Cedrene93, 105, 119(-)-Cedrene93, 105, 119(-)-Cedrene93, 105, 119(-)-Cedrene93, 105, 119(-)-Cedrene93, 105, 110(-)-Cedrene93, 105, 110(-)-Cedrene93, 101, 103(-)-Cedrene93, 94, 123\beta-Cyclocitral91, 91, 233\beta-Cyclocitral109(-)-Cedrene91, 91, 103(-)-Cedrene91, 92, 120(-)-Cedrene91, 92, 120(-)-Cedrene91, 92, 120(-)-Cedrene91, 92, 120(-)-Certal109(-)-Cedrene91, 92, 120(-)-Citral91, 92, 120(-)-Citral91, 93, 95, 67, 93(-)-Citral91, 93, 95, 67, 93(-)-Citral91, 93, 95, 103(-)-Citral91, 108(-)-Methyl salicylate91, 108(-)-Methyl acetate91, 108(-)-Citral91, 108(-)-Methyl acetate91, 108(-)-Methyl acetate91, 108(-)-Methyl acetate91, 108(-)-Methyl acetate91, 108(-)-Methyl acetate91, 108(-)-Methyl acetate91, 108<$	36	3,5-Octadiene-2-one	43, 81, 95	1502	1046	MS, RI, Std	33.00b	2.41	38.90a	3.64	6.41c	0.53
Berzaldehyde $77, 105, 106$ Linalool $55, 71, 93$ Linalyl acetate $55, 71, 93$ Linalyl acetate $53, 109, 110$ $(-)$ -Cedrene $93, 105, 119$ $(-)$ -Cedrene $93, 105, 110$ $(-)$ -Cedrene $93, 105, 123$ $(-)$ -Cedrene $91, 92, 120$ $(-)$ -Cedrene $91, 21, 123$ $(-)$ -Cedrene $91, 21, 123$ $(-)$ -Citral $110, 105, 122$ $(-)$ -Citral $11, 108, 124$ <	37	Decanal	41, 43, 57	1503	1193	MS, RI, Std	15.42a	1.03	0.70c	0.01	3.27b	0.17
Linalool55, 71, 93Linalyl acetate53, 109, 110($F. E. J3.5$ -Octadien-2-one43, 81, 955-Methyl furfural53, 109, 110($f. E. E. J3.5$ -Octadien-2-one53, 109, 110($f. E. E. J3.5$ -Octadien-2-one93, 105, 119($f. E. E. P3.5$ -Octadien-2-one93, 105, 119($f. E. E. P3.5$ -Octadien-2-one93, 105, 119($f. E. E. P3.5$ -Octadien-2-one93, 105, 110($f. E. E. P3.5$ -Octadien-2-one93, 105, 110($f. E. E. P3.5$ -Octadien-493, 94, 123 $P - Cyclocitral109Ferpinen-4-ol71, 93, 111Phenylacetadehyde91, 92, 120Ethyl benzoate88, 101Actophenone71, 71, 105Ethyl benzoate88, 101Actophenone51, 77, 105Ethyl benzoate121, 122Salicyladehyde51, 77, 105Ethyl benzoate91, 92, 120Salicyladehyde90, 91, 1083.Methylnonane-2,4-dione91, 1083.Methylnonane-2,4-dione92, 120, 152Geranyl actate92, 120, 152Geranyl actate91, 108Henylenylpropanoate92, 120, 108Henylenyl alcohol65, 91, 92Soctalactone42, 71, 99\gamma-Octanolactone29, 85$	38	Benzaldehyde	77, 105, 106	1507	942	MS, RI, Std	126.27b	10.56	267.13a	24.63	60.66c	5.09
Linalyl acetate43, 80, 93(E. E.) -3.5-Octaditen-2-one53, 109, 110(-)-Cedrene53, 105, 119(-)-Cedrene39, 94, 123 β -Cyclocitral39, 94, 123 β -Cyclocitral109 β -Cyclocitral101 β -Cyclocitral109 β -Cyclocitral101 β -Cyclocitral121, 125 β -Dythenouce24, 69 β -Methyl saticylate21, 105 β -Methylnonane-2,4-dione21, 105 β -Methylnonane-2,4-dione21, 106 β -Methylnonane-2,4-dione21,	39	Linalool	55, 71, 93	1532	1077	MS, RI, Std	385.8a	25.67	246.37b	21.07	145.18c	13.35
(E. E) -3.5-Octadien-2-one43, 81, 955-Methyl furfural53, 109, 110(-)-Cedrene93, 105, 1191-Ethyl-2-pyrrolecarboxaldehyde39, 94, 123 β -Cyclocitral111Horrienol31, 41, 123 β -Cyclocitral109Ethyl furoate91, 92, 120Ethyl decanoate91, 92, 120Ethyl decanoate91, 92, 120Ethyl benzoate91, 92, 120Ethyl benzoate91, 92, 120Ethyl benzoate91, 92, 120Ethyl benzoate91, 92, 120Salicylaldehyde91, 92, 120Salicylaldehyde91, 92, 120Salicylaldehyde91, 92, 120C/Ortral41, 69C/Ortral41, 69C/Ortral90, 91, 108Benzyl acetate90, 91, 1083-Methylnonane-2,4-dione92, 120, 152Geranyl acetate92, 120, 152Geranyl acetate92, 120, 152Geranyl acetate91, 1083-Methyl salicylate92, 120, 152Geranyl acetate91, 1083-Methyl salicylate92, 121, 136Geranyl acetate91, 1083-Methyl alcohol41, 69, 93Ethyl 3-phenylpropanoate92, 120, 152Geranyl acetate91, 1083-Methyl alcohol91, 1083-Methyl alcohol91, 1093-Methyl alcohol91, 1093-Methyl alcohol91, 1093-Methyl alcohol91, 1093-Methyl alcohol91, 1093-Methyl alcohol91, 1093	40	Linalyl acetate	43, 80, 93	1547	1220	MS, RI, Std	22.25	1.84	I		I	
5-Methyl furfural 53, 109, 110 (-)-Cedrene 93, 105, 119 1-Ethyl-2-pyrrolecarboxaldehyde 94, 123 β -Cyclocitral 31, 41, 123 Hotrienol 95, 112, 140 Terpinen 4-ol 91, 92, 120 Ethyl accanoate 95, 112, 140 Terpinen 4-ol 91, 92, 120 Ethyl benzoate 88, 101 Acetophenone 71, 105, 122 Ethyl benzoate 88, 101 Acetophenone 51, 77, 105 Ethyl benzoate 121, 122 Salicylaldehyde 90, 91, 108 Zherpineol 11, 122 Actophenone 51, 77, 105 Ethyl benzoate 121, 122 Salicylaldehyde 90, 91, 108 Zhortylonane-2,4-dione 90, 91, 108 Benzyl acctate 90, 91, 108 J-Methyl salicylate 92, 120, 152 Geranyl acctate 92, 120, 152 Geranyl acctate 92, 100 Benzyl acchol 91, 108 Benzyl acchol 91, 108 Hotol 91, 108 Geranyl acctate 92, 120, 152	41	(E, E)-3,5-Octadien-2-one	43, 81, 95	1556	1069	MS, RI, Std	15.69	1.74	33.26	2.74	I	
(-)-Cedrene93, 105, 1191-Ethyl-2-pyrrolecarboxaldehyde39, 94, 123 β -Cyclocitral31, 41, 123Hotrienol31, 41, 123 β -Cyclocitral09, 111Hotrienol95, 112, 140Terpinen 4-ol91, 92, 120Ethyl furoate91, 92, 120Ethyl benzoate88, 101Acetophenone51, 77, 105Ethyl benzoate121, 122Salicylaldehyde51, 77, 105Ethyl benzoate121, 122Salicylaldehyde59, 67, 93(Z)-Citral41, 69, 84Benzyl acetate90, 91, 1083-Methylnonane-2,4-dione92, 120, 152Geranyl acetate92, 120, 152Geranyl acetate92, 120, 152Geranyl acetate92, 120, 152Geranyl acetate91, 108Horol41, 69, 93Ethyl 3-phenylpropanoate91, 108Benzyl alcohol81, 109, 124Phenylethyl alcohol65, 91, 92ô-Ottalactone20, 85 γ -Octanolactone20, 85	42	5-Methyl furfural	53, 109, 110	1564	939	MS, RI, Std	1.81c	0.12	6.04a	0.45	3.54b	0.20
1-Ethyl-2-pyrrolecarboxaldehyde39, 94, 123 β -Cyclocitral1, 1, 123Hotrienol31, 41, 123Hotrienol95, 112, 140Terpinen-4-ol91, 92, 120Terpinen-4-ol91, 92, 120Tehyl decanoate88, 101Acetophenone51, 77, 105, 122Salicylaldehyde53, 67, 93 α -Terpineol77, 105, 122Salicylaldehyde121, 122Salicylaldehyde90, 91, 108 α -Terpineol77, 105, 122Salicylaldehyde121, 122 α -Terpineol59, 67, 93 α -Terpineol90, 91, 108 α -Terpineol90, 91, 108 α -Terpineol90, 91, 108 α -Terpineol90, 91, 108 α -Tornone91, 108 β -Methylnonane-2,4-dione92, 120, 152 α -Iernone92, 120, 152 α -Ionone91, 108 α -Ionone91, 108 α -Ionone91, 104 β -Denylpropanoate91, 104 β -Denylpropanoate91, 104 β -Octalactone81, 109, 124 γ -Octanolactone20, 85	43	(-)-Cedrene	93, 105, 119	1582	1396	MS, RI, Std	I		I		29.03	2.27
β -Cyclocitral31, 41, 123Horrienol109Ethyl furoate95, 112, 140Terpinen-4-ol91, 92, 120Terpinen-4-ol71, 93, 111Phenylacetaldehyde88, 101Acetophenone51, 77, 105Ethyl benzoate88, 101Acetophenone51, 77, 105Ethyl benzoate51, 77, 105Ethyl benzoate121, 122Salicylaldehyde59, 67, 93(Z)-Citral41, 69Citral90, 91, 1083-Methylnonane-2,4-dione91, 1083-Methyl acetate92, 120, 152Geranyl acetate92, 121, 136Geraniol41, 69, 93Ethyl 3-phenylpropanoate91, 104Benzyl alcohol81, 109, 124Phenylethyl alcohol65, 91, 92 δ -Ottalactone29, 85 γ -Octanolactone29, 85	44	1-Ethyl-2-pyrrolecarboxaldehyde	39, 94, 123	1595	1025	MS, RI, Std	14.56b	1.44	42.79a	3.01	5.92c	0.32
Hotrienol109Ethyl furoate95, 112, 140Terpinen 4-ol91, 92, 120Terpinen 4-ol91, 92, 120Ethyl benzoate88, 101Acetophenone51, 77, 105Ethyl benzoate51, 77, 105Ethyl benzoate51, 77, 105Ethyl benzoate51, 77, 105Ethyl benzoate51, 77, 105Salicylaldehyde51, 77, 105Citral77, 105, 122Salicylaldehyde77, 105, 122Salicylaldehyde77, 105, 122Gerand41, 69, 84Benzyl acetate90, 91, 1083-Methylnonane-2,4-dione92, 120, 152Geranyl acetate92, 120, 152Geraniol41, 68, 69Nerol92, 121, 136Geraniol41, 69, 93Ethyl 3-phenylpropanoate91, 104Benzyl alcohol81, 109, 124Phenylethyl alcohol65, 91, 92 δ -Ottalactone29, 85 γ -Octanolactone29, 85	45	β-Cyclocitral	31, 41, 123	1598	1193	MS, RI, Std	15.42	1.06	I		0.48	0.02
Ethyl furoate $95, 112, 140$ Terpinen-4-ol $71, 93, 111$ Phenylacetaldehyde $91, 92, 120$ Ethyl decanoate $88, 101$ Acetophenone $51, 77, 105$ Ethyl benzoate $51, 77, 105, 122$ Salicylaldehyde $77, 105, 122$ Salicylaldehyde $90, 91, 108$ Salicylate $90, 91, 108$ Benzyl acetate $90, 91, 108$ 3-Methyl salicylate $92, 120, 152$ Geranyl acetate $92, 120, 152$ Geranyl acetate $92, 120, 152$ Geranyl acetate $92, 121, 136$ Geranyl acetate $92, 120, 152$ Geranyl acetate $92, 121, 136$ Geranyl acetate $91, 08, 09$ Nerol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $91, 107, 108$ Phoylet	46	Hotrienol	109	1602	1080	MS, RI, Std	40.48	3.10	30.87	2.22	I	
Terpinen-4-ol71, 93, 111Phenylacetaldehyde91, 92, 120Ethyl decanoate88, 101Acetophenone51, 77, 105Ethyl benzoate51, 77, 105, 122Salicylaldehyde77, 105, 122Salicylaldehyde121, 122Salicylaldehyde121, 122Salicylaldehyde90, 91, 108Salicylate90, 91, 108Benzyl acetate90, 91, 1083-Methylnonane-2,4-dione92, 120, 152Geranyl acetate92, 120, 152Geranyl acetate92, 121, 136Geranyl acetate91, 104Benzyl alcohol81, 109, 124Phenylethyl alcohol65, 91, 92S-Octalactone29, 85 γ -Octanolactone29, 85	47	Ethyl furoate	95, 112, 140	1614	1030	MS, RI, Std	I		I		42.75	3.55
Phenylacetaldehyde $91, 92, 120$ Ethyl decanoate $88, 101$ Acetophenone $51, 77, 105$ Ethyl benzoate $51, 77, 105, 122$ Salicylaldehyde $77, 105, 122$ Salicylaldehyde $121, 122$ Salicylaldehyde $121, 122$ Salicylaldehyde $91, 105, 122$ Salicylaldehyde $91, 108$ Benzyl acetate $90, 91, 108$ 3 -Methylnonane-2,4-dione $91, 108$ 171 Methyl salicylate $91, 108$ 171 Methyl salicylate $92, 121, 136$ Geranyl acetate $91, 108$ Nerol $91, 108$ Benzyl alcohol $81, 109, 124$ Phenylethyl alcohol $92, 81, 02$ $g-Octalactone29, 85$	48	Terpinen-4-ol	71, 93, 111	1616	1158	MS, RI, Std	6.97a	0.32	7.68a	0.66	1.10b	0.06
Ethyl decanoate88, 101Acetophenone51, 77, 105Ethyl benzoate51, 77, 105Ethyl benzoate77, 105, 122Salicylaldehyde121, 122 α -Terpineol59, 67, 93 α -Terpineol59, 67, 93 α -Terpineol90, 91, 108 α -Methyl noane-2,4-dione90, 91, 108 β -Methyl salicylate90, 91, 108 β -Methyl salicylate92, 120, 152 α -Ionone92, 121, 136 α -Ionone92, 121, 136 α -Ionone92, 121, 136 α -Ionone92, 121, 136 α -Ionone91, 104 β -Ionone92, 121, 136 α -Ionone91, 104 β -Ionole81, 109, 124 β -Ionole91, 104 β -Ionole91,	49	Phenylacetaldehyde	91, 92, 120	1631	1021	MS, RI, Std	44.43b	2.90	113.11a	10.27	49.97b	3.20
Acetophenone $51, 77, 105$ Ethyl benzoate $77, 105, 122$ Salicylaldehyde $121, 122$ salicylaldehyde $121, 122$ α -Terpineol $59, 67, 93$ (Z) -Citral $90, 91, 108$ α -Terpineol $41, 69$ (Z) -Citral $90, 91, 108$ α -Terpineol $41, 69, 84$ β -Methyl none-2,4-dione $90, 91, 108$ β -Methyl salicylate $92, 120, 152$ β -Methyl salicylate $92, 120, 152$ β -Methyl salicylate $92, 121, 136$ β -Ionone $41, 69, 93$ β -Ionone $92, 121, 136$ β -Ionone $91, 104$ β -Ionole $91, 92$ <td>50</td> <td>Ethyl decanoate</td> <td>88, 101</td> <td>1631</td> <td>1368</td> <td>MS, RI, Std</td> <td>I</td> <td></td> <td>I</td> <td></td> <td>79.37</td> <td>60.9</td>	50	Ethyl decanoate	88, 101	1631	1368	MS, RI, Std	I		I		79.37	60.9
Ethyl benzoate $77, 105, 122$ Salicylaldehyde121, 122salicylaldehyde59, 67, 93 α -Terpineol59, 67, 93 α -Terpineol41, 69 α -Terpineol41, 69 α -Torla41, 69, 84Benzyl acetate90, 91, 108 3 -Methylnonane-2,4-dione90, 91, 108 3 -Methyl salicylate92, 120, 152Geranyl acetate92, 121, 136 α -Ionone92, 121, 136Geraniol41, 69, 93Nerol91, 104Benzyl alcohol91, 104Benzyl alcohol81, 109, 124Phenylethyl alcohol65, 91, 92 γ -Octalactone29, 85	51	Acetophenone	51, 77, 105	1637	1050	MS, RI, Std	I		I		67.77	5.66
Salicylaldehyde $121, 122$ α -Terpineol $59, 67, 93$ α -Terpineol $59, 67, 93$ (Z) -Citral $41, 69$ (Z) -Citral $41, 69$ (Z) -Citral $90, 91, 108$ Benzyl acetate $90, 91, 108$ 3 -Methylnonane-2,4-dione $92, 120, 152$ Geranyl acetate $92, 120, 152$ Geranyl acetate $92, 120, 152$ Geranyl acetate $92, 121, 136$ Geraniol $41, 69, 93$ Nerol $92, 121, 136$ Nerol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $81, 109, 124$ Phenylethyl alcohol $65, 91, 92$ δ -Octalactone $42, 71, 99$ γ -Octanolactone $29, 85$	52	Ethyl benzoate	77, 105, 122	1647	1145	MS, RI, Std	2.65c	0.14	11.57b	0.89	27.89a	2.74
α -Terpineol59, 67, 93(Z)-Citral41, 69(Z)-Citral41, 69, 84Benzyl acetate90, 91, 1083-Methylnonane-2,4-dione90, 91, 1083-Methyl salicylate92, 120, 152Geranyl acetate92, 121, 136 α -lonone92, 121, 136Geraniol41, 68, 69Nerol92, 121, 136Geraniol41, 68, 69Nerol91, 104Benzyl alcohol91, 104Benzyl alcohol81, 109, 124Phenylethyl alcohol65, 91, 92 γ -Octalactone29, 85	53	Salicylaldehyde	121, 122	1682	1027	MS, RI, Std	I		I		55.31	5.02
(Z)-Citral $41,69$ Citral $41,69,84$ Benzyl acetate $90,91,108$ 3 -Methylnonane-2,4-dione $90,91,108$ 3 -Methyl salicylate $92,120,152$ Geranyl acetate $92,120,152$ Geranyl acetate $92,121,136$ α -Ionone $92,121,136$ Geraniol $41,63,69$ Nerol $92,121,136$ Seroniol $41,69,93$ Benzyl alcohol $91,104$ Benzyl alcohol $91,104$ Benzyl alcohol $81,109,124$ Phenylethyl alcohol $65,91,92$ δ -Octalactone $29,85$	54	α-Terpineol	59, 67, 93	1688	1172	MS, RI, Std	24.69a	1.71	15.08c	1.21	21.21b	1.25
Citral $41, 69, 84$ Benzyl acetate $90, 91, 108$ 3 -Methylnonane-2,4-dione $90, 91, 108$ 3 -Methyl salicylate $92, 120, 152$ Geranyl acetate $92, 120, 152$ Geranyl acetate $92, 121, 136$ α -Ionone $92, 121, 136$ Geraniol $41, 68, 69$ Nerol $92, 121, 136$ Geraniol $41, 69, 93$ Benzyl alcohol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $81, 109, 124$ Phenylethyl alcohol $65, 91, 92$ γ -Octalactone $29, 85$	55	(Z)-Citral	41, 69	1688	1209	MS, RI, Std	6.33b	0.52	18.43a	1.63	5.27b	0.43
Benzyl acetate 90, 91, 108 3-Methylnonane-2,4-dione 171 Methyl salicylate 92, 120, 152 Geranyl acetate 92, 120, 152 Geranyl acetate 92, 121, 136 Graniol 92, 121, 136 Nerol 92, 121, 136 Nerol 92, 121, 136 Kerniol 91, 168, 69 Nerol 41, 69, 93 Ethyl 3-phenylpropanoate 91, 104 Benzyl alcohol 79, 107, 108 Guaiacol 81, 109, 124 Phenylethyl alcohol 65, 91, 92 ô-Octalactone 29, 85	56	Citral	41, 69, 84	1712	1240	MS, RI, Std	9.78b	0.53	31.15a	3.06	28.31a	2.08
3 -Methylnonane-2,4-dione 171 Methyl salicylate $92, 120, 152$ Geranyl acetate $92, 121, 136$ Geraniol $92, 121, 136$ Geraniol $41, 68, 69$ Nerol $41, 69, 93$ Ethyl 3-phenylpropanoate $91, 104$ Benzyl alcohol $79, 107, 108$ Guaiacol $81, 109, 124$ Phenylethyl alcohol $42, 71, 99$ γ -Octanolactone $29, 85$	57	Benzyl acetate	90, 91, 108	1714	1136	MS, RI, Std	3.53c	0.21	6.02b	0.44	22.63a	1.37
Methyl salicylate 9_2 , 120, 152Geranyl acetate $41, 43, 69$ α -lonone $9_2, 121, 136$ Geraniol $41, 68, 69$ Nerol $41, 69, 93$ Nerol $92, 121, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $79, 107, 108$ Guaiacol $65, 91, 92$ δ -Octalactone $42, 71, 99$ γ -Octanolactone $29, 85$	58	3-Methylnonane-2,4-dione	171	1719	1220	MS, RI, Std	0.85a	0.04	0.53c	0.03	0.64b	0.03
Geranyl acetate $41, 43, 69$ α -Ionone $92, 121, 136$ α -Ionone $92, 121, 136$ Geraniol $41, 68, 69$ Nerol $41, 69, 93$ Nerol $91, 104$ Benzyl alcohol $91, 104$ Benzyl alcohol $79, 107, 108$ Guaiacol $81, 109, 124$ Phenylethyl alcohol $65, 91, 92$ δ -Octalactone $42, 71, 99$ γ -Octanolactone $29, 85$	59	Methyl salicylate	92, 120, 152	1765	1170	MS, RI, Std	237.76b	20.81	340.64a	32.71	181.65c	17.05
α-Ionone 92, 121, 136 Geraniol 41, 68, 69 Nerol 41, 69, 93 Nerol 91, 104 Ethyl 3-phenylpropanoate 91, 104 Benzyl alcohol 79, 107, 108 Guaiacol 81, 109, 124 Phenylethyl alcohol 65, 91, 92 δ-Octalactone 42, 71, 99 γ-Octanolactone 29, 85	60	Geranyl acetate	41, 43, 69	1766	1348	MS, RI, Std	10.35	1.14	12.65	1.23	I	
Geraniol 41, 68, 69 Nerol 41, 69, 93 Nerol 91, 104 Ethyl 3-phenylpropanoate 91, 104 Banzyl alcohol 79, 107, 108 Guaiacol 81, 109, 124 Phenylethyl alcohol 65, 91, 92 ô-Octalactone 42, 71, 99 γ -Octanolactone 29, 85	61	α-Ionone	92, 121, 136	1822	1393	MS, RI, Std	4.89b	0.28	5.26b	0.37	16.95a	1.13
Nerol 41, 69, 93 Ethyl 3-phenylpropanoate 91, 104 Benzyl alcohol 79, 107, 108 Guaiacol 81, 109, 124 Phenylethyl alcohol 65, 91, 92 ô-Octalactone 42, 71, 99 γ -Octanolactone 29, 85	62	Geraniol	41, 68, 69	1833	1225	MS, RI, Std	129.69b	10.73	640.62a	55.49	12.49c	1.44
Ethyl 3-phenylpropanoate 91, 104 Benzyl alcohol 79, 107, 108 Guaiacol 81, 109, 124 Phenylethyl alcohol 65, 91, 92 ô-Octalactone 42, 71, 99 y-Octanolactone 29, 85	63	Nerol	41, 69, 93	1840	1200	MS, RI, Std	8.74c	0.41	19.94a	1.32	15.78b	1.37
Benzyl alcohol 79, 107, 108 Guaiacol 81, 109, 124 Phenylethyl alcohol 65, 91, 92 ô-Octalactone 42, 71, 99 γ -Octanolactone 29, 85	64	Ethyl 3-phenylpropanoate	91, 104	1862	1322	MS, RI, Std	1.11	0.09	I		31.60	2.31
Guaiacol $81, 109, 124$ Phenylethyl alcohol $65, 91, 92$ δ -Octalactone $42, 71, 99$ γ -Octanolactone $29, 85$	65	Benzyl alcohol	79, 107, 108	1871	1012	MS, RI, Std	16.59b	1.50	122.99a	10.30	7.10b	0.25
Phenylethyl alcohol $65, 91, 92$ δ -Octalactone $42, 71, 99$ γ -Octanolactone $29, 85$	99	Guaiacol	81, 109, 124	1875	1062	MS, RI, Std	I		I		15.03	1.61
δ -Octalactone 42, 71, 99 γ -Octanolactone 29, 85	67	Phenylethyl alcohol	65, 91, 92	1905	1090	MS, RI, Std	99.91b	6.81	202.43a	19.46	67.88c	6.29
γ -Octanolactone 29, 85	68	ô-Octalactone	42, 71, 99	1918	1254	MS, RI, Std	I		23.86	2.18	I	
	69	γ -Octanolactone	29, 85	1919	1230	MS, RI, Std	I		I		68.91	5.88

Table	Table 2 (continued)										
No	Compounds	Quantitative ions	RI ^a		Identification ^b	Concentral	Concentration (µg/kg)	(
			HP-Innowax	DB-5		DJL	SD^{e}	KM	SD	CL	SD
02	4-Methyl-2-phenyl-2-pentenal	43, 91, 103	1919	1338	MS, RI, Std	8.41	0.93	12.83	1.15	1	
71	2-Phenyl-2-butenal	115, 117, 146	1929	1242	MS, RI, Std	13.41	1.76	26.33	2.34	I	
72	β-Ionone	43, 177	1938	1450	MS, RI, Std	10.72b	0.92	11.79b	1.01	24.47a	2.13
73	<i>cis</i> -Jasmone	79, 110, 164	1944	1367	MS, RI, Std	0.56c	0.02	30.89a	2.48	22.37b	1.94
74	Heptanoic acid	60, 73	1948	1053	MS, RI, Std	I		16.12	1.79	I	
75	(E)-3-Hexenoic acid	41, 55, 60	1955	978	MS, RI, Std	4.95	0.20	3.49	0.31	I	
76	2-Acetylpyrrole	39, 66	1970	1043	MS, RI, Std	9.44c	0.06	27.57b	2.07	34.46a	3.11
LL	β -Ionone epoxide	43, 123	1973	1454	MS, RI, Std	14.31b	1.34	16.97a	1.35	10.57c	0.89
78	γ -Nonalactone	85	2001	1332	MS, RI, Std	17.93	1.88	24.72	2.02	I	
62	2-Formylpyrrole	66, 94, 95	2026	992	MS, RI, Std	1.42	0.12	17.53	1.16	I	
80	2,5-Dimethyl-4-hydroxy-3-(2H)-furanone	129	2028	1055	MS, RI, Std	0.34c	0.01	0.52b	0.01	0.66a	0.03
81	Cinnamaldehyde	103, 131, 132	2033	1246	MS, RI, Std	5.61	0.34	I		41.64	4.46
82	Eugenol	164	2138	1324	MS, RI, Std	4.94b	0.21	5.81a	0.24	1.70c	0.10
83	(z)-7-Decen-5-olide	55, 71, 99	2274	1460	MS, RI, Std	13.23	0.88	44.42	4.01	I	
84	Methyl dihydrojasmonate	83	2288	1609	MS, RI, Std	11.45b	0.83	13.93a	0.95	13.25a	0.82
85	(\pm) -Dihydroactinidiolide	43, 109, 111	2376	1502	MS, RI, Std	39.26a	3.34	34.95a	3.26	40.03a	3.54
86	Coumarin	118, 146	2431	1409	MS, RI, Std	6.26c	0.41	27.55a	2.68	10.69b	1.09
87	Indole	89, 90, 117	2456	1265	MS, RI, Std	4.44b	0.21	8.59a	0.77	3.61b	0.38
88	3-Methylindole	130, 131	2503	1360	MS, RI, Std	2.34b	0.17	4.13a	0.33	0.82c	0.03
89	Vanillin	81, 151, 152	2566	1367	MS, RI, Std	11.99a	1.03	2.62b	0.10	2.10b	0.15
^a Refe	^a Retention index of communds on HP-Innows v and DB-5 columns	DB-5 columns									

Retention index of compounds on HP-Innowax and DB-5 columns

^bMethod of identification: RI: retention index; Std, confirmed by authentic standards; AD: aroma description

^cValues with different Roman letters (a–c) in the same row are significantly different according to the Duncan test (p < 0.05)

^dNot detected in samples ^eThe standard deviation of aroma intensity of compounds

 $\underline{\textcircled{O}}$ Springer

No	Compound	OT (µg/kg) ^a	OAV ^b		
			DJL	KM	CL
1	2-Methylbutanal	0.003	26	29	16
2	3-Methylbutanal	0.0011	37	82	24
3	Pentanal	0.005	3	3	-
4	Hexanal	0.005	12	19	6
5	β-Myrcene	0.015	5	12	4
6	(+)-Limonene	0.034	1	1	1
7	Heptanal	0.0028	2	3	2
8	(3E)-3,7-Dimethyl-1,3,6-octatriene	0.034	1	2	-
9	(Z)-3,7-Dimethylocta-1,3,6,-triene	0.034	<1	1	<1
10	Ethyl hexanoate	0.005	_c	-	1
11	cis-4-Heptenal	0.000025	25	94	-
12	p-Cymene	0.0114	<1	2	2
13	cis-3-Hexen-1-ol	0.05	2	<1	<1
14	(2E)-2-Octenal	0.003	2	2	11
15	cis-Linaloloxide	0.32	<1	1	<1
16	(E,E)-2,4-Heptadienal	0.056	2	2	1
17	Linalool	0.006	64	41	24
18	β-Cyclocitral	0.003	5	-	<1
19	Phenylacetaldehyde	0.0063	7	18	8
20	Ethyl decanoate	0.005	_	-	16
21	Acetophenone	0.065	_	-	1
22	Salicylaldehyde	0.03	_	-	2
23	Citral	0.028	<1	1	1
24	3-Methylnonane-2,4-dione	0.00003	28	18	21
25	Methyl salicylate	0.04	6	9	5
26	α-Ionone	0.00378	1	1	4
27	Geraniol	0.0066	20	97	2
28	Guaiacol	0.0016	_	-	9
30	Phenylethyl alcohol	0.14	<1	1	<1
31	γ-Octanolactone	0.0179	_	-	4
32	β-Ionone	0.0002	54	59	122
33	cis-Jasmone	0.00026	2	119	86
34	γ-Nonalactone	0.0079	2	3	_
35	Eugenol	0.0025	2	2	<1
36	Methyl dihydrojasmonate	0.013	<1	1	1
37	(±)-Dihydroactinidiolide	0.0021	19	17	19
38	3-Methylindole	0.00041	6	10	2

three world-famous black teas

^bThe odor activity values of volatile compounds in black tea infusions

^cThe compounds not detected in samples

compounds identified by GC-O and the OAV calculation were commonly regarded as the key aroma compounds. To verify the accuracy of qualitative and quantitative results, aroma recombination was conducted in this experiment. The 59 aroma compounds with $OAV \ge 1$ or AI > 0 were mixed according to concentrations calculated by standard curves. Sensory evaluations between the aroma recombination models and original black tea models were conducted. The sensory attributes used in this analysis were eight main aromas: sweet, woody, floral, caramel, green, fruity, baked and mushroom-like. The aroma profiles of the aroma recombination models and original black tea models were shown in Fig. 1, which indicated that there were slight differences between the recombination models and original samples. For example, the original model was sweeter and more baked in DJL than in recombination

Table	Table 4 Concentration and OAVs of sulfur compounds detected in three world-famous black teas	ılfur compounds d	etected in th	hree world-famous	black teas									
No	Compound	RI ^a		Identification ^b	OT (µg/kg) ^c	Concentrat	Concentration (µg/kg)					OAV ^d		
		HP-Innowax	HP-5			DIL	SD	KM	SD	CL	SD			
-	Methanethiol	701	<600	Std, RI, FPD	0.2	0.152a ^e	0.019	0.105b	0.009	0.048c	0.004	~	-	~
7	Dimethyl sulfide	751	< 600	Std, RI, FPD	0.3	5.432b	0.384	3.639c	0.243	7.948a	0.587	18	12	26
ю	1-Propanethiol	855	626	Std, FPD	3.1	0.129	0.011	ا		0.024	0.002	$\frac{1}{2}$	I	$\overline{\lor}$
4	2-Methylthiophene	1098	797	Std, RI, FPD	885	0.363b	0.015	1.658a	0.028	0.0292c	0.012	$\frac{1}{2}$	$\overline{\vee}$	$\overline{}$
5	2,5-Dimethylthiophene	1155	871	Std, RI, FPD	0.5	0.027	0.001	I		Ι		$\frac{1}{2}$	I	I
9	2-Isopropyl-4-methylthiazole	1352	1032	Std, RI, FPD	8	0.070	0.005	0.070	0.007	I		$\frac{1}{2}$	$\overline{\lor}$	I
L	Methionol	1720	666	Std, RI, FPD	0.128	0.440b	0.024	1.640a	0.021	0.120c	0.014	б	12	$\overline{\lor}$
8	3-Mercaptohexanol	1874	1127	Std, RI, FPD	0.013	0.042	0.003	0.038	0.001	I		3	ю	I
6	Benzothiazole	1977	1243	Std, RI, FPD	71	0.016	0.002	0.016	0.000	I		\sim	$\frac{1}{2}$	I

^aRetention index of compounds on HP-Innowax and HP-5 columns

^bMethod of identification: RI: retention index; Std, confirmed by authentic standards; FPD: flame photometric detector

^cThe odor threshold (OT) of sulfur compounds in water referred in the literature [26, 27]

^dThe odor activity values of sulfur compounds in black tea infusions

 $^{\circ}$ Values with different Roman letters (a-c) in the same row are significantly different according to the Duncan test (p < 0.05)

f Not detected in samples

model. In KM, the original model was sweeter and fruitier than its recombination model. In CL, the original model was sweeter and greener than recombination. The analysis of the eight sensory attributes between original black tea samples and aroma recombination models showed that aroma characteristics of the black tea recombination model were similar to the original sample. Therefore, the results confirmed the accuracy of identification and quantification of the key aroma compound in black teas.

Besides the sensory evaluation, the electronic nose was also applied to assess the differences between recombination and original samples. The electronic nose was a device that imitated the human sense of smell [43]. Its principle was based on the response signal generated by the interaction between aroma compounds and sensors [44]. Through the application of an electronic nose, the verification of aroma recombination could avoid innate human subjectivity, rending the results more objective. As shown in Fig. 2, the three figures represented the profile of aroma compounds the three black tea specimens and their aroma recombination. From the comparison results between the aroma profiles of original black tea infusions and profiles of aroma recombination, we could see that the aroma peaks of aroma recombination models were almost the same as those of the black tea infusions, which confirmed the accuracy of the qualitative and qualitative results. However, some peaks were slightly different from those of the black tea infusions. This might be due to the lack of unknown compounds, or the complex tea matrix containing tea polyphenol and other non-volatile compounds, which would affect the release of aroma compounds. Overall, the qualitative and quantitative results were almost identical to those of the original black tea infusions; the figures indicated that the aroma recombination models could successfully simulate the aroma of the top three of high-aroma black teas in the world.

Omission experiment

Omission experiments are often used to evaluate the contribution of aroma compounds to overall aroma [45, 46]. The omission experiments were designed to explore the importance of aroma compounds in black tea to the overall aroma. In the omission experiments, compounds with $OAV \ge 1$ or AI > 0 were selected for the omission experiments to explore their influence on the overall aroma. Noticeably, there were many non-volatiles compounds in tea infusions, which might affect the release of volatile organic compounds. Furthermore, the interaction between the aroma compounds might cause masking effect or synergistic effect, resulting in the change of actual OAVs [47]. In addition, the results of GC-O were perceived through air, while the thresholds in air were different from those in water. Therefore, it was necessary to use omission experiments to verify the characteristic aroma compounds in black tea. In the omission experiments, a total of 59 aroma compounds were recombined, and one aroma compound was omitted in each omission experiment. There were 50, 49, and 47 omission models in three black tea samples (DJL, KM, and CL, respectively), and the results of omission experiments were shown in Table 5.

Among the aldehydes of Darjeeling black tea, the omission models of 2-methylbutanal, 3-methylbutanal, benzaldehyde, phenylacetaldehyde, β-cyclocitral, and *cis*-4-heptenal showed extremely significant differences ($p \le 0.1\%$). Among the alcohols, the omission experiments of linalool, geraniol, and phenylethyl alcohol exhibited extremely significant differences ($p \le 0.1\%$). In addition, after removing β -ionone, methyl salicylate, and β -myrcene, the overall aroma also showed significant differences ($p \le 0.1\%$). Similarly, in Keemun black tea, when 2-methylbutanal, 3-methylbutanal, pentanal, hexanal, cis-4-heptenal, and cis-jasmone were omitted, most of the sensory panel members reached correct judgments. While in Ceylon black tea, the results of omission experiments showed significant differences ($p \le 0.1\%$) when omitting 2-methylbutanal, (2E)-2-octenal, salicylaldehyde, linalool, β -ionone, etc.

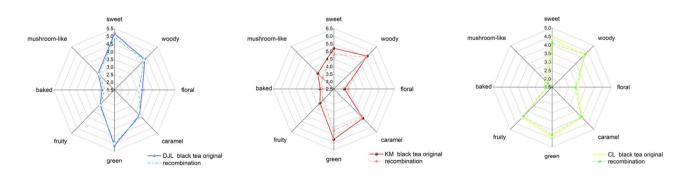


Fig. 1 Comparison of aroma profiles between original black tea infusions and the aroma recombination models by sensory panel

Table 5	Omission	experiments	results	of	each	aroma-active	com-
pound ($OAV \ge 1$ or	AI > 0					

No	Compound ^a	n^{b}		
		DJL	KM	CL
1	Aldehydes			
1–1	2-Methylbutanal	15*** ^c	14***	12***
1–2	3-Methylbutanal	12***	15***	11**
1–3	Pentanal	4=	13***	_d
1–4	Hexanal	9*	15***	8=
1–5	2-Methyl-2-butenal	2=	7=	3=
1–6	Heptanal	10**	12***	11**
1–7	(2E)-2-Octenal	6=	10**	12***
1–8	Furfural	8=	5=	3=
1–9	(E,E)-2,4-Heptadienal	9*	8=	6=
1–10	Benzaldehyde	12***	9*	10**
1–11	5-Methyl furfural	7=	5=	4=
1–12	1-Ethyl-2-pyrrolecarboxaldehyde	6=	7=	_
1–13	Citral	6=	8=	8=
1–14	β-Cyclocitral	13***	_	_
1–15	Phenylacetaldehyde	10**	12***	8=
1–16	Salicylaldehyde	_	_	12***
1–17	γ-Nonalactone	2=	8=	_
1–18	2-Formylpyrrole	5=	6=	_
1–19	Cinnamaldehyde	7=	_	8=
1–20	cis-4-Heptenal	12***	14***	_
2	Alcohols			
2–1	Pentanol	8=	4=	_
2–2	1-Hexanol	9*	_	5=
2–3	cis-3-Hexen-1-ol	10**	7=	8=
2–4	trans-2-Hexen-1-ol	7=	_	4=
2–5	1-Octene-3-ol	8=	5=	_
2–6	Linalool	14***	13***	13***
2–7	α-Terpineol	6=	3=	7=
2-8	Geraniol	12***	14***	9*
2–9	Nerol	3=	5=	7=
2–10	Benzyl alcohol	4=	4=	3=
2–11	Phenylethyl alcohol	12***	14***	11**
2–12	Guaiacol	_	_	11**
2–13	Eugenol	10**	11**	_
3	Ketones	10		
3–1	2-Heptanone	8=	5=	6=
3–2	6-Methylhept-5-en-2-one	7=	9*	10**
3–3	2-Acetylfuran	_	5=	9*
3–4	Acetophenone	_	_	10**
3–5	3-Methylnonane-2,4-dione	7=	4=	6
3–6	α-Ionone	, 5=	3=	11**
3–0 3–7	β-Ionone	12***	5– 12***	15***
3-8	<i>cis</i> -Jasmone	4=	12	12***
3–0 3–9	Coumarin	4– 9*	10**	8=
3–9 4	Easters	/	10	0-
+ 4–1	Ethyl hexanoate	_	_	10**
4–1 4–2	Ethyl decanoate	-	-	10***

No	Compound ^a	n ^b		
		DJL	KM	CL
4–3	Methyl salicylate	13***	15***	11**
4–4	γ-Octanolactone	-	-	12***
4–5	Methyl dihydrojasmonate	-	9*	8=
4–6	Dihydroactinidiolide	10**	8=	11**
5	Terpenes			
5-1	β-Myrcene	12***	15***	12***
5–2	(+)-Limonene	10**	12***	10**
5–3	(Z)-3,7-Dimethylocta-1,3,6,-triene	10**	12***	-
5–4	p-Cymene	-	4=	3=
6	Acids			
6–1	Acetic acid	2=	3=	3=
7	Surfur compounds			
7–1	Dimethyl sulfide	12***	12***	14***
7–2	Methionol	8=	11**	-
7–3	3-Mercaptohexanol	10**	11**	-
8	Others			
8-1	2-Acetylpyrrole	3=	5=	8=
8–2	3-Methylindole	8=	6=	4=
8–3	cis-Linaloloxide	10**	13***	11**

^aThe aroma-active compounds with OAV \geq 1 or AI > 0

^bCorrect judgment numbers for evaluating the difference of aroma by triangle tests

^cThe odor threshold (OT) of sulfur compounds in water referred in the literature.Significance: ***, 0.1% significant level $(p \le 0.1\%)$; **, 1% significant level $(p \le 1\%)$; *, 5% significant level $(\le 5\%)$;=, no significant difference

^dNot aroma-active compound in samples

The results of most of the omission experiments were consistent with the results of GC-O and OAVs. Notably, there were some differences between the results of the omission experiments and OAVs. The OAVs of α -ionone, (E, E)-2,4-heptadienal, (2E)-2-octenal, γ -nonalactone, 3-methylnonane-2,4-dione, 3-methylindole, and methionol in black tea were greater than 1. However, they did not appear important in omission experiments. On the contrary, the OAVs of cis-3-hexen-1-ol and benzaldehyde were less than 1, but they played important roles in the omission experiments. This might be because the thresholds of aroma compounds were determined in water when calculating OAVs, but the omission experiments were conducted in mixed aroma solutions, which meant that the thresholds might have changed. Besides, there might be perceptual interactions among aroma compounds, which affected the thresholds of the aroma compounds [48].

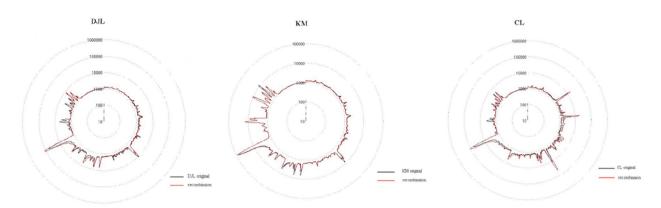


Fig. 2 Comparison of aroma profiles between original black tea infusions and the aroma recombination models by electronic nose

Conclusion

The characteristic aroma compounds in three world-famous black teas were comprehensively analyzed by SBSE/GC-O and OAV methods. A total of 98 volatile compounds were identified, and 87, 83, and 74 aroma compounds were determined in Darjeeling, Keemun, and Ceylon black tea, respectively. In this study, sulfur compounds in black tea were detected for the first time. The comprehensive analysis of the black tea would complement the aroma map of tea. The contributions of characteristic aroma compounds to overall aroma were evaluated through omission experiments. The comprehensive identification of the characteristic aroma compounds of black tea not only provides theoretical guidance to quality control of black tea, but also forms a fundamental basis for recombination and simulation of the black tea aroma.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00217-022-04039-2.

Acknowledgements This work was sponsored by "Shuguang Progran" [20SG55] supported by Shanghai Education Development Foundation and Shanghai Municipal Education Commission, Capacity building project of local universities Science and Technology Commission of Shanghai Municipality [21010503900] and Program of Shanghai Academic Research Leader [21XD1423800].

Declarations

Conflict of interest Authors have declared no conflict of interests.

Compliance with ethics requirements This article does not contain any studies with human or animal subjects.

References

 Wang L-F, Lee J-Y, Chung J-O, Baik J-H, So S, Park S-K (2008) Discrimination of teas with different degrees of fermentation by SPME–GC analysis of the characteristic volatile flavour compounds. Food Chem 109(1):196–206

- Flaten TP (2002) Aluminium in tea—concentrations, speciation and bioavailability. Coord Chem Rev 228(2):385–395
- Bhattacharyya N, Seth S, Tudu B, Tamuly P, Jana A, Ghosh D, Bandyopadhyay R, Bhuyan M, Sabhapandit S (2007) Detection of optimum fermentation time for black tea manufacturing using electronic nose. Sens Actuators, B Chem 122(2):627–634
- Kang S, Yan H, Zhu Y, Liu X, Lv H-P, Zhang Y, Dai W-D, Guo L, Tan J-F, Peng Q-H (2019) Identification and quantification of key odorants in the world's four most famous black teas. Food Res Int 121:73–83
- Yener S, Sánchez-López JA, Granitto PM, Cappellin L, Märk TD, Zimmermann R, Bonn GK, Yeretzian C, Biasioli F (2016) Rapid and direct volatile compound profiling of black and green teas (*Camellia sinensis*) from different countries with PTR-ToF-MS. Talanta 152:45–53
- Larson C (2015) Reading the tea leaves for effects of climate change. Science 348(6238):953–954. https://doi.org/10.1126/ science.348.6238.953
- Ohno A, Oka K, Sakuma C, Okuda H, Fukuhara K (2011) Characterization of tea cultivated at four different altitudes using 1H NMR analysis coupled with multivariate statistics. J Agric Food Chem 59(10):5181–5187
- Zhu M, Li E, He H (2008) Determination of volatile chemical constitutes in tea by simultaneous distillation extraction, vacuum hydrodistillation and thermal desorption. Chromatographia 68(7):603–610
- 9. Lasekan O, Lasekan A (2012) Flavour chemistry of mate and some common herbal teas. Trends Food Sci Technol 27(1):37–46
- Xiao Z, Wang H, Niu Y, Liu Q, Zhu J, Chen H, Ma N (2017) Characterization of aroma compositions in different Chinese congou black teas using GC–MS and GC–O combined with partial least squares regression. Flavour Fragr J 32(4):265–276
- Kumazawa K, Wada Y, Masuda H (2006) Characterization of epoxydecenal isomers as potent odorants in black tea (Dimbula) infusion. J Agric Food Chem 54(13):4795–4801
- Magagna F, Cordero C, Cagliero C, Liberto E, Rubiolo P, Sgorbini B, Bicchi C (2017) Black tea volatiles fingerprinting by comprehensive two-dimensional gas chromatography–Mass spectrometry combined with high concentration capacity sample preparation techniques: toward a fully automated sensomic assessment. Food Chem 225:276–287
- Kraujalytė V, Pelvan E, Alasalvar C (2016) Volatile compounds and sensory characteristics of various instant teas produced from black tea. Food Chem 194:864–872

- Baba R, Amano Y, Wada Y, Kumazawa K (2017) Characterization of the potent odorants contributing to the characteristic aroma of Matcha by gas chromatography–olfactometry techniques. J Agric Food Chem 65(14):2984–2989
- 15. Chen X, Chen D, Jiang H, Sun H, Zhang C, Zhao H, Li X, Yan F, Chen C, Xu Z (2019) Aroma characterization of Hanzhong black tea (*Camellia sinensis*) using solid phase extraction coupled with gas chromatography–mass spectrometry and olfactometry and sensory analysis. Food Chem 274:130–136
- Nickerson GB, Likens S (1966) Gas chromatography evidence for the occurrence of hop oil components in beer. J Chromatogr A 21:1–5
- Lau H, Liu SQ, Xu YQ, Lassabliere B, Sun J, Yu B (2018) Characterising volatiles in tea (*Camellia sinensis*). Part I: comparison of headspace-solid phase microextraction and solvent assisted flavour evaporation. LWT 94:178–189
- Li Q, Li Y, Luo Y, Xiao L, Wang K, Huang J, Liu Z (2020) Characterization of the key aroma compounds and microorganisms during the manufacturing process of Fu brick tea. LWT 127:109355
- Steingass CB, Dickreuter J, Kuebler S, Schweiggert RM, Carle R (2021) Influence of fruit logistics on fresh-cut pineapple (*Ananas comosus* [L.] Merr.) volatiles assessed by HS-SPME–GC–MS analysis. Eur Food Res Technol 247(7):1617–1630
- Ye N, Zhang L, Gu X (2012) Discrimination of green teas from different geographical origins by using HS-SPME/GC–MS and pattern recognition methods. Food Anal Methods 5(4):856–860
- 21. Baltussen E, Sandra P, David F, Cramers C (1999) Stir bar sorptive extraction (SBSE), a novel extraction technique for aqueous samples: theory and principles. J Microcolumn Sep 11(10):737–747
- 22. Kawaguchi M, Ito R, Saito K, Nakazawa H (2006) Novel stir bar sorptive extraction methods for environmental and biomedical analysis. J Pharm Biomed Anal 40(3):500–508
- 23. David F, Ochiai N, Sandra P (2019) Two decades of stir bar sorptive extraction: a retrospective and future outlook. TrAC, Trends Anal Chem 112:102–111
- Wang M-Q, Ma W-J, Shi J, Zhu Y, Lin Z, Lv H-P (2020) Characterization of the key aroma compounds in Longjing tea using stir bar sorptive extraction (SBSE) combined with gas chromatography-mass spectrometry (GC–MS), gas chromatography-olfactometry (GC-O), odor activity value (OAV), and aroma recombination. Food Res Int 130:108908
- Atanasova B, Langlois D, Nicklaus S, Chabanet C, Etiévant P (2004) Evaluation of olfactory intensity: comparative study of two methods. J Sens Stud 19(4):307–326
- 26. Xu X, Xu R, Jia Q, Feng T, Huang Q, Ho C-T, Song S (2019) Identification of dihydro-β-ionone as a key aroma compound in addition to C8 ketones and alcohols in Volvariella volvacea mushroom. Food Chem 293:333–339
- 27. Zhu J, Niu Y, Xiao Z (2021) Characterization of the key aroma compounds in Laoshan green teas by application of odour activity value (OAV), gas chromatography-mass spectrometry-olfactometry (GC-MS-O) and comprehensive two-dimensional gas chromatography mass spectrometry (GC× GC-qMS). Food Chem 339:128136
- Van Gemert L (2003) Odour thresholds. Compilations of odour threshold values in air, water and other media Utrecht (The Netherlands): Oliemans Punter & Partners BV
- Niu Y, Deng J, Xiao Z, Zhu J (2021) Characterization of the major aroma-active compounds in peach (*Prunus persica* L. Batsch) by gas chromatography–olfactometry, flame photometric detection and molecular sensory science approaches. Food Res Int 147:110457
- ISO B 4120 (2004) Sensory analysis-methodology-triangle test (2012)

- 31. Niu Y, Zhang J, Xiao Z, Zhu J (2020) Evaluation of the perceptual interactions between higher alcohols and off-odor acids in Laimao Baijiu by σ - τ plot and partition coefficient. J Agric Food Chem 68(50):14938–14949
- Kanasawud P, Crouzet JC (1990) Mechanism of formation of volatile compounds by thermal degradation of carotenoids in aqueous medium. 1. beta.-Carotene degradation. J Agric Food Chem 38(1):237–243
- Li C, YuLong Y, ChunYan W, JingNa Y, HuaRong T (2019) Research progress on carotenoid aroma precursors in tea. Food Ferment Ind 45(5):266–273
- Mahanta PK, Baruah S, Owuor PO, Murai T (1988) Flavour volatiles of Assam CTC black teas manufactured from different plucking standards and orthodox teas manufactured from different altitudes of Darjeeling. J Sci Food Agric 45(4):317–324
- Alasalvar C, Topal B, Serpen A, Bahar B, Pelvan E, Gökmen V (2012) Flavor characteristics of seven grades of black tea produced in Turkey. J Agric Food Chem 60(25):6323–6332
- Baldermann S, Yang Z, Katsuno T, Tu VA, Mase N, Nakamura Y, Watanabe N (2014) Discrimination of green, oolong, and black teas by GC-MS analysis of characteristic volatile flavor compounds. Am J Anal Chem. https://doi.org/10.4236/ajac.2014. 59070
- 37. Yang Z, Baldermann S, Watanabe N (2013) Recent studies of the volatile compounds in tea. Food Res Int 53(2):585–599
- Gijs L, Perpete P, Timmermans A, Collin S (2000) 3-Methylthiopropionaldehyde as precursor of dimethyl trisulfide in aged beers. J Agric Food Chem 48(12):6196–6199
- Ho C-T, Zheng X, Li S (2015) Tea aroma formation. Food Sci Human Wellness 4(1):9–27
- Sanderson GW, Grahamm HN (1973) Formation of black tea aroma. J Agric Food Chem 21(4):576–585
- Kawabata M, Ohtsuki K, Kokura H, Wakahara Y (1977) Determination of dimethylsulfide in the head space vapor of green tea by gas chromatography. Agric Biol Chem 41(11):2285–2287
- 42. Liu H, Wang Z, Zhang D, Shen Q, Pan T, Hui T, Ma J (2019) Characterization of key aroma compounds in Beijing roasted duck by gas chromatography–olfactometry–mass spectrometry, odoractivity values, and aroma-recombination experiments. J Agric Food Chem 67(20):5847–5856
- Ampuero S, Zesiger T, Gustafsson V, Lundén A, Bosset J (2002) Determination of trimethylamine in milk using an MS based electronic nose. Eur Food Res Technol 214(2):163–167
- Gardner JW, Bartlett PN (1994) A brief history of electronic noses. Sens Actuators, B Chem 18(1–3):210–211
- Niu Y, Sun X, Xiao Z, Wang P, Wang R (2018) Olfactory impact of terpene alcohol on terpenes aroma expression in Chrysanthemum essential oils. Molecules 23(11):2803
- 46. Fischer A, Schieberle P (2009) Characterisation of the key aroma compounds in the peel oil of Pontianak oranges (*Citrus nobilis* Lour. Var. microcarpa Hassk.) by aroma reconstitution experiments. Eur Food Res Technol 229(2):319–328
- 47. Wu C, Liu J, Yan L, Chen H, Shao H, Meng T (2015) Assessment of odor activity value coefficient and odor contribution based on binary interaction effects in waste disposal plant. Atmos Environ 103:231–237
- Xiao Z, Chen L, Niu Y, Zhu J, Zhang J, Deng J (2021) Evaluation of the Interaction Between Esters and Sulfur Compounds in Pineapple Using Feller's Additive Model, OAV, and Odor Activity Coefficient. Food Anal Methods. https://doi.org/10.1007/ s12161-021-02001-8

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.