ORIGINAL PAPER

Nutraceutical properties and phytochemical characterization of wild Serbian fruits

Maja Natić¹ • Aleksandra Pavlović¹ • Fabrizia Lo Bosco² • Nemanja Stanisavljević³ • Dragana Dabić Zagorac⁴ • **Milica Fotirić Akšić5 · Adele Papetti[6](http://orcid.org/0000-0003-1523-7759)**

Received: 31 May 2018 / Revised: 5 October 2018 / Accepted: 14 October 2018 / Published online: 31 October 2018 © Springer-Verlag GmbH Germany, part of Springer Nature 2018

Abstract

Wild fruits grown in Serbia, i.e., elderberry (*Sambucus nigra*), hawthorn (*Crataegus monogyna*), cornelian cherry (*Cornus mas*), and blackthorn (*Prunus spinosa*), are rich in secondary metabolites. In this study, the polyphenolic composition of wild fruit extracts and their antioxidant capacity were investigated by in vitro assays. Elderberry was characterized by the presence of arbutin (a skin protector), and cornelian cherry by syringic and gallic acids. In blackthorn, at least 11 different phenolic compounds were reported for the first time, including vanillic acid and naringin, the metabolite present in the highest amount. Blackthorn extracts were the richest in polyphenols (11.24–18.70 g GAE/kg FW) and had the highest activity in the DPPH radical test (180.93–267.11 mMTE/mL), while cornelian cherry extracts showed the most effective ferric ion chelating (81.37–90.66%) and antityrosinase inhibition capacities (21.75–74.23%). No sample was able to scavenge NO. Using the principal component analysis, wild fruit samples were classified into four separate clusters due to distinctive phenolic profiles and antioxidant capacity. Our investigation showed how every fruit could be considered unique in terms of its phytonutrient content. Thus, Serbian wild fruits may be a great source of bioactive natural compounds and could be therefore considered particularly useful in food supplement production. Particularly, as a source of natural antioxidants, these species could be used to extend the shelf life of food products and replace synthetic antioxidants, avoiding potential health risks and toxicity.

Keywords Wild Serbian fruits · UHPLC–DAD–HESI–MS/MS · Phenolic profile · Antioxidant potential · Functional foods

Electronic supplementary material The online version of this article [\(https://doi.org/10.1007/s00217-018-3178-1\)](https://doi.org/10.1007/s00217-018-3178-1) contains supplementary material, which is available to authorized users.

 \boxtimes Adele Papetti adele.papetti@unipv.it

- ¹ Faculty of Chemistry, University of Belgrade, Belgrade 11158, Serbia
- ² Biophysics Institute, National Research Council, 90143 Palermo, Italy
- Institute of Molecular Genetics and Genetic Engineering, University of Belgrade, Belgrade 11000, Serbia
- ⁴ Innovation Centre, University of Belgrade, Belgrade 11000, Serbia
- ⁵ Faculty of Agriculture, University of Belgrade, Zemun 11080, Serbia
- ⁶ Department of Drug Sciences, University of Pavia, Viale Taramelli 12, 27100 Pavia, Italy

Introduction

Over the last decade, wild fruits have received increasing attention due to their medicinal properties and high nutritional and nutraceutical values, being a rich source of antioxidant compounds, vitamins, minerals, and polyphenols [\[1](#page-8-0)]. Serbia is considered to be a secondary gene center for some representatives of the *Rosaceae* family, as it has a very rich gene pool for indigenous fruits with more than 100 species classified into 15 different families. It is mostly used as rootstocks for fruit production, and as noble-wood production for making furniture in prevention of erosion, for landscape architecture, and in bee keeping [[2](#page-8-1)]. The most widespread species belong to *Crataegus, Prunus, Cornus*, and *Sambucus* genera. Due to their abundance in proanthocyanidins, catechins, phenolic acids, essential oils, and terpenoids, *Crataegus monogyna* (hawthorn, HW) fruits possess strong antioxidant potential [[3\]](#page-8-2). Furthermore, several studies have showed the positive influence of HW fruit extracts on the cardiovascular system, acting as antiarrhythmic, hypolipidemic, and hypotensive agents [[4,](#page-8-3) [5\]](#page-8-4). *Prunus spinosa* (blackthorn or sloe, BL) is mainly used to make jellies, jams, vinegar, wine, ice-cream, and distilled alcoholic and non-alcoholic beverages [[6\]](#page-8-5). BL fruits are rich in polyphenols, vitamins, and minerals [[7\]](#page-8-6). Significant antimicrobial activity was reported, suggesting the usefulness of this fruit in food and pharmaceutical industries [[8\]](#page-8-7). *Cornus mas* L. (cornelian cherry, CC) is mainly used as food and possesses many pharmacological and therapeutic effects [[9–](#page-8-8)[11\]](#page-8-9). *Sambucus nigra* (elderberry, EL) fruits are generally eaten when fully ripe, after cooking, or can be used to make jams and jellies [[12](#page-8-10)]. Literature reports the use of EL in treating several diseases such as diabetes, colic, diarrhea, fever, coughs, colds, congestion, bronchitis, influenza, allergies, rheumatism, swollen limbs, burns, and inflamed mucous membranes [[13\]](#page-8-11).

According to the International Federation of Organic Agriculture Movements (IFOAM) and Mediterranean Organic Agriculture Network (MOAN), wild fruits in Serbia are collected on around 450,000 ha, but just 1550 ha are certified for organic production [\[14](#page-8-12)]. Harvesting of wild berries is traditionally performed in rural economies and at least ten companies are exporting collected wild fruits worth several millions of euros. Wild fruits are processed in households as a part of tradition in the entire Serbia and are sold through specialized retail points such as green markets and specialty shops. Although the domestic market is growing, wild berries collection is export driven. Besides, fruits are used in folk medicine, and the plants as ornamental and honey plants. The aim of including wild berries in Serbian agriculture is the diversification of agriculture throughout the region, the development of new types of agricultural products and their offer in local markets, the introduction of new raw materials in the food industry and of new sources of income for local producers and family farms, as well as the preservation of biodiversity [\[2](#page-8-1)].

Chemical fingerprinting is a valuable method to find distinctive patterns or to identify a specific feature that could be potentially used to verify the identity of a certain plant or food material and is generally strictly related to the antioxidant capacity. Therefore, the aim of this study was to investigate the polyphenolic profile and nutraceutical properties of Serbian hawthorn, blackthorn, cornelian cherry, and elderberry fruits. The total phenolic content (TPC) was evaluated and the quantification of each phenolic compound was performed by ultrahigh performance liquid chromatography (UHPLC) coupled with a diode array detector (DAD) and connected to a triple-quadrupole mass spectrometer. The nutraceutical properties of the wild fruit extracts were evaluated using five assays: DPPH•-scavenging activity, ferric ion-reducing capacity (FRC), ferrous ion-chelating capacity (FCC), nitric oxidescavenging activity (NO), and antityrosinase activity test (Tyr). The characterization of the phenols was reported, and the main quantitative differences in phenolic composition among the berries are emphasized and discussed comparing the results available in literature. Finally, principal component analysis (PCA) was employed to find chemical markers for discrimination among studied berries on the basis of the polyphenolic composition and different nutraceutical properties. The presented approach is in agreement with the most commonly used procedures in the analysis of plant material. Mass spectrometry coupled to liquid chromatography and statistical data analysis in food profiling enables finding key compounds responsible for discrimination and classification of samples. Such an investigation could contribute to the assessment of the possible usage of wild fruit extracts in the formulation of food supplements and/or cosmetic products.

Materials and methods

Wild fruit samples and extract preparation

The autochthonous *Crataegus monogyna* (single seeded hawthorn, HW), *Crataegus oxyacantha* (Northern Europian hawthorn, HW), *Prunus spinosa* (blackthorn, BL), *Sambucus nigra* (elderberry, EL), and *Cornus mas* (cornelian cherry, CC) fruits (identified according to the taxonomical criteria of [\[15\]](#page-8-13)) were selected from the spontaneous flora and collected for experiments from different sites (Table [1\)](#page-2-0). Each plant was authenticated and preserved at the Department of Botany and Systematic at the Faculty of Agriculture, University of Belgrade. The voucher numbers for the analyzed wild species are: *Cornus mas*—PF 2016/10211 (flowers, leaves, fruits) *Crataegus monogyna*—PF 2016/11367 (flowers, leaves, fruits) *Crataegus oxyacantha*—PF 2016/11370 (flowers, leaves, fruits) *Prunus spinosa*—PF 2106/12763 (flowers, leaves, fruits) *Sambucus nigra*—PF 2016/13287 (flowers, leaves, fruits).

Thirty mature fruits of each genotype were picked at the full maturity stage. Three different samples were collected from the same tree/bush, but picked from different sides; each sample was submitted to the extraction procedure and analyzed in triplicate. After harvesting, fruits were placed in AC/DC hand-refrigerator (−20 °C) to prevent oxidation up to the extract preparation. Each sample of frozen fruits were ground up with a Mikro-Dismembrator S (Sartorius, Göttingen, Germany) and extracted under magnetic stirring for 10 min (500 mg/10 mL 70% methanol). The supernatant was collected after centrifugation at 4000 rpm for 5 min, filtered through a 0.45 µm membrane filter (Millex-HV hydrophilic polyvinylidene difluoride (PVDF), Millipore, Billerica, MA, USA) and kept at -20 °C prior to analysis that were all performed in triplicate.

EL5 Tara EL6 Tara EL7 Tara

EL1 Deliblato Sands

EL2 Deliblato Sands EL3 Deliblato Sands EL4 Deliblato Sands

CC2 Fruška Gora CC3 Fruška Gora CC4 Fruška Gora

BL2 Radmilovac BL3 Saranovo BL4 Saranovo BL5 Beograd

HW2 Radmilovac HW3 Gornji Milanovac

CC6 Stara Planina, Radičevac CC7 Stara Planina, Radičevac CC8 Stara Planina, Zlot CC9 Stara Planina, Zlot CC10 Stara Planina, Zlot

Species Code Location

Sambucus nigra Elderberry

Cornus mas Cornelian cherry

Prunus spinosa Blackthorn

Crataegus monogyna

Hawthorn

Hawthorn

The total phenolic content was determined spectrophotometrically by Folin–Ciocalteu method [[25](#page-9-0)] following the procedure reported in [[26](#page-9-1)]. Gallic acid standard solutions (20–100 mg/L) were used for constructing a calibration curve and the results were expressed in grams of gallic acid equivalents (GAE) per kilogram of frozen sample (g GAE/kg FW).

Antioxidant activity assays

The antioxidant activity of the extracts was evaluated by different methods: DPPH $[27]$ $[27]$ $[27]$, Fe²⁺-chelating capacity (FCC) $[28]$ $[28]$ $[28]$, and Fe³⁺-reducing capacity (FRC) $[29]$ $[29]$ assays. Nitric oxide (NO)-scavenging activity [\[30](#page-9-5)] and antityrosinase activity (Tyr) [[31](#page-9-6)] were also evaluated. A more detailed description of the used methods is reported in Supplementary material.

Analysis of polyphenols

Folk medicine, bee pasture, ornamental tree, often as hedges [[8](#page-8-7), [22,](#page-8-19) [23\]](#page-9-7)

BL1 Radmilovac Folk medicine, ornamental plant, syrups, juices, jams, compotes, spirits, pickled like

HW1 Radmilovac Jams, jellies, preserves, wine, vinegar, and distilled alcoholic beverages, digestive and

fresh juice

Crataegus oxyacantha HW4 Stara Planina, Radičevac Folk medicine, jam, yoghurt, juice, wine, liqueurs, marmalade, juice, and pies [\[24\]](#page-9-8)

olives, used to prepare wine and other traditional products

Wood is used to make furniture, jewellery and traditional musical instruments [[21](#page-8-18)]

laxative liqueurs, or ingredients for pastries. Flowers and petals used for tea, syrup,

Polyphenols were separated, quantified and identified using a UHPLC system (Dionex Ultimate 3000, ThermoFisher Scientific, Bremen, Germany) equipped with a diode array detector (DAD) and triple-quadrupole mass spectrometer. Chromatographic separations and quantitative analysis of phenolic compounds were achieved following the optimized method previously reported $[26]$ $[26]$ $[26]$. More details can be found in Supplementary material.

Statistical analysis

Measurements done in triplicate are presented as the mean values \pm standard deviation (SD). Tukey's test was performed using NCSS program. Principal component analysis (PCA) was done using the PLS_Tool Box software package for MATLAB (Version 7.12.0).

Results and discussion

Investigation of *Crataegus monogyna* started long ago, but in most of the studies carried out so far, the analysis of phenolic extracts from leaves and flowers has been done [\[4,](#page-8-3) [5](#page-8-4), [32\]](#page-9-9). Nowadays, oil obtained from the seeds is examined for its antioxidant and antibacterial activities [[33\]](#page-9-10). Even if Cornelian cherry (*Cornus mas*) has been an important plant from ancient time, its analysis has started recently. References [[20](#page-8-17) and [21](#page-8-18)] determined vitamin C, sugars, anthocyanins, organic acids and tannins in its fruits, and [\[9](#page-8-8) and [11](#page-8-9)] continued those studies. Since the consumer awareness of high quality and healthy food has been increasing, blackthorn (*Prunus spinosa*) has become an interesting species for chemical analysis. Reference [[32\]](#page-9-9) proved that it contains substantial quantities of phenolic antioxidants. Besides, [\[34,](#page-9-11) [35](#page-9-12)] showed that extracts and liquors made from blackthorn can treat hypertension and gastrointestinal disturbances. In elderberry, flowers and fruits have been gaining a lot of attention. They have been present in folk medicine for centuries, but the pharmacokinetics of many of their constituents has not been completely understood. Many scientist have reported substantial levels of flavonoids, anthocyanins and hemagglutinin, including sambunigrin, viburnic acid, and vitamins A and C [[36\]](#page-9-13).

Bioactivity assays of wild fruits

The nutraceutical properties of the tested extracts were evaluated using different bioassays providing different information: DPPH about the hydrogen-donating ability of samples, Fe^{2+} -chelating assay about the capacity of extracts to chelate transition metal ions involved in the reactions generating ROS, Fe^{3+} -reducing assay about the capacity of an antioxidant agent to reduce $Fe³⁺$, NO-scavenging assay about the ability of extracts to compete with oxygen, leading to reduced production of nitrite ions, and finally antityrosinase assay about the capacity of extracts to reduce melanin formation through tyrosinase activity.

The results obtained in the different bioassays and the TPC of each tested extract are reported in Fig. [1.](#page-4-0) BL and CC extracts had the highest and the lowest TPC values showing an average content of 14.77 and 7.55 g GAE/ kg FW, respectively. The TPC values obtained for CC are slightly higher than those registered for samples collected in different regions in Turkey and Iran [\[37](#page-9-14), [38\]](#page-9-15), but lower than the results obtained for samples from Greece [\[39\]](#page-9-16); this variability confirms the need to conduct further studies among native CC population (no commercial CC cultivar exists) to determine those higher in nutraceutical properties. HW showed TPC values in the range 5.45–11.27 g GAE/kg, i.e., a mean content higher than that reported by [[40\]](#page-9-17) for ethanolic extracts. Conversely to our results, [[41](#page-9-18)] reported higher TPC for HW than for CC. Therefore, we can conclude that our results agree only in part with those reported in literature for the different wild fruits, as the phenolic profiles depend on cultivars and genotype, and on the environmental conditions and the TPC values are solvent and botanical organ dependent. Considering the bioactivity assays, BL extracts were the most active as DPPH radical scavengers with an average value of 227.08 mM TE/mL, followed by CC (115.96 mM TE/mL) and HW (119.89 mM TE/mL), while EL extracts showed the lowest activity (37.13 mM TE/mL extract). The strongest activity of BL against DPPH could be ascribed to its high TPC content, but also to its high FRC. Total phenolic content is expected to be strongly correlated to the contents of flavonoids and phenolic acids, and to change in dependence on cultivars, environmental conditions and solvent used for the extraction. Various solvent extracts (dichloromethane, ethyl acetate, ethanol, and aqueous solvent) of branches, leaves and fruits from blackthorn (*P. spinosa*) were analyzed spectrophotometrically and the results of TPC and TAC showed solvent and botanical organ dependence [\[42\]](#page-9-19).

Conversely, no activity in scavenging NO was detected independently from the considered fruit. All samples showed Tyr with values ranging from 20.90 to 74.23%. CC samples generally were the most active with the exception of CC4, CC5, and CC9 which showed Tyr values similar to those of EL samples. Intermediate values have been registered for BL and HW fruits. This capacity is interesting considering the fact that tyrosinase enzyme is involved in human melanization process and is also important in the developmental processes in insects. Therefore, potentially, inhibitors could be considered useful in food industry as food additive, in cosmetic industry as whitening agents, and also as insect control agents. The capacity to inhibit tyrosinase mediating melanogeneses was previously reported only for Tunisian *Crataegus monogyna* leaf constituents [[43\]](#page-9-20) and for proanthocyanidins isolated from Chinese HW fruit stone [[44\]](#page-9-21). As shown in Fig. [1](#page-4-0), FCC for all the extracts was in the range of 58.81–97.68%, with the following trend: $CC > EL > BL > HW$. It is worthy to be mentioned that CC extracts exhibiting the strongest FCC showed also the highest Tyr activity. This result can be supported by some earlier findings on the significance of the metal chelation ability when the inhibition of tyrosinase is considered [\[45](#page-9-22)].

Finally, data reported in Fig. [1](#page-4-0) showed divergent bioactivities of the extracts belonging to the same species, but such a behavior is quite usual as differences in the phytochemical profiles exist. Our findings are in agreement with [[46](#page-9-23)] who reported differences in the total phenolic, total **Fig. 1** TPC (**a**) and bioactivity results obtained in DPPH (**b**), Fe2+-chelating capacity (**c**), Fe3+-reducing capacity (**d**), and antityrosinase activity (**e**) assays for the tested samples

flavonoids, phenolic compounds, the mineral content, and the antioxidant activity of fruit extracts of seven wild species (*Crataegus monogyna* Jacq., *Prunus spinosa* L., *Rosa canina* L., *Hippophaë rhamnoides* L., *Rubus fruticosus* L., *Prunus padus, Cornus mas* L.). The analysis of phenolics resulted in quantification of numerous phenolic acids and flavonoids, and it was observed that the activity was more related to the type of individual phenolic compounds than to the total phenolic contents.

Quantification of the polyphenols

The quantitative mean data of individual phenolic compounds are summarized in Table [2](#page-5-0). A set of 12 phenolic

Table 2 Quantitative phenolic composition of the wild fruit extracts obtained by UHPLC-DAD-HESI-MS/MS (mg/kg) **Table 2** Quantitative phenolic composition of the wild fruit extracts obtained by UHPLC–DAD–HESI–MS/MS (mg/kg)

 $\underline{\textcircled{\tiny 2}}$ Springer

geni, *23* kaempferol

acids and 11 flavonoids were determined. The phenolic profiles are quite different both with regard to the composition and the concentration of phenolic acids and flavonoids. BL extracts showed a characteristic profile with a lower number of phenolic acids and flavonoids when compared to other fruit extracts. In fact, among phenolic acids, gallic acid, *p*-hydroxybenzoic acid, caffeic acid (with the exception of BL2 sample), syringic acid, and *p*-coumaric acid were not confirmed as well as arbutin. Syringic acid was not detected, differently from [[47\]](#page-9-24) who identified it in the hydro-alcoholic extract of Southeast Serbian fruits. Interestingly, vanillic acid was detected in the tested samples in the range 1.22–3.17 mg/kg FW: this acid has been never identified in BL extracts before (but only in cranberry [[48\]](#page-9-25)). Catechin, luteolin, kaempferol, and phlorizin (with the exception of BL1 sample) were not detected in BL extracts. Similarly, catechin and kaempferol were not detected in EL extracts which were rich in rutin (187.30–471.51 mg/kg FW), differently from CC, BL, and HW samples. Arbutin was confirmed only in EL extracts in the concentration range 23.12–31.99 mg/ kg FW. This is, most probably, the first report on the presence of arbutin in EL: this compound has been previously reported only in strawberry tree [[49](#page-9-26)]. Arbutin, as a tyrosinase inhibitor, is extensively used as skin-lightening agent in human cosmetic products. However, EL extracts did not show high Tyr values, differently from HW and CC extracts. Such results could be explained by the fact that apart from arbutin, other polyphenols known as tyrosinase inhibitors, such as ellagic and vanillic acid, were present in larger quantities in CC. Aesculin, a natural substance possessing sunscreen, antioxidant, and antitumor activities [[50\]](#page-9-27), was present in all samples in a concentration range of 1.71–14.69 mg/kg FW. All berry fruits (with the exception of HW2 sample) stored naringin (useful for the treatment of several metabolic disorders [[51](#page-9-28)]) and quercetin 3-*O*-galactoside with the highest concentration in BL and HW, respectively. Three HW samples stored *p*-coumaric acid. Interestingly, arbutin was identified in HW3 sample, which was additionally characterized by a slightly higher content of aesculin, rutin, quercetin 3-*O*-galactoside, and phlorizin. The presence of polyphenols in HW fruits was previously reported by [[52](#page-9-29)]. *p*-Coumaric acid in EL samples was in the range 1.08–2.86 mg/kg FW, differently from CC, that contains a lower amount (from 0.33 to 1.19 mg/kg FW), and BL extracts, in which this acid was absent, in accordance with [[49](#page-9-26)]. Gallic acid, so far reported to be present in blackberry and cloudberry fruit [[53](#page-9-30)], was identified only in CC as well as syringic acid. The only exception was HW1 sample, containing 0.58 mg/ kg FW. Recently, syringic acid attracted particular attention since it showed blood sugar regulating properties and was demonstrated to inhibit the development of diabetic cataract [[54](#page-9-31)]. Finally, ellagic acid was identified in all the extracts, with CC showing a notably higher concentration than in all the other extracts.

Principal component analysis

Multivariate statistical analysis in polyphenol profiling enables clustering of extracts according to individual flavonoids, phenolic acids, and other non-flavonoid components. The score plots display trends, groupings and outliers. In this work, PCA on the quantitative data was employed to analyze wild fruits extracts. The content of each determined phenolic compound and each bioactivity assay served as input data. The results of PCA suggested that a seven-component model explained 84.37% of total variance (PC1, PC2, PC3, PC4, PC5, PC6, and PC7 accounted for 29.95%, 16.24%, 13.58%, 8.54%, 7.45%, 5.22%, and 3.39%, respectively). The scores plot and loadings plot for the first three principal components are shown in Fig. [2](#page-7-0). The PCA scores plot discriminated the analyzed wild fruit samples into four separate clusters. From the loadings plot of PCA, it was possible to partly express the influence of the original variables on the separation among samples. EL samples formed separate cluster along PC1 which is in agreement with the distinctive phenolic profile when compared to the other samples investigated. EL fruits were characterized by higher contents of several phenolic acids (compounds 3, 5, 10, and 12) and notably higher content of rutin (14). Compounds 2, 9, and 11 and Tyr were important variables for the separation of CC along PC2. Finally, score plot revealed formation of two clusters of HW and BL samples along PC3. HW was characterized by notably higher contents of compound 15 than other samples, while BL demonstrated the highest DPPH˙ scavenger activity, TPC values and contents of compound 17.

No evidence of relationships was found for most of the studied genotypes according to their geographic location. Our results are not unexpected considering the fact that hawthorns and blackthorn are self-sterile, while Cornelian cherry is partially self-fertile, so characterized by a high degree of heterozygosity. For that reason in each locality, different genotypes can be found.

Conclusion

Serbian wild fruits are considered to be a great source of phytonutrients and natural antioxidant compounds. Our investigation showed how every fruit could be considered unique in terms of its phytonutrient content. The HPLC profile of BL revealed the presence of at least 11 different phenolic compounds, including vanillic acid, reported for the first time in this paper. Alongside vanillic acid, the

Fig. 2 Principal component analysis: scores plot of the first three principal components showing the sample clustering and loadings plot reflecting the influence of a particular parameter. Numbers correspond to the detected compound as given in Table [2](#page-5-0)

abundance of naringin was the highest among all the extracts investigated. EL extracts were unique due to the presence of arbutin, a well-known skin protector, while CC was characterized by syringic and gallic acids. PCA managed to classify berry fruit samples on the basis of variations in the content of individual flavonoids, phenolic acids and antioxidant capacity into four separate clusters. In conclusion, this work proved wild fruits to be an interesting field in the search for compounds with potential nutraceutical properties as they contain phytonutrients that could be used as potential benefits in blood sugar regulation and skin protection. However, additional bioactivity testing is needed, especially bearing in mind the synergistic effect among polyphenols as an important factor determining the functional properties.

Acknowledgements This work was supported by the Serbian Ministry of Education, Science and Technological Development (Projects 172017 and TR 31063). Partly, the research was supported by the CICOPS scholarship foreseen for research collaboration at the University of Pavia. The authors thank Prof. Milanese for her help in English revision.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Compliance with ethics requirements This article does not contain any studies with animal or human subjects.

References

- 1. Li Y, Zhang J-J, Xu D-P, Zhou T, Zhou Y, Li S, Li H-B (2016) Bioactivities and health benefits of wild fruits. Int J Mol Sci. [https](https://doi.org/10.3390/ijms17081258) [://doi.org/10.3390/ijms17081258](https://doi.org/10.3390/ijms17081258)
- 2. Mratinić E, Fotirić-Akšić M (2014) Indigenous fruit species as a significant resource for sustainable development. Bull Fac For 2:181–193
- 3. Barros L, Carvalho AM, Ferreira IC (2011) Comparing the composition and bioactivity of *Crataegus monogyna* flowers and fruits used in folk medicine. Phytochem Anal 22:181–188. [https://doi.](https://doi.org/10.1002/pca.1267) [org/10.1002/pca.1267](https://doi.org/10.1002/pca.1267)
- 4. Yang B, Liu P (2012) Composition and health effects of phenolic compounds in hawthorn (*Crataegus* spp.) of different origins. J Sci Food Agric 92:1578–1590.<https://doi.org/10.1002/jsfa.5671>
- 5. Zhang Y, Zhang L, Geng Y, Geng Y (2014) Hawthorn fruit attenuates atherosclerosis by improving the hypolipidemic and antioxidant activities in apolipoprotein E-deficient mice. J Atheroscler Thromb 21:119–128.<https://doi.org/10.5551/jat.19174>
- 6. Yuksel AK (2015) The effects of blackthorn (*Prunus Spinosa* L.) addition on certain quality characteristics of ice cream. J Food Qual 38:413–421.<https://doi.org/10.1111/jfq.12170>
- 7. Gironés-Vilaplana A, Valentaõ P, Moreno DA, Ferreres F, Garcıá-Viguera C, Andrade PB (2012) New beverages of lemon juice enriched with the exotic berries maqui, açaí, and blackthorn: bioactive components and in vitro biological properties. J Agric Food Chem 60:6571–6580.<https://doi.org/10.1021/jf300873k>
- 8. Veličković JM, Kostić DA, Stojanović GS, Mitić SS, Mitić MN, Ranđelović SS, Đorđević AS (2014) Phenolic composition,

antioxidant and antimicrobial activity of the extracts from *Prunus spinosa* L. fruit. Hemijska Industrija 68:297–303. [https://](https://doi.org/10.2298/HEMIND130312054V) doi.org/10.2298/HEMIND130312054V

- 9. Antolak H, Czyzowska A, Sakač M, Mišan A, Đuragić O, Kregiel D (2017) Phenolic compounds contained in littleknown wild fruits as antiadhesive agents against the beveragespoiling bacteria *Asaia* spp. Molecules 22:E1256. [https://doi.](https://doi.org/10.3390/molecules22081256) [org/10.3390/molecules22081256](https://doi.org/10.3390/molecules22081256)
- 10. Hosseinpour-Jaghdani F, Shomali T, Gholipour-Shahraki S, Rahimi-Madiseh M, Rafieian-Kopaei M (2017) *Cornus mas*: a review on traditional uses and pharmacological properties. J Complement Integr Med. [https://doi.org/10.1515/](https://doi.org/10.1515/jcim-2016-0137) [jcim-2016-0137](https://doi.org/10.1515/jcim-2016-0137)
- 11. Cornescu FC, Cosmulescu SN (2017) Morphological and biochemical characteristics of fruits of different cornelian cherry (*Cornus mas* L.) genotypes from spontaneous flora. Not Sci Biol 9(4):577–581.<https://doi.org/10.15835/nsb9410161>
- 12. Mratinić E, Fotirić M (2007) Selection of black elderberry (*Sambucus nigra* L.) and evaluation of its fruits usability as biologically valuable food. Genetika 39:305–314
- 13. Gray AM, Abdel-Wahab YHA, Flatt PR (2000) The traditional plant treatment, *Sambucus nigra* (elder), exhibits insulin-like and insulin-releasing actions in vitro. J Nutr 130:15–20. [https://doi.](https://doi.org/10.1093/jn/130.1.15) [org/10.1093/jn/130.1.15](https://doi.org/10.1093/jn/130.1.15)
- 14. Baraibar B, Willer H (2007) Classification of land use data. In: Willer H, Yussefi Y (eds) The world of organic agriculture statistics and emerging trend 2007, 9th edn. International Federation of Organic Agriculture Movements (IFOAM), Bonn, Germanny & Research Institute of Organic Agriculture (FiBL), Frick, Switzerland, pp 21–25
- 15. Mratinić E, Kojić M (1998) Indigenous species of Serbia (in Serbian). Beograd: Institit za istraživanja u poljoprivredi "Srbija", p 595
- 16. Tadić VM, Dobrić S, Marković GM, Ðorđević SM, Arsić IA, Menković NAR, Stević T (2008) Anti-inflammatory, gastroprotective, free-radical-cavenging, and antimicrobial activities of hawthorn berries ethanol extract. J Agric Food Chem 56:7700–7709. <https://doi.org/10.1021/jf801668c>
- 17. Rodrigues S, Calhelha RC, Barreira JC, Dueñas M, Carvalho AM, Abreu RM, Santos-Buelga C, Ferreira IC (2012) Crataegus monogyna buds and fruits phenolic extracts: growth inhibitory activity on human tumor cell lines and chemical characterization by HPLC–DAD–ESI/MS. Food Res Int 49:516–523. [https://doi.](https://doi.org/10.1016/j.foodres.2012.07.046) [org/10.1016/j.foodres.2012.07.046](https://doi.org/10.1016/j.foodres.2012.07.046)
- 18. Kao E, Wang CJ, Lin WL, Chu CY, Tseng TH (2007) Effects of polyphenols derived from fruit of *Crataegus pinnatifida* on cell transformation, dermal edema and skin tumor formation by phorbol ester application. Food Chem Toxicol 45:1795–1804. [https://](https://doi.org/10.1016/j.fct.2007.03.016) doi.org/10.1016/j.fct.2007.03.016
- 19. Long SR, Carey RA, Crofoot KM, Proteau PJ, Filtz TM (2006) Effect of hawthorn (*Crataegus oxycantha*) crude extract and chromatographic fractions on multiple activities in a cultured cardiomyocyte assay. Phytomedicine 13:643–650. [https://doi.](https://doi.org/10.1016/j.phymed.2006.01.005) [org/10.1016/j.phymed.2006.01.005](https://doi.org/10.1016/j.phymed.2006.01.005)
- 20. Demir F, Kalyoncu I (2003) Some nutritional, pomological and physical properties of cornelian cherry (*Cornus mas* L.). J Food Eng 60(3):335–341. [https://doi.org/10.1016/S0260](https://doi.org/10.1016/S0260.-8774(03)00056-6) [.-8774\(03\)00056-6](https://doi.org/10.1016/S0260.-8774(03)00056-6)
- 21. Seeram NP, Schutzki R, Chandra A, Nair MG (2002) Characterization, quantification, and bioactivities of anthocyanins in Cornus species. J Agric Food Chem 50(9):2519–2523. [https://doi.](https://doi.org/10.1021/jf0115903) [org/10.1021/jf0115903](https://doi.org/10.1021/jf0115903)
- 22. Dawidowicz AL, Wianowska D, Baraniak B (2006) The antioxidant properties of alcoholic extracts from *Sambucus nigra* L. (antioxidant properties of extracts). LWT 39:308–315. [https://doi.](https://doi.org/10.1016/j.lwt.2005.01.005) [org/10.1016/j.lwt.2005.01.005](https://doi.org/10.1016/j.lwt.2005.01.005)
- 23. Kaack K, Christensen LP, Hughes M, Eder R (2006) Relationship between sensory and volatile compounds of elderflower extracts. Eur Food Res Technol 223:57–70. [https://doi.org/10.1007/s0021](https://doi.org/10.1007/s00217-005-0122-y) [7-005-0122-y](https://doi.org/10.1007/s00217-005-0122-y)
- 24. Rigelsky JM, Sweet BVA (2002) Hawthorn: pharmacology and therapeutic uses. J Health Syst Pharm 59(5):417–422
- 25. Singleton VL, Orthofer R, Lamuela-Raventos RM (1999) Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin–Ciocalteu reagent. Meth Enzymol 299:152–178. [https://doi.org/10.1016/S0076-6879\(99\)99017-1](https://doi.org/10.1016/S0076-6879(99)99017-1)
- 26. Natić MM, Dabić D, Papetti A, Fotirić Akšić MM, Ognjanov V, Ljubojević M, Tešić ŽLj (2015) Analysis and characterisation of phytochemicals in mulberry (*Morus alba* L.) fruits grown in Vojvodina, North Serbia. Food Chem 171:128–136. [https://doi.](https://doi.org/10.1016/j.foodchem.2014.08.101) [org/10.1016/j.foodchem.2014.08.101](https://doi.org/10.1016/j.foodchem.2014.08.101)
- 27. Contreras-Guzman ES, Strong FC (1982) Determination of tocopherols (vitamin E) by reduction of cupric ion. JAOAC 65:1215–1222
- 28. Zhu LJ, Chen J, Tang XY, Xiong YL (2008) Reducing, radical scavenging, and chelation properties of in vitro digest of alcalase-treated zein hydrolysate. J Agric Food Chem 56:2714–2721. [https://doi.](https://doi.org/10.1021/jf703697e) [org/10.1021/jf703697e](https://doi.org/10.1021/jf703697e)
- 29. Yildirim A, Mavi A, Kara AA (2001) Determination of antioxidant and antimicrobial activities of *Rumex crispus* L. extracts. J Agric Food Chem 49:4083–4089.<https://doi.org/10.1021/jf0103572>
- 30. Sreejayan N, Rao MNA (1997) Nitric oxide scavenging by curcuminoids. J Pharm Pharmacol 49:105–107. [https://doi.](https://doi.org/10.1111/j.2042-7158.1997.tb06761.x) [org/10.1111/j.2042-7158.1997.tb06761.x](https://doi.org/10.1111/j.2042-7158.1997.tb06761.x)
- 31. Vanni A, Gastaldi D, Giunata G (1990) Kinetic investigations on the double enzymatic activity of the tyrosinase mushroom. Anal Chim 80:35–60
- 32. Ruíz-Rodríguez BM, de Ancos B, Sánchez-Moreno C, Fernández-Ruíz V, Sánchez-Mata MC, Cámara M, Tardío J (2014) Wild blackthorn (*Prunus spinosa* L.) and hawthorn (*Crataegus monogyba* Jacq.) fruits as valuable sources of antioxidants. Fruits 69:61–73. <https://doi.org/10.1051//fruits/2013102>
- 33. Bechkri S, Berrehal D, Semra Z, Bachari K, Kabouche A, Kabouche Z (2017) Composition and biological activities of seeds oils of two *Crataegus* species growing in Algeria. JMES 8:1023–1028
- 34. Tardio J, Pardo de Santayana M, Morales R (2006) Ethnobotanical review of wild edible plants in Spain. Bot J Linn Soc 152:27–71. <https://doi.org/10.1111/j.1095-8339.2006.00549.x>
- 35. Calvo MI, Akerreta S, Cavero RY (2013) The pharmacological validation of medicinal plants used for digestive problems in Navarra, Spain. Eur J Integr Med 5:537–546. [https://doi.org/10.1016/j.eujim](https://doi.org/10.1016/j.eujim.2013.07.002.538) [.2013.07.002.538](https://doi.org/10.1016/j.eujim.2013.07.002.538)
- 36. Mikulic-Petkovsek M, Samoticha J, Eler K, Stampar F, Veberic R (2015) Traditional elderflower beverages: a rich source of phenolic compounds with high antioxidant activity. J Agric Food Chem 63:1477–1487.<https://doi.org/10.1021/jf506005b>
- 37. Yilmaz KU, Ercisli S, Zengin Y, Sengul M, Kafkas EY (2009) Preliminary characterisation of Cornelian cherry (*Cornus mas* L.) genotypes for their physico-chemical properties. Food Chem 114:408–412.<https://doi.org/10.1016/j.foodchem.2008.09.055>
- 38. Hashempour A, Ghazvini RF, Bakhshi D, Ghasemnezhad M, Sharafti M, Ahmadian H (2010) Ascorbic acid, anthocyanins, and phenolic contents and antioxidant activity of ber, azarole, raspberry and cornelian cherry genotypes growing in Iran. Hortic Environ Biotechnol 51:83–88
- 39. Pantelidis GE, Vasilakakis M, Manganaris GA, Diamantidis G (2007) Antioxidant capacity, phenol, anthocyanin and ascorbic acid contents in raspberries, blackberries, red currants, gooseberries and cornelian cherries. Food Chem 102:777–783. [https://doi.](https://doi.org/10.1016/j.foodchem.2006.06.021) [org/10.1016/j.foodchem.2006.06.021](https://doi.org/10.1016/j.foodchem.2006.06.021)
- 40. Rezaei-Golmisheh A, Malekinejad H, Asri-Rezaei S, Farshid AA, Akbari P (2015) Hawthorn ethanolic acid extracts with triterpenoids and flavonoids exert hepatoprotective effects and suppress

 $\circled{2}$ Springer

the hypercholesterolemia—induced oxidative stress in rats. Iran J Basic Med Sci 18:691–699

- 41. Šamec D, Piljac-Žegarac J (2011) Postharvest stability of antioxidant compounds in hawthorn and cornelian cherries at room and refrigerator temperatures—Comparison with blackberries, white and red grapes. Sci Hortic 131:15–21. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.scienta.2011.09.021) [scienta.2011.09.021](https://doi.org/10.1016/j.scienta.2011.09.021)
- 42. Pinacho R, Yolanda R, Icíar C, Diana A, María A, Calvo I (2015) Phenolic compounds of blackthorn (*Prunus spinosa* L.) and influence of in vitro digestion on their antioxidant capacity. J Funct Foods 19:49–62. <https://doi.org/10.1016/j.jff.2015.09.015>
- 43. Mustapha N, Mokdad-Bzèouich I, Maatouk M, Ghedira K, Hennebelle T, Chekir-Ghedira L (2016) Antitumoral, antioxidant, and antimelanogenesis potencies of Hawthorn, a potential natural agent in the treatment of melanoma. Melanoma Res 26:211–222. [https://](https://doi.org/10.1097/CMR.0000000000000240) doi.org/10.1097/CMR.0000000000000240
- 44. Chai WM, Chen C-M, Gao Y-S, Feng H-L, Ding Y-M, Shi Y, Zhou HT, Chen Q-X (2014) Chinese hawthorn with potent antityrosinase and antioxidant activity. J Agric Food Chem 62:123–129. [https://](https://doi.org/10.1021/jf405385j) doi.org/10.1021/jf405385j
- 45. Kubo I, Kinst-Hori I, Chaudhuri SK, Kubo Y, Sánchez Y, Ogura T (2000) Flavonols from *Heterotheca inuloides*: tyrosinase inhibitory activity and structural criteria. Bioorg Med Chem 8:1749–1755. [https://doi.org/10.1016/S0968-0896\(00\)00102-4](https://doi.org/10.1016/S0968-0896(00)00102-4)
- 46. Cosmulescu S, Trandafir I, Nour V (2017) Phenolic acids and flavonoids profiles of extracts from edible wild fruits and their antioxidant properties. Int J Food Prop 20(12):3124–3134. [https://doi.](https://doi.org/10.1080/10942912.2016.1274906) [org/10.1080/10942912.2016.1274906](https://doi.org/10.1080/10942912.2016.1274906)
- 47. Radovanović BC, Milenković Anđelković AS, Radovanović AB, Anđelković MZ (2013) Antioxidant and antimicrobial activity of polyphenol extracts from wild berry fruits grown in Southeast Serbia. Trop J Pharm Res 12:813–819. [https://doi.org/10.4314/tjpr.](https://doi.org/10.4314/tjpr.v12i5.23) [v12i5.23](https://doi.org/10.4314/tjpr.v12i5.23)
- 48. Zuo Y, Wang C, Zhan J (2002) Separation, characterization and quantitation of benzoic and phenolic antioxidants in American cranberry fruit by GC-MS. J Agric Food Chem 50:3789–3794. [https://](https://doi.org/10.1021/jf020055f) doi.org/10.1021/jf020055f
- 49. Jurica K, Karaconji IB, Segan S, Opsenica DM, Kremer D (2015) Quantitative analysis of arbutin and hydroquinone in strawberry tree (*Arbutus unedo* L., Ericaceae) leaves by gas chromatographymass spectrometry. Arh Hig Rada Toksikol 66:197–202. [https://doi.](https://doi.org/10.1515/aiht-2015-66-2696) [org/10.1515/aiht-2015-66-2696](https://doi.org/10.1515/aiht-2015-66-2696)
- 50. Lee BC, Lee SY, Lee HJ, Sim GS, Kim JH, Kim JH, Cho YH, Lee DH, Pyo HB, Choe TB, Moon DC, Yun YP, Hong JT (2007) Anti-oxidative and photo-protective effects of coumarins isolated from *Fraxinus chinensis*. Arch Pharm Res 30:1293–1301. [https://](https://doi.org/10.1007/BF02980270) doi.org/10.1007/BF02980270
- 51. Alam MA, Subhan N, Rahman MM, Uddin SJ, Reza HM, Sarker SD (2014) Effect of citrus flavonoids, naringin and naringenin, on metabolic syndrome and their mechanisms of action. Adv Nutr 5:404–417.<https://doi.org/10.3945/an.113.005603>
- 52. Han X, Li W, Huang D, Yang X (2016) Polyphenols from hawthorn peels and fleshes differently mitigate dyslipidemia, inflammation and oxidative stress in association with modulation of liver injury in high fructose diet-fed mice. Chem Biol Interact 257:132–140. [https://doi.](https://doi.org/10.1016/j.cbi.2016.08.002) [org/10.1016/j.cbi.2016.08.002](https://doi.org/10.1016/j.cbi.2016.08.002)
- 53. Maatta-Riihinen KR, Kamal-Eldin A, Torronen AR (2004) Identification and quantification of phenolic compounds in berries of *Fragaria* and *Rubus* species (family Rosaceae). J Agric Food Chem 52:6178–6187.<https://doi.org/10.1021/jf049450r>
- 54. Wei X, Chen D, Yi Y, Qi H, Gao X, Fang H, Gu Q, Wang L, Gu L (2012) Syringic acid extracted from *Herba dendrobii* prevents diabetic cataract pathogenesis by inhibiting aldose reductase activity. Evid Based Complement Alternat Med. [https://doi.](https://doi.org/10.1155/2012/426537) [org/10.1155/2012/426537](https://doi.org/10.1155/2012/426537)