# REVIEW

# The D<sub>3</sub> dopamine receptor in cellular and organismal function

Received: 3 June 1997 / Final version: 15 August 1997

Abstract The D<sub>3</sub> dopamine receptor is a member of the family of D<sub>2</sub>-like dopamine receptors. Since the cloning and identification of the D<sub>3</sub> receptor in 1990. considerable progress has been made towards understanding the function of this novel site. Although some avenues of investigation have yielded more definitive results than others, studies to date indicate the  $D_3$ receptor is localized preferentially in limbic brain areas and affects locomotion and perhaps reinforcement and reward. A subpopulation of the receptors appear to be autoreceptors which modulate dopamine synthesis, release, and neuronal activity. These observations have led to the hypothesis that the  $D_3$  receptor may be an appropriate target in the treatment of neuropsychiatric disorders such as schizophrenia and drug addiction. The role of  $D_3$  sites in disease, however, remains to be established. Genetic association of D<sub>3</sub> receptor polymorphisms with neuropsychiatric disorders have been proposed. Alterations in expression of  $D_3$  sites may occur in some diseases. Although the study of this receptor is clearly in the early stages, these findings lay the foundation for future investigation. In this review, dopamine D<sub>3</sub> receptor brain localization, cellular signaling mechanisms, and associated behavior will be discussed. The potential role of the D<sub>3</sub> site in neuropsychiatric disorders and as a therapeutic target is also addressed.

Key words Limbic system · Signal transduction · Autoreceptor · Receptor binding · Behavior · Locomotor activity · Reward · Development · Schizophrenia · Parkinson's disease · Cocaine · Antipsychotic

R.A. Shafer · B. Levant (🖂)

Department of Pharmacology, Toxicology, and Therapeutics, University of Kansas Medical Center, Kansas City, KS 66160-7417, USA

#### Introduction

Dopamine receptors are of considerable interest, as they are the principal target of drugs employed in the treatment of neuropsychiatric disorders such as schizophrenia and Parkinson's disease. Until 1990, the dopamine receptor population in the brain and periphery was believed to consist of two subtypes,  $D_1$  and  $D_2$ (for review, see Seeman and Grigoriadis 1987). Currently, the  $D_1$  and  $D_2$  subtypes represent families of dopamine receptors. Several additional low abundance dopamine receptors were identified following the cloning of the  $D_1$  and  $D_2$  receptors (for review, see Sibley et al. 1993). These novel subtypes include the  $D_3$  and  $D_4$  receptors, which are similar to  $D_2$ , and the  $D_5$  receptor, which is similar to  $D_1$ . The  $D_3$  receptor was initially cloned from a rat cDNA library by Sokoloff and colleagues (1990) using probes derived from the  $D_2$  dopamine receptor sequence. The cloning of the human D<sub>3</sub> receptor was reported shortly thereafter (Giros et al. 1990), followed by the murine  $D_3$ receptor (Fishburn et al. 1993). Due to the limbic location of this receptor, D<sub>3</sub> receptors have been postulated as a therapeutic target in the treatment of schizophrenia and drug abuse.

Although still in the early stages, considerable progress has been made in the study of the  $D_3$  receptor. This article reviews the progress made to date in assessing the neurobiological role of this novel receptor. Its relevance in disease and as a potential therapeutic target are also discussed.

# **Molecular** biology

Based on amino acid sequence and gene organization, the  $D_3$  receptor has been classified as a member of the family of  $D_2$ -like dopamine receptors. Unlike genes for the  $D_1$ -like receptors which do not contain introns, the

Fax (+1) 913-588-7501, e-mail: blevant@kumc.edu

D<sub>2</sub>, D<sub>3</sub>, and D<sub>4</sub> receptor genes contain discontinuous gene sequences. Also, in contrast with the D1-like receptors, which have short third intracellular domains and long carboxy termini, the D<sub>2</sub>-like receptors have relatively long third intracellular domains and short carboxy termini. The D<sub>2</sub>-like receptors possess moderate sequence homology with the  $D_1$ -like receptors. For example, the rat  $D_1$  and  $D_2$  receptors possess only 41%homology in the transmembrane domains (Monsma et al. 1990). In comparison, the overall homology between the  $D_2$  and  $D_3$  receptor is 52%, but increases to 75% when only the transmembrane spanning domains are considered. The D<sub>2</sub> and D<sub>3</sub> receptors exhibit 39% and 41% overall homology with the D<sub>4</sub> receptor, respectively (Van Tol et al. 1991). The rat and human D<sub>3</sub> receptors contain 446 and 400 amino acids, respectively, and exhibit 78% homology (Sokoloff et al. 1990; Giros et al. 1990) (Table 1). Although biochemical evidence for the secondary structure of this receptor has yet to be generated, hydrophobicity analysis indicates that the most probable secondary structure of the D<sub>3</sub> receptor is consistent with those of the seven transmembrane spanning, G-protein coupled receptors.

In addition to aiding in the detection of novel receptor subtypes, the use of molecular methods has enabled the elucidation of receptor splice-variants. For example, the D<sub>2</sub> receptor exists in two functional alternate splice variants D<sub>2L</sub> and D<sub>2S</sub>, which vary in the length of the third intracellular loop (Dal Toso et al. 1989; Giros et al. 1989). The murine  $D_3$  dopamine receptor undergoes alternative RNA splicing to produce two mRNA species ( $D_{3L}$  and  $D_{3S}$ ) which differ by 63 bp in the putative third intracellular loop (Fishburn et al. 1993). An internal acceptor site is used for the  $D_3$ dopamine receptor splicing mechanism; this is different from the existence of a separate exon, as occurs for  $D_2$ receptor splicing. Alternate splice variants of the third intracellular loop have yet to be detected in the rat or human; however, several truncated isoforms of the D<sub>3</sub> receptor have been reported (Giros et al. 1991; Snyder et al. 1991; Nagai et al. 1993; Pagliusi et al. 1993;

Table 1 Characteristics of the D3 dopamine receptor

Amino acids Human	400
Rat	446
Homology with D <sub>2</sub> receptor (%) D <sub>4</sub> receptor (%) Rat D <sub>3</sub> with human D <sub>3</sub> receptor (%)	52 41 78
Brain distribution	Olfactory tubercle Islands of Calleja Nucleus accumbens
Putative selective agonists	7-OH-DPAT PD 128907
Putative selective antagonists	Nafadotride (+)-S-14297 U-99194A

Schmauss et al. 1993; Griffon et al. 1996). Although the functional properties of only some of the truncated  $D_3$  receptor variants have been assessed, all of those tested were found to lack binding activity in transfected cell lines (Giros et al. 1991; Schmauss et al. 1993). It is likely that the other truncated forms are also nonfunctional. The physiological role of these receptor variants is also unclear. It is speculated that the truncated receptors could be expressed under certain circumstances as a mechanism for controlling the density of functional  $D_3$  sites or might occur in certain disease states (Giros et al. 1991).

#### Localization of D<sub>3</sub> receptors in brain

#### Distribution in rat brain

Although present in significantly lower levels than  $D_1$ or D<sub>2</sub> receptor mRNAs, in situ hybridization studies in the rat brain demonstrate that mRNA for the  $D_3$ receptor appears to be expressed preferentially in limbic brain regions. The highest density is reported in the islands of Calleja where D<sub>3</sub> mRNA is expressed by granule cells (Diaz et al. 1995). High levels of D<sub>3</sub> mRNA are also observed in the nucleus accumbens and olfactory tubercles (Sokoloff et al. 1990; Bouthenet et al. 1991; Mengod et al. 1992; Landwehrmeyer et al. 1993a; Curran and Watson 1995; Diaz et al. 1995). Additional brain regions reported to exhibit dense expression of D<sub>3</sub> mRNA include the medial division of the bed nucleus of the stria terminalis, the nucleus of the vertical limb of the diagonal band of Broca, the paracentral thalamic nucleus, the medial and ventral lateral geniculate nuclei, the magnocellular preoptic nucleus, mammallary nucleus, the lateral part of the substantia nigra pars compacta, the dorsal cochear nucleus, and the Purkinje cell layer of the vestibulocerebellum (Bouthenet et al. 1991; Mengod et al. 1992; Diaz et al. 1995).

Moderately dense levels of  $D_3$  mRNA expression have been observed in areas such as the ventral pallidum, the amygdala, the nucleus of the horizontal limb of the diagonal band of Broca, certain thalamic and hypothalamic nuclei, the superior colliculus, and the inferior olivary nucleus (Bouthenet et al. 1991). Moderate to dense expression of  $D_3$  mRNA has also been reported in the dentate gyrus, olfactory bulb and the anterior and intermediate lobes of the pituitary in in situ hybridization studies (Bouthenet et al. 1991); however, no  $D_3$  mRNA was detected in these brain regions in other studies (Sokoloff et al. 1990; Mengod et al. 1992).

Low densities of  $D_3$  mRNA have been reported in the cerebral cortex, caudate/putamen, ventral pallidum, substantia nigra pars reticulata, ventral tegmental area, and cerebellar cortex (Bouthenet et al. 1991; Mengod et al. 1992).

The distribution of D<sub>3</sub> receptors in rat brain has not been mapped in detail. As the pharmacological specificity of radioligands remains to be extensively validated, data obtained to date must be considered to indicate "putative" D<sub>3</sub> sites. With these limitations in mind, the localization of "putative" D<sub>3</sub> binding sites appears to parallel the distribution of D<sub>3</sub> mRNA. D<sub>3</sub> receptors appear to be expressed in highest density in brain regions such as the islands of Calleja, olfactory bulb, and the pituitary intermediate lobe. Moderately dense D<sub>3</sub> binding is observed in the nucleus accumbens, the molecular layer of the vestibulocerebellum, and substantia nigra pars compacta. Relatively little D<sub>3</sub> binding is observed in the caudate/ putamen (Gehlert et al. 1992; Lévesque et al. 1992; Landwehrmeyer et al. 1993a; Levant et al. 1993; Parsons et al. 1993; Booze and Wallace 1995). D<sub>3</sub>-like immunoreactivity was associated with neuronal-type cells and was concentrated at the cell body perimeter (Ariano and Sibley 1994; Larson and Ariano 1995).

#### Distribution in human brain

Although not as extensively characterized, the distribution of  $D_3$  mRNA in the human brain appears to be generally similar to that observed in the rat. Enrichment of  $D_3$  mRNA was observed in the nucleus accumbens and islands of Calleja with relatively low levels of expression in the anterior caudate and putamen (Landwehrmeyer et al. 1993b).  $D_3$  mRNA has also been observed in the granular cell layer of the dentate gyrus (Meador-Woodruff et al. 1994).

The distribution of "putative"  $D_3$  receptors in human brain is generally similar to that observed in the rat; however, the overall pattern of distribution appears to be somewhat less restricted (Herroelen et al. 1994). Highest densities of putative  $D_3$  binding sites are reported in the nucleus accumbens and islands of Calleja (Murray et al. 1994; Hall et al. 1996). Moderate amounts of  $D_3$  binding were observed in the basal ganglia, parietal, temportal and occipital cortex, and cerebellar cortex, followed by substantia nigra, hippocampus, and the basolateral, lateral and basomedial amygdaloid nuclei (Herroelen et al. 1994; Murray et al. 1994; Lahti et al. 1995).  $D_3$  receptors were also detected in moderate density in the pituitary, with somewhat greater labeling in the posterior lobe than the anterior (Herroelen et al. 1994).

# **Cellular signaling mechanisms**

Signal transduction in expression systems

Determining the signal transduction pathways associated with the dopamine receptor in vivo was initially problematic because of the lack of selective pharmacological tools. The expression of the  $D_3$  receptor in transfected cell lines was the primary means for studying the functional properties of this receptor. However, host cells come from a variety of sources and may not express the appropriate G proteins or effectors that are physiologically relevant for the receptor. Thus, data obtained from the same receptor in different cell lines may be different depending on the endogenous signaling molecules expressed by the cells (for review, see Kenakin 1996). Accordingly, it is not surprising that reports on the coupling of the D<sub>3</sub> receptor to signal transduction systems have varied considerably. The initial cloning report indicated that the D<sub>3</sub> receptor expressed in Chinese hamster ovary (CHO) cells did not exhibit a decrease in affinity for agonists in the presence of guanyl nucleotides, or G-shift, as would be

Cell line	Effector system	Reference
CCL1.3	⇔ Adenylate cyclase	MacKenzie et al. (1994)
	⇔ Adenylate cyclase	Tang et al. (1994)
CHO 10001	$\downarrow$ Adenylate cyclase	Chio et al. (1994)
	Na <sup>+</sup> /H <sup>+</sup> exchange	Chio et al. (1994)
CHO-K1	$\Leftrightarrow$ Adenylate cyclase	MacKenzie et al (1994)
СНО	$\Leftrightarrow$ arachadonic acid release	Freedman et al. (1994)
	Na <sup>+</sup> /H <sup>+</sup> exchange	Boyfield et al. (1996)
GH <sub>4</sub> C <sub>1</sub>	$\Leftrightarrow$ K <sup>+</sup> current	Seabrook et al. (1992)
	⇔ Adenylate cyclase	Seabrook et al. (1992)
	$\Leftrightarrow$ arachadonic acid release	Seabrook et al. (1992)
	$\Leftrightarrow Ca^{2+}$ current	Seabrook et al. (1992)
MN9D	⇔ Adenylate cyclase	Tang et al. (1994)
NG108-15	↓ Adenylate cyclase	Freedman et al. (1994)
	$\downarrow$ Ca <sup>2+</sup> current	Freedman et al. (1994)
	⇔ Adenylate cyclase	Pilon et al. (1994)
		Pilon et al. (1994)
SK-N-MC	$\Leftrightarrow$ Adenylate cyclase	Mackenzie et al. (1994)
Xenopus melanophores	↓ Adenylate cyclase	Potenza et al. (1994)
293	↓ Adenylate cyclase	McAllister et al. (1995)
C6 glioma	Na <sup>+</sup> /H <sup>+</sup> exchange	Cox et al. (1995)

**Table 2** Effect of  $D_3$  dopaminreceptor stimulation on signaltransduction pathways

expected for a G-protein-coupled receptor (Sokoloff et al. 1990). This suggested that the  $D_3$  receptor might not be functionally coupled to G-proteins. Other groups studying the receptor expressed in other cell lines, including neuronal mesencephalic MN9D cells, neuroblastoma NG108-15 cells, and insect Sf21 cells, observed a similar lack of G-shift in D<sub>3</sub> binding (Freedman et al. 1994; Tang et al. 1994a; Woodcock et al. 1995). G-shifts in D<sub>3</sub> receptor binding, however, were observed in a number of studies using a variety of other cell lines. Interestingly, the magnitude of the decrease in agonist affinity observed in the presence of guanyl nucleotides ranged from 5- to 10-fold (Seabrook et al. 1992; Sokoloff et al. 1992; Chio et al. 1994; MacKenzie et al. 1994) to 50- to 100-fold, similar to the roughly 100-fold shifts observed for the D<sub>2</sub> receptor (Grigoriadis and Seeman 1985; Castro and Strange 1993; Pilon et al. 1994).

Observations on the coupling of the  $D_3$  receptor in expression systems to specific signal transduction cascades have also varied (Table 2). In some systems, a G-shift in D<sub>3</sub> binding was observed, but alterations in second messengers such as cAMP, phosphoinositides, or arachidonic acid were not detected (Seabrook et al. 1992; MacKenzie et al. 1994). Other groups observed a variety of D<sub>3</sub>-initiated signaling events, including stimulation or inhibition of adenylyl cyclase, increased extracellular acidification, alterations in Ca<sup>2+</sup> and K<sup>+</sup> currents, and induction of c-fos expression (Chio et al. 1994; Pilon et al. 1994; Potenza et al. 1994; Seabrook et al. 1994; Cox et al. 1995; Griffon et al. 1996, 1997; Liu et al. 1996; Werner et al. 1996). D<sub>3</sub> receptors have also been shown to induce aggregation in melanocytes (Potenza et al. 1994) and to initiate mitogenesis in CHO cells (Chio et al. 1994; Pilon et al. 1994; Pugsley et al. 1995) and NG108-15 glioma cell lines (Griffon et al. 1997). Several of the D<sub>3</sub>-mediated signaling events, including stimulation of adenylyl cyclase, mitogenesis, alterations in  $Ca^{2+}$  and  $K^+$  currents, and increased rate of extracellular acidification were blocked by pertussis toxin, suggesting coupling to a Gi or Go isoform (Chio et al. 1994; Pilon et al. 1994; Potenza et al. 1994; Seabrook et al. 1994; Liu et al. 1996; Werner 1996). In another study, however, D<sub>3</sub>-induced increases in the rate of extracellular acidification were not blocked by pertussis toxin (Cox et al. 1995). Thus, these studies demonstrate functional coupling of the D<sub>3</sub> receptor to a variety of signaling cascades in some expression systems. The cellular signaling pathways affected, however, vary depending on the host cell and may not necessarily reflect the signaling pathways associated with the receptor in brain.

# Coupling in brain

While coupling of the  $D_3$  receptor has been shown in some expression systems, signaling pathways associated

with the receptor in brain have been more difficult to determine. Although all of the D<sub>3</sub>-selective radioligands identified to date, such as [<sup>3</sup>H]7-OH-DPAT (7-hydroxy-diphenylaminotetralin), <sup>[3</sup>H]PD 128907,  $[^{125}I]$ 7-OH-PIPAT ((R)-trans-7-hydroxy-2-[Nand propyl-N-(3'-iodo-2'-propenyl)aminoltetralin), have been agonists, which presumably label the high affinity state of a G-protein coupled receptor, most studies indicate that the binding of these ligands at putative  $D_3$ sites is insensitive to guanyl nucleotides (Lévesque et al. 1992; Burris et al. 1994; Kung et al. 1994; Bancroft et al. 1997). In fact, the binding of several non-selective D<sub>2</sub>-like receptor agonists remaining in the presence of guanyl nucleotides has been suggested to represent labeling of D<sub>3</sub> sites in autoradiographic studies (Gehlert 1992; Levant et al. 1993; Kung et al. 1994). One study, however, has reported the inhibition of [<sup>3</sup>H]7-OH-DPAT by guanyl nucleotides and the sulfhydryl alkylating agent N-ethylmaleimide, indicating G-protein coupling (Liu et al. 1994c). This observation, however, is most likely the result of the non-selective labeling of both  $D_2$  and  $D_3$  sites due to the presence of  $Mg^{2+}$  in the assay buffer (see below).

While these findings may suggest that the D<sub>3</sub> receptor in brain may lack functional G-protein coupling, there are other possible explanations. Whereas the high affinity state of the D<sub>2</sub> receptor exhibits about 100fold higher affinity for agonists than the low affinity state, the high affinity conformation of the cloned  $D_3$ receptor in expression systems has been most often reported to exhibit only about 5- to 10-fold higher affinity for agonists than the low affinity state. Thus, while agonist radioligands are presumed preferentially to label the high affinity conformation of G-protein coupled receptors, the putative D<sub>3</sub> binding observed in brain, albeit of nanomolar affinity, may be to receptors in the low affinity state. As such, the binding of either agonist or antagonist ligands to these sites would be unaltered in the presence of guanyl nucleotides.

There are several possible reasons why the observed  $D_3$  binding in brain tissue may represent receptors in the low affinity state. The first and simplest explanation is that the affinity state of these sites is a function of the in vitro assay conditions used to obtain putatively selective labeling of D<sub>3</sub> sites. Specifically, obtaining selective labeling of the D<sub>3</sub> site with the radioligands currently available appears to be dependent on the use of assay conditions which disfavor agonist binding at the  $D_2$  site. The greatest  $D_3/D_2$  selectivity for these ligands has been obtained in the absence of  $Mg^{2+}$  and the presence of EDTA (Lévesque et al. 1992; Akunne et al. 1995) in concordance with previous studies indicating that the high affinity agonist state of D2-like receptors is not favored in the absence of  $Mg^{2+}$  (Sibley and Creese 1983). Although these conditions may also affect the affinity state of the D<sub>3</sub> receptor, the low affinity conformation of the D<sub>3</sub> site exhibits much higher affinity for agonists than the low affinity state of the  $D_2$  receptor. As such, selective labeling of  $D_3$  sites occurs.

Alternatively, D<sub>3</sub> sites in rat brain may exist predominantly in the low affinity state under basal conditions as has been suggested for the  $D_1$  receptor (Richfield et al. 1989). Although "D3-selective" radioligands, such as [<sup>3</sup>H]7-OH-DPAT, label a single putative D<sub>3</sub> site in rat brain (Lévesque et al. 1992; Akunne et al. 1995; Levant 1995), depletion of endogenous catecholamines resulted in the detection of an additional [<sup>3</sup>H]7-OH-DPAT binding site ex vivo. This additional binding site exhibited roughly 10-fold higher affinity than the single binding site detected in control animals without a significant increase in the total number of sites (Levant 1995). Although the higher affinity sites may have been occupied, and thus masked, by endogenous dopamine in control animals (Schotte et al. 1992, 1996), preincubation and extensive washing of membranes from control animals to remove any residual dopamine failed to alter binding of [<sup>3</sup>H]7-OH-DPAT (Levant 1995). These observations suggest that in the absence of dopamine, some D<sub>3</sub> sites, which under normal conditions are predominantly in the low affinity state, assume a high affinity conformation. This hypothesis is supported by the observation that the high affinity component of [<sup>3</sup>H]7-OH-DPAT binding in catecholamine-depleted rats is inactivated by the alkylating agent 1-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ), while the low affinity component is not, a difference that might well be conferred by a conformational change. In fact, when catecholamine-depleted rats are treated with EEDQ, only low affinity sites remain and the density of these sites is reduced by roughly the same amount as the density of high affinity sites present in the catecholaminedepleted animals (Levant 1995).

Although coupling to classical second messengers has yet to be demonstrated for the  $D_3$  receptor in the brain, modulation of expression of the transcription factor *c-fos* has been observed following stimulation or blockade of the receptor. 7-OH-DPAT and quinpirole attenuated clozapine-induced Fos expression in the nucleus accumbens, suggesting the possible involvement of dopamine D3 receptors (Vahid-Ansari and Robertson 1996). U-99194A, a D<sub>3</sub>-preferring antagonist, produced an increase in *c-fos* mRNA expression in the medial prefrontal cortex (Merchant et al. 1996). Similarly, another D<sub>3</sub>-preferring antagonist, *l*-nafadotride, induced Fos protein expression in the nucleus accumbens and olfactory tubercles (Shafer and Levant 1997). However, no induction of c-fos mRNA was observed after treatment with the selective  $D_3$ receptor antagonist GR 103691 (Hurley et al. 1996b). Further study is required to establish the exact role D<sub>3</sub> receptors play in the induction of Fos protein expression.

# The $D_3$ receptor as an autoreceptor

#### Anatomical studies

One of the issues of interest regarding the  $D_3$  receptor is whether, like the  $D_2$  site, this receptor is localized pre- or postsynaptically. The detection of  $D_3$  mRNA in the substantia nigra and ventral tegmental areas and putative binding sites in dopaminergic terminal fields suggests that a subset of  $D_3$  receptors may be presynaptic. In keeping with this hypothesis, unilateral dopaminergic lesions produced a marked decrease in  $D_3$  receptor density in the nucleus accumbens, suggesting the loss of presynaptic sites (Lévesque et al. 1995). While this observation could also result from a decrease in postsynaptic sites, Tepper et al. (1997) also report a marked decrease in  $D_3$  receptor binding in the substantia nigra in rats treated with  $D_3$  receptor antisense oligonucleotides.

#### Neurochemical studies

In vivo and in vitro neurochemical studies also suggest a role for the  $D_3$  site as an autoreceptor that modulate dopaminergic activity. Stimulation of D<sub>3</sub> receptors expressed in neuronal mesencephalic MN9D cells resulted in a dose-dependent inhibition of dopamine release (Tang et al. 1994b). Likewise, the D<sub>3</sub>-preferring agonist 7-OH-DPAT produced a decrease in dopamine release in vivo as assessed by microdialysis or voltametry, as well as in accumbal slice preparations (Damsma et al. 1993; Rivet et al. 1994; Yamada et al. 1994; Devoto et al. 1995; Gilbert et al. 1995; Patel et al. 1995; Gainetdinov et al. 1996). Similar effects were also reported for PD 129807, another D<sub>3</sub> receptor-preferring agonist (Pugsley et al. 1995; Gobert et al. 1996). Both 7-OH-DPAT and PD 128907 have also been shown to decrease extracellular DOPAC concentrations as assessed by in vivo microdialysis, consistent with a decrease in dopamine release (Pugsley et al. 1995; Gainetdinov et al. 1996). In addition, mice deficient in  $D_3$  sites resulting from a targeted mutation of the  $D_3$ receptor gene, or "knockout" animals, exhibited higher basal levels of extracellular dopamine (Cooper et al. 1996). It is difficult, however, to ascertain that the inhibition of dopamine release observed in heterogeneous tissues results from selective actions at the D<sub>3</sub> receptor; as stimulation of D<sub>2</sub> receptors expressed in MN9D cells also inhibits dopamine release (Tang et al. 1994b). Likewise, care must be taken in the interpretation of data obtained with "knockout" animals because of the considerable plasticity in the developing nervous system, compensation changes may occur such as the expression of other receptors in place of the original D<sub>3</sub> receptor. It must also be noted that similar inhibitory responses to PD 128907 were observed for both D<sub>3</sub> "knockout" and wild-type mice (Cooper et al. 1996).

The D<sub>3</sub> receptor has also been implicated in the modulation of dopamine synthesis. In D<sub>3</sub>-expressing MN9D cells, the application of agonist produced a decrease in K<sup>+</sup>-stimulated tyrosine hydroxylase activity, the enzyme responsible for dopamine synthesis (O'Hara et al. 1996). In vivo, the D<sub>3</sub>-selective agonists 7-OH-DPAT and PD 128907 have been reported to decrease dopamine synthesis (Aretha et al. 1995: Gobert et al. 1995; Pugsley et al. 1995; Gainetdinov et al. 1996). This effect appears to be presynaptic, as it is observed in both normal rats and in rats treated with y-butyrolactone, which blocks impulse flow in nigrostriatal and mesolimbic dopamine neurons (Aretha et al. 1995; Pugsley et al. 1995). The involvement of the D<sub>3</sub> receptor in this effect is supported by the observation that 7-OH-DPAT produced a greater decrease in dopamine synthesis in the nucleus accumbens, where  $D_3$  sites are relatively abundant, than in the caudate nucleus where  $D_3$  sites are sparse (Aretha et al. 1995). A preliminary report by Nissbrandt et al. (1995) also suggests that reduction in the density of  $D_3$  sites by intracerebroventricular infusion of antisense oligonucleotides for the D3 receptor may result in increased DOPA accumulation, indicating a possible increase in dopamine synthesis. However, a preliminary report on D<sub>3</sub> receptor deficient mice indicated no alteration in dopamine synthesis compared to wild-type animals (Cooper et al. 1996). Taken together, these observations suggest a potential role for the  $D_3$  receptor as an autoreceptor modulating dopamine release and/or synthesis. Further study, however, must confirm the role of specific dopamine receptor subtypes in these observations.

#### Electrophysiological studies

The D<sub>3</sub>-preferring agonist 7-OH-DPAT has been shown to inhibit firing of neurons in both the substantia nigra and ventral tegmental areas as well as in brain slice preparations by activation of an 85 pS K<sup>+</sup> channel (Bowery et al. 1994; Liu et al. 1994a; Devoto et al. 1995; Kreiss et al. 1995; Lejeune and Millan 1995). Although selective action of 7-OH-DPAT at D<sub>3</sub> receptors cannot be assumed in these studies, Kreiss et al. (1995) have shown that the potencies of ten dopamine agonists in inhibiting firing of neurons in the substantia nigra pars compacta correlated with their affinities at  $D_3$ , but not  $D_2$  receptors. Caution, of course, must be exercised in the interpretation of such findings in view of the significant variability in the in vitro pharmacological profile of the  $D_2$  and  $D_3$  sites in various assay systems (see below). Recent electrophysiological data demonstrates, by D<sub>3</sub> receptor antisense knockout, that dopaminergic neurons possess functional D<sub>3</sub> autoreceptors at their axonal terminal and somatodentritic regions (Tepper et al. 1997). D<sub>3</sub> antisense knockouts produced an attenuation of the inhibitory

response to apomorphine, decreased  $[^{3}H]$ 7-OH-DPAT binding in the substantia nigra, and exhibited spontaneous contralateral rotations which suggest an increased dopamine release. D<sub>3</sub> autoreceptors may modulate terminal excitability and presumably the synthesis/release of dopamine from nerve terminals. On the somatodendrites, D<sub>3</sub> autoreceptors may play a role in modulating the excitability of local dendritic regions.

#### **Mediation of drug effects**

Radioligand binding studies

The  $D_3$  receptor is of interest particularly because of its relatively restricted distribution in brain. Unlike the  $D_2$  receptor, which is abundant in the caudate/putamen and pituitary as well as in limbic brain regions (for review, see Levant 1996), very low levels of expression of the  $D_3$  receptor are detected in either the caudate/putamen or pituitary, brain areas associated with the untoward neurological and endocrine effects produce by most conventional antipsychotics. These observations suggest that the  $D_3$  receptor, alone or in conjunction with other receptors, may be a target for novel antipsychotic drugs which might be free of extrapyramidal and neuroendocrine effects (Sokoloff et al. 1990).

Considering the extensive homology between the  $D_2$ and D<sub>3</sub> sites, it is not unexpected that the pharmacological profile of the D<sub>3</sub> receptor is generally similar to that of the  $D_2$  receptor. As such, the  $D_3$  receptor exhibits high affinity for non-selective and D<sub>2</sub>-selective agonists, such as dopamine, quinpirole, and apomorphine, and significantly lower affinity for the D<sub>1</sub>-selective agonist SKF 38393 (Sokoloff et al. 1990). The D<sub>3</sub> site also possess significantly higher affinity for D<sub>2</sub>selective antagonists, such as spiperone and haloperidol, than the D<sub>1</sub>-selective antagonist SCH 23390 (Freedman et al. 1994). Likewise, the D<sub>3</sub> receptor exhibits stereospecificity with higher affinity for (+)butaclamol than (-)-butaclamol and (-)-sulpiride than (+)-sulpiride (Freedman et al. 1994; Kula et al. 1994; MacKenzie et al. 1994).

A number of studies have also examined the relative affinities of dopaminergic compounds for  $D_2$  and  $D_3$ receptors in various expression systems and in brain. These studies suggest that some dopaminergic agonists, such as dopamine and quinpirole, possess higher affinity for the  $D_3$  site whereas antagonists, such as haloperidol, spiperone, and domperidone have higher affinity for  $D_2$  (Sokoloff et al. 1990). However, results of these studies vary considerably, depending, at least in part, on the expression system or tissue, the radioligand, and the in vitro assay conditions used (Tang et al. 1994a; Burris et al. 1995; Levant et al. 1995). For example, quinpirole was found to have more than

100-fold higher affinity for the  $D_3$  receptor than the  $D_2$ receptor in some assay systems (Sokoloff et al. 1990; Lèvesque et al. 1992; Burris et al. 1995) but roughly equal affinity for these sites in others (Levant and DeSouza 1993; Tang et al. 1994a; Burris et al. 1995). Similar variation in  $D_2/D_3$  selectivity in vitro has been observed for several other agonists including dopamine, quinelorane, 7-OH-DPAT, pramipexole, PD 128907 and 7-trans-OH-PIPAT (Sokoloff et al. 1990, 1992; Levant and DeSouza 1993; Chumpradit et al. 1994; Freedman et al. 1994; Kula et al. 1994; MacKenzie et al. 1994; Tang et al. 1994a; Burris et al. 1995; Levant et al. 1995; Lévesque et al. 1995; Mierau et al. 1995; Pugsley et al. 1995; Sautel et al. 1995a; Flietstra and Levant 1996). In fact, the high affinity state of the  $D_2$  receptor appears to have similar affinity for agonists as the  $D_3$  site (Burris et al. 1995). As such, the observed D<sub>3</sub>-selectivity of many agonists may have resulted from the use of in vitro conditions which disfavor the high affinity conformation of the D<sub>2</sub> receptor, such as the inclusion of  $Na^+$  in in vitro assay systems used for benzamide radioligands (Grigoriadis and Seeman 1985; Burris et al. 1995; Levant et al. 1995). In accordance with this hypothesis, the  $D_2/D_3$  selectivity of antagonists has exhibited much less variability between studies. Compounds such as domperidone, haloperidol, spiperone, chlorpromazine, clozapine and risperidone exhibit slight to substantial selectivity for the  $D_2$  receptor over the  $D_3$  while others, such as (-)sulpiride, amisulpiride, (+)-UH 232 exhibit little selectivity between these sites (Sokoloff et al. 1990, 1992; Levant and DeSouza 1993; Malmberg et al. 1993; Freedman et al. 1994; Kula et al. 1994; MacKenzie et al. 1994; Tang et al. 1994a; Burris et al. 1995; Levant et al. 1995; Mierau et al. 1995; Millan et al. 1995b; Pugsley et al. 1995; Sautel et al. 1995a; Flietstra and Levant 1996). Several antagonist possessing modest selectivity for the  $D_3$  receptor over the  $D_2$  have been identified, including l-nafadotride, (+)-S-14297, U-99194A and GR103691 (Waters et al. 1993; Sautel et al. 1995b; Millan et al. 1995; Flietstra and Levant 1996). What is clear from these studies is that under certain conditions, a number of compounds exhibit significant selectivity between the D<sub>2</sub> and D<sub>3</sub> dopamine receptors. This information is likely to be of considerable utility in the design and interpretation of in vitro studies, particularly for the determination of the localization and density of  $D_3$  sites. On the other hand, the attribution of in vivo pharmacological effects of these drugs to specific receptor subtypes based on these data is, in most instances, premature.

# Functional assays

Several functional assays have established the agonist or antagonist activity of a variety of dopaminergic compounds at the  $D_3$  receptor.  $D_2$  agonists, such as dopamine, quinpirole, and bromocriptine, have been shown to possess agonist activity at the D<sub>3</sub> receptor as assessed by the induction of CHO cell mitogenesis, melanocyte aggregation, or extracellular acidification (Chio et al. 1994; Pilon et al. 1994; Potenza et al. 1994; Sautel et al. 1995a; Boyfield et al. 1996). The putatively D<sub>3</sub>-selective compounds 7-OH-DPAT and PD 128907 also exhibit agonist activity in the mitogenesis test (Chio et al. 1994; Pilon et al. 1994; Potenza et al. 1994; Pugsley et al. 1995; Sautel et al. 1995). In contrast, antagonists, such as spiperone, (±)-sulpiride, (+)-S-14297 and nafadotride, block agonist-induced activity in these tests (Potenza et al. 1994; Sautel et al. 1995b; Gobert et al. 1996). The  $D_2/D_3$  ligand (+)-UH 232 has been shown to be a partial agonist at the  $D_3$  receptor in the mitogenesis assay (Griffon et al. 1995).

In addition to elucidating the agonist or antagonist activity of compounds at the  $D_3$  receptor, the assays described above are also useful in determining the  $D_2/D_3$ -selectivity of various compounds. In contrast to the significant D<sub>3</sub>-selectivity reported in some binding studies, the agonists tested, including dopamine, quinpirole, and 7-OH-DPAT, exhibited only modest, if any, D<sub>3</sub>-selectivity in the mitogenesis, melanocyte aggregation, or extracellular acidification assays (Chio et al. 1994; Pilon et al. 1994; Potenza et al. 1994; Sautel et al. 1995a; Boyfield et al. 1996). In contrast, the antagonists spiperone and  $(\pm)$ -sulpiride were roughly 65-fold more potent in inhibiting agonist-induced melanocyte aggregation at  $D_2$  receptors than  $D_3$  (Potenza et al. 1994). These observations further underscore the need of caution in the use of in vitro binding data in the interpretation of in vivo or in vitro functional studies.

Modulation by antidopaminergic drugs

The up-regulation of  $D_2$  receptors by antidopaminergic drugs is well established (Seeman 1981) and is believed to result from the blockade of those receptors. Because the  $D_3$  receptor has been proposed as a potential antipsychotic site, there has been a great deal of interest in whether antipsychotic drugs produce a similar up-regulation in D<sub>3</sub> sites. To date, several studies have been performed to examine the effects of antipsychotic drugs on the expression of D<sub>3</sub> receptor mRNA with differing results. Chronic treatment with haloperidol, sulpiride, and clozapine has been reported to produce increases in D<sub>3</sub> receptor mRNA in whole brain of 3- to 5-fold as assessed by RNAse protection (Buckland et al. 1992, 1993). More modest increases in D<sub>3</sub> receptor mRNA (40-60%) were observed in olfactory tubercle following treatment with haloperidol and sulpiride, but not clozapine, for 14 days as assessed by polymerase chain reaction (Wang et al. 1996). Hurley et al. (1996c) reported significant increases in D<sub>3</sub> receptor mRNA expression following 21 days of treatment with clozapine in striatal and limbic brain areas by in situ hybridization. Clozapine also produced a decrease in  $D_3$  mRNA in the islands of Calleja. The density of [<sup>3</sup>H]7-OH-DPAT-labeled sites were unaltered after treatment with either clozapine or sulpiride. In contrast, haloperidol did not produce significant alterations in D<sub>3</sub> mRNA expression but an increase in D<sub>3</sub> binding was observed in the islands of Calleja. Other studies, using different treatment paradigms, reported no change in D<sub>3</sub> receptor mRNA expression (Fishburn et al. 1994; Fox et al. 1994; Lévesque et al. 1995) or  $D_3$  binding (Lèvesque et al. 1995: Levant 1997). Clearly, the conflicting results observed in these studies may result from the different treatment paradigms used and must be resolved by further study. Moreover, these studies did not assess whether the observed antipsychotic-induced alterations in D<sub>3</sub> receptor mRNA expression resulted in changes in the density or affinity of  $D_3$  receptors.

# Role in behavior

One of the primary aims in the study of the novel dopamine receptors is the elucidation of their role in organismal function. To date, pharmacological and molecular methods have been used in attempt to selectively study these novel sites. In the case of the D<sub>3</sub> receptor, numerous pharmacological studies have been performed as well as several studies using targeted mutation and antisense technologies. To date, a large body of data has been generated. Care, however, must be taken in the interpretation of these findings. It must be noted that the  $D_2/D_3$  selectivity of many drugs, particularly agonists, varies considerably in in vitro binding studies. In addition, compensatory adaptations must be considered in the interpretation of studies using "knockout" or antisense approaches. While all of methodologies currently available for the study of the functional role of the D<sub>3</sub> receptor possess certain limitations, several themes are gradually emerging from the body of data collected thus far.

#### Locomotor activity

Although the  $D_3$  receptor has been implicated in numerous behaviors, the receptor is most widely cited in the modulation of locomotor activity. In contrast to the  $D_2$  receptor, where stimulation is believed to increase locomotion, stimulation of the  $D_3$  site appears to inhibit locomotor activity. This effect was initially reported in several studies using the  $D_3$ -preferring drug 7-OH-DPAT. This drug produced a biphasic effect on locomotor activity in rats in which locomotion was inhibited at lower doses and stimulated at higher doses (Daly and Waddington 1993; McElroy et al. 1993; Ahlenius and Salmi 1994; Svensson et al. 1994a,b; Khroyan et al. 1995; Depoortere et al. 1996; Kagaya et al. 1996). The inhibitory effects of the drug were attributed to activity of the drug at the D<sub>3</sub> receptor; the stimulatory effects to the actions of higher doses of the drug at the D<sub>2</sub> receptor (Daly and Waddington 1993; Ahlenius and Salmi 1994; Svensson et al. 1994a). This interpretation was supported by the demonstration that the inhibitory effects of 7-OH-DPAT were produced by doses of the drug which do not produce significant occupancy of D<sub>2</sub> receptors in vivo (Levant et al. 1996). Several studies observed inhibition of locomotor activity after microinjection of 7-OH-DPAT into the nucleus accumbens (Gilbert and Cooper 1995; Kling-Petersen et al. 1995). Inhibition of locomotor activity by 7-OH-DPAT has also been reported in mice (Starr and Starr 1995). The D<sub>3</sub>-preferring agonist PD 128907 produced similar biphasic effects on locomotor activity in rats (Pugsley et al. 1995).

Consistent with the effects of  $D_3$  agonists on locomotor activity, the  $D_3$ -preferring antagonist nafadotride produced biphasic effects on locomotor activity in rats, stimulating locomotion at lower doses, inhibiting at higher doses (Sautel et al. 1995b). As with 7-OH-DPAT, doses of nafadotride which increased locomotor activity were shown to produce negligible occupancy of  $D_2$  receptors, while those which inhibited locomotion produced significant  $D_2$  occupancy (Levant and Vansell 1997). Another  $D_3$ -preferring antagonist, U-99194A, has also been reported to increase locomotor activity (Waters et al. 1993, 1994). Finally, increased locomotor activity, rearing behavior, and hyperactivity in an exploratory test were observed in one strain of  $D_3$  "knock-out" mice (Accili et al.

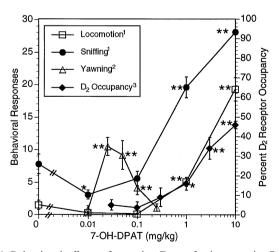


Fig. 1 Behavioral effects of putative D<sub>3</sub>-preferring agonist 7-OH-DPAT in rat. Comparison with in vivo occupancy of D<sub>2</sub> dopamine receptors. Data are reported as number of exhibitions of a behavior during observation periods. In vivo occupancy of D<sub>2</sub> dopamine receptor by systemically administered (SC) 7-OH-DPAT was defined as percent protection of receptors from inactivation by EEDQ as determined by ex vivo [<sup>3</sup>H] spiperone binding in striatal membranes. \*\*P < 0.01, P < 0.05, <sup>1</sup>Daly and Waddington (1993). <sup>2</sup>Kurashima et al. (1995). <sup>3</sup>Levant et al. (1995)

1996). Likewise, a preliminary report on another strain of  $D_3$ - deficient mice indicated a transient increase in activity in a novel environment compared to wild-type mice although no alterations in agonist stimulated locomotor behavior were observed (Koeltzow et al. 1995; Xu et al. 1995). Taken together, these findings indicate the probable involvement of the  $D_3$  receptor in the modulation of locomotor activity in a manner opposite of that of the  $D_2$  receptor.

#### Reinforcement and reward

Stimulation of the D<sub>3</sub> receptor has also been implicated in intriguing behavioral effects involving reinforcement and reward. Of note, 7-OH-DPAT has been reported to decrease self-administration of cocaine (Caine and Koob 1993; Parsons et al. 1996; Caine et al. 1997) and self-stimulation of the ventral tegmental area (Depoortere et al. 1996). Likewise, stimulation of D<sub>3</sub> sites is implicated in blocking the reinforcing effects of cocaine and *d*-amphetamine (Kling-Petersen et al. 1994), decreasing the rate of food-reinforced responding in a fixed-ratio operant paradigm (Sanger et al. 1996), and producing an aversive effect in a conditioned place preference paradigm (Chaperon and Thiebot 1996). In addition, the subjective effects of 7-OH-DPAT and other  $D_3$ -preferring agonists generalize to cocaine in drug-discrimination paradigms (Lamas et al. 1996). These observations have important implications for the understanding and treatment of drug addiction. However, as discussed above, further study must determine the role of specific dopamine receptor subtypes in these observations.

# Other behaviors

Based on the somewhat limited in vivo and in vitro pharmacological data currently available, it is possible that the  $D_3$  site may play a role in several additional behaviors. These include agonist-induced yawning and hypothermia (Damsma et al. 1993; Ahlenius and Salmi 1994; Millan et al. 1994, 1995a,b; Ferrari and Guiliani 1995; Khroyan et al. 1995; Kurashima et al. 1995), decreased sniffing (Daly and Waddington 1993), decreased alcohol consumption (Meert and Clincke 1994), and increased penile erection and ejaculatory behavior (Ahlenius and Larsson 1995; Ferrari and Guiliani 1995). Clearly, further study must confirm the role of the  $D_3$  receptor in these behaviors, particularly in light of the significant variability in the in vitro pharmacological profile of the  $D_2$  and  $D_3$  sites on which much of the interpretation of these data is based.

In addition to the pharmacological effects discussed above, a variety of other behavioral and physiological effects of putatively  $D_3$ -preferring compounds have been reported. These effects include conditioned taste aversion (Bevins et al. 1996), disruption of huddling behavior in rats (Kagaya et al. 1996), decreased grooming (Khroyan et al. 1995), alterations in performance in an elevated maze test (Rodgers et al. 1996), decreased prepulse inhibition (Caine et al. 1995), catalepsy (Millan et al. 1995b; Sautel et al. 1995b), enhancement of morphine-induced conditioned place preference (Rodriguez De Fonseca et al. 1995), inhibition of pilocarpine-induced limbic seizures (Alam and Starr 1994), induction of depressant EEG patterns (Popoli et al. 1996), increased oxytocin secretion (Uvnas Moberg et al. 1995), and decreased gastric acid secretion (Glavin 1994). The  $D_3$  receptor has also been suggested to play a role in emesis in the dog (Yoshida et al. 1995) and decreased climbing in mice (Sautel et al. 1995b). The involvement of dopamine receptors in these effects is likely; however, evidence for the selective involvement of the D<sub>3</sub> site is currently lacking.

# **Role in development**

Expression of the  $D_3$  receptor in the brain occurs quite early in development. In rat, D<sub>3</sub> receptor mRNA is detectable by polymerase chain reaction as early as embryonic day 11 and is clearly detectable by embryonic day 14 (Cadoret et al. 1993). Similarly, in the mouse, D<sub>3</sub> receptor mRNA is detectable on embryonic day 9.5, 4 days before the detection of  $D_2$  receptor mRNA (Fishburn et al. 1996). D<sub>3</sub> receptor binding, as assessed with [<sup>3</sup>H]7-OH-DPAT, is detectable in the islands of Calleja and olfactory tubercle at birth in the mouse brain. D<sub>3</sub> binding in the nucleus accumbens is detectable on postnatal day 4, substantia nigra on postnatal day 8, and in the vestibulocerebellum on postnatal day 11. Binding in these brain areas was observed to increase in density through development until adult levels were reached. In addition, transient expression of D<sub>3</sub> binding was observed in the dorsolateral parietal cortex between postnatal days 6 and 15 (Demotes-Mainard et al. 1996). In contrast to the mouse,  $D_3$ receptors in the developing rat appear to be expressed later in development than D<sub>2</sub> receptors in the forebrain, islands of Calleja, and nucleus accumbens. D2 receptor binding occurred as early as postnatal day 3, whereas, significant levels of [125I]trans-7-OH-PIPATlabeled D<sub>3</sub> sites were not detected until postnatal days 14 -21 (Stanwood et al. 1997).

Interestingly, during the second trimester of gestation,  $D_2$ -like receptors are transiently expressed in the cortical plate of the developing human (Todd 1992; Unis 1993). Preliminary reports also indicate transient, dense expression of  $D_3$  receptor mRNA in the cortical plate of human brain at midgestation (Unis and Dorsa 1993; Unis et al. 1995) suggesting that these receptors are of the  $D_3$  subtype. This transient expression of  $D_3$ receptors suggests a role for dopamine in orchestrating 10

neuronal migration and differentiation during this period of accelerated cortical development that is mediated by the  $D_3$  receptor. This hypothesis is supported by the observation that stimulation of  $D_3$  sites induces increased branching and extension of neurites in both mesencephalic MN9D cells and primary mesencephalic neuronal cultures (Swarzenski et al. 1994).

# Role in neuropsychiatric disorders

# Schizophrenia

Investigation into alterations in the expression of the  $D_3$  receptor in disease is still in the early stages. Accordingly, data available at present is relatively limited. Initial studies examining the densities of the D<sub>2</sub>-like dopamine receptors in post-mortem putamen tissues from schizophrenic patients using relatively nonselective radioligands suggested an elevation in  $D_4$  sites but not  $D_2$  or  $D_3$  (Seeman et al. 1993). Another postmortem study of schizophrenic brain reported that D<sub>3</sub> mRNA was lost in certain cortical regions of their brains, whereas a mRNA encoding a splice variant of the  $D_3$  receptor protein ( $D_{3nf}$ ) could readily be detected in the same anatomic regions (Schmauss 1996; Schmauss et al. 1993).  $D_{3nf}$ , a  $D_3$ -like protein with a different C terminus, results from a deletion of 98 nucleotides that constitute the C-terminal region of the putative third cytoplasmic domain of the D<sub>3</sub> receptor (Schmauss et al. 1993; Liu et al. 1994b). The predominance of D<sub>3nf</sub> protein in schizophrenic brain appears to result from increased D<sub>3nf</sub>-specific splicing activity which could potentially contribute to the pathogenesis of the disease.

Although at present there is no evidence of genetic linkage between the D<sub>3</sub> gene and schizophrenia (Coon et al. 1993; Wiese et al. 1993; Nanko et al. 1994a; Sabate et al. 1994), several studies suggest that a distinct polymorphism in the first coding exon of the  $D_3$  gene, the BalI or MscI restriction fragment length polymorphism, may increase the suspectibility to disease (Crocq et al. 1992; Mant et al. 1994; Kennedy et al. 1995; Griffon et al. 1996; Shaikh et al. 1996). This polymorphism corresponds to a point mutation that results in a serine-to-glycine substitution in the receptor's extracellular N-terminal domain. This substitution is likely to affect the way in which the protein is incorporated into the membrane and appears to increase the affinity for dopamine (Lundstrom and Turpin 1996). Although other studies have failed to support an association between the BalI polymorphism and schizophrenia (Jonsson et al. 1993; Nanko et al. 1993; Nothen et al. 1993; Yang et al. 1993; Bella et al. 1994; Macciardi et al. 1994; Saha et al. 1994; Higuchi et al. 1995; Gaitonde et al. 1996; Nimgaonkar et al. 1996; Rietschel

et al. 1996; Tanaka et al. 1996; Chen et al. 1997), this genotype may be associated with certain subsets of schizophrenics. For example, the association between the *Bal*I polymorphism and schizophrenia was found to be particularly strong in patients with a good response to antipsychotic treatment (Asherson et al. 1996). Other studies found a positive association only in male subjects (Asherson et al. 1996) or in patients with a familial history of schizophrenia (Nimgaonkar et al. 1993). Likewise, the *Bal*I polymorphism has been associated with the onset of the disease in certain age groups (Durany et al. 1996; Maziade et al. 1997).

#### Parkinson's disease

Investigation into alterations in the D<sub>3</sub> receptor in Parkinson's disease suggest that while decreases in expression of D<sub>3</sub> receptor mRNA are observed in certain brain regions, such as the olfactory tubercle, during aging (Valerio et al. 1994), densities of both D<sub>3</sub> receptor mRNA and D<sub>3</sub> binding are not altered in post-mortem brain from patients afflicted with Parkinson's disease (Hurley et al. 1996a). Although no significant alterations in D<sub>3</sub> receptor expression in the brains of Parkinson's patients have yet been reported, a decrease in D<sub>3</sub> receptor mRNA expression in lymphocytes of Parkinson's patients has been observed. In fact, the magnitude of the decrease in  $D_3$  receptor mRNA expression correlated with the severity of the disease suggesting that this might serve as a marker for monitoring disease progression (Nagai et al. 1996). The detection of D<sub>3</sub> receptor mRNA in human lymphocytes, however, has not been observed in some studies (Vile and Strange 1996). No association has been made between the  $D_3$  receptor gene and polymorphisms in its alleles in the incidence of Parkinson's disease (Nanko et al. 1994b; Higuchi et al. 1995).

#### Cocaine use

Finally, alterations in the density of the  $D_3$  receptor have been reported with cocaine use. No changes in D<sub>3</sub> receptor mRNA expression were observed in the dorsal or ventral striata of post-mortem brains of chronic cocaine abusers (Meador-Woodruff et al. 1995). However, the density of D<sub>3</sub> sites was observed to be increased from 2- to 3-fold in the caudate, nucleus accumbens, and substantia nigra of persons who died from a cocaine over-dose (Staley and Mash 1996). In addition, a recent study of gene polymorphisms associated with reward dependence suggests that the BalI D<sub>3</sub> receptor polymorphism, as well as D<sub>4</sub> receptor polymorphisms, appears to interact with certain serotonin receptor polymorphisms to produce this personality trait which may contribute to the susceptibility to drug addiction (Ebstein et al. 1997).

Clearly, investigation of alterations in the  $D_3$  receptor in disease is in the early stages. Further study must verify whether alterations in the density or the regulation of this receptor exist in various pathological conditions.

### Conclusion

In the short time since the identification of the dopamine  $D_3$  receptor, considerable progress has been made towards understanding the function of this novel site. Although some avenues of investigation have yielded more definitive results than others, studies to date indicate the  $D_3$  receptor is localized preferentially in limbic brain areas and affects locomotion. The receptor may also play a role in reinforcement and reward. A subpopulation of the receptors appear to be autoreceptors which modulate dopamine synthesis, release, and neuronal activity. These observations have led to the hypothesis that the  $D_3$  receptor may be an appropriate target in the treatment of neuropsychiatric disorders, such as schizophrenia and drug addiction. The role of  $D_3$  sites in disease, however, remains to be established. Genetic association of D<sub>3</sub> receptor polymorphisms with neuropsychiatric disorders have been proposed but remain controversial. Alterations in expression of D<sub>3</sub> sites may occur in some diseases. Although some study of this receptor is clearly in the early stages, these findings lay the foundation for future investigation. Further study may ultimately aid in the elucidation of the role of the D<sub>3</sub> receptor in health and disease and their potential utility in the treatment of neuropsychiatric disorders.

#### References

- Accili D, Fishburn CS, Drago J, Steiner H, Lachowicz JE, Park BH, Gauda EB, Lee EJ, Cool MH, Sibley DR, Gerfen CR, Westphal H, Fuchs S (1996) A targeted mutation of the D<sub>3</sub> dopamine receptor gene is associated with hyperactivity in mice. Proc Natl Acad Sci USA 93:1945–1949
- Ahlenius S, Larsson K (1995) Effects of the dopamine D<sub>3</sub> receptor ligand 7-OH-DPAT on male rat ejaculatory behavior. Pharmacol Biochem Behav 51:545–547
- Ahlenius S, Salmi P (1994) Behavioral and biochemical effects of the dopamine D<sub>3</sub> receptor-selective ligand, 7-OH-DPAT, in the normal and the reserpine-treated rat. Eur J Pharmacol 260:177–181
- Akunne HC, Towers P, Ellis GJ, Dijkstra D, Wikstrom H, Heffner TG, Wise LD, Pugsley TA (1995) Characterization of binding of [<sup>3</sup>H]PD 128907, a selective dopamine D<sub>3</sub> receptor agonist ligand, to CHO-K1 cells. Life Sci 57:1401–1410
- Alam AM, Starr MS (1994) Effects of dopamine D<sub>3</sub> receptor agonists on pilocarpine-induced limbic seizures in the rat. Neuroscience 60:1039–1047
- Aretha CW, Sinha A, Galloway MP (1995) Dopamine D<sub>3</sub>-preferring ligands act at synthesis modulating autoreceptors. J Pharmacol Exp Ther 274:609–613

- Ariano MA, Sibley DR (1994) Dopamine receptor distribution in the rat CNS: elucidation using anti-peptide antisera directed against D<sub>1A</sub> and D<sub>3</sub> subtypes. Brain Res 649:95–110
- Asherson P, Mant R, Holmans P, Williams J, Cardno A, Murphy K, Jones L, Collier D, McGuffin P, Owen MJ (1996) Linkage, association and mutational analysis of the dopamine D<sub>3</sub> receptor gene in schizophrenia. Mol Psychiatry 1:125–132
- Bancroft GN, Morgan KA, Flietstra RJ, Levant B (1997) Binding of [<sup>3</sup>H]PD 128907, a putatively selective ligand for the  $D_3$ dopamine receptor, in rat brain: a receptor binding and quantitative autoradiographic study. Neuropsychoparmacology (in press)
- Bevins RA, Delzer TA, Bardo MT (1996) Characterization of conditioned taste aversion produced by 7-OH-DPAT in rats. Pharmacol Biochem Behav 53:695-699
- Booze RM, Wallace DR (1995) Dopamine  $D_2$  and  $D_3$  receptors in the rat striatum and nucleus accumbens: use of 7-OH-DPAT and [<sup>125</sup>I]-iodosulpride. Synapse 19:1–13
- Bouthenet ML, Souil E, Martres MP, Sokoloff P, Giros B, Schwartz JC (1991) Localization of dopamine D<sub>3</sub> receptor mRNA in the rat brain using in situ hybridization histochemistry: comparison with dopamine D<sub>2</sub> receptor mRNA. Brain Res 564: 203–219
- Bowery B, Rothwell LA, Seabrook GR (1994) Comparison between the pharmacology of dopamine receptors mediating the inhibition of cell firing in rat brain slices through the substantia nigra pars compacta and ventral tegmental area. Br J Pharmacol 112:873–880
- Boyfield I, Winn F, Coldwell M (1996) Comparison of agonist potencies at human dopamine  $D_2$  and  $D_3$  receptors, expressed in the same cell line, using the cytosensor microphysiometer. Biochem Soc Transact 24:S57
- Buckland PR, O'Donovan MC, McGuffin P (1992) Changes in dopamine  $D_1$ ,  $D_2$  and  $D_3$  receptor mRNA levels in rat brain following antipsychotic treatment. Psychopharmacology 106:479-483
- Buckland PR, O'Donovan Mc, McGuffin P (1993) Clozapine and sulpiride up-regulate dopamine D<sub>3</sub> receptor mRNA levels. Neuropharmacology 32:901–907
- Burris KD, Filtz TM, Chumpradit S, Kung MP, Foulon C, Hensler JG, Kung HF, Molinoff PB (1994) Characterization of [<sup>125</sup>I](*R*)-trans-7-hydroxy-2-[*N*-propyl-*N*(3'-iodo-2'-propenyl) amino] tetralin binding to dopamine D<sub>3</sub> receptors in rat olfactory tubercle. J Pharmacol Exp Ther 268:935–942
- Burris KD, Pacheco MA, Filtz TM, Kung MP, Kung HF, Molinoff PB (1995) Lack of discrimination by agonists for D<sub>2</sub> and D<sub>3</sub> dopamine receptors. Neuropsychopharmacology 12:335–345
- Cadoret MA, Jaber M, Bloch B (1993) Prenatal  $D_1$ ,  $D_{1b}$  and  $D_3$ dopamine receptor gene expression in the rat forebrain: detection by reverse polymerase chain reaction. Neurosci Lett 155:92–95
- Caine SB, Koob GF (1993) Modulation of cocaine self-administration in the rat through D-3 dopamine receptors. Science 260:1814–1816
- Caine SB, Koob GF, Parsons LH, Everitt BJ, Schwartz JC, Sokoloff P (1997) D<sub>3</sub> receptor test in vitro predicts decreased cocaine selfadministration in rats. Neuroreport 8:2373–2377
- Caine SB, Geyer MA, Swerdlow NR (1995) Effects of  $D_3/D_2$  dopamine receptor agonists and antagonists on prepulse inhibition of acoustic startle in the rat. Neuropsychopharmacology 12:139–145
- Castro SW, Strange PG (1993) Coupling of D<sub>2</sub> and D<sub>3</sub> dopamine receptors to G-proteins. FEBS Lett 315:223–226
- Chaperon F, Thiebot MH (1996) Effects of dopaminergic D<sub>3</sub> receptor preferring ligands on the acquisition of place conditioning in rats. Behav Pharmacol 7:105–109
- Chen C-H, Liu MY, Wei F-C, Koong F-J, Hwu H-G, Hsiao K-J (1997) Further evidence of no association between ser-gly polymorphism of dopamine  $D_3$  receptor gene and schizophrenia. Am J Med Genet 74:40–43

- Chio CL, Lajiness ME, Huff RM (1994) Activation of heterologously expressed D<sub>3</sub> dopamine receptors: comparison with D<sub>2</sub> dopamine receptors. Mol Pharmacol 45:51–60
- Chumpradit S, Kung MP, Vessotskie J, Foulon C, Mu M, Kung HF (1994) Iodinated 2-aminotetralins and 3-amino-1-benzopyrans: ligands for dopamine D<sub>2</sub> and D<sub>3</sub> receptors. J Med Chem 37:4245-4250
- Coon H, Byerly W, Hoik J, Hoff M, Myles-Worsley M, Lannfelt L, Sokoloff P, Schwartz J-C, Waldo, M. (1993) Linkage analysis of schizophrenia with five dopamine receptor genes in nine pedigrees. Am J Hum Genet 52:327–334
- Cooper DC, Loeltzow TE, Xu M, Tonegawa S, White FJ, Wolf ME (1996) Regulation of dopamine release and synthesis in dopamine D<sub>3</sub>R receptor knockout mice. Soc Neurosci Abstr 22:1319
- Cox BA, Rosser MP, Kozlowski MR, Duwe KM, Neve RL, Neve KA (1995) Regulation and functional characterization of a rat recombinant dopamine  $D_3$  receptor. Synapse 21:1-9
- Crocq MA, Mant R, Asherson P, Williams J, Hode Y, Mayerova A, Collier D, Lannfelt L, Sokoloff P, Schwartz J-C, Gill M, Macher J-P, McGuffin P, Owen MJ (1992) Association between schizophrenia and homozygosity at the D<sub>3</sub> dopamine receptor gene. J Med Genet 29:858–860
- Curran EJ, Watson SJ Jr (1995) Dopamine receptor mRNA expression patterns by opioid peptide cells in the nucleus accumbens of the rat: a double in situ hybridization study. J Comp Neurol 361:57–76
- Dal Toso R, Sommer B, Ewert M, Herb A, Pritchett DB, Bach A, Shivers BD, Seeberg PH (1989) The dopamine D<sub>2</sub> receptor: two molecular forms generated by alternative splicing. EMBO J 8:4025-4034
- Daly SA, Waddington JL (1993) Behavioural effects of the putative D-3 dopamine receptor agonist 7-OH-DPAT in relation to other "D-2-like" agonists. Neuropharmacology 32: 509-510
- Damsma G, Bottema T, Westerink BH, Tepper PG, Dijkstra D, Pugsley TA, MacKenzie RG, Heffner TG, Wikstrom H (1993) Pharmacological aspects of *R*-(+)-7-OH-DPAT, a putative dopamine D<sub>3</sub> receptor ligand. Eur J Pharmacol 249: R9–10
- Demotes-Mainard J, Henry C, Jeantet Y, Arsaut J, Arnauld E (1996) Postnatal ontogeny of dopamine D<sub>3</sub> receptors in the mouse brain: autoradiographic evidence for a transient cortical expression. Dev Brain Res 94:166–174
- Depoortere R, Perrault G, Sanger DJ (1996) Behavioral effects in the rat of the putative dopamine D<sub>3</sub> receptor agonist 7-OH-DPAT: comparison with quinpirole and apomorphine. Psychopharmacology 124: 231–240
- Devoto P, Collu M, Muntoni AL, Pistis M, Serra G, Gessa GL, Diana M (1995) Biochemical and electrophysiological effects of 7-OH-DPAT on the mesolimbic dopaminergic system. Synapse 20:153–155
- Di Bella D, Catalano M, Strukel A, Nobile M, Novelli E, Smeraldi E (1994) Distribution of the MscI polymorphism of the dopamine D<sub>3</sub> receptor in an Italian psychotic population. Psychiatr Genet 4:39–42
- Diaz J, Levesque D, Lammers CH, Griffon N, Martres MP, Schwartz JC, Sokoloff P (1955) Phenotypical characterization of neurons expressing the dopamine  $D_3$  receptor in the rat brain. Neuroscience 65:731–745
- Durany N, Thome J, Palomo A, Foley P, Riedere P, Cruz-Sanchez FF (1996) Homozygosity at the dopamine D<sub>3</sub> receptor gene in schizophrenic patients. Neurosci Lett 220:151–154
- Ebstein RP, Segman R, Benjamin J, Osher Y, Nemanov L, Belmaker RH (1997) 5- $HT_{2C}$  (HTR2C) serotonin receptor gene polymorphism associated with the human personality trait of reward dependence: interaction with dopamine D<sub>4</sub> receptor (D4DR) and dopamine D<sub>3</sub> (D3DR) polymorphisms. Am J Med Genet 74:65–72

- Ferrari F, Guiliani D (1995) Behavioral effects of the dopamine D<sub>3</sub> receptor agonist 7-OH-DPAT in rats. Pharmacol Res 32: 63-68
- Fishburn CS, Belleli D, David C, Carmon S, Fuchs S (1993) A novel short isoform of the  $D_3$  dopamine receptor generated by alternative splicing in the third cytoplasmic loop. J Biol Chem 268:5872–5878
- Fishburn CS, David C, Carmon S, Fuchs S (1994) The effect of haloperidol on  $D_2$  dopamine receptor subtype mRNA levels in the brain. FEBS Lett 339:63-66
- Fishburn CS, Bedford M, Lonai P, Fuchs S (1996) Early expression of D<sub>3</sub> dopamine receptors in murine embryonic development. FEBS Lett 381:257–261
- Flietstra RJ, Levant B (1996) Simultaneous comparison of D<sub>2</sub>/D<sub>3</sub> affinity in rat brain. Soc Neurosci Abstr 22:827
- Fox CA, Mansour A, Watson SJ Jr (1994) The effects of haloperidol on dopamine receptor gene expression. Exp Neurol 130:288–303
- Freedman SB, Patel S, Marwood R, Emms F, Seabrook GR, Knowles MR, McAllister G (1994) Expression and pharmacological characterization of the human D<sub>3</sub> dopamine receptor. J Pharmacol Exp Ther 286:417–426
- Gainetdinov RR, Sotnikova TD, Grekhova TV, Rayevsky KS (1996) In vivo evidence for preferential role of dopamine D<sub>3</sub> receptor in the presynaptic regulation of dopamine release but not synthesis. Eur J Pharmacol 308:261–269
- Gaitonde EJ, Morris A, Sivagnanasundaram S, McKenna PJ, Hunt DM, Mollon JD (1996) Assessment of association of D<sub>3</sub> dopamine receptor mscI polymorphism with schizophrenia: analysis of symptom ratings, family history, age at onset, and movement disorders. Am J Med Genet 67:455–458
- Gehlert DR (1992) Quantitative autoradiography of Gpp(NH)p sensitive and insensitive [<sup>3</sup>H]quinpirole binding sites in the rat brain. Synapse 14:113–120
- Gehlert DR, Gackenheimer SL, Seeman P, Schaus J (1992) Autoradiographic localization of  $[^{3}H]$ quinpirole binding to dopamine D<sub>2</sub> and D<sub>3</sub> receptors in rat brain. Eur J Pharmacol 211:189–194
- Gilbert DB, Cooper SJ (1995) 7-OH-DPAT injected into the accumbens reduces locomotion and sucrose ingestion: D<sub>3</sub> autoreceptor-mediated effects? Pharmacol Biochem Behav 52:275-280
- Gilbert DB, Millar J, Cooper SJ (1995) The putative dopamine D<sub>3</sub> agonist, 7-OH-DPAT, reduces dopamine release in the nucleus accumbens and electrical self-stimulation to the ventral tegmentum. Brain Res 681:1–7
- Giros B, Sokoloff P, Martres MP, Riou JF, Emorine LJ, Schwartz JC (1989) Alternative splicing directs the expression of two D<sub>2</sub> dopamine receptors isoforms. Nature 342:923–926
- Giros B, Martres MP, Sokoloff P, Schwartz JC (1990) Clonage gu gène du récepteur dopaminergique D<sub>3</sub> humain et identification de son chromosome. CR Acad Sci III 311:501–508
- Giros B, Martres MP, Pilon C, Sokoloff P, Schwartz JC (1991) Shorter variants of the D<sub>3</sub> dopamine receptor produced through various patterns of alternative splicing. Biochem Biophys Res Commun 176:1584–1592
- Glavin GB (1994) A dopamine D<sub>3</sub> receptor agonist, 7-hydroxy-*N*,*N*di-*n*-propyl-2-aminotetralin, reduces gastric acid and pepsin secretion and experimental gastric mucosal injury in rats. Life Sci 56:287–393
- Gobert A, Rivet JM, Audinot V, Cistarelli L, Spedding M, Vian J, Peglion JL, Millan MJ (1995) Functional correlates of dopamine D<sub>3</sub> receptor activation in the rat in vivo and their modulation by the selective antagonist, (+)-S 14297: II. Both D<sub>2</sub> and "silent" D<sub>3</sub> autoreceptors control synthesis and release in mesolimbic, mesocortical and nigrostriatal pathways. J Pharmacol Exp Ther 275:899–913
- Gobert A, Lejeune F, River J-M, Cistarelli L, Millan MJ (1996) Dopamine  $D_3$  (auto)receptors inhibit dopamine release in the frontal cortex of freely moving rats in vivo. J Neurochem 66:2209-2212

- Griffon N, Crocq MA, Pilon C, Martres MP, Mayerova A, Uyanik G, Burgert E, Duval F, Macher JP, Javoyagid F, Tamminga CA, Schwartz JC, Sokoloff P (1996) Dopamine D<sub>3</sub> receptor gene: organization, transcript variants, and polymorphism associated with schizophrenia. Am J Med Genet 67:63–70
- Griffon N, Pilon C, Schwartz JC, Sokoloff P (1995) The preferential dopamine D<sub>3</sub> receptor ligand, (+)-UH232, is a partial agonist. Eur J Pharmacol 282:R3-4
- Griffon N, Sautel F, Pilon C, Levesque D, Sokoloff P, Schwartz JC, Diaz J, Simon P, Costentin J, Nann A, Wermuth CG (1996) Functional models for the dopamine D<sub>3</sub> receptor. Biochem Soc Transact 24:193–198
- Griffon N, Pilon C, Sautel F, Schwartz JC, Sokoloff P (1997) Two intracellular signaling pathways for the dopamine D<sub>3</sub> receptor: opposite and synergistic interactions with cyclic AMP. J Neurochem 68:1–9
- Grigoriadis DE, Seeman P (1985) Complete conversion of brain D<sub>2</sub> dopamine receptors from the high- to low-affinity state for dopamine agonists using sodium ions and guanine nucleotides. J Neurochem 44:1925–1935
- Hall H, Halldin C, Dijkstra D, Wikstrom H, Wise LD, Pugsley TA, Sokoloff P, Pauli S, Farde L, Sedvall G (1996) Autoradiographic localisation of D<sub>3</sub>-dopamine receptor in the human brain using the selective D<sub>3</sub>-dopamine receptor agonist (+)-[<sup>3</sup>H]PD 128907. Psychopharmacology 128:240–247
- Herroelen L, De Backer J-P, Wilczak N, Flamez A, Vaquelin G, De Keyser J (1994) Autoradiographic distribution of D<sub>3</sub>-type dopamine receptors in human brain using [3H]7-hydroxy-N,Ndi-n-propyl-2-aminotetralin. Brain Res 648:222–228
- Higuchi S, Muramatsu T, Arai H, Hayashida M, Sasaki H, Trojanowski JQ (1995) Polymorphisms of dopamine receptor and transporter genes and Parkinson's disease. J Neural Transm [Park Dis Dement Sect] 10:107–113
- Hurley MJ, Stubbs CM, Jenner P, Marsden CD (1996a) D<sub>3</sub> receptor expression within the basal ganglia is not affected by Parkinson's disease. Neurosci Lett 214:75–78
- Hurley MJ, Stubbs CM, Jenner P, Marsden CD (1996b) Dopamine D<sub>3</sub> receptor are not involved in the induction of c-fos mRNA by neuroleptic drugs: comparison of the dopamine D<sub>3</sub> receptor antagonist GR 103691 with typical and atypical neuroleptics. Eur J Pharmacol 318:283–293
- Hurley MJ, Stubbs CM, Jenner P, Marsden CD (1996c) Effect of chronic treatment with typical and atypical neuroleptics on the expression of dopamine  $D_2$  and  $D_3$ receptor in rat brain. Psychopharmacology 128:362–270
- Jonsson E, Lannfelt L, Sokoloff P, Schwartz J-C, Sedvall G (1993) Lack of association between schizophrenia and alleles in the dopamine D<sub>3</sub> receptor gene. Acta Psychiatr Scand 87:543–549
- Kagaya T, Yonaga M, Furuya Y, Hashimoto T, Kuroki J, Nishizawa Y (1996) Dopamine D<sub>3</sub> agonists disrupt social behavior in rats. Brain Res 721:229–232
- Kenakin T (1996) The classification of seven transmembrane receptors in recombinant expression systems. Pharmacol Rev 48:413-463
- Kennedy JL, Billett EA, Macciardi FM, Verga M, Parsons TJ, Meltzer HY, Lieberman J, Buchanan JA (1995) Association study of dopamine D<sub>3</sub> receptor gene and schizophrenia. Am J Med Genet 60:558–562
- Khroyan TV, Baker DA, Neisewander JL (1995) Dose-dependent effects of the D<sub>3</sub>-preferring agonist 7-OH-DPAT on motor behaviors and place conditioning. Psychopharmacology 122:351–357
- Kling-Petersen T, Ljung E, Svensson K (1994) The preferential dopamine autoreceptor antagonist (+)-UH232 antagonizes the positive reinforcing effects of cocaine and *d*-amphetamine in the ICSS paradigm. Pharmacol Biochem Behav 49:345–351
- Kling-Petersen T, Ljung E, Svensson K (1995) Effects on locomotor activity after local application of D<sub>3</sub> preferring compounds in discrete areas of the rat brain. J Neural Transm Gen Sect 102:209–220

- Koeltzow T, Cooper DC, Hu XT, Xu M, Tonegawa S, White FJ (1995) In vivo effects of dopaminergic ligands on dopamine D<sub>3</sub> receptor deficient mice. Soc Neurosci Abstr 21:364
- Kreiss DS, Bergstrom DA, Gonzalez AM, Huang KX, Sibley DR, Walters JR (1995) Dopamine receptor agonist potencies for inhibition of cell firing correlate with dopamine D<sub>3</sub> receptor binding affinities. Eur J Pharmacol 277:209–214
- Kula NS, Baldessarini RJ, Kebabian JW, Neumeyer JL (1994) S-(+)-Aporphines are not selective for human D<sub>3</sub> dopamine receptors. Cell Mol Neurobiol 14:185–191
- Kung MP, Chumpradit S, Frederick D, Garner S, Burris KD, Molinoff PB, Kung HF (1994) Characterization of binding sites for [1251] R(+) trans-7-OH-PIPAT in rat brain. Naunyn-Schmiedeberg's Arch Pharmacol 350:611–617
- Kurashima M, Yamada K, Nagashima M, Shirakawa K, Furukawa T (1995) Effects of putative dopamine  $D_3$  receptor agonists, 7-OH-DPAT, and quinpirole, on yawning, stereotypy, and body temperature in rats. Pharmacol Biochem Behav 52: 503–508
- Lahti RA, Roberts RC, Tamminga CA (1995) D<sub>2</sub>-family receptor distribution in human postmortem tissue: an autoradiographic study. NeuroReport 6:2505–2512
- Lamas X, Negus SS, Nader MA, Mello NK (1996) Effects of the putative dopamine D<sub>3</sub> receptor agonist 7-OH-DPAT in rhesus monkey trained to discriminated cocaine from saline. Psychopharmacology 124:306–314
- Landwehrmeyer B, Mengod G, Palacios JM (1993a) Differential visualization of dopamine  $D_2$  and  $D_3$  receptor sites in rat brain. A comparative study using in situ hybridization histochemistry and ligand binding autoradiography. Eur J Neurosci 5:145-153
- Landwehrmeyer B, Mengod G, Palacios JM (1993b) Dopamine D<sub>3</sub> receptor mRNA and binding sites in human brain. Mol Brain Res 18:187–192
- Larson ER, Ariano MA (1995) D<sub>3</sub> and D<sub>2</sub> dopamine receptors: visualization of cellular expression patterns in motor and limbic structures. Synapse 20:325–337
- Lejeune F, Millan MJ (1995) Activation of dopamine D<sub>3</sub> autoreceptors inhibits firing of ventral tegmental dopaminergic neurones in vivo. Eur J Pharmacol 275:R7–9
- Levant B (1995) Differential sensitivity of [<sup>3</sup>H]7-OH-DPAT-labeled binding sites in rat brain to inactivation by *N*-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline. Brain Res 698:146–154
- Levant B (1996) Distribution of dopamine receptor subtypes in the CNS. In: Stone TW (ed) CNS neurotransmitters and neuromodulators. Dopamine. CRC Press, Boca Raton, pp 77–87
- Levant B (1997) Up regulation of D<sub>2</sub>, but not D<sub>3</sub> dopamine receptors, after chronic antipsychotic drug treatment. Soc Neurosci Abstr 23:683
- Levant B, DeSouza EB (1993) Differential pharmacological profile of striatal and cerebellar dopamine receptors labeled by [<sup>3</sup>H]quinpirole: identification of a discrete population of D<sub>3</sub> receptors. Synapse 14:90–95
- Levant B, Vansell NR (1997) In vivo occupancy of D<sub>2</sub> dopamine receptors by nafadotride. Neuropsychopharmacology 17:67–71
- Levant B, Grigoriadis DE, DeSouza EB (1993)  $[^{3}H]$ Quinpirole binding to putative D<sub>2</sub> and D<sub>3</sub> dopamine receptors in rat brain and pituitary gland: a quantitative autoradiographic study. J Pharamacol Exp Ther 264:991–1001
- Levant B, Grigoriadis DE, DeSouza EB (1995) Relative affinities of dopaminergic drugs at  $D_2$  and  $D_3$  dopamine receptors. Eur J Pharmacol 278:243–247
- Levant B, Bancroft GN, Selkirk CM (1996) In vivo occupancy of D<sub>2</sub> dopamine receptors by 7-OH-DPAT. Synapse 24:60–64
- Lévesque D, Diaz J, Pilon C, Martres M-P, Giros B, Souil E, Schott D, Morgat J-L, Schwartz J-C, Sokoloff P (1992) Identification, characterization, and localization of the dopamine D<sub>3</sub> receptor in rat brain using 7-[<sup>3</sup>H]hydroxy-N,N-di-n-propyl-2-aminotetralin. Proc Natl Acad Sci USA 89:8155–8159

- Lévesque D, Martres MP, Diaz J, Griffon N, Lammers CH, Sokoloff P, Schwartz JC (1995) A paradoxical regulation of the dopamine D<sub>3</sub> receptor expression suggests the involvement of an anterograde factor from dopamine neurons. Proc Natl Acad Sci USA 92:1719–1723
- Liu JC, Cox RF, Greif GJ, Freedman JE, Waszczak BL (1994a) The putative dopamine D<sub>3</sub> receptor agonist 7-OH-DPAT: lack of mesolimbic selectivity. Eur J Pharmacol 264: 269–278
- Liu K, Bergson C, Levenson R, Schmauss C (1994b) On the origin of mRNA encoding the truncated dopamine  $D_3$ -type receptor  $D_{3nf}$  and detection and  $D_{3nf}$ -like immunoreactivity in human brain. J Biol Chem 269:29220–29226
- Liu Y, Hillefors-Berglund M, von Euler G (1994c) Modulation of dopamine  $D_3$  receptor binding by N-ethylmaleimide and neurotensin. Brain Res 643:343-348
- Liu LX, Monsma FJ, Sibley DR, Chiodo LA (1996) D<sub>2L</sub>, D<sub>2S</sub>, and D<sub>3</sub> dopamine receptors stably transfected into NG108-15 cells couple to a voltage dependent potassium current via distinct g protein mechanisms. Synapse 24:156–164
- Lundstrom K, Turpin MP (1996) Proposed schizophrenia related gene polymorphism: expression of the Ser-Gly mutant human dopamine D<sub>3</sub> receptor with the Semliki forest virus system. Biochem Biophys Res Commun 225:1068–1072
- Macciardi F, Verga M, Kennedy JL, Petronis A, Bersani G, Pancheri P, Smeraldi E (1994) An association study between schizophrenia and the dopamine receptor genes DRD3 and DRD4 using haplotype relative risk. Hum Hered 44: 328–336
- MacKenzie RG, VanLeeuwan D, Pugsley TA, Shih YH, Demattos S, Tang L, Todd RD, O'Malley KL (1994) Characterization of the human dopamine D<sub>3</sub> receptor expressed in transfected cell lines. Eur J Pharmacol 266:79–85
- Malmberg A, Jackson DM, Ericksson A, Mohell N (1993) Unique binding characteristics of antipsychotic agents interacting with human D<sub>2A</sub>, D<sub>2B</sub> and D<sub>3</sub> receptors. Mol Pharmacol 43:749–754
- Mant R, Williams J, Asherson P, Parfitt E, McGuffin P, Owen MJ (1994) Relationship between homozygosity at the dopamine D<sub>3</sub> receptor gene and schizophrenia. Am J Med Genet 54:21–26
- Maziade M, Martinez M, Rodrigue C, Gauthier B, Tremblay G, Fournier C, Bissonnette L, Sinard C, Roy M-A, Ruillard E, Mérette C (1997), Childhood/early-onset and adult-onset schizophrenia. Heterogeneity at the dopamine D<sub>3</sub> receptor gene. Br J Pharmacol 17:27–30
- McElroy J, Zeller KL, Amy KA, Ward KA, Cawley JF, Mazzola AL, Keim W, Rohrbach K (1993) In vivo agonist properties of 7-hydroxy-*N*,*N*-di-*n*-propyl-2-aminotetralin, adopamine D<sub>3</sub>selective receptor ligand. Drug Dev Res 30:257–259
- Meador-Woodruff JH, Grandy DK, Van Tol HH, Damask SP, Little KY, Civelli O, Watson SJ, Jr (1994) Dopamine receptor gene expression in the human medial temporal lobe. Neuropsychopharmacology 10:239–248
- Meador-Woodruff JH, Little KY, Damask SP, Watson SJ (1995) Effects of cocaine on D<sub>3</sub> and D<sub>4</sub> receptor expression in the human striatum. Biol Psychiatry 38:263–266
- Meert TF, Clincke GH (1994) 7-OH-DPAT and alcohol consumption, withdrawal and discriminative stimulus properties in rats. Alcohol 29:489–492
- Mengod G, Villaro MT, Landwehrmeyer GB, Martinez Mir MI, Niznik HB, Sunahara RK, Seeman P, O'Dowd BF, Probst A, Palacios JM (1992) Visualization of dopamine D<sub>1</sub>, D<sub>2</sub> and D<sub>3</sub> receptor mRNAs in human and rat brain. Neurochem Int 20 [Suppl]:33s-43s
- Merchant KM, Figur LM, Evans DL (1996) Induction of dos mRNA in rat medial prefrontal cortex by antipsychotic drugs: role of dopamine D<sub>2</sub> and D<sub>3</sub> receptors. Cerebral Cortex 6:561–570
- Mierau J, Schneider FJ, Ensinger HA, Chio CL, Lajiness ME, Huff RM (1995) Pramipexole binding and activation of cloned and expressed dopamine D<sub>2</sub>, D<sub>3</sub> and D<sub>4</sub> receptors. Eur J Pharmacol 290:29–36

- Millan MJ, Audinot V, Rivet J-M, Gobert A, Vian J, Prost J-F, Spedding M, J.-L.P (1994) S 14297, a novel selective ligand a cloned human dopamine D<sub>3</sub> receptors blocks 7-OH-DPAT-induced hypothermia in rats. Eur J Pharmacol 260: R3–R5
- Millan MJ, Audinot V, Melon C, Newman Tancredi A (1995a) Evidence that dopamine D<sub>3</sub> receptors participate in clozapineinduced hypothermia. Eur J Pharmacol 280: 255–229
- Millan MJ, Peglion JL, Vian J, Rivet JM, Brocco M, Gobert A, Newmann Tancredi A, Dacquet C, Bervoets K, Girardon S, et al. (1995b) Functional correlates of dopamine D<sub>3</sub> receptor activation in the rat in vivo and their modulation by the selective antagonist, (+)-S 14297:1. Activation of postsynaptic D<sub>3</sub> receptors mediates hypothermia, whereas blockade of D<sub>2</sub> receptors elicits prolactin secretion and catalepsy. J Pharmacol Exp Ther 275:885–898
- Monsma FJ Jr, Mahan LC, McVittie LD, Gerfen CR, Sibley DR (1990) Molecular cloning and expression of a D<sub>1</sub> receptor linked to adenylyl cyclase activation. Proc Natl Acad Sci USA 87:6723–6727
- Murray AM, Ryoo HL, Gurevich E, Joyce JN (1994) Localization of dopamine  $D_3$  receptors to mesolimbic and  $D_2$  receptors to mesostriatal regions of human forebrain. Proc Natl Acad Sci USA 91:11271–11275
- Nagai Y, Ueno S, Saeki Y, Soga F, Yanagihara T (1993) Expression of the  $D_3$  dopamine receptor gene and a novel variant transcript generated by alternative splicing in human peripheral blood lymphocytes. Biochem Biophys Res Commun 194: 368–374
- Nagai Y, Ueno S, Saeki Y, Soga F, Hirano M, Yanagihara T (1996) Decrease of the D<sub>3</sub> dopamine receptor mRNA expression in lymphocytes from patients with Parkinson's disease. Neurology 46:791–795
- Nanko S, Sasaki T, Fukuda R, Hattori M, Dai, XY, Kazamatsuri H, Kuwata S, Juji T, Gill M (1993) A study of the association between schizophrenia and the dopamine  $D_3$  receptor gene. Hum Genet 92:336–338
- Nanko S, Fukuda R, Hattori M, Sasaki T, Dai XY, Yamaguchi K, Kazamatsuri H (1994a) Further evidence of no linkage between schizophrenia and the dopamine D<sub>3</sub> receptor gene locus. Am J Med Genet 54:264–267
- Nanko S, Ueki A, Hattori M, Dai XY, Sasaki T, Fukuda R, Ikeda K, Kazamatsuri H (1994b) No allelic association between Parkinson's disease and dopamine D<sub>2</sub>, D<sub>3</sub>, and D<sub>4</sub> receptor gene polymorphisms. Am J Med Genet 54:361–364
- Nimgaonkar VL, Zhang XR, Caldwell JG, Ganguli R, Chakravarti A (1993) Association study of schizophrenia with dopamine D<sub>3</sub> receptor gene polymorphisms: probable effects of family history of schizophrenia? Am J Med Genet 48:214–217
- Nimgaonkar VL, Sanders AR, Ganguli R, Zhang XR, Brar J, Hogge W, Fann WE, Patel PI, Chakravarti A (1996) Association study of schizophrenia and the dopamine D<sub>3</sub> receptor gene locus in two independent samples. Am J Med Genet 67: 505–514
- Nissbrandt H, Ekman A, Eriksson E, Heilig M (1995) Dopamine D<sub>3</sub> receptor antisense influences dopamine synthesis in rat brain. Neuroreport 6:573–576
- Nothen MM, Cichon S, Propping P, Fimmers R, Schwab SG, Wildenauer DB (1993) Excess of homozygosity at the dopamine D<sub>3</sub> receptor gene in schizophrenia not confirmed [letter; comment]. J Med Genet 30:708–709
- O'Hara CM, Uhland Smith A, O'Malley KL, Todd RD (1996) Inhibition of dopamine synthesis by dopamine  $D_2$  and  $D_3$  but not  $D_4$  receptors. J Pharmacol Exp Ther 277:186–192
- Pagliusi S, Chollet Daemerius A, Losberger C, Mills A, Kawashima E (1993) Characterization of a novel exon within the D<sub>3</sub> receptor gene giving rise to an mRNA isoform expressed in rat brain. Biochem Biophys Res Commun 194:465–471
- Parsons B, Stanley M, Javitch J (1993) Differential visualization of dopamine  $D_2$  and  $D_3$  receptors in rat brain. Eur J Pharmacol 234:269–272

- Parsons LH, Caine SB, Sokoloff P, Schwartz JC, Koob GF, Weiss F (1996) Neurochemical evidence that postsynaptic nucleus accumbens D<sub>3</sub> receptor stimulation enhances cocaine reinforcement. J Neurochem 67:1078–1089
- Patel J, Trout SJ, Palij P, Whelpton R, Kruk ZL (1995) Biphasic inhibition of stimulated endogenous dopamine release by 7-OH-DPAT in slices of rat nucleus accumbens. Br J Pharmacol 115:421–426
- Pilon C, Levesque D, Dimitriadou V, Griffon N, Martres MP, Schwartz JC, Sokoloff P (1994) Functional coupling of the human dopamine D<sub>3</sub> receptor in a transfected NG 108-15 neuroblastoma-glioma hybrid cell line. Eur J Pharmacol 268:129–139
- Popoli P, Pezzola A, Reggio R, Decarlis AS (1996) Evidence for the occurrence of depressant EEG effects after stimulation of dopamine D<sub>3</sub> receptors: a computerized study in rabbits. Life Sci 59:1755–1761
- Potenza MN, Graminski GF, Schmauss C, Lerner MR (1994) Functional expression and characterization of human  $D_2$  and  $D_3$  dopamine receptors. J Neurosci 14:1463–1476
- Pugsley TA, Davis MD, Akunne HC, MacKenzie RG, Shih YH, Damsma G, Wikstrom H, Whetzel SZ, Georgic LM, Cooke LW et al. (1995) Neurochemical and functional characterization of the preferentially selective dopamine D<sub>3</sub> agonist PD 128907. J Pharmacol Exp Ther 275:1355–1366
- Richfield EK, Penney JB, Young AB (1989) Anatomical and affinity state comparisons between dopamine  $D_1$  and  $D_2$  receptors in the rat central nervous system. Neuroscience 30:767–777
- Rietschel M, Nothen MM, Albus M, Maier W, Minges J, Bondy B, Korner J, Hemmer S, Fimmers R, Moller HJ, Wildenauer D, Propping P (1996) Dopamine D<sub>3</sub> receptor Gly(9)/Ser(9) polymorphism and schizophrenia: no increased frequency of homozygosity in German familial cases. Schizophr Res 20:181–186
- Rivet JM, Audinot V, Gobert A, Peglion JL, Millan MJ (1994) Modulation of mesolimbic dopamine release by the selective dopamine D<sub>3</sub> receptor antagonist, (+)-S 14297. Eur J Pharmacol 265:175–177
- Rodgers RJ, Johnson NJT, Champion AJ, Mills S (1996) Modulation of plus-maze behavior in mice by the Preferential D<sub>3</sub>-receptor agonist 7-OH-DPAT. Pharmacol Biochem Behav 54:79–84
- Rodriguez De Fonseca F, Rubio P, Martin Calderon JL, Caine SB, Koob GF, Navarro M (1995) The dopamine receptor agonist 7-OH-DPAT modulates the acquisition and expression of morphine-induced place preference. Eur J Pharmacol 274:47–55
- Sabaté O, Campion D, d'Amato T, Martres MP, Sokoloff P, Giros B, Leboyer M, Jay M, Guedj F, Thibaut F et al. (1994) Failure to find evidence for linkage or association between the dopamine D3 receptor gene and schizophrenia. Am J Psychiatry 151:107–111
- Saha N, Tsoi WF, Low PS, Basair J, Tay JS (1994) Lack of association of the dopamine D<sub>3</sub> receptor gene polymorphism (Ball) in Chinese schizophrenic males. Psychiatr Genet 4: 201–204
- Sanger DJ, Depoortere R, Perrault G (1996) Evidence for a role dopamine D<sub>3</sub> receptors in the effects of dopamine agonists on operant behaviour in rats. Behav Pharmacol 7:477–482
- Sautel F, Griffon N, Levesque D, Pilon C, Schwartz JC, Sokoloff P (1995a) A functional test identifies dopamine agonists selective for D<sub>3</sub> versus D<sub>2</sub> receptors. Neuroreport 6:329–332
- Sautel F, Griffon N, Sokoloff P, Schwartz JC, Launay C, Simon P, Costentin J, Schoenfelder A, Garrido F, Mann A, Wermuth CG (1995b) Nafadotride, a potent preferential dopamine D<sub>3</sub> receptor antagonist, activates locomotion in rodents. J Pharmacol Exp Ther 275:1239–1246
- Schmauss C (1996) Enhanced cleavage of an atypical intron of dopamine D<sub>3</sub> receptor pre mRNA in chronic schizophrenia. J Neurosci 16:7902–7909
- Schmauss C, Haroutunian V, Davis KL, Davidson M (1993) Selective loss of dopamine D<sub>3</sub>-type receptor mRNA expression in parietal and motor cortices of patients with chronic schizophrenia. Proc Natl Acad Sci USA 90:8942–8946

- Schotte A, Janssen PFM, Bonaventure P, Leysen JE (1996) Endogenous dopamine limits the binding of antipsychotic drugs to D<sub>3</sub> receptors in the rat brain: a quantitative autoradiographic study. Histochem J 28:791–799
- Schotte A, Janssen PFM, Gommeren W, Luyten WHLM, Leysen JE (1992) Autoradiographic evidence for the occlusion of rat brain dopamine D<sub>3</sub> receptors in vivo. Eur J Pharmacol 218:373–375
- Seabrook GR, Patel S, Marwood R, Emms F, Knowles MR, Freedman SB, McAllister G (1992) Stable expression of human  $D_3$  dopamine receptors in  $GH_4C_1$  pituitary cells. FEBS Lett 312:123-126
- Seabrook GR, Kemp JA, Freedman SB, Patel S, Sinclair HA, McAllister G (1994) Functional expression of human D<sub>3</sub> dopamine receptors in differentiated neuroblastoma x glioma NG108–15 cells. Br J Pharmacol 111:391–393
- Seeman P (1981) Brain dopamine receptors. Pharmacol Rev 32:229-313
- Seeman P, Grigoriadis D (1987) Dopamine receptors in brain and periphery. Neurochem Int 10:1–25
- Seeman P, Guan HC, Van Tol HH (1993) Dopamine D<sub>4</sub> receptors elevated in schizophrenia. Nature 365:441–445
- Shafer RA, Levant B (1997) FOS protein expression is induced by nafadotride, a dopamine D<sub>3</sub> receptor-preferring antogonist. Soc Neurosci Abstr 23:682
- Shaikh S, Collier DA, Sham PC, Ball D, Aitchison K, Vallada H, Smith I, Gill M, Kerwin RW (1996) Allelic association between a Ser-Gly polymorphism in the dopamine D<sub>3</sub> receptor gene and schizophrenia Hum Genet 97:714–719
- Sibley DR, Creese I (1983) Regulation of ligand binding to pitutary D-2 dopaminergic receptors. J Biol Chem 258: 4957–4965
- Sibley DR, Monsma FJ, Jr, Shen Y (1993) Molecular neurobiology of dopaminergic receptors. Int Rev Neurobiol 35:391–415
- Synder LA, Roberts JL, Sealfon SC (1991) Alternative transcripts of the rat and human dopamine D<sub>3</sub> receptor. Biochem Biophys Res Commun 180:1031–1035
- Sokoloff P, Giros B, Martres MP, Bouthenet ML, Schwartz JC (1990) Molecular cloning and characterization of a novel dopamine receptor  $(D_3)$  as a target for neuroleptics. Nature 347:146-151
- Sokoloff P, Andrieux M, Besançon R, Pilon C, Martres M-P, Giros B, Schwartz J-C (1992) Pharmacology of human dopamine D<sub>3</sub> receptor expressed in a mammalian cell line: comparison with D<sub>2</sub> receptor. Eur J Pharmacol 225:331–337
- Staley JK, Mash DC (1996) Adaptive increase in  $D_3$  dopamine receptors in the brain reward circuits of human cocaine fatalities. J Neurosci 16:6100–6106
- Stanwood GD, McElligot S, Lu L, McGonigle P (1997) Ontogeny of dopamine D<sub>3</sub> receptors in the nucleus accumbens of the rat. Neurosci Lett 223:13–16
- Starr MS, Starr BS (1995) Motor actions of 7-OH-DPAT in normal and reserpine-treated mice suggest involvement of both dopamine  $D_2$  and  $D_3$  receptors. Eur J Pharmacol 277: 151-158
- Svensson K, Carlsson A, Huff RM, Kling Petersen T, Waters N (1994a) Behavioral and neurochemical data suggest functional differences between dopamine  $D_2$  and  $D_3$  receptors. Eur J Pharmacol 263:235–243
- Svensson K, Carlsson A, Waters N (1994b) Locomotor inhibition by the D<sub>3</sub> ligand R-(+)-7-OH-DPAT is independent of changes in dopamine release. J Neural Transm Gen Sect 95:71–74
- Swarzenski BC, Tang L, Oh YJ, O'Malley KL, Todd RD (1994) Morphogenic potentials of D<sub>2</sub>, D<sub>3</sub> and D<sub>4</sub> dopamine receptors revealed in transfected neuronal cell lines. Proc Natl Acad Sci USA 91:649–653
- Tanaka T, Igarashi S, Onodera O, Tanaka H, Takahashi M, Maeda M, Kameda K, Tsuji S, Ihda S (1996) Association study between schizophrenia and dopamine D<sub>3</sub> receptor gene polymorphism. Am J Med Genet 67:366–368

- Tang L, Todd RD, Heller A, O'Malley KL (1994a) Pharmacological and functional characterization of D<sub>2</sub>, D<sub>3</sub> and D<sub>4</sub> dopamine receptor in fibroblast and dopaminergic cell lines. J Pharmacol Exp Ther 268:495–502
- Tang L, Todd RD, O'Malley KL (1994b) Dopamine  $D_2$  and  $D_3$  receptors inhibit dopamine release. J Pharmacol Exp Ther 270:475–479
- Tepper JM, Sun BC, Martin LP, Creese I (1997) Functional roles of dopamine  $D_2$  and  $D_3$  autoreceptors on nigrostriatal neurons analyzed by antisense knockdown in vivo. J Neurosci 17:2519-1530
- Todd RD (1992) Neural development is regulated by classical neurotransmitters: dopamine  $D_2$  receptor stimulation enhances neurite outgrowth. Biol Psychiatry 31:794–807
- Unis AS (1993) Concentration and distribution of [<sup>3</sup>H]-SCH23390 and [<sup>3</sup>H]-YM O9151-2 binding sites in midgestational human fetal cortex. Psychopharmacol Bull 29:415–425
- Unis AS, Dorsa DM (1993) Localization of human D<sub>3</sub> receptor mRNA in midgestational human fetal cortex by in situ hybridization. Soc Neurosci Abstr 19:1111
- Unis AS, Roberson MD, Hill K, Hamblin MW, Dorsa DW (1995) Differential localization of D<sub>2</sub> versus D<sub>3</sub> mRNA in midgestational human forebrain by in situ hybridization. Soc Neurosci Abstr 21:1620
- Uvnas Moberg K, Alster P, Hillegaart V, Ahlenius S (1995) Suggestive evidence for a DA D<sub>3</sub> receptor-mediated increase in the release of oxytocin in the male rat. Neuroreport 6:1338–1340
- Vahid-Ansari F, Robertson GS (1996) 7-OH-DPAT differentially reverses clozapine-and haloperidol-induced increases in Foslike immunoreactivity in the rodent forebrain. Eur J Neurosci 8:2605–2611
- Valerio A, Belloni M, Gorno ML, Tinti C, Memo M, Spano P (1994) Dopamine D<sub>2</sub>, D<sub>3</sub> and D<sub>4</sub> receptor mRNA levels in rat brain and pituitary during aging. Neurobiol Aging 15:713–719
- Van Tol HMM, Bunzow Jr, Guan H-C, Sunahara RK, Seeman P, Niznik HB, Civelli O (1991) Cloning the gene for a human dopamine D<sub>4</sub> receptor with high affinity for the antipsychotic clozapine. Nature 350:610–614
- Vile JM, Strange PG (1996) D<sub>2</sub> like dopamine receptors are not detectable on human peripheral blood lymphocytes. Biol Psychiatry 40:881–885

- Wang W, Hahn K-H, Bishop JF, Gao D-Q, Jose PA, Mouradian MM (1996) Up-regulation of D<sub>3</sub> dopamine receptor mRNA by neuroleptics. Synapse 23:232–235
- Waters N, Lofberg L, Haadsma Svensson S, Svensson K, Sonesson C, Carlsson A (1994) Differential effects of dopamine D<sub>2</sub> and D<sub>3</sub> receptor antagonists in regard to dopamine release, in vivo receptor displacement and behaviour. J Neural Transm [Gen Sect] 98:39–55
- Waters N, Svensson K, Haadsma Svensson SR, Smith MW, Carlsson A (1993) The dopamine D<sub>3</sub>-receptor: a postsynaptic recptor inhibitory on rat locomotor activity. J Neural Transm [Gen Sect] 94:11–19
- Werner P, Hussy N, Buell G, Jones KA, North RA (1996)  $D_2$ ,  $D_3$ , and  $D_4$  dopamine receptors couple to G protein-regulated potassium channels in Xenopus oocytes. Mol Pharmacol 49:656-661
- Wiese C, Lannfelt L, Kristbjarnarnson H, Yang L, Zoega T, Sokoloff P, Ivarsson O, Schwartz J-C, Moisses HW, Helgason T (1993) No evidence of linkage between schizophrenia and D<sub>3</sub> dopamine receptor gene locus in Icelandic pedigrees. Psychiatry Res 46:69–78
- Woodcock C, Graber SG, Rooney BC, Strange PG (1995) Expression of the rat  $D_2$  and  $D_3$  dopamine receptors in insect cells using the baculovirus system. Biochem Soc Transact 23:93s
- Xu M, Caine SB, Cooper DC, Gold LH, Graybiel AM, Hu XT, Koeltzow T, Koob GF, Moratalla R, White FJ, Tonegawa S (1995) Analysis of dopamine D<sub>3</sub> and D<sub>1</sub> receptor mutant mice. Soc Neurosci Abstr 21:363
- Yamada S, Yokoo H, Nishi S (1994) Differential effects of dopamine agonists on evoked dopamine release from slices of striatum and nucleus accumbens in rats. Brain Res 648:176–179
- Yang L, Li T, Wiese C, Lannfelt L, Sokoloff P, Xu CT, Zeng Z, Schwartz JC, Liu X, Moises HW (1993) No association between schizophrenia and homozygosity at the D<sub>3</sub> dopamine receptor gene. Am J Med Genet 48:83–86
- Yoshida N, Yoshikawa T, Hosoki K (1995) A dopamine  $D_3$  receptor agonist, 7-OH-DPAT, causes vomiting in the dog. Life Sci 57:1347-1350