ORIGINAL INVESTIGATION

The interactive effects of ketamine and nicotine on human cerebral blood flow

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Abstract

Aim The purpose of this study was to determine if acute nicotine attenuated ketamine-induced regional cerebral blood flow (rCBF).

Method Following 2–4 h of nicotine abstinence, healthy chronic smokers participated in four sets of rCBF studies, $H_2^{15}O$ positron emission tomography, during a simple sensory motor control task. The four drug conditions studied were placebo, ketamine alone, nicotine alone, and ketamine+nicotine.

Results Intravenous ketamine increased rCBF in frontal, orbital–frontal, and anterior cingulate areas. Nicotine alone induced marked rCBF elevations in the lateral occipital cortex and rCBF suppressions in the basal ganglia and anterior cingulate cortex. Nicotine added to ketamine attenuated the ketamine-induced elevated rCBF in the anterior cingulate cortex but caused a marked rCBF increase in the orbital frontal region.

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H. H. Holcomb Department of Psychiatry, Johns Hopkins University, Baltimore, MD, USA *Conclusion* This study illustrates the interactive effects of ketamine, an NMDA receptor antagonist, and nicotine in multiple brain regions. Nicotine substantially ameliorated the effects of ketamine on anterior cingulate rCBF and, when given alone, markedly suppressed anterior cingulate rCBF. The enhanced, synergistic orbitofrontal effects observed with ketamine and nicotine together suggest a marked increase in excitatory neurotransmission in a brain region often linked to psychosis, reward, and addictive behaviors.

Keywords Nicotine · Glutamate · Dopamine · PET · NMDA receptor · Cingulate · Schizophrenia · Blood flow · Frontal cortex

Introduction

Accumulating evidence indicates that hypofunction of the glutamate N-methyl-D-aspartate receptor (NMDAR) is involved in the pathophysiology of schizophrenia (Coyle et al. 2002; Krystal et al. 2003;Olney et al. 1999; Tamminga and Holcomb 2005). NMDAR antagonists, such as phencyclidine (PCP) and ketamine, induce positive, negative, and cognitive symptoms in healthy adult humans similar to that observed in schizophrenia (Krystal et al. 1994; Newcomer et al. 1999; Rowland et al. 2005a,b; Rowland 2005). Ketamine exacerbates existing or dormant symptoms in patients with schizophrenia (Lahti et al. 1995a,b; Malhotra et al. 1997). In nonhuman animal research, NMDAR antagonists induce behavioral alterations (Jentsch et al. 1997) and neurochemical changes (Adams et al. 2002; Gao et al. 1993) that translate well to features of schizophrenia. Hence, NMDAR antagonism appears to be a good model of schizophrenia and provides a framework to test novel drug agents for the treatment of schizophrenia.

Glutamatergic and cholinergic systems, specifically the NMDA and nicotinic receptors, interact in a complex manner (Deutsch et al. 2003; McGehee et al. 1995; Toth et al. 1992). Nicotine has been shown to increase frontal extracellular glutamate concentrations through the stimulation of $\alpha 4\beta 2$ nicotinic receptors (Gioanni et al. 1999). Animal studies show that nicotine ameliorates some behavioral and electrophysiological (Cromwell and Woodward 2007) effects of NMDAR antagonism (Levin et al. 1996; Tizabi et al. 1998). As such, drugs that target the nicotinic system may be good candidates to test with the NMDAR hypofunction model (Buccafusco and Terry 2009). This is supported by the implication that the nicotinic system is involved in the pathophysiology of schizophrenia. Patients exhibit a higher smoking rate, heavier smoking patterns, and extract more nicotine when smoking than the general population (Dalack et al. 1998: Strassnig et al. 2006). Postmortem studies have revealed decreased nicotinic receptor number in brains of patients with schizophrenia (Ripoll et al. 2004). Acute nicotine administration has been shown to improve negative symptoms (Smith et al. 2002) and various cognitive impairments (Depatie et al. 2002; Harris et al. 2004; Murphy and Glanzman 1999; Smith et al. 2002) in patients with schizophrenia. In another study of non-psychiatric subjects, chronic smokers and nonsmokers exhibited markedly different electrophysiological responses to intravenous ketamine (Knott et al. 2006). These studies point toward important relationships between nicotine and glutamate interactions, especially as they relate to negative symptoms and cognition.

Nicotine's action on local cerebral blood flow in human studies has demonstrated consistent results (Domino et al. 2000; London 1990; Stapleton et al. 2003; Zubieta et al. 2005). Its administration to chronic smokers reliably enhances blood flow in visual cortex and suppresses flow to the anterior cingulate cortex. In marked contrast, intravenous ketamine, which elicits robust glutamate release, promotes blood flow elevations in anterior cingulate, orbital frontal, and dorsal frontal regions (Breier et al. 1997; Holcomb et al. 2005; Rowland 2005; Vollenweider et al. 1997).

The purpose of this study was to determine if nicotine attenuated the ketamine-induced regional cerebral blood flow (rCBF) alterations in healthy humans (Holcomb et al. 2001). We hypothesized that acute nicotine would normalize the frontal rCBF elevations and diminish psychomimetic behaviors associated with NMDAR antagonism. Nicotine alone was expected to induce those patterns described above. But because nicotine's actions are diverse, we also expected to observe nicotine's indirect actions through its stimulation of dopamine release in the basal ganglia. Whereas ketamine is expected to precipitate glutamate release by blocking NMDA receptors on inhibitory GABA interneurons, dopamine released secondary to nicotine action is expected to inhibit glutamate release. This dynamic interaction, one drug-promoting glutamate release and the other suppressing it, was the focus of this study.

This study is unique in providing brain activity information arising from the interaction between glutamatergic and cholinergic systems, which are especially important to schizophrenia and drug abuse research.

Methods

The University of Maryland School of Medicine Human Institutional Review Board and the Johns Hopkins Joint Committee on Clinical Investigation approved this study.

Subjects

Nine healthy subjects (five female and four male; mean age, 30.8; right-handed; chronic smokers for 10 years or longer, five cigarettes per day or more) participated in this study. Inclusion/exclusion criteria were as follows: (1) no past or present psychiatric disorder as determined with the Structured Clinical Interview for Diagnostic and Statistical Manual of Mental Disorders, fourth edition, Non-Patient Version (Spitzer et al. 1992); (2) no first-degree relatives with a diagnosis of a psychotic disorder; (3) no current medical illnesses as determined with a physical exam and laboratory tests; and (4) no previous exposure to ketamine or PCP. All subjects gave written informed consent prior to the study and were paid for their participation.

Task description

A simple visual task, with low cognitive demand, was employed (Fig. 1). Pilot testing showed that ketamine did not disrupt accuracy or response time on this task. We expected the low demand task to provide a common behavioral state among subjects. The potential confound of performance-related rCBF activity differing across the drug conditions was minimized. For this task, subjects indicated by right- or left-hand button press the displacement side of a rectangular block on the computer screen when the "test



Fig. 1 Subjects were required to press a button in the right or left hand to indicate the displacement side of the rectangle

image" was presented following a cueing image. The hand used to press the button was congruent with the direction of the displaced object's location. The number of right and left trials were balanced within a given scan. Accuracy and reaction time were recorded for each trial.

Ketamine screening

Prior to scanning, all subjects underwent a ketamine test infusion to ensure drug tolerability. Subjects performed variations of the visual task during placebo and ketamine conditions. A test dose of nicotine spray (0.5 mg in each nostril) was administered 20 min after the ketamine infusion was stopped. This helped to demonstrate the subject's tolerance of the intranasal nicotine. Blood pressure and heart rate were monitored throughout the session.

PET scanning

Positron emission tomography (PET) procedures were conducted at the Johns Hopkins University Cyclotron Facility. We studied participants on two separate days and used radiolabeled water (H₂¹⁵O) to measure rCBF (Herscovitch et al. 1983; Raichle et al. 1983). Subjects fasted overnight and refrained from smoking for at least 2 h (2-4 h) before the study. Tobacco withdrawal symptoms were not assessed prior to the PET scans. Subjects also abstained from alcohol, caffeine, and medications overnight prior to the study. PET procedures were performed on a GE Advance Tomograph (Waukesha, WI, USA) in 3D acquisition mode. Catheters were placed in the antecubital veins of both arms: one for isotope injection and drug level blood sampling and one for i.v. ketamine/saline infusions. Transmission scans were obtained prior to tracer infusions to provide attenuation correction information: 12 mCi of $H_2^{15}O$ were injected per scan. Task performance began 20 s before isotope infusion and continued throughout each scan, which lasted 90 s. Scans were separated by 7 to 8 min except when nicotine was given. On those occasions, the interscan interval was approximately 12 min.

Placebo (saline), nicotine, ketamine, and ketamine+ nicotine conditions were assessed (Fig. 2). Four PET blood flow scans were obtained in association with each drug condition. In the analyses discussed here, only two scans from each drug condition are considered. This is due to the fact that only two of the four drug condition scans were associated with a control behavior. The other two scans were associated with a match-to-sample task that required working memory and are not presented.

During one session subjects received placebo infusion followed by intranasal nicotine administration. During the other session, subjects received ketamine infusion followed by intranasal nicotine spray. Session order, placebo-first, or ketamine-first session was pseudo-randomized and counterbalanced across subjects. Ketamine was administered in a double-blinded fashion. Ketamine was administered with a computerized infusion pump starting with a loading dose of 0.2 mg/kg over 10 min followed by a maintenance dose of 0.4 mg kg⁻¹h⁻¹. After four ketamine scans were obtained, subjects were removed from the scanner and given nicotine intranasal spray (0.5 mg/nostril=1.0 mg total); scanning resumed immediately afterward. Psychiatric assessment was completed at baseline, after the ketamine loading dose, during ketamine steady-state infusion, post nicotine, and during recovery. The rating scale consisted of the Brief Psychiatric Rating Scale (Overall and Gorham 1962). Blood samples for ketamine level determination were collected at the same time as the ratings. A second

Nicotine

Fig. 2 The number of subjects first given PET session 1 was balanced with those given session 2 initially. The entire PET session lasted about 80 min



transmission PET scan was acquired at the end of each day's session to provide additional attenuation information for image reconstruction.

Behavioral assessment

To determine if participants experienced schizophrenia-like features associated with ketamine administration, a 2 (drug group: ketamine and placebo)×5 (time: baseline, loading, steady-state, post-nicotine, and recovery) repeated measures analysis of variance (ANOVA) was performed for the BPRS total rating scores and positive and negative subscale scores. When appropriate, significant findings were investigated with follow-up tests (Table 1).

PET data analysis

All scans were realigned and spatially normalized into the stereotactic space of Talairach and Tournoux (1988). Images were smoothed with a full width at half maximum of $10 \times 10 \times 10$ mm in the *x*, *y*, and *z* planes. Pixel rCBF values were scaled using the ratio adjustment method. The image data were analyzed using Statistical Parametric Mapping (SPM99; Welcome Department of Cognitive Neurology, London, England), where voxel by voxel comparisons determined significant changes in rCBF between the drug conditions ($p \le 0.01$, corrected at the cluster level). The significance of each cluster of activated voxels was based on the magnitude of activation (z=2.36) and spatial extent (corrected, $p \le 0.05$, or uncorrected at 0.005; Friston et al. 1994, 1996; Poline et al. 1997).

Results

Behavioral and psychiatric rating scales

Three 2 (drug group: ketamine, placebo) \times 5(time: baseline, loading, steady state, post-nicotine, and recovery) repeated measures ANOVAs were performed for the BPRS total rating scores and positive and negative symptom subscale scores.

Results revealed significant interactions for the total score [F(1, 8)=7.0, p<0.05] and negative subscale [F(1, 8)=11.7, p<0.05]p < 0.05] and a trend interaction for the positive subscale [F(1, 8)=3.8, p<0.06]. These scores increased during the ketamine loading dose and steady-state time points and decreased following nicotine administration, but these scores did not change in the placebo condition. Follow-up tests revealed a significant difference between ketamine and placebo conditions during the loading dose time point [total, t (8)=3.1, p<0.05; negative symptoms, t (8)=3.6, p<0.05]. Measurements taken at the steady-state time points approached trend significance [total, t(8)=1.8, p=0.1; negative, t(8)=2.3, p=0.051]. There were no significant differences on psychiatric ratings between the ketamine+nicotine and placebo+nicotine conditions. This suggests that nicotine may attenuate the psychotomimetic reaction associated with ketamine administration. Means and standard deviations (SD) are presented in Table 2. No subject reported adverse effects or recreational ketamine-like substance use during follow-up phone contact.

The visual task elicited similar accuracy and response times across all four conditions. The accuracy ranged from 97.5% to 100%, and the reaction times ranged from 731 to 748 ms.

Ketamine and nicotine blood levels

Ketamine plasma levels were not detectable at baseline but rose to 65 ± 24 ng/ml (mean \pm SD) following the loading dose. At the mid-point of the study, immediately prior to nicotine administration, the level averaged 88 ± 22 ng/ml, and at the completion of the nicotine+ketamine phase, the blood level averaged 110 ± 29 ng/ml. Nicotine blood levels did not change significantly across conditions. In the four conditions, the blood levels ranged from 5.2 to 6.5, ±0.5 ng/ml.

PET data

Regional activations are described in terms of Brodmann areas from the Talairach atlas (Talairach and Tournoux 1988). Detailed activation results are shown in Table 2 and

Table 1 The main effect of drug group [F(1, 8)=8.7, p<0.05] was evident with ketamine administration compared to placebo

A main effect of time [F(4, 32)= 6.8, p<0.05] was also evident. BPRS scores increased during ketamine loading dose and steady-state time points and decreased following nicotine administration

| Group/rating | Baseline | Post-loading | Steady state | Post-nicotine | Recovery |
|-------------------|------------|--------------|--------------|---------------|----------|
| Placebo/nicotine | | | | | |
| BPRS positive | 5 (0) | 5 (0) | 5 (0) | 5 (0) | 5 (0) |
| BPRS negative | 4 (0) | 4 (0) | 4 (0) | 4 (0) | 4 (0) |
| BPRS total | 20.1(0.4) | 20.1 (0.4) | 20.1(0.4) | 20 (0) | 20 (0) |
| Ketamine/nicotine | | | | | |
| BPRS positive | 5.1 (0.3) | 5.7 (1.1) | 5.1 (0.3) | 5 (0) | 5 (0) |
| BPRS negative | 4 (0) | 6.1 (1.8) | 4.6 (0.7) | 4.2 (0.67) | 4 (0) |
| BPRS total | 20.2(0.45) | 23.8 (3.7) | 21.3 (2.4) | 20.2 (0.67) | 20 (0) |

Table 2 Image contrast statistics are based on a minimum cluster size of 100 voxels, above a threshold of T=2.36, p=0.01 (voxel size= $2 \times 2 \times 2$ mm, one resel=164 voxels)

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| $\begin{array}{c c c c c c c c c c c c c c c c c c c $ | Gyrus rectus (R) | 11 | 1,685 | 0.0001 | 0.0001 | 4 | 8 | 48 | -24 |
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| Decrease Hippocampus/pandippocampus (). 36 1,228 0.0001 0.0001 5.1 -34 -38 -10 Dentate nucleus (cerebellum) (R) * 5,583 0.0001 0.0001 5.1 -34 -38 -10 Increase - - - - - - - - -24 Microanes (L) 7 and 19 327 0.18 0.003 3.9 -14 -56 34 Decrease - <t< td=""><td>Middle frontal cortex (L)</td><td>9</td><td>544</td><td>0.019</td><td>0.0001</td><td>3.7</td><td>-30</td><td>48</td><td>36</td></t<> | Middle frontal cortex (L) | 9 | 544 | 0.019 | 0.0001 | 3.7 | -30 | 48 | 36 |
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| | Dentate nucleus (cerebellum) (R) | * | 5,583 | 0.0001 | 0.0001 | 5 | 24 | -50 | -24 |
| $\begin{array}{llllllllllllllllllllllllllllllllllll$ | (Ketamine+nicotine)-placebo | | | | | | | | |
| $\begin{array}{c c c c c c c c c c c c c c c c c c c $ | Increase | | | | | | | | |
| $\begin{array}{c c c c c c c c c c c c c c c c c c c $ | Orbital frontal cortex (midline) | 11 and 25 | 7,195 | 0.0001 | 0.0001 | 6.8 | -4 | 30 | -24 |
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| | Superior Temporal Gyrus (L) | 38 | 1,932 | 0.0001 | 0.0001 | 4.8 | -42 | 8 | -16 |

Fig. 3. Increased activity associated with ketamine, relative to placebo, was observed primarily in ventral frontal brain regions. Ketamine caused bilateral rCBF activation in the superior (BA 8/10/11/21), middle (BA 11), and inferior frontal (BA 47) regions as well as the insula. In the right hemisphere, orbitofrontal (BA 11) and anterior cingulate (BA 32) regions were increased. In the left hemisphere, middle temporal (BA 21) regions were increased. Decreased activity with ketamine, relative to placebo, was observed in bilateral cerebellum and hippocampal/parahippocampal systems.

Nicotine, relative to placebo, evoked increased rCBF activity bilaterally in the inferior occipital region (BA 18). Decreased rCBF activity was observed in the right medial frontal (BA 8/32), right anterior cingulate (BA 24), left

Fig. 3 Regional cerebral blood flow maps were generated by various pharmacological contrasts. Whereas nicotine's actions are predominantly suppressive and ketamine's predominantly enhancing, the two drugs together caused a marked elevation in OFC rCBF inferior frontal (BA 47), ventral striatum, left caudate/ putamen, and left cerebellum.

Ketamine+nicotine, relative to placebo, resulted in increased activity in the bilateral orbitofrontal (BA 11), right superior frontal (BA 9/10), bilateral superior temporal (BA 22/38), and right inferior temporal (BA 20). Decreased activity was observed in bilateral cerebellum, amygdala, precentral gyrus, and anterior cingulate.

Discussion

The purpose of this study was to determine whether acute nicotine administration diminishes the neural and behavioral effects of intravenous ketamine. Because ketamine



induces psychomimetic, "schizophrenia" symptoms, by eliciting asynchronous and elevated glutamatergic output, (Krystal et al. 2003; Moghaddam 2003), and because subjects with schizophrenia are disproportionately addicted to cigarettes, we asked whether nicotine might ameliorate some of ketamine's actions. The results show that nicotine affects ketamine's neural actions in two disparate ways. First, it reverses and normalizes the neural hyperactivity of the rostral and dorsal anterior cingulate cortex. Second, it enhances excitatory activity in the orbital frontal cortex. The study does not, however, permit a clear assessment of nicotine's effect on ketamine-induced behavioral changes. Although BPRS total and subscale scores diminished after nicotine was added during the ketamine infusion, this may have occurred coincidentally due to the time it was given. Psychomimetic symptoms tend to resolve with time even when ketamine elicited blood flow changes are still present and ketamine blood levels are still high (Newcomer et al. 1999). Nonetheless, this study is noteworthy in showing robust nicotine-ketamine interactions in the orbital frontal cortex (OFC) and the anterior cingulate cortex, two regions that are extensively implicated in addiction and psychiatric illness.

When given alone, intranasal nicotine administration induced marked rCBF suppression throughout the rostral and dorsal anterior cingulate cortex, the ventral striatum and the globus pallidus. The only area that showed enhanced activity with nicotine was the lateral occipital cortex. In marked contrast with the suppressant actions of nicotine, ketamine elicited widespread rCBF elevations, especially in the orbital frontal and anterior cingulate cortices. Although noteworthy, these individual changes have been described by investigators previously (Breier et al. 1997; Holcomb et al. 2001; Vollenweider et al. 1997). The interactions between nicotine and ketamine, however, have only been studied in experimental animal models (Fu et al. 2000; Rodvelt et al. 2008), not in human subjects

Ketamine+nicotine induces greater rCBF elevations in the OFC than ketamine alone. The following discussion briefly considers how nicotine, ketamine and the two drugs combined might alter glutamate release, the crucial determinant of local blood flow in the brain (Patel et al. 2003; Rothman et al. 1999; Shulman et al. 2002). The OFC, anterior cingulate, and striatum are emphasized due to their well-documented roles in addiction, reward, psychiatric illness, and planning behaviors.

Replicated by multiple investigators, nicotine's suppressant action on local brain blood flow has been documented over the last 10 years (Domino et al. 2000; Ghatan et al. 1998; Rose et al. 2003; Stapleton et al. 2003; Zubieta et al. 2005). Nicotine lowered rCBF in the anterior cingulate, orbital frontal cortex, and ventral striatum/nucleus accumbens. Although an extensive body of work has demonstrated nicotine's pharmacological actions on dopamine and glutamate release (Fu et al. 2000; Jones and Wonnacott 2004; Lambe et al. 2003; Mansvelder and McGehee 2000; Reid et al. 2000), neuroimaging studies with nicotine have not offered mechanistic models to suggest how and why nicotine would suppress rCBF in OFC, cingulate, and striatal regions. There is now overwhelming evidence for glutamate's primary role in local vascular regulation and neuroimaging signal generation (Patel et al. 2003; Rothman et al. 1999; Shulman et al. 2002). This consensus prompts us to hypothesize how nicotine and ketamine interact to modify glutamate release.

Nicotinic stimulation of presynaptic cholinergic receptors located on glutamatergic afferents in the ventral tegmental area elicits robust dopamine release in the ventral striatum and frontal cortex (Jones and Wonnacott 2004; Mansvelder and McGehee 2000). Dopaminergic modulation of glutamatergic networks in forebrain regions may inhibit (Seamans and Yang 2004) excitatory transmission. This may occur by direct inhibitory dopaminergic action on pyramidal neurons (Bandyopadhyay and Hablitz 2007; Mair and Kauer 2007) or by dopamine's activation of GABA interneurons, which subsequently inhibit pyramidal neurons (Tseng and O'Donnell 2007).

Ketamine, an NMDAR antagonist, promotes elevated blood flow and glucose metabolism secondary to enhanced glutamate release throughout the human frontal cortex (Breier et al. 1997; Holcomb et al. 2005; Vollenweider et al. 1997). For unknown reasons, ketamine-induced glutamate release is particularly robust in the OFC and ventral cingulate cortices. NMDAR antagonism has a potent effect on GABA inhibitory control of glutamatergic pyramidal neurons. By antagonizing the NMDARs on inhibitory neurons ketamine indirectly promotes excitatory neurotransmission (Farber 2003; Moghaddam 2003). The resulting elevated, asynchronous glutamatergic activity is apparently associated with diminished cognitive abilities and a worsening of psychotic symptoms in subjects with schizophrenia and induced psychotic and cognitive symptoms in healthy volunteers (Krystal et al. 1994; Lahti et al. 1995a,b; Newcomer et al. 1999).

Our study measured changes in local cerebral blood flow in subjects treated with ketamine who were given nicotine 40 min after starting the ketamine infusion. Nicotine, therefore, acted on the central nervous systems of subjects with altered glutamatergic activity. Nicotine administration may have further enhanced dopaminergic and glutamatergic release in the OFC. In this "activated" state, dopamine may have contributed to greater glutamatergic transmission. It may have amplified the state initially created by ketamine. These mixed neurotransmitter actions may have contributed to an intensely active glutamatergic state in the OFC. A previous, unpublished ketamine continuous infusion study of three healthy volunteers, by our group, followed rCBF responses out to 55 min. That pilot study did not find elevations in the OFC beyond the first 30 min of the infusion. It is, therefore, extremely unlikely that the OFC's response to ketamine–nicotine together is a "late" response to ketamine alone.

The rostral and dorsal anterior cingulate response to nicotine, during ketamine infusion, did not reflect greater glutamate transmission. Instead, the anterior cingulate had a reduction in rCBF. This suggests that nicotine may be especially potent in its inhibitory action in the anterior cingulate cortex. It also suggests that nicotine may be particularly helpful in reversing some of the attentional and memory deficits induced by NMDAR antagonism (Buccafusco and Terry 2009).

Nicotine's suppression of glutamate release in the anterior cingulate occurs in conjunction with robust glutamate suppression in the basal ganglia. This dual suppression pattern may be an important indicator of what the person addicted to nicotine is "seeking" from her smoking habit. To the extent that cortico-striate glutamate neurotransmission is controlled by nicotine, it may provide the smoker with a way to modulate her mood and anxiety state, and this may be an important reason why those with schizophrenia are especially heavy nicotine users. Excessive glutamate release may sustain schizophrenic symptoms, and nicotine acquired from cigarettes may help modulate those symptoms by inhibiting glutamate transmission in limbic cortex. In kind, the effectiveness of antipsychotic medication may depend, in part, on suppression of glutamate release in the anterior cingulate cortex (Holcomb et al. 1996; Lahti et al. 2009).

NMDAR antagonism may promote greater dopamine release in some brain regions than in others (Kosowski and Liljequist 2004; Lorrain et al. 2003). If ketamine enhances dopamine and glutamate release in the OFC more than in the anterior cingulate, then nicotine's addition may substantially facilitate dopamine's excitatory actions in the OFC. Clarity on this question will only come through animal and neuroimaging studies specifically designed to assess metabolite changes in this region under multiple drug treatment combinations created across a range of temporal epochs.

The shortcomings of this study are important. First, the results are preliminary. Data must be acquired on more subjects to substantiate these findings. Second, because subjects were given the ketamine–nicotine only when in the PET scanning apparatus, it was difficult to evaluate their subjective drug response. Furthermore, nicotine was given after ketamine's most robust behavioral actions (evident during the first 20 min) had subsided. This makes it impossible to know to what extent nicotine might directly antagonize ketamine's psychomimetic actions. The ran-

domization was incomplete. Ketamine was not given following a placebo scan, but nicotine was. This may make the comparison between placebo and nicotine more accurate than the comparison between ketamine and placebo.

The study does, however, draw attention to a fundamentally important interaction between dopamine, glutamate, and nicotine in the OFC and cingulate cortices. In light of the extensive body of work associated with these structures in the behavioral/psychiatric literature, it is reasonable to pursue additional studies that control for time of administration and the environment of the intervention.

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