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Differential sensitivity to acute administration of Ritalin, apomorphine, SCH 23390, but not raclopride in mice selectively bred for hyperactive wheel-running behavior

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Abstract *Rationale:* Previous studies of mice (*Mus domesticus*) selectively bred for high voluntary wheel running have suggested that the hyperactivity is associated with dysfunction in the dopaminergic neuromodulatory system and that high-running mice may represent a useful genetic model for attention deficit hyperactivity disorder (ADHD). *Objectives:* We tested the hypothesis that mice from the four replicate hyperactive lines would respond differently to methylphenidate (Ritalin), apomorphine (non-selective dopamine agonist), SCH 23390 (selective D1-like dopamine antagonist), and raclopride (selective D2-like dopamine antagonist) than individuals from the four replicate, randomly bred, control lines. *Methods:* After animals were habituated (3 weeks) to their cages with attached wheels, drugs were administered via intraperitoneal injections, at night, during peak wheel-running activity. Revolutions on wheels 10–70 min post-injection were used to quantify drug responses. *Results:* Ritalin (15 mg/kg and 30 mg/kg) increased wheel running in control lines but decreased running in selected lines. A low-dose (0.125 mg/kg) of apomorphine reduced wheel running by a similar amount in control and selected lines; however, higher doses of apomorphine (0.25 mg/kg and 0.5 mg/kg) produced greater reductions in wheel running in the control lines. SCH 23390 (0.025, 0.05, and 0.1 mg/kg) caused greater reductions in wheel running in control than in selected lines. Raclopride (0.5, 1, and 2 mg/kg) reduced wheel running by a similar amount in

control and selected lines. *Conclusions:* These results support the interpretation that genetically determined hyperactive wheel-running behavior is associated with altered dopaminergic function in this mouse model. More specifically, results suggest that D1-like (D1 or D5), but not D2-like (D2, D3, or D4), dopamine receptors have reduced function in the high-running mice. The fact that Ritalin decreased wheel running in selected lines further supports their use as an animal model of ADHD.

Keywords ADHD · Dopamine · Hyperactivity · Locomotor activity · Selective breeding · Wheel running

Introduction

Dopamine plays a prominent role in the voluntary control of locomotor behavior (Messier et al. 1992; Baik et al. 1995), and is implicated in drug addiction (Robinson and Berridge 1993) as well as such mental disorders as attention deficit hyperactivity disorder (ADHD) (Solanto et al. 2001). Because Ritalin increases the function of dopamine and norepinephrine (Solanto 1998) and ameliorates symptoms of ADHD, many have proposed that ADHD is caused by reduced function of dopamine (Russell et al. 1995; Sagvolden and Sergeant 1998; Papa et al. 2000) or norepinephrine (Arnsten 2001). Moreover, the dopamine hypothesis of ADHD is supported by neuroimaging (Castellanos 2001; Levy and Swanson 2001) and genetic (Cook et al. 1995; Swanson et al. 2000) studies in humans. A variety of animal models [e.g., spontaneously hypertensive rats (Sagvolden et al. 1992), dopamine transporter (DAT) knockout mice (Gainetdinov et al. 1999), dopamine-depleted rats (Shaywitz et al. 1978)] have been used in attempts to elucidate the molecular basis of altered dopamine function in ADHD.

A good animal model of ADHD (the combined or predominantly hyperactive subtype, DSM-IV, Solanto et al. 2001) should represent the key features of the disorder (Sagvolden 2000; Solanto 2000; Ferguson 2001), such as

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24-h hyperactivity in the habituated environment (Porrino et al. 1983) and motor impulsiveness (Sagvolden and Sergeant 1998). Another consideration is that ADHD may not be a discrete behavioral/physiological state, but rather represents one end of a continuum of behavior. Moreover, ADHD is likely affected by multiple environmental factors and multiple genetic loci that may interact with each other in complex ways (Todd 2000).

We have developed a unique animal model to study the neurobiology of genetically determined hyperactive wheel-running behavior and have argued that it has considerable potential as a model for ADHD (Rhodes et al. 2001). Beginning from an outbred base population, selective breeding was used to increase levels of voluntary wheel running in four replicate, genetically variable lines of house mice (Swallow et al. 1998). Four additional lines of mice were randomly bred to serve as controls. The response to selective breeding for a complex phenotype (such as voluntary activity) typically involves changes in allele frequencies at multiple loci, and these alleles may interact in complex ways to produce the phenotype (Falconer and Mackay 1996; Marley et al. 1998; Britton and Koch 2001). The (presumed) polygenic nature of high activity in our selected lines may make them a better model for ADHD than animal models that involve manipulation of single genes. Moreover, the existence of the replicate selected lines allows the possibility of multiple genetic bases for high voluntary wheel-running behavior (Garland 2003; Garland et al. 2002). Finally, because the lines are replicated, differences in drug sensitivity between the selected and control lines can truly be attributed to genetic differences that are related to the selected trait, i.e., hyperactivity (Henderson 1989, 1997). In other models that compare only one selected and one control line (e.g., the spontaneously hypertensive rat–Wistar-Kyoto comparison, Sagvolden et al. 1992), phenotypic differences may result from genes that are entirely unrelated to hyperactivity (or hypertension). This is because, when populations are initially established and then separately propagated for several generations, genetic differences arise through random processes such as genetic drift (Henderson 1989, 1997), in addition to whatever selection is imposed by the investigator.

Similarities between some of the biological properties of our selected lines and human ADHD are striking. First, each of the selected lines run further on wheels than controls by running shorter, faster bouts (Girard et al. 2001), consistent with motor impulsiveness (Sagvolden and Sergeant 1998). Second, hyperactivity is not specific to running wheels because the selected lines are also hyperactive after acclimating for 24 h to cages without wheels (Rhodes et al. 2001). Third, the dopamine reuptake blockers cocaine and GBR 12909 reduce wheel running in selected lines (Rhodes et al. 2001). In the present study, we investigated the effects of Ritalin, which shares a similar mode of action as cocaine and GBR 12909, in order to explore further the potential of the selected lines as a model of ADHD.

The aims of this study were, first, to determine whether the high wheel running in the selected mice was reduced by behaviorally appropriate doses of Ritalin, the most common drug used to treat ADHD (Solanto et al. 2001), and, second, to determine whether the D1 or D2 classes of dopamine receptors are altered in hyperactive mice by determining whether the selected-line mice respond differently to drugs that interact specifically with these receptor families.

Methods

Animals

We studied mice from generations 27 and 29 of an artificial selection experiment for high voluntary wheel-running behavior (Swallow et al. 1998). The original progenitors were outbred, genetically variable laboratory house mice (*Mus domesticus*) of the Hsd:ICR strain. After two generations of random mating, mice were randomly paired and assigned to eight closed lines (ten pairs in each). In each subsequent generation, when the offspring of these pairs were 6–8 weeks old, they were housed individually with access to a running wheel for 6 days. Daily wheel-running activity was monitored using an automated system.

In the four “selected” lines, the highest-running male and female from each family were selected as breeders to propagate the lines to the next generation. Wheel running was quantified as the total number of revolutions run on day 5 and day 6 of the 6-day test. In the four “control” lines, a male and a female were randomly chosen from each family. Within all lines, the chosen breeders were randomly paired except that sibling matings were not allowed.

To supply animals for the experiments presented here, generation-26 and -28 parents were allowed to produce a second litter (i.e., generations 27 and 29). Different groups of mice ($n=48$, 6 animals per line, 24 control and 24 selected mice) were assigned to each drug. The Ritalin and apomorphine trials were composed of generation-27 mice, whereas generation 29 was the source for the SCH 23390 and raclopride trials. Given that the selected lines appear to have been at a selection limit since approximately generation 16 (Garland 2003), cross-generation comparisons should be valid. To maintain comparability with our previous pharmacological (Rhodes et al. 2001) and locomotor (Girard et al. 2001) studies, only females were studied.

The Principles of Laboratory Animal Care (NIH publication no. 85–23, revised 1985) were followed, and all experiments were approved by the University of Wisconsin Animal Care Committee. Throughout the selection experiment and during this study, water and food [Harlan Teklad Laboratory Rodent Diet (8604); after generation 23, breeding females were given Harlan Teklad Mouse Breeder Diet (7004)] were available ad libitum. Rooms were controlled for temperature (~22°C) and photoperiod 12-h/12-h light/dark (lights on at 0400 hours, Central Standard Time).

Drug protocols and wheel running

Drug trials were conducted according to Rhodes et al. (2001). Drug responses were measured as changes in voluntary wheel running at night during peak activity. Animals were acclimated to cages with running wheels attached (Swallow et al. 1998; Rhodes et al. 2001) for approximately 3 weeks prior to drug injections (age at time of injections was approximately 3 months).

Seven drug trials were conducted. A trial consisted of a series of three intraperitoneal injections, including a vehicle control (0.9% saline) and two different drug doses. The three injections were administered every other day for a total of three injection days and were given in a volume adjusted to the body mass of the animal (5 ml/kg). 5 ml/kg was used instead of the more typical 10 ml/kg

because anecdotal evidence suggested that the reduced volume is more comfortable for the mice. Each individual received all three types of injections (vehicle, dose 1, dose 2) over the course of the three injection days (6 days total, including days off). Each mouse per line received the three injections in a different order (one of the six permutations of the three doses), such that possible effects of injection order did not need to be considered in statistical analyses (cross-over design, Milliken and Johnson 1992).

Mice were injected in the same sequence each day so that a mouse always received its injection at approximately the same time of day. It usually took less than 2 min to capture, inject, and return a mouse to its home cage. Injections began 2 h after lights off, during peak activity (unpublished observations; see also Fig. 1 in Girard et al. 2001).

Drugs were dissolved in sterile saline, then filtered through a 0.22- μ m syringe-driven filter unit (Millipore, Bedford, MA), and were prepared immediately before injections each day. Ritalin was administered at 15 mg/kg and 30 mg/kg. Apomorphine was administered in two separate trials (separated by a 1-week interval), a low-dose (0.125 mg/kg and 0.25 mg/kg) and a high-dose trial (0.5 mg/kg, and 1 mg/kg). SCH 23390 was also administered in two separate trials (0.025 mg/kg and 0.05 mg/kg then 0.05 mg/kg and 0.1 mg/kg) to extend the dose range. Similarly, raclopride was administered in two trials (0.5 mg/kg and 1.0 mg/kg then 1.0 mg/kg and 2.0 mg/kg). Doses were chosen after consulting the literature [for Ritalin (Gainetdinov et al. 1999), for apomorphine (Cabib and Puglisi-Allegra 1985), for SCH 23390 (Ichihara et al. 1993; Womer et al. 1994), for raclopride (Simon et al. 2000)]. However, in all of these studies, drugs were administered during the day and wheel running was not used to measure drug responses. Therefore, we also conducted preliminary studies to determine behaviorally equivalent doses for all four drugs.

As in our previous study (Rhodes et al. 2001), wheel rotations were monitored via computer in 1-min intervals throughout each trial. Running distance was computed as revolutions multiplied by 1.12 m, the circumference of the wheels (Swallow et al. 1998). An average running speed was calculated as total revolutions divided by number of 1-min intervals with any revolutions (Koteja and Garland 2001).

To compare responses between selected and control lines, proportional, rather than absolute changes in wheel rotations following drug injections were statistically analyzed because baseline running differed substantially between selected and control lines (by a factor of approximately three). Individual proportional responses for each drug dose were calculated by dividing total revolutions in the 10–70 min period after drug administration by total revolutions in the 10–70 min period after the saline injection. The first 10 min were not included because wheel running was significantly suppressed by the vehicle injection during this period (see Figures and Rhodes et al. 2001). Data beyond 70 min were excluded because inspection of the data suggested that the greatest effects of all the drugs occurred prior to 70 min except for Ritalin, where the greatest effect extended to 100 min in control mice. However, for Ritalin, we decided the 10–70 min was a good compromise because, in selected mice, the effect of Ritalin vanished after 50 min, and we did not wish to dilute the effect in selected mice by including 30 extra minutes of noise.

Statistics

SAS (SAS Institute Inc.) PROC MIXED (which employs Restricted Maximum Likelihood) was used to analyze the data. Line was always entered as a random effect, nested within the fixed effect of "linetype" (selected or control). The lines were separately propagated for 27 generations; therefore, individuals in a given generation do not represent independent data points and must be nested within the populations from which they arose (Henderson 1989, 1997). Degrees of freedom for testing the linetype effect were always 1 and 6. Stage of the estrus cycle was not measured and hence was not entered as a cofactor in any analyses.

Baseline wheel running was compared between selected and control lines by considering mean total revolutions during the 2 days preceding injections in a pooled analysis of all the animals. Variance was estimated separately for control and selected lines. Data were analyzed using a two-way analysis of covariance (ANCOVA), with linetype and drug trial as factors and wheel freeness (total number of revolutions produced by the wheel after being accelerated to constant velocity, an inverse measurement of how resistant the wheel is to continued rotation) as a covariate.

Proportional drug responses (total revolutions 10–70 min after drug injection divided by total revolutions 10–70 min after saline injection) were analyzed separately for each drug dose to test for differences between selected and control lines. In the cases where a dose was replicated in two separate drug trials (SCH 23390 0.05 mg/kg, raclopride 1.0 mg/kg), individual responses were averaged over the replicates prior to analysis. To improve normality of residuals, the proportional responses were transformed by raising to an exponent <1 (residuals were highly positively skewed otherwise).

Absolute responses (total revolutions 10–70 min post-injection) were analyzed separately for selected and control lines to determine the effect of the drug doses in each linetype. One-way repeated-measures ANCOVAs were used, with dose as the repeated measures factor (because the doses were applied to the same individual) and wheel freeness as the covariate. The compound symmetric covariance structure was used, which assumes that the correlation between responses within individuals is the same for all pair-wise comparisons between drug doses (e.g. correlation between responses for dose 1 and dose 2 is equal to correlation between responses for dose 1 and dose 3, within individuals).

Results

Baseline wheel running

As expected, mice from selected lines ran farther than controls throughout the study. Considering the 2 days preceding drug injections (in a pooled analysis of all the animals), selected mice ran on average 18.0 ± 0.95 km/day (least square adjusted mean \pm standard error) versus 6.4 ± 0.42 km/day in control mice ($F_{1,6} = 123.44$, $P < 0.0001$). For unknown reasons, animals in the apomorphine trial (both selected and control) ran approximately 2 km less than in other drug trials ($F_{3,172} = 5.32$, $P = 0.002$). Otherwise, average levels of wheel running 2 days preceding injections were similar among Ritalin, SCH 23390, and raclopride trials. Wheel freeness was positively correlated with running distance ($F_{1,172} = 9.24$, $P = 0.003$), but this covariate explained only a small fraction (approximately 2%) of the total variation in wheel running.

Ritalin

Selected and control mice responded differently to Ritalin (Table 1, Fig. 1). Ritalin increased running distance in control lines (one-way repeated-measures ANCOVA, $F_{2,46} = 8.73$, $P < 0.001$; Table 2, Fig. 1) but decreased it in selected lines ($F_{2,44} = 6.76$, $P = 0.003$; Table 2, Fig. 1). The decreased distance in selected mice resulted from decreased average running speed, not duration of running; whereas, in control lines, both speed and duration contributed to the increased running distance (Table 2).

Table 1 Mean proportional responses (95% confidence intervals in parentheses) during a 1-h period (10–70 min) after drug injections, shown separately for control and selected mice ($n=48$; 24 control and 24 selected mice). Individual proportional responses for each drug dose were calculated by dividing total revolutions after drug administration by total revolutions after the saline injection. Prior to statistical analyses, the proportional responses were transformed to normality by raising to an exponent <1 ; otherwise, residuals were

highly positively skewed. The F statistic and P value are for comparisons of selected with control lines and were obtained using SAS Proc Mixed, with linetype as a fixed effect, and line as a random effect. The least-squares adjusted means were then transformed back to the original scale (and LS standard errors were converted to confidence intervals, which are asymmetric about the back-transformed means); these values are shown in the table and were used for comparison and interpretation

Drug	Dose	Control	Selected	Transform	$F_{1,6}$	P value
Ritalin	15 mg/kg	1.50 (1.16,1.90)	0.91 (0.68,1.18)	$\wedge^{0.25}$	7.14	0.04
	30 mg/kg	1.25 (0.95,1.62)	0.76 (0.56,1.01)	$\wedge^{0.25}$	6.03	0.05
Apomorphine	0.125 mg/kg	0.58 (0.36,0.87)	0.73 (0.47,1.08)	$\wedge^{0.25}$	0.60	0.47
	0.25 mg/kg	0.20 (0.11,0.33)	0.62 (0.40,0.94)	$\wedge^{0.25}$	10.65	0.02
	0.5 mg/kg	0.27 (0.15,0.42)	0.64 (0.47,0.81)	$\wedge^{0.75}$	10.57	0.02
	1 mg/kg	0.14 (0.06,0.29)	0.22 (0.11,0.42)	$\wedge^{0.20}$	0.72	0.43
SCH 23390	0.025 mg/kg	0.54 (0.33,0.76)	0.87 (0.66,1.09)	None	4.63	0.07
	0.05 mg/kg	0.23 (0.13,0.38)	0.72 (0.47,1.06)	$\wedge^{0.25}$	11.55	0.01
	0.1 mg/kg	0.10 (0.03,0.21)	0.45 (0.28,0.66)	$\wedge^{0.5}$	12.33	0.01
Raclopride	0.5 mg/kg	0.89 (0.66,1.14)	0.82 (0.59,1.06)	$\wedge^{0.75}$	0.18	0.68
	1 mg/kg	0.54 (0.37,0.77)	0.69 (0.48,0.96)	$\wedge^{0.25}$	0.91	0.38
	2 mg/kg	0.40 (0.22,0.67)	0.50 (0.29,0.82)	$\wedge^{0.25}$	0.34	0.58

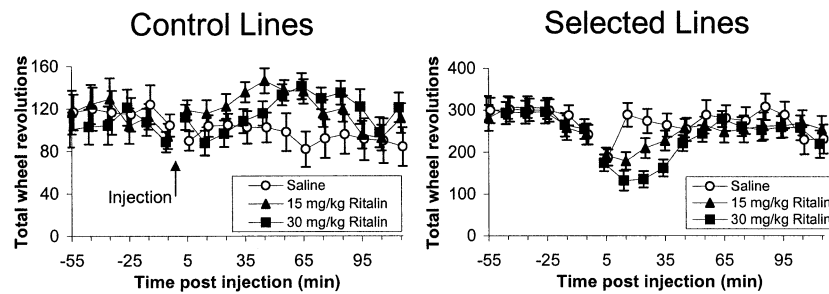


Fig. 1 Mean wheel running (revolutions) \pm SEM is plotted in 10-min increments 1 h before and 2 h after an injection of either saline or Ritalin (15 mg/kg or 30 mg/kg) ($n=24$ per data point). Data points are plotted at the mid-point of the interval (i.e., 5 on the x-axis represents the 0-min to 10-min interval). Data for control-line

mice are plotted on the left, selected on the right. The range of the y-axes differ between left and right graphs to facilitate comparison of proportional responses between control and selected mice. Ritalin increased wheel running in control lines and decreased running in selected lines

In selected lines, Ritalin had the greatest effect 10–40 min post-injection (Fig. 1), when Ritalin decreased average running speed from 2.00 ± 0.17 km/h (after saline) to 1.38 ± 0.15 km/h (after 15 mg/kg) to 1.01 ± 0.12 km/h (after 30 mg/kg). In control lines, Ritalin had the greatest effect during the 50- to 100-min time period (Fig. 1), when it increased total distance (over the 50-min time period) from 0.51 ± 0.08 km (saline) to 0.68 ± 0.05 km (15 mg/kg) to 0.74 ± 0.06 km (30 mg/kg).

Apomorphine

Apomorphine decreased wheel-running distance in both control (one-way repeated-measures ANCOVA, $F_{4,92}=21.22$, $P<0.0001$) and selected lines ($F_{4,92}=19.57$, $P<0.0001$; Table 2). However, selected mice were less sensitive to the activity-reducing effects of medium doses (0.25 mg/kg and 0.5 mg/kg) of apomorphine than controls (Table 1, Fig. 2). The lowest dose (0.125 mg/kg) decreased wheel running by approximately 40% in

control lines and 30% in selected lines, and there were no significant differences in the proportional responses between selected and control lines for this low dose (Table 1; Fig. 2A, B). However, the medium doses (0.25 mg/kg and 0.5 mg/kg) caused a proportionally greater response in control than selected lines (Table 1, Fig. 2C, D). In control mice, 0.25, 0.5, and 1 mg/kg all produced severe, comparable reductions in wheel running (approximately 80% reductions). In contrast, in selected mice, 0.5 mg/kg and 0.25 mg/kg produced only a slight reduction in wheel running (approximately 40%) comparable to the response after the 0.125-mg/kg dose. Substantial reductions in wheel running (approximately 80%) in selected mice required the highest dose, 1 mg/kg, and at this high dose there were no significant differences in the proportional response between selected and control lines (Table 1).

Table 2 Mean distance, duration, and average speed (standard errors in parentheses) run by control and selected mice in the 1-h period (10–70 min) after injection of saline or drug ($n=48$; 24 control and 24 selected mice). For distance, SAS least-square means, adjusted for wheel resistance, are shown, and the standard errors are pooled estimates from the analysis. For duration (measured as the number of 1-min intervals with at least one full wheel rotation divided by 60) and speed (distance divided by

duration), raw means and raw standard errors are shown, because these data were not statistically analyzed, but instead were used to interpret results for distance. Distance was analyzed separately for selected and control mice, with dose as a repeated-measures factor, line as a random effect, and wheel freeness (inverse measurement of wheel resistance) as a covariate. The F-statistic and *P* value refer to the effect of drug dose on running distance

Drug	Dose	Distance (km)		Duration and average speed			
		Control	Selected	Control		Selected	
				(h)	(km/h)	(h)	(km/h)
Ritalin	Saline	0.66 (0.07)	1.85 (0.15)	0.72 (0.05)	0.86 (0.08)	0.87 (0.03)	2.06 (0.16)
	15 mg/kg	0.89 (0.07)	1.55 (0.15)	0.93 (0.04)	0.92 (0.06)	0.96 (0.01)	1.60 (0.12)
	30 mg/kg	0.76 (0.07)	1.31 (0.15)	0.92 (0.03)	0.82 (0.06)	0.92 (0.03)	1.39 (0.11)
		$F_{2,46}=8.73, P<0.001$	$F_{2,44}=6.76, P=0.003$				
Apomorphine	Saline	0.50 (0.06)	1.73 (0.16)	0.55 (0.06)	0.76 (0.07)	0.85 (0.02)	2.00 (0.10)
	0.125 mg/kg	0.36 (0.06)	1.32 (0.16)	0.47 (0.06)	0.63 (0.06)	0.78 (0.04)	1.67 (0.11)
	0.25 mg/kg	0.16 (0.06)	1.29 (0.16)	0.30 (0.05)	0.35 (0.06)	0.75 (0.05)	1.57 (0.16)
	0.5 mg/kg	0.14 (0.06)	1.15 (0.16)	0.37 (0.05)	0.27 (0.05)	0.70 (0.06)	1.42 (0.15)
	1 mg/kg	0.11 (0.06)	0.62 (0.16)	0.37 (0.05)	0.22 (0.04)	0.58 (0.06)	0.84 (0.12)
		$F_{4,92}=21.22, P<0.0001$	$F_{4,92}=19.57, P<0.0001$				
SCH 23390	Saline	0.61 (0.07)	2.35 (0.22)	0.64 (0.04)	0.86 (0.07)	0.91 (0.02)	2.51 (0.16)
	0.025 mg/kg	0.37 (0.07)	2.04 (0.22)	0.40 (0.06)	0.72 (0.07)	0.81 (0.06)	2.28 (0.17)
	0.05 mg/kg	0.22 (0.07)	1.77 (0.22)	0.28 (0.05)	0.60 (0.08)	0.78 (0.04)	2.10 (0.15)
	0.1 mg/kg	0.11 (0.07)	1.16 (0.22)	0.17 (0.04)	0.37 (0.07)	0.64 (0.05)	1.64 (0.15)
		$F_{3,68}=43.59, P<0.0001$	$F_{3,65}=17.24, P<0.0001$				
Raclopride	Saline	0.56 (0.06)	2.07 (0.10)	0.66 (0.04)	0.78 (0.06)	0.93 (0.01)	2.31 (0.13)
	0.5 mg/kg	0.43 (0.06)	1.83 (0.10)	0.58 (0.05)	0.68 (0.07)	0.82 (0.06)	1.94 (0.16)
	1 mg/kg	0.35 (0.06)	1.46 (0.10)	0.52 (0.05)	0.56 (0.04)	0.87 (0.02)	1.64 (0.09)
	2 mg/kg	0.24 (0.06)	1.08 (0.11)	0.53 (0.05)	0.42 (0.04)	0.87 (0.03)	1.21 (0.08)
		$F_{3,66}=10.10, P<0.0001$	$F_{3,67}=38.03, P<0.0001$				

Raclopride

Raclopride decreased wheel running in both control ($F_{3,66}=10.10, P<0.0001$) and selected lines ($F_{3,67}=38.03, P<0.0001$; Table 2), and there were no significant differences in the proportional responses between selected and control lines (Table 1, Fig. 2E, F).

SCH 23390

SCH 23390 decreased wheel running in both control ($F_{3,68}=43.59, P<0.0001$) and selected lines ($F_{3,65}=17.24, P<0.0001$; Table 2). However, the proportional responses were significantly greater in control than in selected lines (Table 1, Fig. 2G, H), indicating that selected mice were less sensitive to the activity-reducing effects of SCH 23390.

Wheel freeness was occasionally a significant positive predictor of wheel running distance after drug injections, but explained very little of the total variation in wheel running. The random effect of line nested within linetype was not significant in any analyses.

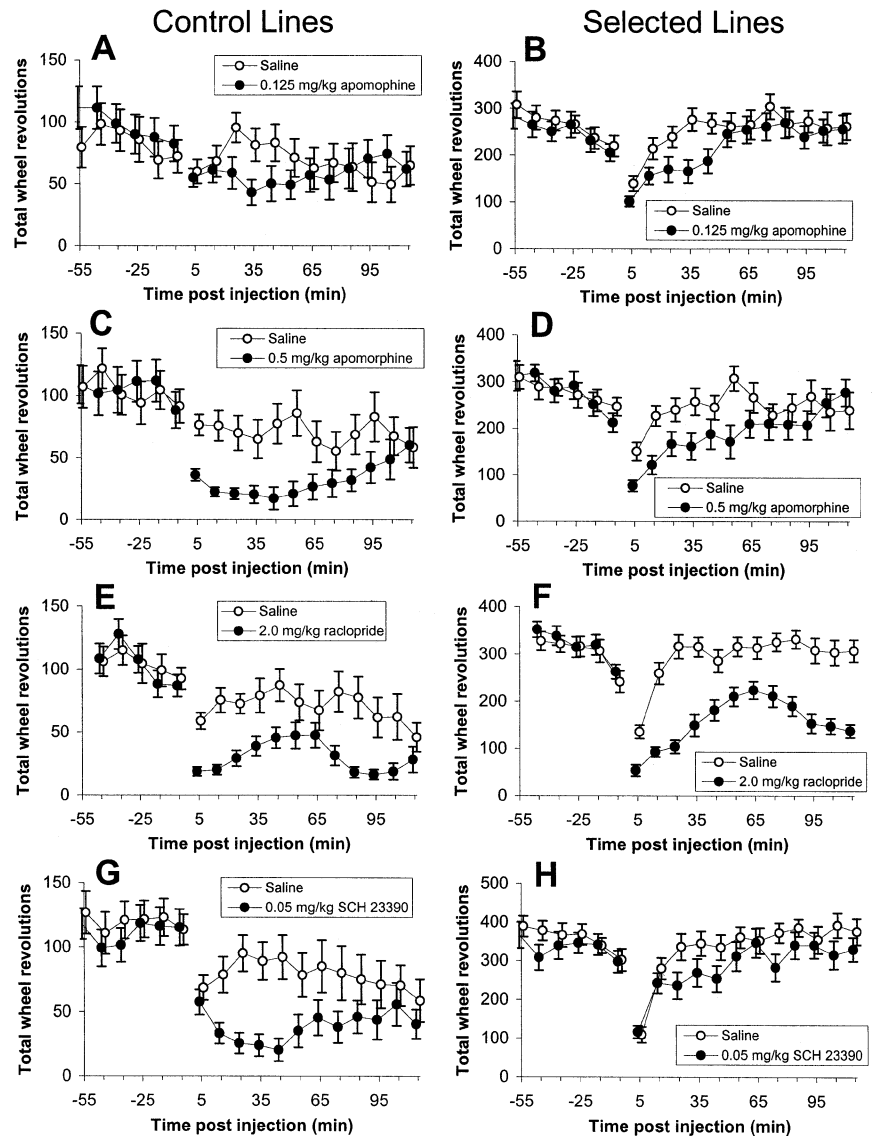
Discussion

Results support the hypothesis that genetically determined hyperactive wheel-running behavior is associated with altered dopaminergic function (Rhodes et al. 2001). The selected, hyperactive mice were less sensitive than controls to the mixed D1-/D2-like dopamine receptor agonist, apomorphine (Thal et al. 1978). Selected mice were also less sensitive than controls to the specific D1-like antagonist, SCH 23390 (Andersen 1988), but were normally sensitive to the specific D2-like antagonist, raclopride (Andersen 1988). Taken together, these results suggest that hyperactive wheel running is associated with reduced function of D1-like (D1 or D5) but not D2-like (D2, D3, or D4) dopamine receptors (see Jarvie and Caron 1993 for a review of dopamine receptor families). Reduced function of D1-like receptors in hyperactive mice could explain why drugs that facilitate dopamine function (e.g., Ritalin, cocaine, GBR 12909) ameliorate the hyperactivity (Rhodes et al. 2001). The fact that Ritalin decreased wheel running ($P=0.003$) in selected lines supports their use as a model of ADHD.

Ritalin

Ritalin might have reduced wheel-running in the selected, hyperactive lines by increasing the function of dopamine

Fig. 2 Mean wheel running (revolutions) \pm SEM is plotted in 10-min increments 1 h before and 2 h after an injection of a low dose of apomorphine (A, B), a medium dose of apomorphine (C, D), raclopride (E, F), and SCH 23390 (G, H) ($n=24$ per data point). Data points are plotted at the mid-point of the interval (i.e., 5 on the x-axis represents the 0-min to 10-min interval). Data for control-line mice are plotted in the left column, selected on the right. The range of the y-axes differ between left and right columns to facilitate comparison of proportional responses between control and selected mice. Control and selected mice responded similarly to a low dose of apomorphine (mixed D1-/D2-like agonist), but were less sensitive than controls to a medium dose of apomorphine. Selected and control mice responded similarly to raclopride (D2-like antagonist), but were less sensitive than controls to SCH 23390 (D1-like antagonist). Taken together, results suggest that D1-like but not D2-like receptors have reduced function in selected, hyperactive wheel running mice



and/or norepinephrine (Solanto 1998). Previous studies support a role for dopamine, because dopamine reuptake blockers GBR 12909 and cocaine also reduced wheel running in hyperactive but not control lines (Rhodes et al. 2001). It is possible that reduced function of D1-like receptors underlies hyperactive wheel-running, and that Ritalin corrects the deficit by increasing dopaminergic function in these regions. Reduced dopaminergic function is implicated in other animal models of genetic hyperactivity (Shaywitz et al. 1978; Russell et al. 1998). Furthermore, dopaminergic function may have been altered in selected mice in association with increased motivation (Koob 1996; Salamone 1996) for running, or possibly addiction to wheel running (Berridge and Robinson 1998; Nestler et al. 2001; Werme et al. 2002).

The doses of Ritalin chosen for this study demonstrate what can be viewed as a classic “paradoxical” effect in hyperactive versus control mice: activity was reduced in hyperactive mice but increased in control mice. This

paradoxical effect is consistent with the observation in rodents that responses to stimulant drugs (e.g., cocaine, Ritalin, amphetamine) depend on baseline level of response, with low-baseline responders being stimulated and high-baseline responders being depressed (inverse rate-dependent effect) (Sanger and Blackman 1976; Robbins and Sahakian 1979; Solanto 1998; Rhodes et al. 2001). In humans, it is widely believed that there is no paradoxical effect for clinically relevant doses of stimulants (Solanto 1998). However, this is based on one report that amphetamine decreased activity in 14 “normal” boys between the ages of 6 years and 12 years selected by the following criteria: superior school performance, good coordination and good peer relationships (Rapoport et al. 1978). More recently it was found that clinical doses of amphetamine increased activity in 12 “normal” adults (8 males and 4 females, mean age 32 years) chosen using newspaper advertisements and by word of mouth (Greenwald et al. 1998). Taken together, results are consistent

with an inverse rate-dependent effect of stimulants in humans over the clinically relevant dose range. One body-mass-scaling-equation (Morris 1999) translates the human dose range for Ritalin [0.3–0.7 mg/kg (Solanto et al. 2001)] into 2–5 mg/kg for mice. Although the doses we administered were higher, 15 mg/kg and 30 mg/kg, they were behaviorally relevant because they produced opposite effects in selected and control mice, did not reduce wheel running in selected lines to levels below controls (Fig. 1), and reduced the speed not the duration of running in selected mice (Table 2).

Reduced function of D1-like receptors in hyperactive mice

It has been proposed that wheel running is naturally rewarding and addictive (Sherwin 1998; Nestler et al. 2001; Werme et al. 2002), and recent evidence suggests that D1-like receptors play an important role in mediating neuronal plasticity associated with addiction (Beurrier and Malenka 2002). Therefore, it is not surprising that D1-like receptors are implicated in the high wheel-running behavior of the selected mice. Several mechanisms could contribute to the reduced sensitivity of selected mice to SCH 23390, including reduced expression of the D1-like receptors, altered coding sequence or post-transcriptional or post-translational modification of D1-like receptors, differences in the numerous second messengers which are down-stream from the D1-like receptors (Vallone et al. 2000), structural differences in the anatomy of dopaminergic projections in the brain, or changes in other receptor systems that interact with D1-receptor signaling (Kelley and Cadore 1988; Smith-Roe and Kelley 2000).

Agonists and antagonists

Consistent with effects of apomorphine on locomotor activity (as measured by photobeams in cages without wheels) in DBA and BALB inbred mouse strains (Cabib and Puglisi-Allegra 1985), the non-selective dopamine agonist apomorphine dose dependently reduced wheel running in both control and selected mice over a range of doses (Table 2). However, in C57BL/6, the same doses had a biphasic effect on photobeam activity, with the lowest dose decreasing activity and the highest doses increasing it (Cabib and Puglisi-Allegra 1985). Therefore, comparisons across studies using different strains of mice should be made with caution. Moreover, apomorphine can have opposite effects on different measures of locomotion in the same strain (Cabib and Puglisi-Allegra 1985). Wheel running and cage activity are different behaviors that could be mediated by different dopaminergic mechanisms (as suggested previously by Rhodes et al. 2001). We measured drug responses at night, during peak activity, after animals had acclimated to their cages with attached wheels for 3 weeks. Typically, studies employ-

ing photobeam cages measure locomotion during the day, when nocturnal rodents are normally sleeping. Moreover, animals are typically moved from their home cage to the experimental cage for the testing period. We have argued previously (Rhodes et al. 2001) that photobeam activity measured in this manner represents reactivity to a novel environment, which is different from activity in an acclimated environment.

The dopamine receptor agonist apomorphine and antagonists SCH 23390 or raclopride, seemingly opposing treatments, both reduced wheel running in selected and control mice. Agonists and antagonists of dopamine receptors have produced behaviorally similar responses in other model systems, such as cocaine-seeking behavior in rats (Weissenborn et al. 1996; Alleweireldt et al. 2002). Alleweireldt et al. (2002) attributed similar effects of dopamine agonists and antagonists, which reduce cocaine-seeking behavior, to complex interactions of receptor subtypes. Dopamine receptors can modulate excitatory or inhibitory neurons (Pennartz et al. 1992) via inhibitory or excitatory signaling pathways (Smialowski and Bijak 1987), and concurrent activation of combinations of dopamine receptors determine behavioral phenotypes (Capper-Loup et al. 2002). Therefore, it is difficult to predict the behavioral effect of an agonist (such as apomorphine) based on knowledge of an antagonist effect.

Selected, hyperactive mice, displayed significantly reduced sensitivity to apomorphine only for the medium (0.25, 0.5 mg/kg) doses, not for the low (0.125 mg/kg) or high (1 mg/kg) dose. The low dose may have been too low to affect wheel running substantially enough in control lines to detect differences between control and selected mice, and the high dose may have produced near maximum (i.e., ceiling) effects in both control and selected mice (Table 1), again obscuring differences. In addition, it is possible that the behavioral response to the low dose was primarily mediated by activation of high-affinity D2-like dopamine receptors (Vallone et al. 2000), which may not function differently in selected mice.

Summary

Results support the hypothesis that genetic hyperactivity on running wheels is associated with reduced dopamine function. Mice from lines that had been selectively bred for high wheel running were less sensitive than controls to drugs that interact with D1-like but not D2-like receptors, which suggests that hyperactive mice have reduced function of D1-like receptors in the brain. The differential response to Ritalin in control versus hyperactive mice supports the use of the selected lines as a model of ADHD. The replicate, selectively bred, hyperactive lines represent a powerful animal model to investigate the underlying neural basis of genetic hyperactivity. Future investigations are needed to investigate the possible role of other neurotransmitter systems, such as norepinephrine (Arnsten 2001). In addition, future investigations are

needed to determine which genes have changed in the selectively bred lines. Genes that affect dopamine function, in particular D1-like receptors (D1 or D5), are likely candidates.

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