# The role of oxidative stress in anticancer activity of sesquiterpene lactones

Katarzyna Gach · Angelika Długosz · Anna Janecka

Received: 11 December 2014 /Accepted: 21 January 2015 / Published online: 6 February 2015  $\oslash$  Springer-Verlag Berlin Heidelberg 2015

Abstract Sesquiterpene lactones (SLs) are plant-derived compounds that are abundant in plants of the Asteraceae family and posses a broad spectrum of biological activities, ranging from anti-inflammatory, phytotoxic, antibacterial, and antifungal to cytotoxic/anticancer. In recent years, anticancer properties of these compounds and molecular mechanisms of their action have been studied extensively on numerous cell lines and also on experimental animals. SLs have been shown to disrupt cellular redox balance and induce oxidative stress in cancer cells. Oxidative stress is associated with increased production of reactive oxygen species (ROS) which in turn can promote many aspects of cancer development and progression. On the other hand, ROS, which initiate apoptosis via the mitochondrialdependent pathway, can also be used to kill cancer cells, if they can be generated in cancer. One of the most important regulators of the redox equilibrium in the cells is reduced glutathione (GSH). In cancer cells, GSH levels are higher than in normal cells. Therefore, SL can induce apoptosis of cancer cells by decreasing intracellular GSH levels. The use of SL which can affect intracellular redox signaling pathways can be considered an interesting approach for cancer treatment. In this review, we give a brief description of the mechanisms and pathways involved in oxidative stress-induced anticancer activity of SL.

Keywords Alkylating agents . Apoptosis . GSH . α-Methylene-γ-lactones . Oxidative stress . ROS . Sesquiterpene lactones

K. Gach · A. Długosz · A. Janecka ( $\boxtimes$ ) Department of Biomolecular Chemistry, Medical University of Lodz, Mazowiecka 6/8, 92-215 Lodz, Poland e-mail: anna.janecka@umed.lodz.pl

# Introduction

Natural products isolated from plants have for centuries been and still are an important source of pharmaceutical agents with diverse chemical structures and bioactivities. Today, natural products and their derivatives or analogs still represent over 50 % of all drugs in clinical use. With cancer being the second leading cause of death worldwide, it is no wonder that new therapeutic agents to fight cancer are often being sought in the kingdom of plants.

In the past, organized collection of plants for evaluation as potential sources of new drugs resulted in the development of several important anticancer agents. One of the well-known examples of such screening programs resulted in the discovery of taxol in the bark of the Pacific Jew tree. Taxol and related compounds enhance the polymerization of tubulin to microtubules stabilizing them against depolymerization and thus interfere with the ability of cancer cells to divide. Generally, plant-derived anticancer drugs can be classified on the basis of their mechanism of action as compounds interfering with the process of mitosis (Taxol, vinca alkaloids), antioxidant agents (thymoquinone, vincristine), inhibitors of DNA modifying agents (camptothecin), and angiogenesisinhibiting agents (flavopiridol, epigallocatechin gallate).

A new group of compounds, named sesquiterpene lactones (SLs), is emerging as a promising alternative to already known anticancer therapies. In the recent years, the anticancer potential of SLs has drawn attention of chemists and pharmacologists to this interesting group of natural compounds. Several studies have provided convincing evidence of the anticancer properties of SLs on numerous human cancer cell lines (Merfort [2011;](#page-8-0) Kreuger et al. [2012](#page-8-0); Zhang et al. [2005](#page-9-0)) and also on experimental animals. But so far, only one compound in this group, arglabin, is already used as a drug in oncological clinics in Kazakhstan (Zhangabylov et al. [2004](#page-9-0)); others are in various stages of clinical or pre-clinical trials.

In this review, we concentrated on one aspect of SL anticancer activity which is their involvement in the induction of oxidative stress triggering the apoptosis process. We also discussed the differences in the sensitivity of normal versus cancer cells to SLs that may be caused by their different oxidative states.

#### Origin and structure of SLs

SLs constitute a large and diverse group of over 5000 biologically active compounds of plant origin that have been identified in several families of flowering plants, such as Cactaceae, Solanaceae, and Araceae, but are most abundant in Asteraceae (Chadwick et al. [2013](#page-7-0)). SLs were used in traditional medicine, especially for the treatment of inflammatory diseases. However, they possess a broad spectrum of other biological activities, including cytotoxic, antibacterial, antifungal, and antiviral.

SLs are 15-carbon terpenoids, consisting of three isoprene (5C) units and a lactone ring. Most, but not all of them, are characterized by an α-methylene-γ-lactone motif with an exocyclic double bond conjugated with a carbonyl function (Zhang et al. [2005\)](#page-9-0) (Fig. 1). The major categories of SLs and the examples of their important representatives are given in Table [1.](#page-2-0) The well-known examples of SLs lacking an exocyclic double bond are artemisinin (currently used as an antimalarial drug) and thapsigargin (Janecka et al. [2012](#page-8-0)).

# Mode of action of SLs

Although the exact mechanisms of action of SLs are not well understood, it is believed that  $\alpha$ -methylene- $\gamma$ -lactone group is the one responsible for their biological effects, especially for their anticancer activity. The exocyclic double bond conjugated with a carbonyl function is a strong alkylating agent and can act on transcription factors and enzymes in the human body causing steric and chemical changes that affect the ability of the targets to function appropriately. Accordingly, the methylene group of SLs reacts by the Michael-type addition



with various bionucleophiles (Fig. [2](#page-2-0)), especially mercaptyl groups of cysteine residues in proteins and in the free intracellular glutathione (GSH), leading to reduction of enzyme activity and the disruption of GSH metabolism and intracellular redox balance (Pati et al. [2007](#page-8-0); Lee et al. [1977\)](#page-8-0). Such alkylation of cellular thiols disrupts the key biological processes (Zhang et al. [2004,](#page-9-0) [2005](#page-9-0); Heilmann et al. [2001](#page-8-0); Knight [1995](#page-8-0)) resulting in the controlled cell death, apoptosis. SLs were shown in vitro to induce apoptosis, inhibit cell cycle and proliferation, and diminish metastasis in various cancer cell lines (Zhang et al. [2005](#page-9-0); Janecka et al. [2012\)](#page-8-0). Conducted research showed that cytotoxic activity of SLs involves different signaling pathways and affects multiple targets in cancer cells (Janecka et al. [2012;](#page-8-0) Kreuger et al. [2012\)](#page-8-0). Emerging data suggest that the underlying mechanism for anti-tumor effects of SLs seems to be mediated by oxidative stress (Wen et al. [2002\)](#page-9-0). The SL-induced apoptosis, described in many cancer cell lines, was found to be associated with reduced glutathione (GSH) depletion, reactive oxygen species (ROS) generation, mitochondrial transmembrane potential dissipation, cytochrome c release, and activation of caspases (caspase 9 and 3) (Wen et al. [2002](#page-9-0); Khan et al. [2012\)](#page-8-0).

It seems that the exo-methylene group conjugated with a carbonyl in a lactone ring is essential for the cytotoxic function of SLs. However, other factors, such as lipophilicity, molecular geometry, and the chemical environment of the target sulfhydryl group can also influence the activity of SLs (Beekman et al. [1997](#page-7-0); Scotti et al. [2007](#page-8-0)). The differences in activity can be also explained by different numbers of alkylating structural elements. Compounds with two alkylating centers are termed "bifunctional." Parthenolide (PTL), which in addition to its  $\alpha$ -methylene- $\gamma$ -lactone moiety also contains an epoxide, is an example of such bifunctional structure, with two sites for potential nucleophilic attack. Another example is helenalin, with an additional endocyclic α,β-unsaturated ketone (Lee and Furukawa [1972\)](#page-8-0). In general, compounds with two alkylating centers are more potent inhibitors of tumor cell proliferation but sometimes are also more toxic.

The exocyclic double bond is also responsible for such effects as regulation of gene expression by activating and deactivating transcription (Wong and Menendez [1999;](#page-9-0) Mazor et al. [2000](#page-8-0); Schomburg et al. [2013](#page-8-0)).

#### Parthenolide—the most studied sesquiterpene lactone

Parthenolide (PTL) is the major SL found in feverfew (Tanacetum parthenium), a herbal plant used in traditional medicine. For centuries, PTL was known for its antiinflammatory activity (Mathema et al. [2012\)](#page-8-0), but the more recent evidence points at its potential as a novel anti-tumor Fig. 1 The general structure of α-methylene-γ-lactones agent. PTL was shown to induce cytotoxicity in various

#### <span id="page-2-0"></span>Table 1 Sesquiterpene lactones with anticancer potential



cancer cell types, including prostate (Hayashi et al. [2011](#page-7-0)), pancreatic (Liu et al. [2010\)](#page-8-0), breast (Nakshatri et al. [2004\)](#page-8-0) and colorectal cancers (Zhang et al. [2004](#page-9-0)), multiple myeloma



Fig. 2 The mode of action of sesquiterpene lactones with the exocyclic methylene group conjugated with carbonyl function

(Suvannasankha et al. [2008\)](#page-9-0), and leukemia (Zunino et al. [2007](#page-9-0)). More recently, it has been demonstrated in in vivo studies using mice xenografts of childhood acute lymphoblastic leukemia that PTL eliminates leukemia-initiating cell populations (LICs) and improves survival and restoration of normal murine hemopoiesis (Diamanti et al. [2013](#page-7-0)).

Anticancer effect of PTL involves inhibition of proliferation, induction of apoptosis, cell cycle arrest, and inhibition of metastasis (Janecka et al. [2012;](#page-8-0) Koprowska and Czyz [2010;](#page-8-0) Zhang et al. [2005\)](#page-9-0). The broad array of biological activities exerted by PTL is due to its ability to bind proteins that are overexpressed in various pathophysiological states, including cancer. The molecular mechanisms of PTL action are strongly associated with DNA-binding inhibition of two transcription factors, the nuclear factor κB (NF-κB) (Rüngeler et al. [1999](#page-8-0); Steele et al. [2006\)](#page-8-0) and the signal transducer and activator of transcription 3 (STAT3), as well as the proapoptotic activation of p53, together with reduced glutathione (GSH) depletion (D'Anneo et al. [2013\)](#page-7-0), reactive oxygen species (ROS) generation (Wen et al. [2002;](#page-9-0) Zunino et al. [2007](#page-9-0); Guzman et al. [2005\)](#page-7-0), and c-Jun N-terminal kinase (JNK) activation (Nakshatri et al. [2004](#page-8-0)). PTL decreases also the activity of histone deacetylase (HDAC) (Gopal et al. [2007](#page-7-0)) and DNA methyltranspherase 1 (DNMT1) (Liu et al. [2009\)](#page-8-0). Furthermore, PTL can interfere with microtubule function through tubulin binding (Fonrose et al. [2007\)](#page-7-0). However, the main mechanism of PTL action is linked to the inhibition of NF-κB (Rüngeler et al. [1999\)](#page-8-0). NF-κB is an extracellular signal-activated transcription factor that governs the expression of various important genes involved in cytokine production, cell proliferation and differentiation, cellular adhesion, inflammatory processes and apoptosis (Karin and Lin [2002\)](#page-8-0). Constitutive activation of NF-κB is a relatively common feature of many cancers, including leukemias (Guzman et al. [2001\)](#page-7-0) and mediates the cellular transformation, proliferation, invasion, angiogenesis, and metastasis of cancer (Fujioka et al. [2003\)](#page-7-0). Antiapoptotic activity of NF-κB is one of the major mechanisms of cancer cell resistance to chemotherapy and radiation (Montagut et al. [2006\)](#page-8-0). NF-κB is localized in the cytoplasm and consists of two subunits, p50 and p65, which are inactive due to their association with the inhibitory protein IκB-α. PTL inhibits NF-κB activity by preventing IκB-α degradation and p50 and p65 NF-κB subunit modification. Blocking NF-κB can cause tumor cells to stop proliferating, to die, or to become more sensitive to the action of anti-tumor agents. Thus, a very important new direction in PTL-based studies is sensitization tumor cells to antineoplastic agents (Wyrebska et al. [2014\)](#page-9-0).

However, the major drawback that restricts efficacy of PTL as a drug is its poor solubility in water. To overcome the solubility problem, a series of PTL derivatives was obtained through the diastereoselective addition of several primary and secondary amines to the exocyclic double bond (Nasim and Crooks [2008;](#page-8-0) Neelakantan et al. [2009\)](#page-8-0). Based on the favorable pharmacokinetic and pharmacodynamic properties, N,Ndimethylaminoparthenolide (DMAPT) (Fig. [3\)](#page-4-0) with improved solubility and bioavailability was selected as a lead compound (Neelakantan et al. [2009](#page-8-0); Guzman et al. [2007](#page-7-0)). When formulated as a fumarate salt, DMAPT demonstrated more than 1000-fold greater solubility in water than PTL and maintained the anticancer activity of the parent compound because in body fluids, DMAPT is rapidly converted back to PTL. In

vitro studies on cancer cells confirmed that DMAPT maintains basic characteristics of its parent compound with the ability to generate ROS, activate p53 protein as well as inhibit NF-κB DNA binding and proliferation of cancer cells. Recently, inhibition of STAT3 signaling pathway was also highlighted in anticancer activity of DMAPT (Song et al. [2014\)](#page-8-0). DMAPT was shown to eradicate in vitro stem and progenitor cells of acute myeloid leukemia (AML). Pharmacologic experiments using both mouse xenograft models and spontaneous acute canine leukemias demonstrated its high in vivo bioactivity (Guzman et al. [2007\)](#page-7-0). Further in vivo studies demonstrated that DMAPT significantly suppressed the growth of prostate (Shanmugam et al. [2010](#page-8-0)), lung, bladder (Shanmugam et al. [2011](#page-8-0)), and breast cancers (D'Anneo et al. [2013](#page-7-0)). Moreover, it inhibited metastasis in mouse xenograft model of breast cancer and enhanced survival of the treated mice.

# Reactive oxygen species

ROS are chemically reactive molecules containing oxygen. It is a collective term used for oxygen-derived free radicals such as superoxide anion, hydroxyl, peroxyl, and alkoxyl, as well as  $O<sub>2</sub>$ -derived non-radical species such as hydrogen peroxide, singlet oxygen, alkyl peroxide, etc. (Fig. [4\)](#page-4-0) (Halliwell and Cross [1994\)](#page-7-0). ROS are produced intracellularly by eukaryotic cells during normal oxidative metabolism through multiple mechanisms. Depending on the cell and tissue type, the major sources of ROS are NADPH oxidase (NOX) complexes in the cell membranes, mitochondria, peroxisomes, and endoplasmic reticulum.

At physiological low levels, ROS function as "redox messengers" for normal signaling transduction. Recently, the role of ROS in regulation of different cellular processes, such as gene expression, metabolism, cell differentiation, proliferation, and apoptosis, has been emphasized. However, at higher concentrations, ROS become toxic and induce cell depth by various signaling pathway.

# Oxidative stress

Oxidative stress reflects an imbalance between the intracellular production of ROS and a cell ability to readily detoxify the reactive intermediates or to repair the resulting damage (Betteridge [2000](#page-7-0); Sharma et al. [2007](#page-8-0)). The balance between ROS formation and anti-oxidative defense is crucial for normal cellular function and determines the fate of the cell. Cellular redox balance is maintained by a powerful cell's endogenous antioxidant systems, including glutathione and thioredoxin and antioxidant enzymes such as superoxide dismutase, catalase, etc., which are able to eliminate ROS. Disturbance in the redox balance and excess ROS induce lipid

<span id="page-4-0"></span>Fig. 3 The structure of parthenolide and micheliolide and their water-soluble derivatives— DMAPT and DMAMCL, respectively



and protein oxidative modifications and DNA damage, leading to apoptotic cell death or cancerogenic cell transformation (Valko et al. [2007](#page-9-0)). Therefore, oxidative stress is involved in most of pathological states and diseases, including cancer. Recent evidence has demonstrated that enhanced ROS generation and oxidative stress may trigger cell transformation and contribute to cancer progression by increasing DNA mutations or inducing DNA damage and amplifying genomic instability (Reuter et al. [2010](#page-8-0); Visconti and Grieco [2009](#page-9-0)).

It is well documented that cancer cells contain higher level of endogenous ROS than normal cells. Increased generation of ROS in cancer cells is caused largely by the products of their highly metabolic nature. When compared with the normal cells, cancer cells are in a persistent prooxidative state that can lead to intrinsic oxidative stress (Szatrowski and Nathan [1991;](#page-9-0) Toyokuni et al. [1995](#page-9-0)). The higher levels of endogenous ROS in cancer cells increase their susceptibility to oxidative stress-induced cell death and could be exploited for cancer therapy (Nogueira and Hay [2013\)](#page-8-0). Chemotherapeutic agents



#### Cellular redox system

Cellular redox balance is maintained by a powerful antioxidant system that includes enzymes, such as superoxide dismutase, catalase, glutathione peroxidase, or glutathione reductase, and non-enzymatic scavengers, such as glutathione, flavonoids, and minerals (Se, Mn, Cu, and Zn) (Irshad and Chaudhuri [2002\)](#page-8-0). The tripeptide glutathione (γ-glutamyl-cysteinyl-glycine, GSH) is the most abundant intracellular free thiol in eukaryotic







extracellular

Fig. 5 Mechanism of enzymatic GSH oxidation and GSSG reduction



<span id="page-5-0"></span>

cells that plays the major role in maintaining the intracellular redox status and defense against oxidative stress (Circu and Aw [2008](#page-7-0)). Reduced GSH is the biologically active form which is oxidized to glutathione disulfide (GSSG) during oxidative stress. Typically, cells exhibit a high ratio of GSH to GSSG and more than 90 % of total GSH is maintained in a reduced

<span id="page-6-0"></span>

Fig. 6 Schematic mechanism of PTL action in the cell

form through de novo GSH synthesis, enzymatic reduction of GSSG, or exogenous GSH uptake (Circu and Aw [2010\)](#page-7-0) (Fig. [5](#page-4-0)). A shift in the cellular GSH/GSSG ratio constitutes an important signal that could decide the fate of a cell. Depletion of intracellular GSH concentration results in oxidative stress and increased formation of ROS which initiate apoptosis via the mitochondria-dependent pathway (Circu and Aw [2008\)](#page-7-0). On the contrary, elevated GSH levels afford protection against stress-induced apoptosis (Friesen et al. [2004](#page-7-0)) and have been associated with drug resistance to chemotherapy.

#### Sesquiterpene lactones as oxidative stress inducers

It has been shown in several studies that anticancer activity of PTL and other SLs is associated with their ability to induce oxidative stress in cancer cells. Numerous cellular factors and signaling pathways linked to oxidative stress were already described in various cancer cell lines (Table [2\)](#page-5-0). Even though many of them were discussed in detail, the overall molecular mechanism of oxidative stress-induced cell death is still under investigation.

Recently, D'Anneo and co-workers (D'Anneo et al. [2013\)](#page-7-0) described the possible mechanism of the oxidative stress-

Fig. 7 Redox state alternations during cancer progression

induced effects of PTL in MDA-MB231 cells. In the time course experiment of ROS generation, they showed that the level of ROS rapidly increased in the first hours (1–3 h) of the treatment with PTL, when the drug induced the production of superoxide anion by stimulation of NOX activity. In this phase, the increased ROS generation caused activation of JNK and inhibition of NF-κB activity. During the prolonged PTL treatment (5–20 h), ROS generation led to the dissipation of mitochondrial membrane potential and the appearance of necrotic events (Fig. 6).

Agents targeting the redox state of cancer cells by GSH depletion may be considered for the development of specific anticancer therapies. In tumor cells exposed to SLs, reduction of intracellular GSH levels has been observed both in vitro and in vivo (Woerdenbag et al. [1989](#page-9-0)). In the study of Wen (Wen et al. [2002](#page-9-0)), it was shown that intracellular GSH depletion in hepatoma cell lines is crucial for triggering the oxidative stress-mediated apoptosis after PTL treatment. The authors suggested also that the tumor cell sensitivity to PTL seemed to correlate well with GSH metabolism.

Although most attention has been focused on the intracellular redox state as a target for cancer therapy, the recent studies suggest that extracellular redox-related proteins may be potential therapeutic targets for cancer treatment (Chaiswing and Oberley [2010](#page-7-0)). Extracellular redox state is an important factor that affects cell behavior and plays an important role in the regulation of critical cellular functions and microenvironmental cell interactions (Chaiswing and Oberley [2010;](#page-7-0) Skalska et al. [2009\)](#page-8-0). For example, the redox status of exofacial thiols on T cells regulates their activation and proliferation (Gelderman et al. [2006\)](#page-7-0). Extracellular redox environment is a dynamic state determined by several known variables, including redox-modulating proteins and extracellular protein thiol groups (i.e., exofacial thiols), and is sculpted by intracellular metabolism. Under physiological conditions, the extracellular space is relatively more oxidized than the interior



<span id="page-7-0"></span>of the cell. During pathologic conditions, such as cancer, the extracellular redox state may be altered (Chaiswing and Oberley 2010) (Fig. [7\)](#page-6-0). Recent evidence suggests that in contrast to the oxidized extracellular state of normal cells, malignant cells exist in a reduced environment (Nogueira and Hay [2013\)](#page-8-0). Different cancer cell types or stages may demonstrate different and unique extracellular redox states. Therefore, the redox state of the extracellular micro-environment may be important in terms of the response of cancer cells to chemotherapy. It has been shown that PTL can modulate the redox state of cancer cell exofacial thiols and induce selective apoptosis in acute myelogenous leukemia (AML) and chronic lymphocytic leukemia (CLL) cells without affecting normal hematopoietic stem cells or T cells (Steele et al. [2006;](#page-8-0) Guzman et al. 2005). It seems that the selectivity of PTL against cancer cells might be due to the difference in extracellular redox state between normal and cancer cells. Exofacial thiols are reduced on cancer cells and can readily interact with PTL but are oxidized on normal cells thus unavailable for interaction with PTL (Skalska et al. [2009\)](#page-8-0).

Detailed in vitro investigations of the anti-leukemic activity of PTL and its water-soluble analog DMAPT have been carried out against cultured primary AML, CML, and CLL cells and demonstrated potent eradication of leukemic stem and progenitor cells, as well as the overall blast population of both myeloid and lymphoid leukemias. It has also been shown that DMAPT specifically ablated the primitive human leukemia cells without impairing their normal counterparts (Neelakantan et al. [2009;](#page-8-0) Guzman et al. 2007; Zhang et al. [2012\)](#page-9-0). Similar selectivity in anticancer action was observed with another representative of SLs, micheliolide (Fig. [3\)](#page-4-0) and its water-soluble derivative dimethylaminomicheliolide (DMAMCL) (Zhang et al. [2012\)](#page-9-0).

It seems that reduction of cell surface protein free thiols on cancer cells is emerging as a novel mechanism of SL action. DMAPT has already entered clinical trials in the UK for the treatment of AML, ALL, and CLL (Kevin [2010](#page-8-0)).

Acknowledgments This work was financed by the Medical University of Lodz (No 502-14-191 to KG and No 503/1-156-02/503-01) and by the Ministry of Science and Higher Education (Project No. N N204 005736).

Conflict of interest None declared.

### **References**

- Beekman AC, Woerdenbag HJ, van Uden W, Pras N, Konings AW, Wikström HV, Schmidt TJ (1997) Structure-cytotoxicity relationships of some helenanolide-type sesquiterpene lactones. J Nat Prod 60:252–257
- Berges C, Fuchs D, Opelz G, Daniel V, Naujokat C (2009) Helenalin suppresses essential immune functions of activated  $CD4<sup>+</sup>$  T cells by multiple mechanisms. Mol Immunol 46:2892–2901
- Betteridge DJ (2000) What is oxidative stress? Metabolism 49(2 Suppl 1):3–8
- Chadwick M, Trewin H, Gawthrop F, Wagstaff C (2013) Sesquiterpenoids lactones: benefits to plants and people. Int J Mol Sci 14:12780–12805
- Chaiswing L, Oberley TD (2010) Extracellular/microenvironmental redox state. Antioxid Redox Signal 13:449–465
- Choi JH, Lee KT (2009) Costunolide-induced apoptosis in human leukemia cells: involvement of c-jun N-terminal kinase activation. Biol Pharm Bull 32:1803–1808
- Circu ML, Aw TY (2008) Glutathione and apoptosis. Free Radic Res 42: 689–706
- Circu ML, Aw TY (2010) Reactive oxygen species, cellular redox systems, and apoptosis. Free Radic Biol Med 48:749–762
- D'Anneo A, Carlisi D, Lauricella M, Puleio R, Martinez R, Di Bella S, Di Marco P, Emanuele S, Di Fiore R, Guercio A, Vento R, Tesoriere G (2013) Parthenolide generates reactive oxygen species and autophagy in MDA-MB231 cells. A soluble parthenolide analogue inhibits tumour growth and metastasis in a xenograft model of breast cancer. Cell Death Dis 4:e891
- Diamanti P, Cox CV, Moppett JP, Blair A (2013) Parthenolide eliminates leukemia-initiating cell populations and improves survival in xenografts of childhood acute lymphoblastic leukemia. Blood 121:1384– 1393
- Fang J, Seki T, Maeda H (2009) Therapeutic strategies by modulating oxygen stress in cancer and inflammation. Adv Drug Deliv Rev 61: 290–302
- Fonrose X, Ausseil F, Soleilhac E, Masson V, David B, Pouny I, Cintrat JC, Rousseau B, Barette C, Massiot G, Lafanechère L (2007) Parthenolide inhibits tubulin carboxypeptidase activity. Cancer Res 67:3371–3378
- Friesen C, Kiess Y, Debatin KM (2004) A critical role of glutathione in determining apoptosis sensitivity and resistance in leukemia cells. Cell Death Differ 11(Suppl 1):S73–S85
- Fujioka S, Sclabas GM, Schmidt C, Frederick WA, Dong QG, Abbruzzese JL, Evans DB, Baker C, Chiao PJ (2003) Function of nuclear factor kappaB in pancreatic cancer metastasis. Clin Cancer Res 9:346–354
- Gelderman KA, Hultqvist M, Holmberg J, Olofsson P, Holmdahl R (2006) T cell surface redox levels determine T cell reactivity and arthritis susceptibility. Proc Natl Acad Sci U S A 103:12831–12836
- Gopal YN, Arora TS, Van Dyke MW (2007) Parthenolide specifically depletes histone deacetylase 1 protein and induces cell death through ataxia telangiectasia mutated. Chem Biol 14:813–823
- Guzman ML, Neering SJ, Upchurch D, Grimes B, Howard DS, Rizzieri DA, Luger SM, Jordan CT (2001) Nuclear factor-kappaB is constitutively activated in primitive human acute myelogenous leukemia cells. Blood 98:2301–2307
- Guzman ML, Rossi RM, Karnischky L, Li X, Peterson DR, Howard DS, Jordan CT (2005) The sesquiterpene lactone parthenolide induces apoptosis of human acute myelogenous leukemia stem and progenitor cells. Blood 105:4163–4169
- Guzman ML, Rossi RM, Neelakantan S, Li X, Corbett CA, Hassane DC, Becker MW, Bennett JM, Sullivan E, Lachowicz JL, Vaughan A, Sweeney CJ, Matthews W, Carroll M, Liesveld JL, Crooks PA, Jordan CT (2007) An orally bioavailable parthenolide analog selectively eradicates acute myelogenous leukemia stem and progenitor cells. Blood 110:4427–4435
- Halliwell B, Cross CE (1994) Oxygen-derived species: their relation to human disease and environmental stress. Environ Health Perspect 102(Suppl 10):5–12
- Hayashi S, Koshiba K, Hatashita M, Sato T, JujoY SR, Tanaka Y, Shioura H (2011) Thermosensitization and induction of apoptosis or cellcycle arrest via the MAPK cascade by parthenolide, an NF-κB inhibitor, in human prostate cancer androgen-independent cell lines. Int J Mol Med 28:1033–1042
- <span id="page-8-0"></span>Heilmann J, Wasescha MR, Schmidt TJ (2001) The influence of glutathione and cysteine levels on the cytotoxicity of helenanolide type sesquiterpene lactones against KB cells. Bioorg Med Chem 9:2189– 2194
- Irshad M, Chaudhuri PS (2002) Oxidant-antioxidant system: role and significance in human body. Indian J Exp Biol 40:1233–1239
- Itoh T, Ito Y, Ohguchi K, Ohyama M, Iinuma M, Otsuki M, Nozawa Y, Akao Y (2008) Eupalinin A isolated from Eupatorium chinense L. induces autophagocytosis in human leukemia HL60 cells. Bioorg Med Chem 16:721–731
- Itoh T, Ohguchi K, Nozawa Y, Akao Y (2009) Intracellular glutathione regulates sesquiterpene lactone-induced conversion of autophagy to apoptosis in human leukemia HL60 cells. Anticancer Res 29:1449– 1457
- Janecka A, Wyrębska A, Gach K, Fichna J, Janecki T (2012) Natural and synthetic α-methylenelactones and α-methylenelactams with anticancer potential. Drug Discov Today 17:561–572
- Karin M, Lin A (2002) NF-kappaB at the crossroads of life and death. Nat Immunol 3:221–227
- Kevin P (2010) New agents for the treatment of leukemia: discovery of DMAPT (LC-1). Drug Discov Today 15:322
- Khan M, Yi F, Rasul A, Li T, Wang N, Gao H, Gao R, Ma T (2012) Alantolactone induces apoptosis in glioblastoma cells via GSH depletion, ROS generation, and mitochondrial dysfunction. IUBMB Life 64:783–794
- Khan M, Li T, Ahmad Khan MK, Rasul A, Nawaz F, Sun M, Zheng Y, Ma T (2013) Alantolactone induces apoptosis in HepG2 cells through GSH depletion, inhibition of STAT3 activation, and mitochondrial dysfunction. Biomed Res Int 2013:719858
- Knight DW (1995) Feverfew: chemistry and biological activity. Nat Prod Rep 12:271–276
- Koprowska K, Czyz M (2010) Molecular mechanisms of parthenolide's action: old drug with a new face. Postepy Hig Med Dosw (Online) 64:100–114
- Kreuger MR, Grootjans S, Biavatti MW, Vandenabeele P, D'Herde K (2012) Sesquiterpene lactones as drugs with multiple targets in cancer treatment: focus on parthenolide. Anticancer Drugs 23:883–896
- Lee KH, Furukawa H (1972) Antitumor agents. 3. Synthesis and cytotoxic activity of helenalin amine adducts and related derivatives. J Med Chem 15:609–611
- Lee KH, Hall IH, Mar EC, Starnes CO, ElGebaly SA, Waddell TG, Hadgraft RI, Ruffner CG, Weidner I (1977) Sesquiterpene antitumor agents: inhibitors of cellular metabolism. Science 196:533–536
- Lee MG, Lee KT, Chi SG, Park JH (2001) Costunolide induces apoptosis by ROS-mediated mitochondrial permeability transition and cytochrome C release. Biol Pharm Bull 24:303–306
- Liu Z, Liu S, Xie Z, Pavlovicz RE, Wu J, Chen P, Aimiuwu J, Pang J, Bhasin D, Neviani P, Fuchs JR, Plass C, Li PK, Li C, Huang TH, Wu LC, Rush L, Wang H, Perrotti D, Marcucci G, Chan KK (2009) Modulation of DNA methylation by a sesquiterpene lactone parthenolide. J Pharmacol Exp Ther 329:505–514
- Liu JW, Cai MX, Xin Y, Wu QS, Ma J, Yang P, Xie HY, Huang DS (2010) Parthenolide induces proliferation inhibition and apoptosis of pancreatic cancer cells in vitro. J Exp Clin Cancer Res 29:108
- Mathema VB, Koh YS, Thakuri BC, Sillanpää M (2012) Parthenolide, a sesquiterpene lactone, expresses multiple anti-cancer and antiinflammatory activities. Inflammation 35:560–565
- Mazor RL, Menendez IY, Ryan MA, Fiedler MA, Wong HR (2000) Sesquiterpene lactones are potent inhibitors of interleukin 8 gene expression in cultured human respiratory epithelium. Cytokine 12: 239–245
- Merfort I (2011) Perspectives on sesquiterpene lactones in inflammation and cancer. Curr Drug Targets 12:1560–1573
- Montagut C, Tusquets I, Ferrer B, Corominas JM, Bellosillo B, Campas C, Suarez M, Fabregat X, Campo E, Gascon P, Serrano S, Fernandez L, Rovira A, Albanell J (2006) Activation of nuclear factor-κB is

linked to resistance to neoadjuvant chemotherapy in breast cancer patients. Endocr Relat Cancer 13:607–616

- Nakshatri H, Rice SE, Bhat-Nakshatri P (2004) Antitumor agent parthenolide reverses resistance of breast cancer cells to tumour necrosis factor-related apoptosis-inducing ligand through sustained activation of c-Jun N-terminal kinase. Oncogene 23:7330–7344
- Nasim S, Crooks PA (2008) Antileukemic activity of aminoparthenolide analogs. Bioorg Med Chem Lett 18:3870–3873
- Neelakantan S, Nasim S, Guzman ML, Jordan CT, Crooks PA (2009) Aminoparthenolides as novel anti-leukemic agents: discovery of the NF-kappaB inhibitor, DMAPT (LC-1). Bioorg Med Chem Lett 19: 4346–4349
- Nogueira V, Hay N (2013) Molecular pathways: reactive oxygen species homeostasis in cancer cells and implications for cancer therapy. Clin Cancer Res 19:4309–4314
- Pati HN, Das U, Sharma RK, Dimmock JR (2007) Cytotoxic thiol alkylators. Mini Rev Med Chem 7:131–139
- Rasul A, Bao R, Malhi M, Zhao B, Tsuji I, Li J, Li X (2013) Induction of apoptosis by costunolide in bladder cancer cells is mediated through ROS generation and mitochondrial dysfunction. Molecules 18: 1418–1433
- Reuter S, Gupta SC, Chaturvedi MM, Aggarwal BB (2010) Oxidative stress, inflammation, and cancer: how are they linked? Free Radic Biol Med 49:1603–1616
- Rüngeler P, Castro V, Mora G, Gören N, Vichnewski W, Pahl HL, Merfort I, Schmidt TJ (1999) Inhibition of transcription factor NFkappaB by sesquiterpene lactones: a proposed molecular mechanism of action. Bioorg Med Chem 7:2343–2352
- Salla M, Fakhoury I, Saliba N, Darwiche N, Gali-Muhtasib H (2013) Synergistic anticancer activities of the plant-derived sesquiterpene lactones salograviolide A and iso-seco-tanapartholide. J Nat Med 67:468–479
- Schomburg C, Schuehly W, da Costa FB, Klempnauer K-H, Schmidt TJ (2013) Natural sesquiterpene Lactones as inhibitors of Mybdependent gene expression: structure-activity relationships. Eur J Med Chem 63:313–320
- Schumacker PT (2006) Reactive oxygen species in cancer cells: live by the sword, die by the sword. Cancer Cell 10:175–176
- Scotti MT, Fernandes MB, Ferreira MJ, Emerenciano VP (2007) Quantitative structure-activity relationship of sesquiterpene lactones with cytotoxic activity. Bioorg Med Chem 15:2927–2934
- Shanmugam R, Kusumanchi P, Cheng L, Crooks P, Neelakantan S, Matthews W, Nakshatri H, Sweeney CJ (2010) A water-soluble parthenolide analogue suppresses in vivo prostate cancer growth by targeting NFkappaB and generating reactive oxygen species. Prostate 70:1074–1086
- Shanmugam R, Kusumanchi P, Appaiah H, Cheng L, Crooks P, Neelakantan S, Peat T, Klaunig J, Matthews W, Nakshatri H, Sweeney CJ (2011) A water soluble parthenolide analog suppresses in vivo tumor growth of two tobacco-associated cancers, lung and bladder cancer, by targeting NF-κB and generating reactive oxygen species. Int J Cancer 128:2481–2494
- Sharma V, Joseph C, Ghosh S, Agarwal A, Mishra MK, Sen E (2007) Kaempferol induces apoptosis in glioblastoma cells through oxidative stress. Mol Cancer Ther 6:2544–2553
- Skalska J, Brookes PS, Nadtochiy SM, Hilchey SP, Jordan CT, Guzman ML, Maggirwar SB, Brieh MM, Bernstein SH (2009) Modulation of cell surface protein free thiols: a potential novel mechanism of action of the sesquiterpene lactone parthenolide. PLoS One 4:e8115
- Song JM, Qian X, Upadhyayya P, Hong KH, Kassie F (2014) Dimethylaminoparthenolide, a water soluble parthenolide, suppresses lung tumorigenesis through down-regulating the STAT3 signaling pathway. Curr Cancer Drug Targets 14:59–69
- Steele AJ, Jones DT, Ganeshaguru K, Duke VM, Yogashangary BC, North JM, Lowdell MW, Kottaridis PD, Mehta AB, Prentice AG, Hoffbrand AV, Wickremasinghe RG (2006) The sesquiterpene

<span id="page-9-0"></span>lactone parthenolide induces selective apoptosis of B-chronic lymphocytic leukemia cells in vitro. Leukemia 20:1073–1079

- Suvannasankha A, Crean CD, Shanmugam R, Farag SS, Abonour R, Boswell HS, Nakshatri H (2008) Antimyeloma effects of a sesquiterpene lactone parthenolide. Clin Cancer Res 14:1814–1822
- Szatrowski TP, Nathan CF (1991) Production of large amounts of hydrogen peroxide by human tumor cells. Cancer Res 51:794–798
- Toyokuni S, Okamoto K, Yodoi J, Hiai H (1995) Persistent oxidative stress in cancer. FEBS Lett 358:1–3
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J (2007) Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 39:44–84
- Visconti R, Grieco D (2009) New insights on oxidative stress in cancer. Curr Opin Drug Discov Dev 12:240–245
- Wang W, Adachi M, Kawamura R, Sakamoto H, Hayashi T, Ishida T, Imai K, Shinomura Y (2006) Parthenolide-induced apoptosis in multiple myeloma cells involves reactive oxygen species generation and cell sensitivity depends on catalase activity. Apoptosis 11:2225– 2235
- Wen J, You KR, Lee SY, Song CH, Kim DG (2002) Oxidative stressmediated apoptosis. The anticancer effect of the sesquiterpene lactone parthenolide. J Biol Chem 277:38954–38964
- Woerdenbag HJ, Lemstra W, Malingre TM, Konings AW (1989) Enhanced cytostatic activity of the sesquiterpene lactone eupatoriopicrin by glutathione depletion. Br J Cancer 59:68–75
- Wong HR, Menendez IY (1999) Sesquiterpene lactones inhibit inducible nitric oxide synthase gene expression in cultured rat aortic smooth muscle cells. Biochem Biophys Res Commun 262:375–380
- Wyrebska A, Gach K, Janecka A (2014) Mini Rev combined effect of parthenolide and various anti-cancer drugs or anticancer candidate

substances on malignant cells in vitro and in vivo. Med Chem 14: 222–228

- Zhang S, Ong CN, Shen HM (2004) Critical roles of intracellular thiols and calcium in parthenolide-induced apoptosis in human colorectal cancer cells. Cancer Lett 208:143–153
- Zhang S, Won YK, Ong CN, Shen HM (2005) Anti-cancer potential of sesquiterpene lactones: bioactivity and molecular mechanisms. Curr Med Chem Anticancer Agents 5:239–249
- Zhang Q, LuY DY, Zhai J, Ji Q, Ma W, Yang M, Fan H, Long J, Tong Z, Shi Y, Jia Y, Han B, Zhang W, Qiu C, Ma X, Li Q, Shi Q, Zhang H, Li D, Zhang J, Lin J, Li LY, Gao Y, Chen Y (2012) Guaianolide sesquiterpene lactones, a source to discover agents that selectively inhibit acute myelogenous leukemia stem and progenitor cells. J Med Chem 55:8757–8769
- Zhang Y, Bao YL, Wu Y, Yu CL, Huang YX, Sun Y, Zheng LH, Li YX (2013) Alantolactone induces apoptosis in RKO cells through the generation of reactive oxygen species and the mitochondrial pathway. Mol Med Rep 8:967–972
- Zhangabylov NS, Yu Dederer L, Gorbacheva LB, Vasil'eva SV, Terekhov AS (2004) Adekenov SM Sesquiterpene lactone arglabin influences DNA synthesis in P388 leukemia cells in vivo. Pharm Chem J 38: 651–653
- Zheng B, Wu L, Ma L, Liu S, Li L, Xie W, Li X (2013) Telekin induces apoptosis associated with the mitochondria-mediated pathway in human hepatocellular carcinoma cells. Biol Pharm Bull 36:1118– 1125
- Zunino SJ, Ducore JM, Storms DH (2007) Parthenolide induces significant apoptosis and production of reactive oxygen species in highrisk pre-B leukemia cells. Cancer Lett 254:119–127