

Serotonin pharmacology in the gastrointestinal tract: a review

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Abstract Serotonin (5-hydroxytryptamine or 5-HT) plays a critical physiological role in the regulation of gastrointestinal (GI) function. 5-HT dysfunction may also be involved in the pathophysiology of a number of functional GI disorders, such as chronic constipation, irritable bowel syndrome and functional dyspepsia. This article describes the role of 5-HT in the enteric nervous system (ENS) of the mammalian GI tract and the receptors with which it interacts. Existing serotonergic therapies that have proven effective in the treatment of GI functional disorders and the potential of drugs currently in development are also highlighted. Advances in our understanding of the physiological and pathophysiological roles of 5-HT in the ENS and the identification of selective receptor ligands bodes well for the future development of more efficacious therapies for patients with functional GI disorders.

Keywords 5-HT · Gastrointestinal · Enteric nervous system

Introduction

The gastrointestinal (GI) tract is unique among organ systems in that, although under the influence of the central nervous system (CNS), it can function in isolation. A complex and highly organized enteric nervous system (ENS) regulates and coordinates GI absorption, secretion, motility, and sensation (Gershon and Tack 2007). The ENS consists of submucosal and myenteric ganglia and an

intricate network of intrinsic and extrinsic afferent neurons, interneurons, and motor neurons, which interact with longitudinal and circular smooth muscle and mucosal endocrine cells. Submucosal intrinsic primary afferent neurons (IPANs), which innervate the secretory epithelium, are critical in the initiation of both secretory and peristaltic reflexes, while myenteric IPANs may participate in the regulation of peristalsis through the initiation of giant migrating contractions. Myenteric motor neurons innervate smooth muscle via the interstitial cells of Cajal and regulate mechanical activity (Gershon 2004; Gershon and Tack 2007). Ascending and descending interneurons play an important role in the activation of excitatory and inhibitory motor neurons, respectively. Considering the complexity inherent in regulating GI function and its physiological importance, it is not surprising that malfunctions occur and that when they do, the quality of life of affected individuals is impacted significantly. It is estimated that digestive diseases, as a whole, affect 60–70 million people in the USA with direct and indirect costs totaling approximately \$107 billion (Gershon and Tack 2007). In many cases, no organic etiology can be identified to explain the clinical problem, and a functional disorder is diagnosed. A diverse assortment of functional GI disorders, defined most recently by the Rome III committee, has been identified (Drossman 2006). Of those disorders in which serotonergic dysfunction and/or therapy is indicated, albeit with different levels of validation, the most common are chronic constipation, irritable bowel syndrome (IBS), gastroparesis, functional dyspepsia, and functional heartburn.

In the USA, 10–20% of adults have symptoms consistent with IBS, and the disorder accounts for 2.5–3.5 million physician visits per year and 20–40% of all visits to the gastroenterologist. The burden of IBS in the USA was estimated to be approximately \$20 billion in direct and

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indirect costs in 2000 (American Gastroenterological Association 2002). IBS is defined as a “functional bowel disorder in which abdominal pain or discomfort is associated with defecation or a change in bowel habit, with features of disordered defecation” (Longstreth et al. 2006). It is generally acknowledged that three categories of IBS exist: diarrhea-predominant (IBS-D), constipation-predominant (IBS-C), or IBS with a mixed stool pattern (IBS-M). Chronic constipation, which affects up to 27% of the population, depending on demographics and definition, presents as persistently difficult, infrequent, or seemingly incomplete defecation, which does not meet the IBS criteria (Longstreth et al. 2006). Gastroparesis is a chronic disorder characterized by delayed gastric emptying, which results in early satiety, nausea and vomiting, bloating, and upper abdominal discomfort (Parkman et al. 2004). Functional dyspepsia, consisting of chronic or recurrent epigastric pain or burning with postprandial fullness and early satiation, is extremely common, affecting 20–30% of the general population each year (Tack et al. 2006c). In contrast to IBS, the pain or discomfort of functional dyspepsia is not relieved by defecation, nor is it associated with a change in defecation frequency or stool consistency (Saad and Chey 2006). Functional heartburn consists of retrosternal burning in the absence of gastroesophageal reflux disease (GERD; Galmiche et al. 2006).

Of all the endogenous factors influencing GI function, both physiologically and pathophysiologically, serotonin (5-hydroxytryptamine or 5-HT) is particularly important. This article discusses the role of 5-HT in the mammalian GI tract and the receptors with which it interacts. In addition, existing and potential future serotonergic therapies for the treatment of GI functional disorders are highlighted. Emphasis is placed on human pharmacological data, where available, and some of the many preclinical animal studies that have been performed. While it is evident that 5-HT activity within the CNS can influence GI function (e.g., gastric tone, intestinal motility, and visceral sensitivity) via an interaction with multiple receptor subtypes (Crocì et al. 1995; Sivarao et al. 2004), the focus of this review is serotonin receptor pharmacology within the GI tract.

The physiological and pathophysiological roles of 5-HT in the GI tract

The coordinated movement of food along the GI tract is dependent on 5-HT-mediated regulation of smooth muscle tone, peristalsis, mucosal secretion, and visceral perception (Baker 2005; Hansen and Skadhauge 1997; Jin et al. 1999) via an interaction with intrinsic enteric and extrinsic afferent neurons, the interstitial cells of Cajal, smooth muscle cells, and enterocytes (Gershon and Tack 2007; Read and Gwee 1994; Wouters et al. 2007a,b). Enterochromo-

affin cells, located primarily at the base of the epithelial crypts in the GI tract, contain more than 90% of the total 5-HT within the human body and contain the enzymes required for its synthesis, including the rate-limiting enzyme, tryptophan hydroxylase 1 (TPH1), which catalyzes the conversion of dietary tryptophan to 5-hydroxytryptophan. In response to mechanical stimulation, as provided, for example by a bolus of food, 5-HT is released from the enterochromaffin cells into the lamina propria (Gershon 2004). The released 5-HT stimulates IPANs containing calcitonin gene-related peptide (CGRP) in the mucosa, which synapse with ascending and descending interneurons and, in turn, with motor neurons (Gershon and Tack 2007; Pan and Gershon 2000). Ascending interneurons activate excitatory motor neurons, which release acetylcholine, substance P, and neurokinin A causing contraction of circular smooth muscle, while descending interneurons stimulate inhibitory motor neurons, which release nitric oxide (NO), vasoactive intestinal peptide (VIP), and pituitary adenylate cyclase-activating peptide (PACAP), producing circular smooth muscle relaxation. Longitudinal smooth muscle contracts and relaxes in reverse fashion to circular muscle, under regulation by excitatory neurotransmitters such as VIP, PACAP, and NO (Grider 2003a). This coordinated neuromuscular activity or peristaltic reflex promotes the oral–aboral transit of food along the GI tract (Grider 2003b; Grider and Makhlof 1990; Pan and Gershon 2000). It is now appreciated that the interstitial cells of Cajal, which regulate smooth muscle rhythmic electrical activity and participate in neuromuscular transmission, are influenced by 5-HT (Wouters et al. 2007a,b). Activation of submucosal IPANs initiates mucosal secretion and peristalsis, while those in the myenteric plexus are thought to be responsible for giant migrating complexes. Extrinsic afferent neurons, which are activated directly by 5-HT following its release from enterochromaffin cells and indirectly via the IPANs, transmit sensation from the GI tract to the CNS. Enteric serotonergic neurons represent only a small proportion of the total number of neurons in the ENS. For example, in the myenteric plexus, 5-HT-containing neurons constitute only 1% of neurons (Costa et al. 1996). 5-HT is, however, an important enteric neurotransmitter in the GI tract. Neuronally released 5-HT mediates fast synaptic transmission in some IPANs (Galligan et al. 2000). 5-HT contained in interneurons and motor neurons participates in a variety of reflex pathways, such as the descending pathways in the myenteric plexus (Galligan 2002; Young and Furness 1995).

Data from many studies indicate that 5-HT is likely to play an important role in the pathophysiology of functional disorders and their symptomatology. Thus, in carcinoid syndrome, a disease resulting from tumors of the enterochromaffin cells, there is excessive release of 5-HT, which

contributes to the severe diarrhea and abdominal discomfort associated with the disease (Carling et al. 2002; Von Der Ohe et al. 1993). Furthermore, elevations in either fasting or postprandial plasma 5-HT concentrations have been noted in patients with IBS-D and in functional dyspepsia (Atkinson et al. 2006; Houghton et al. 2003; Lea et al. 2002). In IBS-C patients, there are reductions in the plasma concentrations of 5-HT and 5-hydroxyindole acetic acid (5-HIAA), its primary metabolite (Atkinson et al. 2006; Dunlop et al. 2005). Moreover, mucosal 5-HT and TPH1 messenger ribonucleic acid (mRNA) or immunoreactivity is reduced in rectal biopsy samples from patients with IBS-C and IBS-D compared to healthy subjects (Coates et al. 2004). In contrast, an increase in 5-HT-containing enterochromaffin cells has been observed in patients with post-infectious IBS (Dunlop et al. 2005). Caution is, however, warranted as some studies have failed to demonstrate significant differences in 5-HT levels between healthy subjects and IBS patients (Mawe et al. 2006). The impact of acute depletion of tryptophan, the dietary source of 5-HT, on GI function of IBS patients and healthy subjects is unclear. In one study, tryptophan depletion was associated with fewer GI symptoms in IBS patients compared to acute tryptophan increase, while in control subjects neither manipulation had any effect (Shufflebotham et al. 2006). However, in another study, visceral perception was increased by tryptophan depletion in patients with IBS-D and healthy control subjects alike (Kilkens et al. 2004). Overall, these data support the hypothesis that 5-HT dysfunction may be involved in the pathophysiology of functional GI disorders, such as IBS. The contributions made by 5-HT receptors and by the serotonin transporter (SERT), in the physiological and pathophysiological actions of 5-HT, will now be addressed.

5-HT and its receptors

5-HT was discovered in the 1930s and named “enteramine” as it was first extracted from the intestine (Erspamer and Vialli 1937). Since its discovery, it has become clear that 5-HT serves many diverse physiological functions, such as the regulation of sleep, appetite, mood, neuroendocrine secretion, sexual behavior, cognition, and GI function via an interaction with multiple 5-HT receptors (Barnes and Sharp 1999; Hoyer et al. 2002). To date, 14 5-HT receptors, belonging to seven families (5-HT₁, 5-HT₂, 5-HT₃, 5-HT₄, 5-HT₅, 5-HT₆, and 5-HT₇), have been identified (Hoyer et al. 1994; 2002; Table 1). With the exception of the 5-HT₃ receptor, which is a ligand-gated ion channel, each of the 5-HT receptors identified is a seven-transmembrane domain, G-protein-coupled receptor (Hoyer et al. 1994; Hartig et al. 1996). The 5-HT₁ and 5-HT₅ receptor families are

negatively and the 5-HT₄, 5-HT₆, and 5-HT₇ receptors are positively coupled to adenylyl cyclase. Binding of 5-HT to the Gq-coupled 5-HT₂ receptor activates phospholipase C resulting in the release of inositol triphosphate and an elevation of cytosolic calcium. Selective agonists and antagonists for the majority of the receptor families and their subtypes are now available as pharmacological tools. The use of these ligands and data generated by molecular biology studies have provided a clearer understanding of the roles of each 5-HT receptor in GI function (Table 1). As a result of its diverse physiological and pathophysiological functions and the multiplicity of its receptor subtypes, the serotonergic system has served the pharmaceutical industry well in the search for novel therapies for a variety of human disorders (Jones and Blackburn 2002). Thus, 5-HT₁ receptor agonists, 5-HT₃ receptor antagonists, and selective serotonin reuptake inhibitors (SSRIs) are effective therapies for migraine, cancer chemotherapy-induced emesis, and major depressive disorder, respectively. With regard to GI disorders, various 5-HT receptor ligands have been evaluated clinically to treat conditions such as IBS-D, IBS-C, chronic constipation, functional dyspepsia, and gastroparesis (Table 2). In the USA, however, there are currently only four serotonergic agents approved for the treatment of functional GI disorders (i.e., the 5-HT₃ receptor antagonist, alosetron [Lotronex[®]], the 5-HT₄ receptor agonist, tegaserod [Zelnorm[®]], and the dual 5-HT₄ receptor agonist and 5-HT₃ receptor antagonists, metoclopramide [Reglan[®]] and cisapride [Propulsid[®]]). Each has significant safety concerns that limit their general utility (see below).

5-HT receptors in the GI tract

5-HT₁ receptor family

The 5-HT₁ receptor family consists of the 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1D}, 5-HT_{1E}, and 5-HT_{1F} subtypes (Hoyer et al. 2002). Data from receptor distribution studies and preclinical or clinical pharmacodynamic investigations with 5-HT₁ receptor-selective ligands such as sumatriptan, 8-hydroxy-2-(di-*n*-propylamino)tetralin (8-OH DPAT), buspirone, and flesinoxan suggest that several of the 5-HT₁ receptor subtypes have physiological and pathophysiological roles in the GI tract. Radioligand binding and mRNA expression studies in the rat have provided evidence that 5-HT_{1A} receptors are expressed on submucosal and myenteric neurons throughout the GI tract, with a particularly high density in the stomach (Kirchgessner et al. 1993). Submucosal 5-HT_{1A}-immunoreactive cells are principally interneurons within the submucosal plexus while myenteric 5-HT_{1A}-immunoreactive neurons project to submucosal ganglia (Kirchgessner et al. 1996). A subset of 5-HT-

Table 1 Summary of 5-HT receptor characteristics in the GI tract

Receptor type	Receptor subtype	Signal transduction mechanism(s)	Primary localization	Functional role	Subtype selective ligands ^a
5-HT ₁	5-HT _{1A}	Gi/o	Intrinsic sensory neurons, interneurons, excitatory motor neurons, enterocytes	Relaxation, modulation of visceral sensitivity	Agonists: 8-OH DPAT, BP554 Antagonists: (S)WAY100135, NAN-190
	5-HT _{1B}	Gi/o	Intrinsic and extrinsic sensory neurons, smooth muscle cells	Contraction/relaxation	Agonists: L694247, CP94253 Antagonists: SB236057, SB224289
	5-HT _{1D}	Gi/o	Intrinsic and extrinsic sensory neurons, smooth muscle cells	Contraction/relaxation	Agonists: PNU10929, L694247 Antagonists: BRL15572
	5-HT _{1E}	Gi/o	Not present	Not applicable	Agonists: none Antagonists: none
	5-HT _{1F}	Gi/o	Cellular localization not described	Relaxation	Agonists: LY334370, LY334864 Antagonists: none
	5-HT ₂	5-HT _{2A}	Gq/11	Enteric neurons, enterocytes, smooth muscle cells	Contraction/relaxation
5-HT ₃	5-HT _{3B}	Gq/11	Enteric neurons, smooth muscle cells	Contraction	Agonists: BW723C86 Antagonists: RS127445
	5-HT _{3C}	Gq/11	Not present	Not applicable	Agonists: Ro600175, WAY629 Antagonists: SB242084, RS102221
	5-HT _{3A/B}	Cation channel (Na ⁺ , Ca ²⁺ , K ⁺)	Intrinsic and extrinsic sensory neurons, interstitial cells of Cajal, secretomotor neurons, enterocytes	Modulation of visceral sensitivity and motility, secretion	Agonists: SR57227 Antagonists: ondansetron, granisetron, alosetron
5-HT ₄	Ten 5-HT ₄ splice variants	Gs; increase [Ca ²⁺] _i (5-HT _{4(a)}); Gi/o (5-HT _{4(b)})	Intrinsic sensory neurons, interneurons, interstitial cells of Cajal, excitatory and inhibitory motor neurons, smooth muscle cells, enterocytes	Contraction/relaxation, stimulation of motility, secretion	Agonists: ML10302, TD-5108 Antagonists: GR113808, piboserod
	5-HT ₅	5-HT _{5A} 5-HT _{5B}	Not present Not present	Not applicable Not applicable	Agonists: none Antagonists: none
5-HT ₆	5-HT ₆	None identified	Limited expression mRNA in stomach, cellular localization not described	No definitive role	Agonists: EMD386088 Antagonists: SB271046, SB357134
	5-HT ₇	Gs	Intrinsic sensory neurons, inhibitory motor neurons, smooth muscle cells	Relaxation, modulation of visceral sensitivity	Agonists: LP44, AS19 Antagonists: SB656104, SB269970

^a From Alexander et al. 2004, Toctris Bioscience catalog, or this review

Table 2 Selected serotonergic agents in development for the treatment of GI disorders (source: Thompson Pharma, company websites and FDA.gov)

Drug name	Company	Target mechanism(s)	Status ^a	Indication
R-137696	Johnson & Johnson	5-HT _{1A} agonist	Phase 1	Dyspepsia
TZB-30878	Aska-Pharma	5-HT _{1A} agonist, 5-HT ₃ antagonist	Preclinical	IBS-D
M0007	Movetis	5-HT _{1A} antagonist	Preclinical	GERD, dyspepsia
Espindolol	AGI Therapeutics	5-HT _{1A/B} antagonist, β -antagonist	Phase 2	Functional dyspepsia, IBS
R-1 (IBS)	Asterand	5-HT _{2B} antagonist	Preclinical	IBS
M0005, M0006	Movetis	5-HT ₃ agonist	Preclinical	GERD, dyspepsia
Pumosestrag (DDP-733)	Dynogen Pharm.	5-HT ₃ partial agonist	Phase 2 (Canada)	IBS-C, nocturnal GERD
Alosetron (Lotronex [®])	GSK	5-HT ₃ antagonist	Launched	IBS-D
Ramosetron	Astellas	5-HT ₃ antagonist	Pre-registration (Japan)	IBS
Cilansetron	Solvay	5-HT ₃ antagonist	Pre-registration (Europe)	IBS-D
DDP-225	Dynogen Pharm.	5-HT ₃ antagonist; NET inhibitor	Phase 2	IBS-D
Prucalopride	Movetis	5-HT ₄ agonist	Phase 3 (Europe)	Chronic constipation
TD-5108	Theravance	5-HT ₄ agonist	Phase 2	Chronic constipation
ATI-7505	Procter & Gamble	5-HT ₄ agonist	Phase 2	Gastroparesis, GERD
M0003, M0004	Movetis	5-HT ₄ agonist	Phase 1 (preclinical)	Gastroparesis, gastric regurgitation
Mosapride (Gasmotin [®])	Takeda Pharm.	5-HT ₄ partial agonist	Launched (Japan)	Chronic gastritis GI symptoms
Tegaserod (Zelnorm [®])	Novartis	5-HT ₄ partial agonist	Restricted use	IBS-C, chronic constipation
Cisapride (Propulsid [®])	Johnson & Johnson	5-HT ₄ agonist, 5-HT ₃ antagonist	Restricted use	Nocturnal GERD
Renzapride	Alizyme	5-HT ₄ agonist, 5-HT ₃ antagonist	Phase 3	IBS-C, IBS-M
Metoclopramide (Reglan [®])	Schwarz Pharma	5-HT ₄ agonist, 5-HT ₃ and D ₂ antagonist	Launched	Diabetic gastroparesis, GERD
M0008	Movetis	5-HT ₄ antagonist	Preclinical	GERD, dyspepsia
Venlafaxine (Effexor [®])	Wyeth	Dual SERT, NET inhibitor	Phase 4	Functional dyspepsia
Duloxetine (Cymbalta [®])	Lilly	Dual SERT, NET inhibitor	Phase 4	IBS
LX-1031	Lexicon Pharm.	5-HT release inhibitor (TPH1 inhibitor)	Phase 1	IBS
Tianeptine	Pharmos	5-HT uptake stimulator, opioid agonist	Preclinical	Dyspepsia, IBS

^a Most advanced phase in the USA, unless otherwise indicated

containing enterochromaffin cells also express 5-HT_{1A} immunoreactivity (Kirchgeßner et al. 1996). 5-HT_{1A} receptor activation produces presynaptic inhibition of fast and slow excitatory neurotransmission in the ENS and hyperpolarization of myenteric IPANs resulting in a reduction in the amplitude of excitatory postsynaptic potentials (Galligan 1996; Pan and Galligan 1994). Many preclinical studies using isolated GI smooth muscle preparations have demonstrated 5-HT_{1A} receptor-mediated activity. Thus, 5-HT_{1A} receptor activation is associated with inhibition of electrically evoked contractions of guinea pig ileum and stomach circular smooth muscle (Buchheit and Buhl 1994; Mir et al. 1988), and relaxation of the dog proximal stomach and mouse fundus (Janssen et al. 2003; Xue et al. 2006). The 5-HT_{1A} receptor may also have a role in visceral sensitivity; in rats, buspirone attenuates colorectal distension-mediated abdominal withdrawal and changes in blood pressure, although its site of action is unclear (Sivarao et al. 2004). Data on the pharmacological effects of 5-HT_{1A} receptor activation in humans are rather limited. Relaxation of the human gastric fundus has been observed following application of R137696, a selective 5-HT_{1A} receptor agonist (Boeckxstaens et al. 2006). In addition, modest antiemetic activity of buspirone has been reported

clinically in cancer patients (Alfieri and Cubeddu 1995), although it remains unclear whether a CNS or peripheral site of action is involved. In healthy human subjects, buspirone fails to affect sensorimotor functions, nor does it influence gastric emptying, colonic compliance, or tone, although it reduces postprandial aggregate symptom and nausea scores (Chial et al. 2003a,b). Clinical utility of 5-HT_{1A} receptor agonists in patients with functional dyspepsia remains a possibility, although this requires further investigation (Talley 2003a). The selective 5-HT_{1A} receptor agonist, R137696, developed by Janssen, had no significant efficacy in patients with functional dyspepsia (Tack et al. 2004). It has been speculated that this was a consequence of receptor desensitization, and other companies continue to pursue the development of 5-HT_{1A} receptor agonists or partial agonists for indications such as functional dyspepsia and GERD. TZB-30878 (Aska Pharmaceutical) possesses 5-HT_{1A} receptor agonist activity and 5-HT₃ receptor antagonist activity and, on the basis of preclinical rodent data, may have potential in the treatment of IBS-D, at least in part via an action within the CNS (Tamaoki et al. 2007).

Although the human 5-HT_{1B} and 5-HT_{1D} receptors were cloned and characterized in the early 1990s (Hamblin et al. 1992; Jin et al. 1992; Weinschank et al. 1992), a detailed

analysis of their expression in the GI tract is lacking. Expression of 5-HT_{1B} and to a lesser extent 5-HT_{1D} mRNA has been described in the bovine ileum and colon (Engel et al. 2006). mRNA for both receptor subtypes is present in human and rat sensory neurons consistent with an involvement in sensory neurotransmission generally (Pierce et al. 1996; 1997). Definition of the GI pharmacology of 5-HT_{1B} and 5-HT_{1D} receptor ligands is based almost exclusively on data generated with sumatriptan, an agent marketed for the acute treatment of migraine headache. Although sumatriptan has significant affinity, for and agonist activity at, the 5-HT_{1F}, in addition to 5-HT_{1B} and 5-HT_{1D} receptors, there is only limited evidence for involvement of the former receptor subtype in regulating GI function (see below). Sumatriptan produces relaxation of the gastric fundus and inhibition of antral contractility in a number of species including humans, actions attributed to activation of the 5-HT_{1B} or 5-HT_{1D} receptor (Coulie et al. 1997, 1999; Tack et al. 2000). In the dog, gastric accommodation has also been described (De Ponti et al. 2003). There is a corresponding delay in the gastric emptying of solids and liquids following sumatriptan administration to healthy human subjects (Coulie et al. 1997). Sumatriptan increases the periodicity of the migrating motor complex and reduces the occurrence of postprandial coordinated motor activity involving the gastric antrum, pylorus, and duodenum (Calvert et al. 2004). It is interesting to note that in healthy human subjects, sumatriptan increases postprandial lower esophageal sphincter pressure but, seemingly paradoxically, increases the frequency of reflux, actions that may stem from prolonged fundic relaxation and retention of proximal stomach contents (Sifrim et al. 1999). 5-HT_{1D} receptor activation may be responsible for the 5-HT-mediated contraction of the circular and longitudinal muscles of the human ileum (Borman and Burleigh 1997a). On the basis of cost alone, sumatriptan is unlikely to be utilized by gastroenterologists in chronic functional GI disorders, and it remains to be determined whether the available data are sufficiently promising to stimulate the development of other 5-HT_{1B} or 5-HT_{1D} receptor agonists in this therapeutic area.

The 5-HT_{1E} receptor (McAllister et al. 1992) appears to be localized entirely within the CNS (Bai et al. 2004; Barnes and Sharp 1999). The lack of selective ligands for the 5-HT_{1E} receptor precludes a definition of its GI pharmacology, although a functional role in the ENS seems unlikely based on the absence of 5-HT_{1E} receptor mRNA in guinea pig intestine and rat sensory nerves, at least at the level of the dorsal root ganglion (Bai et al. 2004; Chen et al. 1998; Nicholson et al. 2003). The 5-HT_{1F} receptor is closely related to the 5-HT_{1E} subtype, possessing greater than 70% sequence homology across the seven transmembrane domains and like the latter appears to be largely restricted to the CNS, although limited expression in peripheral tissues

has been described (Adham et al. 1993; Barnes and Sharp 1999; Hoyer et al. 2002). Data indicative of a role for the 5-HT_{1F} receptor in the GI tract are sparse. In the cat stomach, LY-344864 and BRL-54443, selective 5-HT_{1F} receptor agonists, produce relaxation (Janssen et al. 2004), and it is therefore possible that some of the gastric effects of sumatriptan, described above, are 5-HT_{1F} receptor mediated, although this requires further investigation.

A number of studies have indicated an important physiological role in GI function for a peripheral 5-HT receptor characterized by high affinity for [³H]5-HT, and termed the 5-HT_{1P} subtype (Mawe et al. 1986). The 5-HT_{1P} receptor has not been cloned, and its molecular identity therefore remains elusive. 6-Hydroxyindalpine and *N*-acetyl-5-hydroxytryptophyl-5-hydroxytryptophan amide have agonist and antagonist activities, respectively, at the 5-HT_{1P} receptor, with some selectivity. Interaction of sumatriptan with the 5-HT_{1P} receptor on myenteric neurons of the guinea pig gastric antrum has also been proposed (Tack et al. 2007). It has been suggested that the 5-HT_{1P} receptor is either the 5-HT₇ receptor (see below) or a heterodimer of the dopamine D₂ receptor with either the 5-HT_{1B} or 5-HT_{1D} receptor (Liu and Gershon 2005b; Monro et al. 2005; Tonini 2005). 5-HT_{1P} receptors are localized on IPANs in the submucosal and myenteric plexuses and in the intestinal mucosa (Branchek et al. 1988; Kirchgessner et al. 1993), and data from a number of studies suggest that the 5-HT_{1P} receptor has a functional role in the ENS of a variety of species (e.g., rat, guinea pig, and human; Cooke et al. 1997; Foxx-Orenstein et al. 1996; Tack et al. 1992). 5-HT, released from enterochromaffin cells, following mucosal stimulation, initiates reflexes via activation of 5-HT_{1P} receptors on the submucosal IPAN terminals (Pan and Gershon 2000). 5-HT_{1P} receptor activation and subsequent stimulation of submucosal VIP- and CGRP-containing afferent neurons are considered to have a critical role in initiation and maintenance of the peristaltic reflex. To date, no drugs specifically targeting the 5-HT_{1P} receptor have been developed for GI disorders. It has been postulated that while a 5-HT_{1P} receptor agonist should increase and an antagonist decrease GI transit, intractable diarrhea and paralytic ileus, respectively, may be unavoidable (Gershon 2004).

5-HT₂ receptor family

The 5-HT₂ receptor class comprises the 5-HT_{2A}, 5-HT_{2B}, and 5-HT_{2C} subtypes, which possess 46–50% sequence identity. The 5-HT_{2A} and 5-HT_{2B} receptor subtypes are present in the CNS and periphery, while 5-HT_{2C} receptors appear to be restricted to the CNS (Hoyer et al. 2002; Leysen 2004). 5-HT_{2C} receptors are considered to lack a role in GI physiology and pathophysiology, in contrast to the 5-HT_{2A} and 5-HT_{2B} receptor subtypes.

Following its cloning from a variety of species, including humans (Saltzman et al. 1991), 5-HT_{2A} receptor mRNA and immunoreactivity have been localized to myenteric and submucosal neurons, enterocytes, and longitudinal and circular muscle cells of the GI tract (Bonaventure et al. 2005; Fiorica-Howells et al. 2002; Leysen 2004). 5-HT_{2A} receptor activation mediates secretory responses in human and rat isolated colonic mucosa (Borman and Burleigh 1996; Hansen and Skadhauge 1997; Imada-Shirakata et al. 1997). In the rat and guinea pig stomach antrum or corpus, 5-HT_{2A} receptor activation results in contraction (Komada and Yano 2007; Tamura et al. 1996), while in the rat fundus, a relaxation is observed (Komada and Yano 2007). Activation of 5-HT_{2A} receptors in canine and guinea pig isolated colonic longitudinal muscle results in contraction (Briejer et al. 1995a; Prins et al. 1997), and in vivo, motility is stimulated in the middle and distal colon (Nagakura et al. 1996a). 5-HT_{2A} receptor activation also produces contraction of human isolated jejunal smooth muscle cells (Kuemmerle et al. 1995). It is interesting to note that in mice, ENS morphology and resting GI transit or colorectal motility are unaffected by “knocking out” the 5-HT_{2A} receptor despite the fact that 5-HT-evoked contraction of the isolated colon of these animals is absent (Fiorica-Howells et al. 2002). This observation may indicate that 5-HT_{2A} receptors are not important physiologically or pathophysiologically in the GI tract. 5-HT_{2A} receptor agonist activity should increase transit in humans with disorders of reduced GI transit (e.g., IBS-C or chronic constipation), although the anticipated generalized vasoconstriction associated with this approach is likely to preclude any therapeutic opportunity.

The 5-HT_{2B} receptor was originally cloned from the rat stomach fundus (Foguet et al. 1992) with the human receptor cloned in 1994 (Kursar et al. 1994). mRNA for the 5-HT_{2B} receptor is widely expressed in the human and rodent GI tracts (Fiorica-Howells et al. 2000; Borman et al. 2002). In human colon, 5-HT_{2B} receptor mRNA and protein are present in the longitudinal and, to a lesser extent, circular muscle layers and in myenteric neurons (Borman et al. 2002). The 5-HT_{2B} receptor is also expressed on interstitial cells of Cajal in the human and mouse intestine and is postulated to have a proliferative role (Wouters et al. 2007a,b). Rodent studies implicate activation of 5-HT_{2B} receptors in the development of the ENS (Fiorica-Howells et al. 2000). Indeed, it has been suggested that the primary role of the 5-HT_{2B} receptor in the human GI tract may be in the developmental regulation of the ENS (Gershon 2000). However, the 5-HT_{2B} receptor may serve other functions too. It has long been recognized that 5-HT contracts the rat stomach fundus (Vane 1957), an effect attributed to 5-HT_{2B} receptor activation (Komada and Yano 2007). 5-HT also produces a 5-HT_{2B} receptor-

mediated contraction of the longitudinal smooth muscle of the human ileum (Borman and Burleigh 1995) and augmentation of neuronally mediated contraction of human colonic longitudinal smooth muscle (Borman and Burleigh 1997a; Borman et al. 2002). It is conceivable, based on preclinical data (Beattie et al. 2004; Borman and Burleigh 1997a; Borman et al. 2002), that 5-HT_{2B} receptor agonism and antagonism will, respectively, increase and decrease GI motility in humans, although this remains to be substantiated. The potential clinical utility of a 5-HT_{2B} receptor agonist in chronic constipation or IBS-C is unlikely on the basis of anticipated cardiac valvular toxicity (Hofmann et al. 2002; Horowski et al. 2004). However, the use of a 5-HT_{2B} receptor antagonist for diarrhea and IBS-D may present a feasible yet, at this stage, still speculative therapeutic opportunity. It has been postulated that 5-HT_{2B} receptor antagonism may inhibit visceral hypersensitivity (Borman et al. 2002), although rodent data are inconsistent with this proposal (Greenwood-Van Meerveld et al. 2006a).

5-HT₃ receptor

Molecular biology and receptor localization

In common with the nicotinic acetylcholine, γ -amino butyric acid_A (GABA_A), and glutamatergic *N*-methyl-D-aspartate receptors, the 5-HT₃ receptor is a ligand-gated ion channel (Maricq et al. 1991). Several subunits of the 5-HT₃ receptor have been identified (5-HT_{3A}, 5-HT_{3B}, 5-HT_{3C}, 5-HT_{3D}, and 5-HT_{3E}; Sanger and Andrews 2006). Native 5-HT₃ receptors are pentamers, which are homo-oligomeric assemblies of 5-HT_{3A} or hetero-oligomeric assemblies of 5-HT_{3A} and 5-HT_{3B} subunits. The 5-HT_{3B} subunit imparts distinct biophysical properties upon the hetero-oligomeric assembly relative to the homo-oligomer but has negligible influence on the apparent affinity of agonists or antagonists (Davies et al. 1999). Variants of the human 5-HT_{3B} subunit that differ in the extracellular N-terminal domain have been postulated to be present in the GI tract as a result of polymorphisms in the 5-HT_{3B} gene promoter (Tzvetkov et al. 2007). The role of the other subunits in the heteromeric complexes is unknown at present. Expression analysis of the different 5-HT₃ receptor subunit genes has indicated that each is present in the human intestine and that the 5-HT_{3E} is expressed specifically at this location (Niesler et al. 2003). In the ENS, 5-HT₃ receptor immunoreactivity is expressed on neurons of the myenteric and submucosal plexuses, interstitial cells of Cajal and fibers in the circular and longitudinal muscle layers, submucosa, and mucosa (Glatzle et al. 2002). In addition, 5-HT₃ receptor-immunopositive mucosal terminals of vagal and spinal afferent neurons are also evident (Glatzle et al. 2002; Raybould et al. 2003).

5-HT₃ receptor-mediated activity in the gastrointestinal tract

Data from numerous preclinical studies have demonstrated 5-HT₃ receptor-mediated activity in the GI tract. Thus, 5-HT₃ receptor activation is associated with increased electrically evoked contractions of guinea pig and mouse stomach corpus or fundus circular smooth muscle (Buchheit and Buhl 1994; Xue et al. 2006). In the guinea pig isolated antrum, 5-HT₃ receptor activation produces a contraction (Tamura et al. 1996). Data from electrophysiology studies using the guinea pig ileum indicate that the mucosal processes of myenteric IPANs are excited via 5-HT₃ receptor activation and play a role in initiation or enhancement of myenteric reflexes (Bertrand et al. 2000). The majority of myenteric and submucosal neurons maintained in primary culture respond to 5-HT with a fast inward current that is inhibited by 5-HT₃ receptor antagonists (Galligan et al. 2000). While the role of the 5-HT₃ receptor on the interstitial cells of Cajal is unclear, it is postulated that it participates in the regulation of pacemaker activity (Wouters et al. 2007a). Preclinical studies have demonstrated that the 5-HT₃ receptor antagonists, ondansetron, granisetron, tropisetron, ramosetron, and alosetron, increase whole gut, small intestinal, or colonic transit time in mice, rats, and guinea pigs, presumably by attenuating endogenous, 5-HT₃ receptor-mediated prokinetic activity (Brown et al. 1993; Clayton et al. 1999; Nagakura et al. 1996b; Sanger and Wardle 1994). Similarly, azasetron inhibits 5-HT, corticotrophin-releasing factor, and stress-induced increases in colonic transit in rats (Haga et al. 1995). Preclinical data also support a role for the 5-HT₃ receptor in visceral nociception. Thus, ondansetron, alosetron, and cilansetron attenuate visceromotor and nociceptive responses to mechanical rectal distension or chemical stimulation in rats (Mori et al. 2004; Morteau et al. 1994). It is interesting to note however, that the efficacy of 5-HT₃ receptor antagonists in colorectal distension studies in rats pretreated with 5-hydroxytryptophan to sensitize the colon and rectum is inconsistent. Granisetron, zatosetron, and bemesetron increase the threshold for the distension-induced visceromotor reflex, while tropisetron and ondansetron have little or no effect (Banner and Sanger 1995). The 5-HT₃ receptor may also have a particularly important role in regulating GI mucosal secretion. Activation of 5-HT₃ receptors results in intestinal chloride secretion (Hansen and Skadhauge 1997) and 5-HT release from enterochromaffin cells (Schworer and Ramadori 1998). Furthermore, ondansetron inhibits 5-HT-induced secretion in the rat distal colon (Budhoo et al. 1996) and 5-hydroxytryptophan-induced diarrhea in mice (Pascual et al. 2002). Cholera toxin and *Salmonella typhimurium*-induced secretion in the porcine jejunum is attributed to the activation of 5-HT₃ receptors (Jensen et al. 1997).

Clinical activity in gastrointestinal disorders

Of the many 5-HT₃ receptor-selective antagonists identified (e.g., ondansetron, granisetron, alosetron, cilansetron, ramosetron, azasetron, tropisetron, dolasetron, and palonosetron), several have proved of considerable value, not only as pharmacological tools but also as clinically efficacious therapies for nausea and vomiting and IBS-D (Aapro 2005; Camilleri et al. 2000; Haga et al. 1995; Humphrey et al. 1999; Nagakura et al. 1996a, b; Sanger and Andrews 2006). 5-HT₃ receptor antagonists have proven particularly useful in reducing the incidence and severity of acute emetic episodes during cancer chemotherapy, providing, upon combination with dexamethasone, complete protection from vomiting in up to 90% of patients (Roila and Fatigoni 2006). The degree of protection is, however, dependent on the chemotherapeutic agent used and its dosing regimen. 5-HT₃ receptor antagonists are less effective in controlling delayed emesis that occurs, for example, more than 24 h after cisplatin administration (Roila and Fatigoni 2006). In general, clinical trial data suggest that the marketed 5-HT₃ receptor antagonists have similar therapeutic profiles with respect to efficacy and safety (McNulty 2007). One possible exception is palonosetron (Aloxi®), which was launched in 2003 for the prevention of chemotherapy-induced nausea and vomiting and is currently in phase 2 trials for the prevention of postoperative nausea and vomiting. Differentiating features of palonosetron from other marketed 5-HT₃ receptor antagonists are its markedly higher 5-HT₃ receptor-binding affinity and antagonist potency and its extended plasma half-life (approximately 40 h in healthy subjects; Navari 2006; Rubenstein 2004). Superiority of palonosetron over ondansetron and dolasetron in the prevention of both acute and delayed chemotherapy-induced nausea and vomiting has been observed in phase 3 clinical trials (Rubenstein 2004), although additional studies are warranted.

The mode of action of 5-HT₃ receptor antagonists in preventing emesis is well understood. Cancer chemotherapeutic drugs release 5-HT from enterochromaffin cells in the GI tract, resulting in activation of 5-HT₃ receptors on vagal sensory afferent neurons. The vagal afferent neurons project to the emetic center in the brainstem via the area postrema, located at the floor of the fourth ventricle (Hesketh 2004; Tyers and Freeman 2002). In ferrets, vagotomy abolishes cisplatin-mediated emesis indicating the importance of the vagus nerve in the emetic reflex (Hawthorn et al. 1988). In patients receiving cisplatin chemotherapy, a rise in urinary 5-HIAA levels has been demonstrated, which correlates with the onset and development of emesis (Cubeddu et al. 1990). The inability of ondansetron to affect the increased 5-HIAA urinary levels is consistent with an action on vagal afferent neurons rather than on the release of 5-HT (Cubeddu et al. 1990).

With respect to human GI motility effects, 5-HT₃ receptor antagonists such as alosetron, ondansetron, and cilansetron are associated with reductions in colonic and whole-gut transit or motility, increased fluid absorption, and stool consistency or attenuation of postprandial dyspepsia-like symptoms (Clemens et al. 2002; Gore et al. 1990; Houghton et al. 2000; Kuo et al. 2002; Stacher et al. 2000). The clinical efficacy of alosetron in IBS-D patients is generally attributed to its ability to prolong intestinal transit and reduce secretions; although some evidence exists for a direct inhibitory effect on visceral sensitivity in humans, this remains uncertain (Baker 2005; Gershon and Tack 2007; Mayer and Bradesi 2003). Data from rodent studies indicate that a direct, peripheral action on visceral nociceptive transmission may be important (Jiang et al. 2000; Kozlowski et al. 2000). Human positron emission tomography (PET) imaging studies have provided evidence that alosetron acts principally within discrete areas of the brain, particularly those associated with emotion, to reduce visceral sensitivity (Berman et al. 2002; Mayer et al. 2002). A central site of action for alosetron is also implicated in rodent studies of stress-induced sensitization of visceral nociception (Bradesi et al. 2007). An analysis of clinical data indicates that to achieve demonstrable efficacy in IBS, the calculated “number needed to treat” (NNT) with alosetron is seven (Cremonini et al. 2003).

While constipation, sometimes severe in nature, is an important adverse effect associated with alosetron, a more serious issue has been its association with a low (0.15% vs. 0% for placebo; Chang et al. 2006) but significant incidence of ischemic colitis, which, in some cases, has led to hospitalization, surgical intervention, and even death (Chang et al. 2006; Gallo-Torres et al. 2006). IBS patients have significantly higher rates of colonic ischemia compared to healthy subjects (Chang et al. 2006; Cole et al. 2004), and it is possible that ischemic colitis constitutes a distinct part of the natural progression of IBS in some patients or is a manifestation of another bowel pathology. However, it is evident that, although rare, there is indeed an association of alosetron with ischemic colitis. The majority of the reports of ischemic colitis occur in the first month of alosetron treatment, although specific risk factors remain to be identified (Chang et al. 2006). Following its withdrawal from the market by Glaxo SmithKline in 2000, the Food and Drug Association (FDA) announced, in June 2002, the approval of a supplemental new drug application that allows restricted marketing of alosetron, with a risk management plan and a revised indication for women with severe IBS-D who have chronic symptoms (generally lasting 6 months or longer), no discernable anatomic or biochemical abnormalities of the GI tract, and a failure to respond to conventional therapy. Development of another 5-HT₃ receptor antagonist, cilansetron, was suspended in

the USA despite clinical efficacy in IBS-D patients, as a result of an increased incidence of ischemic colitis (Chey and Cash 2005). The question remains as to whether ischemic colitis is limited to alosetron and cilansetron or whether it is inextricably linked to 5-HT₃ receptor antagonism. Ondansetron and granisteron do not appear to be associated with ischemic colitis when used as antiemetic therapies. However, as these agents are not approved for IBS therapy, it is unclear whether ischemic colitis would occur upon their use in this apparently more susceptible patient population. Therefore, although the efficacy of 5-HT₃ receptor antagonists in IBS-D is established, it remains to be determined whether or not this class of agent will play a significant role in the future given the perceived safety risks. DDP-225 is a 5-HT₃ receptor antagonist, under development by Dynogen, which also exhibits affinity for the norepinephrine transporter (NET). This dual mechanism of action is proposed to offer complementary efficacy in IBS-D with respect to symptoms of visceral hypersensitivity and accelerated motility, although this remains to be seen. It is also unclear at present whether DDP-225 will be associated with a reduced incidence of ischemic colitis relative to 5-HT₃ receptor-selective antagonists.

As 5-HT₃ receptor antagonists reduce GI transit, it is reasonable to assume that a 5-HT₃ receptor agonist would be prokinetic. This appears to be the case as a clinical study in healthy men has shown that pumosestrag (MKC-733), a selective 5-HT₃ receptor partial agonist, stimulates antroduodenal-migrating motor complex activity and accelerates small intestinal transit (Coleman et al. 2003). In the same study, pumosestrag delayed liquid gastric emptying in association with relaxation of the proximal stomach. In constipated women, pumosestrag increases stool frequency suggesting that it may have clinical potential in chronic constipation and IBS-C patients (Fujita et al. 2005). The apparent absence of adverse effects such as nausea and vomiting may be consistent with little or no systemic exposure of pumosestrag (Evangelista 2007) or reflect its partial agonist activity. It has been proposed that a 5-HT₃ receptor partial agonist is more likely to have clinical utility compared to a receptor agonist with high intrinsic activity as the former may be less susceptible to desensitization (Costedio et al. 2007).

5-HT₄ receptor

Molecular biology and receptor localization

The 5-HT₄ receptor was first cloned in 1995 (Gerald et al. 1995), and in recent years, many splice variants of the receptor, which differ only in the sequence of their intracellular COOH-terminal domain, have been identified (i.e.,

5-HT_{4(a)}, 5-HT_{4(b)}, 5-HT_{4(c)}, 5-HT_{4(d)}, 5-HT_{4(e)}, 5-HT_{4(f)}, 5-HT_{4(g)}, 5-HT_{4(i)}, and 5-HT_{4(n)}; Hoyer et al. 2002; Kaumann and Levy 2006; Liu et al. 2005a; Bender et al. 2000). A splice variant with a 14-amino acid insert in the second extracellular domain also has been described and designated 5-HT_{4(hb)} since it was found only in combination with the 5-HT_{4(b)} C terminal tail (Bender et al. 2000). In the mouse, mRNA for the 5-HT_{4(a)}, 5-HT_{4(b)}, 5-HT_{4(c)}, and 5-HT_{4(f)} splice variants is expressed, albeit to different extents, in the submucosal plexus (Liu et al. 2005a). The 5-HT_{4(a)} and 5-HT_{4(b)} but not 5-HT_{4(c)} and 5-HT_{4(f)} isoforms are also expressed in the myenteric plexus of the small and large intestine (Liu et al. 2005a). Nerve fibers in the intestinal circular muscle (but not mucosa) and the neuropil of the submucosal and myenteric plexuses are 5-HT₄-immunoreactive (Liu et al. 2005a). In the human small intestine, 5-HT_{4(b)} receptor splice variant mRNA is expressed at the highest level, followed by 5-HT_{4(c)}, 5-HT_{4(a)}, 5-HT_{4(g)}, and 5-HT_{4(d)} (Medhurst et al. 2001). In the human stomach, expression of these splice variants is generally low, as is the case for 5-HT_{4(a)}, 5-HT_{4(b)}, 5-HT_{4(c)}, and 5-HT_{4(n)} mRNA in the esophagus (Medhurst et al. 2001; Vilaro et al. 2002). In the human colon, 5-HT_{4(a)}, 5-HT_{4(b)}, and 5-HT_{4(i)} but not 5-HT_{4(g)} mRNA are expressed (Brattelid et al. 2004). It should be noted that, to date, mRNA encoding human 5-HT_{4(e)} and 5-HT_{4(f)} splice variants has not been demonstrated (Kaumann and Levy 2006). While the 5-HT₄ splice variants may have different tissue distributions, the functional significance of these isoforms is unclear. Some subtle differences in agonist potency, efficacy, constitutive receptor activity, desensitization, and signal transduction have been reported (e.g., Claeysen et al. 1999; Bender et al. 2000; Pindon et al. 2002; Mialet et al. 2003; Castro et al. 2005; Ponimaskin et al. 2002). It is interesting to note that 5-HT₄ receptor-mediated responses in human colonic smooth muscle can be demonstrated despite low or even undetectable levels of 5-HT₄ receptor mRNA (Irving et al. 2007). 5-HT₄ receptor immunoreactivity has been identified in IPANs and in the interstitial cells of Cajal in several species (Poole et al. 2006).

5-HT₄ receptor-mediated activity in the gastrointestinal tract

The 5-HT₄ receptor is considered to have a particularly important role, both physiologically and pathophysiologically in the regulation of GI function (Baker 2005; Kim and Camilleri 2000). Activation of neuronal 5-HT₄ receptors, which are located exclusively presynaptically on the terminals of IPANs, at synapses within the myenteric plexus and at the neuromuscular junction, results in prokinetic activity throughout the GI tract (Gershon 2005). The secretion of acetylcholine and CGRP from stimulated submucosal IPANs is enhanced by 5-HT₄ receptor activation (Gershon 2005; Gershon and Tack

2007; Grider 2003b) resulting in a potentiation of excitatory neurotransmission. Augmentation of acetylcholine release also occurs at nerve–nerve synapses in the myenteric plexus and at motor neuron–smooth muscle synapses (Galligan et al. 2003; Grider 2003b; Pan and Galligan 1994; Pan and Gershon 2000). 5-HT₄ receptor-mediated enhancement of neurotransmitter release from myenteric neurons occurs via cyclic adenosine monophosphate- and protein kinase A-dependent facilitation of fast excitatory postsynaptic potentials in the guinea pig ileum/myenteric plexus (Galligan et al. 2003; Pan and Galligan 1994). Data from studies in which mechanical activity is measured using isolated tissues demonstrates the prejunctional role of the 5-HT₄ receptor in the GI tract. Thus, activation of 5-HT₄ receptors on enteric motor neurons produces an acetylcholine-mediated contraction of ileal and colonic longitudinal smooth muscle (Taniyama et al. 2000; Wardle and Sanger 1993). Furthermore, 5-HT₄ receptor activation is associated with augmentation of electrically evoked contractions of guinea pig stomach circular smooth muscle (Buchheit and Buhl 1994). While 5-HT₄ receptor stimulation most frequently results in neurogenically mediated inhibition of electrically evoked contractions of human isolated colonic circular smooth muscle, the release of both inhibitory and excitatory neurotransmitters (i.e., NO and acetylcholine, respectively), together with inhibition and augmentation of neurogenic responses, has been demonstrated (Cellek et al. 2006; Leclere et al. 2005). Activation of 5-HT₄ receptors expressed on smooth muscle cells results in relaxation of the rat esophagus and stomach (Komada and Yano 2007; Reeves et al. 1991), canine rectum (Prins et al. 1999b), and human colon (Tam et al. 1994).

The 5-HT₄ receptor has a prosecretory role in the GI tract (Hansen and Skadhauge 1997). Activation of 5-HT₄ receptors on submucosal enteric neurons or enterocytes promotes chloride and bicarbonate secretion from duodenal, colonic, or jejunal epithelial cells (Budhoo et al. 1996; Kellum et al. 1994; Ning et al. 2004; Säfsten et al. 2006). In human ileum and ascending colon, 5-HT produces mucosal chloride secretion via activation of 5-HT₄ receptors (Borman and Burleigh 1996, 1997b). Similarly, in the rat distal colon, antagonism of 5-HT-mediated chloride secretion by SC 53606, a selective 5-HT₄ receptor antagonist, and the agonist rank order of potency are consistent with a prosecretory role for the 5-HT₄ receptor (Budhoo et al. 1996). In mice, 5-hydroxytryptophan-induced diarrhea is inhibited by the selective 5-HT₄ receptor antagonists, RS 39604, GR 113808, and SB 204070, an action attributed to attenuation of 5-HT₄ receptor-mediated secretion (Hegde et al. 1994; Pascual et al. 2002).

Rodent data suggest that 5-HT₄ receptor activation may reduce visceral hyperalgesia. Thus, the 5-HT₄ receptor agonist, tegaserod, attenuates the visceromotor response

evoked by colorectal distension in control animals and in those in which the colon has been rendered hypersensitive with intracolonic infusion of acetic acid or trinitrobenzenesulfonic acid (Greenwood-Van Meerveld et al. 2006b). This activity is attributed to 5-HT₄ receptor agonist activity of tegaserod as 5-HT₄ receptor-selective antagonists reverse, albeit incompletely, its actions (Greenwood-Van Meerveld et al. 2006a, b). Another study demonstrated that intragastric dosing of tegaserod to rats reduces the number of Fos-labeled neurons and substance P immunoreactivity in the dorsal horn of the lumbarsacral spinal cord following intracolonic instillation of trinitrobenzenesulfonic acid (Sun and Luo 2004). In contrast, 5-HT₄ receptor agonists have been shown to lack a direct antinociceptive effect on visceral afferents (Hicks et al. 2001). In healthy human subjects, tegaserod attenuated inhibition of the RIII nociception reflex caused by rectal distension, although its effect on corresponding symptom ratings was not significantly different from placebo (Coffin et al. 2003). In comparison, another 5-HT₄ receptor agonist, prucalopride, appears to lack an effect on rectal sensitivity (Emmanuel et al. 1998; Poen et al. 1999). The precise role of the 5-HT₄ receptor in visceral sensitivity remains unclear.

Appreciation of the role played by the 5-HT₄ receptor in the GI tract has advanced considerably due to the availability of 5-HT₄ receptor ligands. Several 5-HT₄ receptor-selective agonists (e.g., prucalopride, TS-951, and TD-5108) and antagonists (e.g., piboserod and GR 113808) have been described (Beattie et al. 2007; Briejer et al. 2001b; Gale et al. 1994; Kajita et al. 2001; Smith et al. 2007; Wardle et al. 1996). Tegaserod, cisapride, mosapride, BIMU-1, and renzapride have also been used clinically or as pharmacological tools to probe the function of the 5-HT₄ receptor, although each lacks selectivity for the 5-HT₄ receptor. Thus, tegaserod has significant binding affinity at the 5-HT_{1B}, 5-HT_{1D}, 5-HT_{2A}, and 5-HT₇ receptors and potent 5-HT_{2B} receptor antagonist activity, while cisapride, mosapride, BIMU-1, and renzapride have varying levels of affinity for the 5-HT₃ receptor (Beattie et al. 2004; Freeman et al. 1992; Rizzi et al. 1994; Taniyama et al. 1991; Theravance, unpublished observations). Tegaserod also has some affinity for SERT (K_i of approximately 3 μM; Ismail et al. 2007). Cisapride has significant affinity at dopamine D₂, α₁ adrenergic, and 5-HT_{2B} receptors (pK_i values of 7.7, 8.1, and 8.4, respectively; Theravance, unpublished observations).

Clinical activity in gastrointestinal disorders

Based on the robust prokinetic activity of 5-HT₄ receptor agonists in the upper or lower GI tract of healthy human subjects (Bouras et al. 1999; Coffin et al. 2003; Degen et al. 2001; Emmanuel et al. 1998; Goldberg et al. 2007a, b; Poen et al. 1999), the utility of this class of agents for the

treatment of patients with disorders of reduced GI motility has been investigated.

Clinical efficacy of cisapride, prucalopride, renzapride, mosapride, tegaserod, and TD-5108, among others, has been reported in patients with IBS-C, chronic constipation, functional dyspepsia, or gastroparesis (Abell et al. 1991; Camilleri 2001; Camilleri et al. 2004; Deruyttere et al. 1987; Emmanuel et al. 2002; Johanson 2004; Muller-Lissner 1987). A meta-analysis of clinical trials performed with cisapride in functional dyspepsia concluded that there was significant benefit in favor of cisapride with respect to global assessment of efficacy by investigators and patients alike, in addition to improvement in individual measures of epigastric pain and discomfort, early satiation, abdominal distension, and nausea (Veldhuyzen van Zanten et al. 2001). As cisapride lacks 5-HT₄ receptor selectivity, the extent of the involvement of this receptor in its clinical efficacy is unclear. With respect to tegaserod, some data suggest that it provides modest benefit in improving functional dyspepsia symptoms (Tack et al. 2001). A more robust effect of tegaserod has been demonstrated in many placebo-controlled studies in healthy subjects and IBS-C patients; a reduction in oral–cecal transit time accompanied by an increased frequency of spontaneous bowel movements and softness of stools results (Degen et al. 2005; Muller-Lissner et al. 2001; Prather et al. 2000). Tegaserod also alleviates abdominal discomfort and pain in IBS-C patients (Camilleri 2001; Muller-Lissner et al. 2001). Reductions in visceral sensitivity, when reported, may reflect normalization of bowel function rather than a direct effect on sensory neurons. However, analysis of data from all randomized, placebo-controlled trials performed prior to 2003 indicates that although tegaserod offers significant benefit in the treatment of IBS-C, the NNT ranges from 14 to 20 patients depending on the dose evaluated (Evans et al. 2004). The limited effectiveness of tegaserod may reflect its mixed pharmacology, discussed above. It has been postulated that antagonism of 5-HT_{2A} and 5-HT_{2B} receptors may reduce GI motility (Briejer et al. 1995b; Beattie et al. 2004) and thus interfere with 5-HT₄ receptor-mediated prokinetic activity. Tegaserod and other more selective 5-HT₄ receptor agonists (e.g., prucalopride and TD-5108) have also proven effective in patients with chronic constipation (Emmanuel et al. 2002; Sloots et al. 2002; Goldberg et al. 2007b).

Some limited clinical data suggest that tegaserod and therefore potentially other 5-HT₄ receptor agonists may have a beneficial role in the treatment of functional heartburn, GERD, and gastroparesis. Tegaserod improves the esophageal pain threshold to mechanical distension and upper GI symptoms in patients with functional heartburn (Rodriguez-Stanley et al. 2006). Case reports suggest that tegaserod may also be useful in the treatment of gastroparesis (Banh et al. 2005; Friedenbergs and Parkman 2006;

Zuberi et al. 2005). Additionally, tegaserod is associated with a significant reduction in postprandial esophageal acid exposure in GERD patients, possibly as a result of improved esophageal acid clearance, enhanced gastric emptying, or reduced transient lower esophageal sphincter relaxations (Kahrilas et al. 2000). The use of scintigraphy in healthy human subjects has demonstrated that tegaserod can indeed accelerate gastric emptying (Degen et al. 2005). Similarly, the 5-HT₄ receptor-selective agonist, ATI-7505, showed a trend toward increased gastric emptying in healthy volunteers (Camilleri et al. 2006). In a small clinical study, the 5-HT₃ receptor antagonist/5-HT₄ receptor agonist, renzapride, increased liquid gastric emptying in patients with diabetic gastroparesis but not in healthy control subjects (Mackie et al. 1991). Future large-scale clinical studies with more selective 5-HT₄ receptor agonists should establish the clinical activity, if any, attributable to this mechanism in upper GI disorders. The unique dual pharmacological profile of renzapride, which differentiates it from other drugs currently in development for the treatment of IBS, may also help clarify the relative roles of the 5-HT₃ and 5-HT₄ receptors in this disorder. Renzapride produces increases in colonic transit, improvement in stool form, and ease of passage and also reduces abdominal pain in patients with IBS-C (Camilleri et al. 2004; Tack et al. 2006b). Renzapride is currently in phase 3 clinical trials for IBS-C.

The clinical use of cisapride and tegaserod is now restricted on the basis of cardiovascular safety concerns (Barbey et al. 2002; Pasricha 2007). Restrictions on the use of cisapride stem from reports of serious cardiac arrhythmias including ventricular tachycardia, ventricular fibrillation, and torsades de pointes. The occurrence of these adverse effects is further compounded by metabolism of cisapride by the cytochrome P450 3A4 isoenzyme leading to drug–drug interactions with coadministered substrates (Zhou et al. 2005). Cisapride is now restricted to the treatment of severe nighttime heartburn in patients with GERD who do not adequately respond to other therapies and in consequence has limited use. In March 2007, the marketing of tegaserod was suspended in the USA due to concerns with respect to an increased risk of serious ischemic cardiovascular events. An analysis of pooled clinical data from 29 studies involving 11,614 patients on tegaserod and 7,031 on placebo revealed that 13 patients treated with tegaserod (or 0.1%) had serious and life-threatening cardiovascular insults; four had a myocardial infarction (one death), six had unstable angina, and three had a stroke. In the placebo-treated patients, only one (or 0.01%) had a stroke. All of the affected patients had preexisting cardiovascular disease (e.g., prior coronary artery disease) and/or risk factors, namely advancing age, tobacco use, hypertension, or hyperlipidemia (Gerson

2007). While the cardiovascular adverse effects of cisapride are attributed to QT prolongation of the electrocardiogram (ECG) signal, as a result of cardiac human ether-a-go-go related gene (hERG) potassium channel blockade, this is unlikely to be true for tegaserod as it is devoid of such activity (Drici et al. 1999). Furthermore, its adverse effects are ischemic in nature rather than arrhythmogenic. ECG analysis indicates that there is a higher frequency of ST-segment depression in high-risk patients on tegaserod compared to placebo (3.7% vs. 1.3%), although postmarketing cardiac adverse events are claimed to occur at a level similar to that of background (Pasricha 2007).

Assuming that the perceived cardiovascular risk associated with tegaserod is genuine, what is the potential mechanism responsible, and what are the implications for other 5-HT₄ receptor agonists? In cardiac muscle, 5-HT₄ receptor activation may be associated with tachycardia and potentially arrhythmias in a pathophysiological setting (Kaumann and Levy 2006). With the exception of renzapride (Meyers and Hickling 2007), tachycardia is not evident in the clinical literature with a 5-HT₄ receptor agonist devoid of significant hERG activity. Several 5-HT receptor types exist in the heart, at the level of the myocardium and its neuronal innervation, but their roles are unclear (Doggrell 2003; Jordan 2005; Kaumann and Levy 2006). With respect to the vasculature, 5-HT₄ receptors are present in some blood vessels and, for example, have been shown to mediate dilation of pulmonary veins (Cocks and Arnold 1992). Non-5-HT₄ receptors are expressed in the vasculature; 5-HT_{1B} and 5-HT_{2A} receptor activation results in vasoconstriction (Nilsson et al. 1999), while 5-HT_{1B}, 5-HT_{1D}, 5-HT_{2B}, and 5-HT₇ receptor activation in either the endothelium or vascular smooth muscle is associated with vasodilation (Centurion et al. 2000; Ellis et al. 1995; Schoeffter and Hoyer 1990; Ishida et al. 1998). 5-HT_{1B} or 5-HT_{2B} receptor activation on vascular endothelium may, under pathophysiological conditions, provide protective vasodilator and platelet antiaggregatory effects via release of NO (Ishida et al. 1998). Furthermore, there is evidence indicating that the 5-HT_{2B} receptor has an important trophic role in cardiac development and that 5-HT_{2B} receptors on cardiac fibroblasts may be involved in tissue regenerative processes and therefore be of potential importance in some pathological scenarios (Jaffré et al. 2004; Nebigil et al. 2000; Nebigil and Maroteaux 2003). As stated above, tegaserod has significant 5-HT_{1B}, 5-HT_{1D}, 5-HT_{2A}, 5-HT_{2B}, and 5-HT₇ receptor affinity and/or antagonist potency. An interaction of tegaserod with one or more of these receptors in the heart or vasculature may be relevant to its perceived cardiovascular risk, although this requires further investigation. It also remains to be established whether other 5-HT₄ receptor agonists currently in development, particularly those with a higher degree of 5-HT₄

receptor selectivity, will prove to be more efficacious and/or better tolerated than tegaserod. TD-5108, under development by Theravance, is a 5-HT₄ receptor agonist with a high degree of selectivity over other 5-HT receptors (Goldberg et al. 2007a, b; Smith et al. 2007). TD-5108 was recently found to be highly effective and well tolerated in a 4-week phase 2a trial in patients with chronic constipation (Goldberg et al. 2007b). Prucalopride, a 5-HT₄ receptor-selective agonist (Briejer et al. 2001a), has GI prokinetic activity in healthy human subjects and patients with chronic constipation (Coremans et al. 2003; Emmanuel et al. 2002; Poen et al. 1999). Following completion of multiple phase 3 trials in chronic constipation, development of prucalopride was halted by Janssen for several years, allegedly on the basis of carcinogenicity concerns (Cash and Chey 2005; Kamm 2002). Recently, the compound was licensed to Movetis whose objective is to gain registration in Europe. Thus, the efficacy of several 5-HT₄ receptor agonists in treating patients with IBS-C or chronic constipation is established. While it remains to be determined whether the modest clinical efficacy of tegaserod outweighs its small, possible risk of cardiovascular adverse effects, the development of more selective 5-HT₄ receptor agonists such as TD-5108 and prucalopride may provide much needed benefit to patients requiring GI prokinetic therapy.

On the basis of the perceived excitatory influence of 5-HT₄ receptor agonists on GI motility, it has been postulated that 5-HT₄ receptor antagonism could confer benefit in patients with IBS-D. In healthy human subjects, piboserod, a selective 5-HT₄ receptor antagonist, inhibits cisapride-mediated increases in oral–cecal transit and tends to delay colonic transit in its own right without affecting gastric emptying, small intestinal transit, colonic motor activity, or colonic perception to balloon distension (Bharucha et al. 2000). These findings suggest that 5-HT₄ receptors may have only a modest tonic role in GI function under resting physiological conditions. This conclusion is supported by animal data from studies indicating that piboserod and SDZ 205-557 have no effect on stool production in mice or on dog colonic contractile activity per se but can inhibit 5-hydroxytryptophan or 5-HT₄ receptor agonist-induced responses (Nagakura et al. 1996a; Sanger et al. 1998). However, in 5-HT₄ “knockout” mice, gastric emptying, small and large intestinal transit, and the number of myenteric neurons in the colon are all reduced compared to wild-type littermates (Cuenca et al. 2006). Furthermore, in a small clinical study with IBS-D patients, piboserod significantly increased oral–cecal transit time and tended to reduce rectal sensitivity, with the majority of patients reporting symptomatic improvement (Houghton et al. 1999). It remains to be determined whether 5-HT₄ receptor antagonists will be efficacious in disorders such as

carcinoid syndrome where a profound overstimulation of the 5-HT₄ receptor may be anticipated (Bharucha et al. 2000; De Ponti and Tonini 2001). With respect to a potential clinical utility in upper GI disorders, the selective 5-HT₄ receptor antagonist, R216073, was found to have no significant effect on fundic relaxation, drinking capacity, and upper abdominal symptoms in patients with functional dyspepsia (Van Lelyveld et al. 2006).

Metoclopramide is approved for the treatment of GERD and diabetic gastroparesis. In addition to its established, potent dopamine D₂ receptor antagonist activity, 5-HT₄ receptor agonist and 5-HT₃ receptor antagonist activities may contribute to its clinical efficacy (Tonini et al. 1999). However, metoclopramide-mediated extrapyramidal symptoms and tardive dyskinesias, resulting from dopamine D₂ receptor antagonism in the CNS, limit its utility in the treatment of functional GI disorders. The clinical utility of a 5-HT₄ receptor-selective agonist for the treatment of upper GI disorders has yet to be established. Proctor & Gamble and Movetis have such agents in early clinical development for the treatment of GERD and gastroparesis (Table 2). Furthermore, the Movetis company website indicates that they are pursuing the development of 5-HT₄ receptor agonists that are specific for 5-HT₄ receptors in the upper GI tract. The pharmacological basis for this upper GI selectivity is unclear, although the 5-HT₄ receptor splice variants have region-specific expression in the GI tract (see above) and so perhaps differential activation is achievable. Alternatively, differences in 5-HT₄ receptor reserve may confer regional selectivity on the activity of 5-HT₄ receptor partial agonists.

5-HT₅ and 5-HT₆ receptor families

The 5-HT₅ receptor family consists of two members, designated 5-HT_{5A} and 5-HT_{5B}; only the former member is expressed in the human as the coding sequence of the 5-HT_{5B} receptor is interrupted by stop codons (Nelson 2004). Both receptor subtypes are essentially limited in distribution to the CNS, although the 5-HT_{5A} receptor has also been found in the carotid body. There are no data suggesting a role for the 5-HT₅ receptor in GI function. The 5-HT₆ receptor has been cloned from several species including humans (Kohen et al. 1996; Ruat et al. 1993). Within the CNS, there is considerable interest in the role played by the 5-HT₆ receptor in the regulation of feeding, cognition, affective states, and seizures (Woolley et al. 2004), although there appears to be extremely limited 5-HT₆ receptor expression in the periphery. Despite the interaction of the 5-HT₆ receptor with many neurotransmitter systems known to influence GI function (e.g., cholinergic, dopaminergic, and GABAergic; Mitchell and Neumaier

2005) and detection of weak receptor expression in the rat stomach (Ruat et al. 1993), there are no definitive data demonstrating a functional role for the 5-HT₆ receptor in the GI tract.

5-HT₇ receptors

The 5-HT₇ receptor has been cloned from a number of species (e.g., rat, guinea pig, and human; Bard et al. 1993; Shen et al. 1993; Tsou et al. 1994), and three human splice variants, with similar pharmacology, have been identified. The 5-HT₇ receptor has been detected in the CNS and periphery (Vanhoenacker et al. 2000), although the majority of physiological effects attributed to 5-HT₇ receptor activation occur in the CNS (e.g., increased excitability of hippocampal neurons, regulation of prefrontal cortex development, and thermoregulation; Bacon and Beck 2000; Beique et al. 2004; Hedlund et al. 2003). However, the 5-HT₇ receptor is present in the ENS and is postulated to play a role in GI physiology. Each of the 5-HT₇ receptor splice variants is present in the human stomach, small intestine, and colon (Irving et al. 2007; Jasper et al. 1997; Krobert et al. 2001), while 5-HT₇ receptor immunoreactivity is present in myenteric and submucosal IPANs, NO synthase- and VIP-immunoreactive descending neurons, and in smooth muscle cells of the ileum (Tonini et al. 2005). Activation of 5-HT₇ receptors on IPANs results in a slow excitatory postsynaptic potential, while stimulation of 5-HT₇ receptors in the smooth muscle of the ileum and colon produces relaxation (Carter et al. 1995; Prins et al. 1999a). The ability of SB-269970, a selective 5-HT₇ receptor antagonist, to inhibit 5-HT-induced slow depolarization and excitatory postsynaptic potentials of myenteric IPANs and to reduce accommodation of the circular muscle of the guinea pig ileum during the preparatory phase of peristalsis, is consistent with a physiological role for the 5-HT₇ receptor (Monro et al. 2005; Tonini et al. 2005). With respect to a pathophysiological role for 5-HT₇ receptor activation, it has been postulated that overstimulation of this receptor may lead to exaggerated accommodation of colonic circular muscle and abdominal bloating, a common symptom in many functional bowel disorders (Tonini et al. 2005), although this will remain speculative until supporting clinical data are available.

Serotonin transporter

The paracrine and neurotransmitter actions of 5-HT are terminated rapidly and efficiently by SERT (Ramamoorthy et al. 1993). SERT plays a critical role in ensuring that 5-HT-mediated toxicity and receptor desensitization, likely consequences of maintained agonist exposure, are avoided.

Rat, guinea pig, and human mucosal epithelial cells, in common with serotonergic neurons, express mRNA encoding SERT, display SERT immunoreactivity, and transport 5-HT with high specificity (Camilleri et al. 2007; Chen et al. 1998; Coates et al. 2004; Takayanagi et al. 1995; Wade et al. 1996). SERT is expressed at both the apical and basolateral sides of the mucosal cell membrane in the human and rodent GI tract (Van Lelyveld et al. 2007; Martel et al. 2003; Martel 2006). Variation in the density of SERT expression is apparent in human mucosal biopsy samples; levels in the duodenum are markedly higher than those in the stomach fundus and antrum (Van Lelyveld et al. 2007). Any 5-HT that is not “captured” by the mucosa either overflows into the lumen of the GI tract or enters the bloodstream and is rapidly transported into platelets, which also express SERT (Chen et al. 2001; Grønstad et al. 1985; Gershon and Tack 2007). Considering the importance of SERT in removing 5-HT from the vicinity of its many receptors in the GI tract, interference with its function has a major impact on GI physiology. Analysis of rectal biopsies from patients with IBS has demonstrated reductions in SERT mRNA and immunoreactivity in patients with IBS-C and IBS-D (Coates et al. 2004). Furthermore, significant association is observed between an insertion/deletion polymorphism of the promoter region of the human SERT gene and IBS-D suggesting that this and/or other polymorphisms may contribute to GI functional disorders such as IBS-D at least in some individuals (Yeo et al. 2004). Polymorphisms at the SERT promoter also appear to influence colonic responsiveness to the 5-HT₃ receptor antagonist, alosetron, in IBS-D patients (Camilleri et al. 2002).

It is noteworthy that the SSRIs, paroxetine, sertraline, citalopram, and fluoxetine, which are used widely in the treatment of depression, panic disorder, and obsessive-compulsive disorder, are associated with a relatively high incidence of nausea and diarrhea (approximately 20%; Spigset 1999). These adverse effects would be consistent with increased 5-HT-mediated activation of 5-HT₃ and 5-HT₄ receptors in the ENS, respectively. The impact of SERT inhibition on GI function will presumably be dictated by the relative contributions and interplay of each 5-HT receptor at distinct locations in the digestive system, together with the degree and rapidity of any mucosal or neuronal 5-HT receptor desensitization, among other factors. Thus, fluoxetine-induced contraction of the mouse isolated stomach fundus is mediated via 5-HT_{1B} or 5-HT_{1D} receptor activation on the basis of the inhibitory effect of the selective antagonist, GR 127935 (Xue et al. 2006), while in guinea pig isolated antrum and fundus, 5-HT₄ receptors are implicated (James et al. 2005). The 5-HT₄ receptor agonist, tegaserod, was shown recently to inhibit SERT with micromolar potency (Ismair et al. 2007). It has been suggested that inhibition of SERT by high local

concentrations of tegaserod in the GI tract may synergize with 5-HT₄ receptor agonist-induced prokinetic activity (Ismair et al. 2007).

The data from several human studies have demonstrated SSRI-mediated alterations in GI function, presumably as a consequence of an enhancement of 5-HT exposure in close proximity to its receptors. Paroxetine stimulates motor activity in the human small intestine, increasing oral–cecal motility and transit, but appears to have no significant effect on gastric emptying or colonic transit in healthy human subjects (Chial et al. 2003a,b; Gorard et al. 1994). Similarly, sertraline has no effect on gastric compliance or sensitivity in healthy subjects (Ladabaum and Glidden 2002). However, citalopram is associated with increased colonic compliance and phasic activity and increased colonic tone in response to a meal (Tack et al. 2000). In the esophagus, citalopram reduces chemical (acid perfusion) and mechanical (balloon distension) sensitivity without altering motility in healthy humans possessing established esophageal hypersensitivity (Broekaert et al. 2006). In addition to human data, preclinical observations indicate that SSRIs influence GI physiology. Chronic administration of paroxetine to mice reduces upper GI transit, defecation, and colonic sensitivity in response to balloon distension (Coates et al. 2006), while stool water content and colonic motility are increased in the majority of mice with a targeted deletion of SERT. In SERT knockout mice, a subset of animals is constipated, and analysis of GI function in the longer term indicates an alternating pattern of diarrhea and constipation, a phenomenon reminiscent of that in a subset of patients with IBS (Chen et al. 2001). Although SERT is absent in these mice, other mechanisms (e.g., the dopamine transporter) appear to compensate for the deletion, albeit with lower 5-HT-binding affinity compared to SERT.

Clinical data indicate beneficial effects of tricyclic antidepressants in IBS and to a lesser extent, functional dyspepsia (Mertz et al. 1998; Talley 2003b). The clinical efficacy of amitriptyline in functional dyspepsia appears unrelated to any changes in perception of gastric distension, and increased tolerance to aversive visceral sensations may be responsible (Mertz et al. 1998). While tricyclic antidepressants inhibit SERT, they also inhibit NET and interact with a variety of non-5-HT receptors. In consequence, it is difficult to attribute any clinical efficacy definitively to inhibition of SERT. There are, however, some limited clinical data with the SSRIs suggesting that SERT inhibition can confer a therapeutic benefit in patients with functional GI disorders (Creed et al. 2003; Tack et al. 2006a; Talley 2004). As many patients with functional GI disorders have coexisting psychiatric conditions (e.g., depression and anxiety), there has been some discussion related to the precise mechanism of any GI clinical efficacy where it has

been noted (Talley et al. 2004). While SERT inhibition in both the CNS and periphery may provide benefits, it remains unclear whether antidepressant or anxiolytic activity contributes to a successful outcome in patients with functional GI disorders. Thus, in one study, citalopram improved abdominal pain, bloating, and quality-of-life measures in nondepressed IBS patients with no correlation in efficacy to changes in depression or anxiety scores (Tack et al. 2006a), but in another, fluoxetine had no effect on rectal sensitivity or global symptom relief in nondepressed IBS patients (Kuiken et al. 2003). There are limited clinical data regarding the efficacy of the new generation of dual 5-HT and norepinephrine reuptake inhibitors, such as venlafaxine and duloxetine. Two novel development compounds of interest are LX-1031 (Lexicon Pharmaceuticals), a selective inhibitor of TPH1, which is responsible for 5-HT synthesis in the GI tract, and tianeptine (Pharmos), which is reported to stimulate 5-HT uptake.

Concluding remarks

In recent years, our understanding of the physiological and pathophysiological roles of the serotonergic system in the GI tract has increased significantly and continues to do so. A number of 5-HT receptor agonists and antagonists have demonstrable clinical efficacy in a variety of functional GI disorders, and preclinical and clinical data suggest that additional opportunities in the future can be anticipated as a result of manipulation of the serotonergic system (Table 2). As with all drug development, clinical failure in terms of inadequate efficacy or unacceptable tolerability has been and will continue to be a challenge. However, advances in our appreciation of the complexity of the ENS and the identification of more selective receptor ligands with suitable properties for therapeutic development bodes well for the future in addressing the clear unmet medical need that remains in the treatment of functional GI disorders.

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References

- Aapro M (2005) 5-HT(3)-receptor antagonists in the management of nausea and vomiting in cancer and cancer treatment. *Oncology* 69:97–109

- Abell TL, Camilleri M, DiMugno EP, Hench VS, Zinsmeister AR, Malagelada JR (1991) Long-term efficacy of oral cisapride in symptomatic upper gut dysmotility. *Dig Dis Sci* 36:616–620
- Adham N, Kao HT, Schechter LE, Bard J, Olsen M, Urquhart D, Durkin M, Hartig PR, Weinschank RL, Branchek TA (1993) Cloning of another human serotonin receptor (5-HT_{1F}): a fifth 5-HT₁ receptor subtype coupled to the inhibition of adenylate cyclase. *Proc Natl Acad Sci USA* 90:408–412
- Alexander SPH, Mathie A, Peters JA (2004) Guide to receptors and channels. *Br J Pharmacol* 141(Suppl 1):S36–S37
- Alfieri AB, Cubeddu LX (1995) Comparative efficacy of a single oral dose of ondansetron and of buspirone against cisplatin-induced emesis in cancer patients. *Br J Cancer* 72:1013–1015
- American Gastroenterological Association (2002) The burden of gastrointestinal diseases. American Gastroenterological Association, Bethesda, MD
- Atkinson W, Lockhart S, Whorwell PJ, Keevil B, Houghton LA (2006) Altered 5-hydroxytryptamine signaling in patients with constipation- and diarrhea-predominant irritable bowel syndrome. *Gastroenterology* 130:34–43
- Bacon WL, Beck SG (2000) 5-Hydroxytryptamine(7) receptor activation decreases slow afterhyperpolarization amplitude in CA3 hippocampal pyramidal cells. *J Pharmacol Exp Ther* 294:672–679
- Bai F, Yin T, Johnstone EM, Su C, Varga G, Little SP, Nelson DL (2004) Molecular cloning and pharmacological characterization of the guinea pig 5-HT_{1E} receptor. *Eur J Pharmacol* 484:127–139
- Baker DE (2005) Rationale for using serotonergic agents to treat irritable bowel syndrome. *Am J Health-Syst Pharm* 62:700–711
- Banh HL, MacLean C, Topp T, Hall R (2005) The use of tegaserod in critically ill patients with impaired gastric motility. *Clin Pharmacol Ther* 77:583–586
- Banner SE, Sanger GJ (1995) Differences between 5-HT₃ receptor antagonists in modulation of visceral hypersensitivity. *Br J Pharmacol* 114:558–562
- Barbey JT, Lazzara R, Zipes DP (2002) Spontaneous adverse event reports of serious ventricular arrhythmias, QT prolongation, syncope, and sudden death in patients treated with cisapride. *J Cardiovasc Pharmacol Ther* 7:65–76
- Bard JA, Zgombick J, Adham N, Vaysse P, Branchek TA, Weinschank RL (1993) Cloning of a novel human serotonin receptor (5-HT₇) positively linked to adenylate cyclase. *J Biol Chem* 268:23422–23426
- Barnes NM, Sharp T (1999) A review of central 5-HT receptors and their function. *Neuropharmacology* 38:1083–1152
- Beattie DT, Smith JA, Marquess D, Vickery RG, Armstrong SR, Pulido-Rios T, McCullough JL, Sandlund C, Richardson C, Mai N, Humphrey PPA (2004) The 5-HT₄ receptor agonist, tegaserod, is a potent 5-HT_{2B} receptor antagonist in vitro and in vivo. *Br J Pharmacol* 143:549–560
- Beattie DT, Armstrong SR, Marquess D, Shaw J, Smith JA, Humphrey PP (2007) The in vivo preclinical profile of TD-5108, a selective, high intrinsic activity 5-HT₄ receptor agonist. *Digestive Disease Week, Washington, DC, W1228*
- Beique JC, Chapin-Pennick EM, Mladenovic L, Andrade R (2004) Serotonergic facilitation of synaptic activity in the developing rat prefrontal cortex. *J Physiol* 556:739–754
- Bender E, Pindon A, Van Oers I, Zhang YB, Gommeren W, Verhasselt P, Jurzak M, Leysen J, Luyten W (2000) Structure of the human serotonin 5-HT₄ receptor gene and cloning of a novel 5-HT₄ splice variant. *J Neurochem* 74:478–489
- Berman SM, Chang L, Suyenobu B, Derbyshire SW, Stains J, Fitzgerald L, Mandelkern M, Hamm L, Vogt B, Naliboff BD, Mayer EA (2002) Condition-specific deactivation of brain regions by 5-HT₃ receptor antagonist alosetron. *Gastroenterology* 123:969–977
- Bertrand PP, Kunze WA, Furness JB, Bornstein JC (2000) The terminals of myenteric intrinsic primary afferent neurons of the guinea-pig ileum are excited by 5-hydroxytryptamine acting at 5-hydroxytryptamine-3 receptors. *Neuroscience* 101:459–469
- Bharucha AE, Camilleri M, Haydock S, Ferber I, Burton D, Cooper S, Tompson D, Fitzpatrick K, Higgins R, Zinsmeister AR (2000) Effects of a serotonin 5-HT₄ receptor antagonist SB-207266 on gastrointestinal motor and sensory function in humans. *Gut* 47:667–674
- Boeckxstaens GE, Tytgat GN, Wajs E, Van Nueten L, De Ridder F, Meulemans A, Tack J (2006) The influence of the novel 5-HT_{1A} agonist R137696 on the proximal stomach function in healthy volunteers. *Neurogastroenterol Motil* 18:919–926
- Bonaventure P, Nepomuceno D, Miller K, Chen J, Kuei C, Kamme F, Tran DT, Lovenberg TW, Liu C (2005) Molecular and pharmacological characterization of serotonin 5-HT_{2A} and 5-HT_{2B} receptor subtypes in dogs. *Eur J Pharmacol* 513:181–192
- Borman RA, Burleigh DE (1995) Functional evidence for a 5-HT_{2B} receptor mediating contraction of longitudinal muscle in human small intestine. *Br J Pharmacol* 114:1525–1527
- Borman RA, Burleigh DE (1996) Human colonic mucosa possesses a mixed population of 5-HT receptors. *Eur J Pharmacol* 309:271–274
- Borman RA, Burleigh DE (1997a) 5-HT_{1D} and 5-HT_{2B} receptors mediate contraction of smooth muscle in human small intestine. *Ann NY Acad Sci* 812:222–223
- Borman RA, Burleigh DE (1997b) Heterogeneity of 5-HT receptors mediating secretion in the human intestine. *Ann NY Acad Sci* 812:224–225
- Borman RA, Tilford NS, Harmer DW, Day N, Ellis ES, Sheldrick RL, Carey J, Coleman RA, Baxter GS (2002) 5-HT_(2B) receptors play a key role in mediating the excitatory effects of 5-HT in human colon in vitro. *Br J Pharmacol* 135:1144–1151
- Bouras EP, Camilleri M, Burton DD, McKinzie S (1999) Selective stimulation of colonic transit by the benzofuran 5HT₄ agonist, prucalopride, in healthy humans. *Gut* 44:682–686
- Bradesi S, Lao L, McLean PG, Winchester WJ, Lee K, Hicks GA, Mayer EA (2007) Dual role of 5-HT₃ receptors in a rat model of delayed stress-induced visceral hyperalgesia. *Pain* 130:56–65
- Branchek TA, Mawe GM, Gershon MD (1988) Characterization and localization of a peripheral neural 5-hydroxytryptamine receptor subtype (5-HT_{1P}) with a selective agonist, ³H-5-hydroxyindalpine. *J Neurosci* 8:2582–2595
- Brattellid T, Kvingsdal AM, Krobert KA, Andressen KW, Bach T, Hystad ME, Kaumann AJ, Levy FO (2004) Cloning, pharmacological characterisation and tissue distribution of a novel 5-HT₄ receptor splice variant, 5-HT_{4(i)}. *Naunyn Schmiedeberg's Arch Pharmacol* 369:616–628
- Briejer MR, Akkermans LM, Schuurkes JA (1995a) Interactions of serotonin with multiple receptors and neurotransmitters in the guinea-pig isolated colon. *Arch Int Pharmacodyn Ther* 329:121–133
- Briejer MR, Akkermans LMA, Schuurkes JAJ (1995b) Gastrointestinal prokinetic benzamides: the pharmacology underlying stimulation of motility. *Pharmacol Rev* 47:631–651
- Briejer MR, Bosmans JP, Van Daele P, Jurzak M, Heylen L, Leysen JE, Prins NH, Schuurkes JAJ (2001a) The in vitro pharmacological profile of prucalopride, a novel enterokinetic compound. *Eur J Pharmacol* 423:71–83
- Briejer MR, Prins NH, Schuurkes JA (2001b) Effects of the enterokinetic prucalopride (R093877) on colonic motility in fasted dogs. *Neurogastroenterol Motil* 13:465–472
- Broekaert D, Fischler B, Sifrim D, Janssens J, Tack J (2006) Influence of citalopram, a selective serotonin reuptake inhibitor, on oesophageal hypersensitivity: a double-blind, placebo-controlled study. *Aliment Pharmacol Ther* 23:365–370

- Brown NJ, Horton A, Rumsey RD, Read NW (1993) Granisetron and ondansetron: effects on the ileal brake mechanism in the rat. *J Pharm Pharmacol* 45:521–524
- Buchheit KH, Buhl T (1994) Stimulant effects of 5-hydroxytryptamine on guinea pig stomach preparations in vitro. *Eur J Pharmacol* 262:91–97
- Budhoo MR, Harris RP, Kellum JM (1996) 5-Hydroxytryptamine-induced Cl⁻ transport is mediated by 5-HT₃ and 5-HT₄ receptors in the rat distal colon. *Eur J Pharmacol* 298:137–144
- Calvert EL, Whorwell PJ, Houghton LA (2004) Inter-digestive and post-prandial antro-pyloro-duodenal motor activity in humans: effect of 5-hydroxytryptamine agonism. *Aliment Pharmacol Ther* 19:805–815
- Camilleri M (2001) Review article: tegaserod. *Aliment Pharmacol Ther* 15:277–289
- Camilleri M, Northcutt AR, Kong S, Dukes GE, McSorley D, Mangel AW (2000) Efficacy and safety of alosetron in women with irritable bowel syndrome: a randomised, placebo-controlled trial. *Lancet* 355:1035–1040
- Camilleri M, Atanasova E, Carlson PJ, Ahmad U, Kim HJ, Viramontes BE, McKinzie S, Urrutia R (2002) Serotonin-transporter polymorphism pharmacogenetics in diarrhea-predominant irritable bowel syndrome. *Gastroenterology* 123:425–432
- Camilleri M, McKinzie S, Fox J, Foxx-Orenstein A, Burton D, Thomforde G, Baxter K, Zinsmeister AR (2004) Effect of renzapride on transit in constipation-predominant irritable bowel syndrome. *Clin Gastroenterol Hepatol* 2:895–904
- Camilleri M, Burton D, Vasquez-Roque MI, Ford T, McKinzie S, Zinsmeister AR (2006) Effects of a novel 5-HT₄ agonist, ATI-7505, on gastrointestinal and colonic transit in humans. Abstract T2029, Digestive Disease Week, Los Angeles, CA, May 20–25
- Camilleri M, Andrews CN, Bharucha AE, Carlson PJ, Ferber I, Stephens D, Smyrk TC, Urrutia R, Aerssens J, Thielemans L, Gohlmann H, Van den Wyngaert I, Coulie B (2007) Alterations in expression of p11 and SERT in mucosal biopsy specimens of patients with irritable bowel syndrome. *Gastroenterology* 132:17–25
- Carling RS, Degg TJ, Allen KR, Bax ND, Barth JH (2002) Evaluation of whole blood serotonin and plasma and urine 5-hydroxyindole acetic acid in diagnosis of carcinoid disease. *Ann Clin Biochem* 39:577–582
- Carter D, Champney M, Hwang B, Eglen RM (1995) Characterization of a postjunctional 5-HT receptor mediating relaxation of guinea pig isolated ileum. *Eur J Pharmacol* 280:243–250
- Cash BD, Chey WD (2005) Review article: the role of serotonergic agents in the treatment of patients with primary chronic constipation. *Aliment Pharmacol Ther* 22:1047–1060
- Castro L, Mialet-Perez J, Guillemeau A, Stillitano F, Zolk O, Eschenhagen T, Lezoualch F, Bochet P, Fischmeister R (2005) Differential functional effects of two 5-HT₄ receptor isoforms in adult cardiomyocytes. *J Mol Cell Cardiol* 39:335–344
- Cellek S, John AK, Thangiah R, Dass NB, Bassil AK, Jarvie EM, Lalude O, Vivekanandan S, Sanger GJ (2006) 5-HT₄ receptor agonists enhance both cholinergic and nitergic activities in human isolated colon circular muscle. *Neurogastroenterol Motil* 18:853–861
- Centurion D, Sanchez-Lopez A, Ortiz MI, De Vries P, Saxena PR, Villalon CM (2000) Mediation of 5-HT-induced internal carotid vasodilation in GR127935- and ritanserin-pretreated dogs by 5-HT₇ receptors. *Naunyn-Schmiedeberg's Arch Pharmacol* 362:169–176
- Chang L, Chey W, Harris L, Olden K, Surawicz C, Schoenfeld P (2006) Incidence of ischemic colitis and serious complications of constipation among patients using alosetron: systematic review of clinical trials and post-marketing surveillance data. *Am J Gastroenterol* 101:1069–1079
- Chen JJ, Zhishan L, Pan H, Murphy DL, Tamir H, Koepsell H, Gershon MD (2001) Maintenance of serotonin in the intestinal mucosa and ganglia of mice that lack the high-affinity serotonin transporter (SERT): abnormal intestinal motility and the expression of cation transporters. *J Neurosci* 21:6348–6361
- Chen JX, Pan H, Rowbotham TP, Wade PR, Gershon (1998) Guinea-pig 5-HT transporter: cloning, expression, distribution and function in intestinal sensory reception. *Am J Physiol* 275:G433–G448
- Chey WD, Cash BD (2005) Cilansetron: a new serotonergic agent for the irritable bowel syndrome with diarrhoea. *Exp Opin Invest Drugs* 14:185–193
- Chial HJ, Camilleri M, Burton D, Thomforde G, Olden KW, Stephens D (2003a) Selective effects of serotonergic psychoactive agents on gastrointestinal functions in health. *Am J Physiol* 284:G130–G137
- Chial HJ, Camilleri M, Ferber I, Delgado-Aros S, Burton D, McKinzie S, Zinsmeister AR (2003b) Effects of venlafaxine, buspirone, and placebo on colonic sensorimotor functions in healthy humans. *Clin Gastroenterol Hepatol* 1:211–218
- Claeyssen S, Sebben M, Becamel C, Bockaert J, Dumuis A (1999) Novel brain-specific 5-HT₄ receptor splice variants show marked constitutive activity: role of the C-terminal intracellular domain. *Mol Pharmacol* 55:910–920
- Clayton NM, Sargent R, Butler A, Gale J, Maxwell MP, Hunt AA, Barrett VJ, Cambridge D, Bountra C, Humphrey PP (1999) The pharmacological properties of the novel selective 5-HT₃ receptor antagonist, alosetron, and its effects on normal and perturbed small intestinal transit in the fasted rat. *Neurogastroenterol Motil* 11:207–217
- Clemens CHM, Samson M, Van Berge Henegouwen GP, Fabri M, Smout AJPM (2002) Effect of alosetron on left colonic motility in non-constipated patients with irritable bowel syndrome and healthy volunteers. *Aliment Pharmacol Ther* 16:993–1002
- Coates MD, Mahoney CR, Linden DR, Sampson JE, Chen J, Blaszyk H, Crowell MD, Sharkey KA, Gershon MD, Mawe GM, Moses PL (2004) Molecular defects in mucosal serotonin content and decreased serotonin reuptake transporter in ulcerative colitis and irritable bowel syndrome. *Gastroenterology* 126:1657–1664
- Coates MD, Johnson AC, Greenwood-Van Meerveld B, Mawe GM (2006) Effects of serotonin transporter inhibition on gastrointestinal motility and colonic sensitivity in the mouse. *Neurogastroenterol Motil* 18:464–471
- Cocks TM, Arnold PJ (1992) 5-Hydroxytryptamine (5-HT) mediates potent relaxation in the sheep isolated pulmonary vein via activation of 5-HT₄ receptors. *Br J Pharmacol* 107:591–596
- Coffin B, Farmachidi JP, Rueegg P, Bastie A, Bouhassira D (2003) Tegaserod, a 5-HT₄ receptor partial agonist, decreases sensitivity to rectal distension in healthy subjects. *Aliment Pharmacol Ther* 17:577–585
- Cole JA, Cook SF, Sands BE, Ajene AN, Miller DP, Walker AM (2004) Occurrence of colon ischemia in relation to irritable bowel syndrome. *Am J Gastroenterol* 99:486–491
- Coleman NS, Marciani L, Blackshaw E, Wright J, Parker M, Yano T, Yamazaki S, Chan PQ, Wilde K, Gowland PA, Perkins AC, Spiller RC (2003) Effect of a novel 5-HT₃ receptor agonist MKC-733 on upper gastrointestinal motility in humans. *Aliment Pharmacol Ther* 18:1039–1048
- Cooke HJ, Sidhu M, Wang YZ (1997) Activation of 5-HT_{1P} receptors on submucosal afferents subsequently triggers VIP neurons and chloride secretion in the guinea pig colon. *J Auton Nerv Syst* 66:105–110
- Coremans G, Kerstens R, De Pauw M, Stevens M (2003) Prucalopride is effective in patients with severe chronic constipation in whom laxatives fail to provide adequate relief. Results from a double-blind, placebo-controlled clinical trial. *Digestion* 67:82–89

- Costa M, Brookes SJH, Steele PA, Gibbins I, Burcher E, Kandiah CJ (1996) Neurochemical classification of myenteric neurons in the guinea pig ileum. *Neuroscience* 75:949–967
- Costedio MM, Hyman N, Mawe GM (2007) Serotonin and its role in colonic function and in gastrointestinal disorders. *Dis Colon Rectum* 50:376–388
- Coulie B, Tack J, Maes B, Geypens B, De Roo M, Janssens J (1997) Sumatriptan, a selective 5-HT₁ receptor agonist, induces a lag phase for gastric emptying of liquids in humans. *Am J Physiol* 272:G902–G908
- Coulie B, Tack J, Sifrim D, Andrioli A, Janssens J (1999) Role of nitric oxide in fasting gastric fundus tone and in 5-HT₁ receptor-mediated relaxation of gastric fundus. *Am J Physiol* 276:G373–377
- Creed F, Fernandes L, Guthrie E, Palmer S, Ratcliffe J, Read N, Rigby C, Thompson D, Tomenson B (2003) The cost-effectiveness of psychotherapy and paroxetine for severe irritable bowel syndrome. *Gastroenterology* 124:303–317
- Cremonini F, Delgado-Aros S, Camilleri M (2003) Efficacy of alosetron in irritable bowel syndrome: a meta-analysis of randomized controlled trials. *Neurogastroenterol Motil* 15:79–86
- Croci T, Landi M, Bianchetti A, Manara L (1995) Drug-induced defaecation in rats: role of central 5-HT_{1A} receptors. *Br J Pharmacol* 115:203–209
- Cubeddu LX, Hoffman IS, Fuenmayor NT, Finn AL (1990) Efficacy of ondansetron (GR 38032F) and the role of serotonin in cisplatin-induced nausea and vomiting. *New Eng J Med* 322:810–816
- Cuenca A, Liu MT, Holick K, Hen R, Gershon MD (2006) Physiological dependence of gastric emptying and intestinal motility on 5-HT₄ receptors: analyses in wild-type and 5-HT₄ knockout mice. *Gastroenterology* 130(Suppl S2):A5
- Davies PA, Pistis M, Hanna MC, Peters JA, Lambert JJ, Hales TG, Kirkness EF (1999) The 5-HT_{3B} subunit is a major determinant of serotonin-receptor function. *Nature* 397:359–363
- Degen L, Matzinger D, Merz M, Appel-Dingemanse S, Osborne S, Luchinger S, Bertold R, Maecke H, Beglinger C (2001) Tegaserod, a 5-HT₄ receptor partial agonist, accelerates gastric emptying and gastrointestinal transit in healthy male subjects. *Aliment Pharmacol Ther* 15:1745–1751
- Degen L, Petrig C, Studer D, Schroll S, Beglinger C (2005) Effect of tegaserod on gut transit in male and female subjects. *Neurogastroenterol Motil* 17:821–826
- De Ponti F, Tonini M (2001) Irritable bowel syndrome: new agents targeting serotonin receptor subtypes. *Drugs* 61:317–332
- De Ponti F, Crema F, Moro E, Nardelli G, Frigo G, Crema A (2003) Role of 5-HT_{1B/D} receptors in canine gastric accommodation: effect of sumatriptan and 5-HT_{1B/D} receptor antagonists. *Am J Physiol* 285:G96–G104
- Deruyttere M, Lepoutre L, Heylen H, Samain H, Pennoit H (1987) Cisapride in the management of chronic functional dyspepsia: a multicenter, double-blind, placebo-controlled study. *Clin Ther* 10:44–51
- Doggrell SA (2003) The role of 5-HT on the cardiovascular and renal systems and the clinical potential of 5-HT modulation. *Expert Opin Invest Drugs* 12:805–823
- Drici MD, Ebert SN, Wang WX, Rodriguez I, Liu XK, Whitfield BH, Woosley RL (1999) Comparison of tegaserod (HTF 919) and its main human metabolite with cisapride and erythromycin on cardiac repolarization in the isolated rabbit heart. *J Cardiovasc Pharmacol* 34:82–88
- Drossman DA (2006) The functional gastrointestinal disorders and the Rome III process. *Gastroenterology* 130:1377–1390
- Dunlop SP, Coleman NS, Blackshaw E, Perkins AC, Singh G, Marsden CA, Spiller RC (2005) Abnormalities of 5-hydroxytryptamine metabolism in irritable bowel syndrome. *Clin Gastroenterol Hepatol* 3:349–357
- Ellis ES, Byrne C, Murphy OE, Tilford NS, Baxter GS (1995) Mediation by 5-hydroxytryptamine_{2B} receptors of endothelium-dependent relaxation in rat jugular vein. *Br J Pharmacol* 114:400–404
- Emmanuel AV, Kamm MA, Roy AJ, Antonelli K (1998) Effect of a novel prokinetic drug, R093877, on gastrointestinal transit in healthy volunteers. *Gut* 42:511–516
- Emmanuel AV, Roy AJ, Nicholls TJ, Kamm MA (2002) Prucalopride, a systemic enterokinetic, for the treatment of constipation. *Aliment Pharmacol Ther* 16:1347–1356
- Engel L, Kobel B, Ontsouka EC, Graber HU, Blum JW, Steiner A, Meylan M (2006) Distribution of mRNA coding for 5-hydroxytryptamine receptor subtypes in the intestines of healthy dairy cows and dairy cows with cecal dilatation–dislocation. *Am J Vet Res* 67:95–101
- Erspamer V, Vialli M (1937) Ricerche sul secreto delle cellule enterochromaffini. *Boll Soc Med Chir Pavia* 51:357–363
- Evangelista S (2007) Drug evaluation: Pumosetrag for the treatment of irritable bowel syndrome and gastroesophageal reflux disease. *Curr Opin Invest Drugs* 8:416–422
- Evans BW, Clark WK, Moore DJ, Whorwell PJ (2004) Tegaserod for the treatment of irritable bowel syndrome. *Cochrane Database Syst Rev* 1:CD003960
- Fiorica-Howells E, Maroteaux L, Gershon MD (2000) Serotonin and the 5-HT_{2B} receptor in the development of enteric neurones. *J Neurosci* 20:294–305
- Fiorica-Howells E, Hen R, Gingrich J, Li Z, Gershon MD (2002) 5-HT_{2A} receptors: location and functional analysis in intestines of wild-type and 5-HT_{2A} knockout mice. *Am J Physiol* 282:G877–G893
- Foguet M, Hoyer D, Pardo LA, Parekh A, Kluxen FW, Kalkman HO, Stühmer W, Lübbert H (1992) Cloning and functional characterization of the rat stomach fundus serotonin receptor. *EMBO J* 11:3481–3487
- Foxx-Orenstein AE, Kuemmerle JF, Grider JR (1996) Distinct 5-HT receptors mediate the peristaltic reflex induced by mucosal stimuli in human and guinea pig intestine. *Gastroenterology* 111:1281–1290
- Freeman AJ, Cunningham KT, Tyers MB (1992) Selectivity of 5-HT₃ receptor antagonists and anti-emetic mechanisms of action. *Anti-Cancer Drugs* 3:79–85
- Friedenberg FK, Parkman HP (2006) Delayed gastric emptying: whom to test, how to test, and what to do. *Curr Treat Options Gastroenterol* 9:295–304
- Fujita T, Yokota S, Sawada M, Majima M, Ohtani Y, Kumagai Y (2005) Effect of MKC-733, a 5-HT receptor partial agonist, on bowel motility and symptoms in subjects with constipation: an exploratory study. *J Clin Pharm Ther* 30:611–622
- Gale JD, Grossman CJ, Whitehead JW, Oxford AW, Bunce KT, Humphrey PP (1994) GR113808: a novel, selective antagonist with high affinity at the 5-HT₄ receptor. *Br J Pharmacol* 111:332–338
- Galligan JJ (1996) Electrophysiological studies of 5-hydroxytryptamine receptors on enteric neurons. *Behav Brain Res* 73:199–201
- Galligan JJ (2002) Ligand-gated ion channels in the enteric nervous system. *Neurogastroenterol Motil* 14:611–623
- Galligan JJ, LePard KJ, Schneider DA, Zhou X (2000) Multiple mechanisms of fast excitatory synaptic transmission in the enteric nervous system. *J Auton Nerv Syst* 81:97–103
- Galligan JJ, Pan H, Messori E (2003) Signalling mechanism coupled to 5-hydroxytryptamine₄ receptor-mediated facilitation of fast synaptic transmission in the guinea pig ileum myenteric plexus. *Neurogastroenterol Motil* 15:523–529
- Gallo-Torres H, Brinker A, Avigan M (2006) Alosetron: ischemic colitis and serious complications of constipation. *Am J Gastroenterol* 101:1080–1083
- Galmiche JP, Clouse RE, Bálint A, Cook I, Kahrilas PJ, Paterson WG, Smout AJPM (2006) Functional esophageal disorders. *Gastroenterol* 130:1459–1465
- Gerald C, Adham N, Kao HT, Olsen MA, Laz TM, Schechter LE, Bard JA, Vaysse PJJ, Hartig PR (1995) The 5-HT₄ receptor:

- molecular cloning and pharmacological characterization of two splice variants. *EMBO J* 14:2806–2815
- Gershon MD (2000) 5-HT (serotonin) physiology and related drugs. *Curr Opin Gastroenterol* 16:113–120
- Gershon MD (2004) Review article: serotonin receptors and transporters-roles in normal and abnormal gastrointestinal motility. *Aliment Pharmacol Ther* 20(Suppl 7):3–14
- Gershon MD (2005) Nerves, reflexes, and the enteric nervous system. *J Clin Gastroenterol* 39:S184–S193
- Gershon MD, Tack J (2007) The serotonin signaling system: from basic understanding to drug development for functional GI disorders. *Gastroenterol* 132:397–414
- Gerson L (2007) Comment. *Gastroenterology* 133:721–722
- Glatzle J, Sternini C, Robin C, Zittel TT, Wong H, Reeve JR, Raybould HE (2002) Expression of 5-HT₃ receptors in the rat gastrointestinal tract. *Gastroenterology* 123:217–226
- Goldberg MR, Wong SL, Ganju J, Li YP, Ballow CH, Kitt MM (2007a) TD-5108, a selective 5-HT₄ agonist with high intrinsic activity, shows immediate and sustained prokinetic activity in healthy subjects. Abstract 318363, *Digestive Disease Week*, Washington, DC, May 19–24
- Goldberg M, Li YP, Garofalo B, Valmonte A, Kitt M (2007b) TD-5108, a selective 5-HT₄ agonist with high intrinsic activity, increases bowel movement frequency and provides significant overall relief in patients with chronic constipation. In: American College of Gastroenterology Annual Scientific Meeting and Postgraduate Course, Philadelphia PA, October 12–17
- Gorard DA, Libby GW, Farthing MJ (1994) 5-Hydroxytryptamine and human small intestinal motility: effect of inhibiting 5-hydroxytryptamine reuptake. *Gut* 35:496–500
- Gore S, Gilmore IT, Haigh CG, Brownless SM, Stockdale H, Morris AI (1990) Colonic transit in man is slowed by ondansetron (GR38032F), a selective 5-hydroxytryptamine receptor (type 3) antagonist. *Aliment Pharmacol Ther* 4:139–144
- Greenwood-Van Meerveld B, Campbell-Dittmeyer K, Johnson AC, Hicks GA (2006a) 5-HT_{2B} receptors do not modulate sensitivity to colonic distension in rats with acute colorectal hypersensitivity. *Neurogastroenterol Motil* 18:343–345
- Greenwood-Van Meerveld B, Venkova K, Hicks G, Dennis E, Crowell MD (2006b) Activation of peripheral 5-HT receptors attenuates colonic sensitivity to intraluminal distension. *Neurogastroenterol Motil* 18:76–86
- Grider JR (2003a) Reciprocal activity of longitudinal and circular muscle during intestinal peristaltic reflex. *Am J Physiol Gastrointest Liver Physiol* 284:G768–G775
- Grider JR (2003b) Neurotransmitters mediating the intestinal peristaltic reflex in the mouse. *J Pharmacol Exp Ther* 307:460–467
- Grider JR, Makhlof GM (1990) Regulation of the peristaltic reflex by peptides of the myenteric plexus. *Arch Int Pharmacodyn Ther* 303:232–251
- Grønstad KO, DeMagistris L, Dahlstrom A, Nilsson O, Price B, Zinner MJ, Jaffe BM, Ahlman H (1985) The effects of vagal nerve stimulation on endoluminal release of serotonin and substance P into the feline small intestine. *Scand J Gastroenterol* 20:163–169
- Haga K, Asano K, Fukuda T, Kobayakawa T (1995) The function of 5-HT₃ receptors on colonic transit in rats. *Obesity Res* 3(Suppl 5):801S–810S
- Hamblin MW, Metcalf MA, McGuffin RW, Karpells S (1992) Molecular cloning and functional characterization of a human 5-HT_{1B} serotonin receptor: a homologue of the rat 5-HT_{1B} receptor with 5-HT_{1D}-like pharmacological specificity. *Biochem Biophys Res Commun* 184:752–759
- Hansen MB, Skadhauge E (1997) Signal transduction pathways for serotonin as an intestinal secretagogue. *Comp Biochem Physiol A Physiol* 118:283–290
- Hartig PR, Hoyer D, Humphrey PP, Martin GR (1996) Alignment of receptor nomenclature with the human genome classification of 5-HT_{1B} and 5-HT_{1D} receptor subtypes. *Trends Pharmacol Sci* 17:1003–1005
- Hawthorn J, Ostler KJ, Andrews PL (1988) The role of the abdominal visceral innervation and 5-hydroxytryptamine M-receptors in vomiting induced by the cytotoxic drugs cyclophosphamide and cis-platin in the ferret. *Q J Exp Physiol* 73:7–21
- Hedlund PB, Danielson PE, Thomas EA, Slanina K, Carson MJ, Sutcliffe JG (2003) No hypothermic response to serotonin in 5-HT₇ receptor knockout mice. *Proc Natl Acad Sci USA* 100:1375–1380
- Hegde SS, Moy TM, Perry MR, Loeb M, Eglen RM (1994) Evidence for the involvement of 5-hydroxytryptamine 4 receptors in 5-hydroxytryptophan-induced diarrhea in mice. *J Pharmacol Exp Ther* 271:741–747
- Hesketh PJ (2004) Understanding the pathobiology of chemotherapy-induced nausea and vomiting. Providing a basis for therapeutic progress. *Oncology* 18(Suppl 6):9–14
- Hicks GA, Clayton NM, Gaskin PJ, Kirkup AJ, Su X, Joshi S, Friedrich A, Conner HE, Cox B, Grundy D, Gebhart GF, Humphrey PPA (2001) 5-HT₄ receptor agonists stimulate small intestinal transit but do not have direct visceral antinociceptive effects in the rat. *Gastroenterology* 120(Suppl 1):A–6
- Hofmann C, Penner U, Dorow R, Pertz HH, Jahnichen S, Horowski R, Latte KP, Palla D, Schurad B (2002) Lisuride a dopamine receptor agonist with 5-HT_{2B} receptor antagonist properties: absence of cardiac valvulopathy adverse drug reaction reports supports the concept of a crucial role for 5-HT_{2B} receptor agonism in cardiac valvular fibrosis. *Clin Neuropharmacol* 29:80–86
- Horowski R, Jahnichen S, Pertz HH (2004) Fibrotic valvular heart disease is not related to chemical class but to biological function: 5-HT_{2B} receptor activation plays crucial role. *Mov Disord* 19:1523–1524
- Houghton LA, Jackson NA, Whorwell PJ, Cooper SM (1999) 5-HT₄ receptor antagonism in irritable bowel syndrome: effect of SB-207266-A on rectal sensitivity and small bowel transit. *Aliment Pharmacol Ther* 13:1437–1444
- Houghton LA, Foster JM, Whorwell PJ (2000) Alosetron, a 5-HT₃ receptor antagonist, delays colonic transit in patients with irritable bowel syndrome and healthy volunteers. *Aliment Pharmacol Ther* 14:775–782
- Houghton LA, Atkinson W, Whitaker RP, Whorwell PJ, Rimmer MJ (2003) Increased platelet depleted plasma 5-hydroxytryptamine concentration following meal ingestion in symptomatic female subjects with diarrhoea predominant irritable bowel syndrome. *Gut* 52:663–670
- Hoyer D, Clarke DE, Fozard JR, Hartig PR, Martin GR, Mylecharane EJ, Saxena PR, Humphrey PP (1994) International Union of Pharmacology classification of receptors for 5-hydroxytryptamine (serotonin). *Pharmacol Rev* 46:157–203
- Hoyer D, Hannon JP, Martin GR (2002) Molecular, pharmacological and functional diversity of 5-HT receptors. *Pharmacol Biochem Behav* 71:533–554
- Humphrey PP, Bountra C, Clayton N, Kozlowski K (1999) Review article: the therapeutic potential of 5-HT₃ receptor antagonists in the treatment of irritable bowel syndrome. *Aliment Pharmacol Ther* 13(Suppl 2):31–38
- Imada-Shirakata Y, Kotera T, Ueda S, Okuma M (1997) Serotonin activates electrolyte transport via 5-HT_{2A} receptor in rat colonic crypt cells. *Biochem Biophys Res Commun* 230:437–441
- Irving HR, Tan YY, Tochon-Danguy N, Liu H, Chetty N, Desmond PV, Pouton CW, Coupar IM (2007) Comparison of 5-HT₄ and 5-HT₇ receptor expression and function in the circular muscle of the human colon. *Life Sci* 80:1198–1205

- Ishida T, Kawashima S, Hirata K, Yokoyama M (1998) Nitric oxide is produced via 5-HT_{1B} and 5-HT_{2B} receptor activation in human coronary artery endothelial cells. *Kobe J Med Sci* 44:51–63
- Ismair MG, Kullak-Ublick GA, Blakely RD, Fried M, Vavricka SR (2007) Tegaserod inhibits the serotonin transporter SERT. *Digestion* 75:90–95
- Jaffré F, Callebert J, Sarre A, Etienne N, Nebigil CG, Launay JM, Maroteaux L, Monassier L (2004) Involvement of the serotonin 5-HT_{2B} receptor in cardiac hypertrophy linked to sympathetic stimulation. *Circulation* 110:969–974
- James AN, Ryan JP, Parkman HP (2005) Effects of the selective serotonin reuptake inhibitor, fluoxetine, on regional gastric contractility. *Neurogastroenterol Motil* 17:76–82
- Janssen P, Prins NH, Moreaux B, Meulemans AL, Lefebvre RA (2003) In vivo characterization of 5-HT_{1A} receptor-mediated gastric relaxation in conscious dogs. *Br J Pharmacol* 140:913–920
- Janssen P, Tack J, Sifrim D, Meulemans AL, Lefebvre RA (2004) Influence of 5-HT₁ receptor agonists on feline stomach relaxation. *Eur J Pharmacol* 492:259–267
- Jasper JR, Kosaka A, To ZP, Chang DJ, Eglen RM (1997) Cloning, expression and pharmacology of a truncated splice variant of the human 5-HT₇ receptor (h5-HT_{7b}). *Br J Pharmacol* 122:126–132
- Jensen GM, Gronndahl ML, Nielsen CG, Skadhauge E, Olsen JE, Hansen MB (1997) Effect of ondansetron on Salmonella typhimurium-induced net fluid accumulation in the pig jejunum in vivo. *Comp Biochem Physiol* 118:297–299
- Jiang W, Kreis ME, Eastwood C, Kirkup AJ, Humphrey PP, Grundy D (2000) 5-HT₍₃₎ and histamine H(1) receptors mediate afferent nerve sensitivity to intestinal anaphylaxis in rats. *Gastroenterol* 119:1267–1275
- Jin H, Oksenberg D, Ashkenazi A, Peroutka SJ, Duncan AMV, Rozmahel R, Yang Y, Mengod G, Palacios JM, O'Dowd BF (1992) Characterization of the human 5-hydroxytryptamine_{1B} receptor. *J Biol Chem* 267:5735–5738
- Jin JG, Foxx-Orenstein AE, Grider JR (1999) Propulsion in guinea pig colon induced by 5-hydroxytryptamine (HT) via 5-HT₄ and 5-HT₃ receptors. *J Pharmacol Exp Ther* 288:93–97
- Johanson JF (2004) Review article: tegaserod for chronic constipation. *Aliment Pharmacol Ther* 20(Suppl 7):20–24
- Jones BJ, Blackburn TP (2002) The medical benefit of 5-HT research. *Pharmacol Biochem Behav* 71:555–568
- Jordan D (2005) Vagal control of the heart: central serotonergic (5-HT) mechanisms. *Exp Physiol* 90:175–181
- Kahrilas PJ, Quigley EMM, Castell DO, Spechler SJ (2000) The effects of tegaserod (HTF 919) on oesophageal acid exposure in gastro-oesophageal reflux disease. *Aliment Pharmacol Ther* 14:1503–1509
- Kajita S, Ito C, Kawamura R, Yasuda S, Isobe Y, Fukushima K (2001) Pharmacological characterization of a novel 5-HT₄ receptor agonist, TS-951, in vitro. *Pharmacology* 63:8–16
- Kamm MA (2002) Review article: the complexity of drug development for irritable bowel syndrome. *Aliment Pharmacol Ther* 16:343–351
- Kaumann AJ, Levy FO (2006) 5-hydroxytryptamine receptors in the human cardiovascular system. *Pharmacol Ther* 111:674–706
- Kellum JM, Budhoo MR, Siriwardena AK, Smith EP, Jebraili SA (1994) Serotonin induces Cl⁻ secretion in human jejunal mucosa in vitro via a nonneural pathway at a 5-HT₄ receptor. *Am J Physiol* 267:G357–G363
- Kilkens TOC, Honig A, Van Niewenhoven MA, Riedel WJ, Brummer RJM (2004) Acute tryptophan depletion affects brain–gut responses in irritable bowel syndrome patients and controls. *Gut* 53:1794–1800
- Kim DY, Camilleri M (2000) Serotonin: a mediator of the brain–gut connection. *Am J Gastroenterol* 95:2698–2709
- Kirchgessner AL, Liu MT, Howard MJ, Gershon MD (1993) Detection of the 5-HT_{1A} receptor and 5-HT_{1A} receptor mRNA in the rat bowel and pancreas: comparison with 5-HT_{1P} receptors. *J Comp Neurol* 327:233–250
- Kirchgessner AL, Liu MT, Raymond JR, Gershon MD (1996) Identification of cells that express 5-hydroxytryptamine 1A receptors in the nervous systems of the bowel and pancreas. *J Comp Neurol* 364:439–455
- Kohen R, Metcalf MA, Khan N, Druck T, Huebner K, Lachowicz JE, Meltzer HY, Sibley DR, Roth BL, Hamblin MW (1996) Cloning, characterization and chromosomal localization of a human 5-HT₆ serotonin receptor. *J Neurochem* 66:47–56
- Komada T, Yano S (2007) Pharmacological characterization of 5-Hydroxytryptamine-receptor subtypes in circular muscle from the rat stomach. *Biol Pharm Bull* 30:508–513
- Kozłowski CM, Green A, Grundy D, Boissonade FM, Bountra C (2000) The 5-HT₍₃₎ receptor antagonist alosetron inhibits the colorectal distension induced depressor response and spinal c-fos expression in the anaesthetised rat. *Gut* 46:474–480
- Krobert KA, Bach T, Syversveen T, Kvingedal AM, Levy FO (2001) The cloned human 5-HT₇ receptor splice variants: a comparative characterization of their pharmacology, function and distribution. *Naunyn-Schmiedeberg's Arch Pharmacol* 363:620–632
- Kuemmerle JF, Murthy KS, Grider JR, Martin DC, Makhlof GM (1995) Coexpression of 5-HT_{2A} and 5-HT₄ receptors coupled to distinct signaling pathways in human intestinal muscle cells. *Gastroenterology* 109:1791–1800
- Kuiken SD, Tytgat GN, Boeckxstaens GE (2003) The selective serotonin reuptake inhibitor fluoxetine does not change rectal sensitivity and symptoms in patients with irritable bowel syndrome: a double blind, randomized, placebo-controlled study. *Clin Gastroenterol Hepatol* 1:219–228
- Kuo B, Camilleri M, Burton D, Viramontes B, McKinzie S, Thomforde G, O'Connor MK, Brinkmann BH (2002) Effects of 5-HT₃ antagonism on postprandial gastric volume and symptoms in humans. *Aliment Pharmacol Ther* 16:225–233
- Kursar JD, Nelson DL, Wainscott DB, Baez M (1994) Molecular cloning, functional expression, and mRNA tissue distribution of the human 5-hydroxytryptamine_{2B} receptor. *Mol Pharmacol* 46:227–234
- Ladabaum U, Glidden D (2002) Effect of the selective serotonin reuptake inhibitor sertraline on gastric sensitivity and compliance in healthy humans. *Neurogastroenterol Motil* 14:395–402
- Lea R, Houghton LA, Whorwell PJ, Whitaker RP (2002) Evidence for increased plasma 5-hydroxytryptamine (5-HT) concentration following meal ingestion in patients with functional dyspepsia (FD) and its relationship to gender and symptoms. *Gastroenterology* 122:304
- Leclere PG, Prins NH, Schuurkes JA, Lefebvre RA (2005) 5-HT₄ receptors located on cholinergic nerves in human colon circular muscle. *Neurogastroenterol Motil* 17:366–375
- Leysen JE (2004) 5-HT₍₂₎ receptors. *Curr Drug Target CNS Neurol Disord* 3:11–26
- Liu M, Gershon MD (2005b) Slow excitatory ('5-HT_{1P}'-like) responses of mouse myenteric neurons to 5-HT: mediation by heterodimers of 5-HT_{1B/1D} and Drd₂ receptors. *Gastroenterology* 128(4 Suppl 2):A87
- Liu M, Geddis MS, Wen Y, Setlik W, Gershon MD (2005a) Expression and function of 5-HT₄ receptors in the mouse enteric nervous system. *Am J Physiol* 289:G1148–G1163
- Longstreth GF, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC (2006) Functional bowel disorders. *Gastroenterology* 130:1480–1491
- Mackie AD, Ferrington C, Cowan S, Merrick MV, Baird JD, Palmer KR (1991) The effect of renzapride, a novel prokinetic agent, in diabetic gastroparesis. *Aliment Pharmacol Ther* 5:135–142

- Maricq AV, Peterson AS, Brake AJ, Myers RM, Julius D (1991) Primary structure and functional expression of the 5HT₃ receptor, a serotonin-gated ion channel. *Science* 254:432–437
- Martel F (2006) Recent advances on the importance of the serotonin transporter SERT in the rat intestine. *Pharmacol Res* 54:73–76
- Martel F, Monteiro R, Lemos C (2003) Uptake of serotonin at the apical and basolateral membranes of human intestinal epithelial (Caco-2) cells occurs through the neuronal serotonin transporter (SERT). *J Pharmacol Exp Ther* 306:355–362
- Mawe GM, Branchek TA, Gershon MD (1986) Peripheral neural serotonin receptors: Identification and characterization with specific antagonists and agonists. *Proc Natl Acad Sci USA* 83:9799–9803
- Mawe GM, Coates MD, Moses PL (2006) Review article: intestinal serotonin signalling in irritable bowel syndrome. *Aliment Pharmacol Ther* 23:1067–1076
- Mayer EA, Bradesi S (2003) Alosetron and irritable bowel syndrome. *Exp Opin Pharmacother* 4:2089–2098
- Mayer EA, Berman S, Derbyshire SWG, Suyenobu B, Chang L, Fitzgerald L, Manelkern M, Hamm L, Vogt B, Naliboff BD (2002) The effect of the 5-HT₃ receptor antagonist, alosetron, on brain responses to visceral stimulation in irritable bowel syndrome patients. *Aliment Pharmacol Ther* 16:1357–1366
- McAllister G, Charlesworth A, Snodin C, Beer MS, Noble AJ, Middlemiss DN, Iversen LL, Whiting P (1992) Molecular cloning of a serotonin receptor from human brain (5HT_{1E}): a fifth 5HT₁-like subtype. *Proc Natl Acad Sci USA* 89:5517–5521
- McNulty R (2007) Are all 5-HT₃ receptor antagonists the same? *J Natl Comp Cancer Netw* 5:35–43
- Medhurst AD, Lezoualch F, Fischmeister R, Middlemiss DN, Sanger GJ (2001) Quantitative mRNA analysis of five C-terminal splice variants of the human 5-HT₄ receptor in the central nervous system by TaqMan real time RT-PCR. *Brain Res Mol Brain Res* 90:125–134
- Mertz H, Fass R, Kodner A, Yan-Go F, Fullerton S, Mayer EA (1998) Effect of amitriptyline on symptoms, sleep, and visceral perception in patients with functional dyspepsia. *Am J Gastroenterol* 93:160–165
- Meyers NL, Hickling RI (2007) The cardiovascular safety profile of renzapride, a novel treatment for irritable bowel syndrome. *J Int Med Res* 35:848–866
- Mialet J, Fischmeister R, Lezoualch F (2003) Characterization of human 5-HT_{4(d)} receptor desensitization in CHO cells. *Br J Pharmacol* 138:445–452
- Mir AK, Hibert M, Tricklebank MD, Middlemiss DN, Kidd EJ, Fozard JR (1988) MDL 72832: a potent and stereoselective ligand at central and peripheral 5-HT_{1A} receptors. *Eur J Pharmacol* 149:107–120
- Mitchell ES, Neumaier JF (2005) 5-HT₆ receptors: a novel target for cognitive enhancement. *Pharmacol Ther* 108:320–333
- Monro RL, Bornstein JC, Bertrand PP (2005) Slow excitatory post-synaptic potentials in myenteric AH neurons of the guinea pig ileum are reduced by the 5-hydroxytryptamine(7) receptor antagonist SB 269970. *Neuroscience* 134:975–986
- Mori T, Kawano K, Shishikura T (2004) 5-HT₃-receptor antagonist inhibits visceral pain differently in chemical and mechanical stimuli in rats. *J Pharmacol Sci* 94:73–76
- Morteau O, Julia V, Eeckhout C, Bueno L (1994) Influence of 5-HT₃ receptor antagonists in visceromotor and nociceptive responses to rectal distension before and during experimental colitis in rats. *Fundam Clin Pharmacol* 8:553–562
- Muller-Lissner SA (1987) Treatment of chronic constipation with cisapride and placebo. *Gut* 28:1033–1038
- Muller-Lissner SA, Fumagalli I, Bardhan KD, Pace F, Pecher E, Nault B, Ruegg P (2001) Tegaserod, a 5-HT₄ receptor partial agonist, relieves symptoms in irritable bowel syndrome patients with abdominal pain, bloating and constipation. *Aliment Pharmacol Ther* 15:1655–1666
- Nagakura Y, Kamato T, Nishida A, Ito H, Yamano M, Miyata K (1996a) Characterization of 5-hydroxytryptamine (5-HT) receptor subtypes influencing colonic motility in conscious dogs. *Naunyn-Schmiedeberg's Arch Pharmacol* 353:489–498
- Nagakura Y, Naitoh Y, Kamato T, Yamano M, Miyata K (1996b) Compounds possessing 5-HT₃ receptor antagonistic activity inhibit intestinal propulsion in mice. *Eur J Pharmacol* 311:67–72
- Navari RM (2006) Palonosetron: a second-generation 5-hydroxytryptamine receptor antagonist. *Future Oncol* 2:591–602
- Nebigil CG, Maroteaux L (2003) Functional consequences of serotonin/5-HT_{2B} receptor signaling in heart. *Circulation* 108:902–908
- Nebigil CG, Launay JM, Hickel P, Tournois C, Maroteaux L (2000) 5-Hydroxytryptamine 2B receptor regulates cell-cycle progression: cross-talk with tyrosine kinase pathways. *Proc Natl Acad Sci USA* 97:2591–2596
- Nelson D (2004) 5-HT₃ receptors. *Curr Drug Targets* 3:53–58
- Nicholson R, Small J, Dixon AK, Spanswick D, Lee K (2003) Serotonin receptor mRNA in dorsal root ganglion neurons. *Neurosci Lett* 337:119–122
- Niesler B, Frank B, Kapeller J, Rappold GA (2003) Cloning, physical mapping and expression analysis of the human 5-HT₃ serotonin receptor-like genes HTR3C, HTR3D and HTR3E. *Gene* 310:101–111
- Nilsson T, Longmore J, Shaw D, Pantev E, Bard JA, Branchek T, Edvinsson L (1999) Characterisation of 5-HT receptors in human coronary arteries by molecular and pharmacological techniques. *Eur J Pharmacol* 372:49–56
- Ning Y, Zhu JX, Chan HC (2004) Regulation of ion transport by 5-hydroxytryptamine in rat colon. *Clin Exp Pharmacol Physiol* 31:424–428
- Pan H, Galligan JJ (1994) 5-HT_{1A} and 5-HT₄ receptors mediate inhibition and facilitation of fast synaptic transmission in enteric neurons. *Am J Physiol* 266:G230–G238
- Pan H, Gershon MD (2000) Activation of intrinsic afferent pathways in submucosal ganglia of the guinea pig small intestine. *J Neurosci* 20:3295–3309
- Parkman HP, Hasler WL, Fisher RS (2004) American gastroenterological association technical review on the diagnosis and treatment of gastroparesis. *Gastroenterology* 127:1592–1622
- Pascual D, Alsasua A, Goicoechea C, Martin MI (2002) The involvement of 5-HT₃ and 5-HT₄ receptors in two models of gastrointestinal transit in mice. *Neurosci Lett* 326:163–166
- Pasricha PJ (2007) Desperately seeking serotonin...a commentary on the withdrawal of tegaserod and the state of drug development for functional and motility disorders. *Gastroenterology* 132:2287–2290
- Pierce PA, Xie GX, Levine JD, Peroutka SJ (1996) 5-Hydroxytryptamine receptor subtype messenger RNAs in rat peripheral sensory and sympathetic ganglia: a polymerase chain reaction study. *Neuroscience* 70:553–559
- Pierce PA, Xie GX, Meuser T, Peroutka SJ (1997) 5-Hydroxytryptamine receptor subtype messenger RNAs in human dorsal root ganglia: a polymerase chain reaction study. *Neuroscience* 81:813–819
- Pindon A, Van Hecke G, Van Gompel P, Lesage AS, Leysen JE, Jurzak M (2002) Differences in signal transduction of two 5-HT₄ receptor splice variants: compound specificity and dual coupling with Galphas- and Galphai/o-proteins. *Mol Pharmacol* 61:85–96
- Poen AC, Felt-Bersma RJF, Van Dongen PAM, Meuwissen SGM (1999) Effect of prucalopride, a new enterokinetic agent, on gastrointestinal transit and anorectal function in healthy volunteers. *Aliment Pharmacol Ther* 13:1493–1497
- Ponimaskin EG, Heine M, Joubert L, Sebben M, Bickmeyer U, Richter DW, Dumuis A (2002) The 5-hydroxytryptamine(4a)

- receptor is palmitoylated at two different sites, and acylation is critically involved in regulation of receptor constitutive activity. *J Biol Chem* 277:2534–2546
- Poole DP, Xu B, Koh SL, Hunne B, Coupar IM, Irving HR, Shinjo K, Furness JB (2006) Identification of neurons that express 5-hydroxytryptamine₄ receptors in intestine. *Cell Tissue Res* 325:413–422
- Prather CM, Camilleri M, Zinsmeister AR, McKinzie S, Thomforde G (2000) Tegaserod accelerates orocecal transit in patients with constipation-predominant irritable bowel syndrome. *Gastroenterology* 118:463–468
- Prins NH, Briejer MR, Schuurkes JA (1997) Characterization of the contraction to 5-HT in the canine colon longitudinal muscle. *Br J Pharmacol* 120:714–720
- Prins NH, Briejer MR, Van Bergen PJ, Akkermans LM, Schuurkes JA (1999a) Evidence for 5-HT₇ receptors mediating relaxation of human colonic circular smooth muscle. *Br J Pharmacol* 128:849–852
- Prins NH, Van Haselen JF, Lefebvre RA, Briejer MR, Akkermans LM, Schuurkes JA (1999b) Pharmacological characterization of 5-HT₄ receptors mediating relaxation of canine isolated rectum circular smooth muscle. *Br J Pharmacol* 127:1431–1437
- Ramamoorthy S, Bauman AL, Moore KR, Han H, Yang-Feng T, Chang AS, Ganapathy V, Blakely RD (1993) Antidepressant- and cocaine-sensitive human serotonin transporter: molecular cloning, expression, and chromosomal localization. *Proc Natl Acad Sci USA* 90:2542–2546
- Raybould HE, Glatzle J, Robin C, Meyer JH, Phan T, Wong H, Sternini C (2003) Expression of 5-HT₃ receptors by extrinsic duodenal afferents contribute to intestinal inhibition of gastric emptying. *Am J Physiol* 284:G367–G372
- Read NM, Gwee KA (1994) The importance of 5-hydroxytryptamine receptors in the gut. *Pharmacol Ther* 62:159–173
- Reeves JJ, Bunce KT, Humphrey PP (1991) Investigation into the 5-hydroxytryptamine receptor mediating smooth muscle relaxation in the rat oesophagus. *Br J Pharmacol* 103:1067–1072
- Rizzi CA, Sagrada A, Schiavone A, Schiantarelli P, Cesana R, Schiavi GB, Ladinsky H, Donetti A (1994) Gastroprokinetic properties of the benzimidazolone derivative BIMU 1, an agonist at 5-hydroxytryptamine₄ and antagonist at 5-hydroxytryptamine₃ receptors. *Naunyn-Schmiedeberg's Arch Pharmacol* 349:338–345
- Rodriguez-Stanley S, Zubaidi S, Proskin HM, Kralstein JR, Shetzline MA, Miner PB, Philip B (2006) Effect of tegaserod on esophageal pain threshold, regurgitation, and symptom relief in patients with functional heartburn and mechanical sensitivity. *Clin Gastroenterol Hepatol* 4:442–450
- Roila F, Fatigoni S (2006) New antiemetic drugs. *Ann Oncol* 17(Suppl 2):96–100
- Ruat M, Traiffort E, Arrang JM, Tardivellacombe J, Diaz J, Leurs R, Schwartz JC (1993) A novel rat serotonin (5-HT₆) receptor: molecular cloning, localization and stimulation of cAMP accumulation. *Biochem Biophys Res Commun* 193:268–276
- Rubenstein EB (2004) Palonosetron: a unique 5-HT₃ receptor antagonist indicated for the prevention of acute and delayed chemotherapy-induced nausea and vomiting. *Clin Adv Hematol Oncol* 2:284–288
- Saad RJ, Chey WD (2006) Review article: current and emerging therapies for functional dyspepsia. *Aliment Pharmacol Ther* 24:475–492
- Säfsten B, Sjöblom M, Flemström G (2006) Serotonin increases protective duodenal bicarbonate secretion via enteric ganglia and a 5-HT₄-dependent pathway. *Scand J Gastroenterol* 41:1279–1289
- Saltzman AG, Morse B, Whitman MM, Ivanshchenko Y, Jaye M, Felder S (1991) Cloning of the human serotonin 5-HT₂ and 5-HT_{1C} receptor subtypes. *Biochem Biophys Res Commun* 181:1469–1478
- Sanger GJ, Andrews PLR (2006) Treatment of nausea and vomiting: gaps in our knowledge. *Autonom Neurosci Basic Clin* 129:3–16
- Sanger GJ, Wardle KA (1994) Constipation evoked by 5-HT₃-receptor antagonism: evidence for heterogeneous efficacy among different antagonists in guinea pigs. *J Pharm Pharmacol* 46:666–670
- Sanger GJ, Banner SE, Smith MI, Wardle KA (1998) SB-207266: 5-HT₄ receptor antagonism in human isolated gut and prevention of 5-HT-evoked sensitization of peristalsis and increased defecation in animal models. *Neurogastroenterol Motil* 10:271–279
- Schoeffter P, Hoyer D (1990) 5-Hydroxytryptamine (5-HT)-induced endothelium-dependent relaxation of pig coronary arteries is mediated by 5-HT receptors similar to the 5-HT_{1D} receptor subtype. *J Pharmacol Exp Ther* 252:387–395
- Schworer H, Ramadori G (1998) Autoreceptors can modulate 5-hydroxytryptamine release from porcine and human small intestine in vitro. *Naunyn-Schmiedeberg's Arch Pharmacol* 357:548–552
- Shen Y, Monsma FJ, Metcalf MA, Jose PA, Hamblin MW, Sibley DR (1993) Molecular cloning and expression of a 5-hydroxytryptamine₇ serotonin receptor subtype. *J Biol Chem* 268:18200–18204
- Shufflebotham J, Hood S, Hendry J, Hince DA, Morris K, Nutt D, Probert C, Potokar J (2006) Acute tryptophan depletion alters gastrointestinal and anxiety symptoms in irritable bowel syndrome. *Am J Gastroenterol* 101:2582–2587
- Sifrim D, Holloway RH, Tack J, Zelter A, Missotten T, Coulie B, Janssens J (1999) Effect of sumatriptan, a 5-HT₁ agonist, on the frequency of transient lower esophageal sphincter relaxations and gastroesophageal reflux in healthy subjects. *Am J Gastroenterol* 94:3158–3164
- Sivarao DV, Newberry K, Lodge NJ (2004) Effect of the 5-HT_{1A} receptor partial agonist buspirone on colorectal distension-induced pseudoaffective and behavioral responses in the female Wistar rat. *Eur J Pharmacol* 494:23–29
- Sloots CE, Poen AC, Kerstens R, Stevens M, De Pauw M, Van Oene JC, Meuwissen SG, Felt-Bersma RJ (2002) Effects of prucalopride on colonic transit, anorectal function and bowel habits in patients with chronic constipation. *Aliment Pharmacol Ther* 16:759–767
- Smith JA, Beattie DT, Cuthbert AW, Ropenga A, Marquess D, Shaw J, Vickery R, Humphrey PP (2007) TD-5108, a selective, high intrinsic activity 5-HT₄ receptor agonist - in vitro profile at human recombinant 5-HT₄ receptor splice variant and human isolated colon. *Digestive Disease Week, Washington, DC, W1222*
- Spigset O (1999) Adverse reactions of selective reuptake inhibitors: reports from a spontaneous reporting system. *Drug Saf* 20:277–287
- Stacher G, Weber U, Stacher-Janotta G, Bauer P, Huber K, Holzappel A, Krause G, Steinborn C (2000) Effects of the 5-HT₃ antagonist cilansetron vs placebo on phasic sigmoid colonic motility in healthy man: a double-blind crossover trial. *Br J Clin Pharmacol* 49:429–436
- Sun YN, Luo JY (2004) Effects of tegaserod on Fos, substance P and calcitonin gene-related peptide expression induced by colon inflammation in lumbar sacral spinal cord. *World J Gastroenterol* 10:1830–1833
- Tack J, Janssens J, Vantrappen G, Wood JD (1992) Actions of 5-hydroxytryptamine on myenteric neurons in guinea pig gastric antrum. *Am J Physiol* 263:G838–G846
- Tack J, Coulie B, Wilmer A, Andrioli A, Janssens J (2000) Influence of sumatriptan on gastric fundus tone and on the perception of gastric distension in man. *Gut* 46:468–473
- Tack J, Bisschops R, DeMarchi B (2001) Causes and treatment of functional dyspepsia. *Curr Gastroenterol Rep* 3:503–508
- Tack J, Van Elzen B, Tytgat G (2004) A placebo-controlled trial of the 5-HT_{1A} agonist R-137696 on symptoms, visceral hypersensitivity and on impaired accommodation in functional dyspepsia. *Gastroenterology* 126:A70

- Tack J, Broekaert D, Corsetti M, Fischler B, Janssens J (2006a) Influence of acute serotonin reuptake inhibition on colonic sensorimotor function in man. *Aliment Pharmacol Ther* 23:265–274
- Tack J, Middleton SJ, Horne MC, Piessevaux H, Bloor JS, Meyers NL (2006b) Pilot study of the efficacy of renzapride on gastrointestinal motility and symptoms in patients with constipation-predominant irritable bowel syndrome. *Aliment Pharmacol Ther* 23:1655–1665
- Tack J, Talley NJ, Camilleri M, Holtmann G, Hu P, Malagelada J-R, Stanghellini V (2006c) Functional gastroduodenal disorders. *Gastroenterology* 130:1466–1479
- Tack J, Vanden Berghe P, Coulie B, Janssens J (2007) Sumatriptan is an agonist at 5-HT receptors on myenteric neurones in the guinea-pig gastric antrum. *Neurogastroenterol Motil* 19:39–46
- Takayanagi S, Hanai H, Kumagai J, Kaneko E (1995) Serotonin uptake and its modulation in rat jejunal enterocyte preparation. *J Pharmacol Exp Ther* 272:1151–1159
- Talley NJ (2003a) Update on the role of drug therapy in non-ulcer dyspepsia. *Rev Gastroenterol Disord* 3:25–30
- Talley NJ (2003b) Pharmacologic therapy for the irritable bowel syndrome. *Am J Gastroenterol* 98:750–758
- Talley NJ (2004) Antidepressants in IBS: are we deluding ourselves? *Am J Gastroenterol* 99:921–923
- Tamaoki S, Yamauchi Y, Nakano Y, Sakano S, Asagarasu A, Sato M (2007) Pharmacological properties of 3-amino-5,6,7,8-tetrahydro-2-[4-[4-(quinolin-2-yl)piperazin-1-yl]butyl]quinazolin-4(3H)-one (TZB-30878), a novel therapeutic agent for diarrhea-predominant irritable bowel syndrome (IBS) and its effects on an experimental IBS model. *J Pharmacol Exp Ther* 322:1315–1323
- Tam FS, Hillier K, Bunce KT (1994) Characterization of the 5-hydroxytryptamine receptor type involved in inhibition of spontaneous activity of human isolated colonic circular muscle. *Br J Pharmacol* 113:143–150
- Tamura T, Sano I, Satoh M, Mizumoto A, Itoh Z (1996) Pharmacological characterization of 5-hydroxytryptamine-induced motor activity (in vitro) in the guinea pig gastric antrum and corpus. *Eur J Pharmacol* 308:315–324
- Taniyama K, Nakayama S, Takeda K, Matsuyama S, Shirakawa J, Sano I, Tanaka C (1991) Cisapride stimulates motility of the intestine via the 5-hydroxytryptamine receptors. *J Pharmacol Exp Ther* 258:1098–1104
- Taniyama K, Makimoto N, Furuichi A, Sakurai-Yamashita Y, Nagase Y, Kaibara M, Kanematsu T (2000) Functions of peripheral 5-hydroxytryptamine receptors, especially 5-hydroxytryptamine₄ receptor, in gastrointestinal motility. *J Gastroenterol* 35:575–582
- Tonini M (2005) 5-Hydroxytryptamine effects in the gut: the 3, 4, and 7 receptors. *Neurogastroenterol Motil* 17:637–642
- Tonini M, De Ponti F, Di Nucci A, Crema F (1999) Review article: cardiac adverse effects of gastrointestinal prokinetics. *Aliment Pharmacol Ther* 13:1585–1591
- Tonini M, Vicini R, Cervio E, De Ponti F, De Giorgio R, Barbara G, Stanghellini V, Dellabianca A, Sterini C (2005) 5-HT₇ receptors modulate peristalsis and accommodation in the guinea pig ileum. *Gastroenterology* 129:1557–1566
- Tsou AP, Kosaka A, Bach C, Zuppan P, Yee C, Tom L, Alvarez R, Ramsey S, Bonhaus DW, Stefanich E, Jakeman L, Eglen RM, Chan HW (1994) Cloning and expression of a 5-hydroxytryptamine₇ receptor positively coupled to adenylyl cyclase. *J Neurochem* 63:456–464
- Tyers MB, Freeman AJ (2002) Mechanism of the anti-emetic activity of 5-HT₃ receptor antagonists. *Oncology* 49:263–268
- Tzvetkov MV, Meineke C, Oetjen E, Hirsch-Ernst K, Brockmoller J (2007) Tissue-specific alternative promoters of the serotonin receptor gene HTR3B in human brain and intestine. *Gene* 386:52–62
- Vane JR (1957) A sensitive method for the assay of 5-hydroxytryptamine. *Br J Pharmacol* 12:344–349
- Vanhoeacker P, Haegeman G, Leysen JE (2000) 5-HT₇ receptors: current knowledge and future prospect. *Trends Pharmacol Sci* 21:70–77
- Van Lelyveld N, Ter Linde J, Baron A, Mundt M, Wajs E, Samsom M (2006) The 5-HT₄ antagonist R216073 does not affect gastric motor and sensory function in patients with functional dyspepsia. *Aliment Pharmacol Ther* 24:669–677
- Van Lelyveld N, Ter Linde J, Schipper ME, Samsom M (2007) Regional differences in expression of TPH-1, SERT, 5-HT₍₃₎ and 5-HT₍₄₎ receptors in the human stomach and duodenum. *Neurogastroenterol Motil* 19:342–348
- Veldhuyzen van Zanten SJO, Jones MJ, Verlinden M, Talley NJ (2001) Efficacy of cisapride and domperidone in functional (nonulcer) dyspepsia: a meta-analysis. *Am J Gastroenterol* 96:689–696
- Vilaro MT, Domenech T, Palacios JM, Mengod G (2002) Cloning and characterization of a novel human 5-HT₄ receptor variant that lacks the alternatively spliced carboxy terminal exon. RT-PCR distribution in human brain and periphery of multiple 5-HT₄ receptor variants. *Neuropharmacology* 42:60–73
- Von Der Ohe MR, Camilleri M, Kvols LK, Thomforde GM (1993) Motor dysfunction of the small bowel and colon in patients with the carcinoid syndrome and diarrhea. *New Eng J Med* 329:1073–1078
- Wade PR, Chen J, Jaffe B, Kassem IS, Blakely RD, Gershon MD (1996) Localization and function of a 5-HT transporter in crypt epithelia of the gastrointestinal tract. *J Neurosci* 16:2352–2364
- Wardle KA, Sanger GJ (1993) The guinea pig distal colon—a sensitive preparation for the investigation of 5-HT₄ receptor-mediated contractions. *Br J Pharmacol* 110:1593–1599
- Wardle KA, Bingham S, Ellis ES, Gaster LM, Rushant B, Smith MI, Sanger GJ (1996) Selective and functional 5-hydroxytryptamine₄ receptor antagonism by SB 207266. *Br J Pharmacol* 118:665–670
- Weinshank RL, Zgombick JM, Macchi MJ, Branchek TA (1992) Human serotonin 1D receptor is encoded by a subfamily of two distinct genes: 5-HT_{1Da} and 5-HT_{1Db}. *Proc Natl Acad Sci USA* 89:3630–3634
- Woolley ML, Marsden CA, Fone KC (2004) 5-HT₆ receptors. *Current Drug Targets* 3:59–79
- Wouters MM, Farrugia G, Schemann M (2007a) 5-HT receptors on interstitial cells of Cajal, smooth muscle and enteric nerves. *Neurogastroenterol Motil* 19(Suppl 2):5–12
- Wouters MM, Gibbons SJ, Roeder JL, Distad M, Ou Y, Strege PR, Szurszewski JH, Farrugia G (2007b) Exogenous serotonin regulates proliferation of interstitial cells of Cajal in mouse jejunum through 5-HT_{2B} receptors. *Gastroenterology* 133:897–906
- Xue L, Camilleri M, Locke GR, Schuurkes JA, Meulemans A, Coulie BJ, Szurszewski JH, Farrugia G (2006) Serotonergic modulation of murine fundic tone. *Am J Physiol* 291:G1180–G1186
- Yeo A, Boyd P, Lumsden S, Saunders T, Handley A, Stubbins M, Knaggs A, Asquith S, Taylor I, Bahari B, Crocker N, Rallan R, Varsani S, Montgomery D, Alpers DH, Dukes GE, Purvis I, Hicks GA (2004) Association between a functional polymorphism in the serotonin transporter gene and diarrhoea predominant irritable bowel syndrome in women. *Gut* 53:1452–1458
- Young HM, Furness JB (1995) Ultrastructural examination of the targets of serotonin-immunoreactive descending reflexes to the circular muscle of guinea pig small intestine. *J Comp Neurol* 356:101–114
- Zhou S, Yung Chan S, Cher Goh B, Chan E, Duan W, Huang M, McLeod HL (2005) Mechanism-based inhibition of cytochrome P450 3A4 by therapeutic drugs. *Clin Pharmacokinet* 44:279–304
- Zuberi BF, Quraishy MS, Faisal N, Ahmed S (2005) Idiopathic gastroparesis. *J Coll Physicians Surgeons Pakistan* 15:566–577