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Characterization of phospholipase C activity at h5-HT_{2C} compared with h5-HT_{2B} receptors: influence of novel ligands upon membrane-bound levels of [³H]phosphatidylinositols

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Abstract Employing a novel, rapid and sensitive method for evaluation of phospholipase C (PLC) activity, the present study characterized the actions of diverse agonists and antagonists at human (h)5-HT_{2C} receptors expressed in Chinese Hamster Ovary (CHO) cells. In addition, affinities and efficacies at these sites were compared with those obtained at h5-HT_{2B} receptors.

5-HT elicited a robust and rapid reduction in levels of the pre-labelled, membrane-bound substrate of PLC, [³H]phosphatidylinositols ([³H]PI). The time-course of [³H]PI depletion paralleled that of [³H]inositol phosphate ([³H]IP) accumulation, as determined by conventional anion exchange chromatography. Inactivation of h5-HT_{2C} receptors with the alkylating agent, *N*-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ), revealed a large receptor reserve, with half-maximal PLC activation induced by a concentration of 5-HT occupying only 5% of sites. In analogy to 5-HT ($E_{max}=100\%$), DOI, MK212 and mCPP, as well as the novel ligands, Ro600332, Ro600175 and BW723C86, showed “full” efficacy at h5-HT_{2C} sites. Their efficacies were similar at h5-HT_{2B} sites, with the exception of mCPP and MK212, which acted as partial agonists. Further, lisuride and Ro600869 behaved as partial agonists and antagonists at h5-HT_{2C} and h5-HT_{2B} receptors, respectively. As concerns functional selectivity (potency for induction of [³H]PI depletion), only Ro600175 preferentially activated h5-HT_{2B} sites. In contrast, Ro600332 preferentially activated h5-HT_{2C} receptors. Amongst antagonists, RS102221 and SB242084 displayed a marked preference for h5-HT_{2C} sites, whereas LY266097, S33526 and SB204741 behaved as selective antagonists at h5-HT_{2B}

receptors. At both h5-HT_{2C} and h5-HT_{2B} receptors, antagonist potency (pK_b) and binding affinity (pK_i) were highly correlated.

In conclusion, this rapid and innovative method for determination of PLC activity permitted characterization of an extensive range of novel ligands at h5-HT_{2C} receptors. Although several antagonists clearly differentiated h5-HT_{2C} from h5-HT_{2B} receptors under these conditions, highly selective agonists remain to be identified.

Keywords 5-HT_{2C} receptors · 5-HT_{2B} receptors · Phospholipase C · Phosphatidylinositols · Receptor reserve

Introduction

5-HT_{2C} receptors are currently attracting considerable interest in view of their broad implication in the control of mood, motor behaviour and cognition, and their potential importance as targets for the improved treatment of anxiety, depression and other psychiatric and neurologic disorders (Stefanski and Goldberg 1997; Jenck et al. 1998a; Roth et al. 1998; Fox and Brotchie 1999; Meltzer 1999; Millan et al. 2000a). Of particular interest, the potent antagonist properties of the atypical antipsychotic, clozapine, at 5-HT_{2C} receptors (Canton et al. 1994; Millan et al. 1998; Cussac et al. 2000a) may contribute to its low extrapyramidal potential, superior control of negative symptoms and improvement of mood (Roth and Meltzer 1995; Meltzer 1999; Reavill et al. 1999). Further, blockade of 5-HT_{2C} receptors may participate in the therapeutic actions of several antidepressant agents, including nefazodone and mirtazapine (Millan et al. 2000a, 2000b). The role of closely related 5-HT_{2B} receptors remains unclear inasmuch as their density is low in the CNS (Foguet et al. 1992; Kursar et al. 1992, 1994; Bonhaus et al. 1995; Duxon et al. 1997a; Hoyer and Martin 1997). However, they may be involved, together with 5-HT_{2C} receptors, in the modulation of appetite (Kennett et al. 1997a). Further, activation of 5-HT_{2B} receptors may, in contrast to 5-HT_{2C} receptor, attenuate anxious states (Kennett et al. 1996a, 1998a;

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Duxon et al. 1997b), and, more recently, agonist properties at 5-HT_{2B} receptors have been associated with cardiopathy (Fitzgerald et al. 2000).

In light of the above, much effort has been devoted to identification of ligands differentiating 5-HT_{2C} from 5-HT_{2B} receptors. In the characterization of such agents, behavioural paradigms of actions at 5-HT_{2C} sites have been extensively exploited. However, in vivo models of actions at central 5-HT_{2B} receptors remain to be established. Thus, studies of 5-HT_{2C} and 5-HT_{2B} receptor-coupled signal transduction are of particular importance.

In this regard, the 5-HT₂ receptor family is coupled via a heterotrimeric GTP binding protein, G_q, to phospholipase C (PLC; Conn et al. 1986; Sanders-Bush and Canton 1995; Gerhardt and Heerikhuizen 1997). Activation of PLC elevates cytosolic levels of inositol phosphates, and subsequently increases levels of intracellular calcium, two major parameters exploited for characterization of drug efficacy at 5-HT₂ receptor subtypes (Conn et al. 1986; Porter et al. 1999; Cussac et al. 2000b; Jerman et al. 2001). Although calcium levels show rapid kinetics and yield important information, they are subject to modulation by a plethora of intracellular signals. Further, alterations of calcium levels may reflect actions at 5-HT_{2C} receptors mediated via G-proteins other than G_q (notably G_i; Lucaites et al. 1996; Alberts et al. 1999). This can complicate interpretation of drug actions since, in line with the concept of agonist-directed trafficking, 5-HT_{2C} receptor ligands may differentially modify coupling to distinct effectors (Berg et al. 1998, 1999; Alberts et al. 1999; Miller et al. 2000). Correspondingly, determination of [³H]IP levels offers a more direct and straightforward approach to evaluation of drug actions at 5-HT_{2C} receptors. In support of this contention, employing membrane-permeable peptides, 5-HT_{2C} receptors in choroid plexus were shown to couple to PLC via G_q independently of G_i (Chang et al. 2000).

Standard methodology for determination of [³H]IP levels is, however, cumbersome. We recently developed, therefore, a complementary approach quantifying membrane-bound [³H]phosphatidylinositols ([³H]PI), the substrate of PLC (Cussac et al. 2000a, 2000b). In the present study, we exploited the rapidity of this novel technique for characterization of the actions of an extensive series of "classical" and novel ligands at h5-HT_{2C} receptors in comparison with h5-HT_{2B} receptors.

In these studies, we exploited the VSV (edited) isoform of 5-HT_{2C} receptors which is highly expressed both in rat and in human brain (Burns et al. 1997; Herrick-Davis et al. 1999; Niswender et al. 1999; Wang et al. 2000). This isoform does not, in contrast to the non-edited (INI) isoform, exhibit constitutive activity (Burns et al. 1997; Herrick-Davis et al. 1999). In addition to mRNA editing, a further important factor influencing drug actions and, in particular, coupling efficacy at 5-HT_{2C} receptors, is receptor density (Lucaites et al. 1996; Burns et al. 1997; Berg et al. 1998, 1999; Herrick-Davis et al. 1999; Wang et al. 2000). Thus, we investigated the influence of the high expression level of h5-HT_{2C} receptors in this cell line by quantifying receptor reserve employing irreversible inactivation with

the alkylating agent, EEDQ (Atkinson and Minneman 1992; Adham et al. 1993; Cussac et al. 1999).

Materials and methods

Phospholipase C activity: [³H]PI depletion assay. CHO cells stably expressing ~3 pmol/mg of h5-HT_{2B} receptors and ~20 pmol/mg of edited h5-HT_{2C} receptors (VSV isoform), were obtained from Euroscreen (Brussels, Belgium) and grown in adherent culture in 225-cm² flasks with UltraCHO medium (BioWhittaker, Verviers, Belgium) containing sodium pyruvate (1 mM), dialysed fetal calf serum (0.1%) and geneticin (400 µg/ml). At confluence, cells were labelled with 2 µCi/ml of [³H]myo-inositol (10–20 Ci/mmol; Amersham, Les Ulis, France) for 24 h in serum-free UltraCHO medium. Adherent cells were rinsed twice in Krebs-LiCl buffer (NaH₂PO₄ 15.6 mM pH 7, NaCl 120 mM, KCl 4.8 mM, MgSO₄ 1.2 mM, CaCl₂ 1.2 mM, glucose 0.6% w/v, bovine serum albumin 0.04% w/v, LiCl 10 mM), scraped from the flask and washed once again by slow centrifugation. The cells were then resuspended in Krebs-LiCl and left to stand for 15 min at 37°C before use, or kept at –80°C in Krebs-LiCl/DMSO 10% until assay. The resulting suspension of cells loaded with [³H]myo-inositol is referred to as 'loaded cells'.

For determination of [³H]PI depletion, incubation of loaded cells was performed in 96-well plates (typically, one 225-cm² culture flask yielded sufficient cells for one 96-well plate containing 2.5×10⁵ cells per well) with agonist ligands at 37°C for 20 min (h5-HT_{2C} receptors) and for 30 min (h5-HT_{2B} receptors) in 0.4 ml final volume. Antagonists were preincubated with the cells for 15 min before addition of the agonist. Assays were stopped by addition of 0.4 ml of methanol/HCl (88 ml of 100% methanol + 12 ml of HCl 1 N) and cells were placed at –20°C for at least 2 h to facilitate cell lysis. The 96-well plates were then sonicated for 2 min and membranes recovered using a 'Filtermate' harvester (Packard, Meriden, Conn., USA) by filtration through GF/B filters impregnated with 0.1% v/v polyethyleneimine followed by three washes with distilled, deionized water. Radioactivity was determined using a 'Top-Count' microplate scintillation counter (Packard, Meriden, Conn., USA). Conventional determination of PLC activity was performed with the same loaded cell preparations in the same buffer and incubation conditions as above, and soluble total [³H]IP generation was determined using Dowex anion exchange chromatography as described by Dickenson and Hill (1996). Isotherms were analysed by non-linear regression, using the program 'PRISM' (Graphpad Software, San Diego, Calif., USA) to yield EC₅₀ and IC₅₀ (concentration of agonist and antagonist, respectively, that produced a half-maximal response) values. K_B values for inhibition of 5-HT (30 nM and 100 nM for h5-HT_{2B} and h5-HT_{2C}, respectively)-stimulated [³H]PI depletion, were calculated according to Lazareno and Birdsall (1993).

$K_B = IC_{50} / (1 + (Agonist/EC_{50}))$ where IC₅₀=Inhibitory Concentration₅₀ of the antagonist; Agonist=5-HT concentration; EC₅₀=Effective Concentration₅₀ of 5-HT alone.

Competition binding assays. Binding affinity at h5-HT_{2B} and h5-HT_{2C} receptors was determined essentially as described (Bonhaus et al. 1995) by competition binding with [³H]mesulergine (1 nM; Amersham, Les Ulis, France) in a buffer containing HEPES (20 mM) pH 7.7, EDTA (2 mM) and ascorbic acid (0.1% w/v). Incubations lasted 2 h at 22°C and non-specific binding was defined by 5-HT (10 µM). Isotherms were analysed by non-linear regression to yield IC₅₀ values. Inhibition constants (K_i values) were derived from IC₅₀ values according to Lazareno and Birdsall (1993).

5-HT_{2C} receptor alkylation with EEDQ. Intact CHO-h5-HT_{2C} cells were treated in Krebs-LiCl with EEDQ at a final concentration of 100 µM for different incubation times at 37°C followed by application of 5-HT and quantification of [³H]PI depletion as described above. For saturation experiments following EEDQ treatment, CHO-h5-HT_{2C} cells were washed twice by slow centrifugation in

an excess volume of Krebs-LiCl and [^3H]mesulergine binding performed on membrane preparation as described above. K_A values were determined by Furchgott analysis, as described by Atkinson and Minneman (1992) and Adham et al. (1993). Plots were derived of $1/[A]$ vs. $1/[A']$; where $[A]$ and $[A']$ are equiactive concentrations for stimulation of [^3H]PI depletion before and after receptor alkylation, respectively. K_A was calculated from $K_A = (\text{slope} - 1) / y\text{-intercept}$. Percentage receptor occupancy (O) was calculated by $O = 100 \times L / (L + K_A)$; where L is the concentration of agonist. The curve is fitted by a rectangular hyperbola. All data are expressed as means \pm SEM of ≥ 3 independent determinations performed in triplicate. Protein concentrations were determined colourimetrically using a bicinchonic acid assay kit (Sigma, Saint Quentin Fallavier, France).

Drugs. 5-HT was purchased from Sigma (Saint Quentin Fallavier, France). mCPP (1-(3-chlorophenyl)piperazine), DOI (1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane), lisuride, mesulergine and metergoline were purchased from Research Biochemicals International (Natick, Mass., USA), and RS102221 (*N*-[5-[5-(2,5-dioxo-spiro[imidazolidine-4,4'-piperidin]-1'-yl)pentanoyl]-2,4-dimethoxyphenyl]-4-(trifluoromethyl)benzenesulfonamide) from Tocris Cookson (Southampton, UK). Ro600175 (2(*S*)-1-(6-chloro-5-fluoro-1*H*-indol-1-yl)-2-propanamine fumarate), Ro600332 (1(*S*)-methyl-2-(4,4,7-trimethyl-1,4-dihydroindeno[1,2-*b*]pyrrol-1-yl)ethylamine fumarate), Ro600869 (*cis*-7-ethyl-6-hydroxy-8-methoxy-2-methyl-1,2,3,3a,4,9b-hexahydro-5*H*-benzo[*e*]isoindol-5-one), BW723C86 (1-methyl-2-[5-(2-thienylmethoxy)-1*H*-indole-3-yl]ethylamine hydrochloride), SB242084 (6-chloro-5-methyl-*N*-[6-(2-methylpyridin-3-yloxy)pyridin-3-yl]indoline-1-carboxamide), ketanserin, mianserin and ritanserin were synthesized by G. Lavielle, Servier. MK212 (4-(6-chloro-2-pyrazinyl)piperazine), SB215505 (6-chloro-5-methyl-*N*-(5-quinolinyl)-2,3-dihydro-1*H*-indole-1-carboxamide), LY266097 (1-(2-chloro-3,4-dimethoxybenzyl)-6-methyl-1,2,3,4-tetrahydro-9*H*-pyrido[3,4-*b*]indole hydrochloride) and S33526 ((6-chloro-2,3,4,9-tetrahydro-1*H*- β -carbolin-1-yl)-phenyl-acetic acid ethyl ester) were synthesized by S. Goldstein, J.G. Parmentier and G. Poissonnet, Servier. SB206553 (5-methyl-*N*-(3-pyridyl)-1,2,3,5-tetrahydrobenzo[1,2-*b*:4,5-*b'*]dipyrrole-1-carboxamide) and SB204741 (*N*-(1-methyl-5-indolyl)-*N'*-(3-methylisothiazol-5-yl)urea) were synthesized by J.-L. Peglion, Servier.

Results

Kinetics of [^3H]PI depletion as compared to [^3H]IP generation

At CHO-h5-HT_{2B} cells, 5-HT elicited [^3H]PI depletion that was expressed linearly over 40 min before attaining a plateau (Fig. 1A). The depletion of membrane-bound [^3H]PI at CHO-h5-HT_{2C} cells in response to 5-HT reached a plateau at about 30 min (Fig. 1B). The soluble phase of these samples revealed a similar time-course for h5-HT_{2C} receptor-mediated [^3H]IP generation (Fig. 1C). In the absence of 5-HT, no [^3H]PI depletion was observed in either cell line. On the basis of these observations, we selected incubation times of 30 min and 20 min for determination of [^3H]PI depletion at CHO-h5-HT_{2B} and CHO-h5-HT_{2C} cells, respectively. Compared with a total level of approximately 40,000–45,000 dpm (Fig. 1), 5-HT reduced [^3H]PI levels by about 15,000–20,000 dpm in both cell lines, a decrease of ~40% in each case.

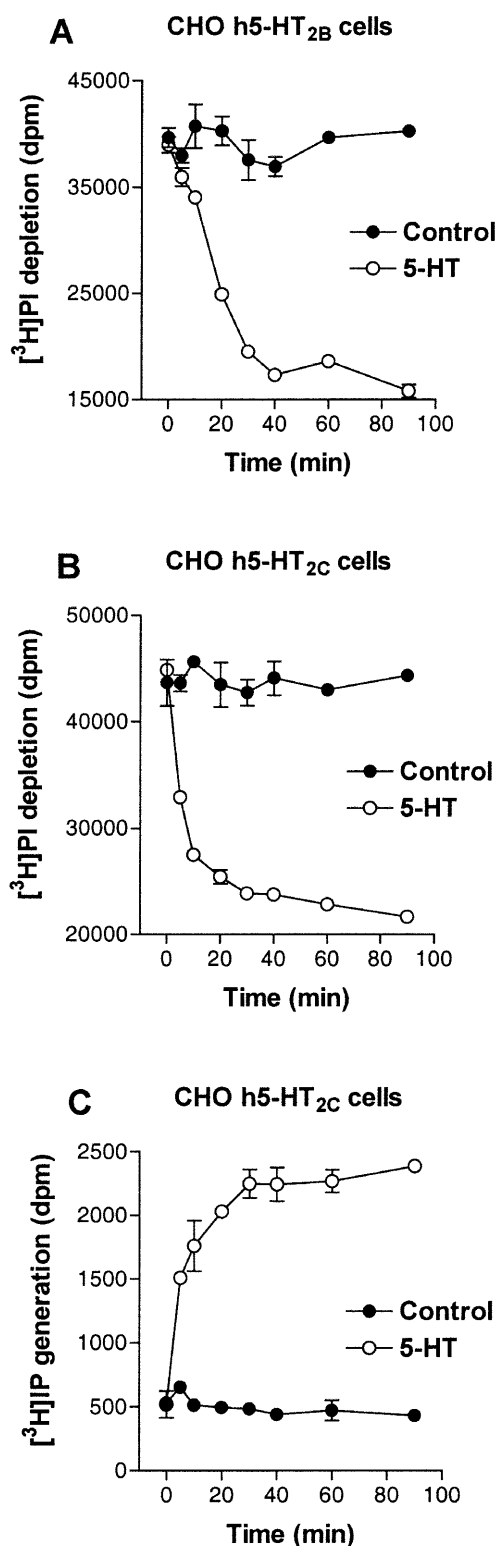


Fig. 1A–C Kinetics of phospholipase C activation at h5-HT_{2B} and h5-HT_{2C} receptors stably expressed in CHO cells. **A, B** Time-course of [^3H]phosphatidylinositol ([^3H]PI) depletion induced by 5-HT (10 μM) at h5-HT_{2B} and at h5-HT_{2C} cells, respectively. **C** Parallel time-course increase of 5-HT (10 μM)-stimulated soluble [^3H]inositol phosphate ([^3H]IP) generation at h5-HT_{2C} cells determined by anion exchange chromatography. Similar data were obtained on three occasions. A representative experiment performed in triplicate is shown

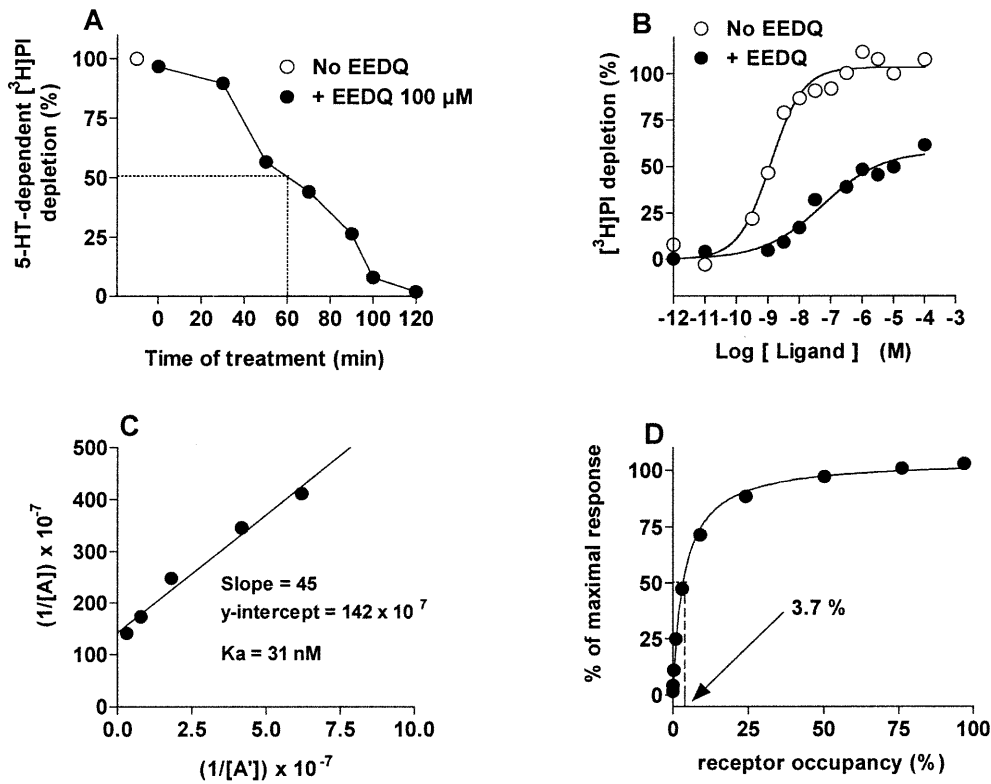


Fig. 2A–D Influence of h5-HT_{2C} receptor inactivation by EEDQ upon 5-HT-stimulated phospholipase C. **A** Time-dependent reduction of 5-HT (10 μM , 20 min)-stimulated $[^3\text{H}]\text{PI}$ depletion at h5-HT_{2C} cells by pre-treatment with 100 μM EEDQ (0–120 min). Similar data were obtained on three occasions. A representative experiment performed in triplicate is shown. **B** Concentration-response of 5-HT-induced $[^3\text{H}]\text{PI}$ depletion at control or EEDQ (100 μM , 60 min)-pre-treated CHO-h5-HT_{2C} cells. $[^3\text{H}]\text{PI}$ depletion is expressed as a percentage of maximal stimulation with 10 μM 5-HT in absence of EEDQ. **C** Double-reciprocal plot of $1/[A']$ vs. $1/[A]$ derived from B, where $[A']$ and $[A]$ are equiactive concentrations of 5-HT for stimulation of $[^3\text{H}]\text{PI}$ depletion with and without EEDQ treatment, respectively. **D** 5-HT occupancy/response relationship, derived using the value of K_A from C. The hyperbolic isotherm indicates the presence of receptor reserve. For B, C and D, points shown are the means of triplicate determinations from a representative experiment repeated on five occasions. The mean K_A value was $53.4 \pm 14.6 \text{ nM}$. Mean half-maximal response to 5-HT was observed at $4.6 \pm 0.9\%$ occupation of h5-HT_{2C} binding sites

presence of a high degree of receptor reserve was demonstrated in occupancy/response plots which yielded hyperbolic curves, with the mean half-maximal response to 5-HT being observed at $4.6 \pm 0.9\%$ occupation of h5-HT_{2C} binding sites (Fig. 2D). EEDQ treatment (60 min at 100 μM) decreased the density of h5-HT_{2C} receptors by over $\sim 97\%$ as determined by $[^3\text{H}]\text{mesulergine}$ saturation binding (Fig. 3; $21.6 \pm 3.4 \text{ pmol/mg}$ without EEDQ compared with $0.6 \pm 0.2 \text{ pmol/mg}$ in the presence of EEDQ). Thus, although only about $\sim 3\%$ of h5-HT_{2C} receptors remained following EEDQ treatment, $\sim 50\%$ of the influence of 5-HT upon $[^3\text{H}]\text{PI}$ levels was retained compared with non-EEDQ-treated cells.

Binding affinities of agonists and induction of $[^3\text{H}]\text{PI}$ depletion at h5-HT_{2B} and h5-HT_{2C} receptors

Influence of EEDQ treatment upon h5-HT_{2C} receptors

Addition of EEDQ (100 μM) to CHO-h5-HT_{2C} cells time-dependently reduced depletion of $[^3\text{H}]\text{PI}$ by 5-HT (Fig. 2A) without altering basal $[^3\text{H}]\text{PI}$ levels (not shown). Subsequent experiments were carried out with a treatment time of 60 min, which reduced the influence of 5-HT upon $[^3\text{H}]\text{PI}$ levels by about 50% (Fig. 2A,B). Under these conditions, the pEC_{50} of 5-HT was reduced to 7.50 ± 0.30 compared with 8.87 ± 0.05 for control cells (Fig. 2B; Table 1). The K_A value determined by Furchgott analysis was $53.4 \pm 14.6 \text{ nM}$ (Fig. 2C), which corresponded approximately to the affinity of 5-HT at h5-HT_{2C} receptors determined in competition binding experiments ($K_i = 18 \text{ nM}$; Table 1). The

Based on competition binding with $[^3\text{H}]\text{mesulergine}$, Ro600332 showed similar affinity at h5-HT_{2B} and h5-HT_{2C} receptors, whereas Ro600869 showed weak selectivity for h5-HT_{2C} receptors (Table 1; Fig. 4). All other agonists showed higher affinity at h5-HT_{2B} receptors. As derived from measures of $[^3\text{H}]\text{PI}$ depletion, the majority of agonists presented functional selectivity for h5-HT_{2C} vs. h5-HT_{2B} receptors. Thus, mCPP, MK212 and Ro600332 induced $[^3\text{H}]\text{PI}$ depletion with potencies $\sim 27\%$, $\sim 13\%$ and $\sim 12\%$ fold greater than at h5-HT_{2B} receptors, respectively. However, mCPP and MK212 displayed full efficacy at h5-HT_{2C} receptors, whereas they were partial agonists at h5-HT_{2B} receptors. In contrast, 5-HT and DOI acted as full agonists at both sites and displayed mild functional selectivity for

Table 1 Actions of agonists at h5-HT_{2C} compared with h5-HT_{2B} receptors: [³H]PI depletion (pEC₅₀; E_{max}) and binding affinities (pK_is). Agonist efficacies were determined by [³H]PI depletion and are expressed relative to the effects of 5-HT at 10 μM (defined as 100%), which were evaluated in all experiments. Lisuride did not

alter [³H]PI depletion at h5-HT_{2B} cells (see Table 2). Affinities (pK_i values) were determined by competition binding experiments with [³H]mesulergine. pEC₅₀ and pK_i values are expressed as means ± SEM of 3–10 independent experiments performed in triplicate (*n.c.* not computable)

Ligand	h5-HT _{2C}			h5-HT _{2B}			Selectivity h5-HT _{2B} /h5-HT _{2C}	
	pEC ₅₀	E _{max} (%)	pK _i	pEC ₅₀	E _{max} (%)	pK _i	EC ₅₀ ratio	K _i ratio
5-HT	8.87±0.05	104.7±3.6	7.74±0.05	8.52±0.03	102.0± 6.0	8.55±0.04	2.3	0.15
mCPP	8.41±0.18	100.6±0.3	7.07±0.17	6.98±0.18	37.9± 9.1	7.92±0.05	26.9	0.14
MK212	7.69±0.16	103.0±3.6	6.05±0.11	6.58±0.21	75.3±10.3	6.82±0.07	12.9	0.17
Ro600332	8.34±0.14	90.9±3.6	7.86±0.03	7.26±0.10	91.0± 5.0	7.80±0.02	12.0	1.14
DOI	8.55±0.03	101.6±3.1	7.73±0.08	7.98±0.07	103.6± 6.4	8.15±0.12	3.7	0.38
BW723C86	8.14±0.08	103.9±4.3	6.94±0.11	8.10±0.10	107.5± 3.6	8.50±0.04	1.1	0.02
Ro600175	8.72±0.07	97.5±3.8	7.57±0.03	9.23±0.09	90.2± 3.4	9.26±0.03	0.3	0.02
Lisuride	8.11±0.10	74.5±5.6	8.19±0.03	<i>n.c.</i> ^a	0	8.86±0.06	<i>n.c.</i> ^a	0.21
Ro600869	7.67±0.16	37.9±5.4	8.56±0.06	<i>n.c.</i> ^a	0	8.10±0.05	<i>n.c.</i> ^a	2.9

^aAntagonist; see Table 2

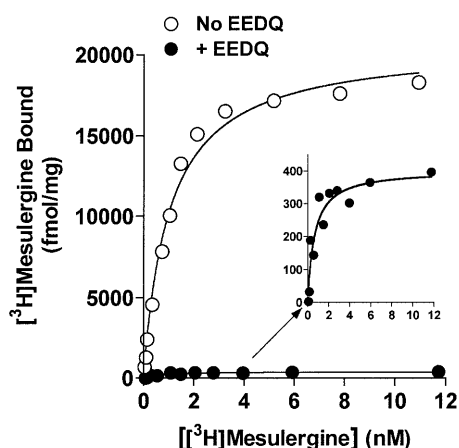


Fig. 3 Reduction of h5-HT_{2C} receptor density by EEDQ. Representative saturation binding isotherms of [³H]mesulergine to CHO-h5-HT_{2C} membranes pre-treated with and without EEDQ (100 μM for 60 min). Points shown are means of triplicate determinations from a representative experiment repeated on three occasions. The mean B_{max} values were 21.6±3.4 pmol/mg without EEDQ and 0.6±0.2 pmol/mg in presence of EEDQ, respectively. K_D values were unchanged, ~0.8 nM (the *insert* represents the EEDQ treatment at a higher scale)

h5-HT_{2C} receptors of ~2.3- and ~3.7-fold, respectively. BW723C86 was equipotent at these sites. Ro600175 was the only ligand which exhibited weak functional selectivity for h5-HT_{2B} receptors (~3.3-fold). Lisuride and Ro600869 did not induce [³H]PI depletion at h5-HT_{2B} receptors. Nevertheless, lisuride was a potent and relatively efficacious (75%) agonist at h5-HT_{2C} receptors, at which Ro600869 displayed low efficacy (38%). Interestingly, for h5-HT_{2C} receptors, pEC₅₀ values (except for partial agonists) were higher than pK_i values.

Binding affinities of antagonists and their influence upon 5-HT-induced [³H]PI depletion at h5-HT_{2B} and h5-HT_{2C} receptors

There was a marked correlation between antagonist potency (pK_b) and binding affinity (pK_i) for both h5-HT_{2C} (*r*=0.95, *P*<0.001) and h5-HT_{2B} sites (*r*=0.92, *P*<0.001; Table 2; Fig. 5). All antagonists tested, at both 5-HT_{2B} and 5-HT_{2C} receptors, completely abolished 5-HT-induced PLC activation, demonstrating that endogenous 5-HT_{1B} receptors expressed in CHO cells (Giles et al. 1996) are not involved in PLC regulation. RS102221 displayed pronounced affinity for h5-HT_{2C} receptors and the highest degree of selectivity vs. h5-HT_{2B} receptors (>100-fold). A pronounced preference for h5-HT_{2C} receptors was similarly displayed by SB242084. Although it was more potent than RS102221 at h5-HT_{2C} receptors, it was somewhat less selective (Table 2). Ketanserin showed a modest preference for h5-HT_{2C} vs. h5-HT_{2B} sites as concerns both its binding affinity and its antagonist potency. On the other hand, mesulergine, metergoline, SB206553, mianserin and ritanserin failed to clearly discriminate h5-HT_{2C} and h5-HT_{2B} receptors, based on both their pK_i and pK_b values. While SB215505 behaved as a modestly preferential antagonist at h5-HT_{2B} vs. h5-HT_{2C} sites, this difference was considerably more pronounced for several other agents. Thus, both SB204741 and S33526 manifested a high degree of selectivity for h5-HT_{2B} sites (>30-fold), although the former was less potent at this site. LY266097 was the most potent and selective antagonist at h5-HT_{2B} vs. h5-HT_{2C} sites of all ligands evaluated. Interestingly, lisuride and Ro600869 behaved as h5-HT_{2B} antagonists although, as described above, they possess agonist properties at h5-HT_{2C} receptors. None of the antagonists provoked [³H]PI depletion at h5-HT_{2B} or h5-HT_{2C} sites upon application alone (not shown). Several antagonists, including the h5-HT_{2C}-selective ligands, SB242084 and RS102221, and the h5-HT_{2B}-selective ligands, SB204741, S33526 and LY266097, showed low binding affinities at h5-HT_{2A} receptors la-

Fig. 4 Concentration-response curves of agonist actions at h5-HT_{2C} and h5-HT_{2B} receptor-mediated phospholipase C activation. [³H]PI depletion is expressed as a percentage of maximal stimulation evoked by 10 μM 5-HT. Points shown are means of triplicate determinations from representative experiments repeated on at least 3–10 occasions. *E*_{max} and pEC₅₀ data from these experiments are shown in Table 1

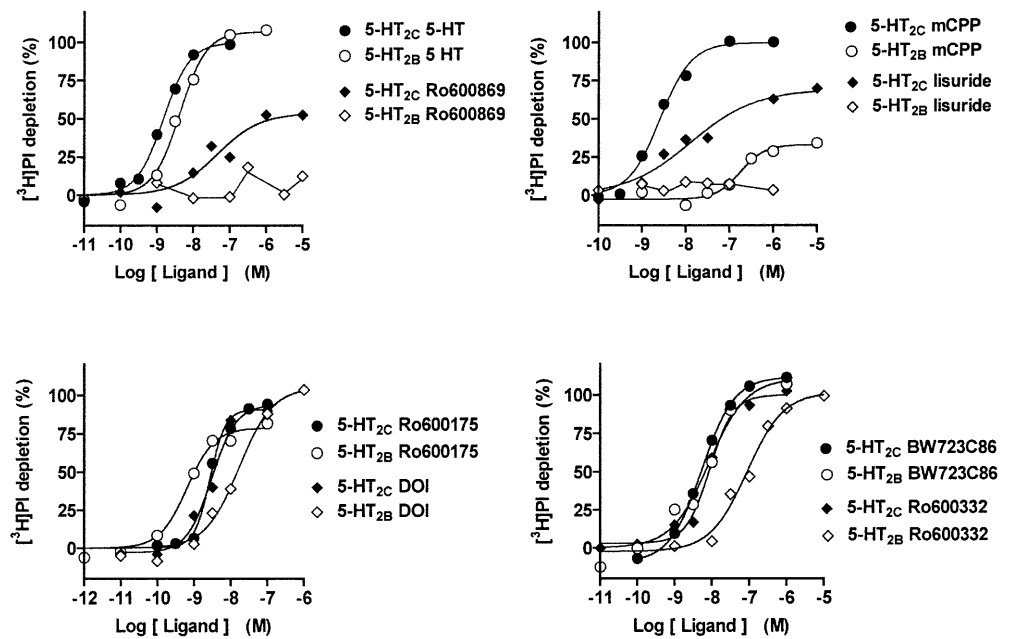


Table 2 Actions of antagonists at h5-HT_{2C} compared with h5-HT_{2B} receptors: inhibition of 5-HT-induced [³H]PI depletion (p*K*_B) and binding affinities. Antagonist potencies (p*K*_B values) were calculated from IC₅₀ values for inhibition of 5-HT-stimulated [³H]PI depletion (30 nM for h5-HT_{2B} and 100 nM or 30 nM for h5-HT_{2C}).

Affinities (p*K*_i values) were determined by competition binding experiments with [³H]mesulergine. p*K*_B and p*K*_i values are expressed as means ± SEM of 3–6 independent experiments performed in triplicate (*n.c.* not computable)

Ligand	h5-HT _{2C}		h5-HT _{2B}		Selectivity h5-HT _{2B} /h5-HT _{2C}	
	p <i>K</i> _B	p <i>K</i> _i	p <i>K</i> _B	p <i>K</i> _i	<i>K</i> _B ratio	<i>K</i> _i ratio
RS102221	8.47±0.10	8.83±0.04	5.99±0.07	6.63±0.05	302	158
SB242084	9.53±0.14	9.32±0.06	7.70±0.03	7.34±0.07	67	95
Ketanserin	7.17±0.13	7.77±0.03	6.36±0.05	6.63±0.06	6.4	14
Mesulergine	9.14±0.08	8.95±0.06	8.58±0.13	8.71±0.02	3.6	1.73
Metergoline	9.30±0.06	9.33±0.07	9.03±0.20	9.15±0.01	1.9	1.51
SB206553	8.35±0.09	8.50±0.13	8.44±0.06	8.26±0.17	0.81	1.73
Mianserin	8.14±0.12	8.82±0.04	8.68±0.09	8.26±0.05	0.28	3.63
Ritanserin	8.84±0.05	8.79±0.10	9.49±0.13	8.90±0.13	0.22	0.77
SB215505	8.47±0.06	7.95±0.06	9.16±0.15	8.83±0.09	0.20	0.13
SB204741	5.95±0.05	5.67±0.11	7.53±0.11	7.29±0.04	0.026	0.023
S33526	6.34±0.05	6.34±0.04	8.11±0.17	8.39±0.07	0.016	0.009
LY266097	7.03±0.09	7.17±0.19	9.68±0.05	9.70±0.07	0.002	0.002
Lisuride	<i>n.c.</i> ^a	8.19±0.03	8.94±0.06	8.86±0.06	<i>n.c.</i> ^a	0.21
Ro600869	<i>n.c.</i> ^a	8.56±0.06	7.02±0.10	8.10±0.05	<i>n.c.</i> ^a	2.88

^aAgonist; see Table 1

belled by [³H]ketanserin; selectivity of at least 100-fold vs. h5-HT_{2C} and h5-HT_{2B}, respectively (not shown).

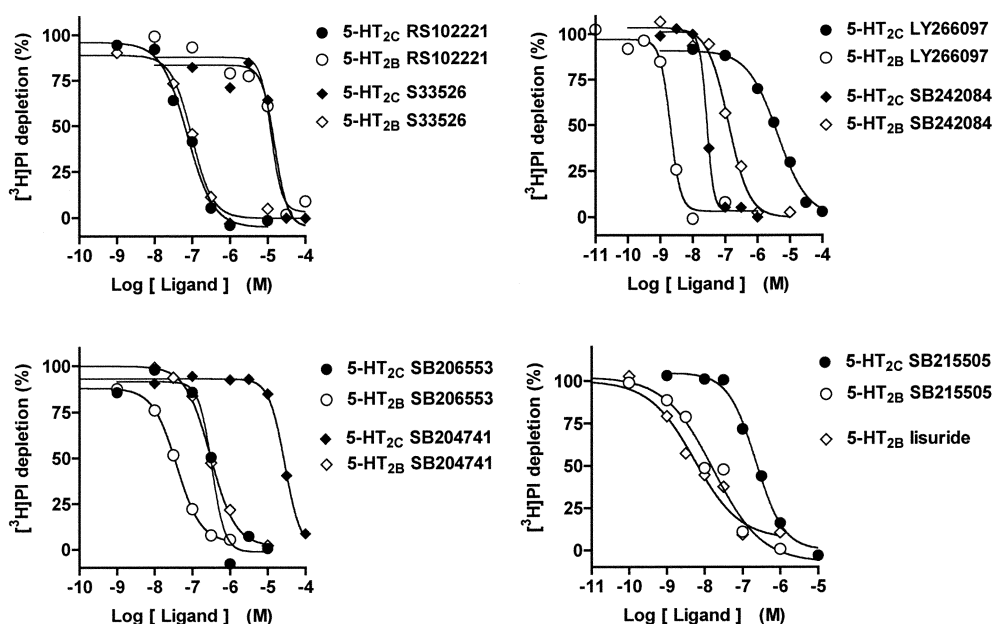
Discussion

Technical aspects

Determination of membrane-bound [³H]phosphatidylinositol ([³H]PI) depletion, as compared to generation of cytosolic [³H]inositol phosphates ([³H]IP), offers a rapid technique for evaluation of drug activity at h5-HT_{2C}, h5-HT_{2B}

receptors and, in principle, other sites coupled to PLC. As discussed elsewhere (Cussac et al. 2000b), addition of LiCl prevents the recycling of inositol phosphates (Berridge 1993), permitting measurement of the influence of ligands upon membrane-bound [³H]PI. [³H]PI consists, thus, of the substrate of PLC, phosphatidylinositol-4,5 biphosphate, as well as its precursors, phosphatidylinositol-4-phosphate and phosphatidylinositol (Berridge 1993). Emphasizing the pharmacological specificity of this novel approach, measures of [³H]PI generated results identical to a conventional [³H]IP measurement as concerns actions of 5-HT and the antagonist, SB242084, at h5-HT_{2C} receptors (Cussac

Fig. 5 Concentration-response curves of antagonism actions at h5-HT_{2C} and h5-HT_{2B} receptor-mediated phospholipase C activation. [³H]PI depletion is expressed as a percentage of maximal stimulation elicited by 10 μM 5-HT. Points shown are means of triplicate determinations from representative experiments repeated on at least 3–5 occasions. pK_B data from these experiments are shown in Table 2



et al. 2000b). Moreover, for diverse antagonists examined herein, pK_b values for blockade of 5-HT-induced [³H]PI depletion correlated tightly with pK_i values at h5-HT_{2C} and h5-HT_{2B} receptors. The present observation that, at h5-HT_{2C} receptors, the time-course of [³H]PI depletion by 5-HT parallels that for generation of [³H]IP as monitored by anion exchange chromatography under the same assay conditions (Dickenson and Hill 1996) further underpins the validity of the present approach.

Pre-loaded CHO-h5-HT_{2B} and CHO-h5-HT_{2C} cells can be stored frozen (see Materials and methods) without affecting drug actions (not shown). Together with the advantage of a simple filtration step for determination of [³H]PI, this allows for use of the present protocol in the rapid characterization of large numbers of ligands, which would be prohibitively time-consuming employing a standard [³H]IP procedure.

h5-HT_{2C} receptors alkylation with EEDQ reveals receptor reserve

With the exception of lisuride and Ro600869, all agonists at h5-HT_{2C} receptors exhibited high K_i/EC₅₀ ratios (7–40) and “full” efficacy relative to 5-HT, suggesting that there exists a substantial receptor reserve (Adham et al. 1993; Brink et al. 2000; Table 1). h5-HT_{2C} receptors have been shown to be sensitive to EEDQ treatment (Ni et al. 1997). Indeed, the presence of spare h5-HT_{2C} receptors was revealed upon their inactivation with EEDQ, which reduced their levels to 3% of control values, yet diminished the 5-HT-induced PLC response by 50%. The presence of pronounced receptor reserve was confirmed by Furchgott analysis which yielded a K_A/EC₅₀ ratio of 40 for 5-HT, and a hyperbolic occupancy/response plot with a half-maximal response at 4.6%, similar to the 3% value men-

tioned above. These data indicate that the present cellular expression system (CHO-h5-HT_{2C}) exhibits very high sensitivity to agonist stimulation. In comparison, in a receptor inactivation study employing a different cell line (NIH3T3) expressing 5 pmol/mg of edited VSV or non-edited INI h5-HT_{2C} isoforms, no receptor reserve was detected for PLC activation (Burns et al. 1997). These observations indicate that careful characterisation of signal transduction parameters in each expression system is important for appropriate interpretation of functional responses (Burstein et al. 1997). It is not known what receptor expression levels are physiological. Saturation binding studies using brain tissue homogenates yield measures of receptor number for a heterogeneous mix of cell types. Hence 5-HT_{2C}-receptor expression levels per 5-HT_{2C}-expressing neuron, for instance, can only be roughly estimated. In the case of rat choroid plexus neurons, 5-HT_{2C} receptor expression is reported to be in the pmol/mg range (Yagaloff and Hartig 1985), thus potentially implicating receptor reserve, as described herein.

Inasmuch as the primary focus of the present study was h5-HT_{2C} receptors, the possible presence of spare receptors at h5-HT_{2B} receptors was not evaluated by receptor inactivation herein. In fact the close correspondence of K_i and EC₅₀ values for all agonists except mCPP (see below) suggests that this is unlikely to be pronounced. Nevertheless, this aspect, and the high expression level of h5-HT_{2C} vs. h5-HT_{2B} receptors should be borne in mind in the following comparison of drug actions at h5-HT_{2C} vs. h5-HT_{2B} sites.

Agonist actions at h5-HT_{2B} and h5-HT_{2C} receptors

The high level of receptor reserve in the present CHO-h5-HT_{2C} cell line has important implications for interpretation of agonist properties. Indeed, the potencies and effi-

cacies of several agonists herein were more pronounced compared with previous studies of CHO-expressed h5-HT_{2C} (VSV) receptors (Porter et al. 1999) or SH-SY5Y-expressed h5-HT_{2C} (INI) receptors (Jerman et al. 2001). Thus, while these authors did not directly determine receptor reserve, the expression levels of h5-HT_{2C} receptors in their systems was low (0.2 pmol/mg for VSV and 0.9–6.3 pmol/mg for INI, compared with 20 pmol/mg herein). Notably, these authors did not detect agonist properties of lisuride at h5-HT_{2C} receptors, whereas it displayed substantial efficacy in the present study, in analogy to reports of a highly expressed VNV isoform of h5-HT_{2C} receptors (Egan et al. 1998; Fitzgerald et al. 1999). The novel, selective 5-HT₂ ligand, Ro600869, mimicked the actions of lisuride at h5-HT_{2C} receptors, exhibiting modest partial agonist activity, although it was previously reported to act as an antagonist at h5-HT_{2C} receptors (Boes et al. 1997, 1998). In view of the potential utility of Ro600869 as a selective 5-HT_{2C} ligand and its structural novelty, it would be of interest to extend these observations to other cellular paradigms. Taken together, the present data demonstrate that: (1) a high degree of receptor reserve in the present CHO-h5-HT_{2C} cell line can reveal partial agonist properties of low efficacy ligands and (2) lisuride and Ro600869 display antagonist properties at h5-HT_{2B} receptors.

Inasmuch as mCPP is extensively exploited as an experimental and clinical probe for evaluation of 5-HT_{2C} receptor function, its actions are of particular importance. Indeed, agonist actions of mCPP have been characterised (Kennett and Curzon 1988; Newton et al. 1996; Porter et al. 1999; Miller et al. 2000; Jerman et al. 2001). As concerns actions at recombinant h5-HT_{2B} and h5-HT_{2C} receptors, Thomas et al. (1996) documented antagonist properties at h5-HT_{2B} receptors expressed in HEK293 cells. However, these findings may be characteristic of that expression system inasmuch as, in the present CHO cell line (which expresses a similar level of 5-HT_{2B} receptors), partial agonist properties of mCPP were observed. Moreover, despite a sixfold lower expression of h5-HT_{2B} receptors in another CHO cell line (0.6 pmol/mg compared with 3 pmol/mg herein), partial agonist actions of mCPP were also characterised by an increase in cytosolic Ca²⁺ concentrations (Porter et al. 1999). Further, agonist properties of mCPP at native, rat 5-HT_{2B} receptors have been documented (Baxter et al. 1995).

In contrast to mCPP, BW723C86 was proposed as a tool for exploration of the role of 5-HT_{2B} vs. 5-HT_{2C} receptors based on its selectivity in binding studies (Baxter 1996; Kennett et al. 1996a, 1997a). Indeed, employing a measure of cytosolic Ca²⁺ concentrations, BW723C86 displayed ~100-fold functional selectivity for h5-HT_{2B} sites (Porter et al. 1999; Jerman et al. 2001). In contrast, the present binding studies revealed higher binding affinity of BW723C86 at h5-HT_{2B} sites, but showed comparable potency and efficacy at h5-HT_{2B} and h5-HT_{2C} sites in eliciting [³H]PI depletion. Although the present functional selectivities should be interpreted in the context of the marked h5-HT_{2C} receptor reserve, these data are consistent with the action of BW723C86 in activating PLC at h5-HT_{2C} ex-

pressed in HEK293 cells (Thomas et al. 1996; Wood et al. 1997).

Ro600175 was originally suggested to be a selective agonist for 5-HT_{2C} receptors, principally based upon binding studies (Boes et al. 1997). Subsequently, *in vivo* studies demonstrated that 5-HT_{2C} receptors are indeed involved in its functional actions (Jenck et al. 1998b; Dekeyne et al. 1999; Gobert et al. 2000). However, Ro600175 also behaves as a full agonist at native, rat 5-HT_{2B} receptors (Martin et al. 1998) and it was more potent in elevating cytosolic Ca²⁺ concentrations at CHO-expressed h5-HT_{2B} than h5-HT_{2C} receptors (Porter et al. 1999). The present study corroborates these observations in demonstrating that Ro600175 more potently enhances [³H]PI depletion at h5-HT_{2B} vs. h5-HT_{2C} receptors. In fact, Ro600175 may currently be the most selective h5-HT_{2B} agonist available.

In contrast, Ro600332, while exhibiting equilibrated binding affinity and full agonist properties at both h5-HT_{2B} and h5-HT_{2C} receptors, manifested 12-fold greater potency in enhancing [³H]PI depletion at the latter. In the light of the present results and *in vivo* studies demonstrating robust agonist actions at 5-HT_{2C} receptors in rodents (Jenck et al. 1998a, 1998b; Martin et al. 1998), Ro600032 may, thus, be a more appropriate ligand than Ro600175 for exploration of its functional significance. Nevertheless, as for all the agonists evaluated herein, caution needs to be exercised in the interpretation of their functional selectivity due to the presence of spare receptors in this CHO-h5-HT_{2C} cell line.

Antagonist actions at h5-HT_{2B} and h5-HT_{2C} receptors

The latter point emphasises the importance of studies with antagonists for differentiating actions at 5-HT_{2C} vs. 5-HT_{2B} receptors. In fact, the rapidity of the present technique permitted the – to our knowledge – most extensive comparative study to date of the functional properties of diverse antagonists at h5-HT_{2C} vs. h5-HT_{2B} receptors. In view of the high degree of correlation between pK_B and pK_i values, comparison of ratios (5-HT_{2B}/5-HT_{2C}) in functional tests as well as binding affinities permitted the identification of three groups of antagonists in accordance with their selectivity (Table 2). The most h5-HT_{2C}-selective ligand found herein, RS102221, was also described as a selective 5-HT_{2C} receptor antagonist based on a microphysiometry assay which yielded a pA₂ of 8.1 (Bonhaus et al. 1997). This value is close to the pK_B of 8.4 (present study), confirming its potent and preferential blockade of h5-HT_{2C} compared with h5-HT_{2B} receptors. Previous studies demonstrated antagonist properties of RS102221 at central 5-HT_{2C} sites under certain conditions (Bonhaus et al. 1997; Sevoz-Couche et al. 2000), and the present data support its utility as a tool for differentiation of actions mediated at 5-HT_{2C} vs. 5-HT_{2B} receptors. This is also the case for SB242084 (Kennett et al. 1997b), which exhibited pronounced selectivity for h5-HT_{2C} receptors, underpinning previous observations of its potent – and competitive – antagonist properties at h5-HT_{2C} receptors *in vitro* (Cussac et al. 2000b) and *in vivo* (Dekeyne et al. 1999).

In contrast, the structurally related analogues, SB206553 and SB215505, were potent antagonists at both sites (Kennett et al. 1996b, 1998b; Dekeyne et al. 1999). Although SB215505 was described as a preferential h5-HT_{2B} ligand, the poor selectivity for this site observed herein (<tenfold) encourages caution in the interpretation of its actions. Amongst other weakly discriminative h5-HT_{2C} vs. h5-HT_{2B} receptor ligands, mesulergine, metergoline, mianserin and ritanserin were potent antagonists at both sites, while ketanserin displayed lower potency and a mild preference for h5-HT_{2C} sites (Baxter et al. 1995).

As concerns selective h5-HT_{2B} antagonists, SB204741 was previously described as a selective ligand based on its actions in functional tests in vitro (Baxter et al. 1995; Thomas et al. 1996) and in vivo (Dekeyne et al. 1999; Glusa and Pertz 2000; Gobert et al. 2000). This preference was substantiated herein, although SB204741 showed only modest potency at h5-HT_{2B} sites. On the other hand, LY266097 and the novel ligand, S33526, displayed both high potency and high selectivity for h5-HT_{2B} vs. h5-HT_{2C} sites. These ligands appear, therefore, of particular use for separating actions mediated via h5-HT_{2B} rather than h5-HT_{2C} sites.

Conclusions

Employing an innovative and rapid technique for quantification of the influence of drugs upon PLC activity, the present study comprises the, to date, most comprehensive characterization of novel (and putatively selective) agonists and antagonists at h5-HT_{2C} receptors. The high receptor reserve at h5-HT_{2C} receptors encourages caution as concerns comparisons with h5-HT_{2B} receptors, but the present data indicate that, in contrast to antagonists, selective agonists at these sites remain to be identified. In view of the pleiotropic coupling of 5-HT_{2C} receptors to diverse G-proteins and intracellular cascades (Gerhardt and Heerikhuizen 1997), considerable research is still required to understand mechanisms of intracellular coupling at 5-HT_{2C} receptors, as well as their pathophysiological significance. In this light, the present data provide an extensive framework for further exploration of drug actions at 5-HT_{2C} receptors.

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