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ORIGINAL ARTICLE

Circulating levels of dickkopf-1, osteoprotegerin and sclerostin are higher in old compared with young men and women and positively associated with whole-body bone mineral density in older adults

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Abstract

Summary Bone mineral density declines with increasing older age. We examined the levels of circulating factors known to regulate bone metabolism in healthy young and older adults. The circulating levels of dickkopf-1, osteocalcin, osteoprotegerin and sclerostin were positively associated with whole-body bone mineral density (WBMD) in older adults, despite the average WBMD being lower and circulating dickkopf-1, osteoprotegerin and sclerostin being higher in old than young.

Introduction This study aims to investigate the relationship between whole-body bone mineral density (WBMD) and levels of circulating factors with known roles in bone remodelling during 'healthy' ageing.

Methods WBMD and fasting plasma concentrations of dickkopf-1, fibroblast growth factor-23, osteocalcin, osteoprotegerin, osteopontin and sclerostin were measured in 272 older subjects (69 to 81 years; 52% female) and 171 younger subjects (18–30 years; 53% female).

Results WBMD was lower in old than young. Circulating osteocalcin was lower in old compared with young, while dickkopf-1, osteoprotegerin and sclerostin were higher in old compared with young. These circulating factors were each positively associated with WBMD in the older adults and the relationships remained after adjustment for covariates (*r* values ranging from 0.174 to 0.254, all $p < 0.01$). In multivariate regression, the body mass index, circulating sclerostin and whole-body lean mass together accounted for 13.8% of the variation with WBMD in the older adults. In young adults, dickkopf-1 and body mass index together accounted for 7.7% of variation in WBMD.

Conclusion Circulating levels of dickkopf-1, osteocalcin, osteoprotegerin and sclerostin are positively associated with

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WBMD in community-dwelling older adults, despite the average WBMD being lower and circulating dickkopf-1, osteoprotegerin and sclerostin being higher in old than young.

Keywords DKK1 · MYOAGE · Osteoporosis · Osteoprotegerin · Sclerostin

Introduction

Progressive loss of bone mineral density (BMD) in older age leads to osteoporosis as the balance of bone remodelling favours resorption of mineralised extracellular matrix over formation. This common change is characterised by 'micro-architectural' deterioration of bone tissue and increases the risk of fracture [1]. Circulating factors influencing bone development have been implicated in the age-related changes to BMD. This includes regulatory factors released from osteoblasts and osteocytes involved in bone formation and from osteoclasts with bone resorption. These factors can enter the circulation where their concentrations may be related to BMD in older age.

Some of the candidate circulating factors possibly related to BMD include osteoprotegerin (OPG), which is expressed by osteocytes and osteoblasts and can reduce production of osteoclasts by binding receptor activator of nuclear factor kappa-B ligand (RANKL) [2]. Osteocalcin (OC) is a major non-collagen protein of the bone matrix secreted by osteoblasts for bone formation, but released from the matrix during bone resorption [3]. Dickkopf-1 (DKK1) [4] and sclerostin, released primarily by osteocytes [5], negatively regulate bone formation and have emerged as therapeutic targets to tackle osteoporosis [6]. Fibroblast growth factor 23 (FGF23) is produced by a variety of cell types, including osteoblasts and osteocytes, and released into the circulation where it acts on the kidney to increase excretion of phosphate and reduce production of 1–25 OH vitamin D [7]. Osteopontin (OPN) is an extracellular matrix protein released by osteoblasts, osteocytes and osteoclasts to facilitate bone resorption [8].

It remains unclear how the combination of these circulating markers of bone turnover are related to BMD in older age. Therefore, the purpose of this study was to compare plasma concentrations of these markers between recreationally active, community-dwelling older adults and a reference group of young adults, and to examine the association of these with whole-body bone mineral density (WBMD). It was hypothesised that older adults would have higher circulating levels of factors related to bone resorption compared with young, and higher circulating markers of bone resorption were expected to be associated with lower BMD in old age.

Materials and methods

Study design

The cross-sectional European multi-centre MYOAGE cohort consists of relatively healthy older men and women (aged 69 to 81 years) and young adults (aged 18–30 years) [9]. The study was approved by ethics committees at each institute, and written informed consent was obtained from all participants. Participants were recruited by advertisement in newspapers, the University of the Third Age and Association of Emerti. All measurements were performed according to standard operating procedures that had been unified at the study centres, and data collection was ceased through December–March and July–August. Volunteers were excluded if dependent living, unable to walk a distance of 250 m, presence of morbidity (such as neurologic disorders, metabolic diseases, rheumatic diseases, heart failure, severe chronic obstructive pulmonary disease and haemocoagulative syndromes), immobilisation for 1 week during the last 3 months and orthopaedic surgery during the last 2 years or still causing pain or functional limitations. The inclusion and exclusion criteria were designed to ensure the selection of relatively healthy participants and to minimise the confounding effect of comorbidity on sarcopenia [9], and we recorded the use of bisphosphonates, calcium and vitamin D supplements. The present study included 443 participants [Leiden, The Netherlands (young, $n = 35$; old, $n = 75$); Jyvaskyla, Finland (young, $n = 34$; old, $n = 65$); Tartu, Estonia (young, $n = 39$; old, $n = 60$), Paris, France (young, $n = 35$; old, $n = 30$) and Manchester, UK (young, $n = 28$; old, $n = 42$)] with complete BMD and blood results.

Dual-energy x-ray absorptiometry (DXA)

A whole-body scan was performed using DXA while the participants lay supine, as previously reported [9] (The Netherlands—Hologic QDR 4500, version 12.4, Hologic Inc., Bedford, MA, USA; Finland—Lunar Prodigy, version en-Core 9.30; Estonia—Lunar Prodigy Advanced, version en-Core 10.51.006; France—Lunar Prodigy, version encore 12.30; UK—Lunar Prodigy Advance, version enCore 10.50.086). A trained technician completed the daily equipment calibration and the DXA scans according to local and manufacturers' quality control procedures. Participants wore a light cotton garment to reduce effects of clothing absorption on the scanning results. The whole-body lean mass, fat mass and the WBMD were recorded after manual adjustment of the regions of interest carried out offline.

Blood sample analysis

Blood samples were collected from a vein in the forearm into vacutainer EDTA tubes in the morning when participants were

in a fasted state. Samples remained at room temperature for 15–30 min and were then centrifuged for 15 min at $2000 \times g$ at 4 °C. The plasma was collected and stored at −80 °C until analysis. Plasma concentrations of the selected analytes were determined in the research laboratory in Manchester, UK, using multiplex immunoassays (Millipore, Billerica, MA, USA). The manufacturer instructions were followed and the magnetic bead panels quantified DKK1, OPG, OC, OPN, sclerostin and FGF23 using a 96-well plate after an overnight incubation. The sensitivity of each analyte was 1.4 (DKK1), 1.9 (OPG), 68.5 (OC), 37.7 (OPN), 31.1 (sclerostin) and 9.2 (FGF23) pg/mL. Samples were processed using a Luminex 200 Bioanalyser and protein concentrations were estimated using the xPONENT software (Luminex, v.3.1.871).

Statistical analysis

Participant descriptive characteristics (Table 1) were normally distributed and are presented as mean ± standard error of the mean (SEM). Comparisons between age and gender were assessed using multivariate ANOVA. Relationships between body stature, BMI, total body lean mass and supplement use (independent variables) with WBMD (dependent variable) were assessed using bivariate Pearson's product moment correlation. Data for circulating factors were not normally distributed and are presented as median (25th/75th) centiles. The results were log-transformed and z-scores calculated by expressing each log-transformed value as a standard deviation from the mean of the gender-matched young. Z-scores of WBMD, lean mass and BMI were also calculated for use in subsequent correlation and regression analysis. Spearman's rho partial correlations were performed to assess relationships between the z-score WBMD with z-scores of circulating factors using two models. The first model included adjustment for

country of testing to account for any systematic differences. The second accounted for the positive correlations we observed between WBMD and BMI in men and women (r values ranging from 0.210 to 0.387) and WBMD and lean mass for men (r values in men ranging from 0.268 to 0.357, and women 0.085–0.099) as well as health status and use of bisphosphonates, calcium or vitamin D supplements. Thus, the second model included adjustments for country of testing, z-score of lean mass, z-score of BMI, self-reported health and supplement use. A stepwise multiple linear regression using the self-reported health and supplement use as well as z-scores for BMI, lean mass and circulating factors was then used to evaluate which combination of the independent variables was associated with z-score WBMD (dependent variable) in older adults and in young adults. Data was analysed using SPSS for Windows (v.21; IBM, USA) and significance accepted as $p < 0.05$.

Results

Based on z-scores relative to gender-matched young, 26% of the older participants had WBMD values between −1.5 and −2.49 below the mean for young and 10.6% were ≥−2.5 below the mean of young. There was a significant age-by-gender interaction for WBMD z-scores ($p < 0.0005$).

Table 2 shows concentrations of the circulating factors. Compared with young, older participants had higher concentrations of DKK1, OPG and sclerostin. Concentrations of OC were significantly lower in old compared with young. OPN and FGF23 did not differ significantly between young and older participants although this was after removal of 37% of FGF23 samples (similar proportions of young and old) that fell below the level of assay detection. Compared with men, women had higher circulating concentrations of OPG, but lower OPN and

Table 1 Participant descriptive characteristics

	Old		Young		p value	
	Men ($n = 129$)	Women ($n = 143$)	Men ($n = 82$)	Women ($n = 89$)	Age	Gender
Age (years)	74.6 ± 0.3	74.0 ± 0.3	23.6 ± 0.3	23.2 ± 0.3	<0.0005	
Height (m)	1.74 ± 0.01	1.61 ± 0.01	1.81 ± 0.01	1.67 ± 0.01	<0.0005	<0.0005
Body mass (kg)	78.8 ± 1.0	65.1 ± 0.8	75.4 ± 1.2	62.4 ± 1.0	<0.0005	0.018
BMI (kg/m^2)	25.8 ± 0.3	25.2 ± 0.3	23.1 ± 0.3	22.4 ± 0.3	0.017	<0.0005
Body fat (kg)	20.1 ± 0.7	22.7 ± 0.6	12.9 ± 0.7	18.8 ± 0.7	<0.0005	<0.0005
Lean mass (kg)	55.9 ± 0.6	40.2 ± 0.5	59.9 ± 0.9	41.4 ± 0.6	<0.0005	<0.0005
Body fat (%)	25.5 ± 0.6	34.6 ± 0.6	16.6 ± 0.7	29.6 ± 0.7	<0.0005	<0.0005
Lean mass (%)	71.9 ± 0.6	63.0 ± 0.6	79.8 ± 0.7	67.2 ± 0.7	<0.0005	<0.0005
WBMD (g/cm^2)	1.19 ± 0.01	1.04 ± 0.01	1.25 ± 0.01	1.15 ± 0.01	<0.0005	0.001
WBMD (z-score)	-0.63 ± 0.10	-1.47 ± 0.11	0.00 ± 0.11	0.00 ± 0.11	<0.0005	<0.0005

Values are mean ± SEM

WBMD whole-body bone mineral density

Table 2 Circulating markers of bone remodelling in old and young, men and women

	Old		Young		p value		
	Men	Women	Men	Women	Age	Gender	Age × gender
DKK1 (pg mL^{-1})	577.0 ± 352–804	575.3 ± 346–864	420.6 ± 290–627	494.3 ± 284–703	<0.0005	0.942	0.843
FGF23 (pg mL^{-1})	113.5 ± 72–274 (n = 75)	103.0 ± 64–211 (n = 87)	122.9.7 ± 87–195 (n = 54)	141.7 ± 94–225 (n = 60)	0.792	0.316	0.700
OC (pg mL^{-1})	14,160.5 ± 9911–18,708	16,065.4 ± 11,073–19,933	17,581.1 ± 13,304–21,223	16,733.9 ± 12,013–20,715	<0.0005	0.880	0.036
OPG (pg mL^{-1})	319.2 ± 229–419	306.9 ± 257–392	159.4 ± 114–193	208.5 ± 160–260	<0.0005	<0.0005	<0.0005
OPN (pg mL^{-1})	26,590.1 ± 17,094–38,028	21,350.1 ± 13,971–31,255	24,822.5 ± 16,928–35,662	20,877.5 ± 15,937–27,777	0.700	0.009	0.184
Sclerostin (pg mL^{-1})	5690.3 ± 4348–7556	4147.6 ± 3349–5159	3016.1 ± 2079–3932	2366.0 ± 1923–3134	<0.0005	<0.0005	0.034

Values are median ± 25th–75th percentiles. For FGF23, the n is less than those given in Table 1 due to some samples having values that were below the level of detection. The n for all other analytes is the same as shown in Table 1

sclerostin. There were no significant differences between men and women for DKK1, FGF23 and OC. Age × gender interactions were found for OC, OPG and sclerostin (all $p < 0.05$); the difference between young and old in OC, OPG and sclerostin was greater for men than it was for women.

Table 3 shows the associations between circulating bone regulatory factors and WBMD. When using z-scores of all variables and including all participants, while adjusting for country, WBMD was positively associated with DKK1. This association remained significant after additionally adjusting for lean mass, BMI, self-reported health and supplement use. In older participants only, DKK1, OC, OPG and sclerostin were positively associated with WBMD after adjusting for country. This remained the case when additionally adjusting for lean mass, BMI, self-reported health and supplement use. In younger participants only, DKK1 was positively associated with WBMD after adjusting for country as well as when additionally adjusting for lean mass, BMI, self-reported health and supplement use.

Stepwise multiple linear regression was performed including z-score WBMD as the dependent variable, and independent variables included self-reported health, supplement use and z-scores of the variables BMI and lean mass, as well as the z-scores derived from log-transformed data for DKK1, FGF23, OC, OPG, OPN and sclerostin. Results in the young showed DKK1 accounted for 5.1% of the variation in WBMD (adjusted $r^2 = 0.051$, $p = 0.010$), while DKK1 and BMI accounted for 7.7% of the variation in WBMD (adjusted $r^2 = 0.077$, $p = 0.005$). In the old, BMI alone accounted for 8.9% of the variation in WBMD (adjusted $r^2 = 0.089$, $p < 0.0005$); BMI and sclerostin together accounted for 12.0% of the variation in WBMD (adjusted $r^2 = 0.120$, $p < 0.0005$), while BMI, sclerostin and whole-body lean mass accounted for 13.8% of the variation in WBMD (adjusted $r^2 = 0.138$, $p < 0.0005$).

Discussion

The results of this study showed that circulating factors DKK1, OPG and sclerostin were each higher in old compared with young, but positively associated with WBMD in older adults. Circulating OC was lower in old compared with young and positively associated with WBMD. In multivariate regression, BMI, circulating sclerostin and whole-body lean mass together accounted for 13.8% of the variation with WBMD in the older adults. In young, DKK1 and BMI together accounted for 7.7% of variation in WBMD.

Circulating factors associated with whole-body BMD

Five out of the six circulating factors differed in concentration between old and young (Table 2). Of those, DKK1, OC, OPG and sclerostin were identified from both partial correlation

Table 3 Associations between circulating bone regulatory factors and whole-body bone mineral density

Correlation with z-score WBMD Adjustment models	All participants combined		Old		Young	
	1	2	1	2	1	2
DKK1	**r = 0.107**	**r = 0.129**	**r = 0.167**	**r = 0.174**	**r = 0.263**	**r = 0.282**
	p = 0.026	**p = 0.008**	**p = 0.007**	**p = 0.005**	**p = 0.001**	**p < 0.0005**
FGF-23	r = 0.067	r = 0.051	r = −0.095	r = −0.079	r = −0.086	r = −0.130
	p = 0.274	p = 0.406	p = 0.235	p = 0.330	p = 0.370	p = 0.182
OC	**r = −0.124**	r = −0.083	**r = 0.150**	**r = 0.187**	r = −0.023	r = −0.008
	p = 0.010	p = 0.088	**p = 0.015**	**p = 0.003**	p = 0.767	p = 0.916
OPG	**r = −0.096**	r = −0.039	**r = 0.209**	**r = 0.254**	r = 0.081	r = 0.055
	p = 0.047	p = 0.419	**p = 0.001**	**p < 0.0005**	p = 0.297	p = 0.484
OPN	r = −0.005	r = −0.001	r = 0.055	r = 0.073	r = −0.120	r = −0.122
	p = 0.918	p = 0.980	p = 0.370	p = 0.245	p = 0.124	p = 0.120
Sclerostin	r = −0.091	r = −0.075	**r = 0.241**	**r = 0.240**	r = 0.129	r = 0.135
	p = 0.059	p = 0.126	**p < 0.0005**	**p < 0.0005**	p = 0.096	p = 0.086

Data are shown as Spearman's rho. The circulating bone regulatory factors were log-transformed and their z-scores calculated. The p value indicates the level of significance after statistical analysis. Results were adjusted for (1) country; (2) country, z-score lean mass, z-score BMI, self-reported health and supplement use. Significant relationships are highlighted using bold text

models as associated with WBMD in older participants (Table 3).

Sclerostin and DKK1 are released primarily by osteocytes and inhibit bone formation by blocking the osteoblast Wnt/β-catenin signalling pathway [4, 10], with sclerostin and DKK1 also stimulating bone resorption through RANKL [11]. Down-regulation of sclerostin [6] and DKK1 [4, 6] is associated with markedly increased bone formation. For these reasons, an inverse association between circulating sclerostin and DKK1 with WBMD would be expected, but is not entirely what was observed. In line with expectations, our results revealed, on average, a 1.8-fold higher circulating sclerostin and approximately 1.2-fold higher DKK1 in old compared with young, which is consistent with an inverse association between sclerostin and BMD in older age [12] and with results from a small sample of 36 patients showing an inverse association between DKK1 and lumbar and femur BMD [13]. However, contrary to expectations, the circulating levels of sclerostin and DKK1 were positively associated with WBMD in the older participants (Table 3). Similar positive associations between circulating sclerostin with BMD and bone micro-architecture in old age have been previously reported [14–17].

Similar to the findings for sclerostin and DKK1, a paradoxical relationship existed for OPG and WBMD in older adults: we found higher circulating OPG in old compared with young (Table 2), but circulating OPG was positively associated with WBMD (Table 3). OPG released by osteocytes and osteoblasts promotes bone formation. It has been shown to protect against generalised bone resorption by blocking tumour necrosis factor alpha, (TNFα), in models of chronic inflammation [18] and is considered to be a decoy receptor for RANKL to reduce

osteoclast-driven bone resorption [19]. There are conflicting reports about the direction of association between circulating OPG and BMD. A study of postmenopausal women of mean age 62 years [20] and a study of middle-aged men [21] reported inverse relationships between BMD and OPG, while others reported no relationship [22, 23]. Conversely, and in line with the results of the present work, when adults in their eighth and ninth decades of life were included in the sample population, the relationship between OPG and BMD was positive [24, 25]. These conflicting results cannot be explained by the differences between studies in skeletal site examined. Conflicting results may be related to the differences in the age range of the study samples and possible gender differences. Our results for OPG and sclerostin showed significant age × gender interactions indicating that the differences between young and older men were greater than those between young and older women (Table 2). It is already known that sex hormones can regulate bone turnover and may interact with these circulating factors [26].

It is not clear why circulating sclerostin, DKK1 and OPG were positively associated with WBMD in older age, despite the conflicting overall trend for higher circulating levels and lower WBMD in the old. One possibility is that the older but healthy mature osteocytes generally release higher absolute levels of sclerostin, DKK1 and OPG into the circulation [27, 28]. For example, a positive correlation was found for circulating sclerostin with trabecular density, number and thickness in older men [14, 27], suggesting the more advanced trabecular resorption in osteoporotic bone leaves fewer mature osteocytes and, thus, lower sclerostin release than healthy older bone. However, analysis of bone biopsies showed similar sclerostin mRNA levels in young and old despite higher

circulating sclerostin levels in the old [12] which indicates that the age-related differences in circulating sclerostin may not be due to increased osteocyte sclerostin gene expression, although this does not necessarily equal protein production [29].

Lower circulating OC was found in old compared with young (Table 2) and, consistent with this, circulating OC levels were positively correlated with WBMD in the old (Table 3). OC released by osteoblasts plays a role in bone formation, so the positive correlation with WBMD may be expected. However, others suggest that higher circulating OC indicates greater rates of bone resorption because fragments or whole OC protein is released into the circulation during bone resorption [3]. A previous study of young and middle-aged women suggested that circulating levels peaked soon after menopause and dropped thereafter, although levels were higher in those with osteoporosis than those without [30]. Interestingly, our results also showed a positive association between DKK1 and WBMD in the young adults from univariate and multivariate analyses. This association may be a reflection of the numbers of mature osteocytes or related to total bone mass, but more work is needed to confirm. One previous study of children and adolescents did not find any association between circulating DKK1 and BMD, but the young included in that study of youths were in stages of rapid developmental growth, which could present different results from the steady state of young adults [31].

Strengths and limitations

The MYOAGE study included young and older participants relatively free from lifestyle-related comorbidities for their age, and the results are therefore indicative of age-related effects. Nevertheless, the associations identified in this cross-sectional study cannot be interpreted as causal relationships despite the clear roles for the selected circulating markers in bone remodelling. The results for FGF23 showed no significant age or gender differences, nor correlations with WBMD, but a large proportion of the results were below the level of assay sensitivity, so firm conclusions cannot be drawn for this analyte. We have measured the circulating levels of markers, which may be influenced by release from non-bone cells, so it is not possible to determine the originating cell type. It is possible that altered renal function can affect the levels of the circulating factors, but markers of renal function was not included in the present study due to limitation of plasma sample quantity. A phantom was not used to calibrate the DXA scanners across sites, and we did not adjust the results to derive 'standardised' DXA values, as others have done for hip and femur sites [32]. Instead, all study centres followed the local quality control procedures, including use of phantoms and daily calibration, and the results were adjusted for country of testing to account for possible systematic differences.

Future studies should determine the reasons for the positive relationship between circulating sclerostin, DKK1 and OPG with BMD in older adults, despite the old having on average higher circulating levels of these factors and lower WBMD.

Conclusion

Sclerostin, DKK1, OPG and OC were each positively associated with WBMD in older adults, despite the average WBMD being lower and circulating DKK1, OPG and sclerostin being higher in old than young. Multiple linear regression identified BMI, circulating sclerostin and whole-body lean mass as explaining approximately 14% of all variation on WBMD amongst older adults.

Compliance with ethical standards

Conflicts of interest None.

References

1. Genant HK et al (1999) Interim report and recommendations of the World Health Organization task-force for osteoporosis. Osteoporos Int 10(4):259–264
2. Hofbauer LC et al (1999) Estrogen stimulates gene expression and protein production of osteoprotegerin in human osteoblastic cells. Endocrinology 140(9):4367–4370
3. Ferron M et al (2010) Insulin signaling in osteoblasts integrates bone remodeling and energy metabolism. Cell 142(2):296–308
4. Kamiya N (2012) The role of BMPs in bone anabolism and their potential targets SOST and DKK1. Curr Mol Pharmacol 5(2):153–163
5. Gaudio A et al (2010) Increased sclerostin serum levels associated with bone formation and resorption markers in patients with immobilization-induced bone loss. J Clin Endocrinol Metab 95(5):2248–2253
6. Ke HZ et al (2012) Sclerostin and Dickkopf-1 as therapeutic targets in bone diseases. Endocr rev 33(5):747–783
7. Takei Y, Minamizaki T, Yoshiko Y (2015) Functional diversity of fibroblast growth factors in bone formation. Int J Endocrinol 2015: 729352
8. Standal T, Borset M, Sundan A (2004) Role of osteopontin in adhesion, migration, cell survival and bone remodeling. Exp Oncol 26(3):179–184
9. McPhee JS et al (2013) Physiological and functional evaluation of healthy young and older men and women: design of the European MyoAge study. Biogerontology 14(3):325–337
10. Monroe DG et al (2012) Update on Wnt signaling in bone cell biology and bone disease. Gene 492(1):1–18
11. Brunetti G et al (2016) Impaired bone remodeling in children with osteogenesis imperfecta treated and untreated with bisphosphonates: the role of DKK1, RANKL, and TNF-alpha. Osteoporos Int 27(7):2355–2365

12. Roforth MM et al (2014) Effects of age on bone mRNA levels of sclerostin and other genes relevant to bone metabolism in humans. Bone 59:1–6

13. Butler JS et al (2011) The role of Dkk1 in bone mass regulation: correlating serum Dkk1 expression with bone mineral density. J Orthop res 29(3):414–418

14. Szulc P et al (2013) Correlates of bone microarchitectural parameters and serum sclerostin levels in men: the STRAMBO study. J Bone Miner res 28(8):1760–1770

15. Garnero P et al (2013) Association of serum sclerostin with bone mineral density, bone turnover, steroid and parathyroid hormones, and fracture risk in postmenopausal women: the OFELY study. Osteoporos Int 24(2):489–494

16. Polyzos SA et al (2012) Serum sclerostin levels positively correlate with lumbar spinal bone mineral density in postmenopausal women—the six-month effect of risedronate and teriparatide. Osteoporos Int 23(3):1171–1176

17. Thorson S et al (2013) Sclerostin and bone strength in women in their 10th decade of life. J Bone Miner res 28(9):2008–2016

18. Schett G et al (2003) Osteoprotegerin protects against generalized bone loss in tumor necrosis factor-transgenic mice. Arthritis Rheum 48(7):2042–2051

19. Weitzmann MN (2013) The role of inflammatory cytokines, the RANKL/OPG axis, and the immunoskeletal interface in physiological bone turnover and osteoporosis. Scientifica (Cairo) 2013: 125705

20. Jabbar S et al (2011) Osteoprotegerin, RANKL and bone turnover in postmenopausal osteoporosis. J Clin Pathol 64(4):354–357

21. Oh KW et al (2005) Circulating osteoprotegerin and receptor activator of NF-kappaB ligand system are associated with bone metabolism in middle-aged males. Clin Endocrinol 62(1):92–98

22. Rogers A et al (2002) Circulating estradiol and osteoprotegerin as determinants of bone turnover and bone density in postmenopausal women. J Clin Endocrinol Metab 87(10):4470–4475

23. Liu JM et al (2005) Relationships between the changes of serum levels of OPG and RANKL with age, menopause, bone biochemical markers and bone mineral density in Chinese women aged 20–75. Calcif Tissue Int 76(1):1–6

24. Samelson EJ et al (2008) Increased plasma osteoprotegerin concentrations are associated with indices of bone strength of the hip. J Clin Endocrinol Metab 93(5):1789–1795

25. Stern A et al (2007) The sex-specific association of serum osteoprotegerin and receptor activator of nuclear factor kappaB legend with bone mineral density in older adults: the Rancho Bernardo study. Eur J Endocrinol 156(5):555–562

26. Modder UI et al (2011) Regulation of circulating sclerostin levels by sex steroids in women and in men. J Bone Miner res 26(1):27–34

27. Modder UI et al (2011) Relation of age, gender, and bone mass to circulating sclerostin levels in women and men. J Bone Miner res 26(2):373–379

28. Mazziotti G et al (2006) Increased serum osteoprotegerin values in long-lived subjects: different effects of inflammation and bone metabolism. Eur J Endocrinol 154(3):373–377

29. Moester MJ et al (2010) Sclerostin: current knowledge and future perspectives. Calcif Tissue Int 87(2):99–107

30. Atalay S et al (2012) Diagnostic utility of osteocalcin, undercarboxylated osteocalcin, and alkaline phosphatase for osteoporosis in premenopausal and postmenopausal women. Ann lab med 32(1):23–30

31. Mora S et al (2015) Sclerostin and DKK-1: two important regulators of bone metabolism in HIV-infected youths. Endocrine 49(3): 783–790

32. Fan B et al (2010) Does standardized BMD still remove differences between Hologic and GE-Lunar state-of-the-art DXA systems? Osteoporos Int 21(7):1227–1236