


ORIGINAL



Plasma angiotensin-converting enzyme 2 as a potential causal marker in sepsis-associated ARDS development: evidence from Mendelian randomization and mediation analysis

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Abstract

Purpose: A causal biomarker for acute respiratory distress syndrome (ARDS) could fuel precision therapy options. Plasma angiotensin-converting enzyme 2 (ANG2), a vascular permeability marker, is a strong candidate on the basis of experimental and observational evidence. We used genetic causal inference methods—Mendelian randomization and mediation—to infer potential effects of plasma ANG2.

Methods: We genotyped 703 septic subjects, measured ICU admission plasma ANG2, and performed a quantitative trait loci (QTL) analysis to determine variants in the *ANGPT2* gene associated with plasma ANG2 ($p < 0.005$). We then used linear regression and post-estimation analysis to genetically predict plasma ANG2 and tested genetically predicted ANG2 for ARDS association using logistic regression. We estimated the proportion of the genetic effect explained by plasma ANG2 using mediation analysis.

Results: Plasma ANG2 was strongly associated with ARDS (OR 1.59 (95% CI 1.35, 1.88) per log). Five *ANGPT2* variants were associated with ANG2 in European ancestry subjects ($n = 404$). Rs2442608C, the most extreme *cis* QTL (coefficient 0.22, 95% CI 0.09–0.36, $p = 0.001$), was associated with higher ARDS risk: adjusted OR 1.38 (95% CI 1.01, 1.87), $p = 0.042$. No significant QTL were identified in African ancestry subjects. Genetically predicted plasma ANG2 was associated with ARDS risk: adjusted OR 2.25 (95% CI 1.06–4.78), $p = 0.035$. Plasma ANG2 mediated 34% of the rs2442608C-related ARDS risk.

Conclusions: In septic European ancestry subjects, the strongest ANG2-determining *ANGPT2* genetic variant is associated with higher ARDS risk. Plasma ANG2 may be a causal factor in ARDS development. Strategies to reduce plasma ANG2 warrant testing to prevent or treat sepsis-associated ARDS.

Keywords: Respiratory distress syndrome, adult, Angiotensin-2, Mendelian randomization analysis, Mediation analysis

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Introduction

Acute respiratory distress syndrome (ARDS) is characterized by the failure of the alveolar-capillary barrier resulting in non-cardiogenic pulmonary edema and life-threatening hypoxemia [1, 2]. Although substantial progress has been made in understanding ARDS pathophysiology, therapy for ARDS remains limited, with no consistently proven pharmacologic options for prevention or treatment. Drug development in ARDS has been hindered by many factors, including the clinical and biologic heterogeneity of the syndrome and the lack of biologically defined endotypes [3, 4]. Further limiting pharmacologic breakthroughs for ARDS is the lack of a validated intermediate phenotype or biomarker with a proven causal role in ARDS that could be used to improve clinical trial efficiency.

Genetic tools may help to identify which biomarkers contribute to a clinical outcome, disentangling correlation from causation [5, 6]. For example, genetic variants regulating plasma low density lipoprotein (LDL) levels are also associated with cardiovascular disease (CVD) risk, and the risk is statistically mediated through plasma LDL levels [5, 7]. Hence, pharmacologic targeting of plasma LDL is a major therapeutic goal for CVD. The analyses suggesting causality, termed Mendelian randomization (MR) studies, are a way to infer causality from observational data, considering each individual as “randomized” during gametogenesis to a high- or low-expressing genotype [8–11]. The MR framework is an adaptation of an instrumental variable analysis that tests an observed association while controlling for threats to its internal validity, including confounding variables, measurement error, spuriousness, simultaneity, and reverse causality [12]. The MR framework is increasingly used to assess intermediate traits and is appropriate when a potential causal intermediate is genetically predictable and associated with outcome [8, 12, 13]. Genotype then acts as an instrument to predict the plasma marker to be assessed, with the assumption that if the genetically predicted portion of the marker retains association with outcome then the measured marker has a causal true effect on the outcome.

Plasma angiotensin-2 (ANG2) is an established biomarker of endothelial activation and permeability that is strongly associated with ARDS risk and outcome [14–17] and thus a potential causal intermediate. Supporting its causal potential, mice genetically deficient in *ANGPT2* are more resistant to hyperoxia, exogenous ANG2 placed on endothelial monolayers disrupts barrier integrity, and exogenous ANG2 potentiates lung injury in animal models [18–21]. However, determining whether plasma ANG2 contributes causally to human ARDS risk has

Take-home message

Using a genetic technique that uses genotypes as an instrument to predict plasma angiotensin-2, we demonstrate that genetically predicted plasma angiotensin-2 is associated with sepsis-associated ARDS risk and thus infer that plasma angiotensin-2 may play a causal role in ARDS development. We apply mediation analysis to determine that plasma ANG2 mediates a significant portion (34%) of the association between polymorphisms in *ANGPT2* and ARDS. To our knowledge, this is the first application of a technique known as Mendelian randomization analysis to investigate plasma biomarkers in ARDS.

been limited by potential unmeasured confounding of plasma ANG2 and the possibility that elevated plasma ANG2 concentration may result from, rather than cause, lung injury. We and others have replicated an association between genetic variation in the angiotensin-2 gene (*ANGPT2*) and ARDS risk [22, 23]. We hypothesized that plasma ANG2 acts as a causal intermediate in determining *ANGPT2*-associated ARDS risk during sepsis. We focused on sepsis (pulmonary and non-pulmonary) because it is the most common cause of ARDS and carries a higher mortality than other causes [24, 25]. We used the MR framework in a prospective cohort of subjects with sepsis and we estimated the proportion of the genetic effect on ARDS risk mediated through plasma ANG2. Given the established association between *ANGPT2* variation and ARDS risk [22, 23], we generated a genetic instrument using multiple *ANGPT2* variants to predict plasma ANG2 in ancestry-specific populations. If plasma ANG2 has a causal role in ARDS risk due to sepsis, then reducing plasma ANG2 or inhibiting its signaling warrants testing for ARDS prevention and treatment.

Methods

The Molecular Epidemiology of Sepsis in the ICU (MESSI) cohort at the University of Pennsylvania has been described previously [26, 27]. Patients were eligible if they were admitted to the intensive care unit (ICU) with infection-associated organ failure [28, 29] and excluded if an alternative diagnosis explained SIRS criteria, for declining life support on admission, or for lack of informed consent. Whole blood was collected for DNA and plasma was collected within 24 h of ICU admission, as close to the time of ICU arrival as possible. Clinical data were abstracted from the electronic medical record. All chest imaging studies obtained during the first 6 days [30] were interpreted by trained physician investigators as described [31, 32]. ARDS was adjudicated in accordance with Berlin criteria requiring that chest radiograph and oxygenation criteria be met on the same calendar day while invasively ventilated [1, 27]. Mortality was determined at 30 days. Source of sepsis was adjudicated

by critical care physician investigators. As a replication sample, the iSPAAR consortium study consisted of European-ancestry genotyped subjects whose ARDS risk factor was either sepsis or pneumonia and whose plasma was assayed for ANG2 [33–36]. We used the GTEx Portal to search individual single nucleotide polymorphisms (SNPs) for expression quantitative trait locus (eQTL) significance in three tissues: lung, aorta, and tibial artery. Further details are provided in the online supplement.

MESSI assay procedures

Day 0 plasma ANG2 was measured by an enzyme-linked immunosorbent assay optimized for human plasma (ELISA; R&D Systems, Minneapolis MN). Genomic DNA was extracted from whole blood using the QIAamp DNA Mini kit (Qiagen, Hilden Germany) and assayed with the Affymetrix Axiom TxArray v.1, a genome-wide platform comprising approximately 780,000 markers, of which 184 are within 70 kilobases (kb) of *ANGPT2* [37].

Statistical analysis

Continuous data were compared using nonparametric methods and categorical data by chi-square test. The association between log(plasma ANG2) and ARDS was tested by multivariable logistic regression adjusting for APACHE III score and pulmonary source of infection [4]. Using all markers on the genotyping platform, we performed multidimensional scaling to identify four principal components, allowing for identification of genetic ancestry (Supplemental Fig. E2) [22, 32]. Plasma ANG2 values were log-transformed for normality as a result of a positive skew, and we determined the association between genotypes and logANG2 using linear regression, assuming an additive model of genetic risk. Models were performed separately for genetically European (EA) and African ancestry (AA) subjects. Subjects of other ancestry were excluded because of low numbers. We limited our search for genetic determinants of plasma ANG2 to variants within 70 kb of the *ANGPT2* gene to find *cis* regulators given our prior replicated association between *ANGPT2* intron 2 and ARDS [22, 38]. SNPs demonstrating an association with plasma ANG2 at p values less than 0.005 were considered significant since *ANGPT2* has fewer than 10 linkage disequilibrium blocks [38]. ANG2-associated SNPs were then tested for an association with ARDS using multivariable logistic regression adjusting for potential confounders of the ARDS–ANG2 association, including APACHE III score, pulmonary (versus non-pulmonary) sepsis [4], and genetic ancestry; please see Supplemental Methods for further justification of covariates. To infer a causal association of plasma ANG2 with ARDS, we next regressed transformed ANG2 levels on the ANG2-associated SNPs and used post-estimation

prediction to generate a genetically predicted plasma ANG2 value for each EA subject. Genetically predicted ANG2 values were then tested for an association with ARDS risk using multivariable logistic regression. To ensure that our genetic instrument was truly associated with plasma ANG2, we tested *ANGPT2* SNPs for replication of the plasma ANG2—*ANGPT2* association in the iSPAAR dataset and tested replicating SNPs in the GTEx Project databank. To test whether plasma ANG2 concentration mediated a significant portion of the association between *ANGPT2* SNPs and ARDS, we undertook mediation analysis. This technique is a formal approach to explain the mechanism by which an explanatory variable (SNP) influences the outcome (ARDS) via an intermediate or mediator variable (plasma ANG2) [39, 40]. We used linear regression of SNP on logANG2 to estimate the change in plasma ANG2 per allele, and logistic regression of SNP, logANG2, an interaction term [SNP²logANG2], pulmonary source, APACHE III score, and genetic ancestry to model the total effect of SNP on ARDS [39, 40]. The online data supplement describes the mediation effect in more detail along with a complementary MR framework analysis and a sensitivity analysis. We used R statistical packages for the principal components and mediation analyses, Plink for the QTL analysis, and Stata 15 (College Station, TX) for all other analyses. We estimated through simulations that with 250 subjects per ancestry we would have at least 80% power to detect ARDS odds ratios of at least 1.5 if we could explain approximately 10% of the variance in plasma ANG2 [41].

Results

The MESSI population is depicted in Table 1. Between September 5, 2008 and February 9, 2015, 9265 intensive care unit (ICU) patients were screened, 2163 were identified as having sepsis, and 1263 were enrolled (Fig. E3). Of enrolled subjects, 703 had available DNA and plasma ANG2 measured. Reasons for a lack of DNA included inadequate DNA quantity, poor DNA quality, or failure to collect the DNA sample. The primary reason for a lack of plasma measurement was that the sample was not obtained within the 24-h time period immediately following ICU admission, including for subjects transferred from another facility. As shown in Table 1, patients who developed ARDS were more likely to be of European ancestry (EA), had higher severity of illness at presentation, and were more likely to have a pulmonary source of sepsis. Pneumonia was a major risk factor for ARDS, with an odds ratio of 2.63 (95% CI 1.75, 3.95), $p < 0.001$, compared to non-pulmonary sepsis. Overall mortality was high at 50% in this observational cohort, and ARDS subjects had a significantly higher mortality than non-ARDS subjects. Measured plasma ANG2 was strongly

Table 1 MESSI population with genotype and plasma ANG2 measured

	ARDS (n = 289)	Non-ARDS (n = 414)	p value
Age	60 (51–69)	62 (52–71)	0.095
Female	119 (41.2%)	181 (43.7%)	0.502
European ancestry	178 (61.6%)	226 (54.6%)	0.022
African ancestry	91 (31.5%)	163 (39.4%)	
Asian ancestry	13 (4.8%)	16 (3.9%)	
Genetically admixed	7 (2.4%)	9 (2.2%)	
APACHE III	84 (69–105)	71 (57–88)	<0.001
Infectious source (primary)			
Pulmonary	172 (60%)	137 (33%)	<0.001
Genitourinary	16 (6%)	71 (17%)	
Abdominal	38 (13%)	63 (15%)	
Blood	12 (4%)	44 (11%)	
Unknown	35 (12%)	67 (16%)	
Comorbidities			
Immunocompromised	142 (49%)	190 (46%)	0.397
Malignancy	108 (37%)	141 (34%)	0.366
Cirrhosis, decompensated	23 (8%)	13 (3%)	0.004
Diabetes	76 (26%)	139 (34%)	0.043
SOFA score day 0	5 (3–7)	3 (1–5)	<0.001
PaO ₂ /FiO ₂ ratio day 0; number, % with ABG on day 0	122 (76–190); n = 222, 77%	215 (117–323); n = 119, 29%	<0.001
Intubated on day 0	192 (66%)	69 (17%)	<0.001
Tidal volume per kg PBW day 0	7.1 (6.4–8.1)	7.3 (6.8–8.6)	0.213
PEEP (cmH ₂ O) day 0	5 (5–8)	5 (5–5)	0.012
Plateau pressure (cmH ₂ O) day 0	23 (18–30)	20 (17–24)	0.049
Vasoactive support day 0	181 (63%)	159 (39%)	<0.001
Measured ANG2 ng/ml	10.0 (5.2–21.6)	7.9 (3.9–16.0)	<0.001
Mortality, 30 day	197 (68.2%)	157 (38.0%)	<0.001

Continuous data are compared by the Wilcoxon ranksum test and categorical data by a chi-square test

SOFA sequential organ failure assessment score, ABG arterial blood gas, PBW predicted body weight, PEEP positive end-expiratory pressure

associated with ARDS independent of APACHE III score or pulmonary source of infection. For each log increase in plasma ANG2 measurements, the adjusted odds ratio for ARDS was 1.49 (95% CI 1.20, 1.77), $p < 0.001$. The association of plasma ANG2 with ARDS was significant in both EA and AA subgroups (Table E2).

Results of the *cis*-QTL analysis for EA ($n = 404$) and AA ($n = 254$) subjects are shown graphically in Fig. 1 and Fig. E4, respectively. Forty-five subjects were of Asian ancestry or were genetically admixed and were excluded from the MR analysis. Genetic analysis of EA subjects revealed strong *cis* regulation, whereas analysis of AA subjects did not. In EAs, five SNPs were associated with ANG2 at $p < 0.005$ (Table 2) and these SNPs demonstrated low linkage disequilibrium with one another ($r^2 < 0.10$ for all but one, $r^2 = 0.24$ with opposite directionality) (Fig. 1a) [42, 43]. Individually, each SNP explained approximately 2% of the variance in plasma ANG2 levels (R^2), whereas collectively they explained 8.1% variance.

We tested each SNP for an additive association with ARDS risk, adjusting for genetic ancestry, pulmonary source of infection, and severity of illness (APACHE III score). Two SNPs—rs2442608 and rs2442630—demonstrated a significant ARDS association (Table 2). In addition, rs2442608 has moderate linkage ($r^2 = 0.37$) with the locus we previously identified as associated with trauma-associated ARDS risk, Fig. 1b [22], providing additional replication for this locus now in sepsis-associated ARDS.

For the MR analysis, we genetically predicted plasma ANG2 using post-estimation analysis from linear regression models of QTL on measured plasma ANG2 (Fig. 2). Of 404 EA subjects, 16 were missing a genotype call on one or more of the five QTL and thus did not have a predicted value, leaving 388 EA subjects for the analysis. Genetically predicted ANG2 values were associated with ARDS, adjusting for genetic ancestry, pulmonary source of infection, and APACHE III score (Fig. 2), adjusted odds ratio 2.25 (95% CI 1.06, 4.78), $p = 0.035$. Furthermore, a

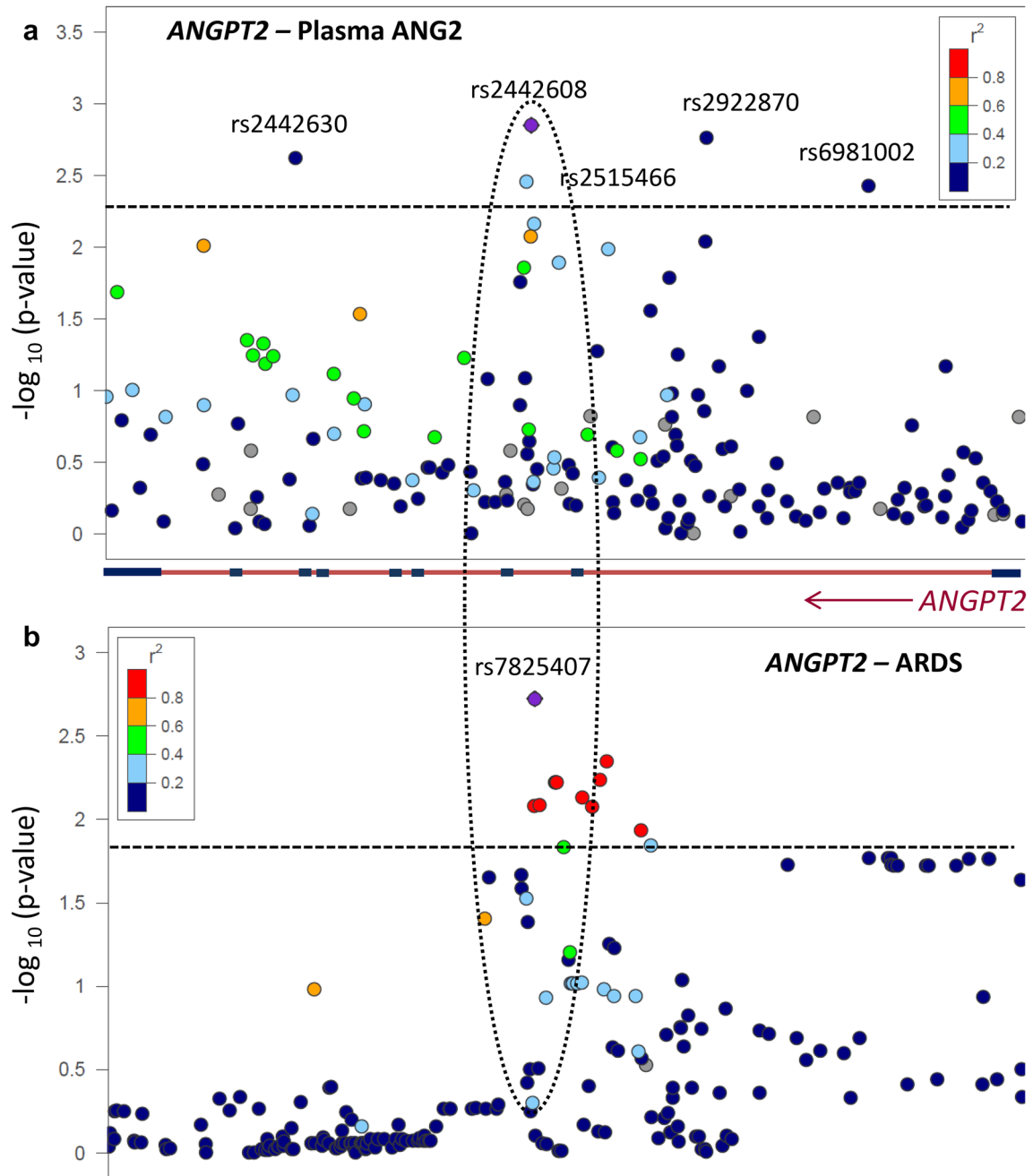
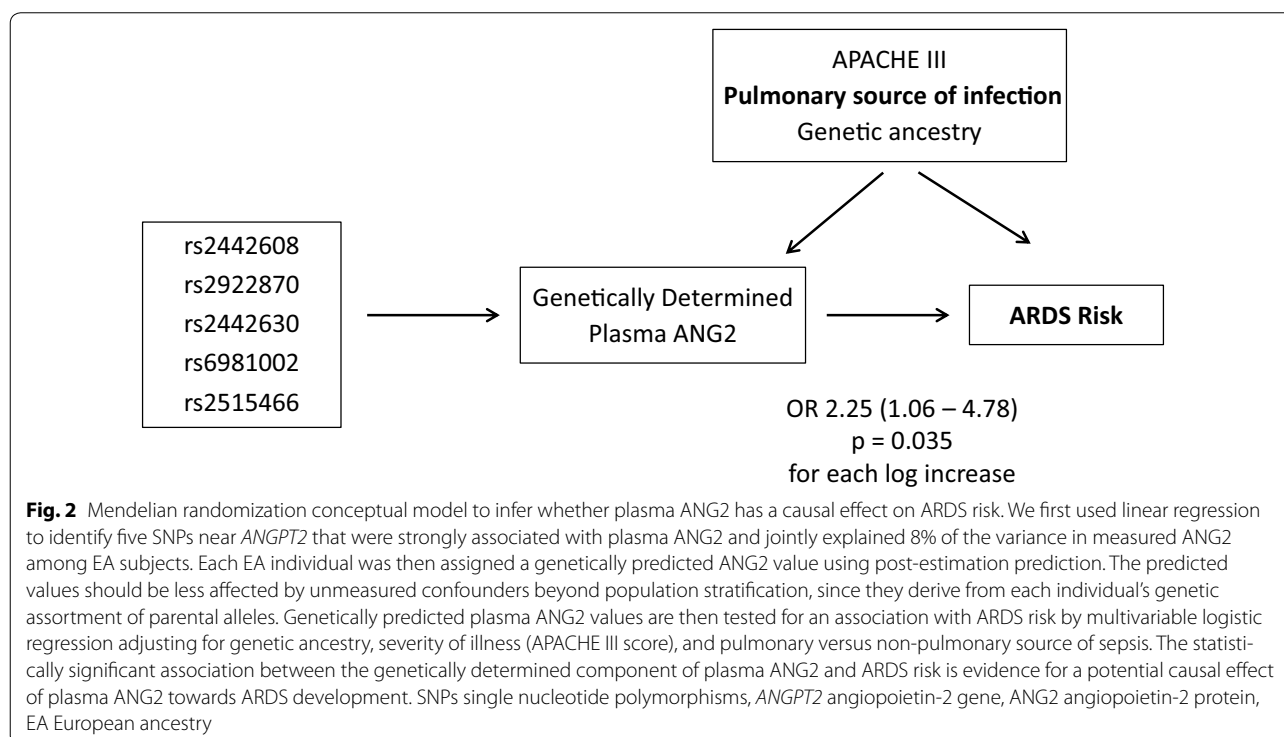


Fig. 1 Regional association plots demonstrate a consistent region of association between the *ANGPT2* gene and plasma ANG2 (**a**) and previously reported trauma-associated ARDS (**b**). **a** Depicts the regional association plot between loci on the *ANGPT2* gene (chromosome 8) on the x-axis and the strength of association with plasma ANG2 in early sepsis among EA shown as $-\log(p$ value for ANG2 association) on the y-axis. Single nucleotide polymorphisms (SNPs) with association more extreme than $p=0.005$ are labeled, with rs2442608 being the most extreme SNP from the QTL analysis. Color-coding depicts the strength of linkage disequilibrium (LD) between rs2442608 and other loci, with increasing LD represented by increasing red. Navy represents minimal LD with rs2442608. A schematic of the *ANGPT2* gene is depicted as a red bar with blue exons and an arrow to indicate the direction of transcription. **b** Depicts the regional association plot between the same region of chromosome 8 with trauma-associated ARDS as reported in our prior study [22]. The most extreme SNP from that association, rs7825407, is in moderate LD with rs2442608 ($r^2 = 0.37$) in EA populations [42], replicating the importance of this *ANGPT2* intron for ARDS and providing functional relevance for this locus. Plots were created using LocusZoom [43]

Table 2 Five unlinked SNPs are *cis* quantitative trait loci (QTL) for plasma ANG2 expression during sepsis in Europeans, and two demonstrate significant ARDS association

SNP	QTL coefficient (95% CI)	<i>p</i> value	ARDS MAF	Non ARDS MAF	Adjusted OR ARDS (95% CI)	<i>p</i> value
rs2442608G	0.22 (0.09, 0.36)	0.001	0.51	0.44	1.38 (1.01, 1.87)	0.042
rs2922870G	-0.21 (-0.34, -0.08)	0.002	0.42	0.41	1.02 (0.76, 1.36)	0.905
rs2442630G	0.76 (0.27, 1.25)	0.002	0.04	0.01	5.48 (1.68, 17.8)	0.005
rs2515466A	-0.22 (-0.36, -0.07)	0.003	0.27	0.31	0.80 (0.58, 1.11)	0.189
rs6981002C	-0.44 (-0.73, -0.14)	0.004	0.04	0.03	1.28 (0.60, 2.72)	0.525

ARDS analyses were adjusted for the first two components of genetic ancestry, APACHE III score, and pulmonary (vs non-pulmonary) source of infection
MAF minor allele frequency, *QTL* quantitative trait loci, *CI* confidence interval



complementary method of MR analysis known as two-stage residual inclusion, described in the online data supplement, also identified a potential causal role for plasma ANG2: OR 2.65 (1.22, 6.05) $p=0.047$.

Because instrumental variable methodology relies upon valid instruments, we sought to replicate the association between plasma ANG2 during sepsis and SNPs in the *ANGPT2* gene in an independent dataset (Table E3). Two of the five SNPs were directly genotyped in the replication population and replicated their association in patient-level meta-analysis (Table E4 and Fig. E5). Both SNPs also exhibited differential expression of *ANGPT2* in one or more relevant tissue, with rs2442608 differentially expressed in lung ($p=0.0060$), aorta ($p=0.048$), and

tibial artery ($p=0.00090$) and rs2515466 differentially expressed in tibial artery ($p=0.0051$) (Table E5).

In mediation analysis, we determined the total effect and the mediation effect for the two replicating *ANGPT2* *cis* QTL (Table 3). For both rs2442608 and rs2515466, the mediation effect was significant and the proportion of ARDS risk explained was greater than 30%, whereas no significant "direct" effect between each SNP independent of plasma ANG2 was detected.

Discussion

We have demonstrated that the genetically determined portion of plasma ANG2 is associated with ARDS risk due to sepsis, suggesting that plasma ANG2 may serve as a causal intermediate phenotype in ARDS. Furthermore,

Table 3 Causal mediation analysis demonstrates that 30–40% of the ARDS risk is mediated through changes in plasma ANG2 for replicating *ANGPT2* SNPs

SNP	Analysis	Adjusted OR ARDS (95% CI)	<i>p</i> value	Proportion mediated by plasma ANG2
rs2442608	Total effect	1.06 (0.99, 1.13)	0.08	
	Mediation effect	1.02 (1.01, 1.04)	0.01	0.34
	Direct effect	1.04 (0.98, 1.10)	0.24	
rs2515466	Total effect	0.95 (0.88, 1.02)	0.12	
	Mediation effect	0.98 (0.96, 0.99)	0.01	0.40
	Direct effect	0.97 (0.91, 1.04)	0.40	

Each effect shown reflects the average value of 1000 iterations. The “mediation effect” in bold reflects the indirect effect of each SNP on ARDS risk that is mediated through plasma ANG2, while the “direct effect” estimates the effect of each SNP on ARDS risk independent of plasma ANG2 variation [39]. Both replicating *ANGPT2* SNPs had significant effects mediated by plasma ANG2, without significant direct effects on ARDS

we provide evidence that a significant proportion, over 30%, of the genetic association between *ANGPT2* variants and ARDS risk is mediated by early plasma ANG2 concentration among septic European ancestry subjects. Thus, efforts to reduce plasma ANG2 or to block its signaling warrant testing to prevent and possibly treat ARDS in sepsis.

Interest in targeting the angiopoietin–TIE2 receptor axis as a potential ARDS strategy is not new, given strong evidence for this pathway’s contribution by in vitro, animal, and human studies. Mice that were genetically deficient in *ANGPT2* were more resistant to hyperoxia-induced inflammation, permeability, cell death, and mortality [19]. Patients with ARDS had significantly elevated plasma and edema fluid ANG2 compared to control patients with hydrostatic edema [19]. Parikh and colleagues demonstrated that sera from septic patients with high circulating ANG2 induced stress fiber formation and endothelial intercellular gaps when applied to an endothelial monolayer [20]. Gap formation was reversed with a competitive inhibitor of ANG2, recombinant human angiopoietin-1 [20], suggesting that circulating ANG2 protein is sufficient to cause vascular permeability in sepsis and ARDS. More recent work has established the prognostic and predictive significance of plasma ANG2 for ARDS in human populations [14, 15]. Numerous approaches to pharmacologically block ANG2’s effects exist and have shown promise for reducing vascular permeability in vitro and in vivo, in some cases improving survival [44–52]. Despite this strong experimental rationale, no therapy targeting the ANG2–TIE2 pathway has yet been tested in human sepsis or ARDS. Trials in both ARDS and sepsis are notable for the failure of many agents to translate to clinical benefit despite strong experimental evidence and evidence of clinical biomarker association, however [53, 54], suggesting that better methods of selecting lead candidates is warranted [55].

In this study, we used the principle of Mendelian randomization (MR) to infer a causal relationship of plasma ANG2 with ARDS in sepsis. Because parental alleles are randomly sorted during gametogenesis, *ANGPT2* genotypes can be considered randomly assigned, theoretically independent of confounders such as propensity for pneumonia or sepsis. We could genetically predict approximately 8% of the variance in plasma ANG2 using five loci close to *ANGPT2*. Similar to measured levels of plasma ANG2, genetically predicted plasma ANG2 was associated with adjusted ARDS risk, suggesting that early plasma ANG2 may contribute to ARDS risk. Because we have demonstrated that plasma ANG2 levels are heterogeneous early in sepsis and that genetically predicted plasma ANG2 concentrations influence ARDS risk, we suspect that the benefits of blocking ANG2 activity will not be uniform. A precision approach using plasma ANG2 to decide which patients should be enrolled in trials to test anti-ANG2 therapy may be superior to an approach whereby all patients are eligible [56], unless the safety profile of new agents favors testing the drug in all septic subjects.

Our work highlights the utility of quantitative traits to maximize power in a complex genetic trait such as ARDS [55, 57]. The regulation of plasma ANG2 is much less complex than the regulation of sepsis-associated ARDS, and the use of a genetic MR approach helps to prioritize plasma ANG2 as a marker that seems to contribute to ARDS risk [8]. To strengthen the validity of our genetic instrument, we used the iSPAAR population to confirm differential plasma expression and the GTEx Portal to confirm differential RNA expression for two SNPs in our instrument. Because the replication population tested plasma ANG2 at variable time points following ICU admission, it may not be surprising that the QTL analysis yielded slightly different results in this population. Nonetheless, the meta-analysis results strengthen the functional significance of our prior replicated locus [22].

Our results also highlight the importance of simulating stress conditions for studies of quantitative traits such as plasma or mRNA expression to reduce the complexity of a trait like ARDS [57–59], as the identified ANG2 QTL during early sepsis differ from the quiescent state [60]. Our mediation analysis suggests that over 30% of rs2442608- and rs2515466-associated ARDS risk is explained by plasma ANG2. We used a multi-SNP predicted ANG2 to explain a higher proportion of plasma ANG2 variance [61], and observed a significant association between genetically predicted plasma ANG2 and ARDS risk. However, our genetic instrument was relatively weak [61] and could have been stronger if we had used genome-wide markers for plasma ANG2 in the QTL analysis, rather than limiting to SNPs close in proximity to the *ANGPT2* gene. A genome-wide approach would have been necessary to test our hypothesis in the AA subgroup, as *cis* variants did not have a strong enough effect on plasma ANG2 in this underpowered subpopulation. However, our study rationale was to test whether prior associations between *ANGPT2* and ARDS risk were mediated by changes in plasma ANG2 concentration.

Our study has limitations. We used two methods of causal inference, instrumental variable and causal mediation, to infer a causal effect of plasma ANG2 on ARDS risk from observational data; however, neither method completely removes confounding and bias. Further, the assumptions of each methodology are somewhat in conflict: whereas Mendelian randomization specifies that the only path between SNP and ARDS travels through plasma ANG2, mediation analysis asks what proportion of the SNP–ARDS association travels through plasma ANG2, and acknowledges that some fraction of the association may be independent of plasma ANG2. However, the fact that each method detects a causal effect for plasma ANG2 is supportive of a true causal relationship [12]. Ultimately, further replication and an experimental approach to modify plasma ANG2 and observe reduced ARDS risk is necessary to prove the causality of human plasma ANG2, which would be consonant with animal and *in vitro* experiments implicating ANG2 as sufficient to provoke lung injury [19, 20]. Further, replication of the causal effect of plasma ANG2 in non-septic precipitants of ARDS such as trauma or inhalational injury is warranted.

Although our intent was to perform plasma ANG2 QTL analysis for both EA and AA subjects with sepsis, our power was stronger for the EA subpopulation because of enrollment trends and the strength of *cis*-QTL observed. Although we observed a significant association between plasma ANG2 and ARDS among AA subjects, none of the individual QTL identified in EA subjects was associated with plasma ANG2 AA subjects. Our

future studies will need to analyze a larger population of AA subjects to determine whether regulation of plasma ANG2 in *cis* is an ancestry-specific finding. Plasma ANG2 was tested at only one time point, and a repeated measures analysis might have captured a larger proportion of plasma ANG2 variance during critical illness. However, this early time point may be most useful to inform a possible biomarker-based approach to therapy. This was a single-center study, and the MESSI population is unique in enrolling over almost a decade, having a high observed mortality—we believe due to the high proportion of comorbidities for which APACHE III may poorly account [62]—and for having a high proportion of severe ARDS. Our phenotyping was consistent with the Berlin definition but did not characterize lung vasculature permeability directly [1, 63]. Finally, it remains unknown if subjects with *ANGPT2*-mediated high plasma ANG2 represent a distinct endotype of septic subjects at risk for ARDS, or whether they will respond differently to therapy [64]. With the appropriate test development and validation, plasma ANG2 could be readily assessed at the bedside, obviating the need to genotype subjects, and our findings suggest that therapies aimed at reducing ANG2 or ANG2 signaling in those with the highest circulating concentrations warrant testing.

In conclusion, we have (1) provided evidence that plasma ANG2 may have a causal role in increasing ARDS risk among European ancestry subjects; (2) demonstrated significant local (*cis*) genetic regulation of plasma ANG2 during early sepsis in EA subjects; and (3) replicated our prior association of the second intron in *ANGPT2* and ARDS in a septic population. Reducing the levels of plasma ANG2 or blocking its signaling should be tested for ARDS prevention and/or therapy, particularly among patients with high circulating concentrations of ANG2.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-018-5328-0>) contains supplementary material, which is available to authorized users.

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Author contributions

Dr. Meyer had access to all data and takes responsibility for the integrity of the work. JPR, JDC, RF, and NJM conceived of and designed the study. JPR, JDC, CSC, MAM, RF, DCC, MMW, and NJM obtained funding. JPR, FW, TKJ, JAP, BJA, MGS, TGD, EDJ, TRR, BL, JA, CAI, and NJM acquired data. JPR, FW, JAP, BJA, MGS, EC, XL, IB, CSC, MAM, JDC, RF, XL, IB, and NJM analyzed and interpreted the data. JPR and NJM drafted the manuscript. All authors made significant contributions to the final manuscript and approve its submission.

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