FOCUSED REVIEW

Selenium in Soil–Plant‑Microbe: A Review

Zhen Wang¹ · Wei Huang1 · Fei Pang1

Received: 11 March 2021 / Accepted: 8 September 2021 / Published online: 6 October 2021 © The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2021

Abstract

Selenium (Se) plays an important role in geochemistry and is an essential trace element for humans and animals. This review summarizes the transformation and accumulation of Se in the plant-soil-microbe system. As one of the important reservoirs of Se, soil is an important material basis of its entry into the food chain through plants. Soil with an appropriate amount of Se is benefcial for plant growth and plays a valuable role in a stress-resistant environment. Among the many migration and transformation pathways, the transformation of Se by microorganisms is particularly important and is the main form of Se transformation in the soil environment. In this review, the role and form transformation of Se in plants, soil, and microorganisms; the role of Se in plants; the form, input, and output of Se in soil; the absorption and transformation of Se by plants; and the role of microorganisms in Se transformation are presented. In addition to describing the migration and transformation laws of Se in the environment, this review expounds on the main directions and trends of Se research in the agricultural feld as well as current gaps and difculties in Se-related research. Overall, this reviews aims to provide necessary information and theoretical references for the development of Se-rich agriculture.

Keywords Se · Plant · Soil · Microbe · Function · Transformation

Selenium (Se) is a necessary trace element for human and animals that participates in many biological metabolic processes in the human body. Se defciency can cause or induce many diseases, such as Keshan disease, skeletal muscle necrosis, and cardio-cerebrovascular disease (Vinceti et al. [2018](#page-13-0)). Although Se is not necessarily a necessary nutrient for plant growth, plants are the main source of Se intake for humans and animals (Natasha et al. [2018](#page-12-0)). Dietary Se supplement is the most common and important route of Se entry into the body. Se content and its existing form in crops determine the amount of Se absorbed by humans through food (Ekumah et al. [2021](#page-10-0)). The Se absorption and transformation abilities of plants and the transfer and enrichment abilities of the plant itself to its edible portion determine the Se enrich-ment effect of the plant (Guignardi and Schiavon [2017](#page-11-0)). According to the standards of the World Health Organization (WHO) and the International Food and Agriculture

 \boxtimes Wei Huang hww908i@foxmail.com

 \boxtimes Fei Pang 876083759@qq.com Organization (FAO), the global population with severe Se deficiency (daily intake of $7-11 \mu$ g) reached 0.5–1 billion, and the population with Se defciency is markedly more than the population with excessive Se (Winkel et al. [2012\)](#page-13-1). In China alone, nearly 2/3 of the planting soil is Se-deficient, posing health risks to people that live in these areas owing to insufficient Se intake (Zhang et al. $2014b$). As in-depth research continues to be carried out, the important role of Se in humans has been discovered. Se can participate in the development of selenoprotein followed by glutathione peroxidase (GSH-Px), thioredoxin reductase (TrxR), iodothyroninedeiodinases, and a series of enzyme systems that display functions, such as anti-disease, anti-cancer, and inhibition of HIV development and other related immune system disorders (Ekumah et al. [2021](#page-10-0)).

Se can be found in diferent geographical features, such as the atmosphere, lithosphere, hydrosphere, and biosphere, and its basic fractions are afected by chemical processes (pH, redox properties, and organic matter content), physical processes (adsorption and deposition efects), and bio-logical processes (microbial effects) (Sharma et al. [2015](#page-13-2)). The form of Se found in the soil of natural Se-rich areas is mainly antimonselite, supplemented by selenate. The form of Se in ores cannot be directly absorbed and utilized by

 1 College of Biology and Pharmacy, Yulin Normal University, Yulin 537000, China

plants. Further, Se salts are produced by long-term leaching and weathering of Se ores, which are highly toxic and unevenly distributed (Deng et al. [2018\)](#page-10-1). The level of Se in plants is determined by the status of Se in the soil (Favorito et al. [2020\)](#page-10-2). The form of Se, especially its efective content, is key to determining its mobility and toxicity (Galić et al. [2021\)](#page-11-1). A series of environmental chemical reactions, such as (biological) oxidation–reduction, precipitation-dissolution, and adsorption–desorption, among others, occur between Se in the soil and soil components, such as iron and manganese oxidation, clay minerals, and organic matter (Wang et al. [2019\)](#page-13-3). An appropriate amount of Se can increase the photosynthetic pigment content in plant leaves under stress, increase the activity of antioxidant enzymes in leaves and fruits, improve the osmotic adjustment ability, increase the organic acid content, and assist in plant growth under stress (Morales-Espinoza et al. [2019](#page-12-1); Zahedi et al. [2019\)](#page-14-1). Se is cycled in soil mainly through the microbial pathway (Mehdi et al. [2013](#page-12-2)). Microorganisms serve as the decisive factor of Se transformation (especially Se reduction) in soil as they participate in the formation and transformation of Se in various valence states (Gómez-Gómez et al. [2019](#page-11-2); Fischer et al. [2020\)](#page-10-3). The circulation of Se in the crop growth environment is shown in Fig. [1.](#page-1-0)

Understanding the migration and role of Se in plant-soilmicroorganisms and clarifying the absorption of Se by plants are of particular importance. Moreover, understanding the process, infuencing factors, and physiological functions of Se absorption and transformation by plants based on agroecology as well as the law of microbial transformation of Se and the migration of Se in soil can better guide production practices. Based on current studies on crop ecology in the development of Se-rich functional agricultural systems, the migration law and role of Se in plants, soil, and microorganisms are summarized in this review. The information presented herein is expected to serve as basic information for future research on the absorption and transformation of Se in agro-ecology and its physiological mechanism as well as a reference and basis for the development of Se-rich functional agricultural systems.

Se has various effects in plants, including promoting plant growth, enhancing plant resistance, and improving crop quality, among others (Table [1\)](#page-2-0). An appropriate concentration of Se can promote the accumulation of starch in plant chloroplasts, which is beneficial to plant growth, as demonstrated in plant studies with lettuce (Rios et al. [2010\)](#page-12-3), chicory (Germ et al. [2007\)](#page-11-3), and potato (Turakainen et al. [2004](#page-13-4)). An appropriate amount of Se can increase the chlorophyll content in plants owing to the ability of Se to restore enzyme activity in the photoresponse and the electron transport chain of stressed plants (Diao et al. [2014](#page-10-4)). The addition of suitable concentration of Se could reduce the damage of chloroplast to some extent and increase the content of chlorophyll (Chu et al. [2010](#page-10-5); Yao et al. [2011;](#page-13-5) Malik et al. [2012\)](#page-12-4). Excessive Se application aggravates the damage to chloroplasts, which is not conducive to plant photosynthesis (Wang et al.

Fig. 1 The transfer and utilization of Se in the crop growth environment

Table 1 The role of Se in plant growth and physiology

Plant	Results after Se application	Mechanism	References
Wheat	Promote plant growth	Increase the dry weight of the above ground part and the yield of grains, and increase the con- centration of organic selenium in the grains	Xia et al. (2020)
Wheat	Enhance cold resistance	Improve the antioxidant capacity of seedlings	Chu et al. (2010)
Wheat	Reduce the damage of UV-B to wheat seedlings	Significant increase in biomass, chlorophyll content, antioxidant content, and antioxidant enzyme activity	Yao et al. (2011)
Wheat	Improve the salt tolerance of wheat	Upregulate the antioxidant system, enhance the synthesis of the permeate, and prevent exces- sive $Na+$ accumulation	Elkelish et al. (2019)
Wheat	Enhance the tolerance of wheat to NaCl stress	Improve antioxidant and sodium manipulation of AOX, SOS1, and NHX1 gene overexpression	Alla et al. (2020)
Tomato	Enhance resistance to salt stress	Regulate the antioxidant defense system in the chloroplast of tomato seedlings to reduce salt- induced oxidative stress	Diao et al. (2014)
Rice	Conducive to the production of rice with higher selenium content	Change the antioxidant enzyme activity and gas exchange in rice leaves	de Lima Lessa et al. (2019)
Rice	Reduce the adverse effects of salinity on the growth of rice plants	Enhance the antioxidant defense system and increase the transcription level of OsNHX 1	Subramanyam et al. (2019)
Rice	Reduce the toxicity of plant mercury	Inhibit the absorption and transport of mercury by plants	Zhang et al. (2012)
Rice	Change the energy metabolism of rice seedlings	Inhibit the growth of rice seedlings, stimulate the reactive oxygen species in rice seedlings, increase ethylene production, and reduce the auxin biosynthesis of rice seedlings	Malheiros et al. (2020)
Rice	Reduce the accumulation of Cd in rice	Increase soil pH, reduce Cd bioavailability, and inhibit Cd transport from roots to shoots	Huang et al. (2018)
Rape	Relieve cadmium toxicity	Increase the unsaturation of fatty acids in the cell Filek et al. (2010) and the fluidity of the cell membrane	
Potato	Positively affect production	Accumulate carbohydrates	Turakainen et al. (2004)
Pak choi	Maintain the stability of the microbial commu- nity in Cr-contaminated soil	Oxidation of Cr(VI) and reduction of Se reduc- tases proportions, and increase the soil pH	Cai et al. (2019)
Mungbean	Relieve cadmium toxicity	Limit arsenic intake and enhance antioxidant activity	Malik et al. (2012)
Microalgae	Enhance photosynthesis	Regulate the photochemical properties of photo- system II (PSII) in chloroplasts	Zhong and Cheng (2017)
Maize	Alleviate the salt stress in corn	Improve photosynthetic capacity and antioxidant enzyme activity, and regulate Na ⁺ homeostasis	Jiang et al. (2017)
Lettuce	Increase nitrogen metabolism	Increase nitrate reductase, nitrite reductase, glu- Rios et al. (2010) tamine synthetase, and glutamate synthase	
Chinese flowering cabbage	Reduce the toxicity of plant cadmium	Increase SOD, CAT, and aseorbateperoxidase levels in plant tissues, and selectively induce glutathione reductase and dehydroascorbate reductase in plant tissues	Wu et al. (2018)
Chicory	Increase the respiratory potential in young plants	Exert a positive role on the photochemistry of PSII	Germ et al. (2007)

[2012a](#page-13-6)). Zhang et al. ([2014b](#page-14-0)) revealed that the application of Se (<50 g hm⁻²) increased the photosynthetic rate (Pn), intercellular $CO₂$ concentration (Ci), electron transport rate (ETR), and chlorophyll fuorescence parameters, such as Fv, Fo, Fv/Fo, and Fv/Fo, in rice; however, the photosynthesis index decreased when 100 g hm⁻² Se was applied. The photosynthetic physiology of plants is particularly sensitive to environmental stress. Further, the mechanistic efect of Se on plant photosynthesis may be similar to that of the antioxidant system; when electron transport is blocked during photosynthesis, reactive oxygen species (ROS) accumula-tion will be induced (Zhang et al. [2007\)](#page-14-2). Se affects plant photosynthesis by inhibiting or inducing ROS accumulation and the photosynthesis-related enzyme system. Se may also

afect electron transport and photosynthetic energy conversion by afecting Fe-S protein synthesis (Van Hoewyk et al. [2007;](#page-13-10) Feng et al. [2013\)](#page-10-9). Freeman et al. [\(2010](#page-10-10)) compared the ETR between Se-hyperaccumulating plants and non-Se-hyperaccumulating plants under Se treatment. The ETR of Se-hyperaccumulating plants was signifcantly increased after treatment with 20 μmol selenate, whereas that of non-Se-hyperaccumulating plants decreased signifcantly under the same conditions. At present, only few studies have assessed the effects of Se on plant photosynthetic physiology, which must be further examined at the physiological level. Further, the molecular mechanism should be detected.

An appropriate amount of Se can promote plant growth and increase the yield of crops, such as rice (Zhang et al. [2014b](#page-14-0)), wheat (Nawaz et al. [2015](#page-12-6)), and lentils (Ekanayake et al. [2015\)](#page-10-11), which is related to an increase in chlorophyll content, promotion of photosynthesis, enhanced antioxidant capacity of crops, and improved stress resistance of crops (Broadley et al. [2010;](#page-9-2) Jiang et al. [2015\)](#page-11-6).

Drought stress can lead to abnormal accumulation of ROS in plant cells, resulting in diferent degrees of oxidative damage to bioflm, protein, and DNA, which in turn afects plant growth, respiration, and photosynthesis, and can even lead to death in severe cases (Mittler [2002](#page-12-7)). Feng et al. ([2013\)](#page-10-9) suggested that Se may control the ROS level of plants under stress via three ways: by mediating the disproportionation of superoxide anion (O_2^-) to H_2O_2 ; through the direct involvement of Se compounds in the scavenging activities of O_2^- and hydroxyl radicals (\cdot OH); and by regulating the antioxidant enzyme system. Se metabolism in plant cells controls the balance of ROS concentration by regulating the concentration of free metal ions (Fe^{2+} , Cu^{+}). Se can directly or indirectly regulate the formation of antioxidant enzymes in plants, especially GSH-Px (Feng et al. [2013](#page-10-9)). Akladious [\(2012](#page-9-3)) indicated that exogenous Se can increase the content of proline and reduce plant stress injury. Se plays an important role in increasing the growth rate of crops (Cartes et al. [2010](#page-10-12)), reducing ultraviolet radiation oxidative damage (Yao et al. [2013](#page-14-5)), increasing chlorophyll and carotenoid contents in plant leaves (Dong et al. [2013](#page-10-13)), increasing the activity of antioxidant enzymes, and regulating the content of osmotic substances under heavy metal stress (Kumar et al. [2012](#page-11-7)). In addition, Se alleviates the adverse efects of drought stress on wheat (Nawaz et al. [2015\)](#page-12-6), barley (Habibi [2013](#page-11-8)), rape (Hasanuzzaman and Fujita [2011](#page-11-9)), and other crops by improving plant photosynthetic capacity, enhancing antioxidant capacity, and increasing the content of osmotic adjustment substances.

Heavy metals in the environment cannot be degraded, and the various forms of these metals will transform into each other as environmental factors change (Nagajyoti et al. [2010\)](#page-12-8). The toxicity of diferent forms of heavy metals is quite diferent, which poses a great challenge to the treatment of heavy metal pollution (Yao et al. [2012](#page-13-11)). An appropriate amount of Se can inhibit the absorption and transport of heavy metals by crop roots (Zhang et al. [2012\)](#page-14-3). Further, Se can directly react with heavy metals in the rhizosphere soil to inhibit its toxicity (Zeng et al. [2005](#page-14-6)). Se can afect the root surface iron plaque, soil solid phase, pH, microorganisms, root exudates and other possible indirect action pathways, convert heavy metals from a more toxic form to a less toxic form, complex/chelate with heavy metals, and reduce the mobility of heavy metals (Huang et al. [2018,](#page-11-4) [2015](#page-11-10); Cai et al. [2019](#page-9-1); Chen et al. [2019\)](#page-10-14), to ultimately reduce the availability of heavy metals in the soil and the accumulation of heavy metals in crops. An appropriate addition of Se could signifcantly reduce the accumulation of heavy metals in crops. Se could inhibit the transport of As from underground to the shoot (Hu et al. [2014](#page-11-11)) and limit the transport of Hg from the root to shoot of garlic (*Allium sativum*), thereby reducing the accumulation of Hg in the shoot (Zhao et al. [2013](#page-14-7)). Selenate could repair damage caused by Cd to the chloroplast membrane structure (Filek et al. [2010\)](#page-10-8). The antagonistic efect of Se on the accumulation of heavy metals is afected by many factors, including the concentration and form of Se and heavy metals. The inhibitory efect of Se on plant Cr (III) absorption was found to be signifcantly stronger than that on Cr (VI); however, the antagonistic efects of diferent fractions of Se on plant Cr absorption were similar (Srivastava et al. [1998\)](#page-13-12). Se may also increase the heavy metal content in plants. For example, Se treatment increased As accumulation in *Thunbergia alata* (Bluemlein et al. [2009\)](#page-9-4) and increased Cd and Cu contents in wheat shoots (Landberg and Greger [1994\)](#page-11-12), which may be due to unreasonable Se addition or different crop varieties (Feng and Wei [2012](#page-10-15)). Se can alleviate the toxic and side efects of heavy metals in plants by: directly inhibiting or forming complexes with heavy metals to inhibit the absorption and transport of heavy metals in plants (Feng et al. [2011;](#page-10-16) Malik et al. [2012](#page-12-4)) or participating in the regulation of the plant antioxidant system to alleviate heavy metal poisoning (Belzile et al. [2006;](#page-9-5) Filek et al. [2010](#page-10-8)).

At present, only few reports have been published on the efect of Se on plants under temperature stress and salt stress. Both high and low temperature induce plant oxidative stress, afect the stability of many types of enzyme activities in plants, and interfere with the normal growth and development of plants (Chiang et al. [2015](#page-10-17)). After soaking in a suitable concentration of Se solution, the contents of auxin, chlorophyll, anthocyanin, proline, and the activities of related antioxidant enzymes under temperature stress increase in wheat seeds to reduce cell membrane damage (Akladious [2012\)](#page-9-3). The application of exogenous Se can enhance the antioxidant system of crops, reduce the concentration of malondialdehyde (MDA), increase the content of proline, strengthen the protection of the mitochondrial electron transport chain, induce an increase in the protective substances in vivo, and alleviate the efects of temperature stress on plants (Djanaguiraman et al. [2018](#page-10-18)).

High salinity will destroy the structure of cytoplasmic membrane, hinder the absorption of mineral elements benefcial for growth, and cause secondary stress efects (Liang et al. [2017\)](#page-12-9). Due to high osmotic pressure, osmotic stress occurs in plants, resulting in ultrastructural damage of chloroplasts, limited or even closure of the respiratory stomata, and a decrease in the rate of photosynthesis and respiration (Hanin et al. [2016\)](#page-11-13). Under salt stress, Se acts as a coenzyme factor of antioxidant enzymes, such as superoxide dismutase (SOD), peroxidase (POD), and catalase (CAT). The increase in Se concentration increases antioxidant enzyme activity, enhances antioxidant levels, and decreases ROS concentration and MDA content in plants. The main efect of exogenous Se is to scavenge ROS, improve plant antioxidation, and reduce the damage caused by salt stress (Hu et al. [2013](#page-11-14)). Under salt stress, exogenous Se increases the absorption of N, K, Ca, and induces more metabolites and stress signals in plants (Elkelish et al. [2019](#page-10-6)). In addition, exogenous Se inhibits the increase in cell membrane permeability of wheat under salt stress, which could efectively alleviate the damage caused by salt stress to plants (Yigit et al. [2012](#page-14-8)). It is inferred that exogenous Se may reduce the expression of $Na⁺$ and $K⁺$ transporter gene, thereby maintaining the balance of cell infltration and improving the salt tolerance of plants. The comprehensive role of Se in plant stress shows that Se can improve the resistance of plants to abiotic stress by afecting ROS and the antioxidant system, interfering with the absorption and transport of heavy metals, changing the transformation process of heavy metals, and repairing the structure of cell membrane and chloroplast as well as the photosynthesis system (Feng et al. [2013](#page-10-9)). Crops are vulnerable to various diseases and insect pests and stress. Less free radicals and increased proline content can help plants resist stress, whereas Se can directly eliminate excessive free radicals, increase proline accumulation, enhance antioxidant enzyme activity, improve plant immunity, and increase its resistance to biotic and abiotic stresses (Kimani et al. [2013](#page-11-15); Priyadarsini et al. [2013;](#page-12-10) Steinbrenner and Sies [2013](#page-13-13); El-Demerdash and Nasr [2014\)](#page-10-19).

Se inhibits the growth of pathogenic microorganisms in vitro. Further, it has a strong inhibitory efect on *Aspergillus funiculosus* isolated from banana and *Alternaria tenuis* and *Fusarium* sp. isolated from tomato; the growth of these pathogenic fungi was terminated in 10 mg kg⁻¹ Na₂SeO₃ solution (Razak et al. [1991\)](#page-12-11). The minimum inhibitory concentration of Se is very low, and it serves as a benefcial essential element at low concentration. If Se is used as an antifungal agent, it will certainly reduce the negative efects of similar pollutants on the environment. When treated with Na_2SeO_3 , the level of ROS in the spores of a fungal pathogen, *Penicillium expansum*, increased, and the antioxidant system was destroyed, thereby weakening the cellular function of the pathogen and directly leading to plasma membrane damage of the pathogen, which inhibits the pathogen growth (Wu et al. [2014\)](#page-13-14). Exogenous Se treatment can not only inhibit the growth of pathogenic microorganisms, but also achieve the efect of Se enrichment in fruits. The preharvest and postharvest treatment of apple and tomato with $Na₂SeO₃$ revealed that although Se delayed plant tissue senescence and inhibited blue mold disease occurrence, the Se content in apple and tomato was 6 and 5.5 times higher than that in the control fruits, respectively, and remained within the safe edible range (Wu et al. [2015](#page-13-15); Wu et al. [2016](#page-13-16)). The use of exogenous Se helps the rape rhizosphere soil to accumulate benefcial rhizosphere microorganisms, thereby promoting plant growth (Cheng et al. [2021\)](#page-10-20). Soil Se inhibits *Sclerotinia sclerotiorum* by afecting the dissolved organic matter in rape straw and upregulating antifungal pathwayrelated genes (Cheng et al. [2020\)](#page-10-21).

Soil adsorption and fxation, leaching and migration, rice absorption, and gaseous volatilization are the four main processes of Se migration and transformation in the soil–plant system (Fernández-Martínez and Charlet [2009](#page-10-22)). Soil Se has diferent fractions, such as soluble state, exchange state and carbonate bound state, iron manganese oxide bound state, organic bound state and residue state (Wang et al. [2012b](#page-13-17)). The proportion of various fractions of Se in the soil is closely related to the physical and chemical properties of the soil, such as redox potential, pH, organic matter (Sharma et al. [2015\)](#page-13-2). The Se form in the soil is the result of the combined effect of soil acidity and alkalinity and redox condi-tions (Neal et al. [1987\)](#page-12-12). Changes in water content affect the redox conditions of soil (Tokunaga et al. [1996;](#page-13-18) Hefting et al. [2004](#page-11-16)), and changes in soil redox characteristics ultimately afect the form of Se in the soil (Gambrell [1994](#page-11-17); Dwire et al. [2006\)](#page-10-23). Soil drought, pH, clay content, and plant transpiration also play an important role in changing the soil's ability to fx Se (Jones et al. [2017](#page-11-18)).

There are fve relatively stable valence states of Se in the environment: -2 , -1 , 0 , $+4$, and $+6$ (Fernández-Martínez and Charlet [2009\)](#page-10-22). In soil, the most common fractions of Se are inorganic, SeO_4^2 ⁻ and SeO_3^2 ⁻, and the proportion of the two in the soil is controlled by the soil redox potential. When soil has alkaline and oxidizing conditions, SeO_4^{2-} is dominant; however, when soil has reducing conditions, SeO_3^{2-} is the main Se compound in the soil (Jacobs [1990\)](#page-11-19). The aerobic soil mainly contains Se in the form of selenite; however, in anoxic soil (such as fooded paddy soil), Se mainly exists in the form of selenite (Zhu et al. [2009\)](#page-14-9). The other organic fractions of Se in the soil depend on Se transformation by plants and microorganisms (Martens and Suarez [1996\)](#page-12-13). The concentration of Se in most soils is $0.01-2$ mg kg⁻¹, with an average value of 0.4 mg kg⁻¹ (Fordyce [2013](#page-10-24)).

The Se in solid materials, such as rocks and soil, only accounts for 30–60% of the total Se in the environment, whereas a large proportion of Se exists in water systems, such as oceans (Zhang et al. [2004](#page-14-10); Winkel et al. [2012](#page-13-1)). Therefore, Se in the atmosphere is mainly derived from the volatilization of marine substances (Blazina et al. [2017](#page-9-6)). Volatile Se is mainly dimethyl Se (DMSe) and dimethyl diselenide (DMDSe), which are easily soluble in water and can enter the soil via rainfall. Volatile Se is an important source of Se in soil (Amouroux et al. [2001](#page-9-7); Wen and Carignan [2009](#page-13-19)). There is a signifcant positive correlation between Se content in soil and precipitation (Sun et al. [2016](#page-13-20)). When the total Se in soil is 0.08–0.12 mg kg⁻¹, water soluble Se accounts for 1–2%. According to the calculation of 100% loss of water-soluble Se in the rainy season each year, the amount of Se loss per hectare can reach 2–8 g a−1 (Wang and Gao [2001](#page-13-21)). Therefore, a large amount of rainfall can accelerate the loss of soluble Se in soil (Jones et al. [2017\)](#page-11-18). Human farming activities also have an important infuence on the migration cycle of Se on the surface (Bailey [2017\)](#page-9-8). However, the aging process of exogenous Se fertilizer in soil markedly varies according to diferent soil types (Wang et al. [2017](#page-13-22)). In general, only 5–30% of Se fertilizers that are directly inputted into the soil can be absorbed and utilized by plants; 70–90% Se remains in the soil or is leached into the surrounding water (Sager [2006\)](#page-12-14).

More studies have found that the distribution of Se in soil is not consistent with that of the parent material, and the distribution of Se in soil cannot be explained by soil parent material alone (Blazina et al. [2014](#page-9-9); Sun et al. [2016\)](#page-13-20). Climate and soil properties are the two most important factors afecting soil Se concentration globally (Winkel et al. [2012](#page-13-1); Jones et al. [2017](#page-11-18)). The shading efect of plants on the soil, and surface vegetation can efectively reduce soil erosion by rainwater (Ravi et al. [2010\)](#page-12-15), thereby reducing Se loss from surface soil. However, plants can also indirectly affect the Se content and the form of soil by changing the physical and chemical properties of the soil. Jones et al. ([2017\)](#page-11-18) found remarkable diferences in soil organic carbon content under diferent vegetation types; the soil organic carbon content of farmland, woodland, and grassland increased, and the increase in organic carbon could fx soil Se to a certain extent, reduced Se loss, and indirectly increased soil Se retention. Therefore, there may be great diferences in soil Se under diferent land use types (Pilon-Smits et al. [2017](#page-12-16)). Among diferent plants, Se-hyperaccumulating plants have a greater effect on soil Se, and they can efectively change the distribution of soil Se and soil Se fractions (Pilon-Smits et al. [2017\)](#page-12-16). This is mainly achieved through the decomposition of litter and the action of root exudates of Se-hyperaccumulating plants (El Mehdawi and Pilon-Smits [2011\)](#page-10-25). Soil and plants are thus an inseparable whole, and good vegetation coverage can play a positive role in the fxation of soil and its nutrients and the reduction of loss induced by leaching.

In the process of regulating the adsorption and desorption of Se and Fe, Al, Ca, Mg ions in the soil, the pH of soil afects the Se form (Goh and Lim [2004](#page-11-20)). When Se chelates with the iron in soil to form an insoluble selenite-iron complex, it is not easily used by plants and reduces the amount of Se migration (Peak and Sparks [2002\)](#page-12-17). Compared with clay minerals, iron oxide has strong adsorption capacity, and its oxides can simultaneously adsorb Se $(+4 \text{ and } +6)$, of which, Se $(+4)$ is the main type adsorbed. However, under alkaline conditions, the negative salts formed by iron oxide and Se converts selenite into soluble selenate, increasing the migration amount of Se, and form a more stable complex under acidic reduction conditions (Gustafsson and Johnsson [1992](#page-11-21)). After Se is applied to the soil, Se can be adsorbed or fxed in a short time, thereby transforming it into a form that is difficult to be absorbed by crops. In terms of the ability to fx Se, oxides are the strongest in all components of soil, followed by organic matter (Hawkes and Kutnink [1996\)](#page-11-22).

Organic Se in soil is mainly recognized through the process of biodegradation and biosynthesis; however, the content of organically-bound Se in the soil is closely related to the content of soil organic matter. When the soil organic matter content is high, the content of organically-bound Se in the soil increases (Gustafsson and Johnsson [1992](#page-11-21)). Some related data show that organically-bound Se is related to its molecular weight. When the combination of soil organic matter and Se exists in the form of low molecular weight mixture, organically-bound Se can be directly absorbed and utilized by plants, whereas organically-bound Se with a larger molecular weight cannot be directly absorbed and utilized by plants (Zayed et al. [1998\)](#page-14-11). Thus, the blind application of organic Se fertilizer and the pursuit of a high-Serich state in the soil to increase the Se content in plants are not desirable. At the same time, organic selenides in soil can be decomposed by microorganisms to form methylation fractions and cause gaseous volatilization. The products are mainly alkyl selenides, such as DMDSe and DMSe complexes, which refect the loss pathway and quantity of soil Se (Gammelgaard et al. [2011](#page-11-23)). Inorganic selenides can also be methylated under the action of molds, and the methylation of Se may be related to the pH of soil (Zawislanski and Zavarin [1996\)](#page-14-12).

Phosphorus has a similar structure to Se. Phosphate fertilizer regulates the migration of Se, which proves that the amount of phosphate fertilizer in the soil signifcantly changes the adsorption of Se in the soil (Altansuvd et al. [2014\)](#page-9-10). When the phosphorus content is high, Se adsorption by soil is inhibited, thereby enhancing Se absorption by plants and further increasing Se mobility (Nakamaru et al. [2006;](#page-12-18) Altansuvd et al. [2014\)](#page-9-10). The Se content in plants is related to the content of S amino acids in plants, where S in the structure of amino acids is often replaced by Se, resulting in an increase in Se content in plants (Ip and Ganther [1994](#page-11-24)).

The structure and chemical properties of Se are similar to those of S. Most plants are unable to distinguish between Se and S, resulting in the absorption of sulfate. Selenate in soil is transported by the S transport pathway through the transporters, SULTR1;1 and SULTR1;2, in plants (Barberon et al. [2008\)](#page-9-11). SULTR2;1 and SULTR2;2 are involved in the selenate transfer process between roots, stems, and leaves, whereas SULTR4;1 and SULTR4;2 are mainly responsible for the phase transfer of Se between roots and stems (Zuber et al. [2010;](#page-14-13) Schiavon et al. [2015](#page-12-19)). SULTR3;1 transporters are mainly present in chloroplasts and are responsible for the transport of selenate across the membrane to chloroplasts (Cao et al. [2013](#page-10-26)). In summary, the pathway of plant absorption of selenate is highly related to the absorption of sulfate. Plant species and nutrition levels in the growing environment affect selenate transport efficiency in roots (White et al. [2004\)](#page-13-23). Some plant species growing in Se-rich soil display Se accumulation, and their Se absorption capacity is more than 100 times higher than that of ordinary plants, up to 1–15 g kg^{-1} (dry matter); these plants are called Se-hyperaccumulating plants (Beath et al. [1939\)](#page-9-12). Besides ordinary plants, there may be specifc selenate transporters responsible for the transport and migration of Se in plants (Schiavon et al. [2015](#page-12-19)). The absorption of Se by plants is an active process, which occurs via two main routes: Se in soil and Se in the atmosphere. Selenate, selenite, and organic Se are the main uptake forms of Se by plants. When selenate is absorbed by plants, the valence state of Se does not change during transport. After being transported to leaves, it is reduced $to +4$ valence state, converted into organic Se compounds, and fnally distributed to other organs and tissues of plants (Li et al. [2010\)](#page-11-25). After selenite is absorbed by plants, Se is converted into organic Se compounds in the roots, and most of the transformed organic Se compounds are retained in the roots, whereas a small portion is transported to the aboveground parts of plants (Keskinen et al. [2010\)](#page-11-26).

Unlike plant selenate absorption, the mechanism of plant selenite absorption is not clear (Zhu et al. [2009\)](#page-14-9). The transfer ability of plants to selenite is relatively poor, as selenite is easily converted into organic selenides after being absorbed by roots (Zhang et al. [2014a](#page-14-14)). Huang et al. [\(2016](#page-11-27)) found that the content of organic Se in wheat treated with selenite for 3 days was close to 90% of the total Se, and most of the substances produced were directly accumulated in the roots and could not be easily transferred to the aboveground parts of plants. The absorption of selenite by wheat is a metabolism-dependent active absorption process, which may be partially mediated by phosphorus transporters (Li et al. [2010](#page-11-25)). The silicon transport carrier, OsNIP2;1, in rice is also suggested to be related to the absorption of selenite. Selenite can enter the plant through OsNIP2;1, but this is regulated by environmental pH (Zhao et al. [2010\)](#page-14-15). Some studies suggest that the absorption of selenite by plants is a passive difusion process (Shrift and Ulrich [1969](#page-13-24); Arvy [1989,](#page-9-13) [1993](#page-9-14)). In particular, Terry et al. ([2000](#page-13-25)) reported that there is no evidence of membrane-mediated selenite uptake by plants. The studies on the absorption mechanism of selenite in plants are far less than that of selenate, which needs to be explored from the point of view of physiology and molecular biology.

Excessive Se has a toxic efect on plants, and there are remarkable diferences in the symptoms of Se poisoning among diferent plants. When non-Se-hyperaccumulating crops are grown in high Se medium, normal growth and development of plants are inhibited. Further, withering and shedding of plant leaves, a decrease in protein synthesis, and dwarfsm of plants occur (Trelease and Beath [1951;](#page-13-26) Mengel et al. [1982](#page-12-20)). Excessive Se leads to general inhibition in crops and reduces crop yield; however, phosphorus has a detoxifcation efect in a certain range (Singh and Singh [1978\)](#page-13-27). Se poisoning has not been reported to occur in Se-hyperaccumulating crops after the absorption of a large amount of Se.

The accumulation of Se in plants will affect plant pollen development and fertilization. The accumulation of a large amount of non-toxic selenomethylcysteine (MeSeCys) in plant fowers inhibits pollen formation. Further, the maternal parents who accumulate a large amount of Se in fowers will have an important effect on their reproductive fertilization (Prins et al. [2011](#page-12-21); Quinn et al. [2011](#page-12-22)).

Under the interaction of appropriate concentrations of S and Se, Se can inhibit or stimulate the absorption of S by plants (Cheng et al. [2016](#page-10-27)). When Se and S are combined, the resistance to cadmium stress is stronger than that of a single element. The ability of plants treated with S and Se to absorb Se was signifcantly higher than that of plants treated with Se alone (Golob et al. [2016\)](#page-11-28). Si and Se have a strong synergistic efect on reducing Cd toxicity (Huang et al. [2020\)](#page-11-29). Si-Se interaction increases the content of glutathione (GSH) and plant chelate (PC), causes more Cd to be distributed in the cell wall and organelles, and decreases the transport coef-ficient of Cd and accumulation in buds (Tang et al. [2015](#page-13-28); Pereira et al. [2018](#page-12-23)).

The process of using plants to transfer Se from the Secontaminated environment to alleviate soil Se pollution is called Se phytoremediation (Pilon-Smits [2005\)](#page-12-24). There are more than 30 species of Se-hyperaccumulating plants, such as *Stanleya*, *Astragalus*, and *Symphyotrichum* (Cappa and Pilon-Smits [2014\)](#page-10-28). Because of the Se enrichment characteristics of the above plants, they are often used in the phytoremediation of Se-contaminated soil (Salt et al. [1998;](#page-12-25) Pilon-Smits [2005\)](#page-12-24). Phytoremediation has the advantage of ecological balance, but the prerequisite for phytoremediation of high Se soil is to screen plants that meet the tolerance conditions. Further, the area to be repaired should have the basic conditions of light, temperature, water, and heat needed for plant growth (Wu et al. [2015\)](#page-13-15).

Microorganisms are the decisive factor of Se transformation (especially Se reduction) in soil (Yanke et al. [1995](#page-13-29); Martens and Suarez [1996](#page-12-13); Kessi and Hanselmann [2004](#page-11-30)). Therefore, understanding the role of soil microorganisms in Se transformation is particularly important to further clarify Se absorption by plants. The process of microbial transformation of Se generally involves dissimilatory reduction, assimilation reduction, oxidation, methylation, and demethylation (Dungan and Frankenberger [1999\)](#page-10-29). Among them, dissimilatory reduction can reduce the toxic oxidized Se $(SeO_3^2$, SeO_4^2) to non-toxic Se⁰ (Wen and Carignan [2007](#page-13-30)). As the synthesis method of the reduced product nano-Se is green, environmentally friendly, safe,

and less toxic, nano-Se is widely used in electrochemical sensing and anticancer (Mehdi et al. [2013](#page-12-2)).

The reduction of selenate to selenite and the reduction of selenite to elemental Se are two separate processes (Kuroda et al. [2011\)](#page-11-31). The mechanism of reduction of selenate to selenite has been thoroughly studied. In this process, there is a signifcant diference between the selenate reductase complex of gram-negative bacteria and gram-positive bacteria, which leads to a signifcant diference in the reduction process (Fig. [2\)](#page-7-0). Membrane-bound molybdate plays an important role in the reduction of selenate to selenite. In Gram-positive bacteria, SrdBCA is a membrane-bound molybdenum enzyme. SrdA contains one [4Fe-4S] cluster, SrdB contains four [4Fe-4S] clusters, and SrdC contains two transmembrane domains, indicating that the protein is located on the cell membrane. The SeO_4^{2-} reduction process is related to the oxidation of hydroquinone. Hydroquinone

Fig. 2 Schematic of selenate reduction by gram-positive bacteria (**a**) and gram-negative bacteria (**b**). MoCo, molybdenum cofactor; SerABC, selenate reductase; SrdBCA, selenate reductase; cytc4, cytochrome *c*4; QCR, quinol-cytochrome *c* oxidoreductase; Q, quinones; QH2, quinols.Adapted from Nancharaiah and Lens ([2015\)](#page-12-26) and Kuroda et al. [\(2011](#page-11-31))

combined with SrdC is oxidized to quinones, releasing 2 protons to the outside of the membrane and providing 2 electrons for SrdB (Nancharaiah and Lens [2015](#page-12-26)). Yee et al. [\(2007](#page-14-16)) inferred that the process of selenate reduction may be related to anaerobic respiration. In Gram-negative bacteria, SerABC is a soluble periplasmic molybdenum enzyme and SerA combines a molybdenum cofactor with catalytic activity and a subunit containing one [4Fe-4S] cluster. SerB contains three [4Fe-4S] clusters and one [3Fe-4S] cluster, and heme b (Nancharaiah and Lens [2015](#page-12-26)) is in SerC. SerABC accepts electrons from cytochrome $c₄$, combines with hydroquinone, and undergoes reduction under the catalysis of quinol-cytochrome *c* oxidoreductase (Lowe et al. [2010\)](#page-12-27).

The mechanism of reduction of selenite to elemental Se has not been clarifed. There are three well-recognized hypotheses regarding the SeO_3^2 ⁻ reduction mechanism: (1) the role of selenite reductase in the periplasmic space (Li et al. [2014](#page-12-28)); (2) sulfde-mediated selenite reduction (Nelson et al. [1996](#page-12-29)); and (3) GSH-mediated selenite reduction (Kessi and Hanselmann [2004](#page-11-30)). Because GSH exists widely in a variety of microorganisms, this process is recognized by most people.

Compared with the dissimilation reduction of Se by microorganisms, there are few reports on other Se transformation methods. Based on microorganism-based Se methylation studies, some microorganisms can use selenate to form DMSe in the process of photoautotrophy as well as selenides to form DMSe (McCarthy et al. [1993](#page-12-30)). The Se methylating microorganisms isolated from soil and sediment include fungi and bacteria, whereas the Se methylating microorganisms in water are mainly bacteria (Swift [2002](#page-13-31)). The bacterial thiopurine methyltransferase (bTPMT) encoded by the *tpm* gene and the novel methylase encoded by the mmtA gene can convert selenite into DMSe and DMDSe (Ranjard et al. [2003](#page-12-31), [2004](#page-12-32), [2002](#page-12-33)). In anoxic sediments or anaerobic conditions, methylSe and dimethyl sulfdes can undergo demethylation under the action of microorganisms; however, the current number of demethylated isolates is relatively small, and the types of reaction products have not been identifed (Francis et al. [1974](#page-10-30)). In the study of Se assimilation by microorganisms, Se combines with amino acids through covalent bonds to form selenomethionine (Se-Met) and selenocysteine (Sec, U); Se-Met can non-specifcally replace Met to participate in protein synthesis, and U specifcally participates in protein synthesis (Böck et al. [2006\)](#page-9-15). At present, few bacterial strains are known to be able to oxidize Se. As Se and S have similar chemical properties, the oxidation methods of Se and S are very similar (Blau [1961\)](#page-9-16). Studies have shown that the oxidation of Se^{0} to SeO_{4}^{2-} and SeO_{3}^{2} is mainly a biological process, and the speed is relatively slow (Torma and Habashi [1972;](#page-13-32) Dowdle and Oremland [1998](#page-10-31)). However, the oxidation mechanism of Se by microorganisms is still unclear and thus should be further examined.

Plant endophytes can also transform Se. Staicu et al. ([2015\)](#page-13-33) isolated the Se-tolerant endophytic strain, *Pseudomonas moraviensis*, from the Se hyperaccumulating plant, *Stanleya pinnata*, which can reduce the Se⁴⁺ of 790 mg L⁻¹ to nanoscale Se^0 below the detection limit within 48 h, indicating that the endophytic strain has a strong ability to metabolize Se. Sura-de Jong et al. [\(2015](#page-13-34)) isolated a variety of Se-tolerant endophytes from the roots, stems, and leaves of *Stanleya pinnata* and *Astragalus bisulcatus* in Se-rich areas of California, which can reduce 15.8 g L^{-1} selenate and selenite to elemental Se. These endophytes can still grow in high Se medium containing 10 mg L^{-1} Se, whereas rhizosphere fungi isolated from non-Se hyperaccumulating plants in the same area cannot grow in high Se medium. Altogether, microorganisms participate in the transformation of Se in various valence states and the formation of organic Se. Further, the transformation of Se in nature cannot be separated from microorganisms.

In recent years, the physiological level and molecular mechanism of Se absorption and transformation in plants have been discovered. A certain basis exists for the study of plant absorption and transformation of selenate; however, the absorption and transformation of selenite and organic Se and the synergistic effect of environmental factors on Se absorption still need to be discussed. Se uptake by plants can induce a series of complex physiological responses in plants. Owing to many factors, such as environmental factors and Se levels, the accumulation of Se in plants can afect their own reproduction and development, and may also afect the habitat of plants. Therefore, it is of great signifcance to study the ecological effects of the interaction between Se and plants in the environment.

The absorption and metabolism of Se in plants are complex, but are closely related to the physical and chemical properties of soil. However, many related mechanisms are still unclear. From the viewpoint of human nutrition and health, how to improve the Se content in the edible part of crops must be urgently established. Se is unevenly distributed in various organs of plants and is mainly distributed in the unharvested parts of crops (Carvalho et al. [2003\)](#page-10-32). With the development of molecular biology and by studying the mechanism of plant metabolism, it is possible to achieve the overexpression of target genes related to Se enrichment in specifc plant tissues, such as grains, to increase the Se content of specifc fractions in specifc tissues of crops.

Although some progress has been made in the study of the interaction between Se and heavy metals, the mechanism of the interaction between Se and heavy metals is not clear. At present, research on the appropriate amount of Se that can alleviate the toxicity of heavy metals is mainly focused on the antioxidation of Se. However, the relationship between the plant antioxidant system and Se form and concentration under heavy metal stress and the relationship between the plant antioxidant system and heavy metal species must be clarifed. In addition to activating the antioxidant system in plants, Se may also antagonize the toxic effects of heavy metals through other mechanisms; however, more experiments are needed to prove this hypothesis.

Further investigations are still needed to elucidate the mechanism of Se metabolism by microorganisms. Furthermore, more microbial resources involved in Se metabolism, especially the discovery of Se oxidizing bacteria, should be explored. However, the process of oxidation of elemental Se is markedly slower than the reduction process of Se. For the reduction and methylation of Se, much room still exists to explore the microbial resources. In addition, the reduction of Se is the most studied at present. However, the general law of selenate to selenite has not been fully clarifed. The role of microorganisms in the transformation of soil Se is yet to be understood. The change in mode of microbial transformation of soil Se and how it afects the way plants absorb Se also need to be demonstrated. These questions need to be simultaneously resolved by researchers. The relationship between the existing fractions of Se in soil and the role of microorganisms must be understood to determine the status and ability of plants to absorb Se.

In this review, we discussed the migration and accumulation of Se in the plant-soil-microbe system. The physical and chemical properties of soil, microorganisms, and vegetation afect the migration of Se. Microbes play an important role in the transformation of Se. Se promotes plant growth and development and enhances plant resistance to adversity stress. Its metabolism in plants is an important physiological process. The circulation route of Se in the environment is relatively clear; however, there are still many aspects to be studied in depth. Understanding the distribution characteristics and migration rules of Se in the environment provides a reference and scientifc basis for the development and utilization of Se resources and the development of Se-enriched agricultural systems.

Funding This work received support from Scientifc Startup Foundation for Doctors of Yulin Normal University (CN) (Grant No. G2020ZK13), Scientific Research and Technology Development Program of Guangxi (Grant Nos. GuiKe AA17202037, GuiKe AD19245169, GuiKe AD18281072), Yulin Science and Technology Program Project (Grant Nos. Yushikeneng 20194301, yushike20204038, yushikete202020001).

References

Akladious SA (2012) Infuence of diferent soaking times with selenium on growth, metabolic activities of wheat seedlings under low temperature stress. Afr J Biotech 11:14792–14804. [https://](https://doi.org/10.5897/AJB12.2140) doi.org/10.5897/AJB12.2140

- Alla MMN, Badran EG, Mohammed FA, Hassan NM, Abdelhamid MA (2020) Overexpression of $Na⁺$ -manipulating genes in wheat by selenium is associated with antioxidant enforcement for enhancement of salinity tolerance. Rendiconti Lincei Scienze Fisiche e Naturali 31:177–187. [https://doi.org/10.1007/](https://doi.org/10.1007/s12210-019-00868-8) [s12210-019-00868-8](https://doi.org/10.1007/s12210-019-00868-8)
- Altansuvd J, Nakamaru YM, Kasajima S, Ito H, Yoshida H (2014) Efect of long-term phosphorus fertilization on soil Se and transfer of soil Se to crops in northern Japan. Chemosphere 107:7–12.<https://doi.org/10.1016/j.chemosphere.2014.02.056>
- Amouroux D, Liss PS, Tessier E, Hamren-Larsson M, Donard OFX (2001) Role of oceans as biogenic sources of selenium. Earth Planet Sci Lett 189:277–283. [https://doi.org/10.1016/S0012-](https://doi.org/10.1016/S0012-821X(01)00370-3) [821X\(01\)00370-3](https://doi.org/10.1016/S0012-821X(01)00370-3)
- Arvy MP (1989) Some factors infuencing the uptake and distribution of selenite in the bean plant (*Phaseolus vulgaris*). Plant Soil 117:129–133.<https://doi.org/10.1007/BF02206265>
- Arvy MP (1993) Selenate and selenite uptake and translocation in bean plants (*Phaseolus vulgaris*). J Exp Bot 44:1083–1087. <https://doi.org/10.1093/jxb/44.6.1083>
- Bailey RT (2017) Review: selenium contamination, fate, and reactive transport in groundwater in relation to human health. Hydrogeol J 25:1191–1217. [https://doi.org/10.1007/](https://doi.org/10.1007/s10040-016-1506-8) [s10040-016-1506-8](https://doi.org/10.1007/s10040-016-1506-8)
- Barberon M, Berthomieu P, Clairotte M, Shibagaki N, Davidian JC, Gosti F (2008) Unequal functional redundancy between the two *Arabidopsis thaliana* high-affinity sulphate transporters *SULTR1;1* and *SULTR1;2*. New Phytol 180:608–619. [https://](https://doi.org/10.1111/j.1469-8137.2008.02604.x) doi.org/10.1111/j.1469-8137.2008.02604.x
- Beath OA, Gilbert CS, Eppson HF (1939) The use of indicator plants in locating seleniferous areas in Western United States. I. General. Am J Bot 26:257–269. [https://doi.org/10.1002/j.1537-2197.](https://doi.org/10.1002/j.1537-2197.1939.tb12900.x) [1939.tb12900.x](https://doi.org/10.1002/j.1537-2197.1939.tb12900.x)
- Belzile N, Wu GJ, Chen YW, Appanna VD (2006) Detoxifcation of selenite and mercury by reduction and mutual protection in the assimilation of both elements by *Pseudomonas fuorescens*. Sci Total Environ 367:704–714. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2006.03.008) [2006.03.008](https://doi.org/10.1016/j.scitotenv.2006.03.008)
- Blau M (1961) Biosynthesis of $[^{75}$ Se]selenomethionine and $[^{75}$ Se] selenocystine. Biochem Biophys Acta 49:389–390. [https://doi.org/](https://doi.org/10.1016/0006-3002(61)90140-8) [10.1016/0006-3002\(61\)90140-8](https://doi.org/10.1016/0006-3002(61)90140-8)
- Blazina T, Sun Y, Voegelin A, Lenz M, Berg M, Winkel LHE (2014) Terrestrial selenium distribution in China is potentially linked to monsoonal climate. Nat Commun 5:4717. [https://doi.org/10.](https://doi.org/10.1038/ncomms5717) [1038/ncomms5717](https://doi.org/10.1038/ncomms5717)
- Blazina T, Läderach A, Jones GD, Sodemann H, Wernli H, Kirchner JW, Winkel LHE (2017) Marine primary productivity as a potential indirect source of selenium and other trace elements in atmospheric deposition. Environ Sci Technol 51:108–118. <https://doi.org/10.1021/acs.est.6b03063>
- Bluemlein K, Klimm E, Raab A, Feldmann J (2009) Selenite enhances arsenate toxicity in *Thunbergia alata*. Environ Chem 6:486–494. <https://doi.org/10.1071/EN09101>
- Böck A, Rother M, Leibundgut M, Ban N (2006) Selenium metabolism in prokaryotes. In: Hatfeld DL, Berry MJ, Gladyshev VN (eds) Selenium. Springer, Boston, MA, pp 9–28. [https://doi.org/10.](https://doi.org/10.1007/0-387-33827-6_2) [1007/0-387-33827-6_2](https://doi.org/10.1007/0-387-33827-6_2)
- Broadley MR et al (2010) Selenium biofortifcation of high-yielding winter wheat (*Triticum aestivum* L.) by liquid or granular Se fertilisation. Plant Soil 332:5–18. [https://doi.org/10.1007/](https://doi.org/10.1007/s11104-009-0234-4) [s11104-009-0234-4](https://doi.org/10.1007/s11104-009-0234-4)
- Cai M et al (2019) Selenium induces changes of rhizosphere bacterial characteristics and enzyme activities afecting chromium/ selenium uptake by pak choi (*Brassica campestris* L. ssp. *Chinensis* Makino) in chromium contaminated soil. Environ Pollut 249:716–727.<https://doi.org/10.1016/j.envpol.2019.03.079>
- Cao MJ, Wang Z, Wirtz M, Hell R, Oliver DJ, Xiang CB (2013) SULTR3;1 is a chloroplast-localized sulfate transporter in *Arabidopsis thaliana*. Plant J 73:607–616. [https://doi.org/10.1111/tpj.](https://doi.org/10.1111/tpj.12059) [12059](https://doi.org/10.1111/tpj.12059)
- Cappa JJ, Pilon-Smits EAH (2014) Evolutionary aspects of elemental hyperaccumulation. Planta 239:267–275. [https://doi.org/10.1007/](https://doi.org/10.1007/s00425-013-1983-0) [s00425-013-1983-0](https://doi.org/10.1007/s00425-013-1983-0)
- Cartes P, Jara AA, Pinilla L, Rosas A, Mora ML (2010) Selenium improves the antioxidant ability against aluminium-induced oxidative stress in ryegrass roots. Ann Appl Biol 156:297–307. <https://doi.org/10.1111/j.1744-7348.2010.00387.x>
- Carvalho KM, Gallardo-Williams MT, Benson RF, Martin DF (2003) Efects of selenium supplementation on four agricultural crops. J Agric Food Chem 51:704–709. [https://doi.org/10.1021/jf025](https://doi.org/10.1021/jf0258555) [8555](https://doi.org/10.1021/jf0258555)
- Chen Y, Zhu Q, Dong X, Huang W, Du C, Lu D (2019) How *Serratia marcescens* HB-4 absorbs cadmium and its implication on phytoremediation. Ecotoxicol Environ Saf 185:109723. [https://doi.](https://doi.org/10.1016/j.ecoenv.2019.109723) [org/10.1016/j.ecoenv.2019.109723](https://doi.org/10.1016/j.ecoenv.2019.109723)
- Cheng B et al (2016) Efects of selenium and sulfur on antioxidants and physiological parameters of garlic plants during senescence. J Integr Agric 15:566–572. [https://doi.org/10.1016/s2095-](https://doi.org/10.1016/s2095-3119(15)61201-1) [3119\(15\)61201-1](https://doi.org/10.1016/s2095-3119(15)61201-1)
- Cheng Q et al (2020) Enhancement and improvement of selenium in soil to the resistance of rape stem against *Sclerotinia sclerotiorum* and the inhibition of dissolved organic matter derived from rape straw on mycelium. Environ Pollut 265:114827. [https://doi.](https://doi.org/10.1016/j.envpol.2020.114827) [org/10.1016/j.envpol.2020.114827](https://doi.org/10.1016/j.envpol.2020.114827)
- Cheng Q, Hu C, Ming J, Cai M, Liu K, Tang Y, Zhao X (2021) Efects of selenium on microorganisms in the rhizosphere soil of oilseed rape. J Agric Res Environ 38:104–110. [https://doi.org/10.](https://doi.org/10.13254/j.jare.2020.0061) [13254/j.jare.2020.0061](https://doi.org/10.13254/j.jare.2020.0061)
- Chiang CM, Chien HL, Chen LFO, Hsiung TC, Chiang MC, Chen SP, Lin KH (2015) Overexpression of the genes coding ascorbate peroxidase from Brassica campestris enhances heat tolerance in transgenic *Arabidopsis thaliana*. Biol Plant 59:305–315. [https://](https://doi.org/10.1007/s10535-015-0489-y) doi.org/10.1007/s10535-015-0489-y
- Chu J, Yao X, Zhang Z (2010) Responses of wheat seedlings to exogenous selenium supply under cold stress. Biol Trace Elem Res 136:355–363.<https://doi.org/10.1007/s12011-009-8542-3>
- de Lima Lessa JH et al (2019) Agronomic biofortifcation of rice (*Oryza sativa* L.) with selenium and its efect on element distributions in biofortifed grains. Plant Soil 444:331–342. [https://](https://doi.org/10.1007/s11104-019-04275-8) doi.org/10.1007/s11104-019-04275-8
- Deng X, Zhao Z, Zhou JJ, Chen JZ, Lv C, Liu X (2018) Compositional analysis of typical selenium ore from Enshi and its efect on selenium enrichment in wetland and dryland crops. Plant Soil 433:55–64.<https://doi.org/10.1007/s11104-018-3822-3>
- Diao M, Ma L, Wang J, Cui J, Fu A, Liu H-y (2014) Selenium promotes the growth and photosynthesis of tomato seedlings under salt stress by enhancing chloroplast antioxidant defense system. J Plant Growth Regul 33:671–682. [https://doi.org/10.1007/](https://doi.org/10.1007/s00344-014-9416-2) [s00344-014-9416-2](https://doi.org/10.1007/s00344-014-9416-2)
- Djanaguiraman M, Belliraj N, Bossmann SH, Prasad PVV (2018) High-temperature stress alleviation by selenium nanoparticle treatment in grain sorghum. ACS Omega 3:2479–2491. [https://](https://doi.org/10.1021/acsomega.7b01934) doi.org/10.1021/acsomega.7b01934
- Dong JZ et al (2013) Selenium increases chlorogenic acid, chlorophyll and carotenoids of *Lycium chinense* leaves. J Sci Food Agric 93:310–315. <https://doi.org/10.1002/jsfa.5758>
- Dowdle PR, Oremland RS (1998) Microbial oxidation of elemental selenium in soil slurries and bacterial cultures. Environ Sci Technol 32:3749–3755.<https://doi.org/10.1021/es970940s>
- Dungan RS, Frankenberger WT (1999) Microbial transformations of selenium and the bioremediation of seleniferous environments.

Bioremediat J 3:171–188. [https://doi.org/10.1080/1088986999](https://doi.org/10.1080/10889869991219299) [1219299](https://doi.org/10.1080/10889869991219299)

- Dwire KA, Kaufman JB, Baham JE (2006) Plant species distribution in relation to water-table depth and soil redox potential in montane riparian meadows. Wetlands 26:131–146. [https://doi.org/10.](https://doi.org/10.1672/0277-5212(2006)26[131:PSDIRT]2.0.CO;2) [1672/0277-5212\(2006\)26\[131:PSDIRT\]2.0.CO;2](https://doi.org/10.1672/0277-5212(2006)26[131:PSDIRT]2.0.CO;2)
- Ekanayake LJ, Vial E, Schatz B, McGee R, Thavarajah P (2015) Selenium fertilization on lentil (*Lens culinaris* Medikus) grain yield, seed selenium concentration, and antioxidant activity. Field Crop Res 177:9–14.<https://doi.org/10.1016/j.fcr.2015.03.002Get>
- Ekumah J-N, Ma Y, Akpali-Tsigbe NDK, Kwaw E, Ma S, Jie H (2021) Global soil distribution, dietary access routes, bioconversion mechanisms and the human health signifcance of selenium: a review. Food Biosci 41:100960. [https://doi.org/10.1016/j.fbio.](https://doi.org/10.1016/j.fbio.2021.100960) [2021.100960](https://doi.org/10.1016/j.fbio.2021.100960)
- El Mehdawi AF, Pilon-Smits EAH (2011) Ecological aspects of plant selenium hyperaccumulation. Plant Biol 14:1-10. [https://doi.org/](https://doi.org/10.1111/j.1438-8677.2011.00535.x) [10.1111/j.1438-8677.2011.00535.x](https://doi.org/10.1111/j.1438-8677.2011.00535.x)
- El-Demerdash FM, Nasr HM (2014) Antioxidant efect of selenium on lipid peroxidation, hyperlipidemia and biochemical parameters in rats exposed to diazinon. J Trace Elem Med Biol 28:89–93. <https://doi.org/10.1016/j.jtemb.2013.10.001>
- Elkelish AA, Soliman MH, Alhaithloul HA, El-Esawi MA (2019) Selenium protects wheat seedlings against salt stress-mediated oxidative damage by up-regulating antioxidants and osmolytes metabolism. Plant Physiol Biochem 137:144–153. [https://doi.](https://doi.org/10.1016/j.plaphy.2019.02.004) [org/10.1016/j.plaphy.2019.02.004](https://doi.org/10.1016/j.plaphy.2019.02.004)
- Favorito JE, Grossl PR, Davis TZ, Eick MJ, Hankes N (2020) Soilplant-animal relationships and geochemistry of selenium in the Western Phosphate Resource Area (United States): a review. Chemosphere 266:128959. [https://doi.org/10.1016/j.chemo](https://doi.org/10.1016/j.chemosphere.2020.128959) [sphere.2020.128959](https://doi.org/10.1016/j.chemosphere.2020.128959)
- Feng RW, Wei CY (2012) Antioxidative mechanisms on selenium accumulation in *Pteris vittata* L., a potential selenium phytoremediation plant. Plant Soil Environ 58:105–110. [https://doi.org/](https://doi.org/10.17221/162/2011-PSE) [10.17221/162/2011-PSE](https://doi.org/10.17221/162/2011-PSE)
- Feng R, Wei C, Tu S, Tang S, Wu F (2011) Detoxifcation of antimony by selenium and their interaction in paddy rice under hydroponic conditions. Microchem J 97:57–61. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.microc.2010.06.003) [microc.2010.06.003](https://doi.org/10.1016/j.microc.2010.06.003)
- Feng R, Wei C, Tu S (2013) The roles of selenium in protecting plants against abiotic stresses. Environ Exp Bot 87:58–68. [https://doi.](https://doi.org/10.1016/j.envexpbot.2012.09.002) [org/10.1016/j.envexpbot.2012.09.002](https://doi.org/10.1016/j.envexpbot.2012.09.002)
- Fernández-Martínez A, Charlet L (2009) Selenium environmental cycling and bioavailability: a structural chemist point of view. Rev Environ Sci Bio/Technol 8:81–110. [https://doi.org/10.1007/](https://doi.org/10.1007/s11157-009-9145-3) [s11157-009-9145-3](https://doi.org/10.1007/s11157-009-9145-3)
- Filek M, Gzyl-Malcher B, Zembala M, Bednarska E, Laggner P, Kriechbaum M (2010) Efect of selenium on characteristics of rape chloroplasts modifed by cadmium. J Plant Physiol 167:28– 33.<https://doi.org/10.1016/j.jplph.2009.07.003>
- Fischer S et al (2020) *Bacillus safensis* JG-B5T affects the fate of selenium by extracellular production of colloidally less stable selenium nanoparticles. J Hazard Mater 384:121146. [https://doi.](https://doi.org/10.1016/j.jhazmat.2019.121146) [org/10.1016/j.jhazmat.2019.121146](https://doi.org/10.1016/j.jhazmat.2019.121146)
- Fordyce FM (2013) Selenium deficiency and toxicity in the environment. In: Selinus O (ed) Essentials of medical geology. Springer, Dordrecht, pp 375–416. [https://doi.org/10.1007/978-94-007-](https://doi.org/10.1007/978-94-007-4375-5_16) [4375-5_16](https://doi.org/10.1007/978-94-007-4375-5_16)
- Francis AJ, Duxbury JM, Alexander M (1974) Evolution of dimethylselenide from soils. Appl Microbiol 28:248–250. [https://doi.org/](https://doi.org/10.1128/aem.28.2.248-250.1974) [10.1128/aem.28.2.248-250.1974](https://doi.org/10.1128/aem.28.2.248-250.1974)
- Freeman JL et al (2010) Molecular mechanisms of selenium tolerance and hyperaccumulation in *Stanleya pinnata*. Plant Physiol 153:1630–1652. <https://doi.org/10.1104/pp.110.156570>
- Galić L, Vinković T, Ravnjak B, Lončarić Z (2021) Agronomic biofortifcation of signifcant cereal crops with selenium—A review. Agronomy 11:1015. <https://doi.org/10.3390/agronomy11051015>
- Gambrell RP (1994) Trace and toxic metals in wetlands—a review. J Environ Qual 23:883–891. [https://doi.org/10.2134/jeq1994.](https://doi.org/10.2134/jeq1994.00472425002300050005x) [00472425002300050005x](https://doi.org/10.2134/jeq1994.00472425002300050005x)
- Gammelgaard B, Jackson MI, Gabel-Jensen C (2011) Surveying selenium speciation from soil to cell—forms and transformations. Anal Bioanal Chem 399:1743–1763. [https://doi.org/10.1007/](https://doi.org/10.1007/s00216-010-4212-8) [s00216-010-4212-8](https://doi.org/10.1007/s00216-010-4212-8)
- Germ M, Stibilj V, Osvald J, Kreft I (2007) Efect of selenium foliar application on chicory (*Cichorium intybus* L.). J Agric Food Chem 55:795–798.<https://doi.org/10.1021/jf0629888>
- Goh K-H, Lim T-T (2004) Geochemistry of inorganic arsenic and selenium in a tropical soil: efect of reaction time, pH, and competitive anions on arsenic and selenium adsorption. Chemosphere 55:849–859.<https://doi.org/10.1016/j.chemosphere.2003.11.041>
- Golob A, Gadžo D, Stibilj V, Djikić M, Gavrić T, Kreft I, Germ M (2016) Sulphur interferes with selenium accumulation in Tartary buckwheat plants. Plant Physiol Biochem 108:32–36. [https://doi.](https://doi.org/10.1016/j.plaphy.2016.07.001) [org/10.1016/j.plaphy.2016.07.001](https://doi.org/10.1016/j.plaphy.2016.07.001)
- Gómez-Gómez B, Pérez-Corona T, Mozzi F, Pescuma M, Madrid Y (2019) Silac-based quantitative proteomic analysis of *Lactobacillus reuteri* CRL 1101 response to the presence of selenite and selenium nanoparticles. J Proteomics 195:53–65. [https://doi.org/](https://doi.org/10.1016/j.jprot.2018.12.025) [10.1016/j.jprot.2018.12.025](https://doi.org/10.1016/j.jprot.2018.12.025)
- Guignardi Z, Schiavon M (2017) Biochemistry of Plant Selenium Uptake and Metabolism. In: Pilon-Smits E, Winkel L, Lin ZQ (eds) Selenium in plants, vol 11. Springer, Cham, Switzerland. https://doi.org/10.1007/978-3-319-56249-0_2
- Gustafsson JP, Johnsson L (1992) Selenium retention in the organic matter of Swedish forest soils. Eur J Soil Sci 43:461–472. [https://](https://doi.org/10.1111/j.1365-2389.1992.tb00152.x) doi.org/10.1111/j.1365-2389.1992.tb00152.x
- Habibi G (2013) Effect of drought stress and selenium spraying on photosynthesis and antioxidant activity of spring barley. Acta Agriculturae Slovenica 101:31–39. [https://doi.org/10.2478/](https://doi.org/10.2478/acas-2013-0004) [acas-2013-0004](https://doi.org/10.2478/acas-2013-0004)
- Hanin M, Ebel C, Ngom M, Laplaze L, Masmoudi K (2016) New insights on plant salt tolerance mechanisms and their potential use for breeding. Front Plant Sci 7:01787. [https://doi.org/10.](https://doi.org/10.3389/fpls.2016.01787) [3389/fpls.2016.01787](https://doi.org/10.3389/fpls.2016.01787)
- Hasanuzzaman M, Fujita M (2011) Selenium pretreatment upregulates the antioxidant defense and methylglyoxal detoxifcation system and confers enhanced tolerance to drought stress in rapeseed seedlings. Biol Trace Elem Res 143:1758–1776. [https://doi.org/](https://doi.org/10.1007/s12011-011-8998-9) [10.1007/s12011-011-8998-9](https://doi.org/10.1007/s12011-011-8998-9)
- Hawkes WC, Kutnink MA (1996) High-performance liquid chromatographic-fuorescence determination of traces of selenium in biological materials. Anal Biochem 241:206–211. [https://doi.org/](https://doi.org/10.1006/abio.1996.0401) [10.1006/abio.1996.0401](https://doi.org/10.1006/abio.1996.0401)
- Hefting M et al (2004) Water table elevation controls on soil nitrogen cycling in riparian wetlands along a European climatic gradient. Biogeochemistry 67:113–134. [https://doi.org/10.1023/B:BIOG.](https://doi.org/10.1023/B:BIOG.0000015320.69868.33) [0000015320.69868.33](https://doi.org/10.1023/B:BIOG.0000015320.69868.33)
- Hu KL, Zhang L, Wang JT, You Y (2013) Infuence of selenium on growth, lipid peroxidation and antioxidative enzyme activity in melon (*Cucumis melo* L.) seedlings under salt stress. Acta Soc Bot Pol 82:193–197. <https://doi.org/10.5586/asbp.2013.023>
- Hu Y, Duan G-L, Huang Y-Z, Liu Y-X, Sun G-X (2014) Interactive efects of diferent inorganic As and Se species on their uptake and translocation by rice (*Oryza sativa* L.) seedlings. Environ Sci Pollut Res 21:3955–3962. [https://doi.org/10.1007/](https://doi.org/10.1007/s11356-013-2321-6) [s11356-013-2321-6](https://doi.org/10.1007/s11356-013-2321-6)
- Huang Q, Yu Y, Wang Q, Luo Z, Jiang R, Li H (2015) Uptake kinetics and translocation of selenite and selenate as afected by iron

plaque on root surfaces of rice seedlings. Planta 241:907–916. <https://doi.org/10.1007/s00425-014-2227-7>

- Huang QQ, Wang Q, Wan YN, Yu Y, Li HF (2016) Application of X-ray absorption near edge spectroscopy to the study of the efect of sulphur on selenium uptake and assimilation in wheat seedlings. Biol Plant 61:726–732. [https://doi.org/10.1007/](https://doi.org/10.1007/s10535-016-0698-z) [s10535-016-0698-z](https://doi.org/10.1007/s10535-016-0698-z)
- Huang Q, Xu Y, Liu Y, Qin X, Huang R, Liang X (2018) Selenium application alters soil cadmium bioavailability and reduces its accumulation in rice grown in Cd-contaminated soil. Environ Sci Pollut Res 25:31175–31182. [https://doi.org/10.1007/](https://doi.org/10.1007/s11356-018-3068-x) [s11356-018-3068-x](https://doi.org/10.1007/s11356-018-3068-x)
- Huang H et al (2020) Synergistic effect of silicon and selenium on the alleviation of cadmium toxicity in rice plants. J Hazard Mater 401:123393. <https://doi.org/10.1016/j.jhazmat.2020.123393>
- Ip C, Ganther HE (1994) Novel strategies in selenium cancer chemoprevention research. In: Burk RF (ed) Selenium in biology and human health. Springer, New York, NY, pp 169–180. [https://doi.](https://doi.org/10.1007/978-1-4612-2592-8_10) [org/10.1007/978-1-4612-2592-8_10](https://doi.org/10.1007/978-1-4612-2592-8_10)
- Jacobs LW (1990) Selenium in Agriculture and the Environment. Soil Sci 149:121. [https://doi.org/10.1097/00010694-19900](https://doi.org/10.1097/00010694-199002000-00008) [2000-00008](https://doi.org/10.1097/00010694-199002000-00008)
- Jiang C, Zu C, Shen J, Shao F, Li T (2015) Efects of selenium on the growth and photosynthetic characteristics of fue-cured tobacco (*Nicotiana tabacum* L.). Acta Soc Bot Pol 84:71–77. [https://doi.](https://doi.org/10.5586/asbp.2015.006) [org/10.5586/asbp.2015.006](https://doi.org/10.5586/asbp.2015.006)
- Jiang C, Zu C, Lu D, Zheng Q, Shen J, Wang H, Li D (2017) Efect of exogenous selenium supply on photosynthesis, Na+ accumulation and antioxidative capacity of maize (*Zea mays* L.) under salinity stress. Sci Rep 7:42039. [https://doi.org/10.1038/srep4](https://doi.org/10.1038/srep42039) [2039](https://doi.org/10.1038/srep42039)
- Jones GD et al (2017) Selenium deficiency risk predicted to increase under future climate change. Proc Natl Acad Sci USA 114:2848– 2853.<https://doi.org/10.1073/pnas.1611576114>
- Keskinen R, Turakainen M, Hartikainen H (2010) Plant availability of soil selenate additions and selenium distribution within wheat and ryegrass. Plant Soil 333:301–313. [https://doi.org/10.1007/](https://doi.org/10.1007/s11104-010-0345-y) [s11104-010-0345-y](https://doi.org/10.1007/s11104-010-0345-y)
- Kessi J, Hanselmann KW (2004) Similarities between the abiotic reduction of selenite with glutathione and the dissimilatory reaction mediated by *Rhodospirillum rubrum* and *Escherichia coli*. J Biol Chem 279:50662–50669. [https://doi.org/10.1074/](https://doi.org/10.1074/jbc.M405887200) [jbc.M405887200](https://doi.org/10.1074/jbc.M405887200)
- Kimani MM, Bayse CA, Stadelman BS, Brumaghim JL (2013) Oxidation of biologically relevant chalcogenones and their Cu(I) complexes: insight into selenium and sulfur antioxidant activity. Inorg Chem 52:11685–11687.<https://doi.org/10.1021/ic401366c>
- Kumar M, Bijo AJ, Baghel RS, Reddy CRK, Jha B (2012) Selenium and spermine alleviate cadmium induced toxicity in the red seaweed *Gracilaria dura* by regulating antioxidants and DNA methylation. Plant Physiol Biochem 51:129–138. [https://doi.org/10.](https://doi.org/10.1016/j.plaphy.2011.10.016) [1016/j.plaphy.2011.10.016](https://doi.org/10.1016/j.plaphy.2011.10.016)
- Kuroda M et al (2011) Molecular cloning and characterization of the srdBCA operon, encoding the respiratory selenate reductase complex, from the selenate-reducing bacterium *Bacillus selenatarsenatis* SF-1. J Bacteriol 193:2141–2148. [https://doi.org/](https://doi.org/10.1128/JB.01197-10) [10.1128/JB.01197-10](https://doi.org/10.1128/JB.01197-10)
- Landberg T, Greger M (1994) Infuence of selenium on uptake and toxicity of copper and cadmium in pea (*Pisum sativum*) and wheat (*Triticum aestivum*). Physiol Plant 90:637–644. [https://doi.org/](https://doi.org/10.1111/j.1399-3054.1994.tb02518.x) [10.1111/j.1399-3054.1994.tb02518.x](https://doi.org/10.1111/j.1399-3054.1994.tb02518.x)
- Li HF, McGrath SP, Zhao FJ (2010) Selenium uptake, translocation and speciation in wheat supplied with selenate or selenite. New Phytol 178:92–102. [https://doi.org/10.1111/j.1469-8137.2007.](https://doi.org/10.1111/j.1469-8137.2007.02343.x) [02343.x](https://doi.org/10.1111/j.1469-8137.2007.02343.x)
- Li D-B et al (2014) Selenite reduction by *Shewanella oneidensis* MR-1 is mediated by fumarate reductase in periplasm. Sci Rep 4:3735. <https://doi.org/10.1038/srep03735>
- Liang W, Ma X, Wan P, Liu L (2017) Plant salt-tolerance mechanism: a review. Biochem Biophys Res Commun 495:286–291. <https://doi.org/10.1016/j.bbrc.2017.11.043>
- Lowe EC et al (2010) Quinol-cytochrome c oxidoreductase and cytochrome c₄ mediate electron transfer during selenate respiration in *Thauera selenatis*. J Biol Chem 285:18433–18442. <https://doi.org/10.1074/jbc.M110.115873>
- Malheiros RS, Gonçalves FC, Brito FA, Zsögön A, Ribeiro DM (2020) Selenomethionine induces oxidative stress and modifes growth in rice (*Oryza sativa* L.) seedlings through efects on hormone biosynthesis and primary metabolism. Ecotoxicol Environm saf 189:109942. [https://doi.org/10.1016/j.ecoenv.](https://doi.org/10.1016/j.ecoenv.2019.109942) [2019.109942](https://doi.org/10.1016/j.ecoenv.2019.109942)
- Malik JA, Goel S, Kaur N, Singh I, Nayyar H (2012) Selenium antagonises the toxic efects of arsenic on mungbean (*Phaseolus aureus* Roxb.) plants by restricting its uptake and enhancing the antioxidative and detoxifcation mechanisms. Environ Exp Bot 77:242– 248. <https://doi.org/10.1016/j.envexpbot.2011.12.001>
- Martens DA, Suarez DL (1996) Selenium speciation of soil/sediment determined with sequential extractions and hydride generation atomic absorption spectrophotometry. Environ Sci Technol 31:133–139. <https://doi.org/10.1021/es960214+>
- McCarthy S, Chasteen T, Marshall M, Fall R, Bachofen R (1993) Phototrophic bacteria produce volatile, methylated sulfur and selenium compounds. FEMS Microbiol Lett 112:93–97. [https://doi.](https://doi.org/10.1111/j.1574-6968.1993.tb06429.x) [org/10.1111/j.1574-6968.1993.tb06429.x](https://doi.org/10.1111/j.1574-6968.1993.tb06429.x)
- Mehdi Y, Hornick J-L, Istasse L, Dufrasne I (2013) Selenium in the environment, metabolism and involvement in body functions. Molecules 18:3292–3311. [https://doi.org/10.3390/molecules1](https://doi.org/10.3390/molecules18033292) [8033292](https://doi.org/10.3390/molecules18033292)
- Mengel K, Kirkby EA, Kosegarten H, Appel T (1982) Principles of plant nutrition. Springer, Dordrecht, Netherlands. [https://doi.org/](https://doi.org/10.1007/978-94-010-1009-2) [10.1007/978-94-010-1009-2](https://doi.org/10.1007/978-94-010-1009-2)
- Mittler R (2002) Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci 7:405–410. [https://doi.org/10.1016/S1360-](https://doi.org/10.1016/S1360-1385(02)02312-9) [1385\(02\)02312-9](https://doi.org/10.1016/S1360-1385(02)02312-9)
- Morales-Espinoza MC et al (2019) Se nanoparticles induce changes in the growth, antioxidant responses, and fruit quality of tomato developed under NaCl Stress. Molecules 24:3030. [https://doi.org/](https://doi.org/10.3390/molecules24173030) [10.3390/molecules24173030](https://doi.org/10.3390/molecules24173030)
- Nagajyoti PC, Lee KD, Sreekanth TVM (2010) Heavy metals, occurrence and toxicity for plants: a review. Environ Chem Lett 8:199– 216. <https://doi.org/10.1007/s10311-010-0297-8>
- Nakamaru Y, Tagami K, Uchida S (2006) Efect of phosphate addition on the sorption-desorption reaction of selenium in Japanese agricultural soils. Chemosphere 63:109–115. [https://doi.org/10.](https://doi.org/10.1016/j.chemosphere.2005.07.046) [1016/j.chemosphere.2005.07.046](https://doi.org/10.1016/j.chemosphere.2005.07.046)
- Nancharaiah YV, Lens PNL (2015) Selenium biomineralization for biotechnological applications. Trends Biotechnol 33:323–330. <https://doi.org/10.1016/j.tibtech.2015.03.004>
- Natasha SM, Niazi NK, Khalid S, Murtaza B, Bibi I, Rashid MI (2018) A critical review of selenium biogeochemical behavior in soilplant system with an inference to human health. Environ Pollut 234:915–934.<https://doi.org/10.1016/j.envpol.2017.12.019>
- Nawaz F, Ahmad R, Ashraf MY, Waraich EA, Khan SZ (2015) Efect of selenium foliar spray on physiological and biochemical processes and chemical constituents of wheat under drought stress. Ecotoxicol Environ Saf 113:191–200. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ecoenv.2014.12.003) [ecoenv.2014.12.003](https://doi.org/10.1016/j.ecoenv.2014.12.003)
- Neal RH, Sposito G, Holtzclaw KM, Traina SJ (1987) Selenite adsorption on alluvial soils: I. Soil composition and pH efects. Soil Sci Soc Am J 51:1161–1165. [https://doi.org/10.2136/sssaj1987.](https://doi.org/10.2136/sssaj1987.03615995005100050012x) [03615995005100050012x](https://doi.org/10.2136/sssaj1987.03615995005100050012x)
- Nelson DC, Casey WH, Sison JD, Mack EE, Ahmad A, Pollack JS (1996) Selenium uptake by sulfur-accumulating bacteria. Geochim Cosmochim Acta 60:3531–3539. [https://doi.org/10.1016/](https://doi.org/10.1016/0016-7037(96)00221-9) [0016-7037\(96\)00221-9](https://doi.org/10.1016/0016-7037(96)00221-9)
- Peak D, Sparks DL (2002) Mechanisms of selenate adsorption on iron oxides and hydroxides. Environ Sci Technol 36:1460– 1466. <https://doi.org/10.1021/es0156643>
- Pereira AS et al (2018) Selenium and silicon reduce cadmium uptake and mitigate cadmium toxicity in *Pfafa glomerata* (Spreng.) Pedersen plants by activation antioxidant enzyme system. Environ Sci Pollut Res 25:18548–18558. [https://doi.org/10.](https://doi.org/10.1007/s11356-018-2005-3) [1007/s11356-018-2005-3](https://doi.org/10.1007/s11356-018-2005-3)
- Pilon-Smits E (2005) Phytoremediation. Annu Rev Plant Biol 56:15– 39. <https://doi.org/10.1146/annurev.arplant.56.032604.144214>
- Pilon-Smits EAH, Winkel LHE, Lin Z-Q (2017) Selenium in Plants. Springer, Cham, Switzerland. [https://doi.org/10.1007/](https://doi.org/10.1007/978-3-319-56249-0) [978-3-319-56249-0](https://doi.org/10.1007/978-3-319-56249-0)
- Prins CN, Hantzis LJ, Quinn CF, Pilon-Smits EAH (2011) Efects of selenium accumulation on reproductive functions in *Brassica juncea* and *Stanleya pinnata*. J Exp Bot 62:5633–5640. [https://](https://doi.org/10.1093/jxb/err247) doi.org/10.1093/jxb/err247
- Priyadarsini KI, Singh BG, Kunwar A, Prabhu P, Jain VK Selenium compounds as antioxidants and radioprotectors. In: Bañuelos GS, Lin Z-Q, Yin X (eds) The 3rd International Conference on Selenium in the Environment and Human Health, Hefei, China, 2013. Taylor & Francis Group, p 37
- Quinn CF et al (2011) Selenium accumulation in fowers and its efects on pollination. New Phytol 192:727–737. [https://doi.](https://doi.org/10.1111/j.1469-8137.2011.03832.x) [org/10.1111/j.1469-8137.2011.03832.x](https://doi.org/10.1111/j.1469-8137.2011.03832.x)
- Ranjard L, Prigent-Combaret C, Nazaret S, Cournoyer B (2002) Methylation of inorganic and organic selenium by the bacterial thiopurine methyltransferase. J Bacteriol 184:3146–3149. <https://doi.org/10.1128/jb.184.11.3146-3149.2002>
- Ranjard L, Nazaret S, Cournoyer B (2003) Freshwater bacteria can methylate selenium through the thiopurine methyltransferase pathway. Appl Environ Microbiol 69:3784–3790. [https://doi.](https://doi.org/10.1128/aem.69.7.3784-3790.2003) [org/10.1128/aem.69.7.3784-3790.2003](https://doi.org/10.1128/aem.69.7.3784-3790.2003)
- Ranjard L, Prigent-Combaret C, Favre-Bonté S, Monnez C, Nazaret S, Cournoyer B (2004) Characterization of a novel selenium methyltransferase from freshwater bacteria showing strong similarities with the calicheamicin methyltransferase. BBA - Gene Str Express 1679:80–85. [https://doi.org/10.1016/j.bba](https://doi.org/10.1016/j.bbaexp.2004.05.001)[exp.2004.05.001](https://doi.org/10.1016/j.bbaexp.2004.05.001)
- Ravi S, Breshears DD, Huxman TE, D'Odorico P (2010) Land degradation in drylands: interactions among hydrologic–aeolian erosion and vegetation dynamics. Geomorphology 116:236–245. <https://doi.org/10.1016/j.geomorph.2009.11.023>
- Razak AA, El-Tantawy H, El-Sheikh HH, Gharieb MM (1991) Infuence of selenium on the efficiency of fungicide action against certain fungi. Biol Trace Elem Res 28:47–56. [https://doi.org/](https://doi.org/10.1007/BF02990462) [10.1007/BF02990462](https://doi.org/10.1007/BF02990462)
- Rios JJ et al (2010) Response of nitrogen metabolism in lettuce plants subjected to diferent doses and forms of selenium. J Sci Food Agric 90:1914–1919. <https://doi.org/10.1002/jsfa.4032>
- Sager M (2006) Selenium in agriculture, food, and nutrition. Pure Appl Chem 78:111–133. [https://doi.org/10.1351/pac2006780](https://doi.org/10.1351/pac200678010111) [10111](https://doi.org/10.1351/pac200678010111)
- Salt DE, Smith RD, Raskin I (1998) Phytoremediation. Annu Rev Plant Physiol Plant Mol Biol 49:643–668. [https://doi.org/10.](https://doi.org/10.1146/annurev.arplant.49.1.643) [1146/annurev.arplant.49.1.643](https://doi.org/10.1146/annurev.arplant.49.1.643)
- Schiavon M, Pilon M, Malagoli M, Pilon-Smits EAH (2015) Exploring the importance of sulfate transporters and ATP sulphurylases for selenium hyperaccumulation-a comparison of *Stanleya pinnata* and *Brassica juncea* (*Brassicaceae*). Front Plant Sci 6:2. <https://doi.org/10.3389/fpls.2015.00002>
- Sharma VK, Mcdonald TJ, Sohn M, Anquandah GAK, Pettine M, Zboril R (2015) Biogeochemistry of selenium. A review. Environ Chem Lett 13:49–58. [https://doi.org/10.1007/](https://doi.org/10.1007/s10311-014-0487-x) [s10311-014-0487-x](https://doi.org/10.1007/s10311-014-0487-x)
- Shrift A, Ulrich JM (1969) Transport of Selenate and Selenite into *Astragalus* Roots. Plant Physiol 44:893–896. [https://doi.org/](https://doi.org/10.1104/pp.44.6.893) [10.1104/pp.44.6.893](https://doi.org/10.1104/pp.44.6.893)
- Singh M, Singh N (1978) Selenium toxicity in plants and its detoxication by phosphorus. Soil Sci 126:255–262. [https://doi.org/](https://doi.org/10.1097/00010694-197811000-00001) [10.1097/00010694-197811000-00001](https://doi.org/10.1097/00010694-197811000-00001)
- Srivastava S, Shanker K, Srivastava S, Shrivastav R, Das S, Prakash S, Srivastava MM (1998) Efect of selenium supplementation on the uptake and translocation of chromium by spinach (*Spinacea oleracea*). Bull Environ Contam Toxicol 60:750–758. <https://doi.org/10.1007/s001289900690>
- Staicu LC et al (2015) *Pseudomonas moraviensis* subsp. stanleyae, a bacterial endophyte of hyperaccumulator *Stanleya pinnata*, is capable of efficient selenite reduction to elemental selenium under aerobic conditions. J Appl Microbiol 119:400–410. <https://doi.org/10.1111/jam.12842>
- Steinbrenner H, Sies H (2013) Selenium homeostasis and antioxidant selenoproteins in brain: Implications for disorders in the central nervous system. Arch Biochem Biophys 536:152–157. <https://doi.org/10.1016/j.abb.2013.02.021>
- Subramanyam K, Du Laing G, Van Damme EJ (2019) Sodium selenate treatment using a combination of seed priming and foliar spray alleviates salinity stress in rice. Front Plant Sci 10:116. <https://doi.org/10.3389/fpls.2019.00116>
- Sun G-X, Meharg AA, Li G, Chen Z, Yang L, Chen S-C, Zhu Y-G (2016) Distribution of soil selenium in China is potentially controlled by deposition and volatilization? Sci Rep 6:20953. <https://doi.org/10.1038/srep20953>
- Sura-de Jong M et al (2015) Selenium hyperaccumulators harbor a diverse endophytic bacterial community characterized by high selenium resistance and plant growth promoting properties. Front Plant Sci 6:113.<https://doi.org/10.3389/fpls.2015.00113>
- Swift MC (2002) Stream ecosystem response to, and recovery from, experimental exposure to selenium. J Aquat Ecosyst Stress Recover 9:159–184.<https://doi.org/10.1023/A:1021299003516>
- Tang H et al (2015) Efects of selenium and silicon on enhancing antioxidative capacity in ramie (*Boehmeria nivea* (L.) Gaud.) under cadmium stress. Environ Sci Pollut Res 22:9999–10008. <https://doi.org/10.1007/s11356-015-4187-2>
- Terry N, Zayed AM, de Souza MP, Tarun AS (2000) Selenium in higher plants. Annu Rev Plant Physiol Plant Mol Biol 51:401– 432.<https://doi.org/10.1146/annurev.arplant.51.1.401>
- Tokunaga TK, Pickering IJ, Brown GE (1996) Selenium transformations in ponded sediments. Soil Sci Soc Am J 60:781–790. <https://doi.org/10.2136/sssaj1996.03615995006000030015x>
- Torma AE, Habashi F (1972) Oxidation of copper (II) selenide by *Thiobacillus ferrooxidans*. Can J Microbiol 18:1780–1781. <https://doi.org/10.1139/m72-278>
- Trelease SF, Beath OA (1951) Selenium: its geological occurrence and its biological efects in relation to botany, chemistry, agriculture, nutrition and medicine. J Geol 59:181. [https://doi.org/](https://doi.org/10.1086/625840) [10.1086/625840](https://doi.org/10.1086/625840)
- Turakainen M, Hartikainen H, Seppänen MM (2004) Efects of selenium treatments on potato (*Solanum tuberosum* L.) growth and concentrations of soluble sugars and starch. J Agric Food Chem 52:5378–5382.<https://doi.org/10.1021/jf040077x>
- Van Hoewyk D, Abdel-Ghany SE, Cohu CM, Herbert SK, Kugrens P, Pilon M, Pilon-Smits EAH (2007) Chloroplast iron-sulfur cluster protein maturation requires the essential cysteine desulfurase CpNifS. Proc Natl Acad Sci USA 104:5686–5691. <https://doi.org/10.1073/pnas.0700774104>
- Vinceti M, Filippini T, Wise LA (2018) Environmental selenium and human health: an update. Curr Environ Health Rep 5:464–485. <https://doi.org/10.1007/s40572-018-0213-0>
- Wang Z, Gao Y (2001) Biogeochemical cycling of selenium in Chinese environments. Appl Geochem 16:1345–1351. [https://doi.org/10.](https://doi.org/10.1016/S0883-2927(01)00046-4) [1016/S0883-2927\(01\)00046-4](https://doi.org/10.1016/S0883-2927(01)00046-4)
- Wang S, Liang D, Wang D, Wei W, Fu D, Lin Z (2012a) Selenium fractionation and speciation in agriculture soils and accumulation in corn (*Zea mays* L.) under feld conditions in Shaanxi Province, China. Sci Total Environ 427–428:159–164. [https://doi.org/10.](https://doi.org/10.1016/j.scitotenv.2012.03.091) [1016/j.scitotenv.2012.03.091](https://doi.org/10.1016/j.scitotenv.2012.03.091)
- Wang Y-D, Wang X, Wong Y-S (2012b) Proteomics analysis reveals multiple regulatory mechanisms in response to selenium in rice. J Proteomics 75:1849–1866. [https://doi.org/10.1016/j.jprot.2011.](https://doi.org/10.1016/j.jprot.2011.12.030) [12.030](https://doi.org/10.1016/j.jprot.2011.12.030)
- Wang D, Zhou F, Yang W, Peng Q, Man N, Liang D (2017) Selenate redistribution during aging in diferent Chinese soils and the dominant infuential factors. Chemosphere 182:284–292. [https://](https://doi.org/10.1016/j.chemosphere.2017.05.014) doi.org/10.1016/j.chemosphere.2017.05.014
- Wang D et al (2019) Efects of straw amendment on selenium aging in soils: mechanism and infuential factors. Sci Total Environ 657:871–881.<https://doi.org/10.1016/j.scitotenv.2018.12.021>
- Wen H, Carignan J (2007) Reviews on atmospheric selenium: emissions, speciation and fate. Atmos Environ 41:7151–7165. [https://](https://doi.org/10.1016/j.atmosenv.2007.07.035) doi.org/10.1016/j.atmosenv.2007.07.035
- Wen H, Carignan J (2009) Ocean to continent transfer of atmospheric Se as revealed by epiphytic lichens. Environ Pollut 157:2790– 2797.<https://doi.org/10.1016/j.envpol.2009.04.021>
- White PJ et al (2004) Interactions between selenium and sulphur nutrition in *Arabidopsis thaliana*. J Exp Bot 55:1927–1937. [https://](https://doi.org/10.1093/jxb/erh192) doi.org/10.1093/jxb/erh192
- Winkel LHE, Johnson CA, Lenz M, Grundl T, Leupin OX, Amini M, Charlet L (2012) Environmental selenium research: from microscopic processes to global understanding. Environ Sci Technol 46:571–579.<https://doi.org/10.1021/es203434d>
- Wu Z-L, Yin X-B, Lin Z-Q, Bañuelos GS, Yuan L-X, Liu Y, Li M (2014) Inhibitory efect of selenium against *Penicillium expansum* and its possible mechanisms of action. Curr Microbiol 69:192–201.<https://doi.org/10.1007/s00284-014-0573-0>
- Wu Z, Bañuelos GS, Lin Z, Liu Y, Yuan L, Yin X, Li M (2015) Biofortifcation and phytoremediation of selenium in China. Front Plant Sci 6:136.<https://doi.org/10.3389/fpls.2015.00136>
- Wu Z et al (2016) Effect of selenium on control of postharvest gray mold of tomato fruit and the possible mechanisms involved. Front Microbiol 6:1441. [https://doi.org/10.3389/fmicb.2015.](https://doi.org/10.3389/fmicb.2015.01441) [01441](https://doi.org/10.3389/fmicb.2015.01441)
- Wu Z et al (2018) Comparison of foliar silicon and selenium on cadmium absorption, compartmentation, translocation and the antioxidant system in Chinese fowering cabbage. Ecotoxicol Environ Saf 166:157–164. [https://doi.org/10.1016/j.ecoenv.2018.09.](https://doi.org/10.1016/j.ecoenv.2018.09.085) [085](https://doi.org/10.1016/j.ecoenv.2018.09.085)
- Xia Q et al (2020) Methods of selenium application diferentially modulate plant growth, selenium accumulation and speciation, protein, anthocyanins and concentrations of mineral elements in purple-grained wheat. Front Plant Sci 11:1114
- Yanke LJ, Bryant RD, Laishley EJ (1995) Hydrogenase I of *Clostridium pasteurianum* functions as a novel selenite reductase. Anaerobe 1:61–67. [https://doi.org/10.1016/S1075-9964\(95\)80457-9](https://doi.org/10.1016/S1075-9964(95)80457-9)
- Yao X, Chu J, He X, Ba C (2011) Protective role of selenium in wheat seedlings subjected to enhanced UV-B radiation. Russ J Plant Physiol 58:283–289. [https://doi.org/10.1134/S10214437110202](https://doi.org/10.1134/S1021443711020257) [57](https://doi.org/10.1134/S1021443711020257)
- Yao Z, Li J, Xie H, Yu C (2012) Review on remediation technologies of soil contaminated by heavy metals. Procedia Environ Sci 16:722–729.<https://doi.org/10.1016/j.proenv.2012.10.099>
- Yao X, Chu J, He X, Liu B, Li J, Yue Z (2013) Efects of selenium on agronomical characters of winter wheat exposed to enhanced ultraviolet-B. Ecotoxicol Environ Saf 92:320–326. [https://doi.](https://doi.org/10.1016/j.ecoenv.2013.03.024) [org/10.1016/j.ecoenv.2013.03.024](https://doi.org/10.1016/j.ecoenv.2013.03.024)
- Yee N, Ma J, Dalia A, Boonfueng T, Kobayashi DY (2007) Se(VI) Reduction and the precipitation of Se(0) by the facultative bacterium *Enterobacter cloacae* SLD1a-1 are regulated by FNR. Appl Environ Microbiol 73:1914–1920. [https://doi.org/10.1128/](https://doi.org/10.1128/AEM.02542-06) [AEM.02542-06](https://doi.org/10.1128/AEM.02542-06)
- Yigit E, Akbulut GB, Gok Y, Bayram D (2012) The efects of organic selenium on some physiological and biochemical parameters in *Hordeum Vulgare* L. and *Triticum Aestivum* L. exposed to salt stress. Fresenius Environ Bull 21:743–747
- Zahedi SM, Abdelrahman M, Hosseini MS, Hoveizeh NF, Tran L-SP (2019) Alleviation of the efect of salinity on growth and yield of strawberry by foliar spray of selenium-nanoparticles. Environ Pollut 253:246–258. [https://doi.org/10.1016/j.envpol.2019.](https://doi.org/10.1016/j.envpol.2019.04.078) [04.078](https://doi.org/10.1016/j.envpol.2019.04.078)
- Zawislanski PT, Zavarin M (1996) Nature and rates of selenium transformations: a laboratory study of kesterson reservoir soils. Soil Sci Soc Am J 60:791–800. [https://doi.org/10.2136/sssaj1996.](https://doi.org/10.2136/sssaj1996.03615995006000030016x) [03615995006000030016x](https://doi.org/10.2136/sssaj1996.03615995006000030016x)
- Zayed A, Lytle CM, Terry N (1998) Accumulation and volatilization of diferent chemical species of selenium by plants. Planta 206:284– 292. <https://doi.org/10.1007/s004250050402>
- Zeng H, Uthus EO, Combs GF Jr (2005) Mechanistic aspects of the interaction between selenium and arsenic. J Inorg Biochem 99:1269–1274.<https://doi.org/10.1016/j.jinorgbio.2005.03.006>
- Zhang Y, Zahir ZA, Frankenberger WT (2004) Fate of colloidal-particulate elemental selenium in aquatic systems. J Environ Qual 33:559–564. <https://doi.org/10.2134/jeq2004.5590>
- Zhang L, Ackley AR, Pilon-Smits EAH (2007) Variation in selenium tolerance and accumulation among 19 *Arabidopsis thaliana* accessions. J Plant Physiol 164:327–336. [https://doi.org/10.](https://doi.org/10.1016/j.jplph.2006.01.008) [1016/j.jplph.2006.01.008](https://doi.org/10.1016/j.jplph.2006.01.008)
- Zhang H et al (2012) Selenium in soil inhibits mercury uptake and translocation in rice (*Oryza sativa* L.). Environ Sci Technol 46:10040–10046.<https://doi.org/10.1021/es302245r>
- Zhang L et al (2014a) OsPT2, a phosphate transporter, is involved in the active uptake of selenite in rice. New Phytol 201:1183–1191. <https://doi.org/10.1111/nph.12596>
- Zhang M, Tang S, Huang X, Zhang F, Pang Y, Huang Q, Yi Q (2014b) Selenium uptake, dynamic changes in selenium content and its infuence on photosynthesis and chlorophyll fuorescence in rice (*Oryza sativa* L.). Environ Exp Bot 107:39–45. [https://doi.org/](https://doi.org/10.1016/j.envexpbot.2014.05.005) [10.1016/j.envexpbot.2014.05.005](https://doi.org/10.1016/j.envexpbot.2014.05.005)
- Zhao XQ, Mitani N, Yamaji N, Shen RF, Ma JF (2010) Involvement of silicon infux transporter OsNIP2;1 in selenite uptake in rice. Plant Physiol 153:1871–1877. [https://doi.org/10.1104/pp.110.](https://doi.org/10.1104/pp.110.157867) [157867](https://doi.org/10.1104/pp.110.157867)
- Zhao J et al (2013) Selenium inhibits the phytotoxicity of mercury in garlic (*Allium sativum*). Environ Res 125:75–81. [https://doi.org/](https://doi.org/10.1016/j.envres.2013.01.010) [10.1016/j.envres.2013.01.010](https://doi.org/10.1016/j.envres.2013.01.010)
- Zhong Y, Cheng JJ (2017) Effects of selenite on unicellular green microalga *Chlorella pyrenoidosa*: bioaccumulation of selenium, enhancement of photosynthetic pigments, and amino acid production. J Agric Food Chem 65:10875–10883. [https://doi.org/](https://doi.org/10.1021/acs.jafc.7b04246) [10.1021/acs.jafc.7b04246](https://doi.org/10.1021/acs.jafc.7b04246)
- Zhu Y-G, Pilon-Smits EAH, Zhao F-J, Williams PN, Meharg AA (2009) Selenium in higher plants: understanding mechanisms for biofortifcation and phytoremediation. Trends Plant Sci 14:436– 442. <https://doi.org/10.1016/j.tplants.2009.06.006>
- Zuber H, Davidian J-C, Wirtz M, Hell R, Belghazi M, Thompson R, Gallardo K (2010) *Sultr4;1* mutant seeds of Arabidopsis have an enhanced sulphate content and modifed proteome suggesting metabolic adaptations to altered sulphate compartmentalization. BMC Plant Biol 10:78.<https://doi.org/10.1186/1471-2229-10-78>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.