



# Preliminary Assessment of Legacy and Current-Use Pesticides in Franciscana Dolphins from Argentina

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## Abstract

The change towards intensive agriculture has led to an increase in the use of pesticides. In addition, legacy pesticides, such as organochlorines are still present in the environment. Ten Franciscana dolphins were accidentally killed by netting in a coastal area of Argentina in Buenos Aires province. From these animals, organochlorine, organophosphate and pyrethroid pesticides were analyzed in liver, bubbler and melon tissues. The concentrations of  $\Sigma$ endosulfan ranged from not detectable values (nd) to 3539 ng g<sup>-1</sup> lw, with the maximum level in melon tissue. DDE was present in 60% of all samples at concentrations from nd to 6672 ng g<sup>-1</sup> lw, indicating historical DDT contamination. The presence of endosulfan and heptachlor in a nursing calf indicated a transfer of these pesticides through lactational and placental transport. The concentrations of organophosphates and pyrethroids were below the limit of detection, reflecting the low persistence of these compounds.

**Keywords** Coastal dolphin · Organochlorines · Organophosphates · Pyrethroids · Southwestern Atlantic Ocean · Biomonitor

Pesticides are one of the hazardous groups of chemicals, which are known to interfere with many vital functions of organisms. Although some organochlorine (OCs) pesticides, such as chlordanes, dieldrin and endosulfan, are banned by the UNEP Stockholm Convention, they are still in the environment due to their high persistence. On the other hand, current use pesticides (CUPs; e.g. chlorpyrifos, diazinon, cypermethrin, thionazin) are large-scale replacements for globally banned legacy OCs pesticides, but relatively few field studies have investigated the presence of these compounds in the marine environment (Morris et al. 2016; Weber et al. 2010).

Argentina is a farming country and the gradual change towards modern and intensive agricultural activities has led to an increase in the use of pesticides (Pórfido 2014).

Estuarine and marine environments are the final receptors of most of these compounds, with an impact on the ecosystem (Arias et al. 2011). Odontocete cetaceans have certain characteristics that contribute to the accumulation of pesticides in their tissues (top predators, longevity, late maturity, large lipid reserves in fat, high metabolic rates) (Dirtu et al. 2016; Fair et al. 2010). Franciscana dolphin (*Pontoporia blainvillei*) is a small and endemic marine mammal that inhabits the Southwestern Atlantic Ocean. Its geographical distribution ranges from Itaúnas (18°25'S, 30°42'W, Brazil; Siciliano 1994) to Golfo Nuevo (42°35'S, 64°48'W, Argentina; Bastida et al. 2007). The International Union for Conservation of Nature (IUCN) has categorized the species as Vulnerable A3d throughout its geographical range being the most anthropogenically impacted small cetacean (Secchi and Wang 2002). Due to their coastal and estuarine habits, Franciscana dolphins from Argentina inhabit areas with intense human activity which poses several threats to their conservation.

The adverse effects of pesticides in marine mammals is difficult to assess (Dirtu et al. 2016); however, several studies have linked effects on the reproductive (Murphy et al. 2015), endocrine (Hoydal et al. 2016) and immune system (Lehnert et al. 2016), and also with carcinogenesis (Martineau et al. 1994) and skeletal abnormalities (Mortensen

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et al. 1992). Few studies have evaluated pesticides in cetaceans in Argentinean waters (Borrell et al. 1990, 1995; Castello et al. 1997; Durante et al. 2016; Torres et al. 2015), and there are no reports on organophosphates (OPs) and pyrethroid pesticides. Therefore, the aim was to determine the presence of OCs, OPs and pyrethroids pesticides in blubber, melon and liver of *Pontoporia blainvillei* from Argentina.

## Materials and Methods

Ten Franciscana dolphins were collected from coastal area of Buenos Aires Province between 2008 and 2011 (Fig. 1). Dolphins were incidentally captured in artisanal fishing nets, being entangled for a period less than 10 h before sampling. The quality of the carcasses was evaluated according to Geraci and Lounsbury (2005), to determine their suitability for the intended study. Total length, weight, and sex were determined for each specimen. Samples of the liver, bubbler and melon were collected and stored at  $-80^{\circ}\text{C}$  until analysis. Age was estimated by Denuncio (2012) using dentine and cementum dental layers to determine growth layer groups (GLGs). Franciscana dolphins were grouped into three age classes: calf, juvenile and adult according to Denuncio 2012.

The extraction and cleaned-up of pesticides in liver was performed by the QuEChERS method described by Anastassiades et al. (2003) with modifications, based on a solid dispersive phase extraction (dSPE). Briefly, a portion of the liver was cut into small pieces (approximately 2 mm) and homogenized until a uniform homogenate was obtained. Approximately 2 g of tissue were weighed into a 50 mL tube, spiked with internal standard *cis*-nonachlor solution (Sigma-Aldrich, Pestanal), and sonicated for 10 min. After this, 15 mL of acetonitrile (ACN)

and 15 mL of hexane were added. The tube was shaken vigorously for 2 min and centrifuged at 5000 rpm. The ACN phase was transferred to a 10 mL beaker and evaporated to 1 mL with gaseous nitrogen. A clean up with previously heat activated florisil (Sigma-Aldrich, Pestanal), PSA y C18 was performed and the supernatant was concentrated under a nitrogen flow until near dryness and re-dissolved in 200  $\mu\text{L}$  of acetone.

QuEChERS technique was developed for samples with a high water content. Due to the high lipid content in blubber and melon, it was not possible to apply the method described above. Therefore, approximately 1 g of tissue was weighed and extracted with sulfur ether in an automated hot Soxhlet extractor for 5 h, and the lipid content was determined gravimetrically. The clean-up was performed in a column (C18, Enviro Clean Extraction Columns), previously activated with ACN. The cleaned eluates were concentrated under a nitrogen flow until near dryness and re-dissolved in 200  $\mu\text{L}$  of acetone. Detection and quantification of compounds in liver, bubbler and melon extracts were performed by Gas Chromatography-Mass Spectrometry (GC-MS, Agilent 7890).

For quality control/quality assurance (QC/QA) purposes, samples were fortified with  $0.05\text{ mg kg}^{-1}$  of the analytical standard *cis*-nonachlor solution (Sigma-Aldrich, Pestanal), and the recovery were between 93.0 and 101.5%. In addition, blanks of solution and matrix were checked in each batch of procedure; both were below the detection limits for all pesticides analyzed. The regression values of the standard curves for the compounds were  $>0.999$  (95% of confidence). The slope and adjustment were checked by the Significance for Linear Regression and Lack of Fit tests, respectively ( $p < 0.01$ ). The precision was checked through the repeatability in batch of 10 determinations, and the %RSD obtained was  $<6.5$  for all pesticides. The limit of detection achieved for OCs was  $10\text{ ng g}^{-1}$ , and for OPs and pyrethroids was  $30\text{ ng g}^{-1}$ . Concentrations lower than these values were considered as not detectable (nd). The results were expressed in  $\text{ng g}^{-1}\text{ lw}$

In order to perform statistical analysis, a value equal to half of the detection limit was arbitrarily assigned for those samples that showed values below this limit. Normality was tested using the Kolmogorov–Smirnov test, whereas homoscedasticity with Levene test. Statistical differences were verified using non-parametric tests (Mann Whitney and Kruskal Wallis tests). All analysis were performed with the STATISTICA® 6.0 program (Statsoft, Inc.).

## Results and Discussion

All carcasses were in good condition (Code 2; Geraci and Lounsbury 2005), indicated by the presence of rigor mortis. Biometric measurements of the specimens and the concentrations of detectable pesticides are shown in Table 1.

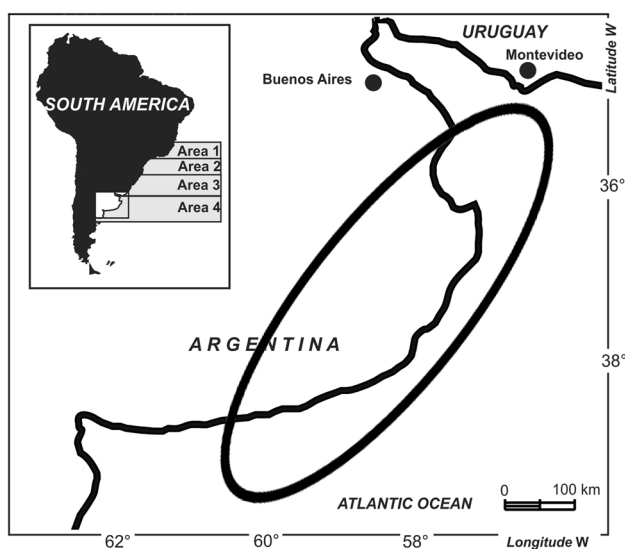


Fig. 1 Study area in Argentinean continental shelf

The following pesticides showed concentrations below the detection limit in all samples: aldrin, *cis*-chlordane, *trans*-chlordane, p,p'-DDT, dieldrin, endrin, heptachlor epoxy, hexachlorobenzene, *cis*-nonaol, *trans*-nonaol, acephate, azinphos methyl, chlorpyrifos ethyl, chlorpyrifos methyl, diazinon, dichlofenthion, dichlorvos, dimethoate, disulfoton, fenitrothion, phosfos, malathion, metamidofos, methidathion, monocrotophos, parathion ethyl, parathion methyl, pirimiphos methyl, thionazin, cypermethrin, deltamethrin, fenvalerate, permethrin.

The concentrations of  $\Sigma$ endosulfan (corresponding to endosulfan I – EI-, endosulfan II – EII- and endosulfan sulfate -ES-) presented range values of nd–275.97 ng g<sup>-1</sup> lw in blubber, nd–3539.57 ng g<sup>-1</sup> lw in melon, and nd–88,341.03 ng g<sup>-1</sup> lw in liver. Endosulfan compounds were found in at least one of the tissues in all Franciscana dolphins; and its presence was related to a wide use of the pesticide in Argentina and recent prohibition in 2013. Information related to endosulfan and its metabolites in cetaceans is limited. In *P.*

*blainvillei* there is only one report of blubber concentrations of EII in Brazil, and they were below the limit of detection (Leonel et al. 2010). Besides, the blubber concentrations of EI and EII in Franciscana dolphin were higher than those reported for bottlenose whale – *Hyperoodon ampullatus* –, bottlenose dolphin – *Tursiops truncatus* –, striped dolphin – *Stenella coeruleoalba* – and white-sided dolphin – *Lagenorhynchus acutus* – (Table 2).

p,p'-DDE metabolite was present as a unique representative of the DDT group in 60% of all samples, p,p'-DDD was present only in one specimen (Table 1), and no DDT was detected. The concentrations of DDE found in Franciscana dolphin were within the range reported for the species and other cetaceans (Table 2). In Argentina, DDT was banned in 1992, and the predominance of the metabolite with respect to the original compound indicates a not recent entry of the pesticide into the environment.

Heptachlor was found only in 30% of dolphins, with values much higher than those reported for other cetaceans

**Table 1** Biological information and concentrations of pesticides (ng g<sup>-1</sup> lw) in the blubber (B), melon (M) and liver (L) of Franciscana dolphins Diet information was obtained from Denuncio (2012)

Code	Sex	Age class	Estimated age (years)	Diet	Total length (cm)	Weight (kg)	Tissue	Pesticide	Concentration
D1	F	Adult	6.7	Solid	140	29.0	B	ES	25.5
							L	ES	1973.6
							B	DDE	267.6
D2	M	Juvenile	2.9	Solid	115	22.1	B	DDE	22.1
							B	Heptachlor	64.2
							M	EI	363.4
D3	F	Calf	<1	Milk	74	4.1	M	EII	545.1
							B	EII	131.8
							L	ES	8773.6
D4	M	Calf	<1	Mixed	84	9.7	B	Heptachlor	70.3
							L	ES	6578.6
							D5	M	Juvenile
B	DDE	246.1							
M	DDE	61.3							
D6	F	Adult	10.5	Solid	139	18.6	M	DDD	735.0
							M	DDE	178.4
							L	DDE	6672.6
D7	F	Juvenil	2.7	Solid	109	na	L	EII	2067.6
							L	ES	8928.1
							B	DDE	24.9
D8	F	Adult	4	Solid	136	31.1	M	EII	230.0
							L	EI	88341.0
							L	ES	12217.4
D9	F	Adult	7	Solid	126	22.5	L	Heptachlor	6484.6
							M	EII	1461.1
							D10	M	Juvenile
L	DDE	1879.6							

M male, F female

**Table 2** Concentrations of pesticides (ng g<sup>-1</sup> lw) reported in other cetacean species

Specie	T	Compound					Reference
		DDE	Endosulfan I	Endosulfan II	Endosulfan sulfate	Heptachlor	
<i>P. blainvillei</i>	B			nd		nd	Leonel et al. (2010)
	B	M: 0.79 ± 0.71 F: 0.75 ± 0.40					Castello et al. (1997)
	B	M: 1.97 ± 1.27 F: 0.3 ± 0.18					Borrell et al. (1990, 1995, 1997)
<i>Tursiops truncatus</i>	B	1.4–22.72	0.85–1.11	2.06–27.39	3.05–37.29	1.11–12.57	Delgado-Estrella et al. (2015)
	B	M: 188–14,300 F: 5500–56,300	M: 0.4–4.1 F: 0.5–5	nd	M: 1.1–2.6 F: 1.1–5.1		Fair et al. (2010)
	B	M: 9952–80,252 F: 1163–15,042		nd	nd		Hansen et al. (2004)
	L	0.041–9.78					Shoham-Frider et al. (2009)
<i>Hyperoodon ampullatus</i>	B	M: 8218 ± 5846 F: 1887 ± 506	M: 0.9 ± 2.3 F: 15.8 ± 28.7	M: 73.2 ± 78.3	M: 28.3 ± 25.8 F: 27.7 ± 18.7	M: 1.6 ± 3.7 F: 1.1 ± 1.3	Hooker et al. (2008)
	B		0.01–0.33				Tuerk et al. (2005)
<i>Lagenorhynchus acutus</i> <sup>a</sup>	B						
<i>Stenella coeruleoalba</i>	B		4.6–134			1.6–17	Wafo et al. (2012)

Data are presented as ranges or as mean ± SD

nd not detectable, T tissue, B blubber, L liver, M male, F female

<sup>a</sup>Concentration expressed in ng g<sup>-1</sup> ww

(Table 1). Its metabolite heptachlor epoxide was not detected, although it is considered the main compound found in marine mammals (Dierauf and Gulland 2001). This pesticide was widely used in Argentina as an insecticide in potato crops (Miglioranza et al. 2003), and although its use was banned in 1992, the presence of the parent compound in *P. blainvillei* tissues indicates a recent contribution of the contaminant to the environment.

The D3 dolphin, a nursing calf, presented endosulfan compounds and heptachlor in its tissues, indicating a transference of the pesticide through milk and/or previous placental exposure. The concentrations of these pesticides were similar to adult Franciscana dolphins. The mother-calf transfer of organochlorine pesticides has been reported for other dolphin species (Stockin et al. 2007).

The OPs and pyrethroid pesticides analyzed were found below the limit of detection in Franciscana dolphins. Information about the presence of these contaminants in cetaceans is scarce, due to the low environmental stability and persistence of these compounds compared to OCs. Hernández et al. (2000) reported non-detectable levels of OPs in the fin whale (*Balaenoptera physalus*), which is in accordance with the results in the Franciscana dolphin. By other way, Alonso et al. (2012, 2015) reported the presence of 12 pyrethroids in tissues of the species in Brazilian waters, although several samples presented levels below the detection limit. Transplacental and lactational transfer was also found. The

authors proposed that the compounds are detected in the first moments of life, during pregnancy and lactation. Although when dolphins reach sexual maturity the pyrethroids would begin to be metabolized, and consequently the decrease or disappearance in the organism would be evident.

The presence of organochlorine pesticides in Franciscana dolphin is related to their persistence in the environment and low metabolic degradation. The concentrations of organophosphates and pyrethroids below the limit of detection reflected the low persistence of these compounds in the environment. In addition, the species could be considered as a good biomonitor of pesticides in Argentinean waters. On the other hand, it is necessary to carry out more detailed in-time studies of the profiles of CUPs in Franciscana dolphin, in order to reach a better understanding of the possible biological effects and the risks for the species in the present.

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## Compliance with Ethical Standards

**Conflict of interest** The authors declare that they have no conflict of interest.

## References

- Alonso MB, Feo ML, Corcellas C, Vidal LG, Bertozzi CP, Marigo J, Sechhi E, Bassoi M, Azevedo A, Dorneles P, Torres JP, Lailson-Brito J, Malm O, Eljarrat E, Barceló D (2012) Pyrethroids: a new threat to marine mammals? *Environ Int* 47:99–106. <https://doi.org/10.1016/j.envint.2012.06.010>
- Alonso MB, Feo ML, Corcellas C, Gago-Ferrero P, Bertozzi CP, Marigo J, Flach L, Meirelles AC, Carvalho VL, Azevedo AF, Torres JP, Lailson-Brito J, Malm O, Diaz-Cruz MS, Eljarrat E, Barceló D (2015) Toxic heritage: maternal transfer of pyrethroid insecticides and sunscreen agents in dolphins from Brazil. *Environ Pollut* 207:391–402
- Anastassiades M, Lehotay S, Štajnbaher D, Schenck F (2003) Fast and easy multiresidue method employing acetonitrile extraction/partitioning and “dispersive solid-phase extraction” for the determination of pesticide residues in produce. *J AOAC Int* 86:412–431
- Arias AH, Pereyra MT, Marcovecchio JE (2011) Multi-year monitoring of estuarine sediments as ultimate sink for DDT, HCH, and other organochlorinated pesticides in Argentina. *Environ Monit Assess* 172:17–32. <https://doi.org/10.1007/s10661-010-1315-9>
- Bastida R, Rodríguez D, Secchi E, da Silva V (2007) Mamíferos Acuáticos de Sudamérica y Antártida. Vazquez Mazzini, Buenos Aires
- Borrell A, Aguilar A, Corcuera J, Monzón F (1990) Distribution of organochlorines in tissues and organs of the franciscana (*Pontoporia blainvillei*). *Eur Res Cetaceans* 4:111–113
- Borrell A, Pastor T, Aguilar A, Corcuera J, Monzón F (1995) DDTs and PCBs in La Plata dolphins (*Pontoporia blainvillei*) from Argentina: age and sex trends. *Eur Res Cetaceans* 8:273–276
- Borrell A, Pastor T, Aguilar A, Corcuera J, Monzón F (1997) Contaminación por DDTs y PCBs en *Pontoporia blainvillei* de aguas argentinas: variación con la edad y el sexo. In: Pinedo MC, Barreto AS (eds) Anais do 2o. Encontro sobre a Coordenação de Pesquisa e Manejamento da Franciscana. Ed. FURG, Rio Grande, pp 62–69
- Castello HP, Junin M, Rotman F, Sartí GC (1997) Análisis de contaminantes organoclorados y metales pesados en Franciscana, *Pontoporia blainvillei*, de Argentina y Brasil. Documento de Trabajo 15 del Tercer Taller para la Coordinación de la Investigación y Conservación de la Franciscana en el Atlántico Sudoccidental
- Delgado-Estrella A, Barreto-Castro MD, Acevedo-Olvera G, Vázquez-Maldonado LE (2015) Effects of pollutant discharges on the aquatic mammal populations of Terminos Lagoon, Campeche, Mexico. *WIT Trans Ecol Environ* 200:229–235. <https://doi.org/10.2495/WS150191>
- Denuncio P (2012) Biología y conservación del delfín del Plata (*Pontoporia blainvillei*) en el sector costero bonaerense. Dissertation, University of Mar del Plata
- Dierauf LA, Gulland FM (2001) Handbook of marine mammal medicine. CRC Press, Boca Raton
- Dirtu AC, Malarvannan G, Das K, Dulau-Drouot V, Kiszka JJ, Lepoint G, Mongin P, Covaci A (2016) Contrasted accumulation patterns of persistent organic pollutants and mercury in sympatric tropical dolphins from the south-western Indian Ocean. *Environ Res* 146:263–273. <https://doi.org/10.1016/j.envres.2016.01.006>
- Durante CA, Santos-Neto EB, Azevedo A, Crespo EA, Lailson-Brito J (2016) POPs in the South Latin America: bioaccumulation of DDT, PCB, HCB, HCH and Mirex in blubber of common dolphin (*Delphinus delphis*) and Fraser’s dolphin (*Lagenodelphis hosei*) from Argentina. *Sci Total Environ* 572:352–360. <https://doi.org/10.1016/j.scitotenv.2016.07.176>
- Fair PA, Adams J, Mitchum G, Hulsey TC, Reif JS, Houde M, Muir D, Wirth E, Wetzel D, Zolman E, McFee W, Bossart GD (2010) Contaminant blubber burdens in Atlantic bottlenose dolphins (*Tursiops truncatus*) from two southeastern US estuarine areas: Concentrations and patterns of PCBs, pesticides, PBDEs, PFCs, and PAHs. *Sci Total Environ* 408:1577–1597. <https://doi.org/10.1016/j.scitotenv.2009.12.021>
- Geraci JR, Lounsbury VJ (2005) Marine mammals ashore: a field guide for strandings. Texas A&M University, Sea Grant College Program, Texas
- Hansen LJ, Schwacke LH, Mitchum GB, Hohn AA, Wells RS, Zolman ES, Fair PA (2004) Geographic variation in polychlorinated biphenyl and organochlorine pesticide concentrations in the blubber of bottlenose dolphins from the US Atlantic coast. *Sci Total Environ* 319:147–172. [https://doi.org/10.1016/S0048-9697\(03\)00371-1](https://doi.org/10.1016/S0048-9697(03)00371-1)
- Hernández F, Serrano R, Roig-Navarro AF, Martínez-Bravo Y, López FJ (2000) Persistent organochlorines and organophosphorus compounds and heavy elements in common whale (*Balaenoptera physalus*) from the Western Mediterranean Sea. *Mar Pollut Bull* 40:426–433. [https://doi.org/10.1016/S0025-326X\(99\)00238-6](https://doi.org/10.1016/S0025-326X(99)00238-6)
- Hooker SK, Metcalfe TL, Metcalfe CD, Angell CM, Wilson JY, Moore MJ, Whitehead H (2008) Changes in persistent contaminant concentration and CYP1A1 protein expression in biopsy samples from northern bottlenose whales, *Hyperoodon ampullatus*, following the onset of nearby oil and gas development. *Environ Pollut* 152:205–216. <https://doi.org/10.1016/j.envpol.2007.05.027>
- Hoydal KS, Ciesielski TM, Borrell A, Wasik A, Letcher RJ, Dam M, Jenssen BM (2016) Relationships between concentrations of selected organohalogen contaminants and thyroid hormones and vitamins A, E and D in Faroese pilot whales. *Environ Res* 148:386–400. <https://doi.org/10.1016/j.envres.2016.04.012>
- Lehnert K, Ronnenberg K, Weijs L, Covaci A, Das K, Hellwig V, Siebert U (2016) Xenobiotic and immune-relevant molecular biomarkers in harbor seals as proxies for pollutant burden and effects. *Arch Environ Contam Toxicol* 70:106–120
- Leonel J, Sericano JL, Fillmann G, Secchi E, Montone RC (2010) Long-term trends of polychlorinated biphenyls and chlorinated pesticides in franciscana dolphin (*Pontoporia blainvillei*) from Southern Brazil. *Mar Pollut Bull* 60:412–418. <https://doi.org/10.1016/j.marpolbul.2009.10.011>
- Martineau D, De Guise S, Fournier M, Shugart L, Girard C, Lagace A, Beland P (1994) Pathology and toxicology of beluga whales from the St. Lawrence Estuary, Quebec, Canada: past, present and future. *Sci Total Environ* 154:201–215. [https://doi.org/10.1016/0048-9697\(94\)90088-4](https://doi.org/10.1016/0048-9697(94)90088-4)
- Miglioranza KS, Aizpún JE, Moreno VJ (2003) Dynamics of organochlorine pesticides in soils from a southeastern region of Argentina. *Environ Toxicol Chem* 22:712–717
- Morris AD, Muir DC, Solomon KR, Letcher RJ, McKinney MA, Fisk AT, McMeans BC, Tomy GT, Teixeira C, Wang X, Duric M (2016) Current-use pesticides in seawater and their bioaccumulation in polar bear–ringed seal food chains of the Canadian Arctic. *Environ Toxicol Chem* 35:1695–1707. <https://doi.org/10.1002/etc.3427>
- Mortensen P, Bergman A, Bignert A, Hansen HJ, Harkonen T, Olsson M (1992) Prevalence of skull lesions in harbor seals (*Phoca vitulina*) in Swedish and Danish museum collections: 1835–1988. *Ambio* 21:520–524
- Murphy S, Barber JL, Learmonth JA, Read FL, Deaville R, Perkins MW, Brownlow A, Davison N, Penrose R, Pierce GJ, Law RJ, Jepson PD (2015) Reproductive failure in UK harbour porpoises *Phocoena phocoena*: legacy of pollutant exposure? *PLoS ONE* 10:e0131085. <https://doi.org/10.1371/journal.pone.0131085>
- Pórfido OD (2014) Los plaguicidas en la República Argentina. Ministerio de Salud de la Nación, Buenos Aires
- Secchi E, Wang J (2002) Assessment of the conservation status of a Franciscana (*Pontoporia blainvillei*) stock in the Franciscana Management Area III following the IUCN red list process. *Lat Am J Aquat Mamm* 1:183–190. <https://doi.org/10.5597/lajam00023>

- Shoham-Frider E, Kress N, Wynne D, Scheinin A, Roditi-Elsar M, Kerem D (2009) Persistent organochlorine pollutants and heavy metals in tissues of common bottlenose dolphin (*Tursiops truncatus*) from the Levantine Basin of the Eastern Mediterranean. *Chemosphere* 77:621–627. <https://doi.org/10.1016/j.chemosphere.2009.08.048>
- Siciliano S (1994) Review of small cetaceans and fishery interactions in coastal waters of Brazil. *Rep Int Whal Comm* 15:241–250
- Stockin KA, Law RJ, Duignan PJ, Jones GW, Porter L, Mirimin L, Meynier L, Orams MB (2007) Trace elements, PCBs and organochlorine pesticides in New Zealand common dolphins (*Delphinus* sp.). *Sci Total Environ* 387:333–345. <https://doi.org/10.1016/j.scitotenv.2007.05.016>
- Torres P, Miglioranza KS, Uhart MM, Gonzalez M, Commendatore M (2015) Organochlorine pesticides and PCBs in southern right whales (*Eubalaena australis*) breeding at Península Valdés, Argentina. *Sci Total Environ* 518–519:605–615. <https://doi.org/10.1016/j.scitotenv.2015.02.064>
- Tuerk KJ, Kucklick JR, McFee WE, Pugh RS, Becker PR (2005) Factors influencing persistent organic pollutant concentrations in the Atlantic white-sided dolphin (*Lagenorhynchus acutus*). *Environ Toxicol Chem* 24:1079–1087
- Wafo E, Mama C, Risoul V, Schembri T, Dhermain F, Portugal H (2012) Chlorinated pesticides in the bodies of dolphins of the French Mediterranean coastal environment. *Adv Environ Sci Int J Bioflux Soc* 4:29–35
- Weber J, Halsall CJ, Muir D, Teixeira C, Small J, Solomon K, Hermanson M, Hung H, Bidleman T (2010) Endosulfan, a global pesticide: a review of its fate in the environment and occurrence in the Arctic. *Sci Total Environ* 408:2966–2984. <https://doi.org/10.1016/j.scitotenv.2009.10.077>