

Lead in the Red-Crowned Cranes (*Grus japonensis*) in Zhalong Wetland, Northeastern China: A Report

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Abstract The dietary uptake of Cd and Pb may contribute to the decline of migratory red-crowned cranes (Grus japonensis) on the Asian mainland. To uncover the relevance of this hypothesis, we determined the concentrations of Pb and Cd as well as further macro and trace elements (Ca, Mg, Cu, Zn, Hg and As) in the gastric contents, gastric wall, intestinal wall, liver, kidney, muscle, and feathers of two individuals found dead in Zhalong Wetland in Northeastern China. Indeed, the Pb concentrations in the liver and kidney tissues was with 31.4 and 60.3 mg kg⁻¹ dry weight (dw), respectively, above concentrations considered as potentially toxic level in common birds (i.e. 30 mg kg^{-1}). These Pb concentration may have possibly been associated with lethal toxicosis in this endangered species suggesting Pb as major threat for G. japonensis populations. Thus, the inputs of Pb into Zhalong Wetland should be reduced to maintain and reestablish environmental conditions supporting the population development of these migratory red-crowned cranes in the Zhalong Wetland, a critical crane habitat for the long-term sustainability of this species.

Keywords Red-crowned crane · Stomach content · Tissue concentration · Toxic effect

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Excessive exposure to trace elements has been extensively investigated because of their toxic effects on biotic systems (Burger and Gochfeld 1993; Eisler 2000). Thus, the assessment of trace element concentrations in birds may provide early warning of changes in their population health (Friend 1985; Dauwea et al. 2005; Scheuhammer 1996). The liver and kidneys of wild animals are commonly used for such evaluations (Wiemeyer and Withers 2004; Orłowski et al. 2012; Kim and Oh 2014), because most toxic trace elements mainly accumulate in these organs (Agusa et al. 2005). However, most baseline tissue levels from environmental exposure and lethal thresholds are described in common birds or mammals. Whether these established thresholds are applicable to rare water birds, such as cranes, remains unknown. For example, Pb toxicosis is a commonly reported cause of toxicity among birds, but lethal levels are variable among tissues and species (Locke and Thomas 1996). For instance, a Pb concentration of 72 mg kg⁻¹ dry weight (dw) in the liver of an endangered whooping crane (Grus americana) is associated with poisoning (Snyder et al. 1992). Pb concentrations ranging from 15.5 to 17.7 mg kg⁻¹ dw have been detected in the kidney of dead nestling rooks (Corvus grugilegus) (Orłowski et al. 2012).

Red-crowned crane (*Grus japonensis*) is an endangered species in East Asia with a small global population of 2750 mature individuals (BirdLife International 2012). The resident population in Japan remains stable (Teraoka et al. 2007), while the migratory population of the Asian mainland has declined due to the loss and degradation of their habitat (i.e. wetlands) for agricultural and industrial development (Harris 2008). Thus, red-crowned cranes increasingly rely on agricultural lands in northeastern China, placing the cranes at greater risk for poisoning (Su and Zhou 2012; Luo et al. 2014a, b). A previous study

Site	Habitat	Finding date	Sex	Adult or not	Body W. (kg)
Control c	ranes				
S 1	Reed marsh	Apr-2014	Male	Adult	9.8
S2	Reed marsh	Apr-2014	Male	Juvenile	6.2
S 3	Reed marsh	Apr-2010	Male	Adult	9.4
S4	Meadow	Oct-2010	Male	Juvenile	6.8
S5	Meadow	Nov-2012	Female	Juvenile	6.5
S6	reed marsh	May-2011	Female	Adult	6.5
S 7	Meadow	Apr-2014	Female	Juvenile	5.8
S 8	Meadow	Oct-2012	Male	Adult	8.3
Poisoned	cranes				
P1	Reed marsh (approx.	Jul-2014	Male	Adult	7.9
	460 m to a corn land)		Female	Adult	6.4

Table 1 Habitat, finding date, and body weight of control cranes and poisoned cranes



Fig. 1 Location of Wuyur catchments (a, b) and sampling sites of poisoned cranes (P1) and control cranes $(S \times)$ (c)

found a common inorganic pesticide and relatively high Pb and Cd concentrations in the sediment and prey of redcrowned cranes in this region (Luo et al. 2013, Luo et al. 2014a, Luo et al. 2014b). These conditions probably enhance the exposure of these cranes to potentially toxic doses of pesticides and trace elements (Luo et al. 2015a). However, published toxicological analyses among redcrowned cranes have been rarely performed.

Our study aimed to determine the distribution and concentration of trace elements in tissues of two dead red-

crowned cranes. The findings from this case study may be used to optimize environmental management.

Materials and Methods

Two moribund red-crowned cranes were collected in the field in July 2014 in Zhalong Wetland (Table 1). Before the cranes' death, they exhibited loss of appetite, lethargy, weakness, emaciation, tremors, and drooped wings. The

cranes had neither wounds nor external parasites, sug- gesting the birds likely consumed toxic substances or suffer from disease. The carcasses of the birds were immediately transported under refrigeration to the labora-
tory for dissection. An infectious disease was ruled out on
the basis of gross pathology beyond the emaciated state of
the birds.
Polyethylene gloves were used throughout the dissec-
tion procedure to prevent external contamination. The
proventriculus and ventriculus (stomach) of the red-
crowned cranes were dissected from between the esopha-
gus and small intestine. Each stomach was incised in half
with a razor blade and the contents were flushed into a
Petri dish for inspection. Likely lead-containing particles,
such as shotgun pellets, were not detected in the stomach.
Food items were examined and sorted using a zoom
stereomicroscope and divided into three categories: ani-
mal, plant, and mineral according to Orłowski et al. (2009).
Additionally, mineral particle size was determined using
the wet sieve method (Gee and Bauder 1982). The
extracted fraction was dried to weigh for contents of 0.2-2,
2-5, and 5-10 mm in size, and classified as coarse sand,
fine gravel, and media gravel, respectively, based on the

as dried to weigh for contents of 0.2-2, extracted f 2-5, and 5 in size, and classified as coarse sand, fine grave dia gravel, respectively, based on the U.S. Department of Agricultural soil texture criteria (Gent 1983). Eight red-crowned crane samples were sampled during 2010–2014 with the aid of fire inspectors at the Zhalong Wetland (Table 1; Fig. 1). The direct causes of death of these cranes were identified as starvation resulting from food shortage under freezing conditions and collision with power lines, as determined by pathological inspection (Luo et al. 2015a). For the purposes of this report, the eight cranes served as negative comparisons to the 2 cases of

suspected poisoning. Approximately 1–2 g of liver, kidney, gastric wall, intestine, pectoral skeletal muscle, and stomach content samples (wet weight) were collected for acid digestion. Several flight feathers were also collected. Each sample was acid-digested in a microwave in accordance with USEPA (1996) methods. The techniques used in our study were described by Luo et al. (2015a). Ca, Mg, Cu, Zn, Pb, Cd, and As concentrations in the six sample types were determined through inductively coupled plasma-mass spectrometry (Agilent 7500ce, Agilent Technologies, Inc., Santa Clara, CA, USA). We estimated the precision and accuracy of the analyses on the basis of two certified reference materials (Beijing Shiji Ouke Bio-tech Co., Ltd): Pseudoscianea (GBW08573) crocea for Cu $(1.36 \pm 0.13 \text{ mg kg}^{-1})$, Zn $(28.8 \pm 1.4 \text{ mg kg}^{-1})$, Pb $(8.8 \pm 1.10 \text{ mg kg}^{-1})$, Cd $(0.014 \pm 0.001 \text{ mg kg}^{-1})$, and As $(5.08 \pm 0.39 \text{ mg kg}^{-1})$. The results agreed with the certified values for all metals, with average recovery rates of 102 %, 94 %, 103 %, 105 %, and 102 % for Cu, Zn, Pb,

Deringer

g) of the red-crowned crane (dw)	Mineral parts
${\bf 2}$ 2 Comparison of the stomach contents (g	Mass of stomach
Fable	

Food parts

	contents (g)	0.2–2 mm	2–5 mm	5-10 mm	Sum	pH _{1:5}		PI	ınt			Animal	
							Seed	Maize	Stem and root	Sum	Fish	Shell	Sum
Control-cranes $(n = 8)$	16.1 ± 7.0	2.7 ± 1.3 (42 ± 14)	4.0 ± 1.6 (69 ± 20)	8.6 ± 6.3 (29 ± 18)	14.0 ± 6.8 (135 ± 27)	5.5 ± 0.1	0.3 ± 0.1	0.3 ± 0.4	0.3 ± 0.2	1.1 ± 0.6	0.6 ± 0.7	0.2 ± 0.2	0.8 ± 0.8
Poisoned cranes $(n = 2)$	10.6 ± 7.0	3.3 ± 0.6 (51 ± 22)	4.6 ± 1.0 (92 ± 23)	7.6 ± 2.5 (36 ± 15)	15.6 ± 2.2 (179 ± 29)	5.2 ± 0.2	I	$4.5 \pm 1.1^{*}$	I	$4.5 \pm 1.1^{*}$	I	I	I
The stomach con	itents of the control-c	stanes were cit	ted from our	early paper	(Luo et al. 20	15b)							

* Indicated significant difference with control (p < 0.05), data in the parentheses are the numbers of the mineral detected in the stomach

represents the item was not detected in the stomach of these avian

; i

Soft tissue	Cu	Zn	Pb	Cd	Hg	As	Ca	Mg
Control of cran	es $(n = 8)$ (Luc	o et al. 2015a, 2	015b)					
Liver	47.3 ± 20.6	469 ± 198	$1.83\pm0.62^{\rm b}$	2.21 ± 1.64	1.64 ± 1.02	0.23 ± 0.03	1.38 ± 0.42	1.32 ± 0.41
	(28.8–72.6)	(209–650)	(0.38–3.21 ^b)	$(0.37 - 4.42^{b})$	$(0.43 - 2.52^{b})$	(0.12-0.52)	(1.07-2.00)	(0.84–1.64)
Kidney	42.5 ± 22.3	370 ± 143	$1.85\pm1.35^{\mathrm{b}}$	1.23 ± 0.80	2.04 ± 1.27	0.18 ± 0.02	1.40 ± 0.40	1.47 ± 0.65
	(29.5–77.5)	(264–577)	(0.50-2.98)	(0.74–2.20)	$(0.52 - 3.41^{b})$	(0.09-0.25)	(0.81-1.70)	(0.79–2.31)
Gut wall	30.1 ± 10.8	321 ± 201	0.81 ± 0.38	0.53 ± 0.34	0.37 ± 0.11	0.07 ± 0.01	2.31 ± 1.51	1.54 ± 1.01
	(15.7–39.9)	(129–598)	(0.09–1.39)	(0.12-0.96)	(0.15-0.50)	(<dl-0.09)< td=""><td>(0.79-4.98)</td><td>(0.49–2.65)</td></dl-0.09)<>	(0.79-4.98)	(0.49–2.65)
Muscle	41.5 ± 20.7	269 ± 128	0.96 ± 0.53	0.53 ± 0.25	0.34 ± 0.28	0.08 ± 0.04	0.95 ± 0.23	0.62 ± 0.47
	(21.3–70.1)	(156–693)	(<dl-1.36)< td=""><td>(0.12-0.90)</td><td>(0.11-0.52)</td><td>(<dl-016)< td=""><td>(0.83-1.15)</td><td>(0.77–1.51)</td></dl-016)<></td></dl-1.36)<>	(0.12-0.90)	(0.11-0.52)	(<dl-016)< td=""><td>(0.83-1.15)</td><td>(0.77–1.51)</td></dl-016)<>	(0.83-1.15)	(0.77–1.51)
Feather	5.98 ± 1.12	174 ± 131	$6.12\pm4.37^{\rm b}$	$0.65\pm0.41^{\rm b}$	$2.51 \pm 1.94^{\rm b}$	<dl< td=""><td>1.19 ± 0.29</td><td>0.75 ± 0.27</td></dl<>	1.19 ± 0.29	0.75 ± 0.27
	(18.2–59.1)	(139–285)	(3.19–14.7 ^b)	(0.41-3.06 ^b)	(1.17–4.23 ^b)		(0.85–1.72)	(0.41–1.18)
Poisoned crane	s (n = 2)							
Resid ^a	9.23 ± 1.28	135 ± 23	$68.5 \pm 10.8^{\rm b}$	0.07 ± 0.03	0.39 ± 0.11	0.32 ± 0.09	0.14 ± 0.03	0.47 ± 0.06
Gastric wall	19.4 ± 3.4	105 ± 34	48.8 ± 7.2^{b}	0.49 ± 0.08	0.26 ± 0.07	0.51 ± 0.11	0.47 ± 0.16	0.83 ± 0.20
Liver	24.2 ± 2.7	263 ± 42	31.4 ± 3.8^{b}	1.14 ± 0.24	$5.69 \pm 1.21^{\rm b}$	0.19 ± 0.04	1.29 ± 0.34	1.35 ± 0.53
Kidney	38.8 ± 5.9	210 ± 37	60.3 ± 5.7^{b}	1.33 ± 0.18	1.57 ± 0.37	0.46 ± 0.11	1.04 ± 0.26	0.99 ± 0.37
Gut wall	12.6 ± 2.3	229 ± 56	25.3 ± 6.2^{b}	1.05 ± 0.26	<dl< td=""><td>0.97 ± 0.20</td><td>0.81 ± 0.10</td><td>0.85 ± 0.41</td></dl<>	0.97 ± 0.20	0.81 ± 0.10	0.85 ± 0.41
Muscle	23.9 ± 4.3	190 ± 43	$3.60\pm0.52^{\rm b}$	1.37 ± 0.24	0.71 ± 0.20	0.67 ± 0.14	0.72 ± 0.29	0.54 ± 0.06
Feather	21.7 ± 3.5	88.5 ± 12.7	$5.41 \pm 0.77^{\mathrm{b}}$	0.12 ± 0.05	$0.53\pm0.09^{\rm b}$	0.11 ± 0.06	0.70 ± 0.15	0.28 ± 0.05

Table 3 Body burden (Mean \pm SD/(min–max)) of six elements in different tissue of the two poisoned red-crowned crane and comparison with control cranes (dw, g kg⁻¹ for Ca and Mg, and mg kg⁻¹ for other elements)

^a Resid was the abbreviation of residues in stomach, ^b means the value to reach the environmental exposure level (0.2 mg kg⁻¹ dw for feather Cd by Pain et al. 2005; 1.7 mg kg⁻¹ dw for Pb by Degernes 2008; and 0.2 mg kg⁻¹ dw for feather Hg by Scheuhammer 1987); <DL means below detection level

Cd, and As, respectively. We determined the T-Hg concentration in internal tissues and feathers by using a mercury analyzer (Tekran 2600 CVAFS, Tekran Instrument Corporation, Knoxville, USA) in accordance with the methods described in our previous study (Luo et al. 2014b). The detection limit for all elements was 5 ng kg⁻¹. Reusable materials, such as Petri dishes for examining stomach contents, were acid-washed. The samples were analyzed in triplicate at a relative standard deviation lower below 1.5 %. The concentrations were expressed as mg kg⁻¹ dw.

Results and Discussion

The stomach contents of the two cranes were dominated by maize compared to available controls, and lacked animal residues (Table 2). The mean tissue element concentrations determined from the two poisoned cranes exhibited the following order: Ca > Mg > Zn > Pb > Cu > Cd > H g > As (Table 3). The concentrations of macro elements Ca and Mg were predominant in the cranes, with concentrations ranging from 0.14 to 1.38 g kg⁻¹ and from 0.28 to 1.54 g kg⁻¹, respectively; these concentrations were similar to control cranes (Luo et al. 2015a). The mean tissue concentrations of the trace elements Cu and Zn were high in the liver (24.2–47.3 mg kg⁻¹ dw for Cu, and

263–469 mg kg⁻¹ dw for Zn), kidney (38.8–42.5 mg kg⁻¹ dw for Cu, and 210–370 mg kg⁻¹ dw for Zn), and muscle $(23.9-41.5 \text{ mg kg}^{-1} \text{ for Cu, and } 190-269 \text{ mg kg}^{-1} \text{ dw for}$ Zn). By contrast, the concentrations were relatively low in feathers $(5.98-21.7 \text{ mg kg}^{-1})$ for the Cu, and 88.5–174 mg kg^{-1} dw for Zn). The concentrations of Cu, Zn, Cd, Hg, and As in the two poisoned cranes were generally similar to those in the eight control birds. The stomach contents and the tissues from the two poisoned cranes contained elevated Pb levels compared with the control cranes. The average Pb concentrations in the kidney and liver tissues of the two examined cranes reached 31.4 and 60.3 mg kg^{-1} dw, respectively. By comparison, the Pb concentrations were 1.83 and 1.85 mg kg⁻¹ dw in the same tissues of the controls. The average Pb concentrations in the tissues of the two poisoned cranes displayed a decreasing pattern: kidney > gastric wall > liver > gut wall > feather > muscle.

In general, the clinical signs of the poisoned cranes before their death and the results of postmortem tissue analyses indicated that excessive intake of Pb might have been associated with the death of the red-crowned cranes. In support of this assumption, a summary of toxic effect Pb level in kidney and liver of birds is provided in Table 4.

In the past 30 years, habitat loss has forced the cranes to rely on crops and farmlands in Zhalong Wetland (Su and

Organ	Concentration	Species	Toxic effect	Ref. source
Kidney	1–10	For common bird	Background level	Scheuhammer (1987)
	Approxi. 25-30	Whooping crane	Death	Snyder et al. (1992)
	15.5-17.7	Rooks (dead nestlings)	Elevated level (mortality effect: unknown)	Orłowski et al. (2012)
	>6	For lead shot poisoned bird	Cause morality	Eisler (1988)
	31.0-34.0	Island red-crowned crane	Death	Teraoka et al. (2007)
	< 6	Raptor	Background level	Clark and Scheuhammer (2003)
	6–20	Raptor	Sub-lethal effect	
	>20		Cause mortality	
	35.4-68.3	Red-crowned crane	Suspect Pb poisoning	In present research
Liver	0.5-5	For common bird	Background level	Scheuhammer (1987)
	5.5-44.3	Whooper swans (Cygnus cygnus)	Death	Ochiai et al. (1992)
	31.8-62.5	Island red-crowned crane	Cause mortality	Teraoka et al. (2007)
	Approxi. 50-70	Whooping crane	Cause mortality	Snyder et al. (1992)
	16.3–17.8	Rooks (dead nestlings)	Elevated level (mortality effect: unknown)	Orłowski et al.(2012)
	>1.5	For common bird	Toxicosis	Degernes (2008)
	>10	Lead shot poisoned bird	Cause mortality	Eisler (1988)
	9.7	White fronted geese	Abnormal exposure	Kim and Oh (2014)
	5.5-6.1	Mallard		
	6.4–7.7	Spot-billed duck		
	<6	Raptor	Background level	Clark and Scheuhammer (2003)
	6–30	Raptor	Sub-lethal effect	
	>30	Raptor	Cause mortality	
	29.4–35.5	Red-crowned crane	Suspect Pb poisoning	In present research

Table 4 Summary of Pb toxic effect level in kidney and liver of avian (dw, mg kg $^{-1}$)

Zhou 2012; Luo et al. 2014a). High corn contents left in the stomach of the red-crowned cranes in this case study supported the point. High anthropogenic inputs from the residential, agricultural, and industrial zones surrounding Zhalong Wetland in the past decades also increased the concentrations of trace elements in the sediments and prey of cranes (Luo et al. 2013, 2014b). These changes may ultimately pose a risk to cranes and other birds that rely on food and water from this habitat (Luo et al. 2013, 2015a). However, the two dead cranes only exhibited high concentrations of Pb among the several trace elements analyzed. The Zn concentration of $<210 \text{ mg kg}^{-1}$ in the liver was significantly below the toxic level of 2100 mg kg⁻¹ (United States Department of the Interior 1998). By contrast, whooping cranes with a Zn concentration of 420 mg kg^{-1} dw in the liver are diagnosed with Zn toxicosis (Spalding et al. 1997). The Zn concentration of 150 mg kg⁻¹ wet weight (approximately 430 mg kg⁻¹ dw) in the kidney and the Zn concentration of 150 mg kg^{-1} wet weight (approximately 450 mg kg^{-1} dw) in the liver of free-flying trumpeter swans (Cygnus buccinator) are considered elevated (Carpenter et al. 2004) compared with the normal range of 30 mg kg⁻¹ wet weight to 100 mg kg⁻¹

wet weight (approximately 100 to 300 mg kg⁻¹ dw) (Puls 1994). In the present study, the detected Zn concentrations in the liver and kidney of crane specimens were within the normal range set by Puls (1994) and below the toxic level previously reported. Similar to Zn, the detected Cu, Hg, and Cd concentrations in the liver did not exceed the toxicity thresholds: 540 mg kg⁻¹ for Cu and 20 to 24 mg kg⁻¹ dw for Hg (United States Department of the Interior 1998), and 40 mg kg⁻¹ dw for Cd (Eisler 2000).

The degree of Pb toxicity varies according to age, sex, and species (Eisler 1988; Burger and Gochfeld 1993; Orłowski et al. 2012). The Pb concentrations in the liver and kidney of the two dead cranes were similar to the lethal threshold of >24 and 20–30 mg kg⁻¹ dw suggested by Clark and Scheuhammer (2003) and Friend (1985). Teraoka et al. (2007) reported similar liver Pb level, ranging from 30 to 60 mg kg⁻¹, in four dead red-crowned cranes in Japan; however, these concentrations were significantly higher than those reported by Takazawa et al. (2004), Friend (1985), and Clark and Scheuhammer (2003). In addition, Snyder et al. (1992) reported a case of lead toxicosis in a whooping crane from Colorado in the United States. Pb was detected in the liver at 20 mg kg⁻¹ wet weight (approximately 50–70 mg kg⁻¹ dw) and in the kidney at 10 mg kg⁻¹ wet weight (approximately 25–30 mg kg⁻¹ dw), similar to those detected in the red-crowned cranes in the present study. The observed symptoms, such as loss of appetite, lethargy, weakness, emaciation, tremors, and drooped wings, of the moribund red-crowned cranes are typical signs of Pb poisoning (Eisler 1988). These symptoms supported our initial assumption of Pb poisoning in this case. Furthermore, Pb concentration of 30 mg kg⁻¹ dw in the liver or kidney appears to be associated with lethal toxicosis in the red-crowned crane species.

In birds, the background concentrations of Ca and Mg in the liver range from 0.2 to 0.4 g kg⁻¹ dw and from to 0.8 g kg^{-1} dw, respectively (Wiemeyer and Withers 2004; Custer et al. 2003). According to Gonzalez (1988) and Scheuhammer (1996), high loads of toxic trace elements, such as Pb and Cd, in the avian body likely reduces Ca and Mg contents; as a consequence, Ca and Mg deficiencies occur. We found that Ca and Mg levels in the liver of the dead samples were approximately two times higher than the background concentrations reported by Wiemeyer and Withers (2004) and Custer et al. (2003) in the present study. Therefore, the increased concentrations of Pb did not induce marked Ca or Mg deficiencies. This conflicting result may be obtained because the high content of minerals in the stomach (Table 2) acted as the source of Ca and Mg for the physiological need of cranes and compensated the possible loss induced by the increased Pb concentration in this case; this observation was supported by the finding of Orłowski et al. (2009).

Potential biases should be considered when interpreting our results because other toxic substances from pesticides were not examined in this study. However, the increased Pb concentrations detected in the tissues of the two redcrowned cranes suggested that the inputs of Pb into Zhalong Wetland should be reduced. Thus, the healthy state of this critical crane habitat can be maintained for the longterm sustainability of the crane population.

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