

Sensitivity of Four Cyanobacterial Isolates from Tropical Freshwaters to Environmentally Realistic Concentrations of Cr^{6+} , Cd^{2+} and Zn^{2+}

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Abstract Sensitivity of four tropical cyanobacteria viz. *Coelosphaerium* sp., *Synechococcus* sp., *Oscillatoria* sp. and *Chroococcus* sp. to environmentally relevant concentrations of Cr^{6+} , Cd^{2+} and Zn^{2+} was assessed based on fluorescence change as a proxy for growth reduction. At 24 h exposure, the growth reduction in the cyanobacteria followed the order: $\text{Zn}^{2+} < \text{Cr}^{6+} \leq \text{Cd}^{2+}$. Of the four cyanobacteria, *Synechococcus* was the most sensitive for Cr^{6+} , whereas *Chroococcus* was the most sensitive for Cd^{2+} and Zn^{2+} . Sensitivity was gradually decreased by 96 h implying the acquisition of tolerance by cyanobacteria to heavy metal ions with prolonged exposure.

Keywords Cyanobacteria · Chromium · Cadmium · Zinc · Fluorescence · Concentration response

Heavy metals are continuously released into the biosphere by natural and anthropogenic sources. Chromium, cadmium and zinc are such heavy metals causing pollution of aquatic ecosystems. Chromium is a transition metal

existing in oxidation states varying from -2 to $+6$, and 0 , $+3$ and $+6$ being the most common oxidation states (Wong and Trevors 1988). Chromium is used in alloys including stainless steel and in chemical industrial processes like leather tanning, pigments and dye, pulp and paper, automobile and electroplating (McGrath and Smith 1990). Being the most abundant, Cr (III) and Cr (VI) attract more attention in terms of their toxicity to aquatic biota. Cadmium usually has an oxidation state of $+2$. Cadmium occurs as a minor component in most zinc ores and therefore is a byproduct of zinc production. It was used for a long time as a pigment and for corrosion resistant plating on steel. Zinc acts as an essential trace element in development, growth and differentiation of all living systems. Zinc occurs exclusively as the Zn^{2+} divalent cation and due to its completely filled *d* orbitals; Zn^{2+} cannot undergo redox changes under biological conditions (Nies 1999). However, Zn^{2+} can be toxic to biological systems at high concentrations. The release of these heavy metals into aquatic ecosystems from industrial waste and other sources has been a serious environmental issue because heavy metals are non-degradable tending to accumulate in the water bodies and cause toxicity to aquatic organisms as well as terrestrial organisms via bioaccumulation through food chains.

Cyanobacteria which are prokaryotic photosynthetic microorganisms, found in almost every terrestrial and aquatic habitats (Waterbury 2006). Cyanobacteria are regarded as bioindicators for environmental monitoring and assessment as they are sensitive to a number of pollutants (Campanella et al. 2000). Recently, use of cyanobacteria as biomonitoring agents for the detection of toxicity of Cr^{6+} , Cd^{2+} , and Zn^{2+} in aquatic environments has been exploited due to their high sensitivity and reproducibility. The most preferred biological process to assess

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the growth reducing effect of Cr^{6+} , Cd^{2+} , Zn^{2+} on microalgae is the photosynthetic activity, estimated by chlorophyll *a* fluorescence of photosystem II (PSII) (Altamirano et al. 2004). During the photosynthesis process a small portion of energy absorbed as sunlight is emitted as fluorescence (Wong et al. 2013). The amount of this fluorescence emission can be changed due to the inhibition of growth in the presence of contaminants like Cr^{6+} , Cd^{2+} , and Zn^{2+} in the environment or their direct effect on PSII. Therefore fluorescence changes in these organisms can be used as an indicator for the monitoring heavy metals in the aquatic systems (Buonasera et al. 2011). Information on sensitivity of tropical cyanobacteria to environmentally realistic levels of heavy metal ions is meager in the scientific literature. The objective of the present study was to assess the effect of environmentally realistic concentrations of Cr^{6+} , Cd^{2+} , and Zn^{2+} on the growth of four cyanobacteria isolated from different tropical freshwater environments based on fluorescence change patterns.

Materials and Methods

Four cyanobacteria viz. *Coelosphaerium* sp., *Synechococcus* sp., *Oscillatoria* sp. and *Chroococcus* sp. used in this study have been isolated from the water samples collected from different freshwater sites in the Gampaha district, Sri Lanka. *Coelosphaerium* sp. and *Synechococcus* sp. were isolated from two domestic freshwater ponds whereas sources of *Oscillatoria* sp. and *Chroococcus* sp. were from Kelani River and a shallow well respectively. For cyanobacteria isolation, flasks containing BG11 medium were inoculated with respective water samples and incubated at $25 \pm 2^\circ\text{C}$ on a bench top orbital shaker (GFL[®] 3005) at 100 rpm, under continuous illumination ($200 \mu\text{E m}^{-2} \text{s}^{-1}$ PPFD). After green color cyanobacterial growth was observed, the organisms were isolated and purified into single axenic cultures using the isolation streak method on the BG11 agar medium. Cyanobacteria were identified to generic level according to Bellinger (1992) based on observable morphological characters under the light microscope (Olympus CX21FS1). Axenic cultures of cyanobacterial isolates were maintained in BG11 medium and incubated at $25 \pm 2^\circ\text{C}$ in flasks on the bench top orbital shaker (GFL[®] 3005), under continuous illumination. Cultures were periodically transferred to fresh media. Chromium, cadmium, and zinc content of water samples where the cyanobacteria were isolated, were determined using graphite furnace Atomic Absorption Spectrometer (Analytik Jena novAA[®] 400P) following the procedure described by APHA (1999). For toxicity assessments, stock solutions of Cr^{6+} , Cd^{2+} and Zn^{2+} were prepared in de-ionized water using $\text{K}_2\text{Cr}_2\text{O}_7$ ($\geq 99\%$

purity, NORMAPUR, Belgium), $\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ ($\geq 99\%$ purity, Sigma-Aldrich, USA), and $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ($\geq 99\%$ purity, Sigma-Aldrich, USA) respectively and working solutions for Cr^{6+} , Cd^{2+} (0.8, 0.4, 0.2, 0.1, 0.05, 0.025, and 0.0125 mg/L) and for Zn^{2+} (16.0, 8.0, 4.0, 2.0, 1.0, 0.5, and 0.25 mg/L) were prepared by appropriate dilutions. The metal levels in the working solutions were analytically verified by atomic absorption spectrometry [Analytik Jena model: novAA 400P atomic absorption spectrometer with a graphite furnace and auto sampler or flame mode (Acetylene/air) where appropriate] following the standard analytical procedures (APHA 1999). Limit of quantification (LOQ) for each analyte was calculated as the metal concentration that correspond to the sum of the mean and ten times the standard deviation of 10 independent measurement of the blank medium (nitric acid). The LOQ for Cr, Cd and Zn were 0.014, 0.001 and 0.1 mg/L respectively. Each sample was analyzed in duplicates. The measured concentrations of metals in the working solutions were for Cr 0.89, 0.42, 0.21, 0.08, 0.044, 0.023 and <0.014 (LOQ) mg/L; for Cd 0.76, 0.41, 0.21, 0.11, 0.045, 0.022 and 0.013 mg/L; for Zn 14, 7, 3.5, 2, 0.75, 0.45, and 0.19 mg/L. In general measured concentrations of the metal did not show much deviations from the nominal concentrations. Metal solutions were sterilized by autoclaving. All glassware was acid washed before use to avoid binding of metal to the glass surface. Sensitivity of the cyanobacteria to Cr^{6+} , Cd^{2+} , and Zn^{2+} wastested individually under BG11 growth medium using seven nominal concentrations of each metal ion (final concentration for Cr^{6+} , Cd^{2+} : 1–66 $\mu\text{g/L}$ and for Zn^{2+} 20–1330 $\mu\text{g/L}$). The control media contained only the growth medium (240 μL). The protocol of ‘Algal microplate toxicity test suitable for heavy metals’ (Peterson et al. 2005) was followed in the toxicity assessments. Bioassays were conducted in quadruplicates in 96-well microplates (Sterilin[®], flat bottom, sterile, with lid). The cyanobacteria were incubated on an orbital shaker (GFL[®] 3005) at 100 rpm under continuous illumination using cool white fluorescent lamps ($200 \mu\text{E m}^{-2} \text{s}^{-1}$ PPFD). Growth patterns of the cyanobacteria were measured as chlorophyll *a* fluorescence (using 440/40 nm excitation filter and 680/30 nm emission filter) at 24 h intervals from the time of initial inoculation up to 96 h using the BioTek SynergyTM HT Microplate Reader using Gen5 software (OECD 2011). Relative growth of cyanobacteria based on fluorescence was calculated as a percentage in relation to the untreated control for each tested concentration of each metal ion. Effective Concentrations for growth reduction (EC_x) $x = 50, 20$ and 10 (estimated metal ion concentration where the organisms show the relevant % reduction in fluorescence compared to the control) were estimated by Probit analysis (Finney 1971), using MINITAB 15 Statistical SoftwareTM. In

addition, no observed effect concentration (NOEC) and lowest observed effect concentration (LOEC) for each metal ion were estimated based on Analysis of Variance test followed by Dunnett's post-hoc test. $p < 0.05$ was considered as statistically significant.

Results and Discussion

Four cyanobacteria (*Coelosphaerium* sp., *Synechococcus* sp., *Oscillatoria* sp. and *Chroococcus* sp.) isolated from tropical freshwaters were tested for their sensitivity to environmentally relevant concentrations of three heavy metal ions, Cr^{6+} , Cd^{2+} and Zn^{2+} was assessed based on fluorescence change as a proxy for growth reduction. The concentration of Cr was 27 $\mu\text{g/L}$ in the water from which *Coelosphaerium* sp. was isolated whereas the waters from which the other three cyanobacteria were isolated contained only 2–3 $\mu\text{g/L}$ of chromium. The water from which *Oscillatoria* sp. was isolated had 20 $\mu\text{g/L}$ cadmium in comparison to the other water samples (below 3 $\mu\text{g/L}$). Zinc levels were relatively low (52 $\mu\text{g/L}$) in the water from which *Coelosphaerium* sp. was isolated compared to the waters from which the other cyanobacteria were collected (110–138 $\mu\text{g/L}$). The range of metal ion concentrations used in this study also covers the metal levels in the natural habitat of the four cyanobacteria. The metal ion concentration–growth response relationships (as % relative fluorescence) for the four cyanobacteria at 24 h exposure are presented in Fig. 1. In all cyanobacteria, relative fluorescence (%) decreased with increasing metal ion concentration in the growth medium at 24 h of exposure. Estimated NOEC and LOEC of the three metal ions for growth reduction of the cyanobacteria at 24 h are presented in Table 1. Of the four cyanobacteria, NOEC and LOEC of Cr^{6+} are 16 and 33 $\mu\text{g/L}$ for *Synechococcus* sp., whereas for the other three cyanobacteria corresponding concentrations are 33 and 66 $\mu\text{g/L}$ respectively. The toxicity thresholds estimated by hypothesis testing indicate that *Synechococcus* sp. is the most sensitive cyanobacteria for Cr^{6+} . Estimated NOEC (16 $\mu\text{g/L}$) and LOEC (33 $\mu\text{g/L}$) for Cd^{2+} are two-fold greater in *Coelosphaerium* sp. and *Synechococcus* sp. in comparison to those (NOEC: 8 $\mu\text{g/L}$ and LOEC: 16 $\mu\text{g/L}$) in *Oscillatoria* sp. and *Chroococcus* sp. For Zn^{2+} , estimated NOEC and LOEC for growth reduction of *Chroococcus* sp. are 166 and 330 $\mu\text{g/L}$ respectively whereas corresponding values for the other three cyanobacteria are two-fold greater (NOEC: 330 and LOEC: 660 $\mu\text{g/L}$) indicating greater sensitivity of *Chroococcus* sp. to Zn^{2+} .

Table 2 shows the estimated 24 h effective concentrations (EC_{50} , EC_{20} and EC_{10}) of metal ions Cr^{6+} , Cd^{2+} and Zn^{2+} , for the growth reduction of cyanobacteria based on

Probit analysis. Of the four cyanobacteria, *Synechococcus* sp. showed the lowest EC_{50} for all three metals (Cr^{6+} , Cd^{2+} and Zn^{2+}). However the differences in EC_{50} s among the four cyanobacteria are not statistically significant as the respective 95 % confidence limits overlap. Moreover except for effects of Cr^{6+} on *Synechococcus* sp., the concentration–response curves did not exceed 50 % decrease in relative fluorescence levels (Fig. 1) and the EC_{50} estimates are based on extrapolations of the measured effects at the tested concentrations. Hence, estimated EC_{50} s have to be interpreted with caution.

When considering the 24 h EC_{20} s for Cr^{6+} (Table 2), *Synechococcus* sp. exhibited the lowest EC_{20} value ($p < 0.05$) indicating the highest sensitivity. *Chroococcus* sp. showed the lowest EC_{20} for Cd^{2+} and Zn^{2+} but the differences among four cyanobacteria are not statistically significant. Toxicity threshold estimates for the three metal ions for the cyanobacteria tested in this study based on two methods viz. LOEC based on hypothesis testing and EC_{10} based on probit analysis correspond well for Zn^{2+} except for *Chroococcus* sp. For Cr^{6+} , EC_{10} estimates are much lower than the LOEC estimates but correspond well with the NOEC estimates for cyanobacteria except in *Synechococcus* sp. where the LOEC agrees with the EC_{10} estimate. For Cd^{2+} , EC_{10} estimates for *Coelosphaerium* sp. and *Synechococcus* sp. correspond well with the LOEC values but LOEC estimates for *Oscillatoria* sp. and *Chroococcus* sp. are much lower than the respective EC_{10} estimates.

The growth response of the four cyanobacterial isolates (as relative fluorescence %) decreased with increasing metal ion concentration for 24 h and the growth reduction followed the increasing order, $\text{Zn}^{2+} < \text{Cr}^{6+} \leq \text{Cd}^{2+}$. Cadmium shows the highest toxicity to the tested cyanobacteria at elevated concentrations. It has been reported earlier that in cyanobacteria, Cd causes severe inhibition of growth, photosynthesis (Zhou et al. 2008) and nitrogen fixation (Singh et al. 2014). Reduction of growth at elevated chromium concentrations may be due to the toxicity of hexavalent chromium to cyanobacteria. Chromium can interfere with the uptake of some essential elements such as Fe and S due to its structural similarity. Once enters the cell, chromium stress can also result in alterations of photosynthetic pigments such as chlorophyll (Pereira et al. 2013). Chromium can also produce reactive oxygen species that cause oxidative damage to cells and cellular mechanisms (Hameed and Hasnein 2014). Hence, changes of chlorophyll fluorescence as observed in this study with Cr^{6+} and Cd^{2+} exposure may be due to inhibition of physiological processes in the cells. Although being an essential metal, zinc is not as toxic as chromium or cadmium, at high concentrations zinc also shows toxic effects to the tested cyanobacteria. In a study by Loez et al.

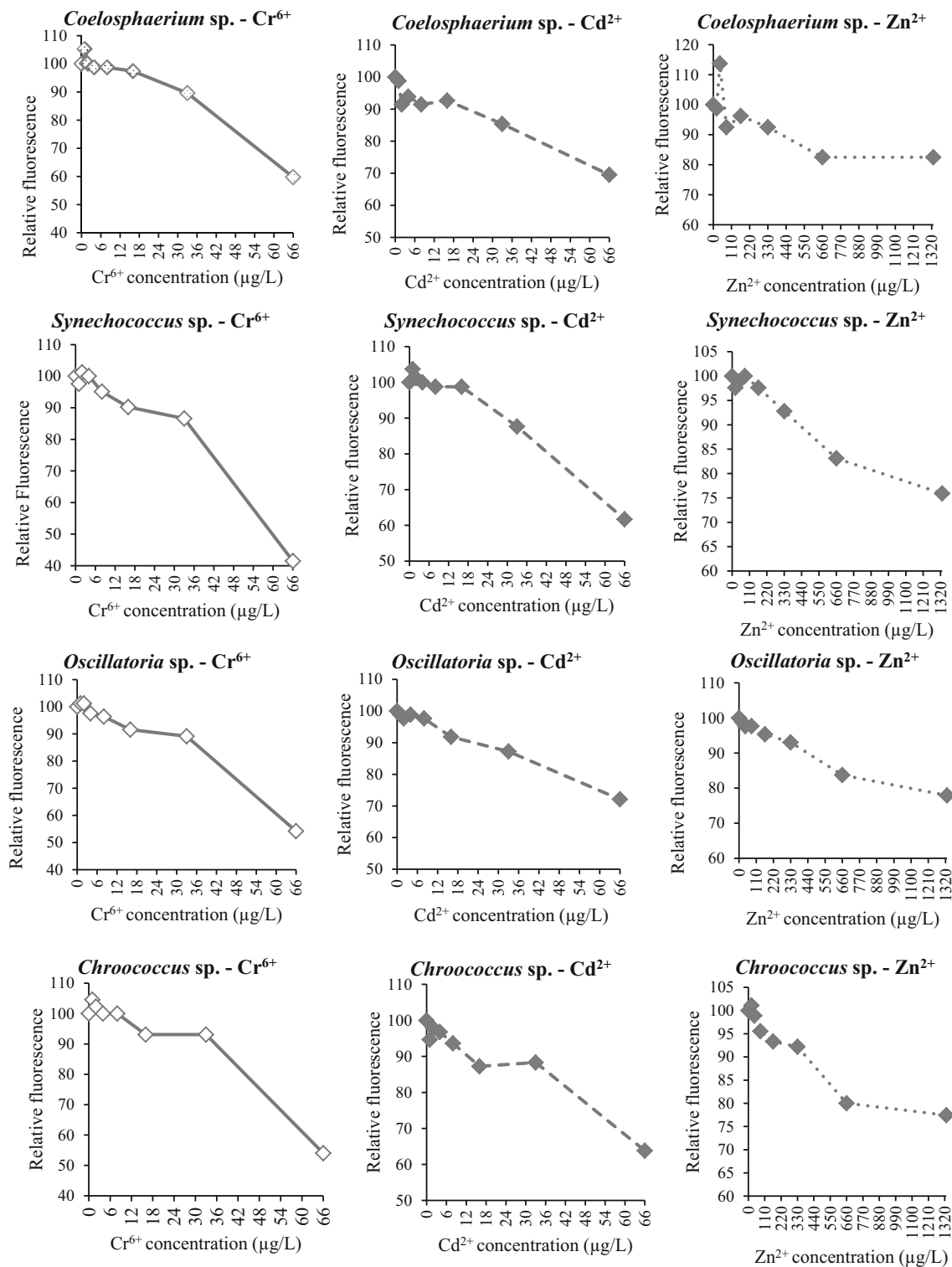


Fig. 1 Metal ion concentration and relative fluorescence patterns of four tropical cyanobacteria following 24 h exposure to a range of Cr⁶⁺, Cd²⁺ and Zn²⁺ concentrations (mean of quadruplicate measurements are plotted for each concentration)

(1995), it was shown that even at low concentrations, Zn was deleterious to the algae *Euglenophyceae*, *Cyanophyceae* and *Xanthophyceae*.

After 24 h, it was observed that sensitivity of the cyanobacteria to the Cr⁶⁺, Cd²⁺ and Zn²⁺ was decreased and growth response of cyanobacterial cultures gradually

Table 1 Estimated 24 h NOEC and LOEC of Cr⁶⁺, Cd²⁺ & Zn²⁺ for growth reduction of four tropical cyanobacteria

Heavy metal ion		Toxicity threshold in µg/L			
		<i>Coelosphaerium</i>	<i>Synechococcus</i>	<i>Oscillatoria</i>	<i>Chroococcus</i>
Cr ⁶⁺	NOEC	33	16	33	33
	LOEC	66	33	66	66
Cd ²⁺	NOEC	16	16	8	8
	LOEC	33	33	16	16
Zn ²⁺	NOEC	330	330	330	166
	LOEC	660	660	660	330

Table 2 Estimated 24 h toxicity thresholds (EC₅₀, EC₂₀ and EC₁₀ in µg/L) of Cr⁶⁺, Cd²⁺ & Zn²⁺ for growth reduction of four cyanobacteria

Heavy metal ion		Toxicity threshold (95 % confidence limits) in µg/L			
		<i>Coelosphaerium</i>	<i>Synechococcus</i>	<i>Oscillatoria</i>	<i>Chroococcus</i>
Cr ⁶⁺	EC ₅₀	73 (66–81)	61 (56–67)	69 (62–78)	69 (63–76)
	EC ₂₀	49 (44–54)	38* (34–42)	43 (39–48)	47 (43–52)
	EC ₁₀	37 (32–42)	26** (22–30)	30 (25–35)	36 (31–40)
Cd ²⁺	EC ₅₀	93 (78–119)	73 (66–81)	87 (75–104)	81 (70–98)
	EC ₂₀	47 (40–59)	50 (46–56)	52 (46–61)	44 (38–52)
	EC ₁₀	24 (16–31)	39 (33–43)	35 (28–41)	25 (19–31)
Zn ²⁺	EC ₅₀	2182 (1762–2969)	1795 (1544–2188)	1950 (1634–2480)	1873 (1576–2361)
	EC ₂₀	1194 (979–1554)	1082 (937–1281)	1122 (951–1378)	1043 (884–1271)
	EC ₁₀	677 (514–871)	710 (582–846)	690 (548–847)	609 (472–753)

95 % confidence limits are given within parentheses

* Significantly different from other three cyanobacteria isolates

** Significantly different from *Coelosphaerium* and *Chroococcus*

reached closer to the control levels by the end of 96 h exposure (Fig. 1S: Supplementary figure). This may be due to the development of metal tolerance mechanisms in the cyanobacterial cells with the increase in exposure time. Prokaryotic cells like cyanobacteria employ ATP-consuming efflux of heavy metals or enzymatic change of speciation to achieve detoxification (Nies 1999). Most cyanobacteria have one or more endogenous plasmids responsible for many cellular functions including heavy metal resistance (Lee et al. 2013). Metallothionein (MT) and efflux ATPases also assist in zinc homeostasis and cadmium tolerance (Chauvat and Chauvat 2015). For example *Synechococcus* PCC7942 have Zn- and Cd-binding MT encoded by *smtA* gene and various *Oscillatoria* strains also have genes for both MT and efflux pump (Blindauer 2011).

In conclusion, sensitivity of four cyanobacteria viz. *Coelosphaerium* sp., *Synechococcus* sp., *Oscillatoria* sp. and *Chroococcus* sp. isolated from tropical freshwaters to environmentally relevant concentrations of Cr⁶⁺, Cd²⁺ & Zn²⁺ based on fluorescence changing patterns followed the increasing order of toxicity, Zn²⁺ < Cr⁶⁺ ≤ Cd²⁺. *Synechococcus* sp. was the most sensitive isolate for Cr⁶⁺,

exhibiting the lowest NOEC/LOEC as well as the lowest 24 h-EC₂₀ of 38 µg/L. With respect to Cd²⁺ and Zn²⁺, statistically significant differences were not found in 24 h-EC₂₀s among the four cyanobacteria but estimated 24 h LOECs/NOECs of Cd²⁺ for growth retardation are lower in *Oscillatoria* sp and *Chroococcus* sp. than those of *Coelosphaerium* sp. and *Synechococcus* sp. Based on NOEC/LOEC estimates *Chroococcus* sp. was the most sensitive cyanobacteria for Zn²⁺. By 96 h exposure, sensitivity to the Cr⁶⁺, Cd²⁺ and Zn²⁺ was decreased implying the acquisition of tolerance by cyanobacteria to the tested metal ions. Of the four tropical cyanobacteria isolates, *Synechococcus* sp. is the most sensitive cyanobacteria for Cr⁶⁺, and *Chroococcus* sp seems to be the most sensitive isolate for Cd²⁺ and Zn²⁺. The results imply that sensitivity of cyanobacteria to specific heavy metal ions can vary depending on the cyanobacterial strain and generalizations cannot be made as to the most sensitive cyanobacteria for all heavy metal ions. Hence the most sensitive consortium of organisms needs to be selected for monitoring and assessments of heavy metal contamination in tropical aquatic environments.

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