

Changes in Microbial Populations and Enzyme Activities During Nitrogen Biodegradation of Domestic Sewage Treatment in the Subsurface Wastewater Infiltration System (SWIS)

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Abstract During the process of domestic sewage treatment in the Subsurface Wastewater Infiltration System (SWIS), changes in the microbial populations (nitrifying and denitrifying bacteria) and enzyme activities (urease, nitrate reductase and nitrite reductase) involved in the nitrogen removal process were evaluated over a 2-year period. The results showed nitrifying bacteria number declined with depths increasing, while denitrifying bacteria increased, both of which increased nearer the inlet. The depth for nitrate reductase activity from high to low in sequence was 0.3, 0.5, 0.7, 0.9 and 1.1 m. For nitrite reductase, the sequence was 0.5, 0.3, 0.7, 0.9 and 1.1 m. Urease and nitrite reductase activities were in positive correlation with the total nitrogen removal efficiency, with correlation coefficients 0.8662 and 0.9140, respectively and could be alternative to monitor the nitrogen biodegradation process in SWIS.

Keywords Soil treatment · Subsurface wastewater infiltration system · Nitrogen removal · Microbial population · Enzyme activity

The idea of using Subsurface Wastewater Infiltration System (SWIS) for the treatment and improving of domestic wastewater emerged in the second half of the last century

(Smiles 2006; Xu and Que-Hee 2007; Arienzo et al. 2009; Tunçsiper et al. 2009). Because of large demand of land area but no requirement of perfect sewage systems, SWIS has recently received considerable attention as low cost and efficient means of cleaning up domestic wastewater at secondary and tertiary levels (Arienzo et al. 2009). Up to now, over ninety million SWISs are in operation and benefit the people in North America, Europe and Asia (Belinda et al. 2007; Kim and Sansalone 2008; Li et al. 2011). In China, hundreds of SWIS projects are constructed in Guizhou, Yunnan, Shenzhen and Shenyang, etc. In the SWIS treatment, wastewater is firstly treated by conventional physico-chemical and/or biological treatment and then allowed to infiltrate through aerated unsaturated zone wherein it gets purified through processes such as filtration, adsorption, chemical reaction and biodegradation.

Despite relatively wide use of this environmentally friendly technology, relatively little is known about the changes of microbial populations and enzyme activities involved in nitrogen biotransformation and removal process in this system (Luanmanee et al. 2002; Parveen et al. 2006). The aim of the current study was to examine the microbial populations and enzyme activities involved in the nitrogen removal process in a SWIS, which was designed to improve the quality of wastewater at tertiary level. Also, the nitrogen removal efficiency of this SWIS was investigated during a 2-year experimental period.

Materials and Methods

The SWIS system in this study is located in Shenyang, northeastern China, which covers 300 m² ($L \times W = 20 \text{ m} \times 15 \text{ m}$) with effective depth (ED) of 1.5 m (Fig. 1). The pre-treated and settled wastewater flows under gravity

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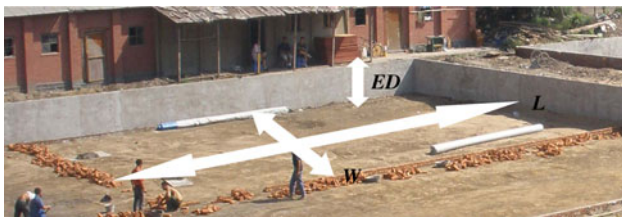


Fig. 1 View of the subsurface wastewater infiltration system

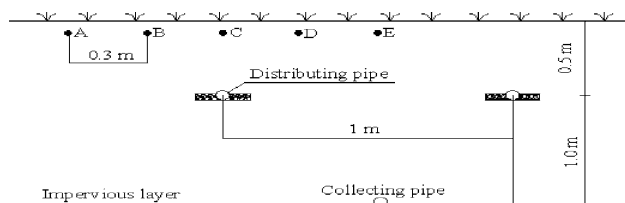


Fig. 2 Schematic diagram of the distributing and collecting pipes and sampling points

action into the distributing pipes of the SWIS, which are 0.08 m in diameter, 0.5 m underneath, then the treated water is collected in the collecting pipe 1.5 m underneath with 0.1 m in diameter. The spacing interval between two distributing pipes is 1 m. In order to monitor the microbial populations and enzyme activities, five sampling points (labeled as A, B, C, D and E) are arranged vertical to the distributing pipes, with intervals of 0.3 m, as shown in Fig. 2.

The influent in this study was a combined wastewater from toilets, bathrooms, etc. The ranges of major water quality indices were pH 7.1–7.4, chemical oxygen demand (COD) 280–353 mg/L, biological oxygen demand (BOD₅) 160–210 mg/L, suspended solid (SS) 150–200 mg/L, total nitrogen (TN) 30–45 mg/L, total phosphorus (TP) 3–4 mg/L, ammonia nitrogen (NH₃-N) 20–30 mg/L, with an average ratio of 0.6 for BOD₅/COD.

The matrix in the SWIS was composed of soil, coal slag and dewatered activated sludge mixed in volume ratio of 13:5:2. The soil used was meadow brown soil, sampled from the top 20 cm from Shenyang Ecological Station, with total organics 22.8 g/kg, TN 1.4 g/kg and TP 0.85 g/kg. The activated sludge was obtained from the aeration tanks in Shenyang Northern Municipal Sewage Treatment Plant, air dried after being centrifuged for 15 min at 1,500 rpm. Other materials (gravel and coal slag) were purchased from a local market (in diameter: gravel 10–25 mm and coal slag 4–8 mm). The infiltration rate, porosity and surface area of the matrix were 0.37 m³/m²·d, 59% and 5.21 m²/g, respectively. In the SWIS, the matrix was filled from the ground surface down to 1.4 m, followed by gravel.

The influent and effluent samples were collected once a week, stored at 4°C and analyzed within 24 h. The water

samples were analyzed according to the Chinese Environmental Protection Agency standard methods (Chinese EPA 2002). Potassium dichromate method was used for COD determination; colorimetric method was used for NH₃-N and NO₃-N measurements. TN of soil was analyzed by Na₂CO₃ fusion method and TP was analyzed by Kjeldahl method.

The nitrifying and denitrifying bacteria in the soil samples were counted using the most probable number (MPN) calculation (Molle et al. 2006). The medium for the nitrifying bacteria contained per litre distilled water: 13.5 g Na₂HPO₄, 0.7 g KH₂PO₄, 0.1 g MgSO₄·7H₂O, 0.5 g NaHCO₃, 2.5 g (NH₄)₂SO₄, 14.4 mg FeCl₃·6H₂O and 18.4 mg CaCl₂·7H₂O, pH 8.0. The medium for the denitrifying bacteria contained per litre distilled water: 1.0 g KNO₃, 0.1 g Na₂HPO₄, 2.0 g Na₂S₂O₇, 0.1 g NaHCO₃ and 0.1 g MgCl₂, pH 8.0. The soil samples were taken from 0.3, 0.5, 0.7, 0.9 and 1.1 m depths, respectively. Aliquot (1 mL) of serial tenfold sterile distilled water dilutions of the soil samples were transferred to 96-cell microtiter plates containing each type of medium, then incubated at 28°C 14 d (for the nitrifying bacteria) and 15 d (for the denitrifying bacteria), respectively. Meanwhile, 10 g of soil samples were oven-dried at 105°C for 12 h to produce a constant weight. The amounts of the nitrifying and denitrifying bacteria were analyzed twice per month during the study. Urease, nitrate reductase (NAR) and nitrite reductase (NIR) activities were analyzed according to the method of Guan (1986) twice per month.

During the whole experimental period, intermittent operation mode was adopted as a passive method for oxygen transfer restoring in the SWIS (Hati et al. 2006; Kamitani and Kaneko 2007). Each cycle of the intermittent operation included a continuous flow period of 24 h (between 9:00 AM and 9:00 AM the next day) and a drying period of 24 h. Statistical analyses were carried out with MicroCal Origin 7.0 (OriginLab) and SPSS 17.0. The SWIS system was operated under hydraulic loading rate of 0.125 m³/(m²·d).

Results and Discussion

Purification processes in the SWIS were gradually established over 4 weeks. After which, soil samples were analyzed for the number of nitrifying and denitrifying bacteria at different depths and positions. As shown in Table 1, the amount of nitrifying bacteria declined with depths increasing. At the same time, the nearer to the distributing pipe, the higher amount. Meanwhile, the number of denitrifying bacteria increased with the depths increasing. Also, the more quantity of denitrifying bacteria achieved nearer the distributing area.

Table 1 Distribution of nitrifying and denitrifying bacteria in the SWIS

Sampling point	0.3		0.5		0.7		0.9		1.1	
	AN ^a	AD ^b	AN	AD	AN	AD	AN	AD	AN	AD
A	$(3.5 \pm 1.2) \times 10^5$	$(6.6 \pm 2.1) \times 10^6$	$(1.6 \pm 1.4) \times 10^5$	$(9.8 \pm 1.9) \times 10^6$	$(5.8 \pm 0.4) \times 10^4$	$(1.5 \pm 0.2) \times 10^7$	$(3.3 \pm 0.1) \times 10^4$	$(6.0 \pm 0.3) \times 10^7$	$(5.5 \pm 0.1) \times 10^3$	$(1.3 \pm 0.3) \times 10^8$
B	$(5.5 \pm 0.1) \times 10^6$	$(2.8 \pm 1.5) \times 10^7$	$(4.2 \pm 0.2) \times 10^6$	$(5.2 \pm 0.8) \times 10^7$	$(7.6 \pm 1.5) \times 10^5$	$(3.3 \pm 0.6) \times 10^8$	$(4.5 \pm 0.4) \times 10^5$	$(7.7 \pm 0.4) \times 10^8$	$(7.8 \pm 0.4) \times 10^4$	$(3.5 \pm 0.4) \times 10^9$
C	$(7.4 \pm 0.7) \times 10^7$	$(6.8 \pm 0.5) \times 10^8$	$(6.7 \pm 0.5) \times 10^7$	$(9.6 \pm 1.2) \times 10^8$	$(6.3 \pm 0.1) \times 10^6$	$(4.5 \pm 1.0) \times 10^9$	$(5.3 \pm 0.2) \times 10^6$	$(8.8 \pm 0.1) \times 10^9$	$(8.9 \pm 0.2) \times 10^5$	$(7.6 \pm 0.2) \times 10^{10}$
D	$(4.3 \pm 0.1) \times 10^6$	$(5.8 \pm 0.7) \times 10^7$	$(3.5 \pm 0.2) \times 10^6$	$(6.9 \pm 0.2) \times 10^7$	$(4.6 \pm 0.5) \times 10^5$	$(1.8 \pm 0.3) \times 10^8$	$(3.4 \pm 0.2) \times 10^5$	$(7.8 \pm 0.3) \times 10^8$	$(6.4 \pm 0.2) \times 10^4$	$(4.8 \pm 0.3) \times 10^9$
E	$(6.6 \pm 1.4) \times 10^5$	$(4.1 \pm 1.2) \times 10^6$	$(2.6 \pm 1.8) \times 10^5$	$(7.3 \pm 1.5) \times 10^6$	$(3.6 \pm 0.7) \times 10^4$	$(3.9 \pm 0.4) \times 10^7$	$(2.0 \pm 0.2) \times 10^4$	$(5.9 \pm 1.4) \times 10^7$	$(4.6 \pm 0.2) \times 10^3$	$(2.9 \pm 1.4) \times 10^8$

^a nitrifying bacteria number (MPN/g)

^b denitrifying bacteria number (MPN/g)

In the SWIS system, nitrification coupled with denitrification is generally thought to be the major method for nitrogen removal. For the fate of nitrification is significantly lower than that of the denitrification, so nitrification is a limiting step for nitrogen removal process. It was reported that for NH₃-N with a concentration of 1 mg/L, nitrification will not occur successfully unless the DO concentration reaches 4.6 mg/L (Molle et al. 2006). Herein, benefiting from the oxygen transferring availability of the surface soil, the zone between 0.3 and 0.7 m depth was the most effective nitrifying reaction region in the SWIS. The denitrifying bacteria were more active between depths of 0.7–1.5 m. Meanwhile, the results implied that flow path of the wastewater in the SWIS was: firstly, the wastewater flew out of the distributing pipe, then went up to the 0.3 m underneath under capillary force, and then diffused to the intervals of the distributing pipes, finally flew under gravity to the collecting pipe. This flow path in the SWIS system was similar to the studies reported before (Head and Oleszkiewicz 2004; Hsu et al. 2006; Babatunde et al. 2008).

From April 2007 to March 2009, the activities of urease, NAR and NIR were investigated, as shown in Table 2.

From Table 2, it can be concluded that urease activity was in positive correlation with the temperature. The correlation equation was $U = 0.3668T + 8.5679$ (correlation coefficient 0.933), where U and T represented the urease activity and temperature, respectively. The reports before informed that once the temperature increased by 10°C, the urease activity will be 1–2 times higher (Head and Oleszkiewicz 2004; Kim and Sansalone 2008). The higher urease activity was achieved nearer the inlet, 0.5 m underneath. NAR activity was influenced by the soil depth. The depth for NAR activity from high to low in sequence was 0.3, 0.5, 0.7, 0.9 and 1.1 m. The sequence for NIR activity from high to low in depth was 0.5, 0.3, 0.7, 0.9 and 1.1 m.

At the same time, NH₃-N and TN influent and effluent concentrations were analyzed and the removal efficiencies were calculated (Fig. 3). After 2 years operation, the average removal efficiencies were $88.7 \pm 1.2\%$ for NH₃-N and $76.2 \pm 1.5\%$ for TN. Compared with the activated sludge method and membrane filtration technology ($89.2 \pm 2.1\%$ and $78.5 \pm 3.4\%$ mean removal efficiencies for NH₃-N and TN) (Luanmanee et al. 2002; Walid and Al-Qodah 2006), the SWIS had comparable nitrogen removal efficiency. In the inflow, NH₃-N was the main form of TN, accounting for $83.3 \pm 1.1\%$. NO₃-N concentration in the influent was 0.2–0.3 mg/L, less than 1% of TN. In the outflow, NO₃-N concentration increased to 2.0–2.5 mg/L, 29.0–30.5% accounting for TN. On the contrary, NH₃-N concentration declined to 2.3–4.4 mg/L, accounting for 63.2–65.6% of TN. The NH₃-N and TN removal efficiencies achieved and

Table 2 Changes in enzyme activities in the SWIS (mg/g·d)

Depth (m)	Intervals											
	2007.04–2007.06 T: 18.7, C ₀ : 37.5		2007.07–2007.09 T: 28.4, C ₀ : 35.0		2007.10–2007.12 T: 15.8, C ₀ : 39.4		2008.01–2008.03 T: 10.6, C ₀ : 38.5					
	Urease	NAR	NIR	NAR	NIR	Urease	NAR	NIR				
0.3	14.05 ± 0.05	0.90 ± 0.01	0.55 ± 0.03	15.03 ± 0.79	0.93 ± 0.20	0.37 ± 0.05	0.88 ± 0.24	0.31 ± 0.01				
0.5	16.35 ± 0.12	0.77 ± 0.01	0.39 ± 0.14	17.37 ± 3.25	0.79 ± 0.14	0.41 ± 0.03	0.61 ± 0.70	0.36 ± 0.17				
0.7	21.88 ± 1.23	0.52 ± 0.05	0.30 ± 0.01	21.27 ± 0.96	0.60 ± 0.73	0.32 ± 0.16	0.47 ± 0.25	0.30 ± 0.02				
0.9	19.77 ± 2.08	0.43 ± 0.11	0.29 ± 0.10	19.93 ± 3.00	0.52 ± 0.01	0.30 ± 0.11	0.34 ± 0.41	0.25 ± 0.08				
1.1	15.32 ± 2.37	0.42 ± 0.03	0.25 ± 0.02	16.98 ± 1.07	0.48 ± 0.08	0.28 ± 0.06	0.38 ± 0.07	0.21 ± 0.01				
Depth (m)	2008.04–2008.06 T: 18.3, C ₀ : 40.1						2008.10–2008.12 T: 11.8, C ₀ : 36.7					
	Urease	NAR	NIR	Urease	NAR	NIR	Urease	NAR	NIR	Urease	NAR	NIR
0.3	15.55 ± 1.24	0.91 ± 0.07	0.34 ± 0.10	16.08 ± 1.10	0.91 ± 0.02	0.35 ± 0.04	11.23 ± 0.77	0.78 ± 0.02	0.33 ± 0.01	13.28 ± 2.14	0.66 ± 0.07	0.29 ± 0.04
0.5	18.56 ± 1.65	0.75 ± 0.03	0.35 ± 0.11	18.43 ± 1.28	0.77 ± 0.34	0.42 ± 0.06	13.46 ± 1.78	0.59 ± 0.04	0.36 ± 0.17	11.26 ± 2.33	0.48 ± 0.03	0.31 ± 0.18
0.7	22.09 ± 1.84	0.53 ± 0.01	0.29 ± 0.06	22.46 ± 0.93	0.55 ± 0.60	0.30 ± 0.09	14.33 ± 2.55	0.38 ± 0.01	0.30 ± 0.10	14.54 ± 1.35	0.41 ± 0.03	0.22 ± 0.03
0.9	21.37 ± 1.49	0.40 ± 0.17	0.26 ± 0.08	19.86 ± 1.65	0.53 ± 0.11	0.28 ± 0.11	12.08 ± 2.06	0.39 ± 0.04	0.26 ± 0.08	12.85 ± 3.27	0.39 ± 0.01	0.20 ± 0.10
1.1	16.76 ± 1.96	0.48 ± 0.04	0.23 ± 0.03	17.23 ± 0.88	0.51 ± 0.27	0.25 ± 0.07	11.55 ± 1.52	0.38 ± 0.03	0.10 ± 0.02	12.32 ± 1.66	0.30 ± 0.03	0.15 ± 0.03

^c Temperature (°C)^d Ammonia nitrogen concentration in the influent (mg/L)

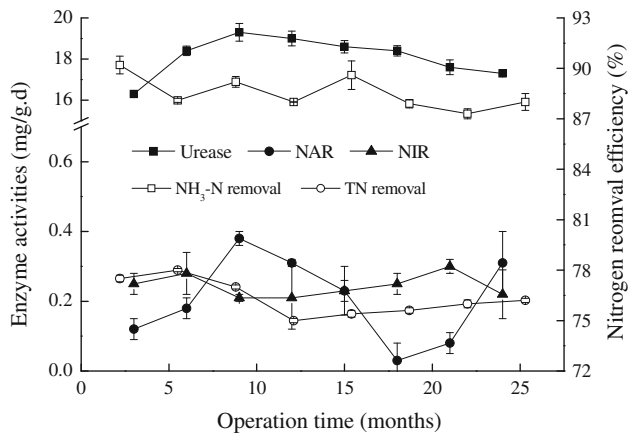


Fig. 3 Correlation between enzyme activities and nitrogen removal efficiency

TN composition analytical results suggested that the nitrification–denitrification process worked well in the SWIS. Therefore, SWIS was proved to be an effective technique for sewage treatment in areas without adequate domestic treatment facilities.

SPSS analysis of the Fig. 3 results implied that the urease and NIR activities were in positive correlation with the TN removal. The correlation equations were $U = 77.08R - 41.407$ for urease and $N = 2.2R - 1.4467$ for NIR, with correlation coefficients 0.8662 and 0.9140, respectively. In the equations, U , N and R represented the activities for urease and NIR, and TN removal efficiency. Therefore, the activities of urease and NIR could be the biological indexes during the nitrogen removal process in the SWIS.

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