## **Trifluralin Residues in Runoff and Infiltration Water from Tomato Production**

G. F. Antonious

Kentucky State University, Land Grant Program, Department of Plant and Soil Science, 218 Atwood Research Facility, Frankfort, KY 40601, USA

Received: 30 July 2003/Accepted: 22 January 2004

Runoff from agricultural watersheds carries enormous amounts of pesticides (Ray et al. 2002). During plant production, pesticides may move from application site into runoff water and runoff sediment following irrigation or natural rainfall. Rainfall intensity and flow rate are critical factors in determining a pesticide movement from application sites into surface runoff, rivers, and streams. Highest concentrations of pesticides are usually detected in the first runoff event after pesticide application (Keese et al. 1994; Antonious and Byers 1997). In recent years, there has been a major shift by U.S. farmers from plow tillage towards systems with reduced tillage. The reduced forms of tillage (conservation tillage) generally minimize soil erosion and water runoff, improve soil physical structure and productivity, but increase water infiltration rates into the vadose zone (Gaynor et al. 1995; Antonious 1999; 2000). Concern has been raised regarding the environmental soundness of conservation tillage because of higher use of pesticides and generally greater rates of water infiltration, leading to leaching of pesticides into groundwater (Antonious 2003a). Conservation tillage, therefore may increase the potential for pesticide leaching. Water infiltration through the soil profile provides the soil with the moisture content needed for seed germination and seedling establishment. Infiltration may also carry substantial amounts of pesticides (originating primarily from surface agricultural use) into the plant root area. However, infiltration (seeping) of pesticides through the soil into the vadose zone, the layer that connects the point of chemical input and surface practices to the groundwater zone (the saturated water zone), must be reduced to ensure the safety of groundwater used as a drinking water supply. Unfortunately, a low water infiltration rate accompanied by a heavy rain leads to flooding or runoff and erosion, particularly when agricultural operations occur on highly erodible lands. This distribution of pesticides throughout the soil profile, as a function of time, represents the integration of several processes such as mass flow, diffusion, adsorption/desorption, degradation, volatility, runoff, and plant uptake. One method of reducing overland flow by soil chemicals (e.g., nutrients, pesticides) is to reduce their concentration at the immediate soil surface. This can be done by incorporating the chemical at lower depths, either by tilling the soil or by irrigation shortly after chemical application and/or development of pesticides that have high degradation rates on the soil surface. Knowledge of how, when, and where pesticide residues reach the edge of the field and streams and how these residues can be minimized is essential in order to reduce the quantity of pesticides reaching surface water and the nation's water resources. Minimizing soil erosion is vital to long-term crop production. Therefore, proper soil management practice is an important key element in reducing soil erosion and pesticide movement. Composting provides an organic amendment useful to improve soil structure and nutrient status and generally stimulates soil microbial activity (Burriuso et al. 1997). It also increases pesticide sorption (Martinez and Almendros 1992; Guo et al. 1993), and decreases pesticide leaching (Zsolnay 1992). Trifluralin, a tertiary aromatic amine and dinitrotoluene



Figure 1. Chemical structure of trifluralin [Treflan, (2.6-dinitro-N, N-dipropyl-4-trifluoromethyl) benzenamine].

derivative (Figure 1), is widely used as a pre-emergence herbicide on a variety of field crops, fruits, vegetables, and ornamentals (Anonymous 2003) for control of annual grasses and broadleaf weeds. Due to its high vapor pressure  $(1.99 \times 10^4 \text{ mm})$ Hg at  $29.5^{\circ}$  C) and sensitivity to photochemical degradation, triflural has to be incorporated into the soil in order to avoid undesirable loss under field application (Francioso et al. 1992). Despite its low solubility in water,  $0.22 \text{ mg} L^{-1}$  at pH 7 (Anonymous 1995), triflural in was investigated in this study because of its intensive use in agriculture and its relatively high toxicity to fish (Anonymous 1994). Pesticide adsorption to soil is related to soil organic matter than other soil chemical and physical properties (Jacques and Harvey 1979; Patel 2002). The objectives of this investigation were 1) to study the effect of mixing soil with yard waste compost (having considerable amount of organic matter) and growing living fescue strips (across the contour of the land slope) on the movement of triflural in from soil into runoff and infiltration water, and 2) to study the impact of each of these two soil management practices (yard waste compost vs. living fescue strips) on tomato yield.

## **MATERIALS AND METHODS**

A field study was conducted on a Lowell silty loam soil (2.8% organic matter, pH 6.9) located at Kentucky State University Research Farm, Franklin County, KY. The soil has an average of 12% clay, 75% silt, and 13% sand. Plots (universal soil loss equation (USLE) standard plots) of  $22 \times 3.7$  m (n=18), on a soil of 10% slope were established. Plots were separated using metal borders 20 cm above the ground level to prevent cross contamination between treated and untreated plots. Three soil management practices were used 1) vard waste compost made from vard and lawn trimmings, and vegetable remains (produced at Kentucky State University Research Farm, Franklin County, KY) mixed with native soil at 50 t. acre<sup>1</sup> (on dry weight basis) with a plowing depth of 15 cm, 2) grass filter strips (tall fescue, Festuca

elatior), 3 feet wide, planted between every two tomato rows perpendicular to the land slope at 5 rows,  $plot^{-1}$  to slow runoff and trap sediment, and 3) no- mulch (NM) treatment (roto-tilled bare soil) for comparison purposes. Sixty day old tomato (Lycopersicon esculentum cv. Fabulous) seedlings were planted at 10 plants. row<sup>-1</sup>. Plots were irrigated by drip tape irrigation and no fertilizer was applied. Trifluralin  $(430 \text{ g.L}^{-1} \text{ EC};$  Treflan) was sprayed on May 31, 2000 and incorporated into the soil surface at the rate of 0.75 lb. acre<sup>-1</sup> using a 4-gallon portable backpack sprayer (Solo) equipped with one conical nozzle operated at 40 p.s.i. Soil samples (6 replicates per treatment) were collected at different time intervals  $(n=11)$  during 45 days following spraying to a depth of 15 cm from the field plots using a soil core sampler equipped with a plastic liner tube (Clements Associates, Newton, IA, USA) of 2.5 cm i.d. for maintenance of sample integrity. Soil samples were air-dried in the dark, sieved to a size of  $\leq 2$  mm. Ten-gram soil samples were used to determine soil moisture content to report the results on dry weight basis. Fifty g soil were shaken with 100 mL mixture of acetonitrile- water (99:1, v/v) for 1 hour using a Multi-wrist shaker (Lab-Line Instruments, Inc., Melrose Park, IL, USA). The solvent was filtered through Whatman 934-AH glass microfibre discs (Fisher Sci, Pittsburgh, PA) of 90 mm diameter, concentrated by rotary vacuum (Buchi Rotavapor Model 461, Switzerland) and  $N_2$  gas stream evaporation. A Supelco Envi-C<sub>18</sub> cartridge was conditioned first with 4 mL of 50% acetonitrile in water and then with isooctane (3 mL). Four mL of the soil extract were loaded onto the cartridge and passed through at a flow rate of 1-2 mL. min<sup>-1</sup>. Finally, the cartridge was eluted with 3 mL of hexane and 3 mL of isooctane, and the eluate was dried under a gentle stream of  $N_2$  gas (99.99% purity) and reconstituted in isooctane for GC/NPD determination.

Runoff (soil-water suspension) was collected and quantified at the lower end of each plot using tipping-bucket runoff metering apparatus (Department of Agricultural Engineering, University of Kentucky, Lexington, KY 40546, USA). Homogeneous samples of runoff were collected in amber borosilicate glass bottles and transported to the laboratory on ice in coolers. Sediment in runoff was determined by weighing the sediments collected from a 1-L sample of runoff using Whatman No.1 filter paper. Trifluralin was extracted from sediment samples using  $C_{18}$  cartridges as described in soil analysis. Total runoff water lost per runoff event, per each 0.02-acre plot was used to measure triflural in concentration, which in turn was dependent on natural rainfall events. To monitor the presence of trifluralin in the vadose zone (the unsaturated water layer below the plant root), pan-lysimeters (Department of Agricultural Engineering, University of Kentucky, Lexington, KY 40546, USA) were installed at the end of the experimental plots down the land slope at a depth of 1.5 m. Infiltration water was collected following each rainfall event in amber borosilicate glass bottles. To monitor triflural in concentration in runoff and infiltration water following each rainfall event, the pH of water samples was adjusted to 2.2 - 2.3 using 15 mL of 2 N HCl per liter of water. Duplicates of 500 mL aliquots were filtered through Whatman 934-AH glass microfibre discs using vacuum filtration. Trifluralin residues in 500 mL of the acidified water samples were extracted three times by liquid-liquid partition with 100, 60, and 40 mL of methylene chloride (CH<sub>2</sub>Cl<sub>2</sub>).  $CH_2Cl_2$  fractions (bottom layer) were combined and passed over anhydrous Na<sub>2</sub>SO<sub>4</sub>, then evaporated to dryness using  $N_2$  stream and reconstituted in isooctane. Trifluralin

residues were determined using a Hewlett Packard model 5890A Series II gas chromatograph (Hewlett Packard Co., Avondale, PA) equipped with a NP detector. Samples were injected onto a BD-5 high resolution column (15 m  $\times$  0.53 mm i.d.) with 0.5  $\mu$ m film thickness (J & W Scientific, Folson, CA). Operating conditions were 230, 250, and 280  $^{\circ}$ C for injector, oven, and detector, respectively. Carrier gas (He) flow rate was 5.2 mL. min<sup>-1</sup>. Peak areas were determined on a Hewlett Packard model 3396 series II integrator. Quantification was based on average peak areas of 1 µL injections obtained from external standards solutions of triflural in ranging from 0.1 to 15 ng.  $\mu L^{-1}$ . Under these conditions retention time (Rt) of triflural in was 6.88 min. Peak identity was confirmed by consistent retention time and coelution with standards under the conditions described. Trifluralin technical material of 96.3% purity was obtained from Dow AgroSciences (Indianapolis, IN, USA). Linearity over the range of concentrations was determined using regression analysis ( $R^2 = 0.99$ ). Standard solutions were used to spike blank soil samples for evaluating the reproducibility and efficiency of the analytical procedures to recover trifluralin. Recoveries (means  $\pm$  SE) of trifluralin from fortified water and soil samples were  $96.6 \pm 2.4$  and  $91.5 \pm 3.8$ , respectively. Quality control (OC) samples included three field blanks to detect possible contamination during sampling, processing, and analysis. The lack of trifluralin residues in the blank samples suggested that there was no contamination from sampling, processing, or laboratory procedures.

## **RESULTS AND DISCUSSION**

Trifluralin residues following application and incorporation into the 10-15 cm top soil, averaged from 0.01 to 0.05  $\mu$ g. g<sup>-1</sup> soil (Figure 2). Concentration of trifluralin was much higher in yard waste compost treatments than the NM treatments (rototilled bare soil). Yard waste compost contains significant concentrations of humic acid, the main constituent of soil organic matter (Patel 2002). Tavares and Rezende (1998) have indicated that functional groups in humic acid, namely carboxylic and phenolic groups appear to be the principal sites for the adsorption and interaction with trifluralin. This may explain why trifluralin residues were higher in compost-amended soil than NM soil.

Runoff water control is highest when living fescue strips are established as repeated barriers to runoff across the land slope. Living fescue strips reduced the volume of runoff water (Figure 3, upper graph) as a pesticide carrier and consequently reduced trifluralin transport and concentration in runoff water by 86.2% (Figure 3, lower graph) down the land slope. Triflural in residues were decreased as the runoff water flowed across the rough soil and grass surfaces along the hill slope that would otherwise have been transported down hill into surface runoff. Filter strips increased water infiltration into the vadose zone as indicated by volume of water collected from the vadose zone (Figure 4, upper graph). Previous results have indicated that living fescue strips reduced runoff but did not reduce leaching of  $\alpha$ -endosulfan (Antonious and Byers 1997) and dacthal into the vadose zone (Antonious 1999). On the contrary, this investigation provides evidence of the low potential leaching of trifluralin towards the vadose zone (Figure 4, lower graph). The low concentration of trifluralin in infiltration water from yard waste compost indicates that the risk of groundwater contamination will be low. This can be explained by the adsorption properties of trifluralin on humic acids in compost-amended soil. Due to mechanical incorporation of the pesticide in the top 10-15 cm of soil, an equilibrium is usually established between the pesticide adsorbed to the soil and that in solution. This equilibrium reduces the transport and movement of strongly adsorbed pesticides such as trifluralin (Baker et al. 2000).



**Figure 2.** Trifluralin residues and organic matter content of soil collected from the rhizosphere of tomato plants grown under three soil management practices. Statistical comparisons were done between three soil management practices for each parameter. Bars accompanied by different letter are significantly different ( $P < 0.05$ ) from each other using the SAS procedure (SAS Institute, 1999, Duncan's multiple range test).

Yard waste compost is very rich in nutrients (Antonious et al. 2003) and organic matter (Figure 2). Treatments high in organic matter produced high tomato yield (Figure 5). This is because organic substances and nutrients in compost support a vast population of soil organisms that "mine" for soil minerals. Evidence of enhanced microbial activity in the rhizosphere of plants grown with yard waste compost has been reported (Anderson et al. 1994; Antonious 2003b). Much of the effects of application of compost on crop yield is derived from availability of nutrients particularly N in compost. Availability of soluble P had also increased following addition of compost (Swiader and Morse 1984). The competition between living fescue strips and tomato plants on the soil nutrients apparently is the cause of the reduced total tomato yield in fescue treatments. Living fescue strips therefore, may not justify the cost of the occupied soil surface used for growing fescue. However, fescue strips reduced runoff water and triflural in residues in runoff. This may justify the cost of possible environmental pollution by triflural and its high toxicity to fish. Yard waste compost reduced trifluralin residues in surface runoff water from June and July rainfall by 76 and 84%, respectively (Figure 3). The sediment masses in runoff were 0.5 to 8.6 g.  $L^{-1}$  with higher amounts in July sampling and lower amounts in June sampling. This corresponds to rainfall intensity during the sampling periods. Rainfall was 5.1 and 6.5 cm with an intensity of 0.9 and 1.3 cm. hr<sup>-1</sup> on June 19 and July 14, 2000, respectively. Trifluralin concentrations in runoff sediment (data not shown) collected from the three soil management practices at the time of the runoff events showed the same trend of triflural in adjoining soil samples. Since pesticide



Figure 3. Volume of runoff water (upper graph) and trifluralin residues detected in runoff water (lower graph) from tomato grown on erodible land under three soil management practices. Statistical comparisons were done between three soil management practices for each runoff event. Bars accompanied by different letter for each runoff event are significantly different ( $P < 0.05$ ) from each other using the SAS procedure (SAS Institute, 1999, Duncan's multiple range test).



Figure 4. Infiltration water (upper graph) and trifluralin residues detected in infiltration water (lower graph) collected from the vadose zone of tomato plants grown under three soil management practices. Statistical comparisons were done between three soil management practices for each sampling date. Bars accompanied by different letter(s) for each date are significantly different ( $P$  < 0.05) from each other using the SAS procedure (SAS Institute, 1999, Duncan's multiple range test).



**Figure 5.** Yield of tomato grown on erodible land with drip irrigation under three soil management practices. Statistical comparisons were done between three soil management practices for each harvest and total yield. Bars accompanied by different letter(s) for each harvest or total yield are significantly different ( $P<$ 0.05) from each other using the SAS procedure (SAS Institute, 1999, Duncan's multiple range test).

adsorption to soil is related more to soil organic matter than to other soil chemical and physical properties (Jacques and Harvey 1979; Sparks 1995; Patel 2002), therefore, addition of soil amendments like yard waste compost (having high organic matter content) to native soil will increase the adsorption of trifluralin and reduce its surface and vertical movement under field conditions. Therefore, pesticides strongly adsorbed to soil amended with compost are much better candidates for this method of soil management.

Acknowledgments. I thank C. Lee, G. Patel, and M. Stone for their kind help in farm operations. This investigation was supported by a grant from USDA/ CSREES to Kentucky State University under agreements No. KYX-9803027 and No. KYX-10-03-37P.

## **REFERENCES**

- Anderson TA, Kruger EL, Coasts JR (1994) Biochemical degradation of pesticides wastes in the roor zone of soils collected at an agrochemical dealership. In: Bioremediation Through Rhizosphere Technology, pp. 199-209. American Chemical Society, Washington, DC.
- Anonymous (1994) The Royal Society of Chemistry. The Agrochemical Handbook, Third Edition, update 5, January 1994, Cambridge CB4 4WF, UK.
- Anonymous (1995) The Pesticide Manual: The Agrochemicals Handbook, British Crop Protection Council, Tenth Edition, Tomblin C., Ed., Farnham, UK.
- Anonymous (2003) Commercial Vegetable Crop Recommendations, Cooperative Extension Service, University of Kentucky, College of Agriculture, ID-36.
- Antonious GF, Byers ME (1997) Fate and movement of endosulfan under field conditions. J Environ Toxicol Chem 16: 644-49.

Antonious GF (1999) Efficiency of grass buffer strips and cropping system on offsite dacthal movement. Bull Environ Contam Toxicol 63: 25-32.

- Antonious GF (2000) Clomazone residues in soil and runoff: Measurement and mitigation. Bull Environ Contam Toxicol 64: 168-175.
- Antonious GF (2003a) Soil infiltration by pesticides. In: Encyclopedia of Pest Management. Pimentel D (ed), volume 3, Marcel Dekker Inc., New York, pp. 1-4.
- Antonious GF (2003b) Impact of soil management and two botanical insecticides on urease and invertase activity. J Environ Sci Health 38 : 479-488.
- Antonious GF, Patterson MA, Snyder JC (2003) Pesticide Residues in soil and quality of potato grown with sewage sludge. Bull Environ Contam Toxicol 71:  $315 - 322$
- Baker JL, Kickelson K, Arora K, Misra AK (2000) The potential of vegetative filter strips to reduce pesticide transport. In: Agrochemical Fate and Movement, Steinheimer TR, Ross LJ, and Spitter TD (Eds.), pp. 272-285, ACS Symposium Series, American Chemical Society, Washington, DC.
- Burriuso E, Houot S, Serra-Wittling C (1997) Influence of compost addition to soil on the behaviour of herbicides. Pestic Sci 49:65-75.
- Francioso O, Bak E, Rossi N, Sequi P (1992) Sorption of atrazine and trifluralin in relation to the physico-chemical characteristics of selected soils. Sci Tot Environ 123/124: 503-512.
- Gaynor JD, MacTavish DC, Findlay WI (1995) Atrazine and metalachlor loss in surface and subsurface runoff from three tillage treatments in corn. J Environ Qual 24: 246-56.
- Guo L, Bicki TJ, Felsot AS, Hinesly TD (1993) Sorption and movement of alachlor in soil modified by carbon-rich wastes. J Environ Qual 22: 186-194.
- Jacques GL, Harvey RG (1979) Adsorption and diffusion of dinitroaniline herbicide in soils. Weed Sci 27: 450-455.
- Keese RJ, Camper ND, Whitwell T, Riley MB, Wilson PC (1994) Herbicide runoff from ornamental container nurseries. J Environ Qual 23: 320-24.
- Martinez MJ, Almendros G (1992) Pesticide sorption on soils treated with evergreen oak biomass at different humification stages. Commun Soil Sci Plant Anal 23: 1717-1729.
- Patel G (2002) Interactions of pyrethrins and piperonyl butoxide with soil organic matter. MS thesis, University of Kentucky, Department of Horticulture, Lexington, KY 40546 (USA).
- Ray C, Soong TW, Lian YO, Roadcap GS (2002) Effect of food-induced chemicals load on filtrate quality at bank filtration sites. J Hydrol 266: 235-258
- SAS Institute (1999) SAS/STAT Guide, Release 6.03 Edition. SAS Institute, Campus Drive, Cary, NC 27513.
- Spraks DL (1995) Chemistry of soil organic matter. Chaper 3, pp. 53-79. In: Environmental Soil Chemistry, Academic Press, San Diego, CA.
- Swiader JM, Morse RD (1984) Influence of organic amendments on phosphorus requirement. J American Soc Hort Sci 109: 150-155.
- Tavares MC, Rezende MO (1998) Effect of humic acid on the sorption of trifluralin by soils. J Environ Sci Health B33: 749-767.
- Zsolnav A (1992) Effect of an organic fertilizer on the transport of the herbicide atrazine in soil. Chemosphere 24: 663-669.