

Association of variants in the fat mass and obesity associated (*FTO*) gene with polycystic ovary syndrome

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Received: 26 January 2008 / Accepted: 8 April 2008 / Published online: 14 May 2008
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Abstract

Aims/hypothesis Variants in the fat-mass and obesity-associated gene (*FTO*) influence susceptibility to type 2 diabetes via an effect on adiposity/obesity. Given the important role of obesity in the aetiology of both polycystic ovary syndrome (PCOS) and type 2 diabetes mellitus, our aim was to establish whether *FTO* variants are also implicated in PCOS susceptibility.

Methods We performed a genetic association study of *FTO* variant rs9939609 using case–control analyses, conducted in 463 PCOS patients (geometric mean BMI 27.5 kg/m²) and 1,336 female controls (geometric mean BMI 25.3 kg/m²) of UK British/Irish origin. We also sought evidence for associations between *FTO* variation and circulating testosterone levels in 324 UK PCOS patients and 1,000 women from the Northern Finland Birth Cohort of 1966. Outcome

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measures included *FTO* rs9939609 genotype frequencies by participant group and androgen measures (testosterone, free androgen index) by genotype.

Results There was a significant association between *FTO* genotype and PCOS status in the UK case–control analysis, which was attenuated by adjustment for BMI (Cochran–Armitage test, odds ratio [per minor allele copy] 1.30 [95% CI 1.12, 1.51], $p=7.2\times 10^{-4}$ [unadjusted], $p=2.9\times 10^{-3}$ [adjusted]). This association was most evident in obese PCOS patients (PCOS patients below median BMI vs UK controls, $p=0.11$; above median BMI vs controls, $p=2.9\times 10^{-4}$). No relationship between *FTO* genotype and androgen levels was seen.

Conclusions/interpretation We provide the first evidence that variants that predispose to common obesity also result in altered susceptibility to PCOS, confirming the mechanistic link between these conditions. The predominant effect of *FTO* variants on PCOS susceptibility is probably mediated through adiposity.

Keywords Androgens · *FTO* variants · Obesity · Polycystic ovary syndrome · Type 2 diabetes mellitus

Abbreviations

| | |
|----------|--|
| FAI | free androgen index |
| NFBC1966 | Northern Finland Birth Cohort of 1966 |
| OBB | Oxford Biobank |
| PCOS | polycystic ovary syndrome |
| SHBG | sex hormone binding globulin |
| UKBS | UK Blood Services |
| WTCCC | Wellcome Trust Case Control Consortium |

Introduction

Polycystic ovary syndrome (PCOS) is a common female endocrinopathy with a prevalence of between 5% and 7% in premenopausal women, and frequently co-exists with both obesity and type 2 diabetes mellitus [1–3]. Substantial evidence implicates obesity as an important factor in the aetiology of PCOS [4–7], though the mechanistic links between these conditions and how they interact with type 2 diabetes mellitus are poorly understood. Both obesity and PCOS are highly heritable [8–10], raising the possibility that a shared genetic predisposition contributes to their co-occurrence. However, until recently there has been little success in the identification of susceptibility genes for either condition [11, 12].

Recent work has shown that common variants in the fat mass and obesity associated gene (*FTO*) influence susceptibility to type 2 diabetes mellitus via a substantial effect on BMI and fat mass, establishing this as the first gene with a

robust effect on individual risk of common polygenic obesity [13, 14]. Though the causal variant has yet to be identified, the association signal maps to a 47 kb region in the first intron of *FTO*. In the UK, the strongest associations are seen for a cluster of highly correlated single-nucleotide polymorphisms (SNPs), of which rs9939609 is representative. These variants are associated with a median per-allele difference in BMI of approximately 0.36 kg/m² and susceptibility to type 2 diabetes mellitus (odds ratio 1.27) [13].

Given the effect of *FTO* on adiposity and susceptibility to the development of type 2 diabetes mellitus, the close relationship between obesity and PCOS, and the common pathophysiological and epidemiological features shared by type 2 diabetes mellitus and PCOS, we set out to establish whether *FTO* variants also impact on the individual risk of PCOS. We did so in the belief that detecting such an association would provide further evidence (complementing that from epidemiology and studies of weight loss intervention) that obesity and PCOS are causally related, as well as providing evidence that genome sequence variation can influence predisposition to PCOS. A further aim was to establish whether *FTO* variation influences circulating levels of testosterone in women with PCOS, given that hyperandrogenaemia is the most consistent endocrine feature and forms part of the diagnostic criteria for PCOS [7, 15].

Methods

Participants We studied 463 UK PCOS patients and 1,336 UK female controls. All UK European British/Irish patients had a confirmed diagnosis of PCOS according to the consensus Rotterdam criteria [15]. More specifically, all UK patients had ultrasound-confirmed polycystic ovarian morphology [16] in addition to a history of hyperandrogenism (defined as previously described, other aetiologies having been excluded) [17] and/or oligo-amenorrhoea (intermenstrual interval >42 days). The vast majority of PCOS patients ($n=459$) had no known history of type 2 diabetes mellitus. All PCOS patients were non-pregnant, and were recruited from St Mary's and Middlesex (University College) Hospitals in London (as previously described) [18], or from Oxford-based endocrine clinics ($n=61$) using the same ascertainment criteria.

Women from two UK population-based control groups (all of European British/Irish origin) were genotyped. These included 565 women in the Oxford Biobank (OBB), drawn from the UK National Health Service population register. Genotypes were also available from 771 women in the UK Blood Services (UKBS) controls, set up by the Wellcome Trust Case Control Consortium (WTCCC) in collaboration

with the UK Blood Services. Height and weight measurements were recorded for all OBB female controls and for 769 UKBS control women (the latter self-reported).

We also examined the relationship between *FTO* genotype and androgen levels (as a continuous trait relevant to PCOS status) in 324 of the UK cases and in 1,000 women from the population-based Northern Finland Birth Cohort of 1966 (NFBC1966, described previously [6, 19–21]), in whom there were available measures of both testosterone and *FTO* genotype. None of these women was on oral hypoglycaemic agents (including metformin) or hormonal therapy, and none was pregnant at the time of study. In addition, none of the women examined from the NFBC1966 had described any symptoms of PCOS in a questionnaire completed at the age of 31 years.

Clinical details of all cases and control groups are shown in Table 1. Serum testosterone and sex hormone binding globulin (SHBG) concentrations were measured as previously described [18] and the free (unbound) androgen index (FAI) was calculated as total testosterone (nmol/l) × 100/SHBG (nmol/l). All clinical investigations were conducted in accordance with the guidelines in the Declaration of Helsinki. All participants provided fully informed written consent and the study was approved by the relevant ethics committees in the UK and Finland.

Genotyping For the UK patients and NFBC1966 women, genotyping of *FTO* rs9939609 was performed using a Taqman assay on demand method (assay ID, C_30090620_10; Applied Biosystems, Warrington, UK). For the OBB control group, genotyping of *FTO* rs9939609 was performed by Kbiosciences (Hoddesdon, UK; <http://www.kbioscience.co.uk>) using a fluorescence-based competitive allele-specific (KASPar) assay. For the UKBS control group, *FTO* rs9939609 genotypes were obtained

from the Affymetrix GeneChip Human Mapping 500k Array Set as part of the WTCCC study, as previously reported [13]. We have established high concordance rates (>99.8%) between rs9939609 genotypes for the same samples generated on these platforms, and (>99.9%) for duplicate genotypes on the same (Taqman) platform. Genotype success rates exceeded 96% in all samples and there were no departures from Hardy–Weinberg equilibrium ($p > 0.05$). Details of all assays are available on request.

Statistical analyses and power calculations Genotype frequency comparisons were conducted using the Cochran–Armitage (additive) test (StatXact v.6; Cytel, Cambridge, MA, USA). Following appropriate distributional logarithmic transformations, one-way ANOVA (conducted in SPSS [v12.0; SPSS, Chicago, IL, USA]) was used for quantitative trait analyses (testosterone and FAI). Testosterone levels were optionally adjusted for BMI (where available). It is important to realise that, in the UK case–control analysis, the use of population-based controls (as opposed to controls in whom a diagnosis of PCOS has been excluded by clinical examination) results in only a modest loss of power and that this can be overcome easily (as here) by increasing the number of controls [22]. As there was no evidence of heterogeneity between genotype counts between women in the two UK control groups ($p = 0.91$), our primary UK case–control analyses were based on a comparison of cases with the (combined) female controls.

Power calculations were performed using Quanto v.0.5.5 (log-additive model) (<http://hydra.usc.edu/GxE/>, accessed April 2008). In the case–control analyses, the available UK sample sizes provided 91% power to detect an allelic odds ratio of 1.3 at an α value of 0.05. For the Finnish (NFBC1966) women whose data were used for the quantitative trait (testosterone) analyses, we had 80%

Table 1 Clinical characteristics of UK and Finnish NFBC1966 participants

| | UK PCOS patients | OBB female controls | UKBS female controls | NFBC1966 women |
|--------------------------|-------------------------------|-----------------------------|--------------------------|-----------------------------|
| Number | 463 ^a | 565 | 771 | 1,000 ^b |
| Age (years) | 32.3 ± 7.0 ^c | 41.5 ± 6.3 ^c | 41.6 ± 12.7 ^c | 31 ^d |
| BMI (kg/m ²) | 27.5 (21.2, 35.7) | 25.2 (21.3, 29.8) | 25.4 (21.5, 30.0) | 24.0 (20.0, 28.8) |
| WHR | 0.79 (0.72, 0.87) | 0.80 (0.73, 0.86) | Not known | 0.81 (0.74, 0.89) |
| Testosterone (nmol/l) | 2.1 (1.4, 3.1) ^e | Not known | Not known | 1.9 (1.2, 2.9) |
| Free androgen index | 6.0 (2.9, 12.4) ^e | Not known | Not known | 3.3 (1.7, 6.3) |
| Glucose (mmol/l) | 4.8 (4.2, 5.4) ^{e,f} | 5.0 (4.6, 5.4) ^f | Not known | 4.9 (4.3, 5.6) ^f |

Quantitative data are presented as geometric mean (SD range) unless otherwise stated. All participants were female

^aUK non-pregnant PCOS patients

^bExcluding women on oral hypoglycaemic agents, metformin or hormonal therapy (oral contraception or hormonal intrauterine device), women pregnant at the time of examination and women with symptoms of PCOS described in the questionnaire completed at the age of 31 years

^cMean ± SD

^dAll women in the NFBC1966 were sampled at the age of 31 years

^eExcluding women on oral hypoglycaemic agents, metformin or hormonal therapy (oral contraception)

^fFasting samples

Table 2 Case–control association analyses for the relationship between variants at *FTO* rs9939609 and PCOS in UK samples

| | <i>n</i> | TT | AT | AA | <i>p</i> value vs PCOS patients ^a |
|-----------------------------|----------|-------------|-------------|-------------|--|
| Alleles in UK PCOS patients | 463 | 133 (28.7%) | 231 (49.9%) | 99 (21.4%) | |
| Alleles in controls | | | | | |
| OBB women | 565 | 204 (36.1%) | 269 (47.6%) | 92 (16.3%) | 4.6×10^{-3} |
| UKBS women | 771 | 276 (35.8%) | 375 (48.6%) | 120 (15.6%) | 1.7×10^{-3} |
| Combined women | 1,336 | 480 (35.9%) | 644 (48.2%) | 212 (15.9%) | 7.2×10^{-4} |

Data shown are genotype counts (and percentages). A allele is associated with an increase in BMI. *p* values represent Cochran–Armitage test results

^a Comparison with UK PCOS patients

power to detect a trait difference between each genotype group (per minor allele copy) exceeding 25.1% of a standard deviation (for $\alpha=0.05$).

Results

Minor allele frequencies for *FTO* rs9939609 in the UK PCOS patients and combined female control groups were 46.3 and 40.0% respectively. Genotype frequency comparisons revealed a significant association of the minor (A) allele with PCOS (Cochran–Armitage test, odds ratio [per minor allele] 1.30, 95% CI 1.12, 1.51, $p=7.2 \times 10^{-4}$; Table 2). The expected relationship between *FTO* genotype and BMI was observed in the UK patients (per allele difference in BMI 1.1 kg/m², 95% CI –0.9, 3.2, $p=0.05$), but less obvious in the controls (per allele difference in BMI 0.5 kg/m², 95% CI –0.1, 1.3, $p=0.33$). Following adjustment for BMI in the comparison between UK patients and the combined control group, the association with PCOS was attenuated but not eradicated ($p=2.9 \times 10^{-3}$). Separate subgroup analyses were performed in UK patients with known BMI ($n=430$),

stratified according to whether their BMI was above or below the median for the case group (26.0 kg/m²). Minor allele frequencies for UK patients in the high- and low-BMI strata were 49.3 and 44.0% respectively, with only the former stratum significantly associated with PCOS when compared with the controls (high-BMI patients, $p=2.9 \times 10^{-4}$; low-BMI patients, $p=0.11$).

Analyses of testosterone and FAI between *FTO* rs9939609 genotype groups were conducted separately in UK patients ($n=324$), and women from the NFBC1966 ($n=1,000$), following exclusion of those taking hormonal therapy, metformin or any other oral hypoglycaemic agents. No significant overall trend was shown for testosterone or FAI in either the UK patients or the NFBC1966 groups (Table 3).

Discussion

This study, involving >1,700 women from the UK and 1,000 from Finland, is the first to demonstrate that variation within the *FTO* gene is significantly associated with PCOS. Although the levels of significance attained are not conclusive on the

Table 3 *FTO* rs9939609 genotypes and analyses of androgen measures

| | TT | AT | AA | Total | <i>p</i> value | <i>p</i> value adjusted for BMI |
|-----------------------|-----------------|-----------------|-----------------|-------|----------------|---------------------------------|
| UK patients | | | | | | |
| Number ^a | 96 | 160 | 68 | 324 | | |
| Testosterone (nmol/l) | 2.1 (1.4, 3.2) | 2.1 (1.4, 3.0) | 2.3 (1.5, 3.4) | | 0.31 | 0.75 |
| FAI ^b | 6.2 (3.1, 12.5) | 5.5 (2.8, 11.8) | 6.9 (3.5, 14.2) | | 0.24 | |
| NFBC1966 | | | | | | |
| Number ^c | 375 | 453 | 172 | 1,000 | | |
| Testosterone (nmol/l) | 1.9 (1.3, 2.9) | 1.8 (1.2, 2.8) | 1.8 (1.2, 2.9) | | 0.22 | 0.05 |
| FAI ^b | 3.5 (1.7, 6.2) | 3.5 (1.7, 6.2) | 3.5 (1.7, 6.6) | | 0.05 | |

A allele is associated with an increase in BMI. Testosterone and FAI values are expressed as geometric mean (SD range). FAI was not adjusted for BMI, given the high correlation between BMI and SHBG concentration

^a All UK PCOS British/Irish patients excluding women on oral hypoglycaemic agents (including metformin) or hormonal therapy

^b Free androgen index calculated as total testosterone (nmol/l)/SHBG (nmol/l) $\times 100$

^c Excluding women on oral hypoglycaemic agents (including metformin), hormonal therapy, women pregnant at the time of examination and women with symptoms of PCOS described in the questionnaire completed at the age of 31 years

genome-wide scale (which would require $p < 10^{-7}$), the strong prior claims for *FTO* (given its incontrovertible association with fat mass and type 2 diabetes mellitus) [13, 14] make this one of the first convincing claims for a relationship between genome sequence variation and PCOS susceptibility.

It seems likely that the effect of *FTO* on PCOS susceptibility is mediated through its effect on fat mass and the risk of obesity, in a fashion analogous to the relationship between *FTO* variation and predisposition to type 2 diabetes mellitus [13]. In line with previous studies (involving over 40,000 participants) [13, 14, 23], we found evidence that *FTO* variants influenced adiposity levels in our sample, though this effect was only nominally significant in the patients. Given previous large-scale studies that have shown no evidence for heterogeneity of *FTO* effect size between samples of diverse origin, including the same controls as used here [13], we regard the failure to detect a relationship between *FTO* rs9939609 and adiposity in the controls as entirely consistent with sampling error (indeed, the 95% CI range for the *FTO* effect in controls includes the mean effect size seen in previous papers) [13, 14, 23].

In the present study, and in contrast to the situation in type 2 diabetes mellitus [13], the PCOS association was not fully abolished by adjusting for BMI. This may well reflect sampling error, and the inadequacy of a single measure of weight and height as an index of chronic excess adiposity. Evidence that the case–control association was far weaker when restricted to the leaner PCOS patients supports this interpretation. To the extent that adjustment for BMI diminishes the PCOS association, our data provide further evidence, complementing that from epidemiology and the effects of weight reduction on PCOS risk [4–7], that obesity and PCOS are causally related. Since the mechanism(s) by which *FTO* variants affect fat mass are not currently known, other possible causal relationships (such as a direct effect of *FTO* variants on the development of PCOS, independent of the effects on fat mass) cannot be excluded, although a distinct non-adiposity effect of *FTO* variants on PCOS susceptibility would seem unlikely. Elucidation of the biological role and mechanism(s) of action of *FTO* variants with respect to weight and obesity should help to confirm how they influence PCOS susceptibility.

In summary, we have shown that variants in *FTO* which are known to influence the metabolic profile through effects on fat mass, BMI and obesity [24], and which are known to affect the risk of type 2 diabetes mellitus, are also associated with PCOS in UK women. The most likely explanation for this finding is an indirect effect on PCOS risk, mediated through an effect on fat mass and resulting in deleterious metabolic consequences. This would be consistent with the data [24] that indicate that the additional fat mass associated with *FTO* variation exerts metabolic effects

which are in line with the epidemiologically defined relationships between fat mass and direct and indirect measures of insulin resistance. *FTO* is one of the first genes for which a robust effect of genome sequence variation on PCOS risk can be claimed. Whether or not *FTO* can be labelled as a PCOS susceptibility gene as opposed to a gene that purely influences fat mass is essentially a semantic argument, analogous to that regarding the status of *FTO* as a susceptibility gene for type 2 diabetes mellitus.

Acknowledgements This work was funded by grants and fellowships from the Wellcome Trust (GR069224MA, 068545/Z/02), the UK Medical Research Council (G9700120, G0000934), the Academy of Finland (M.-R. Järvelin) and NovoNordisk (Clinical Research Fellowship to T. M. Barber). We acknowledge the many patients, relatives, nurses and physicians who contributed to the ascertainment of the various clinical samples used in this study, and the WTCCC.

Duality of interest The authors declare that there is no duality of interest associated with this manuscript.

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